



## Original article

# Weight loss program is associated with decrease $\alpha$ -tocopherol status in obese adults



Jadwiga Hamułka, Magdalena Górnicka\*, Agnieszka Sulich, Joanna Frąckiewicz

Department of Human Nutrition, Faculty of Human Nutrition and Consumer Sciences, Warsaw University of Life Sciences – WULS-SGGW, Warsaw, Poland

## ARTICLE INFO

## Article history:

Received 4 December 2017

Accepted 14 July 2018

## Keywords:

Weight loss  
 $\alpha$ -tocopherol status  
 Adiposity  
 Obese adults

## SUMMARY

**Background & aims:** Studies on changes in plasma  $\alpha$ -tocopherol levels during body fat reduction in obese persons are not clear. The aim of the present study was to assess factors associated with  $\alpha$ -tocopherol status in obese people and to examine changes in  $\alpha$ -tocopherol status after a 6-week AntioxObesity weight loss program.

**Methods:** The study was conducted in 60 overweight or obese adults, aged 18–54 years old. Food intake data were collected using the 3-day record method and a semi-quantitative food-frequency questionnaire. Anthropometric measurements included: height (H), body weight, waist circumference (WC) and hip circumference (HC), body composition: fat mass (FM) and fat-free mass (FFM), subcutaneous fat (SF) and visceral fat (VF). Lipid profile,  $\alpha$ -tocopherol concentration, glutathione peroxidase (GPx) activity, total antioxidant capacity (TAC) in plasma and superoxide dismutase (SOD) activity in erythrocytes were determined.

**Results:** Energy, fat, and carbohydrate intakes decreased significantly in all subjects ( $P < 0.001$ ). Body weight, WC, body mass index (BMI), waist-to-height ratio (WHtR), and FM, VF and SF decreased significantly during the 6 weeks in all subjects. Plasma  $\alpha$ -tocopherol significantly decreased during the program ( $P = 0.006$ ). No changes were observed for SOD activity, but GPx activity and TAC decreased significantly ( $P = 0.001$ ;  $P = 0.023$ , respectively). Plasma  $\alpha$ -tocopherol concentration after 6 weeks of the AntioxObesity program was strongly associated with baseline plasma  $\alpha$ -tocopherol, changes in TC, VF and FM. Low  $\alpha$ -tocopherol status ( $<20 \mu\text{mol/L}$ ) was found in 78% of the women and 68% of the men, after 6 weeks of the AntioxObesity program. Men were characterized by a greater decrease in weight, BMI, WC, FM, VF, SF and TAC compared to women.

**Conclusions:** A 6-week weight loss program lowered  $\alpha$ -tocopherol status in overweight and obese people. Low baseline  $\alpha$ -tocopherol status and adiposity in obese adults negatively affected  $\alpha$ -tocopherol status after 6 weeks weight loss program. These results, coupled with excessive weight and low  $\alpha$ -tocopherol intake, led to the finding that there was an increased risk of oxidative stress diseases in adults on a reduced diet. Long-term dietary restriction program for obese patients should be monitored to avoid  $\alpha$ -tocopherol deficiency, and take into account higher dietary  $\alpha$ -tocopherol requirements for obese people.

© 2018 Elsevier Ltd and European Society for Clinical Nutrition and Metabolism. All rights reserved.

**Abbreviations:** BMI, body mass index; FFM, fat-free mass; FM, fat mass; GPx, glutathione peroxidase; HDL, high-density lipoprotein; LDL, low-density lipoprotein; SF, subcutaneous fat; SOD, superoxide dismutase; TAC, total antioxidant capacity; TC, total cholesterol; TAG, triacylglycerol; VF, visceral fat; WC, waist circumference; WHR, waist-hip ratio; WHtR, waist-to-height ratio.

\* Corresponding author. Department of Human Nutrition, Faculty of Human Nutrition and Consumer Sciences, Warsaw University of Life Sciences – SGGW, Nowoursynowska 159C, 02-776 Warsaw, Poland.

E-mail address: [magdalena\\_gornicka@sggw.pl](mailto:magdalenagornicka@sggw.pl) (M. Górnicka).

## Introduction

Obesity, due to its increasing prevalence in recent years, has become a public health problem, perceived as a global epidemic. Obesity is defined as a disease associated with chronic inflammation, manifesting in abnormal secretion of cytokines, proteins, and mediators of immune response with the activation of inflammatory signaling pathways [1–4]. Excessive body fat causes a series of metabolic (hormonal and immunological) disorders that lead to diabetes, hypertension, and hyperlipidemia (metabolic syndrome), which in turn can accelerate atherosclerosis and increase the risk of

cardiovascular disease. Moreover, chronic inflammation enhances oxidative stress (an impaired balance between oxidants and antioxidants in the body) associated with hyperglycemia [5–7].

Excessive body fat, by exacerbating inflammation and oxidative processes, is linked to decreased plasma lipophilic micronutrients levels and antioxidant capacity [8–11].  $\alpha$ -Tocopherol, like other lipophilic compounds, such as vitamins A, D, and carotenoids is stored in adipose tissue, which affects its distribution to other tissues and their concentration in the blood.  $\alpha$ -Tocopherol is one of eight forms of vitamin E. These are 4 tocopherols and 4 tocotrienols, which are a lipid-soluble and have antioxidant activity related to the protection of lipids from oxidation [12].  $\alpha$ -Tocopherol, is antioxidant with lipoperoxyl radical-scavenging activities and has potentially anti-inflammatory functions [13].  $\alpha$ -Tocopherol, especially the RRR-stereoisomer form, is the most biological active form of vitamin E. It is preferentially absorbed and incorporated into chylomicrons in the small intestine and via the thoracic lymph duct is delivered to the liver, where it is selectively incorporated into very-low-density lipoproteins (VLDL) and low-density lipoproteins (LDL) by the hepatic  $\alpha$ -tocopherol transfer protein and resecreted into the blood-stream. VLDL and LDL exchanges  $\alpha$ -tocopherol with high-density lipoproteins (HDL), which seems to be the most effective  $\alpha$ -tocopherol donor for cellular uptake. Plasma cholesterol, LDL and HDL are the major carriers of  $\alpha$ -tocopherol in the circulation and plasma  $\alpha$ -tocopherol concentration correlates with plasma lipids level [12,14]. Besides lipids protection from oxidation, it was found that  $\alpha$ -tocopherol is involved in the expression of genes associated with glucose and lipid metabolism [14,15]. According to Blat et al. [16], adipose tissue contains 90% of total  $\alpha$ -tocopherol, and  $\alpha$ -tocopherol from adipocytes is released very slowly during  $\alpha$ -tocopherol deficiency or under the specific conditions of hypermetabolism [17]. Several studies have confirmed inverse associations of obesity with plasma carotenoids [11,18], but there is still a lot of ambiguity about the association of obesity and weight loss with  $\alpha$ -tocopherol status. Furthermore, it is known that variations in body composition are responsible for the differences in plasma lipid profile between men and women. Premenopausal women have a less proatherogenic plasma lipid profile than men, and accumulation of excess body fat appears to affect lipid kinetics differently in men and women [19].

Hence, the aim of the present study was to analyze the link between anthropometric and biochemical characteristics of obesity (particularly visceral and subcutaneous adipose tissue content, waist circumference, lipids profile and antioxidant capacity) and plasma  $\alpha$ -tocopherol concentration after a 6-week weight loss program. We also hypothesized that fat mass reduction in overweight and obese women improves  $\alpha$ -tocopherol status at a greater extent than in men.

## Materials and methods

### Ethics statement

This study was conducted according to the principles of the Declaration of Helsinki for experiments involving humans and was approved by the Bioethical Commission of the National Food and Nutrition Institute (No 1805/2011). All the participants provided written consent to participate in the study. Data obtained during the intervention was confidential and restricted to the participating investigators.

### Participants and study design

The study was conducted in 2012–2014 at the Department of Human Nutrition, Warsaw University of Life Sciences (WULS-

SGGW). The study involved 130 males and females, who fulfilled the following inclusion criteria: age 18–54; overweight or obese according to WHO definition: BMI  $\geq 25$  kg/m<sup>2</sup> and WC: men: <94 cm, women: <80 cm [20,21]; lack of hormone replacement therapy and hypolipemic drugs, consent to participate. Participants receiving pharmacological treatment or diagnosed with chronic diseases, allergies, food intolerances, and women in pregnancy, lactation, and menopausal were excluded from the study.

The participants were treated as part of the AntioxObesity weight loss program, involving the development, implementation, and evaluation of the efficacy of a therapeutic program for overweight and obese adults: comprehensive education on nutrition and physical activity. A total of 130 subjects were initially screened, 60 of whom completed the study (Fig. 1).

### AntioxObesity program

The main assumption of the AntioxObesity weight loss program was to investigate how the process of weight reduction influenced biochemical parameters (lipid profile) and oxidative stress biomarkers: the concentration of lipid antioxidants -  $\alpha$ -tocopherol, vitamin A, carotenoids ( $\beta$ -carotene, lycopene, lutein), antioxidant enzyme activities, and total antioxidant capacity in blood serum without changes in the dietary intake of these compounds. Therefore, in the recommendations, particular attention was paid to the appropriate selection of fruits and vegetables, so that the intake of antioxidants was maintained at a similar level.

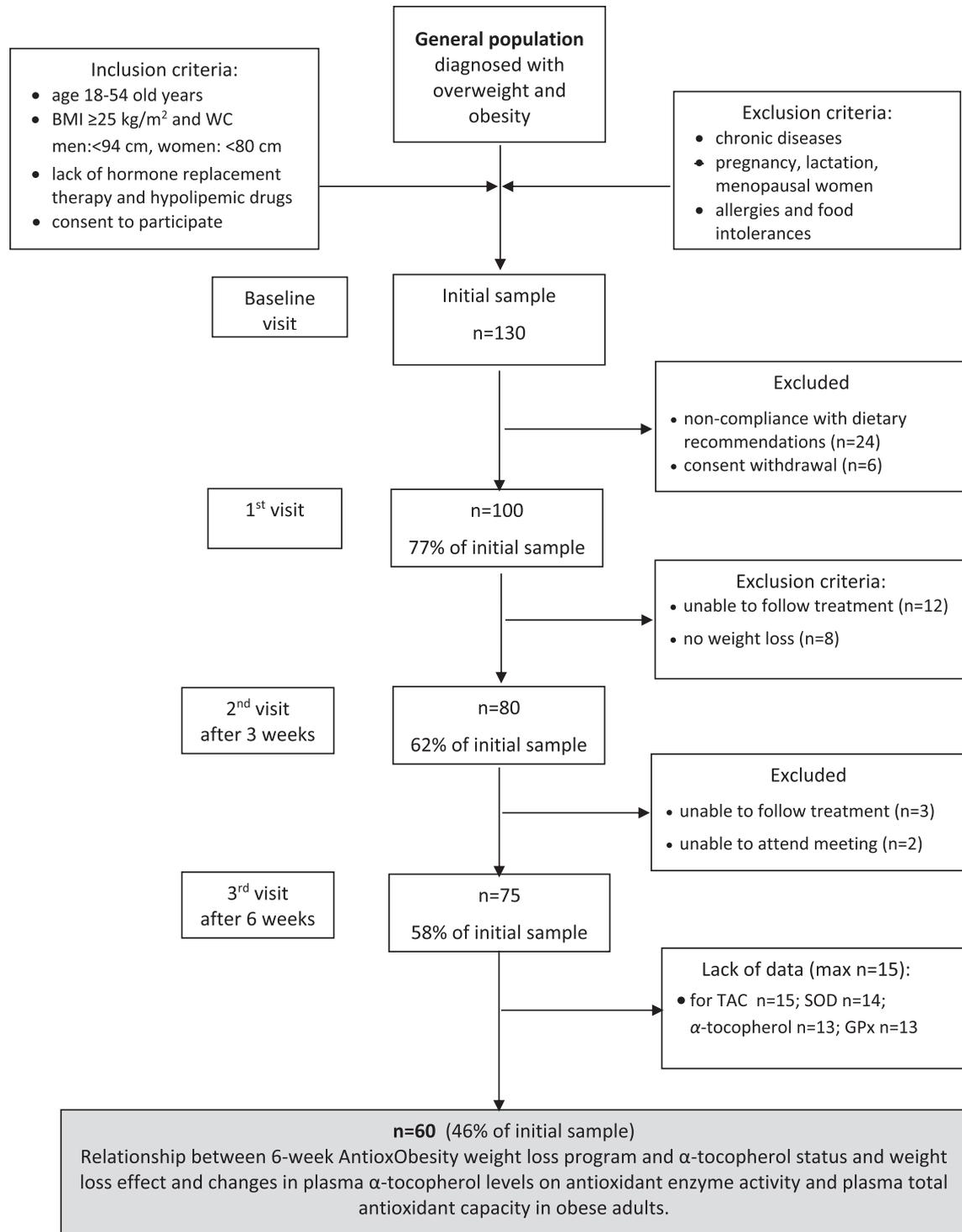
The AntioxObesity weight loss program was an intervention study in overweight and obese adults. The aim of the 6-week AntioxObesity program was weight/fat loss through a diet with a reduced energy value. The six-week weight loss program, divided into 3 stages, included (Table 1): assessment of nutritional value of diet based on 3 days of dietary self-reported records; anthropometric measurements: weight, height, waist and hip circumferences, body composition; biochemical analysis in plasma:  $\alpha$ -tocopherol concentration, lipids profile, antioxidant potential, and SOD and GPx activity. All data from 1st and 3rd stage were used to determine the relationship between the 6-week AntioxObesity weight loss program on  $\alpha$ -tocopherol status in conjunction with lipids profile, antioxidant enzyme activity, and plasma total antioxidant capacity in obese adults.

### Compliance with dietary treatment

According to the main goals of dietary intervention, the dietary compliance criteria were: (1) Adequacy of proposed energy intake according to individual recommendations based on energy restriction (energy value reduced by ca. 500–700 kcal - 20–25% in relation to individuals baseline energy value) according to the individual's BMI; (2) Adequacy of carbohydrate intake; percentage of energy derived from carbohydrates between 50 and 55%  $\pm$  5%; (3) Adequacy of protein intake; percentage of energy from protein between 15 and 20%  $\pm$  5%; (4) Adequacy of fat intake - 25 and 30%  $\pm$  5% of total energy; (5) Adequacy of meal frequency, based on 3 main meals (breakfast, lunch, and dinner) and 2 snacks (mid-morning and mid-afternoon). Participants, who achieved 3 or more of the main goals of the 5 dietary intervention criteria, were considered as showing "global compliance".

### Assessment of dietary variables

Daily intakes of energy, macronutrients, and  $\alpha$ -tocopherol were assessed on the basis of data from the 3-day dietary record method. The dietary record was conducted on the basis of widely accepted and applied rules [22]. Respondents were trained before



**Fig. 1.** Flowchart: study design and data collection.

participating in the survey on how to self-report all foods and beverages consumed daily to provide reliable estimates of dietary intake. In the Department of Human Nutrition, the diary was checked by a nutritionist in the participants' presence. The nutritionist asked for detailed information about the foods and drinks recorded, such as portion sizes and preparation methods using food models of products and dishes (the Polish Atlas of Food Products and Dish Portion Sizes). When necessary, the food diary was corrected by the nutritionist during the visit. After the review, food

intake data were converted to food volume/weight (in mL or g). These data of the 3-days dietary records, three consecutive days (two weekdays and one weekend day) were entered into a nutritional software program (Diet 5.0) based on the latest available information on food composition tables from Poland [23], to evaluate average daily energy value, macronutrients, and  $\alpha$ -tocopherol in the diets. In Polish nutrient database vitamin E content was evaluated as an  $\alpha$ -tocopherol equivalent (mg  $\alpha$ -TE), thus 0.8 conversion factor was used to estimate mg of  $\alpha$ -tocopherol intake.

**Table 1**  
The overall content of the AntioxObesity weight loss program.

| Visit                           | Data collected   |
|---------------------------------|--|
| Baseline                        | 30 min consultation about 6-week AntioxObesity weight loss program; initial measurements and recruitment   |
| 1 <sup>st</sup> visit           | Assessment of nutritional value of diet on the basis of data from 3-day records and a semi-quantitative food-frequency questionnaire<br>Anthropometric measurements (weight, height, waist, and hip circumferences) and body composition (FM, FFM, SF, VF)<br>Blood pressure measurement<br>Blood collection - biochemical analysis: lipid profile, lipid micronutrients, antioxidative biomarkers |
| dietary intervention            | Identifying goals of program, overview of nutritional values, and start reduced caloric diet by ca. 500–700 kcal/day<br>Participants recommended to engage in normal physical activity, suitable to individual abilities and health  |
| 2 <sup>nd</sup> visit (3 weeks) | Assessment of nutritional value of diet on the basis of data from the 3-day record method<br>Anthropometric measurements<br>Blood pressure measurement<br>Dietitian consultation - nutrition recommendations adjusted to individual needs and health status  |
| 3 <sup>rd</sup> visit (6 weeks) | Assessment of nutritional value of diet on the basis of data from 3-day records and a semi-quantitative food-frequency questionnaire<br>Anthropometric measurements (weight, height, waist, and hip circumferences) and body composition (FM, FFM, SF, VF)<br>Blood pressure measurement<br>Blood collection - biochemical analysis: lipid profile, lipid micronutrients, antioxidative biomarkers |

FM, fat mass; FFM, fat-free mass; SF, subcutaneous fat; VF, visceral fat.

Assessing adequacy of  $\alpha$ -tocopherol intake, EAR - 12 mg/d and RDA - 15 mg/d levels were used [24].

#### Anthropometry, body composition

All anthropometric measurements, according to the standardized procedures [25], were taken with light clothing and without shoes twice and averages were calculated. Height (H) was measured with a portable stadiometer with the head in horizontal Frankfurt plane and recorded with a precision of 0.1 cm. Weight was taken in light indoor clothes without shoes using the same electronic digital scale to the nearest 0.1 kg. Waist circumference (WC) was measured using a stretch-resistant tape that provides constant 100 g tension, at the mid-way point between the iliac crest and the costal margin (lower rib) [21] on the anterior axillary line in a resting expiratory position. The hip circumference (HC) measurement was taken at the widest part of the buttocks, with the tape parallel to the floor [26]. The waist-hip ratio (WHR) and the waist-to-height ratio (WHtR) were calculated to determine fat tissue distribution. The WHR was calculated as the WC/HC and interpreted according to WHO criteria, considered 0.90 for men and 0.85 for women to assess central obesity [26]. The WHtR was calculated at the WC/H and value  $\geq 0.5$  was used as a central obesity measure [21]. Body mass index (BMI) was calculated as weight (kg)/height ( $m^2$ ). BMI was categorized according to WHO [20,21]. BMI  $\geq 25$  kg/ $m^2$  was used as overweight and BMI  $\geq 30$  kg/ $m^2$  as obesity measures. Bioelectrical impedance analysis (BIA) (Maltron BioScan 920 ver.1.1) was used to assess fat mass (FM) and fat-free mass (FFM), including subcutaneous fat (SF) and visceral fat (VF). BIA was performed under standardized conditions according to the manufacturer's protocol. All measurements were performed with light clothing and with metal objects (e.g. jewelry, keys) removed. Whole body BIA measurements were performed by placing two adhesive single-use skin electrodes (purchased from Maltron International Ltd, UK) on the right hand and foot, respectively, on the patient when lying in supine position. The device applies a current of 400 mA at a constant frequency of 50 kHz.

The VF and SF measurements were performed in the standing position with four pairs of electrodes positioned on the trunk. The measurements were done at a frequency of 50 kHz, with an impedance range of 5–1100  $\Omega$ .

#### Biochemical analysis

Venous blood samples were taken at baseline and after the 6-week program after an overnight fast (12 h) in the morning

(9–10 a.m.) with minimal stasis and maintained at 4 °C until plasma or serum was separated for biochemical analyses. Plasma and serum samples were collected after centrifugation (1000  $\times$  g for 10 min at 4 °C) and stored frozen (–80 °C) until analysis (no longer than 2 months, except erythrocytes and plasma for SOD and GPx activity, which were analyzed immediately, according to the manufacturer's instructions).

*Lipid profile* (total cholesterol, HDL cholesterol, and triglyceride levels) was determined through standard enzymatic analyses using commercial HYDREX kits (product numbers: total cholesterol - HXB104; HDL cholesterol - HXB106; triglycerides - 17628). LDL level was calculated using the Friedwald formula [27]. The results are expressed as mmol/L and compared with ESC/EAS guidelines [28].

*Plasma  $\alpha$ -tocopherol concentration* was assessed using high-performance liquid chromatography (HPLC - Knauer) with a diode array detector, at scan wavelength of 280–300 nm. LiChro-CART®250-4 RP-18 (4  $\times$  250 mmol; 5  $\mu$ m) with a precolumn (Merc, col. no. 841071, Darmstadt, Germany) was used. Acetonitril/hexane/isopropanol (65:14:21; v/v/v) was applied as the eluent. The flow rate measured 0.8 ml/min. Samples were deproteinized with ethanol with ascorbic acid (1% in 0.1 M HCl) and 20  $\mu$ l of internal standard (15  $\mu$ g/ml,  $\alpha$ -tocopherol acetate in ethanol). Extraction of  $\alpha$ -tocopherol was performed twice with 1 ml hexane by vortex-mixing for 4 min and centrifuging at 3600 g for 10 min at 10 °C. The hexane extracts were collected, and the solvent was evaporated using CentriVap Concentrator (Labconco). The residue was dissolved in 200  $\mu$ l of methanol, vortex-mixed for 30 s and transferred into a vial. The injection volume was 20  $\mu$ l for all analyses. Plasma  $\alpha$ -tocopherol concentrations were calculated by comparing with a corresponding calibration curve (standard of  $\alpha$ -tocopherol from Sigma–Aldrich Inc. in ethanol) in a range of 1–10  $\mu$ g/mL. Analytical recovery of  $\alpha$ -tocopherol was 94%. The mean intra-day coefficients of variation (% CV) for  $\alpha$ -tocopherol at 1.0 and 10  $\mu$ g/mL were <3.3% and <3.2%, respectively. Similarly the corresponding values for the mean inter-day analysis were <3.0 and <3.5%, respectively. All reagents were of highest grade commercially available (HPLC purity) and were purchased from Merc (Darmstadt, Germany).

$\alpha$ -Tocopherol levels are expressed as  $\mu$ mol/L plasma, and lipid adjusted as  $\alpha$ -tocopherol  $\mu$ mol on mmol of total cholesterol (TC), LDL-cholesterol, HDL-cholesterol, triglycerides (TAG), and total lipids (calculated as the sum of total cholesterol and triglycerides).

*$\alpha$ -Tocopherol status* was assessed according the following cut points:  $\alpha$ -tocopherol plasma level >12  $\mu$ mol/L,  $\alpha$ -tocopherol: total TC ratio >2.22  $\mu$ mol/mmol;  $\alpha$ -tocopherol: total lipid ratio

**Table 2**  
Characteristics of study population and changes in investigated parameters during the weight loss program.

| Variables                             | All subjects (n = 60) |                |                                  |          |
|---------------------------------------|-----------------------|----------------|----------------------------------|----------|
|                                       | T <sub>0</sub>        | T <sub>1</sub> | Δ T <sub>1</sub> -T <sub>0</sub> | P value* |
| <b>Age</b>                            | 35.4 ± 8.9            | –              | –                                | –        |
| <b>Diet</b>                           |                       |                |                                  |          |
| Energy (kcal/d)                       | 2040 ± 608            | 1616 ± 705     | -428                             | <0.001   |
| Protein (g/d)                         | 88.5 ± 22.5           | 86.6 ± 34.9    | -1.9                             | 0.271    |
| Fat (g/d)                             | 74.2 ± 32.5           | 52.0 ± 33.7    | -22.1                            | <0.001   |
| Carbohydrate (g/d)                    | 266.4 ± 91.0          | 218.0 ± 96.0   | -48.4                            | <0.001   |
| α-tocopherol (mg/d)                   | 7.6 ± 3.5             | 7.3 ± 4.3      | -0.33                            | 0.679    |
| <b>Anthropometry</b>                  |                       |                |                                  |          |
| Weight (kg)                           | 92.9 ± 17.0           | 88.8 ± 16.9    | -3.9                             | <0.001   |
| BMI (kg/m <sup>2</sup> )              | 31.9 ± 4.5            | 30.1 ± 6.1     | -1.8                             | <0.001   |
| WC (cm)                               | 93.9 ± 1.5            | 89.7 ± 11.3    | -4.2                             | <0.001   |
| WHR                                   | 0.86 ± 0.09           | 0.85 ± 0.09    | -0.01                            | 0.069    |
| WHtR                                  | 0.55 ± 0.06           | 0.53 ± 0.06    | -0.02                            | <0.001   |
| <b>Body composition</b>               |                       |                |                                  |          |
| FFM (%)                               | 58.6 ± 9.0            | 60.0 ± 9.8     | 1.4                              | 0.001    |
| FM (%)                                | 41.4 ± 9.0            | 40.0 ± 9.8     | -1.4                             | 0.001    |
| VF (cm <sup>2</sup> )                 | 169.5 ± 4.7           | 145.5 ± 70.3   | -26.6                            | 0.002    |
| SF (cm <sup>2</sup> )                 | 239.3 ± 0.3           | 224.0 ± 85.8   | -19.2                            | 0.025    |
| <b>Biochemical parameters</b>         |                       |                |                                  |          |
| TC (mmol/L)                           | 5.00 ± 0.83           | 4.74 ± 0.73    | -0.25                            | 0.099    |
| HDL (mmol/L)                          | 1.33 ± 0.23           | 1.31 ± 0.23    | -0.02                            | 0.426    |
| LDL (mmol/L)                          | 3.08 ± 0.80           | 2.84 ± 0.70    | -0.24                            | 0.066    |
| TAG (mmol/L)                          | 1.31 ± 0.43           | 1.29 ± 0.40    | -0.02                            | 0.226    |
| α-tocopherol (μmol/L)                 | 18.2 ± 6.6            | 16.3 ± 6.9     | -1.97                            | 0.006    |
| α-tocopherol:TC ratio (μmol/mmol)     | 3.86 ± 1.77           | 3.70 ± 1.94    | -0.26                            | 0.099    |
| α-tocopherol:HDL ratio (μmol/mmol)    | 14.0 ± 5.29           | 12.97 ± 5.54   | -1.65                            | 0.006    |
| α-tocopherol:LDL ratio (μmol/mmol)    | 6.66 ± 3.67           | 6.64 ± 4.39    | -0.19                            | 0.220    |
| α-tocopherol:TAG ratio (μmol/mmol)    | 15.55 ± 7.62          | 13.66 ± 6.02   | -2.28                            | 0.015    |
| α-tocopherol:lipids ratio (μmol/mmol) | 3.05 ± 1.38           | 2.88 ± 1.40    | -0.25                            | 0.080    |
| SOD (U/g hemoglobin)                  | 161.2 ± 17.2          | 161.9 ± 19.7   | 0.75                             | 0.868    |
| GPx (U/L)                             | 2639 ± 889            | 2182 ± 737     | -448                             | 0.001    |
| TAC (μmol/L)                          | 269.7 ± 21.1          | 263.9 ± 11.2   | -5.72                            | 0.023    |

Results expressed as means ± standard deviation. T<sub>0</sub>, baseline; T<sub>1</sub>, after 6 weeks; Δ T<sub>1</sub>-T<sub>0</sub>, changes; BMI, body mass index; WC, waist circumference; WHR, waist-hip ratio; WHtR, waist-to-height ratio; FFM, fat-free mass; FM, fat mass; VF, visceral fat; SF, subcutaneous fat; TC, total cholesterol; HDL, high-density lipoprotein; LDL, low-density lipoprotein; TAG, triacylglycerol; SOD, superoxide dismutase; GPx, glutathione peroxidase; TAC, total antioxidant capacity.

\*- Student's independent t test (significant difference P ≤ 0.05).

>1.59 μmol/mmol as adequate concentration. α-Tocopherol plasma level >20 μmol/L was used as a strong antioxidant defense indicator and the cut point of decreased risk of cardiovascular disease [13,22,29,30]. In healthy individuals measurement of plasma α-tocopherol concentration is sufficient to establish actual α-tocopherol status, but for patients with metabolic disorders, when α-tocopherol concentration is low, lipid adjustment is necessary [9,30].

Superoxide dismutase (SOD) activity in erythrocytes was determined using the Randox reagent kit (RANSOD - SD-125, UK). Glutathione peroxidase (GPx) activity in blood was determined using the Randox reagent kit (RANSEL - RS-504, UK). Analyzes were performed according to the manufacturer's protocol. Values are expressed as U/g hemoglobin and U/L plasma.

Total antioxidant capacity (TAC) of blood serum was determined using the colorimetric method based on the Trolox Equivalent Antioxidant Capacity (TEAC) assay, according to Re et al. [31].

### Statistical analysis

The parameters analyzed in our study were presented as: means and standard deviation. Differences in all variables between baseline and after 6 weeks were compared by using an independent Student's t-test and Pearson's Chi<sup>2</sup>. Repeated-measures ANOVA was used to assess the effects of gender and effects of time on dietary value, anthropometry, body composition and biochemical parameters.

Linear regression was used to investigate the relationship between changes in selected parameters and α-tocopherol status after 6 weeks the weight loss program. The final 3 multivariate models were specified. Based on the literature research, lipids profile, and adiposity measurements were included, as well models were adjusted on gender, age, α-tocopherol intake. The dependent variable was plasma α-tocopherol after 6 weeks and lipid-corrected plasma α-tocopherol after 6 weeks. The independent variables were baseline plasma α-tocopherol, changes between the end and beginning of the AntioxObesity program for BMI, WC, FM, VF, TC, HDL and for dietary fat. In addition, models were specified for each combination of gender.

For all analyses, P-values ≤ 0.05 were considered statistically significant. Statistical analyses were performed using STATISTICA software (version 12.0 PL; StatSoft Inc., Tulsa, OK, USA).

### Results

General characteristics of the study population and changes in the determined parameters after 6 weeks weight loss program are presented in Table 2. Energy value, dietary fat, and carbohydrate intakes decreased significantly in all subjects (P < 0.001). Mean α-tocopherol intake was about 7.3–7.6 mg α-tocopherol/d and no significantly changes were observed after 6 weeks. Body weight, WC, BMI, WHtR, and FM decreased significantly after the 6 weeks in all subjects (P < 0.05), whereas the percentage of FFM significantly increased. VF and SF decreased significantly (about 27 and 19 cm<sup>2</sup>) in all subjects. Plasma α-tocopherol (P = 0.006), α-tocopherol:HDL

**Table 3**  
Effects of gender and time on anthropometric and biochemical parameters during the weight loss program.

| Variables                             | Women (n = 37) |                |                                  | Men (n = 23)   |                |                                  | P value* |        |        |
|---------------------------------------|----------------|----------------|----------------------------------|----------------|----------------|----------------------------------|----------|--------|--------|
|                                       | T <sub>0</sub> | T <sub>1</sub> | Δ T <sub>1</sub> -T <sub>0</sub> | T <sub>0</sub> | T <sub>1</sub> | Δ T <sub>1</sub> -T <sub>0</sub> | G        | T      | G x T  |
| <b>Age</b>                            | 34.1 ± 8.7     | –              | –                                | 37.9 ± 9.1     | –              | –                                | 0.1226   | –      | –      |
| <b>Diet</b>                           |                |                |                                  |                |                |                                  |          |        |        |
| Energy (kcal/d)                       | 1884 ± 561     | 1460 ± 652     | –424                             | 2340 ± 590     | 1904 ± 665     | –435                             | 0.0046   | 0.0000 | 0.9449 |
| Protein (g/d)                         | 81.4 ± 19.3    | 77.9 ± 30.1    | –3.5                             | 101.7 ± 22.2   | 102.8 ± 38.1   | 1.1                              | 0.0010   | 0.7534 | 0.5575 |
| Fat (g/d)                             | 66.8 ± 28.9    | 45.1 ± 25.2    | –21.8                            | 87.7 ± 34.8    | 64.9 ± 43.2    | –22.8                            | 0.0030   | 0.0004 | 0.9289 |
| Carbohydrate (g/d)                    | 251.2 ± 78.0   | 200.5 ± 95.8   | –50.6                            | 294.5 ± 107    | 250.2 ± 90.0   | –44.3                            | 0.0498   | 0.0000 | 0.7663 |
| α-tocopherol (mg/d)                   | 7.3 ± 3.5      | 6.8 ± 4.1      | –0.50                            | 8.1 ± 3.5      | 8.1 ± 4.7      | –0.02                            | 0.2724   | 0.6837 | 0.6980 |
| <b>Anthropometry</b>                  |                |                |                                  |                |                |                                  |          |        |        |
| Weight (kg)                           | 86.3 ± 14.5    | 82.6 ± 14.3    | –3.3                             | 105.1 ± 14.8   | 100.6 ± 15.3   | –4.9                             | 0.0000   | 0.0000 | 0.0160 |
| BMI (kg/m <sup>2</sup> )              | 31.6 ± 4.6     | 30.4 ± 4.7     | –1.2                             | 32.6 ± 4.3     | 29.8 ± 8.3     | –2.9                             | 0.8653   | 0.0002 | 0.1111 |
| WC (cm)                               | 89.3 ± 8.9     | 85.2 ± 8.9     | –3.8                             | 102.5 ± 10.9   | 97.7 ± 10.9    | –4.8                             | 0.0000   | 0.0000 | 0.2997 |
| WHR                                   | 0.82 ± 0.08    | 0.80 ± 0.06    | –0.01                            | 0.93 ± 0.08    | 0.92 ± 0.08    | –0.01                            | 0.0000   | 0.0415 | 0.5435 |
| WHtR                                  | 0.54 ± 0.05    | 0.52 ± 0.05    | –0.02                            | 0.57 ± 0.07    | 0.55 ± 0.07    | –0.03                            | 0.0640   | 0.0000 | 0.5129 |
| <b>Body composition</b>               |                |                |                                  |                |                |                                  |          |        |        |
| FFM (%)                               | 55.3 ± 8.5     | 56.4 ± 9.2     | 1.1                              | 64.8 ± 6.3     | 66.9 ± 6.7     | 2.0                              | 0.0000   | 0.0014 | 0.2891 |
| FM (%)                                | 44.7 ± 8.5     | 43.6 ± 9.2     | –1.1                             | 35.2 ± 6.3     | 33.2 ± 6.7     | –2.0                             | 0.0000   | 0.0014 | 0.2892 |
| VF (cm <sup>2</sup> )                 | 159.1 ± 54.0   | 139.3 ± 61.9   | –23.6                            | 188.8 ± 78.8   | 156.6 ± 83.9   | –32.3                            | 0.2033   | 0.0000 | 0.1585 |
| SF (cm <sup>2</sup> )                 | 233.6 ± 73.4   | 225.4 ± 88.3   | –14.3                            | 249.9 ± 92.3   | 221.6 ± 83.1   | –28.3                            | 0.7838   | 0.0103 | 0.1535 |
| <b>Biochemical parameters</b>         |                |                |                                  |                |                |                                  |          |        |        |
| TC (mmol/L)                           | 5.06 ± 0.78    | 4.77 ± 0.78    | –0.29                            | 4.89 ± 0.90    | 4.67 ± 0.62    | –0.15                            | 0.4008   | 0.0538 | 0.5270 |
| HDL (mmol/L)                          | 1.30 ± 0.22    | 1.30 ± 0.22    | 0.00                             | 1.38 ± 0.23    | 1.31 ± 0.23    | –0.03                            | 0.8531   | 0.9704 | 0.9556 |
| LDL (mmol/L)                          | 3.16 ± 0.78    | 2.87 ± 0.76    | –0.30                            | 2.91 ± 0.81    | 2.77 ± 0.58    | –0.11                            | 0.3421   | 0.0742 | 0.3855 |
| TAG (mmol/L)                          | 1.31 ± 0.41    | 1.31 ± 0.42    | –0.02                            | 1.31 ± 0.46    | 1.27 ± 0.35    | –0.09                            | 0.8647   | 0.3553 | 0.1964 |
| α-tocopherol (μmol/L)                 | 17.8 ± 6.4     | 16.1 ± 7.0     | –1.76                            | 19.0 ± 7.06    | 16.6 ± 5.77    | –2.39                            | 0.6429   | 0.0035 | 0.6410 |
| α-tocopherol:TC ratio (μmol/mmol)     | 3.72 ± 1.65    | 3.63 ± 2.07    | –0.03                            | 4.11 ± 1.99    | 3.87 ± 1.67    | –0.81                            | 0.2851   | 0.0638 | 0.0821 |
| α-tocopherol:HDL ratio (μmol/mmol)    | 14.03 ± 5.19   | 12.71 ± 5.81   | –1.28                            | 14.04 ± 5.59   | 13.59 ± 4.96   | –2.52                            | 0.3731   | 0.0030 | 0.3132 |
| α-tocopherol:LDL ratio (μmol/mmol)    | 6.32 ± 3.41    | 6.57 ± 4.78    | 0.38                             | 7.23 ± 4.14    | 6.79 ± 3.42    | –1.54                            | 0.3427   | 0.3088 | 0.0965 |
| α-tocopherol:TAG ratio (μmol/mmol)    | 15.18 ± 7.41   | 13.25 ± 6.09   | –1.83                            | 16.20 ± 8.14   | 14.62 ± 5.98   | –3.35                            | 0.3303   | 0.0029 | 0.3597 |
| α-tocopherol:lipids ratio (μmol/mmol) | 2.94 ± 1.28    | 2.81 ± 1.47    | –0.09                            | 3.23 ± 1.56    | 3.03 ± 1.26    | –0.64                            | 0.2692   | 0.0245 | 0.0898 |
| SOD (U/g hemoglobin)                  | 161.3 ± 16.9   | 161.2 ± 18.58  | –0.03                            | 160.9 ± 18.1   | 163.3 ± 22.1   | 2.19                             | 0.8376   | 0.7181 | 0.7107 |
| GPx (U/L)                             | 2625 ± 807     | 2118 ± 585     | –507                             | 2666 ± 1055    | 2307 ± 973     | –341                             | 0.5824   | 0.0000 | 0.4705 |
| TAC (μmol/L)                          | 264.7 ± 90.1   | 263.9 ± 87.2   | –2.00                            | 280.0 ± 32.0   | 267.1 ± 15.2   | –13.2                            | 0.0093   | 0.0167 | 0.0727 |

Results expressed as means ± standard deviation. T<sub>0</sub>, baseline; T<sub>1</sub>, after 6 weeks; Δ T<sub>1</sub>-T<sub>0</sub>, changes; G, gender; T, effects time; G x T, gender x effects time interaction effect; BMI, body mass index; WC, waist circumference; WHR, waist-hip ratio; WHtR, waist-to-height ratio; FFM, fat-free mass; FM fat mass; VF, visceral fat; SF, subcutaneous fat; TC, total cholesterol; HDL, high-density lipoprotein; LDL, low-density lipoprotein; TAG, triacylglycerol; SOD, superoxide dismutase; GPx, glutathione peroxidase; TAC, total antioxidant capacity.

\*- Repeated-measures ANOVA (significant difference  $P \leq 0.05$ ).

**Table 4**  
Percentage of participants who did not meet the reference values for lipids profile values and α-tocopherol status.

|   | All subjects (n = 60) |               |       | Women (n = 37) |               |       | Men (n = 23) |               |       |
|---|-----------------------|---------------|-------|----------------|---------------|-------|--------------|---------------|-------|
|   | Baseline              | After 6 weeks | P*    | Baseline       | After 6 weeks | P*    | Baseline     | After 6 weeks | P*    |
| LDL (<3.0 mmol/L)                                 | 64.8                  | 50.9          | 0.042 | 58.1           | 42.9          | 0.038 | 71.4         | 60.0          | 0.049 |
| HDL (>1.0 mmol/L in men and >1.3 mmol/L in women) | 69.1                  | 68.9          | 0.983 | 57.1           | 58.1          | 0.939 | 90.0         | 92.9          | 0.062 |
| TAG (<1.7 mmol/L)                                 | 85.5                  | 88.9          | 0.611 | 85.7           | 87.1          | 0.870 | 85.0         | 92.9          | 0.384 |
| α-tocopherol in plasma (>12 μmol/L)               | 19.6                  | 23.2          | 0.105 | 21.6           | 27.0          | 0.088 | 15.6         | 15.6          | 1.000 |
| α-tocopherol in plasma (>20 μmol/L)               | 60.7                  | 75.0          | 0.049 | 59.5           | 78.4          | 0.019 | 63.2         | 68.4          | 0.073 |
| α-tocopherol: TC ratio (>2.22 μmol/mmol)          | 5.6                   | 11.4          | 0.038 | 8.6            | 12.9          | 0.047 | 0.0          | 7.7           | 0.219 |
| α-tocopherol: lipids ratio (>1.59 μmol/mmol)      | 25.9                  | 36.4          | 0.026 | 28.6           | 41.9          | 0.009 | 21.1         | 23.1          | 0.069 |

LDL, low-density lipoprotein; HDL, high-density lipoprotein; TAG, triacylglycerol; TC, total cholesterol.

\*Chi<sup>2</sup> Pearson test (significant difference  $P \leq 0.05$ ).

ratio ( $P = 0.006$ ) and α-tocopherol:TAG ratio ( $P = 0.015$ ) significantly decreased after 6 weeks the program in all subjects. No changes were observed for SOD activity, but GPx activity ( $P = 0.001$ ) and TAC ( $P = 0.023$ ) decreased significantly in all participants.

Table 3 shows effects time or/and gender on investigated parameters. For dietary intake (energy,  $P = 0.0046$ ; protein,  $P = 0.0010$ ; fat  $P = 0.0030$  and carbohydrate,  $P = 0.0498$ ), anthropometrics measurements (weight,  $P = 0.0000$ ; WC,  $P = 0.0000$ ; WHR,  $P = 0.0000$  and FM,  $P = 0.0000$ ) and TAC ( $P = 0.0093$ ) statistically significant effects of gender were stated. Men were characterized by a greater decrease in weight, BMI, WC, FM, VF, SF and TAC compared to women. A statistically significant

effects of the time was found for dietary value (energy,  $P = 0.0000$ ; fat,  $P = 0.0004$  and carbohydrate,  $P = 0.0000$ ), anthropometry and body composition (all variables) and biochemical parameters (α-tocopherol level,  $P = 0.0035$ ; α-tocopherol:HDL ratio,  $P = 0.0030$ ; α-tocopherol:TAG ratio,  $P = 0.002$ ; α-tocopherol:lipids ratio,  $P = 0.0245$ ; GPx,  $P = 0.0000$ ) and TAC ( $P = 0.0167$ ). Interaction between gender and effects of the time was found for the subjects' weight ( $P = 0.0160$ ).

Table 4 shows the percentage of participants who did not meet the reference values for the lipids profile and α-tocopherol. The percentage of participants who did not meet the reference values for LDL decreased significantly in all subjects ( $P = 0.0420$ ), women ( $P = 0.0038$ ) and men ( $P = 0.0492$ ) after 6 weeks of weight

**Table 5**

Multivariate models<sup>a</sup> between plasma  $\alpha$ -tocopherol concentration after 6 weeks of the AntioxObesity program and selected predictors.

| Predictors   | $\alpha$ -tocopherol ( $\mu\text{mol/L}$ ) after 6 weeks |                |
|--|--|----------------|
|  | $\beta$ -coefficient                                     | 95% CI         |
| <b>All subjects</b> (Overall model: $r = 0.74$ ; $r^2 = 0.68$ ; $P < 0.0001$ ) |  |                |
| changes TC (mmol/L)  | -0.781   | -0.265; 0.109  |
| changes HDL (mmol/L)   | 0.119  | -0.069; 0.308  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol/L}$ )                            | 0.793*   | 0.612; 0.974   |
| changes dietary fat (g/d)  | 0.084  | -0.119; 0.286  |
| changes VF (cm <sup>2</sup> )  | -0.195   | -0.394; 0.004  |
| changes FM (%)   | 0.208*   | 0.008; 0.408   |
| changes BMI (kg/m <sup>2</sup> )   | 0.037  | -0.194; 0.268  |
| changes WC (cm)  | 0.095  | -0.103; 0.294  |
| <b>Women</b> (Overall model: $r = 0.81$ ; $r^2 = 0.74$ ; $P < 0.0001$ )        |  |                |
| changes TC (mmol/L)  | -0.153   | -0.366; 0.060  |
| changes HDL (mmol/L)   | 0.080  | -0.137; 0.296  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol/L}$ )                            | 0.808*   | 0.592; 1.024   |
| changes dietary fat (g/d)  | 0.232  | 0.010; 0.454   |
| changes VF (cm <sup>2</sup> )  | -0.153   | -0.384; 0.078  |
| changes FM (%)   | 0.201  | -0.028; 0.431  |
| changes BMI (kg/m <sup>2</sup> )   | 0.090  | -0.157; 0.337  |
| changes WC (cm)  | -0.081   | -0.312; 0.150  |
| <b>Men</b> (Overall model: $r = 0.85$ ; $r^2 = 0.79$ ; $P = 0.0087$ )          |  |                |
| changes TC (mmol/L)  | 0.447  | -0.694; 1; 589 |
| changes HDL (mmol/L)   | -0.428   | -1.093; 0.237  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol/L}$ )                            | 1.004*   | 0.444; 1.563   |
| changes dietary fat (g/d)  | -0.373   | -0.915; 0.168  |
| changes VF (cm <sup>2</sup> )  | -0.603*  | -1.199; -0.007 |
| changes FM (%)   | 0.227  | -0.433; 0.886  |
| changes BMI (kg/m <sup>2</sup> )   | -0.064   | -0.456; 0.328  |
| changes WC (cm)  | -0.110   | -0.922; 0.702  |

Linear regression analysis adjusted for age, gender and  $\alpha$ -tocopherol intake.

TC, total cholesterol; HDL, high-density lipoprotein; VF, visceral fat; FM, fat mass; BMI, body mass index; WC, waist circumference.

\*,  $P \leq 0.01$ .

<sup>a</sup>, Each model was controlled for all predictors shown in table.

reduction compared with baseline (Table 4). Moreover, we observed that lipid profile improved; for TC and LDL levels a decreasing tendency ( $P < 0.1$ ) was found (Table 3). However, HDL levels were lower than reference values in close to 60% of women and 90% of men, and TAG level did not meet reference values in 85–93% of the respondents (Table 4).

Mean circulating levels of plasma  $\alpha$ -tocopherol of about 16–19  $\mu\text{mol/L}$  (Table 3) didn't indicate an  $\alpha$ -tocopherol deficiency ( $<12 \mu\text{mol/L}$ ), but we found that 22–27% women and 16% men (Table 4) had inadequate plasma  $\alpha$ -tocopherol concentrations. Low  $\alpha$ -tocopherol status ( $<20 \mu\text{mol/L}$ ) was found at baseline in 60% of the women and 63% of the men, and it increased to 78% and 68% after 6 weeks, respectively, which indicates an increased risk of cardiovascular diseases and weak antioxidant defense.

Plasma  $\alpha$ -tocopherol concentration after 6 weeks of the AntioxObesity program was strongly associated with baseline plasma  $\alpha$ -tocopherol, changes in VF, and changes in FM (Table 5). Lipid-corrected  $\alpha$ -tocopherol (Tables 6 and 7) after the 6-week program was associated with changes in TC, baseline plasma  $\alpha$ -tocopherol, and changes in FM, VF for all subjects. The identified set of correlates explained 74%, 76%, and 74% of total variation in plasma  $\alpha$ -tocopherol, the  $\alpha$ -tocopherol: lipids ratio, and the  $\alpha$ -tocopherol: TC ratio for all subjects. In women baseline plasma  $\alpha$ -tocopherol concentration and changes in TC were statistically significant correlates in models, while a change in VF and in dietary fat were predictors in men (Tables 5–7).

Table 8 shows multivariable-adjusted linear regression models between plasma  $\alpha$ -tocopherol and lipid-corrected  $\alpha$ -tocopherol and selected predictors after the 6-week AntioxObesity program. Changes in TC, baseline plasma  $\alpha$ -tocopherol, VF and FM were predictors of plasma  $\alpha$ -tocopherol and lipid-corrected  $\alpha$ -tocopherol

**Table 6**

Multivariate models<sup>a</sup> between plasma  $\alpha$ -tocopherol: lipids ratio after 6 weeks of the AntioxObesity program and selected predictors.

| Predictors   | $\alpha$ -tocopherol: lipids ratio ( $\mu\text{mol}/\text{mmol}$ ) after 6 weeks |                |
|--|--|----------------|
|  | $\beta$ -coefficient   | 95% CI         |
| <b>All subjects</b> (Overall model: $r = 0.76$ ; $r^2 = 0.71$ ; $P < 0.0001$ ) |  |                |
| changes TC (mmol/L)  | -0.188*  | -0.369; -0.008 |
| changes HDL (mmol/L)   | 0.119  | -0.063; 0.300  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol/L}$ )                            | 0.821*   | 0.647; 0.996   |
| changes dietary fat (g/d)  | 0.018  | -0.178; 0.213  |
| changes VF (cm <sup>2</sup> )  | -0.213*  | -0.404; -0.021 |
| changes FM (%)   | 0.128  | -0.068; 0.317  |
| changes BMI (kg/m <sup>2</sup> )   | 0.108  | -0.114; 0.330  |
| changes WC (cm)  | 0.098  | -0.093; 0.289  |
| <b>Women</b> (Overall model: $r = 0.83$ ; $r^2 = 0.77$ ; $P < 0.0001$ )        |  |                |
| changes TC (mmol/L)  | -0.286*  | -0.486; -0.086 |
| changes HDL (mmol/L)   | 0.064  | -0.139; 0.268  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol/L}$ )                            | 0.822*   | 0.619; 1.024   |
| changes dietary fat (g/d)  | 0.205  | -0.003; 0.414  |
| changes VF (cm <sup>2</sup> )  | -0.199   | -0.416; 0.018  |
| changes FM (%)   | 0.110  | -0.106; 0.325  |
| changes BMI (kg/m <sup>2</sup> )   | 0.167  | -0.065; 0.0399 |
| changes WC (cm)  | -0.106;  | -0.323; 0.112  |
| <b>Men</b> (Overall model: $r = 0.84$ ; $r^2 = 0.80$ ; $P = 0.0061$ )          |  |                |
| changes TC (mmol/L)  | 0.855  | -0.203; 1.907  |
| changes HDL (mmol/L)   | -0.559   | -1.173; 0.053  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol/L}$ )                            | 1.194*   | 0.682; 1.710   |
| changes dietary fat (g/d)  | -0.453   | -0.954; 0.046  |
| changes VF (cm <sup>2</sup> )  | -0.189   | -0.744; 0.360  |
| changes FM (%)   | -0.143   | -0.753; 0.465  |
| changes BMI (kg/m <sup>2</sup> )   | -0.083   | -0.441; 0.278  |
| changes WC (cm)  | -0.308   | -1.064; 0.440  |

Linear regression analysis adjusted for age, gender and  $\alpha$ -tocopherol intake.

TC, total cholesterol; HDL, high-density lipoprotein; VF, visceral fat; FM, fat mass; BMI, body mass index; WC, waist circumference.

\*,  $P \leq 0.01$ .

<sup>a</sup>, Each model was controlled for all predictors shown in table.

in all subjects. In women changes in TC and baseline plasma  $\alpha$ -tocopherol concentration correlated in models. While baseline plasma  $\alpha$ -tocopherol concentration and changes in VF were statistically significant in men.

## Discussion

The main findings of this substudy are: (1) the 6-week weight loss program brought about a decline in fat intake and  $\alpha$ -tocopherol status, (2) baseline plasma  $\alpha$ -tocopherol concentrations and changes in FM, VF proved to be good predictors of changes in  $\alpha$ -tocopherol status, (3) gender no significantly influenced on  $\alpha$ -tocopherol status after 6 weeks weight loss program, but in men more decrease of FM, VF and SF than in women, is associated with lower TAC.

In our study, average intake of  $\alpha$ -tocopherol for about 90% of participants was inadequate at baseline and for 83% at the end of weight loss program. This is in line with the results by Agarwal et al. [32], who reported that compared to normal weight adults, obese adults had lower intakes of vitamin E and a higher prevalence of inadequacy. Most diets assuming a reduction of body mass are based on lowering the fat content in the diet, which decreases intakes of lipophilic micronutrients, like vitamin E. Some previous studies [33,34] reported that increasing fat intake (to 35–50% of calories) improved vitamin E status and did not negatively affect cardiovascular risk factors. Low-fat diets are associated with decreased vitamin E and n-3 fatty acid intakes [35]. In our study, after 6 weeks, the energy from fat decreased by about 3%, but the average intake of  $\alpha$ -tocopherol remained at the same inadequate level. Results indicated that the selection of product groups is

**Table 7**  
Multivariate models<sup>a</sup> between plasma  $\alpha$ -tocopherol:TC ratio after 6 weeks of the AntioxObesity program and selected predictors.

| Predictors   | $\alpha$ -tocopherol: TC ( $\mu\text{mol}/\text{mmol}$ ) after 6 weeks |                |
|--|--|----------------|
|  | $\beta$ -coefficient   | 95% CI         |
| <b>All subjects</b> (Overall model: $r = 0.74$ ; $r^2 = 0.68$ ; $P < 0.0001$ ) |  |                |
| changes TC (mmol/L)  | -0.240*  | -0.427; -0.052 |
| changes HDL (mmol/L)   | 0.158  | -0.031; 0.347  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol}/\text{L}$ )                     | 0.769*   | 0.588; 0.950   |
| changes dietary fat (g/d)  | -0.005   | -0.208; 0.199  |
| changes VF ( $\text{cm}^2$ )   | -0.240*  | -0.439; -0.040 |
| changes FM (%)   | -0.166*  | -0.034; 0.367  |
| changes BMI ( $\text{kg}/\text{m}^2$ )   | 0.166  | -0.066; 0.397  |
| changes WC (cm)  | 0.133  | -0.006; 0.332  |
| <b>Women</b> (Overall model: $r = 0.82$ ; $r^2 = 0.75$ ; $P < 0.0001$ )        |  |                |
| changes TC (mmol/L)  | -0.320*  | -0.531; -0.110 |
| changes HDL (mmol/L)   | 0.105  | -0.108; 0.319  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol}/\text{L}$ )                     | 0.782*   | 0.570; 0.995   |
| changes dietary fat (g/d)  | 0.197  | -0.022; 0.416  |
| changes VF ( $\text{cm}^2$ )   | -0.212   | -0.439; 0.016  |
| changes FM (%)   | 0.141  | -0.085; 0.368  |
| changes BMI ( $\text{kg}/\text{m}^2$ )   | 0.230  | -0.013; 0.473  |
| changes WC (cm)  | -0.027   | -0.255; 0.201  |
| <b>Men</b> (Overall model: $r = 0.96$ ; $r^2 = 0.88$ ; $P = 0.0141$ )          |  |                |
| changes TC (mmol/L)  | 0.594  | -0.690; 1.178  |
| changes HDL (mmol/L)   | -0.453   | -1.204; 0.294  |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol}/\text{L}$ )                     | 1.154*   | 0.521; 1.783   |
| changes dietary fat (g/d)  | -0.667*  | -1.283; -0.058 |
| changes VF ( $\text{cm}^2$ )   | -0.507   | -1.184; 0.164  |
| changes FM (%)   | -0.037   | -0.783; 0.707  |
| changes BMI ( $\text{kg}/\text{m}^2$ )   | 0.141  | -0.302; 0.582  |
| changes WC (cm)  | -0.295   | -1.210; 0.619  |

Linear regression analysis adjusted for age, gender and  $\alpha$ -tocopherol intake. TC, total cholesterol; HDL, high-density lipoprotein; VF, visceral fat; FM, fat mass; BMI, body mass index; WC, waist circumference.

\*  $P \leq 0.01$ .

<sup>a</sup> Each model was controlled for all predictors shown in table.

important to create a future weight loss program, and special attention should be paid on choosing the adequate sources of fat and vitamin E (like oils, olives, dark-green leafy vegetables, whole grains, flax seeds, nuts, almonds) than limiting fat.

Weight loss after 6 weeks was associated with significantly lower plasma  $\alpha$ -tocopherol concentration, which generally depended on its baseline concentration. Moreover, about 90% of men had HDL level lower than references value. After 6 weeks we

observed the tendency in decreasing TC and LDL level. Most subjects (61–78%) had plasma  $\alpha$ -tocopherol concentrations of less than 20  $\mu\text{mol}/\text{L}$ , indicating an increased risk of cardiovascular disease and weak antioxidant defense. How Mah et al. [36] stated that the bioavailability of  $\alpha$ -tocopherol is lower in patients with metabolic syndrome, which obesity is one of its component. Obese individuals have decreased intestinal absorption and hepatic secretion of  $\alpha$ -tocopherol. It was explained by greater intestinal fat storage in adults with greater body fat. Lower  $\alpha$ -tocopherol absorption, “trapping” of  $\alpha$ -tocopherol within enterocytes and delayed secretion into lymphatic system, limit  $\alpha$ -tocopherol availability in individuals with excessive body fat. In addition, inflammation and/or greater oxidation within the enterocyte or hepatocyte is associated with metabolic disorders, may limit  $\alpha$ -tocopherol bioavailability [36,37]. On the other hand, lower  $\alpha$ -tocopherol plasma status in obese people is explained by the accumulation of  $\alpha$ -tocopherol in adipose tissue.  $\alpha$ -Tocopherol, as well as carotenoids or vitamin D are stored in adipose tissue, but in comparison to other lipid soluble micronutrients, weight loss does not affect plasma  $\alpha$ -tocopherol concentration [4]. The differential association of body mass with  $\alpha$ -tocopherol and other lipid-soluble micronutrients argues against a generalized fat effect on oxidative stress and their metabolism. Schaefer et al. [38] found that only triglycerides are mobilized from a fat cell during up to 6 months of weight loss. Cholesterol and  $\alpha$ -tocopherol released from the adipocyte are independent of factors controlling triglycerides afflux. Moreover,  $\alpha$ -tocopherol stimulates the expression of PPAR $\gamma$  and lipid accumulation during adipocyte differentiation, and affects the endogenous synthesis of cholesterol, likely by modulating the cleavage of SREBPs [4]. Traber et al. [39] supposed that plasma  $\alpha$ -tocopherol concentrations mainly are dependent on mechanisms control circulating lipids. In our program, weight loss and changes in diet (lower fat intake) caused a tendency ( $p = 0.066$ ) to lowering LDL-cholesterol concentrations. Because under physiological fasting conditions  $\alpha$ -tocopherol is mostly transported via LDL [40], it may explain the decrease in plasma  $\alpha$ -tocopherol concentration during the 6 weeks.

In addition, it is interesting to note that no effect of weight loss program on SOD activity was found, but significantly lower GPx activity. Similarly, an important reduction in GPx activity in obese patients compared with the control group was confirmed by Monzo-Beltran et al. [41] in a one-year follow-up study, but other authors [42–44] supposed that levels of plasma antioxidants could

**Table 8**  
Multivariable-adjusted linear regression models between plasma  $\alpha$ -tocopherol concentration and lipid-corrected  $\alpha$ -tocopherol after 6 weeks of the AntioxObesity program and selected predictors ( $P \leq 0.01$ , for all presented predictors).

| Predictors   | $\alpha$ -tocopherol ( $\mu\text{mol}/\text{L}$ )          |              | $\alpha$ -tocopherol: lipids ratio ( $\mu\text{mol}/\text{mmol}$ ) |                | $\alpha$ -tocopherol: TC ratio ( $\mu\text{mol}/\text{mmol}$ ) |                |
|--|--|--------------|--|----------------|--|----------------|
|  | $\beta$ -coefficient                                       | 95% CI       | $\beta$ -coefficient   | 95% CI         | $\beta$ -coefficient   | 95% CI         |
| <b>All subjects</b>  |  |              |  |                |  |                |
|  | Overall model: $r = 0.63$ ; $r^2 = 0.60$ ;<br>$P < 0.0000$ |              | Overall model: $r = 0.73$ ; $r^2 = 0.70$ ;<br>$P < 0.0001$         |                | Overall model: $r = 0.85$ ; $r^2 = 0.69$ ;<br>$P < 0.0001$     |                |
| changes TC (mmol/L)  | —  | —            | —  | —              | -0.319   | -0.493; -0.144 |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol}/\text{L}$ ) | 0.706  | 0.520; 0.892 | 0.758  | 0.573; 0.943   | 0.766  | 0.594; 0.939   |
| changes VF ( $\text{cm}^2$ )                               | —  | —            | -0.182   | -0.355; -0.008 | —  | —              |
| changes FM (%)   | 0.208  | 0.030; 0.385 | 0.218  | 0.046; 0.391   | -0.221   | -0.394; -0.048 |
| <b>Women</b>   |  |              |  |                |  |                |
|  | Overall model: $r = 0.66$ ; $r^2 = 0.63$ ;<br>$P < 0.0001$ |              | Overall model: $r = 0.73$ ; $r^2 = 0.71$ ;<br>$P < 0.0001$         |                | Overall model: $r = 0.84$ ; $r^2 = 0.69$ ;<br>$P < 0.0001$     |                |
| changes TC (mmol/L)  | —  | —            | -0.314   | -0.517; -0.111 | -0.434   | -0.643; -0.225 |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol}/\text{L}$ ) | 0.789  | 0.569; 1.008 | 0.778  | 0.575; 0.981   | 0.760  | 0.551; 0.969   |
| <b>Men</b>   |  |              |  |                |  |                |
|  | Overall model: $r = 0.57$ ; $r^2 = 0.48$ ;<br>$P = 0.0052$ |              | Overall model: $r = 0.80$ ; $r^2 = 0.74$ ;<br>$P = 0.0021$         |                | Overall model: $r = 0.81$ ; $r^2 = 0.75$ ;<br>$P = 0.0011$     |                |
| baseline $\alpha$ -tocopherol ( $\mu\text{mol}/\text{L}$ ) | 0.704  | 0.338; 1.070 | 0.935  | 0.561; 1.309   | 0.945  | 0.575; 1.314   |
| changes VF ( $\text{cm}^2$ )                               | —  | —            | —  | —              | -0.411   | -0.787; -0.034 |

Linear regression analysis adjusted for age, gender and  $\alpha$ -tocopherol intake.

TC, total cholesterol; VF, visceral fat; FM, fat mass.

be reduced and antioxidative enzyme activity should be increased, because obesity is positively associated with oxidative stress. The decrease of  $\alpha$ -tocopherol caused a decrease of TAC, which indicated decreased antioxidative defense during fat loss. TAC value after 6 weeks more decrease in men than in women. Our findings indicated the need of further studies to explanation these differences.

In relation to adiposity indicators, we obtained interesting results for plasma  $\alpha$ -tocopherol concentrations, since the decrease in  $\alpha$ -tocopherol level was associated with a decrease in adiposity. Obesity, especially VF, is a confirmed risk factor for metabolic disorders [45–47]. Central adiposity and VF was found to be a stronger predictor of plasma  $\alpha$ -tocopherol concentration. Our results are in line with several prior studies [10,11,48] that reported that  $\alpha$ -tocopherol serum levels were positively related to BMI and central obesity (WC, WHtR, WHR). They explained the role of  $\alpha$ -tocopherol as a gene modulator, especially involved in glucose and lipid metabolism. Waniek et al. [49] observed a strong association of the  $\alpha$ -tocopherol/cholesterol ratio with VF. VF is significantly reduced during weight loss and a greater VF loss was associated with a higher baseline level of VF and higher baseline BMI.

VF is preferentially lost in the early part of weight loss [50]. This may explain our findings regarding decreasing plasma  $\alpha$ -tocopherol, TAC in association with VF and fat loss, especially in men. Adipose tissue is an endocrine organ and especially VF releases more inflammatory markers [50], which could be responsible for increased oxidative stress leading to reduced  $\alpha$ -tocopherol levels [36].

In summary, this study provides further insight regarding factors associated with plasma  $\alpha$ -tocopherol status in overweight and obese adults during 6 weeks weight loss. Nutritional intervention, coupled with nutrition motivational interviewing, should be regarded as an effective dietary plan for weight loss, but special attention should be paid to  $\alpha$ -tocopherol status at the beginning of fat reduction. Baseline  $\alpha$ -tocopherol status is a strong predictor of a decrease in plasma  $\alpha$ -tocopherol and weakened antioxidative defense during weight loss. Traber et al. [13] and Mah et al. [36] pointed out the lack of correlation between dietary  $\alpha$ -tocopherol intakes and circulating  $\alpha$ -tocopherol. In healthy adults, intakes of 12–15 mg  $\alpha$ -tocopherol daily are sufficient to provide adequate vitamin E status [13], but requires further research in adults with metabolic disorders. Future studies should consider gender and obesity-related physiologic effects on circulating  $\alpha$ -tocopherol levels. It is known that serum  $\alpha$ -tocopherol is gender specific [36] due to hormonal differences and gender-dependent differences in the activation of the CYP enzymes involved in vitamin E metabolism [40], but our results didn't confirm this effect. But we found that strong predictors of  $\alpha$ -tocopherol status in women were baseline  $\alpha$ -tocopherol and changes of TC, while in men - baseline  $\alpha$ -tocopherol and changes of VF, which requires explanation in further studies.

Several limitations of the present study should be mentioned. The main limitation of this study was the lack of inflammatory markers and oxidative damage, such as C-reactive protein, interleukins, tumor necrosis factor (TNF $\alpha$ ), and isoprostanes. In future studies on  $\alpha$ -tocopherol status assessment, larger and longer trials are needed to investigate the effect of gender and anthropometric parameters on  $\alpha$ -tocopherol metabolism. In our study, we assessed only  $\alpha$ -tocopherol, which generally predominates in plasma, but other forms of vitamin E, especially  $\gamma$ -tocopherol have antioxidant properties and it is the predominant form in food [51,52]. Moreover additional and better than plasma  $\alpha$ -tocopherol plasma concentration biomarkers of vitamin E status, like adipose tissue  $\alpha$ -tocopherol concentration or urinary  $\alpha$ -tocopherol catabolites:  $\alpha$ -carboxyethyl hydroxychromanol ( $\alpha$ -CEHC) and  $\alpha$ -carboxymethylbutyl hydroxychromanol ( $\alpha$ -CMBHC) and biliary and fecal excretion of  $\alpha$ -tocopherol and its catabolites should be assessed [37].

Although the study had small subject numbers, the findings lay the ground for further studies, and the analyzed relationships can be used as tools in a study of adiposity and the cardiometabolic profile in adults. Moreover, the present study proved that low baseline  $\alpha$ -tocopherol status and adiposity in obese adults negatively affected  $\alpha$ -tocopherol status after 6 weeks weight loss program. These results, coupled with excessive weight and low  $\alpha$ -tocopherol intake, led to the finding that there was an increased risk of oxidative stress diseases in adults on a reduced diet. Long-term dietary restriction program for obese patients should be monitored to avoid  $\alpha$ -tocopherol deficiency, and take into account higher dietary  $\alpha$ -tocopherol requirements for obese people.

### Conflicts of interest

The authors declare no conflicts of interest.

### Author contributions

The paper was written by all authors.

### Statement of authorship

JH, AS conducted research; AS, MG, JF biochemical analysis; JH, MG, JF, AS analysis and interpretation data; MG, JF drafting of manuscript; JH did a critical review of the manuscript; JH, MG had primary responsibility for final content. All authors read and approved the final manuscript.

### Acknowledgements

We would like to thank all the participants in the study group. This study was supported by the Polish Ministry of Science and Higher Education Grant (WULS-SGGW: 505-10-10020030). Part of the study was supported by a designated subsidy for restructuring of research unit in the years 2016–2017.

### References

- [1] Hansen D, Dendale P, Beelen M, Jonkers R, Mullens A, Corluy L, et al. Plasma adipokine and inflammatory marker concentrations are altered in obese, as opposed to non-obese, type 2 diabetes patients. *Eur J Appl Physiol* 2010;109:397–404. <https://doi.org/10.1007/s00421-010-1362-5>.
- [2] Kajitani N, Shikata K, Nakamura A, Nakatou T, Hiramatsu M, Makino H. Microinflammation is a common risk factor for progression of nephropathy and atherosclerosis in Japanese patients with type 2 diabetes. *Diabetes Res Clin Pract* 2010;88:171–6. <https://doi.org/10.1016/j.diabres.2010.01.012>.
- [3] Monteiro R, Azevedo I. Chronic inflammation in obesity and the metabolic syndrome. *Mediat Inflamm* 2010. <https://doi.org/10.1155/2010/289645>. published online Jul 14.
- [4] Landrier JF, Marcotorchino J, Tourniaire F. Lipophilic micronutrients and adipose tissue biology. *Nutrients* 2012;4:1622–49. <https://doi.org/10.3390/nu41116>.
- [5] Vincent HK, Taylor AG. Biomarkers and potential mechanisms of obesity-induced oxidant stress in humans. *Int J Obes (Lond)* 2006;30:400–18. <https://doi.org/10.1038/sj.ijo.0803177>.
- [6] Hopss E, Noto D, Caimi G, Averna MR. A novel component of the metabolic syndrome: the oxidative stress. *Nutr Metab Cardiovasc Dis* 2010;20:72–7. <https://doi.org/10.1016/j.numecd.2009.06.002>.
- [7] Libby P, Okamoto Y, Rocha VZ, Folco E. Inflammation in atherosclerosis: transition from theory to practice. *Circ J* 2010;74:213–20. <https://doi.org/10.1253/circj.CJ-09-0706>.
- [8] Ben Amara N, Tourniaire F, Maraninchi M, Attia N, Amiot-Carlin MJ, Raccach D, et al. Independent positive association of plasma  $\beta$ -carotene concentrations with adiponectin among non-diabetic obese subjects. *Eur J Nutr* 2015;54:447–54. <https://doi.org/10.1007/s00394-014-0728-6>.
- [9] Bonet ML, Canas JA, Ribot J, Palou A. Carotenoids and their conversion products in the control of adipocyte function, adiposity and obesity. *Arch Biochem Biophys* 2015;572:112–25. <https://doi.org/10.1016/j.abb.2015.02.022>.
- [10] Guendiaim M, Mayneris-Perxachs J, Montes R, López-Belmonte G, Martín-Matillas M, Castellote AI, et al. Relation between plasma antioxidant vitamin levels, adiposity and cardio-metabolic profile in adolescents: effects of a

- multidisciplinary obesity program. *Clin Nutr* 2017;36:209–17. <https://doi.org/10.1016/j.clnu.2015.11.001>.
- [11] Kabat GC, Heo M, Ochs-Balcom HM, LeBoff MS, Mossavar-Rahmani Y, Adams-Campbell LL, et al. Longitudinal association of measures of adiposity with serum antioxidant concentrations in postmenopausal women. *Eur J Clin Nutr* 2016;70:47–53. <https://doi.org/10.1038/ejcn.2015.74>.
- [12] Traber MG. Mechanisms for the prevention of vitamin E excess. *J Lipid Res* 2013;54(9):2295–306. <https://doi.org/10.1194/jlr.R032946>.
- [13] Traber MG. Vitamin E inadequacy in humans: causes and consequences. *Adv Nutr* 2014;5:503–14. <https://doi.org/10.3945/an.114.006254>.
- [14] Mardones P, Rigotti A. Cellular mechanisms of vitamin E uptake: relevance in  $\alpha$ -tocopherol metabolism and potential implications for disease. *J Nutr Biochem* 2004;15:252–60. <https://doi.org/10.1016/j.jnutbio.2004.02.006>.
- [15] Zillikens MC, van Meurs JBJ, Rivadeneira F, Hofman A, Oostra BA, Sijbrands E, et al. Interactions between dietary vitamin E intake and SIRT1 genetic variation influence body mass index. *Am J Clin Nutr* 2010;91:1387–93. <https://doi.org/10.3945/ajcn.2009.28627>.
- [16] Blatt DH, Leonard SW, Traber MG. Vitamin E kinetics and the function of tocopherol regulatory proteins. *Nutrition* 2001;17:799–805.
- [17] Traber MG, Leonard SW, Traber DL, Traber LD, Gallagher J, Bobe G, et al.  $\alpha$ -Tocopherol adipose tissue stores are depleted after burn injury in pediatric patients. *Am J Clin Nutr* 2010;92:1378–84. <https://doi.org/10.3945/ajcn.2010.30017>.
- [18] Chai W, Conroy SM, Maskarinec G, Franke AA, Pagano IS, Cooney RV. Associations between obesity and serum lipid-soluble micronutrients among premenopausal women. *Nutr Res* 2010;30:227–32. <https://doi.org/10.1016/j.nutres.2010.04.006>.
- [19] Wang X, Magkos F, Mittendorfer B. Sex differences in lipid and lipoprotein metabolism: it's not just about sex hormones. *J Clin Endocrinol Metab* 2011;96(4):885–93. <https://doi.org/10.1210/jc.2010-2061>.
- [20] World Health Organization. In: Branca F, Nikogosian H, Lobstein T, editors. *The challenge of obesity in the WHO European Region and the strategies for response. Summary*; 2007. Denmark.
- [21] Ashwell M, Gibson S. Waist-to-height ratio as an indicator of 'early health risk': simpler and more predictive than using a 'matrix' based on BMI and waist circumference. *BMJ Open* 2016;6:e010159. <https://doi.org/10.1136/bmjopen-2015-010159>.
- [22] Gibson RS. *Principles of nutritional assessment*. Oxford University Press; 2005.
- [23] Wajszczyk B, Chwojnowska Z, Nasiadko D, Rybaczuk M. *Dieta 5.0 software for individual and group nutrition assessment and diet planning*. Warsaw, Poland: National Food and Nutrition Institute; 2015.
- [24] Institute of Medicine (US). Panel on dietary antioxidants and related compounds. *Dietary reference intakes for vitamin C, vitamin E, selenium, and carotenoids*. vol. 6. Washington (DC): National Academies Press (US); 2000. Vitamin E. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK225461>.
- [25] Lohman TG, Roche AF, Martorell R, editors. *Anthropometric standardization reference manual*. USA: Human Kinetics Publisher; 1988.
- [26] World Health Organization. *Waist circumference and waist-hip ratio: report of a WHO expert consultation*. 2008. Geneva.
- [27] Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma without use of the preparative ultracentrifuge. *Clin Chem* 1972;18:499–502.
- [28] ESC/esc-european society of cardiology and european atherosclerosis society new guidelines. *Pol Heart J* 2016;74(11):1234–318. <https://doi.org/10.5603/KP.2016.0157>.
- [29] Traber MG, Jialal I. Measurement of lipid-soluble vitamins-further adjustment needed? *Lancet* 2000;355:2013–4. [https://doi.org/10.1016/S0140-6736\(00\)02345-X](https://doi.org/10.1016/S0140-6736(00)02345-X).
- [30] Ford L, Farr J, Morris P, Berg J. The value of measuring serum cholesterol-adjusted vitamin E in routine practice. *Ann Clin Biochem* 2006;43:130–4. <https://doi.org/10.1258/000456306776021526>.
- [31] Re R, Pellegrini N, Proteggente A, Pannala A, Yang M, Rice-Evans C. Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radic Biol Med* 1999;26:1231–7. [https://doi.org/10.1016/S0891-5849\(98\)00315-3](https://doi.org/10.1016/S0891-5849(98)00315-3).
- [32] Agarwal S, Reider C, Brooks JR, Fulgoni 3rd VL. Comparison of prevalence of inadequate nutrient intake based on body weight status of adults in the United States: an analysis of NHANES 2001–2008. *J Am Coll Nutr* 2015;34:126–34. <https://doi.org/10.1080/07315724.2014.901196>.
- [33] Johnstone AM, Lobley GE, Horgan GW, Bremner DM, Fyfe CL, Morrice PC, et al. Effects of a high-protein, low-carbohydrate v. high-protein, moderate-carbohydrate weight-loss diet on antioxidant status, endothelial markers and plasma indices of the cardiometabolic profile. *Br J Nutr* 2011;106(2):282–91. <https://doi.org/10.1017/S0007114511000092>.
- [34] Meksawan K, Pendergast DR, Leddy JJ, Mason M, Horvath PJ, Awad AB. Effect of low and high fat diets on nutrient intakes and selected cardiovascular risk factors in sedentary men and women. *J Am Coll Nutr* 2004;23(2):131–40.
- [35] Mueller-Cunningham WM, Quintana R, Kasim-Karakas SE. An ad libitum, very low-fat diet results in weight loss and changes in nutrient intakes in postmenopausal women. *J Am Diet Assoc* 2003;103(12):1600–6. <https://doi.org/10.1016/j.jada.2003.09.017>.
- [36] Mah E, Sapper TN, Chitchumroonchokchai C, Failla ML, Schill KE, Clinton SK, et al.  $\alpha$ -Tocopherol bioavailability is lower in adults with metabolic syndrome regardless of dairy fat co-ingestion: a randomized, double-blind, crossover trial. *Am J Clin Nutr* 2015;102:1070–80. <https://doi.org/10.3945/ajcn.115.118570>.
- [37] Traber MG, Mah E, Leonard SW, Bobe G, Bruno RS. Metabolic syndrome increases dietary  $\alpha$ -tocopherol requirements as assessed using urinary and plasma vitamin E catabolites: a double-blind, crossover clinical trial. *Am J Clin Nutr* 2017 Mar;105(3):571–9. <https://doi.org/10.3945/ajcn.116.138495>.
- [38] Schaefer EJ, Woo R, Kibata M, Bjornsen L, Schreiber PH. Mobilization of triglyceride but not cholesterol or tocopherol from human adipocytes during weight reduction. *Am J Clin Nutr* 1983;37(5):749–54. <https://doi.org/10.1093/ajcn/37.5.749>.
- [39] Traber MG, Leonard SW, Bobe G, Fu X, Saltzman E, Grusak MA, et al.  $\alpha$ -Tocopherol disappearance rates from plasma depend on lipid concentrations: studies using deuterium-labeled collard greens in younger and older adults. *Am J Clin Nutr* 2015;101(4):752–9. <https://doi.org/10.3945/ajcn.114.100966>.
- [40] Schmözl L, Birringer M, Lorkowski S, Wallert M. Complexity of vitamin E metabolism. *World J Biol Chem* 2016;26:14–43. <https://doi.org/10.4331/wjbc.v7.i1.14>.
- [41] Monzo-Beltran L, Vazquez-Tarragón A, Cerdà C, Garcia-Perez P, Iradi A, Sánchez C, et al. One-year follow-up of clinical, metabolic and oxidative stress profile of morbid obese patients after laparoscopic sleeve gastrectomy. 8-oxo-dG as a clinical marker. *Redox Biol* 2017;12:389–402. <https://doi.org/10.1016/j.redox.2017.02.003>.
- [42] Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, et al. Increased oxidative stress in obesity and its impact on metabolic syndrome. *J Clin Invest* 2004;114:1752–61. <https://doi.org/10.1172/JCI20042162>.
- [43] Devries MC, Hamadeh MJ, Glover AW, Raha S, Samjoo IA, Tarnopolsky MA. Endurance training without weight loss lowers systemic, but not muscle, oxidative stress with no effect on inflammation in lean and obese women. *Free Radic Biol Med* 2008;45:503–11. <https://doi.org/10.1016/j.freeradbiomed.2008.04.039>.
- [44] Mittal PC, Kant R. Correlation of increased oxidative stress to body weight in disease-free postmenopausal women. *Clin Biochem* 2009;42:1007–11. <https://doi.org/10.1016/j.clinbiochem.2009.03.019>.
- [45] Castro AV, Kolka CM, Kim SP, Bergman RN. Obesity, insulin resistance and comorbidities? Mechanisms of association. *Arq Bras Endocrinol Metabol* 2014;58:600–9.
- [46] Tatsumi Y, Nakao YM, Masuda I, Higashiyama A, Takegami M, Nishimura K, et al. Risk for metabolic diseases in normal weight individuals with visceral fat accumulation: a cross-sectional study in Japan. *BMJ Open* 2017;7:e013831. <https://doi.org/10.1136/bmjopen-2016-013831>.
- [47] Kim JH, Cho JJ, Park YS. Relationship between sarcopenic obesity and cardiovascular disease risk as estimated by the Framingham risk score. *J Korean Med Sci* 2015;30:264–71. <https://doi.org/10.3346/jkms.2015.30.3.264>.
- [48] Wallström P, Wirfält E, Lahmann PH, Gullberg B, Janson L, Berglund G. Serum concentrations of beta-carotene and alpha-tocopherol are associated with diet, smoking, and general and central adiposity. *Am J Clin Nutr* 2001;73:777–85.
- [49] Waniek S, di Giuseppe R, Plachta-Danielzik S, Ratjen I, Jacobs G, Koch M, et al. Association of vitamin E levels with metabolic syndrome, and MRI-derived body fat volumes and liver fat content. *Nutrients* 2017;9:1143. <https://doi.org/10.3390/nu9101143>.
- [50] Liu FX, Flatt SW, Nichols JF, Pakiz B, Barkai HS, Wing DR, et al. Factors associated with visceral fat loss in response to a multifaceted weight loss intervention. *J Obes Weight Loss Ther* 2017;7(4):346. <https://doi.org/10.4172/2165-7904.1000346>.
- [51] Donnan MS, Heath DD, Flatt SW, Pakiz B, Quintana EL, Rana BK, et al. Factors associated with tocopherol status in obese women: effects of diet composition and weight loss. *Vitam Miner* 2016;5:3. <https://doi.org/10.4172/2376-1318.1000147>.
- [52] Kim YN, Cho YO. Vitamin E status of 20- to 59-year-old adults living in the Seoul metropolitan area of South Korea. *Nutr Res Pract* 2015;9:192–8. <https://doi.org/10.4162/nrp.2015.9.2.192>.