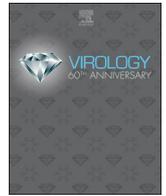




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## *In vitro*, *in cellulo* and structural characterizations of the interaction between the integrase of Porcine Endogenous Retrovirus A/C and proteins of the BET family

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### ABSTRACT

Retroviral integrase (IN) proteins catalyze the permanent integration of the viral genome into host DNA. They can productively recruit cellular proteins, and the human Bromodomain and Extra-Terminal domain (hBET) proteins have been shown to be co-factors for integration of gamma-retroviruses such as Murine Leukemia Virus (MLV) into human cells. By using two-hybrid, co-immunoprecipitation and *in vitro* interaction assays, we showed that IN of the gamma- Porcine Endogenous Retrovirus-A/C (PERV IN) interacts through its C-terminal domain (CTD) with hBET proteins. We observed that PERV IN interacts with the BRD2, BRD3 and BRD4 proteins *in vitro* and that the BRD2 protein specifically binds and co-localizes with PERV IN protein in the nucleus of cells. We further mapped the interaction sites to the conserved Extra-Terminal (ET) domain of the hBET proteins and to several amino acids of the of the C-terminal tail of the PERV IN CTD. Finally, we determined the first experimental structure of an IN CTD – BET ET complex from small-angle X-ray scattering data (SAXS). We showed that the two factors assemble as two distinct modules linked by a short loop which confers partial flexibility. The SAXS-restrained model is structurally compatible with the binding of the PERV intasome to BRD2. Altogether, these data confirm the important role of host BET proteins in the gamma-retroviruses' targeting site and efficiency of integration.

### 1. Background

During the retroviral life cycle, integrases (INs) that catalyze the key step of integration for the provirus of the retrotranscribed DNA into the host cell genome can interact with cellular cofactors to target specific integration sites (Kvaratskhelia et al., 2014). Hence, the Human

Immunodeficiency Virus type 1 (HIV-1) binds the host Lens Epithelium-Derived Growth Factor (LEDGF/p75) protein (Busschots et al., 2005; Cherepanov et al., 2003; Emiliani et al., 2005; Llano et al., 2006; Maertens et al., 2003) to preferentially target the body of actively transcribed genes (Mitchell et al., 2004; Schroder et al., 2002). More recently, the cleavage and polyadenylation specificity factor 6 protein

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(CPSF6) complexed with the HIV-1 capsid (CA) protein has been shown to play a dominant role in directing integration within euchromatin (Sowd et al., 2016). Integration of Avian Sarcoma and Leukemia Viruses (ASLV) is more random (Mitchell et al., 2004) and no integration cell cofactor directing ASLV integration has been reported to date even though the ASLV IN-binding factor FACT (FACilitates Chromatin Transcription) might be a good candidate (Winans et al., 2017). Also, it remains to be seen whether the delta-retroviral IN-binding serine/threonine protein phosphatase 2A (PP2A) protein plays a role in directing integration of delta-retroviruses (Maertens, 2016). Foamy virus (FV) integration seems to also be random, but FVs tend to target promoter-closed regions when integration into a transcribed region has occurred (Nowrouzi et al., 2006).

Concerning gamma-retroviruses, the Murine Leukemia Virus (MLV) favors integration near the promoters, enhancers, CpG islands and DNase I hypersensitive sites (De Ravin et al., 2014; LaFave et al., 2014; Mitchell et al., 2004; Wu et al., 2003). Numerous groups (De Rijck et al., 2013; Gupta et al., 2013; Sharma et al., 2013) have confirmed earlier preliminary data (Studamire and Goff, 2008) indicating a potential binding of MLV IN with the Bromodomain-containing protein Brd2, and they have shown that Brd2, 3 and 4 target the integration of MLV in transcription start sites. These three ubiquitously expressed proteins of the Bromodomain and Extra-Terminal (BET) family specifically bind MLV IN and co-localize in the cell nucleus. Furthermore, the integration site distribution of MLV corresponds to the chromatin-binding profile of BET proteins (De Rijck et al., 2013). Brd4 was reported to interact with other gamma-retroviral INs, such as those of Feline Leukemia Virus (FeLV) (Gupta et al., 2013) and avian Reticulo-Endotheliosis Virus (REV) (Serrao et al., 2015). In addition, a recent paper has pinpointed an important role of the MLV CA protein in facilitating the targeting of the pre-integration complex to nucleosomes via a mechanism that could be similar to the one of the HIV-1 CA-CPSF6 complex (Wanaguru et al., 2018).

Members of the BET protein family contain tandem bromodomains (BD-I and BD-II) and a conserved Extra-Terminal domain (ET) (Thorpe et al., 1996). The BET family contains chromatin-binding factors that have been implicated in the control of cell cycle progression, transcription and DNA replication (Belkina and Denis, 2012; Wu and Chiang, 2007). These proteins bind to the acetylated tails of histones H3 and H4 via their bromodomains. The extra-terminal motif ET in the C-terminal region of BET proteins was shown to interact with numerous cellular — e.g. transcription factors and histone modification enzymes — and viral factors. Interestingly, BRD2 was shown to interact with Kaposi's Sarcoma-Associated Herpesvirus (KSHV) and the latency-associated nuclear antigen 1 (LANA 1), a protein required for the replication of the episomal genome in dividing cells (Viejo-Borbolla et al., 2005). The C-terminal part of the long isoform of BRD4 can bind the bovine papilloma virus type 1 E2 protein (Jang et al., 2009).

Retroviral INs are composed of three conserved domains — i.e. N-terminal domain (NTD), catalytic core domain (CCD) and C-terminal domain (CTD) — joined by flexible linkers (see Fig. 1A) which affect crystallization trials. For this reason structural information was obtained only for one- or two-domain IN fragments for a decade (reviewed by (Andrake and Skalka, 2015)). The NTD was characterized with an all- $\alpha$  fold, the CCD with an RNase-H fold and the CTD with an SH3 fold. Since 2010, crystal and cryo-electron microscopy structures of intasomes (i.e. the entire IN complex with bound viral DNA ends) have been published for spumavirus, lentivirus, alpha-retrovirus and beta-retrovirus genera (Ballandras-Colas et al., 2016, 2017; Hare et al., 2010; Passos et al., 2017; Yin et al., 2016). Concerning gamma-retroviruses, the isolated NTD and CTD structures of MLV IN are known (Aiyer et al., 2014; Guan et al., 2017), and we have recently published a model of the Porcine Endogenous Virus A/C (PERV-A/C) intasome (Demange et al., 2015). BET proteins also display a conserved modular architecture and the isolated structures of BD-I, BD-II and ET have been determined, each with an all- $\alpha$  fold ((Lin et al., 2008); for a review, see (Sanchez and

Zhou, 2009)). The ET binding motif (EBM) of MLV IN was located within the C-terminal tail of the CTD (Aiyer et al., 2014), as observed in the nuclear magnetic resonance (NMR) structure of MLV IN EBM complexed with BRD4 ET (Crowe et al., 2016). The latter study revealed that the 17-long flexible EBM segment (MLV IN aa 389–405) folds into a  $\beta$ -hairpin when it binds the ET domain (Crowe et al., 2016).

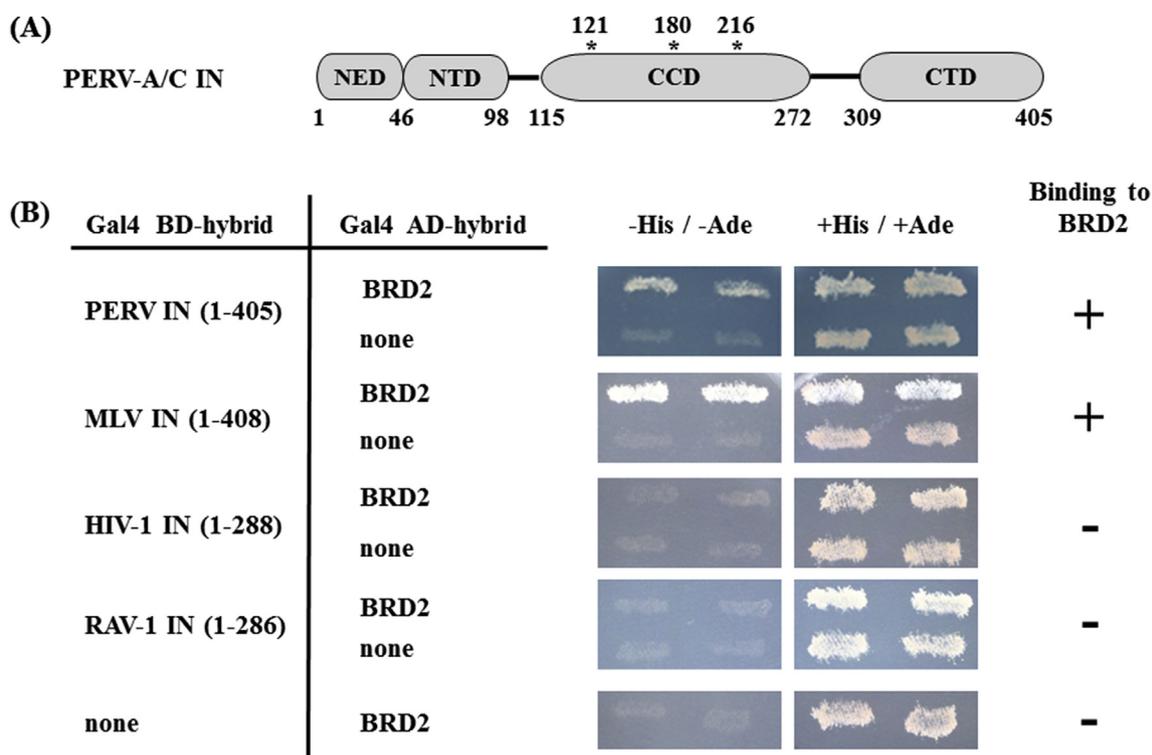
We have previously shown that the integration of the gamma-retrovirus PERV-A/C (hereafter referred to as PERV) in human cells is not random and is strongly favored at transcriptional start sites and CpG islands (Moalic et al., 2006, 2009). These results revealed similarities between PERV and MLV integration site selection, suggesting that gamma-retroviruses have a common pattern of integration and that their cellular integration target cofactors might be similar. However, this assumption remains to be demonstrated. In this study, we show *in vitro* and *in cellulo* that PERV IN interacts strongly with BET proteins like MLV IN, in contrast with the behavior exhibited by ASLV and HIV-1 INs. We further perform small-angle X-ray Scattering (SAXS) experiments on the PERV IN CTD – BRD2 ET complex and generate a 3D model that corroborates our interaction results.

## 2. Materials and methods

### 2.1. Plasmids

**Plasmids for the two-hybrids assay.** PERV, MLV, HIV-1 and RAV-1 INs — as well as BRD2 encoding sequences — were amplified by PCR from the recombinant PERV-A/C clone AY570980 (supplied by Takeuchi, Y.), the pHit60 (supplied by S. P. Goff), pET15b-IN-HIV-1 (Cellier et al., 2013), pET30a-IN-RAV-1 (Moreau et al., 2004), GST-Brd2/RingG3B, GST-Brd4/HUNK, GST-Brd43/ORF XRingG3B (supplied by T. Schulz), respectively, using appropriate primers (Tables S1 and S2). The resulting fragments were cloned into either the pGBKT7 plasmid (for INs) or the pGAD-T7 plasmid (for BRD2, 3, 4) using the Matchmaker Gold Yeast Two Hybrid System (Clontech). Deletion constructs were generated for PERV IN (pGBKT7-PERV IN (1–98), pGBKT7-PERV IN (99–272), pGBKT7-PERV IN (273–405), pGBKT7-PERV IN (1–272), pGBKT7-PERV IN (99–405), pGBKT7-PERV IN (99–394), pGBKT7-PERV IN (99–383), pGBKT7-PERV IN (372–405) and for MLV IN (pGBKT7-MLV IN (1–276), pGBKT7-MLV IN (277–408) by PCR fusion of DNA fragments encoding the corresponding amino acid (aa) residues from either the PERV or the MLV INs, respectively, with the Gal4-DNA binding domain. Point mutations (pGBKT7-PERV INmut (P387A/L388A), pGBKT7-PERV INmut (K389A/L390A), pGBKT7-PERV INmut (R391A/L392A), pGBKT7-PERV INmut (H393A/R394A) of the full-length PERV IN were generated by PCR-directed mutagenesis with the appropriate primers. The oligonucleotides used are listed in Table S1. All constructs were verified by DNA sequencing.

**Cloning of the BRD2-encoding cDNA from HEK293T cells into the pCMV-Myc plasmid.** The full-length BRD2 coding sequences were isolated from HEK293T cells. RNA from HEK293T cells was isolated with the RNeasy kit (Qiagen) and converted to complementary DNA (cDNA) by reverse transcription using the M-MLV RT (Promega) with an oligo (dT) primer. The BRD2 encoding cDNA was then amplified by two independent polymerase chain reactions (PCRs): (i) the first with the hF1 (position 1–22) and hR1 (position 1217–1196) primers, and (ii) the second with hF2 (position 1144–1162) and hR2 (position 2406–2379) primers (Table S3). PCR amplifications were performed with 35 amplification cycles (30 s at 95 °C, 30 s at 60 °C and 105 s at 72 °C) using the Pfu DNA polymerase (Promega). The two overlapping PCR fragments were then denatured (10 min at 95 °C) and hybridized at room temperature for 30 min followed by an elongation step of 15 min at 72 °C using the Pfu polymerase to generate the full-length cDNA of the BRD2 gene. The BRD2 cDNA was then inserted into the mammalian expression vector pCMV-Myc (Invitrogen) according to the Gateway technology (Invitrogen). This plasmid (named pCMV-Myc-BRD2) contains the hCMV enhancer-promoter and an N-terminal Myc tag allowing



**Fig. 1. Interaction of PERV IN with the BRD2 protein.** (A) *Domain organization of the full-length gamma-retroviral PERV IN protein.* The IN proteins are made up of three domains: (i) the N-terminal domain (NTD) which is involved in the binding of the viral DNA and zinc, (ii) the catalytic core domain (CCD) which contains the invariant catalytic triad D,D-35-E (indicated with asterisks), (iii) the C-terminal domain (CTD) involved in non-specific DNA binding. The PERV also contains an N-terminal extension domain (NED). (B) *In the two-hybrid assay, INs from PERV and MLV gamma-retroviruses interact with BRD2, while INs from HIV-1, a lentivirus, and from RAV-1, an alpha-retrovirus, do not.* The full-length ORFs of PERV, MLV, HIV-1 and RAV-1 INs were fused to the Gal4-DNA binding domain (Gal4-BD). The BRD2 ORF was fused to the Gal4-activation domain (Gal4-AD). The yeast reporter strain Y2H-Gold producing the hybrid proteins indicated was analyzed for histidine and adenine auxotrophy (-His/-Ade) acquired by the interaction of the fused proteins.

the expression of the Myc-BRD2 protein. Sequencing was performed to control for the integrity of the cDNA.

**Construction of the pPERV IN CTD – BRD2 (667–808) complex expression plasmid.** An original plasmid construction was built to obtain the protein complex formed by PERV IN CTD (aa 309–405) and BRD2 (aa 667–808), cloned from the pCI-Flag-PERV IN and the pCMV-Myc-BRD2 plasmids respectively. This construction contains three genes encoding: i) TEV protease, ii) PERV IN CTD with an N-terminal Strep-tag II and iii) BRD2 (667–808) with an N-terminal hexahistidine (His) tag (see Fig. 6A). At first, each gene was amplified by PCR using the pRK793b, pGEM-T Easy and pPROEX-Hta plasmids, respectively. The whole insert containing these three genes was generated by PCR. In addition, the genes were separated from each other by TEV protease cleavage sites introduced by the primers. The insert was cloned using an In-Fusion<sup>®</sup> HD-Cloning kit (Clontech) and amplified by PCR in the pET28a vector.

The constructed expression plasmid was also used to generate one single and one double alanine mutant affecting the complexed IN CTD (*i.e.* W379A and R391A/L392A), and to generate the shorter pPERV IN CTD – BRD2 (667–735) complex expression plasmid also termed pPERV IN CTD – BRD2 ET. All plasmids were confirmed by DNA sequencing.

**Construction of the pCMV-Myc-BRD2-, BRD3-, BRD4-Cter plasmids.** The pCMV-Myc-BRD2 plasmid was used for the amplification of the BRD2 C-terminal fragments (BRD2-Cter) by PCR using hF3 (position 1789–1807) and hR3 (position 2406–2389) primers (Table S3). The BRD3 C-terminal fragment (BRD3-Cter) was amplified from pGVB3403 using the hF4 (position 1705–1722) and hR4 (position 2181–2161) primers, and the BRD4 C-terminal fragment (BRD4-Cter) was amplified from pGVB3115 using hF5 (position 1819–1836) and hR5 (position hF5-hR5) primers (Table S3). PCRs were performed with 35 amplification cycles (30 s at 95 °C, 45 s 65 °C and 160 s at 72 °C) using the Pfu

DNA polymerase (*Promega*). The resulting fragment was inserted into the pCMV-Myc vector using the Gateway technology as above and controlled by sequencing.

**Construction of the pCI-Flag-PERV IN plasmids.** The PERV IN coding sequence was amplified from plasmid A14/220 containing the whole genome of PERV (Genbank ref. AY570980) using attB11SP/PERV and attB21ASP/PERV IN primers (Table S3). PCRs were performed with 35 amplification cycles (30 s at 95 °C, 30 s 60 °C and 120 s at 72 °C) using the Pfu DNA polymerase (*Promega*). The resulting fragment was inserted into the pDONR223 and pCI-Flag plasmids using the Gateway technology as above.

The pCI-Flag-PERV IN CTD was obtained by PCR amplification of the CTD of PERV IN (aa 273–405) from pCI-Flag PERV IN by PCR amplification using F and R primers. The resulting fragment was inserted into the pCI-Flag plasmid using the Gateway technology. The pCI-Flag PERV IN CTD (P387A/L388A) plasmid containing a CTD mutated was obtained by PCR-directed mutagenesis with the appropriate primers.

## 2.2. The two-hybrid assay

The yeast reporter strain Y2H-Gold containing the *HIS3* and *ADE2* Gal4-inducible genes were co-transformed and plated on a selective medium lacking tryptophan and leucine. Y2H-Gold double transformants were patched on the same media and then analyzed for histidine and adenine auxotrophy by replica plating on selective medium lacking tryptophan, leucine, histidine and adenine (-His/-Ade).

## 2.3. Cells and immunoassays

**Cells.** Human embryonic kidney HEK293T cells were cultured in

Dulbecco's modified eagle medium (PAA), 10% fetal bovine serum (PAA) and 1% penicillin-streptomycin (Gibco) at 37 °C in 5% CO<sub>2</sub>.

**Confocal immunofluorescence microscopy.** For immunofluorescence microscopy, HEK293T cells were grown in LabTek II chamber slides (*BD Falcon*) for 24 h and transfected with 800 ng of plasmid using Lipofectamine 2000 (Invitrogen). Forty-eight hours later, the transfected HEK293T cells were fixed with 1% paraformaldehyde in phosphate-buffered saline (PBS) overnight at 4 °C, washed with PBS and then incubated in a blocking/permeabilization buffer (BP) (10% FBS and 0.1% saponin in PBS) for 10 min at 4 °C. After incubation, the cells were washed twice with the BP buffer and incubated with the primary antibodies. The Myc-BRD2 fusion protein and the Flag-PERV IN fusion protein were detected using a mix of polyclonal rabbit antibody directed against the Myc tag (*Sigma-Aldrich*, catalog no. C3956, dilution 1/500) and a monoclonal mouse antibody directed against the Flag tag (*Sigma-Aldrich*, catalog no. F3165; dilution 1/500) in the BP buffer for 1 h at 4 °C. After incubation, the cells were washed twice with the BP buffer and incubated with the alexa-488 conjugated goat anti-rabbit antibody (*Invitrogen*, catalog no. A11008, dilution 1/500) or the alexa-633 goat anti-mouse antibody (*Invitrogen*, catalog no. A21053; dilution 1/500), respectively. The cells were then mounted in the Vectashield Mounting Medium (*vector Laboratories*) containing 1.5 µg/ml of 4',6-diamidino-2-phenylindole (DAPI) to stain the nuclear DNA. Images were acquired with a Confocal Spectral SP5 (*Leica*) controlled by LAS AF software (*Leica Microsystems CMS GmbH*) with a 63x oil immersion lens.

**Co-Immuno Precipitation (Co-IP) assays.** HEK293T cells were plated at a confluence of 60–70% in 6-well dishes. The following day, cells were co-transfected with 2 µg of each of two plasmids: (i) the pCI-Flag, the pCI-Flag-PERV IN CTD or the pCI-Flag-PERV IN CTD (P387A/L388A) plasmids and (ii) the pCMV-Myc-BRD2-Cter, pCMV-Myc-BRD3-Cter or pCMV-Myc-BRD4-Cter encoding plasmids. Transfection was carried out using the Lipofectamine 2000 transfection reagent (*Invitrogen*). Forty-eight hours later, transfected cells were trypsinized, pelleted and resuspended in the IP buffer (50 mM Tris, pH 8, 150 mM NaCl, 1% Nonidet-P40, 0.5% sodium deoxycholate) supplemented with a complete EDTA-free protease inhibitor cocktail (*Roche*). The resuspended cells were then disrupted by sonication and incubated on ice for 30 min before being centrifugated for 10 min at 4 °C at 12,000 g. The protein concentration of the supernatant (cell lysate) was determined by the Bradford assay. For the IP reaction, 20 µl of pure proteome Protein G Magnetic Beads (*Millipore*) were washed and incubated for 15 min in the IP buffer with 4 µg of a polyclonal rabbit anti-Myc (*Sigma-Aldrich*, catalog no. C3956) on a rotator at room temperature. The magnetic Myc-beads were then washed three times with 500 µl of the IP buffer before the addition of 500 µg of the cell lysate to 600 µl of the magnetic Myc-beads in the IP buffer. The mixture was then incubated overnight at 4 °C on a rotator. The next day, the magnetic beads were washed three times with 500 µl of IP buffer and then boiled in 60 µl of Laemmli buffer containing 5% β-mercaptoethanol. The protein mixture was analyzed by SDS-PAGE and immuno-detected by Western blotting.

**Immunodetection.** For immunoblotting, 20 µg of the whole cell extracts or 20 µl of the immuno-precipitated proteins were separated on a 10% SDS-PAGE. The proteins were transferred to a nitrocellulose membrane and detected either (i) with a mouse anti-Myc antibody (*Sigma-Aldrich*, catalog no. M4439, dilution 1/5000) and an anti-mouse IgG-Horseradish Peroxidase antibody (*Sigma-Aldrich*, catalog no. A5906, dilution 1/10,000) or (ii) with an anti-Flag M2 Horseradish Peroxidase (HRP) antibody (*Sigma-Aldrich*, catalog no. A8592, dilution 1/10,000). The Western blots were developed using the ECL chemiluminescent HRP substrate (*Thermo Scientific*).

#### 2.4. Protein purification

**Purification of the PERV IN CTD – BRD2 ET complex.** After transformation, the protein complex was expressed in T7 Express

competent *E. coli* in LB broth by induction with 1 mM isopropyl-β-D-thiogalactopyranoside (IPTG) at 37 °C for 3 h. Bacteria were harvested by centrifugation at 6,000 g for 10 min and stored at –20 °C. The thawed bacterial pellet was resuspended in buffer A (20 mM Tris HCl, pH 8.0, 0.5 M NaCl, 25 mM imidazole) in the presence of lysozyme (0.2 mg/ml, *Sigma*) and complete ULTRA EDTA-free protease inhibitor cocktail 1X (*Roche*). Following sonication for 8 min at 40 mA on ice, the lysate was clarified by centrifugation at 14,000 g for 45 min and filtered through a 0.45 µm filter (*VWR International*). The supernatant containing soluble His-tagged BRD2 ET, Strep-tagged II PERV IN CTD and the resulting double-tagged complex was loaded on a Ni<sup>2+</sup> charged 5 ml Protino<sup>®</sup> Ni-NTA (*Macherey Nagel*) equilibrated with buffer A using the ÄKTA purifier system (*GE Healthcare*). The column was extensively washed with buffer B (20 mM Tris HCl, pH 8.0, 1 M NaCl, 25 mM imidazole). The His-tagged BRD2 ET and the double-tagged complex were eluted by a linear gradient of imidazole (0.025–1 M). Eluted fractions were diluted 1:4 with buffer C (20 mM Tris HCl, pH 8, 0.5 M NaCl) and immediately loaded on a 5 ml Strep-Tactin affinity column (Strep-Tactin Superflow<sup>®</sup> cartridge, *IBA*) equilibrated with buffer W (100 mM Tris HCl, pH 8.0, 150 mM NaCl, 1 mM EDTA). The double-tagged complex was eluted with a desthiobiotin-containing buffer (100 mM Tris HCl, pH 8.0, 150 mM NaCl, 1 mM EDTA, 2.5 mM desthiobiotin). After 15% SDS-PAGE analysis, purified protein complex aliquots were flash frozen in liquid N<sub>2</sub> and stored at –80 °C. Each partner of the complex was identified by western blot and MALDI-TOF (Voyager DE-PRO, *AB Sciex*).

**Purification of the PERV IN CTD – BRD2 (667–806) complex.** A similar protocol was used to purify the complex and the two related mutants (*i.e.* W379A, and R391A/L392A mutated on IN).

#### 2.5. Structural studies

**Determination of the oligomeric state of the complex.** PERV IN CTD – BRD2 ET was concentrated at 14 mg/ml using a Vivaspine-PES-10K centrifugal concentrator (*Sartorius*) and loaded on Superdex 200 10/300 GL (*GE Healthcare*) equilibrated with buffer C for analysis by Size Exclusion Chromatography (SEC). The main peak fraction was concentrated to 10 mg/ml and analyzed by Dynamic Light Scattering (DLS) with a Zetasizer Nano S ZEN 1600 instrument (*Malvern Instruments*). Each measure is the mean of 10 runs repeated three times. A cross-linking experiment with glutaraldehyde was performed by mixing 10 µg of purified protein complex, 20 µl of 20 mM Tris HCl, pH 8 and 5 µl of 0.1% glutaraldehyde. After 20 min of incubation at room temperature, the cross-linking reaction was stopped by adding 5 µl of 200 mM glycine. The resulting protein sample was loaded onto a 15% SDS PAGE. A similar protocol was used for PERV IN CTD – BRD2 (667–808).

**SAXS study of the complex.** SAXS data were collected at the European Synchrotron Radiation Facility (ESRF) beamline BM29, Grenoble, France. Sample and buffer were centrifuged at 14,000 g for 10 min before acquisition to remove aggregates. The complex, at a concentration of 15 mg/ml, was injected in SEC coupled mode using an analytical Superdex S200 increase 5/150 GL column (*GE Healthcare*) equilibrated in buffer C. The temperature was maintained at 4 °C. Processing and analysis of SEC-SAXS data were performed with the programs CHROMIXS and SAS-DATA of the ATSAS suite ([Franke et al., 2017](#)). The radius of gyration (R<sub>g</sub>) was calculated from both the Guinier plot and the pair-distribution function P(r). The maximum inter-atomic distance of particle (D<sub>max</sub>) was calculated from P(r) analysis. *Ab initio* bead models were generated using DAMMIN. After superposition and averaging, the filtered SAXS envelope was displayed with CHIMERA ([Pettersen et al., 2004](#)).

A sequence search performed with SWISS-MODEL ([Waterhouse et al., 2018](#)) showed that the C-terminal fragment 736–808 of BRD2 has no homologous sequence of known structure in the Protein Data Bank (PDB). In consequence, the generation of a SAXS-restrained atomic model was possible for PERV IN CTD – BRD2 ET (667–735) only.

First, the NMR structure of MLV IN EBM in complex with BRD4 ET (PDB code 2n3k) was taken as a guide to model with the program SCULPTOR (Bunkoczi and Read, 2011) the structure of PERV IN EBM (aa 378–394) in complex with BRD2 ET. This model of the bound C-terminal tail was linked with an arbitrary relative position and orientation with the structure of the remaining CTD (aa 309–377) extracted from our model of the PERV intasome (Demange et al., 2015). The resulting homology model was fitted against experimental SAXS data with the program DADIMODO (Evrard et al., 2011) using a two domain rigid-body strategy with the loop 376–380 connecting the IN CTD core to the bound EBM segment rendered as flexible. The goodness of fit was estimated by  $\chi^2$  calculation with CRYSOLO (Franke et al., 2017). The resulting SAXS-restrained model was positioned with CHIMERA into the SAXS envelope for observation.

### 3. Results

#### 3.1. The human BRD2 protein interacts specifically with PERV and MLV INs

We first analyzed the interactions between different retroviral INs and BRD2 in the yeast two-hybrid assay (Fig. 1B). We tested the INs of the gamma-retroviruses PERV and MLV, the lentivirus HIV-1 and the alpha-retrovirus Rous Associated Virus type 1 (RAV-1) (Fig. 1B, left table). The full-length PERV IN (aa 1–405), MLV IN (aa 1–408), HIV-1 IN (aa 1–288), and RAV-1 IN (aa 1–286) were fused to the Gal4-DNA binding domain (Gal4-BD). The full length ORF of BRD2 (836 residues long) was fused to the Gal4 activation domain (Gal4-AD). The yeast strain was analyzed for histidine and adenine auxotrophy (-His/-Ade) acquired by the interaction of the fused proteins (Fig. 1B, left table). As expected, no growth was observed if PERV and MLV INs or BRD2 were not fused to their respective Gal4-BD and -AD genes. In accordance with others (Gupta et al., 2013), we detected no interaction between RAV-1 or HIV-1 INs and BRD2. We confirmed that MLV IN interacts with BRD2 and showed that PERV IN gave a positive interaction (Fig. 1B).

#### 3.2. PERV IN and BRD2 co-localize in the nucleus of cells

To further investigate the interaction between BRD2 and PERV IN, we used immunofluorescent microscopy to examine the localization of the full-length Flag-PERV IN and the Myc-BRD2 proteins in HEK 293T cells. In the first experiment, the pCMV-Flag-PERV IN and the pCMV-Myc-BRD2 plasmids were individually transfected into HEK 293T cells. The correct expression of each protein was checked by western blot analysis (data not shown). The cellular distribution of each fusion protein was analyzed using confocal fluorescent microscopy 48 h later (Fig. 2). More than 30 positive cells were observed in each sample. In HEK 293T cells, exogenously expressed Flag-PERV IN was mostly detected in the nucleus (either diffused (top images) or as punctuate nuclear foci (bottom images)) with a minority expression in cytoplasm (Fig. 2A). In HEK 293T cells, exogenously expressed Myc-BRD2 showed diffuse nuclear localization (main pattern) or both a nuclear and cytoplasmic localization (Fig. 2B).

In the second experiment, the pCMV-Flag-PERV IN and the pCMV-Myc-BRD2 plasmids were co-transfected into HEK 293T cells. Several patterns of co-localization (in yellow) were observed (Fig. 2C): (i) at the internal periphery of the nucleus (top images) and (ii) diffuse and punctuate in the nucleus (middle images). We also observed a chromosomal co-localization in a dividing cell (bottom images). Although a  $\chi^2$  statistical test did not reveal any change in the localization of the proteins — whether they were expressed alone or together (data not shown) — these co-localization studies provide further evidence supporting an interaction between PERV IN and BRD2 in cytoplasm and nucleus of cells.

#### 3.3. The BRD2 binding site maps within the C-terminal tail of PERV IN CTD

The retroviral IN protein contains three evolutionarily conserved and functional domains interconnected with flexible linkers: the N-terminal domain (NTD), the catalytic core domain (CCD) and the C-terminal DNA binding domain (CTD) (Fig. 1A), all necessary for the biological activities of the IN protein. The predicted structure of PERV IN also comprises a singular 46-residue long N-terminal extension domain (NED) (Demange et al., 2015). This NED structure is also observed in MLV and prototype foamy virus (PFV) IN.

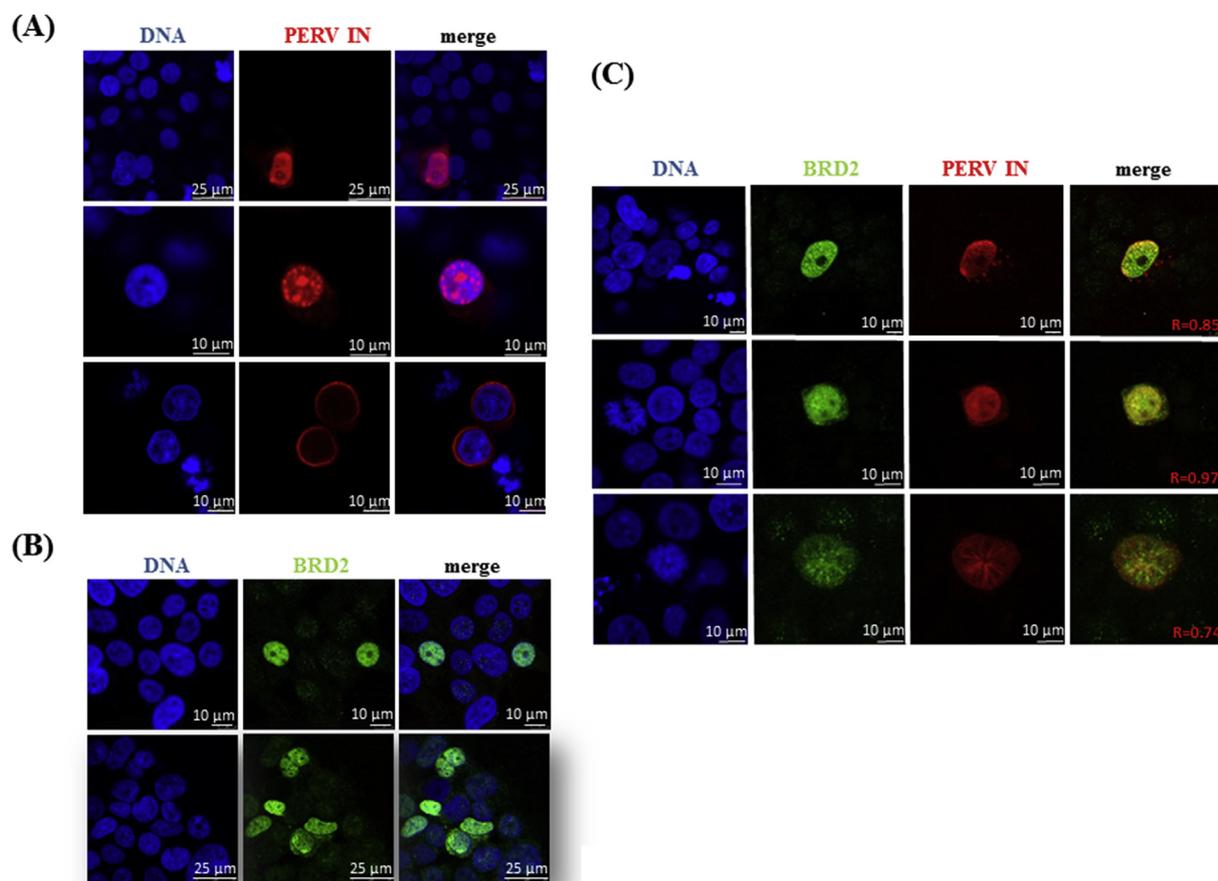
In order to map the region of PERV IN involved in the interaction with BRD2, we constructed a total of five IN truncation mutants fused with Gal4-DB: i.e. the one-domain fragments NTD (aa 1–98), CCD (aa 99–272) and CTD (aa 273–405) and the two-domain fragments NTD + CCD (aa 1–272) and CCD + CTD (aa 99–405) (see Fig. 1A). These five fragments were tested in the yeast two-hybrid system against the Gal4-AD-BRD2 protein (Fig. 3A). BRD2 interacts with PERV IN (99–405) and IN (273–405) but not with IN (1–98), IN (99–272) and IN (1–272). Therefore, only constructs containing PERV IN CTD can bind efficiently to BRD2 and the NTD and CCD are dispensable. Thus, our data showed that the CTD alone (aa 273–405) is necessary and sufficient for the interaction with BRD2. Similar deletion mutants were constructed with MLV IN and tested using the yeast two-hybrid system against Gal4-AD-BRD2 (Supplementary Fig. S1). In accordance with data from other studies (De Rijck et al., 2013; Gupta et al., 2013; Sharma et al., 2013), we observe that the CTD of MLV IN is sufficient for the interaction with BRD2. Altogether, these results demonstrate that INs from gamma-retroviruses interact specifically with BRD2 through their CTD domains.

To characterize more precisely the peptide sequence of IN required for the interaction, we used two CCD + CTD constructs deleted from the C-terminal sequences of the CTD, PERV IN (99–394) and IN (99–372) (Fig. 3B). The truncation of the 11 C-terminal residues of the CTD (PERV IN (99–394)) did not disturb the interaction. By contrast, removal of the 33 C-terminal residues of the CTD in PERV IN (99–372) resulted in a loss of interaction. This deletion study indicated that several residues from the segment 372–394 are necessary to mediate the interaction with BRD2 (Fig. 3B). We also tested positively the short IN fragment 372–405 carrying this interaction sequence (Fig. 3B) showing that the C-terminal tail of PERV IN (residues 372–405), alone, is sufficient for interaction with BRD2.

We then focused on the most C-terminal motif of CTD encompassing residues 378–394 in PERV IN. This segment corresponds to the MLV IN EBM (aa 389–405) which is critical for the interaction with BRD4 (Crowe et al., 2016). The primary sequence of this region is highly conserved among the gamma-retroviral INs (Fig. 4A) and not between retroviral genera (Supplementary Fig. S2). To assess the importance of these conserved residues, we performed a two-hybrid assay using five different mutants: the W379A single mutant (mutant containing an Alanine in place of the W379) and the P387A/L388A, K389A/L390A, R391A/L392A and H393A/R394A double mutants (Fig. 4B left panel). All of these mutants affect the interaction between PERV IN and BRD2 (Fig. 4B right panel). Notably the W379A single mutant disrupts the interaction in accordance with what was previously shown for the equivalent MLV W390A IN mutant and BRD4 (El Ashkar et al., 2014). This means that not only W379 but at least one residue in each double mutant (387–388, 389–390, 391–392 and 393–394) is important for the formation of the PERV IN – BRD2 complex.

PERV IN interacts with the ET domain of different BRD proteins in the two-hybrid assay and in HEK293T cells.

BET proteins such as BRD2 (Fig. 5A), BRD3 and BRD4 contain two conserved tandem bromodomains and a conserved ET domain. The ET from BRD4 is known to be involved in the interaction with MLV IN (Crowe et al., 2016; Gupta et al., 2013). We first tested the interaction of PERV IN with ET from different BET proteins using the two-hybrid system (Fig. 5B). A positive signal was detected with all constructs. The



**Fig. 2. Intracellular co-localization of the full-length Myc-BRD2 and Flag-IN PERV proteins in HEK 293T cells.** HEK 293T cells were transfected with the pCMV-Flag-PERV IN plasmid or/and with the pCMV-Myc-BRD2 plasmids. 48 h after transfection, cells were fixed and confocal laser scanning microscopy was used to study expression of the fluorescent fusion proteins. Cells were also colored with DAPI to show DNA. In (A), cells were transfected with the pCMV-Flag-PERV IN plasmid. The DNA (in blue) and the Flag (in red) signals and their overlays are shown in the left, middle, and right images of each lane, respectively. Flag-PERV IN was localized to both the nucleus and—to a lesser extent—the cytoplasm. In (B), cells were transfected with the pCMV-Myc-BRD2 plasmid. Myc (in green) and DNA (in blue) signals and their overlays are shown in the top left, bottom left and right-hand images of the panel, respectively. Expression of Myc-BRD2 is mainly nuclear (top and bottom images) or both nuclear and cytoplasmic (bottom image). In (C), 293T cells were co-transfected with both the pCMV-Flag-PERV IN and the pCMV-Myc-BRD2 plasmids. The Flag-PERV IN (in red), Myc-BRD2 (in green) and DNA (in blue) signals are shown on the left of the images. A Pearson coefficient (in brackets) greater than 0.75 indicates a good co-localization of the two proteins.

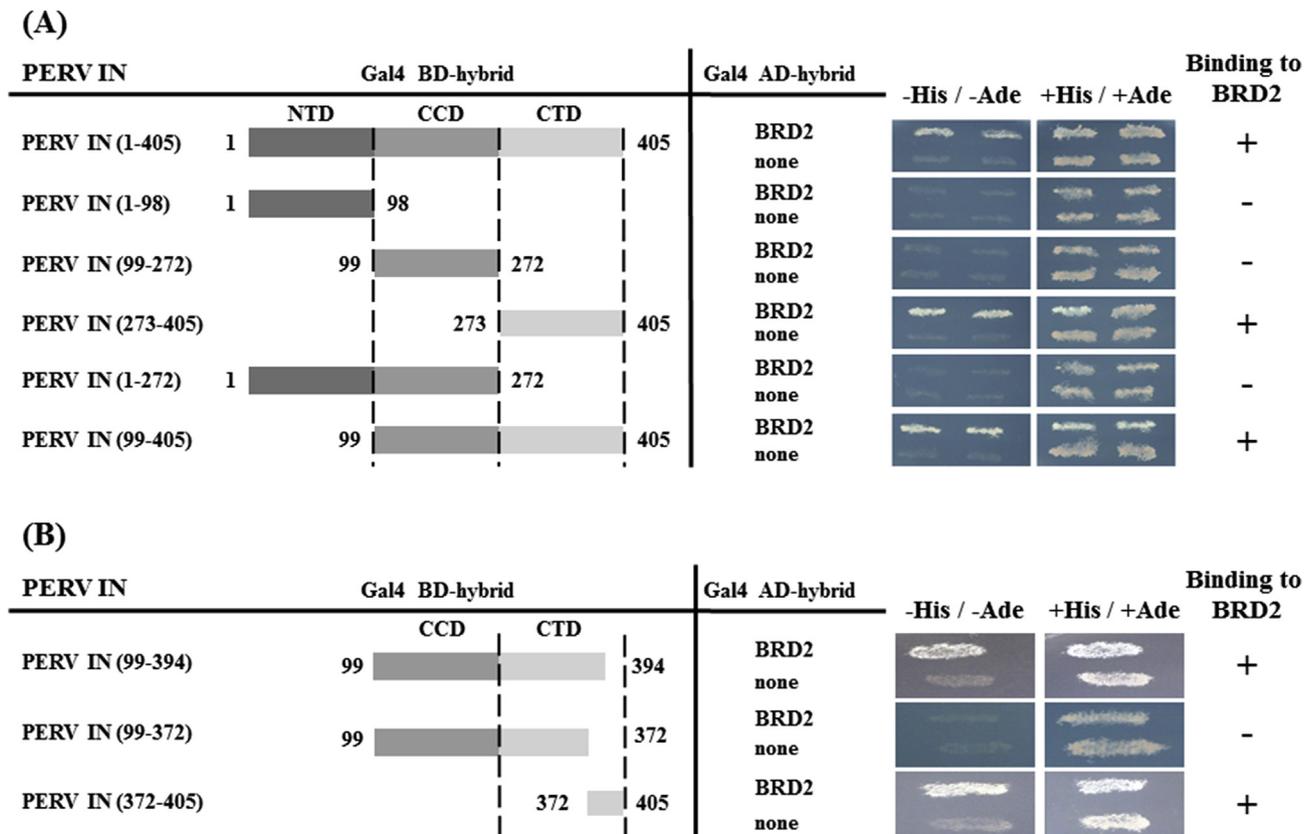
BRD4 ET fragment alone (aa 607–671), was sufficient for this interaction whereas the use of smaller fragments of ET of BRD2 or BRD4 did not show any interaction (data not shown).

We also investigated the interaction of the PERV IN CTD and the hBET C-terminal domain in HEK 293T cells. PERV IN CTD and hBET ET-containing C-terminal region (BRD2-Cter, BRD3-Cter and BRD4-Cter) were tagged with the Flag and Myc tags, respectively. The Flag-PERV IN CTD was tested for its ability to co-immuno-precipitate the Myc-hBET-Cter after transfection in HEK 293T cells (Fig. 5C). As a control, we used a PERV IN CTD (P387A/L388A) mutant containing two alanines in place of P387 and L388 in the interacting domain of IN. The Flag-PERV IN-CTD and the Flag-PERV IN-CTD (P387A/L388A) co-immunoprecipitation with Myc-hBET-Cter were assessed in the Myc-immunoprecipitated fractions using a Flag antibody (Fig. 5C). Data showed that the Flag-PERV IN-CTD was co-immunoprecipitated with Myc-BRD2-Cter, Myc-BRD3-Cter and Myc-BRD4-Cter (panel D, lanes 2, 5 and 8). The mutation of two residues in the Flag-PERV IN-CTD (P387A/L388A) diminished but did not fully abolish the interaction with the Myc-BRD2-Cter and Myc-BRD4-Cter (panel D, lanes 3 and 9, respectively). By contrast, the interaction of Myc-BRD3-Cter with the Flag-PERV IN-CTD (P387A/L388A) was completely lost (panel D, lane 6).

#### 3.4. Co-purification of the two proteins as a complex

With the aim of strengthening our data and obtaining novel structural information about the IN/Brd2 complex, we overexpressed and purified recombinant PERV IN CTD and the BRD2 C-terminal fragment from a prokaryotic expression system and performed *in vitro* interaction assays.

To date, structural knowledge on the C-terminal domain of BET proteins is limited to the ET (Crowe et al., 2016; Sanchez and Zhou, 2009). A prediction of intrinsic disorder by the GLOBPLOT server (Linding et al., 2003) shows that the C-terminal region of BRD2 (aa 667–836) that contains the ET (aa 676–738) is mostly ordered with the exception of the C-terminal extremity which is rich in serine residues (Supplementary Fig. S3). Since this long flexible tail can prevent crystallization of the protein and—in doing so—the use of X-ray crystallography to determine the atomic structure of the complex, we worked on the BRD2 fragment without the C-terminal tail. We produced independently PERV IN CTD (aa 309–405) and the BRD2 fragment (aa 667–808) to form the complex, but an aggregation phenomenon was observed when the two partners were mixed together (data not shown). To overcome this problem, we used a new strategy of protein co-expression inspired by Levy and co-workers that have already expressed HIV-1 IN and LEDGF/p75 recombinant proteins from a



**Fig. 3.** The CTD domain of PERV IN (and more specifically, residues 372–405) interacts with BRD2 in the two-hybrid assay. **(A)** The PERV IN CTD is involved in the interaction with BRD2 protein. The NTD (aa 1–98), CCD (aa 99–272) or CTD (aa 273–405) isolated domains of PERV IN as well as combined domains of PERV IN (NTD + CTD (aa 1–272) and CCD + CTD (aa 99–405)) were fused to the Gal4-BD domain. Interaction with BRD2 was evaluated as described in Fig. 1. **(B)** Refinement of the PERV IN CTD interaction domain. PERV IN fragments 99–394, 99–372 and 372–405 were fused to the Gal4-BD domain. Interaction with BRD2 was evaluated as described in Fig. 1B.

sophisticated polycistronic construction in a mammalian cells expression system (Levy et al., 2016). We optimized the same kind of construction for a bacterial expression system (Fig. 6A). A Strep-tagged-PERV IN fragment (aa 309–405) and a His-tagged-BRD2 fragment (aa 667–808) were cloned in frame with Tobacco Etch Virus (TEV) protease and interspaced with TEV digestion sites to form a unique, cleavable, polypeptide. During overexpression, the TEV protease separates the Strep-PERV IN CTD and the His-BRD2 (667–808) of the polyprotein leading to formation of the complex. The complex was then purified by two successive affinity steps with nickel and Strep-Tactin column (Fig. 6B top left panel). The two partners were identified by western blot using anti-His tag and Anti-Strep tag antibodies (Fig. 6B bottom left panels) and mass spectrometry from gel bands (data not shown). When using a wild type Strep-PERV IN CTD and the His-BRD2 (667–808), we were able to co-purify both proteins in the elution of the nickel column (lane 2) and also in the elution sample of the strep column (lane 3). This indicated that both partners, His-BRD2 (667–808) and Strep-PERV IN CTD were indeed co-purified as a complex form and not as two isolated proteins.

In order to assess the interaction of the previously described IN mutants with BRD2, the PERV IN CTD was modified within this construction to obtain the PERV IN CTD- W379A and PERV IN CTD-R391A/L392A mutants. Using these constructions, both proteins were efficiently co-expressed (Fig. 6B, lanes 4 and 7). However, after passing on the nickel column, although a small proportion of the strep-PERV IN CTD mutants were retained (lanes 5 and 8), the His-BRD2 (667–808) fragment was not retained on the Strep column (lanes 6 and 9). This observation strongly supports the interaction of BRD2 with PERV IN CTD and assesses the critical role of each conserved residue of the IN

EBM region.

We also performed the co-purification of the shorter PERV IN CTD – BRD2 complex, hereafter referred as to PERV IN CTD – BRD2 ET, by using the TEV expression system. We confirmed the interaction of this new two-fragment complex *in vitro*, as described below.

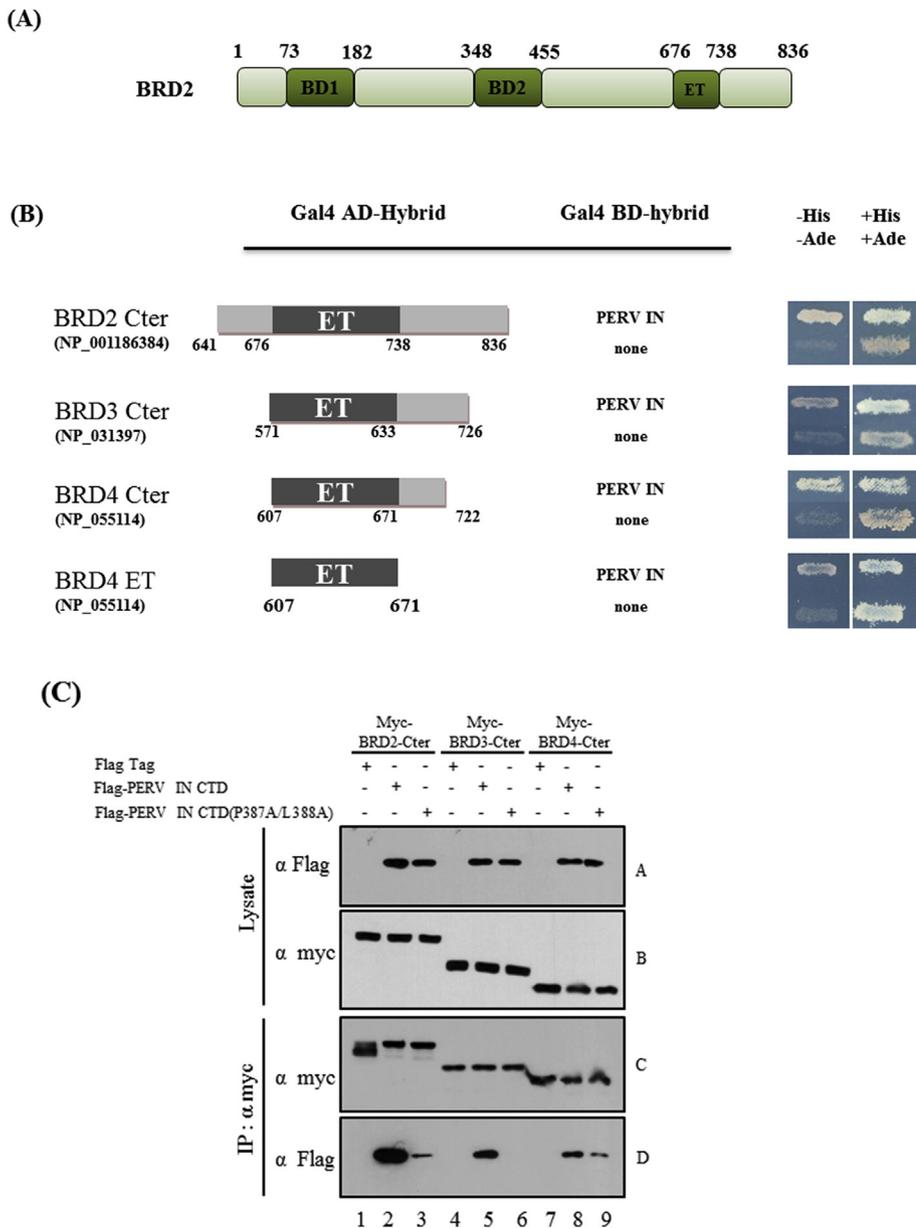
### 3.5. The complex is heterodimeric in solution and assembles as distinct modules

We next investigated the oligomeric state of the complex by Size Exclusion Chromatography (SEC) and cross-linking approaches.

In more details for PERV IN CTD – BRD2 ET, the SEC experiment indicated one main peak at an elution volume of 16.5 ml and a minor peak at an elution volume of 17.5 ml, corresponding to ~23 kDa (heterodimer) and ~13 kDa (IN CTD), respectively (Supplementary Figs. S4A and S4B). This demonstrated that the two-fragment complex is mainly heterodimeric at 14 mg/ml in 20 mM Tris HCl pH 8.0, 500 mM NaCl. Fractions from the main peak were pooled, concentrated and sample homogeneity was assessed by diffuse light scattering, resulting in a satisfactory polydispersity index of 0.2.

The heterodimeric state of PERV IN CTD – BRD2 ET was further confirmed by a chemically cross-linking assay using glutaraldehyde (Supp. Fig. 4C). Indeed, in the presence of glutaraldehyde, the covalently bound PERV IN CTD and BRD2 ET migrate with a higher molecular weight of about 23 kDa (shown on lane 1). The heterodimeric state of the cross-linked sample was confirmed by mass spectrometry from gel bands (data not shown). The heterodimeric state of the cross-linked PERV IN CTD – BRD2 (667–808) was checked with a similar protocol (data not shown).





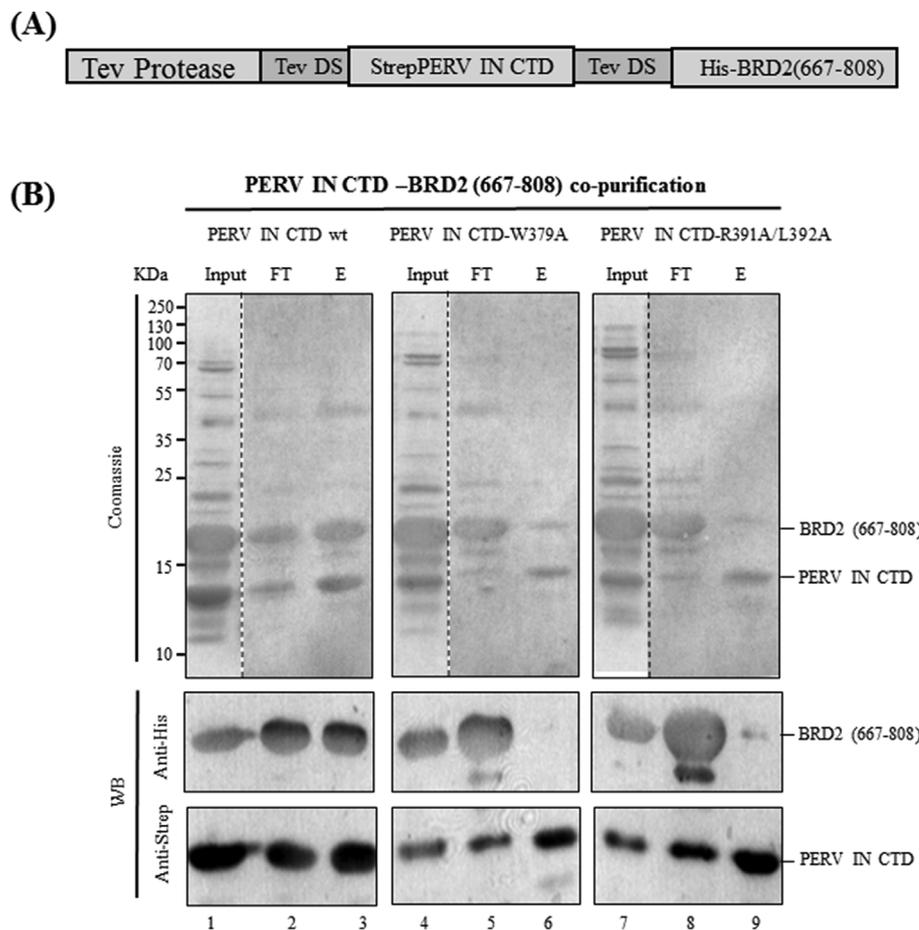
**Fig. 5. The C-terminal domains of BRD2, BRD3 and BRD4 proteins interact with PERV IN in the two-hybrid and *in cellulo* assays. (A) Domain organization of the BRD2 proteins. The hBET proteins contain several functional domains: (i) the bromodomains I and II (BD1 and BD2) bind acetylated marks on histones H3 and H4, (ii) the Extra-Terminal domain (ET) interacts with various viral and cellular proteins. Adapted from (Weidner-Glunde et al., 2010). (B) Two-hybrid assay using the full-length PERV IN and the C-terminal domains of hBET proteins. The C-terminal domains of BRD2, BRD3 and BRD4 proteins encompassing the ET domain, or the BRD4 ET domain alone were fused to the Gal4-activation domain (GAL4-AD). The ability of the full-length PERV IN to interact with these domains was analyzed in the two-hybrid assay as described in Fig. 1. (C) *In cellulo* assay: co-immunoprecipitation. HEK 293T cells were co-transfected: (i) with the pCMV-Myc-BRD2-Cter (lanes 1, 2 and 3), the pCMV-Myc-BRD3-Cter (lanes 4, 5 and 6) or the pCMV-Myc-BRD4-Cter (lanes 7, 8 and 9) and (ii) with the pCI-Flag-PERV IN-CTD (lanes 2, 5 and 8) or the pCI-Flag-PERV IN-CTD (P387A/L388A) (lanes 3, 6 and 9). Forty-eight hours later, cell lysates were collected. Expression of the Myc-BRD2-Cter, Myc-BRD3-Cter and Myc-BRD4-Cter proteins and of the Flag-PERV IN CTD or Flag-PERV IN-CTD (P387A/L388A) proteins was verified in the total cell lysates with anti-Myc and anti-Flag antibodies, respectively (panels A and B). Myc-BRD2-Cter, Myc-BRD3-Cter and Myc-BRD4-Cter proteins were then immuno-precipitated with the anti-Myc antibody and revealed with the Myc antibody (panel C). Co-immunoprecipitation of the PERV IN-CTD and PERV IN-CTD (P387A/L388A) proteins was assessed by a western blot analysis using an anti-Flag antibody (panel D).**

terminal peptide tail, abolish the interaction with BRD2. This segment is equivalent in sequence position with the BET-interaction motif EBM characterized in MLV IN (Crowe et al., 2016; Larue et al., 2014) (Fig. 4A), thereby confirming that gamma-retroviral INs share a common mechanism of selection of BET proteins. We also observed that the W379 PERV IN residue in the CCD is essential for interaction with BRD2 in accordance with other authors that established that the corresponding residue W390 from the MLV IN CTD is a critical hotspot for interaction with the BRD4 protein (El Ashkar et al., 2014). We also showed that, besides W379, at least one residue in each double mutant of the PERV IN EBM 387–394 stretch is also essential for the interaction. Others reported that residues (E266, L268 and Y269 of MLV IN CCD) — located outside the MLV EBM motif — are also important for the interaction with BET proteins (Gupta et al., 2013), but substitution of these residues in the context of the full-length MLV IN did not abolish the interaction with Brd4 (El Ashkar et al., 2014). Finally, PERV IN CTD was shown to interact with the conserved BET ET domain (Fig. 5) as does MLV IN (Aiyer et al., 2014; Crowe et al., 2016).

The interaction with BET protein was shown to be important for the nuclear localization of MLV IN. Indeed, when expressed separately,

MLV IN and BET proteins show a diffuse cytoplasmic and nuclear localization, respectively. When co-expressed with BET proteins, IN MLV changes localization from cytoplasmic to nuclear (Gupta et al., 2013). By contrast, we found that the exogenously expressed full-length Flag-PERV IN and Myc-BRD2 proteins were detected mainly in the nucleus of 293T cells when expressed alone, while only a slight diffuse cytoplasmic localization was observed for both proteins. We did not observe any change in the localization of the Flag-PERV IN when co-expressed with the Myc-BRD2 protein. However, we clearly detected a co-localization of the two protein domains in the nucleus and cytoplasm of cells when expressed together, which confirms the interaction of both proteins *in cellulo* and strengthens the role of the BET protein to direct PERV IN integration (Fig. 2).

All of these results are in accordance with results obtained by others who showed that BET proteins interact with the MLV IN protein and that both proteins co-localize into the nucleus of the cells (De Rijck et al., 2013; Gupta et al., 2013). Interaction between the two partners has been shown to involve the CTD of IN and the ET domain of BET proteins (Gupta et al., 2013; Sharma et al., 2013). Moreover, disruption of the MLV IN – BET interaction by using specific BET inhibitors, blocks



**Fig. 6. Analysis of PERV IN CTD interaction with hBET C-terminal domains *in vitro*.** (A) The pPERV IN CTD-BRD2 (667–808) plasmid is a polycistronic construction for the co-expression and co-purification of PERV IN CTD and BRD2 (667–808). This construction expresses a polyprotein containing the TEV protease and the Strep-PERV IN CTD (309–405) fragment with the His-BRD2 (667–808) fragment, both placed after a TEV digestion site (TEV DS). During overexpression, the TEV protease cleaves the TEV sites, if any, leaving the formation of a complex between PERV IN CTD and BRD2 (667–808) fragments. (B) SDS-PAGE gel illustrating the co-expression and co-purification of the wild-type (WT) Strep-PERV IN CTD, or of the Strep-PERV IN CTD- W379A and - R391A/L392A mutants with His-BRD2 (667–808). Two sequential affinity purifications were performed on nickel and Strep-Trap columns. Non-eluted samples (Lanes 1, 4 and 7; called “Input”) and samples eluted from the nickel column (lanes 2, 5 and 8; called flow-through “FT”) after eluted from the Strep-Trap column (lanes 3, 6 and 9; called “E”) were loaded on an SDS-PAGE gel; proteins were either revealed by Coomassie staining (upper panels) or were visualized by western blot using anti-His and anti-Strep-tag II antibodies targeting His-BRD2 (667–808) and Strep-PERV IN CTD, respectively (lower panels).

**Table 1**

Statistics of SAXS data collection and refinement from the two-fragment complexes.

Data collection	IN CTD - ET	IN CTD – BRD2 (667–808)
Analyzed fragment	IN CTD - ET	IN CTD – BRD2 (667–808)
Instrument	BM29 ESRF	BM29 ESRF
q-range (nm <sup>-1</sup> )	0–5	0–5
Exposure time (s)/nb frames	1/10	1/10
Concentration (mg/ml)	15	6
Temperature (°C)	4	4
Data processing		
Primary data reduction	EDNA	EDNA
Data processing	CHROMIXS	CHROMIXS
q-range (nm <sup>-1</sup> )	0.1–3.2	0.1–2.2
Rg (nm) [from Guinier]	2.4 ± .07	3.1 ± .1
Rg (nm) [from P(r)]	2.4	3.1
Dmax (nm)	7.4	11
Ab initio analysis	DAMMIF	DAMMIF
Number of models	50	50
Validation and averaging	DAMAVER	DAMAVER
NSD	0.87 ± .06	0.74 ± .07
Rigid-body modeling	DADIMODO	N.A.
Model $\chi^2$	1.0	N.A.

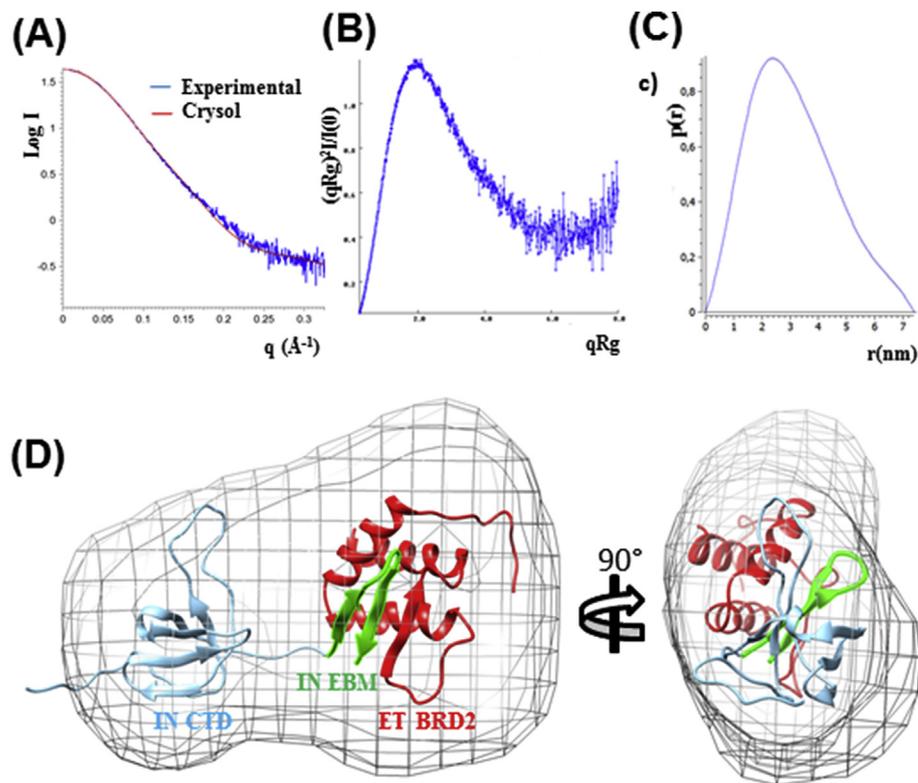
MLV replication at the integration step and — when integration occurs — retargets MLV integration away from transcription start sites (De Rijck et al., 2013; Gupta et al., 2013; Sharma et al., 2013).

We combined SAXS and homology modeling to generate the first experimentally restrained atomic model of a PERV IN CTD – BET ET complex. This structural study reveals that PERV IN CTD and BRD2 ET assemble as heterodimers to form a modular structure (Fig. 7D). The first part of the C-terminal tail of IN bridges the SH3 heart of the CTD to an intermolecular domain that contains the PERV IN EBM and the BET

ET.

Keeping in mind that the estimated resolution of the SAXS-derived envelope is 3 nm and that atomic details are inferred by homology-modeling, the heterodimeric assembly is likely stabilized by an intermolecular  $\beta$ -sheet with two EBM  $\beta$ -strands and one ET  $\beta$ -strand as observed in the NMR structure of bound MLV IN EBM (Crowe et al., 2016), which shares a strong sequence identity with PERV IN EBM – BRD2 ET.

The SAXS-restrained model can also be used with caution (at the atomic level, structural features must be conserved in bound MLV IN EBM) to propose structural explanations to our interaction results: (i) the single mutant W379A is disruptive and W379 is modeled as a buried residue making numerous intramolecular interactions, suggesting that this tryptophan is essential for the stability of the folded tail. Moreover, this residue is strictly conserved in gamma-retroviral INs and known as a critical hotspot for interaction; (ii) the disruptive double mutant P387A/L388A may alter the turn connecting the two  $\beta$ -strands of the modeled bound EBM (Fig. 3) and hinder the induced folding process; (iii) the disruptive mutants K389A/L390A and R391A/L392A (Figs. 4 and 6) cancel the -BHBH- motif of the conserved gamma-retroviral stretch, where -B- and -H- are basic and hydrophobic residues. Thus, the modeled intermolecular interactions between the basic side-chains and conserved acidic residues of the neighboring ET  $\beta$ -strand could be essential to the stability of the heterodimer. Of interest, an amphipathic -BHBH- binding motif has been observed in other viral and cellular partners of BET proteins—such as the transcriptional regulator NSD3—where residues equivalent in position to the pair R391/L392 are essential for ET recognition (Zhang et al., 2016); (iv) the disruptive double mutant H393A/R394A affects the C-terminal arginine of EBM, which is strictly conserved among gamma-retroviral INs and contacts the critical tryptophan W379 in the model.



**Fig. 7.** SAXS analysis of PERV IN CTD – BRD2 ET. (A) Experimental diffraction curve in blue and CRYSOLOG curve in red calculated from the atomic model. (B) Dimensionless Kratky plot with a plateau in the higher  $q$  region reflecting flexibility. (C) Distance distribution function  $P(r)$ . (D) *Ab initio* SAXS envelope and ribbon representation of the SAXS-restrained model with the CTD of PERV IN in blue, the EBM of IN in green and the bound ET domain of BRD2 in red. The figures A, B and C were rendered with ATSAS, and the figure D was rendered with CHIMERA.

Our SAXS data also reveal that the PERV IN CTD – BRD2 ET complex possesses an inherent flexibility due to a short loop connecting the bound IN EBM segment and the IN CTD core. This flexibility is also observed in the bigger PERV IN CTD – BRD2 (667–808) complex and might be advantageous in the context of the full-length proteins for a dynamic selection of the integration sites. Moreover, the recent publications of numerous intasome structures (Ballandras-Colas et al., 2016, 2017; Hare et al., 2010; Passos et al., 2017; Yin et al., 2016) allowed us to gain information on the formation of the PERV IN – BRD2 assembly.

The oligomerization state of the known intasome structure differs between retroviral genera and seems to be related to the length of the IN CCD-CTD linker (for review, see (Engelman and Cherepanov, 2017)). Gamma-retroviral intasomes should be tetrameric, as observed in the PFV intasome (Engelman and Cherepanov, 2017). Our previously published model of the PERV intasome respects this architecture (Demange et al., 2015) with two symmetrical inner and outer monomers of INs. The two inner CTDs are positioned between their corresponding inner NTDs and CCDs (Fig. 8), whereas the outer CTDs are not modeled because they were too flexible to be observed in the PFV intasome. A superposition with our SAXS-restrained model of the PERV IN CTD – BRD2 ET complex shows that the inner CTDs are well accessible to the ET domain of the BET cofactor (Fig. 8). This accessibility remains unchanged if we consider the intasome to be bound with target DNA as observed for PFV (Maertens et al., 2010). Altogether, these data suggest that, before integration, the accessible, flexible C-terminal tail of the inner IN CTD can hook the ET domain of the BET cofactor. The induced  $\beta$ -folding of the IN EBM segment locks the two partners together, while the short loop preceding IN EBM gives partial flexibility to the complex. This structural combination participates in the efficient targeting of the bound vDNA to the promoters of active genes.

## 5. Conclusion

Retroviral vectors are important tools and have been widely used to deliver DNA for therapeutic purposes. However, retroviral integration

can be associated with insertional oncogene activation, especially for gamma-retroviral vectors due to their strong bias of integration into transcription start sites. Eliminating the interaction of gamma-retroviral IN with hBET family members has implications for the safety of human gene therapy and gene delivery when using gamma-retroviral vectors. Besides, it has been shown that the PERV recombinant virus can infect human cells and represents a potential major risk of zoonotic disease in the case of xenotransplantation of pig organs to humans. We have previously shown that Raltegravir (Demange et al., 2015), as well as AZT (Denner, 2017), are potent inhibitors of PERV replication and could be useful for the treatment of PERV infections. Eliminating the interaction of PERV IN with hBET proteins using inhibitors such as JQ1 would be another way to control this zoonotic risk.

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Availability of data and material

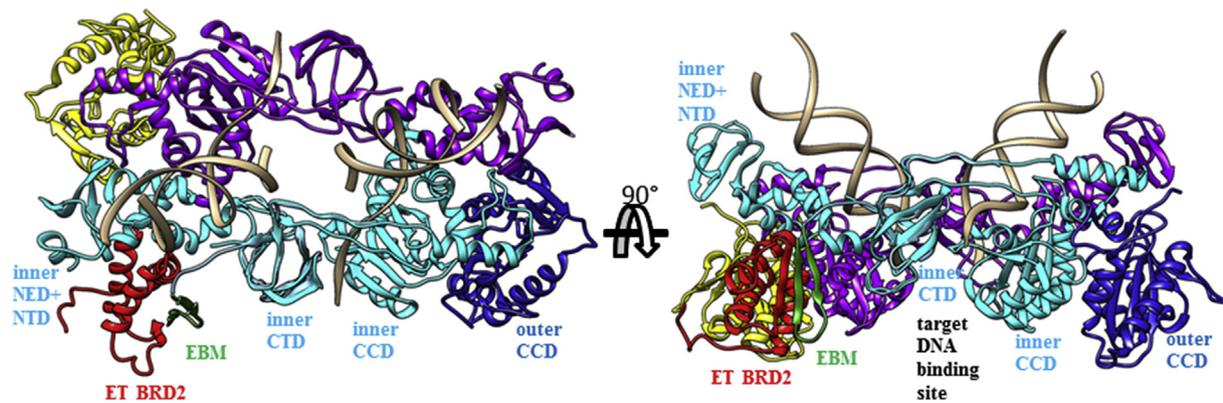
The datasets used and/or analyzed during the current study are available from the corresponding authors on reasonable request.

### Competing interests

The authors declare that they have no competing interests.

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**Fig. 8.** Ribbon representation of the modeled PERV IN intasome superposed with the SAXS-restrained model of PERV IN CTD – BRD2 ET. The two symmetric inner IN monomers are painted cyan and violet, whereas the two outer INs are blue and yellow. The two bound DNA ends are in grey. The PERV IN EBM segments and the bound BRD2 ET of the superposed SAXS-restrained model are colored in green and red, respectively. The inner CTD of the intasome and the CTD of the heterodimeric complex were superimposed with COOT. The figure was rendered with CHIMERA.

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#### Authors' contributions

CR, YB, KM and PG designed the experiments, analyzed data and drafted the manuscript. KG, GB, MC, HY, and FF made substantial contributions to conception, design, acquisition and analysis of data and drafted the manuscript. MPC, CDB, AL, CL performed the experiments. ML and YC discussed the conception and analysis of some experiments. All authors read and approved the final manuscript.

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#### Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.virol.2019.04.002>.

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