



## Neuroinflammation as a risk factor for attention deficit hyperactivity disorder



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### ABSTRACT

Attention Deficit Hyperactivity Disorder (ADHD) is a persistent, and impairing pediatric-onset neurodevelopmental condition. Its high prevalence, and recurrent controversy over its widespread identification and treatment, drive strong interest in its etiology and mechanisms. Emerging evidence for a role for neuroinflammation in ADHD pathophysiology is of great interest. This evidence includes 1) the above-chance comorbidity of ADHD with inflammatory and autoimmune disorders, 2) initial studies indicating an association with ADHD and increased serum cytokines, 3) preliminary evidence from genetic studies demonstrating associations between polymorphisms in genes associated with inflammatory pathways and ADHD, 4) emerging evidence that early life exposure to environmental factors may increase risk for ADHD via an inflammatory mechanism, and 5) mechanistic evidence from animal models of maternal immune activation documenting behavioral and neural outcomes consistent with ADHD. Prenatal exposure to inflammation is associated with changes in offspring brain development including reductions in cortical gray matter volume and the volume of certain cortical areas –parallel to observations associated with ADHD. Alterations in neurotransmitter systems, including the dopaminergic, serotonergic and glutamatergic systems, are observed in ADHD populations. Animal models provide strong evidence that development and function of these neurotransmitter systems are sensitive to exposure to in utero inflammation. In summary, accumulating evidence from human studies and animal models, while still incomplete, support a potential role for neuroinflammation in the pathophysiology of ADHD. Confirmation of this association and the underlying mechanisms have become valuable targets for research. If confirmed, such a picture may be important in opening new intervention routes.

### 1. Introduction

Attention Deficit Hyperactivity Disorder (ADHD) is a neurodevelopmental disorder characterized by pervasive and persistent behavioral symptoms of inattention, hyperactivity, and/or impulsivity that are extreme for age and interfere with social or academic functioning. Diagnosis requires the emergence of symptoms before twelve years of age and that symptoms present in two or more settings (APA, 2013). ADHD is common, affecting about 5% of children worldwide (APA, 2013) with boys being overrepresented, on average, approximately 2:1 (Danielson et al., 2018). The impact of ADHD is far reaching due its early onset and frequent persistence and secondary complications. Those with ADHD suffer reduced academic performance, weakened social and familial relationships, peer rejection, and long-term elevated rates of serious accidental injury, drug addiction, depression, school or

occupational failure, and involvement in the criminal justice system. Cognitive problems such as subtle reduction in IQ, working memory, and executive function are also common, though not universal, in children with ADHD (Dan and Raz, 2012; Nigg, 2005; Willcutt et al., 2005) and may be related to both early environmental stressors (Wiggs et al., 2016) and genetic risk for ADHD (Nigg et al., 2018). Although ADHD is usually thought of as a childhood affliction, 50% of sufferers continue to have impairing symptoms into adolescence and 30–60% have impairing symptoms as adults (Ahmed et al., 2014). Adults with ADHD have lower socioeconomic achievement and reduced life span (Doshi et al., 2012). New insights into ADHD's etiology and new treatment possibilities are thus of keen interest.

The etiology of ADHD is complex, involving both common and rare genetic variants as well as multiple environmental risk factors. Ample evidence indicates that ADHD is highly heritable, but lacks any genes of

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major effect except in very rare cases, although individual loci are reliably associated with ADHD in meta-analysis, and multiple forms of genetic influence are implicated (Faraone and Larsson, 2019; Wiggs et al., 2016). About a third of the heritability is accounted for by common genetic variation. The proportion related to genotype by environment interaction is unknown (discussed further below). Studies of shared SNP heritability indicate that ADHD shares genetic liability with many other common psychiatric conditions including anxiety and mood disorders (Anttila et al., 2018). At the same time, early environmental factors are also implicated in the etiology of ADHD including many factors associated with increased inflammation such as maternal infection (Werenberg Dreier et al., 2016), maternal smoking (Becker et al., 2008; Biederman, 2005; Mick et al., 2002; Neuman et al., 2007; Silva et al., 2014; Thapar et al., 2013), maternal obesity and poor diet (Andersen et al., 2018; Buss et al., 2012; Chen et al., 2014; Rijlaarsdam et al., 2017; Rodríguez, 2010; Rodríguez et al., 2008; Sanchez et al., 2018; Vermiglio et al., 2004), and maternal exposure to pollutants (Thapar et al., 2013). Effect sizes for individual risk factors are generally modest and causality is still being evaluated in novel designs (Nigg et al., 2016; Thapar et al., 2013). Environmental effects are generally non-specific in that other psychiatric outcomes are also associated with these risk factors. At the same time, their association with ADHD in many instances survives statistical adjustment for co-occurring psychiatric symptoms and conditions.

Genotype-by-environment interactions are increasingly established in the field of developmental psychopathology (Kim and Leventhal, 2015; Pietropaolo et al., 2017; Pinto et al., 2015; S.M. Schaafsma et al., 2017) and are emerging in ADHD as well (Elmore et al., 2016; Nigg et al., 2010). While statistical decompositions can distinguish genetic and non-genetic influences on individual differences in a population, from a biological point of view genes and environments always act in concert. Indeed, it is well-established that environmental factors, especially those experienced during early development, can have a long-lasting influence on gene expression via epigenetic signaling and through that route on neural development. Initial evidence supports the idea that genetically susceptible individuals are more likely to develop ADHD (and other complex diseases) if exposed to certain environmental risk factors (Nigg et al., 2016; Stevenson et al., 2010). Many of these risk factors often impact genes involved in the dopamine (Becker et al., 2008; Neuman et al., 2007) and serotonin systems (Cortese, 2012; Pennington et al., 2009). Yet, the mechanism of action of these genetic and environmental influences on ADHD is not known.

In that regard, it is noteworthy that many of the early environmental risk factors for ADHD increase the inflammatory profile of the in-utero environment (Costenbader and Karlson, 2006; Shankar et al., 2011; Terasaki and Schwarz, 2016), supporting the hypothesis that exposure to inflammation during development leads to neuroinflammation and may play a role in the pathophysiology of ADHD (Costenbader and Karlson, 2006; Shankar et al., 2011; Terasaki and Schwarz, 2016). In this review, we discuss the neurobiological developmental changes involved in ADHD, with respect to their possible link to neuroinflammation as a mechanism.

## 2. Peripheral inflammation

Several lines of evidence can be marshaled to support a hypothesis that inflammation is part of a pathway to ADHD. This is not surprising when one considers the numerous associations between inflammation and CNS development and function (Bhattacharya et al., 2016). For example, GWAS studies have identified associations between ADHD and the gene for IL-1RA (Segman et al., 2002) as well as genes involved in regulation of gene expression, cell adhesion, and inflammation (Zayats et al., 2015). As another example, atopic immune disorders such as eczema (Buske-Kirschbaum et al., 2013; Liao et al., 2016; Lin et al., 2016; Schmitt et al., 2009), asthma (Fasmer et al., 2011; Instanes et al., 2017; Lin et al., 2016), rheumatoid arthritis (Instanes et al.,

2017), type 1 diabetes (Instanes et al., 2017), and hypothyroidism (Instanes et al., 2017) have been associated with higher rates of ADHD diagnosis. Meta-analyses (Miyazaki et al., 2017; Schans et al., 2017; Schmitt et al., 2010) confirm that the odds of experiencing atopic immune disorders were slightly higher in children with ADHD than controls, however the effect estimates were quite variable across studies. Although these associations are variable between studies and study quality is sometimes weak, the data are consistent with a connection between the peripheral immune system and ADHD. Interestingly, recent evidence suggests that patients displaying ADHD symptomatology have higher serum cytokine levels than non-ADHD controls (Anand et al., 2017; Darwish et al., 2018; Donfrancesco et al., 2016a; Donfrancesco et al., 2016b; O'Shea et al., 2014). One study reported an association between pro-inflammatory serum cytokines and ADHD symptom severity (Oades et al., 2010b). However, again, these studies are preliminary in that they included small sample sizes and were heterogeneous in the molecules examined and methodology used impairing comparisons across studies (Anand et al., 2017). Further there is evidence that patients taking psychostimulants, such as methylphenidate have lower levels of cytokines than medication-naïve patients suggesting an effect of treatment or improved symptomatology on cytokine levels (Oades et al., 2010a) that was not well-controlled in most studies. Nonetheless, more examination of cytokine levels should prove interesting.

Other isolated but intriguing findings bear note. Higher levels of antibasal ganglia antibodies (Toto et al., 2015) and antibodies against the dopamine transporter (Giana et al., 2015) are reported in populations with ADHD and are noteworthy as signs of inflammation. Furthermore, in one study, ADHD patients were observed to have elevated cerebrospinal fluid levels of the pro-inflammatory cytokine TNF-beta and lower levels of the anti-inflammatory cytokine IL-4 (Mittleman et al., 1997). Besides modulation of the central immune system by cytokines, it is possible that peripheral monocytes and other immune cells penetrate the blood brain barrier to induce neuroinflammation. Under normal conditions the blood-brain barrier separates the peripheral immune system from the CNS and prevents peripheral immune cells from entering the CNS. However, when the blood-brain barrier is compromised from injury, disease, or even marked psychological stress, peripheral monocytes can enter the CNS (Wohleb et al., 2014) and possibly alter function of neurons and other glial cells. The ability of peripheral monocytes to cross the blood brain barrier during development is supported by studies of maternal immune activation (MIA) which detect a compromised blood brain barrier in the developing offspring (Stolp and Dziegielewska, 2009).

Indirect evidence for the role of peripheral inflammation in the etiology of ADHD comes from dietary interventions. Meta-analysis has indicated that children with ADHD tend to have lower blood levels of long chain fatty acids and that ADHD symptoms are partially ameliorated in response to supplemented omega-3 polyunsaturated fatty acids (Chang et al., 2018; Hawkey and Nigg, 2014). Early omega-3 supplementation of mothers causes improved attentional development in infants (Colombo et al., 2016). Omega-3 polyunsaturated fatty acids are well documented to have anti-inflammatory properties and are beneficial in certain chronic inflammatory diseases such as rheumatoid arthritis (Yates et al., 2014). These preliminary findings suggest an association between ADHD and altered peripheral inflammatory profile, supporting the hypothesis that modulation of the neuroinflammatory environment plays a role in the pathophysiology of ADHD.

## 3. Neuroinflammation hypothesis

Based on the preceding, a neuroinflammation hypothesis can be developed for many forms of developmental psychopathology including ADHD. Here we define neuroinflammation as a general term for inflammation of neural tissue. It is characterized by changes in microglia, astrocytes, cytokines, chemokines, and related molecular processes

within the CNS. It is important to note that a strict definition of neuroinflammation may limit it to inflammation in response to bacteria, parasite, or viruses and a more appropriate term for inflammation, in the context we reference throughout this paper, may be microglial activation or neural immune activation. For ADHD post-mortem studies are lacking, but for some disorders post-mortem studies confirm neuroinflammation and not just peripheral tissue inflammation (Faraone and Mick, 2010; Kim et al., 2016; Monji et al., 2013; Vargas et al., 2005). Neuroinflammation is proposed to influence brain development and subsequently increase risk of neurodevelopmental disorders by acting through mechanisms including glial activation (Reus et al., 2015), increased oxidative stress (Hassan et al., 2016), aberrant neuronal development (Belmadani et al., 2006), reduced neurotropic support (Sen et al., 2008), and altered neurotransmitter function (Kronfol and Remick, 2000).

Several early risk factors that have been proposed for ADHD, including maternal infection, low birth weight, fetal alcohol exposure, maternal stress, and maternal obesity, all share an increased maternal inflammatory profile, raising the possibility that inflammation during prenatal development may play a role in the pathophysiology of ADHD (Costenbader and Karlson, 2006; Shankar et al., 2011; Terasaki and Schwarz, 2016).

During prenatal development, neurons rely on concentration gradients of specific chemoattractant factors as well as supporting glial cells to migrate to appropriate locations (Ayala et al., 2007; Barry et al., 2014). Glial cells and the process of neuronal migration are highly sensitive to the inflammatory environment, because changes in inflammatory markers alter chemoattractant gradients and glial cell functioning (Metin et al., 2008). Microglia are the primary immunocompetent cells of the central nervous system (CNS). Initially, believed to be primarily involved in pathologic states, such as reacting to infection and damage to the brain, it is now clear that microglia perform a host of additional functions within the brain including synaptic pruning, neuronal phagocytosis, and refining network connectivity during development (Cunningham et al., 2013; Eyo and Dailey, 2013; Salter and Beggs, 2014). Additionally, recent studies have indicated that microglia exhibit sex-dependent phenotypes. For example, it appears that, in rodent models, male microglia exhibit a more proinflammatory phenotype and develop at delayed rates compared to females (Hanamsagar et al., 2018; Villa et al., 2018). These sex differences in microglia phenotypes in animals raise the intriguing possibility of a mechanism for why ADHD and most other early-onset neurodevelopmental conditions affect males at overrepresented rates.

The extensive involvement of microglia and astrocytes in neurodevelopment makes them a point of interest when discussing mechanistic origins of neurodevelopment disorders, such as ADHD. In rodent models of maternal immune activation (MIA), microglial and astrocytic functionality is altered to an inflammatory state (Mattei et al., 2017). These alterations, along with inflammation directly influence neural precursor cells, can affect neuronal development, potentially giving rise to subsequent behaviors often associated with ADHD such as hyperactivity and anxiety (see the animal models section below, Fig. 1).

#### 4. Neurobiological basis of ADHD: evidence from studies in humans

While a comprehensive coverage of neurobiological correlates of ADHD and other developmental psychopathologies is beyond the scope of this manuscript (for extended discussions see (Aoki et al., 2017; Bilbo and Schwarz, 2009; Dougherty et al., 2016; Lim et al., 2015; Martin et al., 2014; Rommelse et al., 2017), here we highlight selected findings relevant to the current discussion. All the findings discussed below occur at the group level; none are robust enough to be clinically relevant or diagnostic at the individual level as yet.

#### 4.1. Neuroanatomical changes associated with ADHD

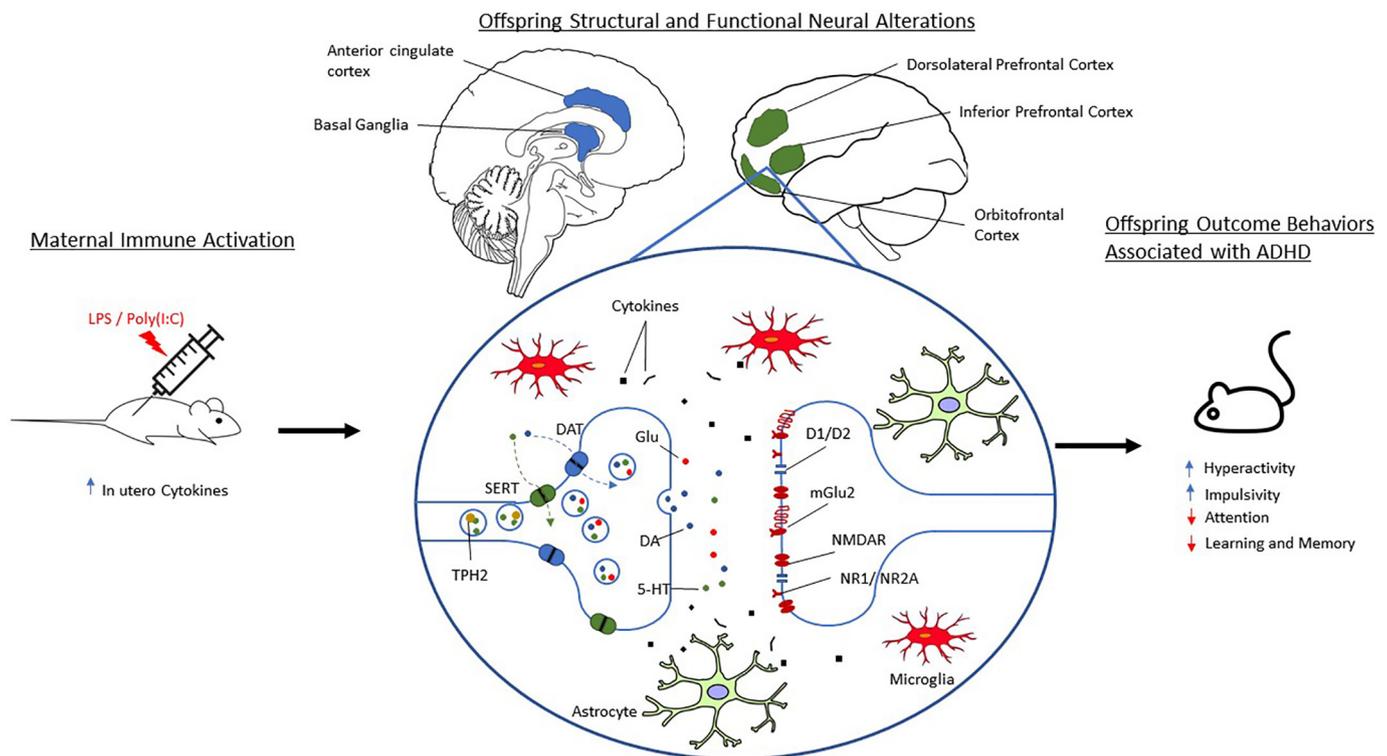
Briefly, CNS development begins 3–4 weeks post-conception with the folding and fusing of the ectoderm to form the neural tube. Subsequently, the prenatal CNS goes through a series of maturational events including neurogenesis and proliferation, neuronal migration, synapse formation and pruning, apoptosis, and finally myelination that extends into postnatal development (Marsh et al., 2008). In typical developing individuals, cortical gray matter development is characterized by increases in volume throughout the brain until puberty followed by reductions until full brain maturation in the third decade of life. The development of cortical volume is non-linear and asynchronous, with the frontal and parietal lobe volumes peaking at about 12 years of age, the temporal lobe at about 16 years of age, and the occipital lobe at about 20 years old (Giedd et al., 1999) while subcortical structures have their own asynchronous developmental timeline (Gilmore et al., 2012). Comparatively, children with ADHD have on average an approximately 4% reduction in overall gray matter volume in both the cerebrum and cerebellum, while developmental trajectories parallel typical development (Castellanos et al., 2002). This translates to key cortical regions reaching peak volume and beginning to prune about 3 years later than what occurs in typically developing children (Shaw et al., 2007). As microglia are highly involved in developmental pruning of synapses it is highly possible that this delayed pruning is related to neuroinflammation and altered microglial function. The neuroanatomical changes discussed here and later are focused on postnatally observed changes. However, alterations in brain development are likely occurring prenatally as well. However, to date there is limited understanding of the alterations in prenatal brain development that are associated with ADHD.

Moving into more specific structures, the primary cortical areas associated with ADHD symptomology are the prefrontal cortex (PFC) (Arnsten and Rubia, 2012), orbitofrontal cortex, and anterior cingulate cortex (Amico et al., 2011). Rubia and Arnsten (Arnsten and Rubia, 2012) point out that the PFC is viewed as important in the “top-down” regulation of attention, inhibition/cognitive control, emotion and motivation. A useful and common heuristic supposition is that attention and cognitive control are regulated relatively more by the dorsolateral and inferior PFC, while motivation and affect are regulated relatively more by the orbital and ventromedial PFC. Through a series of frontal-subcortical connections, these areas are involved in processes such as executive function, motivation, emotion, and evaluative assessment of stimuli. Children with ADHD show abnormalities in the inferior PFC and its connections to striatal, cerebellar, and parietal regions (Arnsten and Rubia, 2012). Findings indicate a reduction in gray matter volume and cortical thinning in these areas in children with ADHD (Li et al., 2015; Makris et al., 2007).

Reduced volume of subcortical structures such as the basal ganglia are also implicated in ADHD. Meta-analyses suggest that differences in structure and function of the basal ganglia represent a core finding in the ADHD literature (Hoogman et al., 2017). Relating to ADHD, these structures are associated with functions such as inhibitory control of action, attention, cognition, hyperactivity, and emotional response inhibition (de Wit et al., 2012; Grabli et al., 2004; van Rooij et al., 2015). The cerebellum has also been seen as potentially important in ADHD. These findings as they relate to ADHD as well other neurodevelopmental disorders have been extensively discussed and reviewed previously in both human studies and animal models (Stoodley, 2014, 2016; Thanellou and Green, 2013).

#### 4.2. Changes in neural function associated with ADHD

Although the monoamines glutamate, and gamma-aminobutyric acid (GABA) interact continually in complex ways, we discuss them individually for the sake of clarity.



**Fig. 1.** Neural and behavioral changes of offspring born from mothers injected with lipopolysaccharide (LPS) or polyinosinic:polycytidylic acid (poly(I:C)) associated with changes observed in ADHD. Maternal immune activation (MIA) by LPS or poly(I:C) results in an increase of in utero cytokines altering the inflammatory environment of the developing offspring. MIA results in structural changes in volume and grey matter volume are observed in the anterior cingulate cortex, basal ganglia, dorsolateral prefrontal cortex, inferior prefrontal cortex, and orbitofrontal cortex. Functional changes observed in MIA offspring are alterations in the homeostatic levels of the neurotransmitters serotonin (5-HT), dopamine (DA), and glutamate (Glu) as well as a reduction in tryptophan hydroxylase (TPH2), the rate limiting step of 5-HT synthesis. In addition, alterations are observed in functionality of serotonin reuptake transporters (SERT) and dopamine transporters (DAT), dopamine receptors (D1/D2), metabotropic glutamate receptors (mGlu2), ionotropic glutamate receptors (NR1/NR2A), and NMDA receptors (NMDAR). Changes in these neural outcomes are associated with behavioral outcomes of increased hyperactivity and impulsivity along side decreases in attention and learning and memory.

#### 4.2.1. Catecholamines (dopamine/norepinephrine)

Historically, ADHD was often considered a hypodopaminergic disorder (Levy, 1991; Robbins and Sahakian, 1979), where a deficiency in synaptic dopamine levels leads to clinical symptoms. Dopamine is a catecholamine synthesized mainly in the ventral tegmental area, substantia nigra, and arcuate nucleus, from the precursor amino acid tyrosine. Dopamine's role in ADHD has been supported circumstantially by pharmacological evidence that medications, like methylphenidate and amphetamines, increase levels of dopamine in the synaptic cleft, and also temporarily ameliorate symptoms of ADHD (Volkow et al., 2005). This “dopamine hypothesis” has undergone multiple, more nuanced revisions in recent years but remains a core line of thinking about ADHD (Volkow et al., 2009). Additionally, the activity of the dopamine derivative, norepinephrine (NE), has also been implicated in ADHD. The role of dopamine and catecholamines in ADHD has been previously and extensively reviewed (Del Campo et al., 2011; Volkow et al., 2009).

#### 4.2.2. Serotonin

Recent clinical, neuroanatomical, and genetic studies provide evidence for a role for serotonin (5-HT) in the etiology of ADHD. Serotonin is a monoamine synthesized from the essential amino acid tryptophan. The main site of 5-HT synthesis in the brain is the dorsal raphe nucleus. In the dorsal raphe 5-HT is synthesized from tryptophan by the rate limiting enzyme tryptophan hydroxylase 2 (TPH2). The 5-HT system is complex, containing 14 known receptor subtypes all of which are G-protein coupled with the exception of the ligand gated 5-HT<sub>3R</sub>. Serotonin is transported from the synaptic cleft to the presynaptic neuron via the serotonin transporter (5-HTT).

Although, drugs that target dopamine and norepinephrine pathways are the first line of treatment for ADHD, up to 30% of ADHD patients do not respond to this treatment and among responders only about 50% show signs of improvement (Arnold et al., 2013). Alternative treatments for ADHD include selective-serotonin re-uptake inhibitors (SSRIs), serotonin norepinephrine re-uptake inhibitors (SNRIs) and tricyclic-antidepressants (TCA) all of which target the 5-HT system (Park et al., 2014). Serotonin has been extensively shown to be an important regulator of behavioral inhibition (Crockett et al., 2009; Dayan and Huys, 2009) which has been postulated to be a core impairment in ADHD (Barkley, 1997). Pharmacological and genetic manipulation of rodent models implicates the 5-HT system in the hyperactive and impulsive dimensions of ADHD (Banerjee and Nandagopal, 2015). Children with ADHD have been reported to have lower levels of 5-HT in the blood (Coleman, 1971; Spivak et al., 1999), and reduced binding of tritiated imipramine (the non-selective 5-HT reuptake inhibitor) to blood platelets (Stoff et al., 1987). These findings led to the serotonergic hypothesis of ADHD which asserts that a reduction in available 5-HT in the synapse may contribute to clinical symptoms of ADHD (Quist and Kennedy, 2001). The serotonin system has been shown to be sensitive to exposure to inflammation with evidence from animal models (discussed in Section 5 below). Another angle by which researchers are understanding serotonin's role in ADHD is through the tryptophan synthesis pathway. These studies show that a reduction in serotonin via tryptophan depletion leads to inattention, reduced behavioral inhibition, and increased impulsivity (Banerjee and Nandagopal, 2015). Elevations in peripheral inflammation have been associated with increases in kynurenine metabolites (Pfaff et al., 2008) which competes

with serotonin for tryptophan resulting in less tryptophan available for 5-HT synthesis.

The serotonin hypothesis of ADHD is in alignment with the more well-known association of serotonin function with mood and anxiety disorders (Adler, 2007), which are strongly associated with reduced 5-HT levels (Coppen and Doogan, 1988; Eison, 1990; Sullivan et al., 2006) and often treated by medications that modulate serotonergic function. Gene by environment interplay involving the serotonin transporter is a fundamental finding in relation to mood disorders (Karg et al., 2011; Risch et al., 2009; Uher and McGuffin, 2008, 2010). However, ADHD is often comorbid with anxiety disorder, and children with ADHD are at a two-fold risk of future depression compared to children without ADHD (Meinzer et al., 2014). Further, emotional dysregulation is increasingly recognized as centrally important in ADHD (Shaw et al., 2014). In other words, it is a misconception that ADHD is purely a cognitive disorder. Thus, as ADHD is reconceptualized increasingly as involving broader aspects of cognitive and emotional dysregulation, the role of serotonin and the frequent diagnostic overlap with emotion-related conditions begins to be meaningful.

Neuroanatomical evidence is consistent with a role for the serotonergic system in ADHD. Serotonergic neurons project from the raphe nuclei throughout the cerebral cortex including robust projections to brain structures associated with ADHD such as the orbital frontal cortex (OFC). A reduction in serotonin levels in the OFC is associated with reduced emotional regulation, inhibition, and reversal learning all of which are correlates of ADHD (Clarke et al., 2007). The dorsomedial PFC is sensitive to reduced tryptophan availability (Evers et al., 2006). These studies suggest that the serotonergic system could be involved in ADHD by influencing orbitofrontal-striatal neurocircuitry.

Serotonin is involved in modulating the dopaminergic system. Serotonergic terminals can take up exogenous L-DOPA and convert it to dopamine (Stansley and Yamamoto, 2013). Also, several serotonin receptor subtypes (5-HT1BR, 5-HT1DR, and 5-HT6R) influence dopamine transmission in the mesolimbic pathway (Valentini et al., 2013; Yan and Yan, 2001). Lastly, 5-HT plays an important role in brain development which is distinct from its role in the mature brain. During development 5-HT is synthesized by the placenta from maternal derived tryptophan (Velasquez et al., 2013) making 5-HT an important link between the early environmental factors, inflammation and brain development. In summary, ample evidence indicates a potential role for 5-HT in ADHD. It is reasonable to propose that 5-HT modulation of the dopamine system and regulation of early brain development are mechanisms by which 5-HT influences risk of ADHD.

#### 4.2.3. Glutamate/GABA

Recent evidence supports a role for the neurotransmitters glutamate (Glu) and gamma-aminobutyric acid (GABA) in the etiology of ADHD. Glutamate is the main excitatory and GABA is the main inhibitory neurotransmitter throughout the brain. GABAergic and glutamatergic neurons project to many areas and structures within the central nervous system (CNS) (Morales and Root, 2014; Soiza-Reilly and Commons, 2011). Importantly for ADHD, glutamatergic and GABAergic neurons project to the striatum where they influence dopamine neurotransmission (Ferreira et al., 2009; Tritsch et al., 2012), suggesting that alterations to these systems could have an important role in ADHD (Miller et al., 2014).

Proton magnetic resonance spectroscopy (MRS) has been used in ADHD research to noninvasively obtain quantitative measurement of differences in regional brain biochemistry between ADHD patients and controls (Novotny et al., 1998). Increased levels of Glu have been detected in children with ADHD using MRS in the frontal (Moore et al., 2006), prefrontal (MacMaster et al., 2003), subcortical (Bollmann et al., 2015) and striatal (Carrey et al., 2007; MacMaster et al., 2003) brain regions as compared with controls. Although most of these differences did not reach significance in a recent meta-analysis (Perlov et al.,

2009). MRS studies in adult ADHD patients report both increases and decreases in Glu. Some studies observed increases in basal ganglia (Ferreira et al., 2009) and left cerebellar (Perlov et al., 2010) regions, while other studies, in contrast, reported decreased levels in the basal ganglia (Maltezos et al., 2014), right ACC (Perlov et al., 2007), and a left mid-frontal region (Dramsahl et al., 2011). A study done by Bollmann et al. (2015) showed differences in glutamate levels in ADHD children which normalizes with brain maturation in adult patients, suggesting a developmental timing effect in ADHD patients as they transition from adolescence to adulthood (Bollmann et al., 2015). The inconsistent reporting of glutamate levels in ADHD patients in the literature could be explained by that fact that glutamate is ubiquitous through-out the CNS with innervation to many regions and nuclei. Additional research is required to provide a more definitive hypothesis to the differences observed in glutamate/glutamine in ADHD patients.

Research on the GABAergic system's involvement in ADHD is relatively limited. GABA has been associated with impulsivity in men suggesting a link to ADHD (Boy et al., 2011). Studies indicate that GABA levels are reduced in ADHD children (Edden et al., 2012), with one study showing increased GABA levels in adult patients (Bollmann et al., 2015). However, these differences did not obtain significance in a meta-analysis comparing measurement of brain GABA levels across psychiatric disorders (Schur et al., 2016). A study utilizing an animal model of ADHD, the spontaneous hypertensive rat (SHR), observed reduced tonic levels of GABA in the hippocampus (Sterley et al., 2013). These studies similarly suggest that a reduction in GABA may be associated with childhood ADHD with a possible transition to increased levels when ADHD persists in adulthood. Additionally, genetic manipulation studies in mice found that knocking out the Gad67 enzyme, an enzyme involved in the synthesis of GABA, and the GABA transporter 1, resulted in mild hyperactivity as well as impaired attention (Chen et al., 2015; Smith, 2018). However, the limited available evidence precludes the ability to draw significant conclusions about the role of GABA in ADHD.

In summary, extensive literature provides support for highlighting both structural and functional changes in the brain associated with ADHD. Functionally, many of the neurotransmitter systems interact in complex ways that may converge on a common end point of aberrant dopamine transmission in ADHD relevant brain structures. In the next sections we will discuss how inflammation experienced during development may influence these structural and functional systems associated with ADHD pathology.

## 5. Animal models support link between developmental exposure to inflammation and changes in cognition and behavior

Evidence for the link between prenatal neuroinflammation and subsequent offspring behavioral changes is substantial. To investigate the association between exposure to inflammation during development and offspring behavior investigators induce inflammation during specific periods of development. The most common models induce maternal immune activation (MIA) by systemic (intraperitoneal (IP) or subcutaneous (SC)) administration of polyinosinic:polycytidylic acid (poly(I:C)) an agonist to the toll-like receptor-3 (TLR3) that models a viral infection, lipopolysaccharide (LPS), an agonist to the TLR4 that models a bacterial infection, bacteria such as streptococcus that commonly cause infection during pregnancy, or a combination of these agents to pregnant animals. These animal models of systemic prenatal infection induce changes in offspring behavior that may be reminiscent of aspects of human neurodevelopmental disorders, including behaviors interpreted as anxiety-like behavior (Arsenault et al., 2014; Makinson et al., 2017; Penteado et al., 2014; W. Schaafsma et al., 2017), sustained attention (Vuillermot et al., 2012), cognitive flexibility (Bitanirwe et al., 2010), and social behavior (Bitanirwe et al., 2010; Machado et al., 2015; Malkova et al., 2012; Smith et al., 2007) establish the proof of concept for maternal inflammation influencing offspring behavior

via alterations in neural development. Below we will examine the evidence linking prenatal inflammation to models of attention, activity level, and impulse control.

### 5.1. Evidence from animal models that exposure to prenatal inflammation leads to behavioral changes modeling aspects of ADHD

Deficits in certain kinds of attention are relevant to ADHD and other neurological disorders associated with dopamine deficiency (Del Campo et al., 2011). However, isolation of different attention systems in animal models are limited, so the relationship between prenatal maternal immune activation and offspring inattention has only been examined in a limited number of studies. One group used a signaled probability sustained attention task to examine cognitive and decision-making processes in adult rats exposed to a single dose of poly(I:C) during fetal development, but failed to detect an effect of MIA on any of the cognitive processes assessed (Bates et al., 2018). Two studies by the same group measured attentional shifting by examining the persistence of latent inhibition in a conditioned freezing paradigm. The first study noted abnormalities in attentional shifting in adult male, but not female, mice treated prenatally with a high dose of poly(I:C) (Bitanirwe et al., 2010). These findings were followed up by the examination of the interaction between prenatal immune activation and deficiency in *Nurr1* a transcription factor essential for the development of dopamine neurons which is also implicated in ADHD (Vuillermot et al., 2012). This study noted impairments in both sustained attention (using the two-choice discrimination test) and attentional shifting in adult male mice that were exposed to prenatal immune activation and had genetic *Nurr1* deficiency (Vuillermot et al., 2012).

Activity level is much more readily simulated in animal models, although animal models to date have not effectively modeled the social cost associated with hyperactivity in human children with ADHD. Also, the setting of the measurement (i.e. home cage versus behavioral testing apparatus) and the novelty of the situation as well as the choice of animal model can greatly influence the results. In children with ADHD hyperactivity is somewhat inhibited in novel situations (Sagvolden and Sergeant, 1998; Sleaford and Ullman, 1981). Thus, it would be ideal if an animal model of ADHD displayed hyperactivity in their home cage or after acclimation to the testing environment, but not necessarily during novel or stressful testing paradigms. A very limited number of studies have assessed locomotor activity in the home cage in rodent offspring exposed prenatally to inflammation. One study reports an increase in locomotor behavior in the home cage as measured by telemetry in young male mouse offspring exposed to poly(I:C) during gestation (Missig et al., 2018). Likewise, a second study observed increased locomotor behaviors in their home cage as documented via coding of videography in male mice offspring exposed to the influenza A virus during gestation (Miller et al., 2013). However, another study which used a photobeam system to measure activity of rat offspring exposed to poly(I:C) observed no group difference in locomotion in the home cage (Missault et al., 2014) and a fourth study observed a reduction in home cage activity in young adult mouse offspring exposed prenatally to LPS as measured by passive infrared monitoring (W. Schaafsma et al., 2017). Several studies report increased locomotor activity in the open field or other novel testing environments such as the elevated plus maze in offspring exposed to inflammation during gestation. These results appear to be gender dimorphic, and context and age dependent.

A study that induced gestational inflammation via prenatal exposure to Group B Streptococcus noted increased total distance travelled in male, but not female offspring at P20 in the open field (Allard et al., 2017), but that female, and not male offspring exhibited increased total distance travelled at P105-110 in an elevated plus maze (Allard et al., 2018). Offspring exposed to LPS during prenatal development were more active in the open field test at 20 months of age (Golan et al., 2006). In the study examining the interaction between

prenatal immune activation and genetic *Nurr1* deficiency, described above, prenatal immune activation and *Nurr1* deficiency exerted additive effects on spontaneous locomotor hyperactivity in the open field test (Vuillermot et al., 2012). Lastly, male, but not female, mouse offspring exposed to inflammation induced by prenatal stress exhibited locomotor hyperactivity in a novel, stressful environment. This behavioral phenotype was ameliorated by treatment of the mother with non-steroidal anti-inflammatory drugs and was associated with alterations in D1 and D2 receptors (Bronson and Bale, 2014).

In summary, these animal studies in rodents suggest that male offspring locomotor activity appears to be particularly responsive to maternal immune activation. However, the current findings are not entirely consistent; only two of the studies that report hyperactivity in maternal immune activation models actually measured locomotor activity in the home cage, which is potentially the most valid measure of changes in activity expected in an animal model of ADHD.

Impulsivity is another key feature of ADHD which has several animal model analogues although each model has notable limitations. Two studies report alterations in an impulsivity in offspring exposed to prenatal inflammation. One study noted that adult female, but not male, rats exposed to group B streptococcus during gestation displayed increased locomotion and entries in the open arms of an elevated plus maze potentially indicating decreased inhibition (Allard et al., 2018). Also, social intrusiveness is a diagnostic feature associated with impulsivity in ADHD (APA, 2013) and a second study noted that young adult female rats exposed in-utero to group B streptococcus displayed hyper-social behavior during late puberty, whereas males were hypo-social compared to same-sex controls (Bergeron et al., 2013).

ADHD is also associated with cognitive problems such as impaired working memory and reduced executive function (Willcutt et al., 2012). A number of animal models of prenatal exposure to inflammation report impairments in learning and memory. Young adult mice prenatally exposed to LPS were slower to learn a T-maze but had similar memory retention for the test as controls (W. Schaafsma et al., 2017). Prenatal LPS treatment impaired neurogenesis and performance in the novel object recognition test in adult rat offspring (Graciarena et al., 2010), learning to associate a cue with the platform in the Morris-water maze (Golan et al., 2005), and learning to avoid a foot shock in the Passive avoidance test (Golan et al., 2005). Gestational LPS exposure was associated with impaired spatial learning and memory in the radial arm water maze primarily in female mice (Li et al., 2016). In murine models, prenatal poly(I:C) exposure impaired working memory in a matching-to-position paradigm in the dry maze (Vuillermot et al., 2012), spatial recognition memory in the Y maze (Giovannoli et al., 2015), performance in the novel object recognition task (Ozawa et al., 2006), and rate of route-based learning when visible cues were unavailable in the Cincinnati water maze (Vorhees et al., 2012).

In summary, rodent models provide evidence of developmental exposure to inflammation being associated with behavioral changes; some of these may be related to features of ADHD (Fig. 1) such as behaviors interpreted as reflecting inattention (Bitanirwe et al., 2010; Vuillermot et al., 2012), motor activity (Miller et al., 2013; Missig et al., 2018), impulsivity (Allard et al., 2018; Bergeron et al., 2013), and impaired learning and memory (Golan et al., 2005; Graciarena et al., 2010; Li et al., 2016; W. Schaafsma et al., 2017; Vuillermot et al., 2012). However, inconsistent behavioral findings across studies remain a limitation, likely due to variations in the gestational age of maternal immune activation, the type of inflammatory event, and doses of the inflammatory inducing agent across studies. Future studies that characterize locomotion in the home cage, reinforcer discounting, unwarmed alerting (arousal), and inattention and impulsivity in maternal immune activation studies will be helpful.

## 5.2. Evidence from animal models that prenatal exposure to inflammation results in changes in brain structure consistent with ADHD

With advancements in brain imaging techniques it is now possible to directly relate changes in brain structure and development associated with neurodevelopmental disorders in humans to animal models. A number of cross-sectional and a few longitudinal MRI studies in rodent models of maternal immune activation provide evidence for subtle, but long-lasting, abnormalities in brain structure consistent with the neuroanatomical changes noted in children with ADHD (Fig. 1). Decreased overall brain volume was reported with prenatal exposure to poly(I:C) in adult mice (da Silveira et al., 2017). Similar to the reduction in gray matter volume documented in children with ADHD (Castellanos et al., 2002), a reduction in cortical gray matter was noted in rhesus monkeys whose mothers were infected with influenza during pregnancy (Short et al., 2010). The most marked reductions in gray matter were observed bilaterally in the cingulate and parietal areas, and white matter was noted to be reduced in the parietal lobe (Short et al., 2010). Prenatal exposure to inflammation has been documented to result in reductions in volume of cortical areas associated with ADHD. Prenatal exposure to poly(I:C) (Piontkewitz et al., 2012) reduced the volume of the prefrontal cortex, consistent with observations in children with ADHD (Arnsten and Rubia, 2012), and reduced the volume of the hippocampus in adult mice. Exposure to polyriboinosinic-polyribocytidylic acid (POL) during gestation decreased the volume of the anterior cingulate cortex, hippocampus, amygdala, striatum, nucleus accumbens and lateral ventricles in rat offspring (Crum et al., 2017). In contrast, prenatal POL exposure resulted in larger volume in the thalamus, ventral mesencephalon, brain stem and major white matter tracts relative to controls (Crum et al., 2017). To date no studies have identified alterations in subcortical structures such as the basal ganglia (Hart et al., 2013; Nakao et al., 2011) associated with ADHD pathology in rodent models of maternal immune activation. It is important to note that the developmental timing of the maternal immune challenge induces distinct changes in the offspring brain and few studies investigate multiple challenge time points in a single study.

## 5.3. Exposure to maternal immune activation during gestation alters the function of neurotransmitter systems associated with ADHD

### 5.3.1. Dopamine

The dopaminergic system is known to be sensitive to inflammation during development as animal studies utilizing a model of maternal immune activation show aberrant dopamine transmission through reduced spontaneous firing rate and population activity, as well as reduced dopamine transport and receptors in both the PFC and subcortical regions (Baharnoori et al., 2013; Luchicchi et al., 2016). Additional evidence is observed in a study performed by Straley et al. (2017), where dopaminergic-driven behaviors are altered in rat offspring. However, the behavioral results were not associated with a loss in dopaminergic neurons suggesting inflammation is influencing behavior by functional changes to the dopamine system during development (Straley et al., 2017). More evidence for dopaminergic system sensitivity to inflammation is observed in a model of maternal stress-induced inflammation that produced behaviors related to ADHD. Researchers observed that administration of maternal non-steroidal anti-inflammatory drugs resulted in amelioration of behaviors associated with dopamine dysregulation (Bronson and Bale, 2014). This study also observed altered expression of the D1 and D2 dopamine transporters in male offspring suggesting a functional change in dopamine transmission rather than a loss of neurons. In a model of maternal inflammation induced via poly(I:C) injection reduced expression of sonic hedgehog protein was observed (Khalil et al., 2013). As sonic hedgehog protein is very important in the maturation of dopaminergic neurons (Wang et al., 1995), a reduction in this protein could lead to an underdeveloped dopamine system. Other neurotransmitters important in the

neurobiology of ADHD including serotonin and glutamate have also been shown to be sensitive to the inflammatory environment (discussed below).

### 5.3.2. Serotonin

Animal models provide direct evidence that inflammatory factors influence the development of the 5-HT system. Much of the evidence comes from mouse models of prenatal exposure to inflammation. Overall, these studies indicate that exposure to inflammation during fetal development reduce neural 5-HT levels. In mouse offspring, LPS exposure decreased overall cerebral levels of 5-HT, down regulated mRNA expression of TPH2, and 5-HTT in whole brain homogenates (Hsueh et al., 2017), and decreased the number and size of TPH2 positive neurons in the dorsal raphe (Hsueh et al., 2017). In murine models, exposure to LPS (Depino, 2015) and poly(I:C) (Winter et al., 2009) decreased 5-HT levels in the hippocampus. In contrast, one study found that prenatal poly(I:C) exposure increased 5-HT levels in the hippocampus (Abazyan et al., 2010). The levels of 5-HT and its metabolite 5-hydroxyindoleacetic acid (5-HIAA) were reduced in the nucleus accumbens and lateral globus pallidus by prenatal poly(I:C) exposure (Winter et al., 2009) and prenatal flu exposure decreased 5-HT and 5-HIAA levels in the cerebellum (Fatemi et al., 2008; Winter et al., 2008). Prenatal exposure to the flu in a mouse model also reduced serum 5-HT levels, particularly in the male offspring, and increased 5-HT metabolism in female offspring (Miller et al., 2013). Interestingly, the vast majority of studies report a decrease in 5-HT synthesis and neuron number with exposure to prenatal inflammation regardless of the developmental timing of the immune challenge (Meyer and Feldon, 2009).

The 5-HT system plays a critical role in brain development, including neurogenesis, neuronal migration and synaptogenesis (Daws and Gould, 2011; Kannan et al., 2011). Thus, alterations in 5-HT synthesis are likely to contribute to impairments in overall brain development. One mechanism by which inflammation reduces 5-HT synthesis is by influencing the availability of its precursor tryptophan through the kynurenine pathway. An increase in kynurenine metabolites has been reported in animal models of maternal inflammation (Pfaff et al., 2008; Zavitsanou et al., 2014). As the kynurenine pathway competes with 5-HT for tryptophan, an increase in this pathway results in less tryptophan available for 5-HT synthesis. Epigenetic mechanisms are also reported to contribute to the reduction in 5-HT synthesis as prenatal exposure to poly(I:C) alters histone acetylation of the 5-HTT promoter in the hippocampus of adult mice (Reisinger et al., 2016).

### 5.3.3. Glutamate/GABA

While there is inconsistency in the degree and direction of change within the glutamatergic and GABAergic systems in ADHD patients, evidence suggests that alterations in these systems occur in the development of ADHD. Animal studies that model maternal inflammation indicate that proper development of the glutamatergic system is sensitive to inflammatory factors. Inflammation during development alters glutamatergic and GABAergic receptor expression and function in the offspring. Increased expression of GABA<sub>A</sub> receptors in the hippocampus and amygdala as well as decreased expression of the metabotropic glutamate receptor, mGlu2, in the frontal cortex is observed in mouse offspring born to either stressed or poly(I:C) injected mothers during pregnancy (Holloway et al., 2013; Nyffeler et al., 2006). Another study found that the ionotropic glutamate receptors NR2A, NR1, and NMDAR are altered in a sex dependent manner. Male offspring displayed increased NMDAR and NR2A binding in the striatum and cortex, which was accompanied by increased mRNA expression for NR2A and NR1 receptors (Rahman et al., 2017). In addition to neuronal changes within functionality of the glutamatergic neurons, glial cells play a crucial role in maintaining and regulating this system. Maternal inflammation disrupts normal glial functioning leading to dysfunctional regulation of glutamate transmission. Multiple studies using different species of

animals document an activation of microglia and astrocytes in response to maternal inflammation that leads to dysregulation in glutamate homeostasis (Roumier et al., 2008; Weaver-Mikaere et al., 2013; Zhang et al., 2016). Finally, evidence suggests that inflammation affects the glutamatergic system through epigenetic means. In a study by Tang et al. (2013), offspring born to mothers injected with LPS or poly(I:C) during pregnancy, showed decreases in promotor-specific histone acetylation and corresponding gene expression (Tang et al., 2013).

## 6. Conclusion

Substantial indirect and circumstantial evidence supports a hypothesis that neuroinflammation plays an important albeit non-specific role in the pathophysiology of ADHD. Promising new technologies and tools such as positron emission tomography (PET) using a radiotracer specific for activated microglia (Boerwinkle and Ances, 2018) will allow future studies to more directly assess neuroinflammation in patients with ADHD in a noninvasive manner. Alterations in the peripheral inflammatory profile are observed in several neurodevelopmental disorders (Kim et al., 2016; Landaas et al., 2010; Monji et al., 2013; Vargas et al., 2005), including ADHD, but this non-specificity is not surprising in light of increasing evidence that the psychiatric nosology does not carve nature at its joints but rather reflects multiple overlapping dimensions and syndromes which may have overlapping etiologic pathways.

ADHD appears to be associated with increased serum cytokines and comorbidity with atopic immune disorders. Additional mechanistic evidence is gathered from observations of the behavioral and neural outcomes of animal models of MIA (Fig. 1). These studies noted abnormalities in behavior consistent with ADHD such as hyperactivity, impulsivity, and inattention. Along with these behavioral changes prenatal exposure to inflammation is associated with changes in brain structures consistent with those observed in imaging studies of patients with ADHD. Animal models of prenatal exposure to inflammation observed a reduction in cortical gray matter volume and reductions in the volume of cortical areas associated with ADHD including the PFC and anterior cingulate cortex. In addition to structural changes, alterations in neurotransmitter systems including the dopaminergic, serotonergic, glutamatergic and GABAergic systems are observed in the ADHD population. Animal models of MIA provide strong evidence that the development and function of these neurotransmitters systems are sensitive to exposure to in utero inflammation. Prenatal exposure to inflammation produced changes in receptors, enzymes, and homeostatic levels of related neurotransmitters.

While ADHD is functionally characterized by a reduction in available dopamine in the synapse, dysfunction of other neurotransmitter systems may modulate dopamine system function. Serotonin, glutamate, and GABA modulate dopamine transmission and are hypothesized here to be important as well. If one or all of these systems are altered, it could lead to aberrant dopamine transmission. Evidence from animal models of maternal immune activation are intriguing but preliminary due to inconsistencies across experiments. One important question concerns the timing of the maternal immune activation during development. The developmental timing of the maternal immune challenge induces distinct changes in the offspring brain and behavior and few studies investigate multiple challenge time points in a single study.

This review presents accumulating evidence for a possible role of neuroinflammation in the progression of ADHD. Specifically, we focus on the impacts of exposure to inflammation during in-utero development. However, exposure to environmental factors, such as maternal smoking and pollutant exposure, that trigger inflammation during the early postnatal period and childhood also influence risk for neuropsychiatric disorders including ADHD. It is noteworthy that in addition to ADHD a number of neuropsychiatric disorders including ASD (Vargas et al., 2005), schizophrenia (Monji et al., 2013), depression

(Kim et al., 2016), anxiety, and bipolar disorder (Landaas et al., 2010) have also been associated with exposure to an increased peripheral and/or central inflammatory response during perinatal development. This elevated inflammatory response is thought to be triggered by a number of environmental factors during perinatal development. The environmental insults associated with ADHD appear to be similarly shared in predicting other neuropsychiatric disorders. In the case of ADHD, the majority of the evidence pertains to peripheral inflammation due to an absence of post-mortem studies. It is interesting that for schizophrenia and ASD both post-mortem studies (Bayer et al., 1999; De Picker et al., 2017; Vargas et al., 2005) and neuroimaging studies using PET and a radiotracer for microglia (Doorduyn et al., 2009; Suzuki et al., 2013) provide direct evidence for persistent neuroinflammation in some subjects compared to controls. However, this evidence is still preliminary as most of these studies have a small sample size making them prone to bias and confounding factors.

We therefore propose that these perinatal insults are acting through a common inflammatory pathway. That is consistent with the artifact of comorbidity among neuropsychiatric disorders (notably, here, ADHD and mood and emotion disorders (Adler, 2007)). In addition to the need for prospective and confirmatory studies of several kinds, several issues need more exploration. First, how does the developmental timing, duration and degree of the in utero inflammatory insult influence certain brain areas and neurotransmitter systems, and thus the behavioral phenotype. Second, how do genetic and environmental risk factors interact or synergize to mediate the influence on brain development and the risk for each specific neuropsychiatric disorder or its sub-domains (e.g., negative valence, controlled attention)? Third, how do sex-specific phenotypes of microglia and sex-differences in the timing of microglial development seen in animals (Hanamsagar et al., 2018; Villa et al., 2018) relate to sex differences in neurodevelopmental conditions like ADHD?

Overall, understanding how inflammation fits in to ADHD pathophysiology holds promise for an exciting new formulation of the disorder. If we can clarify how these processes are functioning in the case of a common and pervasive condition like ADHD, new avenues will be opened for novel therapeutic interventions.

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## References

- Abazyan, B., Nomura, J., Kannan, G., Ishizuka, K., Tamashiro, K.L., Nucifora, F., Pogorelov, V., Ladenheim, B., Yang, C., Krasnova, I.N., Cadet, J.L., Pardo, C., Mori, S., Kamiya, A., Vogel, M.W., Sawa, A., Ross, C.A., Pletnikov, M.V., 2010. Prenatal interaction of mutant DISC1 and immune activation produces adult psychopathology. *Biol. Psychiatry* 68 (12), 1172–1181.
- Adler, B., 2007. Managing ADHD in children, adolescents, and adults with comorbid anxiety in primary care. *Prim Care Companion J Clin Psychiatry* 9 (2), 129–138.
- Ahmed, R., Borst, J.M., Yong, C.W., Aslani, P., 2014. Do parents of children with attention-deficit/hyperactivity disorder (ADHD) receive adequate information about the disorder and its treatments? A qualitative investigation. *Patient Preference and Adherence* 8, 661–670.
- Allard, M.J., Bergeron, J.D., Bahamoori, M., Srivastava, L.K., Fortier, L.C., Poyart, C., Sebire, G., 2017. A sexually dichotomous, autistic-like phenotype is induced by Group B Streptococcus maternofetal immune activation. *Autism Res.* 10 (2), 233–245.
- Allard, M.J., Brochu, M.E., Bergeron, J.D., Sebire, G., 2018. Hyperactive behavior in female rats in utero-exposed to group B Streptococcus-induced inflammation. *Int. J. Dev. Neurosci.* 69, 17–22.
- Amico, F., Stauber, J., Koutsouleris, N., Frodl, T., 2011. Anterior cingulate cortex gray matter abnormalities in adults with attention deficit hyperactivity disorder: a voxel-based morphometry study. *Psychiatry Res.* 191 (1), 31–35.
- Anand, D., Colpo, G.D., Zeni, G., Zeni, C.P., Teixeira, A.L., 2017. Attention-deficit/hyperactivity disorder and inflammation: what does current knowledge tell us? *A*

- systematic review. *Front Psychiatry* 8, 228.
- Andersen, C.H., Thomsen, P.H., Nohr, E.A., Lemcke, S., 2018. Maternal body mass index before pregnancy as a risk factor for ADHD and autism in children. *Eur Child Adolesc Psychiatry* 27 (2), 139–148.
- Anttila, V., Bulik-Sullivan, B., Finucane, H.K., Walters, R.K., Bras, J., Duncan, L., Escott-Price, V., Falcone, G.J., Gormley, P., Malik, R., Patsopoulos, N.A., Ripke, S., Wei, Z., Yu, D., Lee, P.H., Turley, P., Grenier-Boley, B., Chouraki, V., Kamatani, Y., Berr, C., Letenneur, L., Hannequin, D., Amouyel, P., Boland, A., Deleuze, J.F., Duron, E., Vardarajan, B.N., Reitz, C., Goate, A.M., Huentelman, M.J., Kamboh, M.I., Larson, E. B., Rogaeva, E., St George-Hyslop, P., Hakonarson, H., Kukull, W.A., Farrer, L.A., Barnes, L.L., Beach, T.G., Demirci, F.Y., Head, E., Hulette, C.M., Jicha, G.A., Kauwe, J. S.K., Kaye, J.A., Leverenz, J.B., Levey, A.I., Lieberman, A.P., Pankratz, V.S., Poon, W. W., Quinn, J.F., Saykin, A.J., Schneider, L.S., Smith, A.G., Sonnen, J.A., Stern, R.A., Van Derlin, V.M., Van Eldik, L.J., Harold, D., Russo, G., Rubinsztein, D.C., Bayer, A., Tzolaki, M., Proitsi, P., Fox, N.C., Hampel, H., Owen, M.J., Mead, S., Passmore, P., Morgan, K., Nöthen, M.M., Rossor, M., Lupton, M.K., Hoffmann, P., Kornhuber, J., Lawlor, B., McQuillin, A., Al-Chalabi, A., Bis, J.C., Ruiz, A., Boada, M., Seshadri, S., Beiser, A., Rice, K., van der Lee, S.J., De Jager, P.L., Geschwind, D.H., Riemenschneider, M., Riedel-Heller, S., Rotter, J.L., Ransmayr, G., Hyman, B.T., Cruchaga, C., Aegret, M., Winsvold, B., Palta, P., Farh, K.H., Cuenca-Leon, E., Furlotte, N., Kurth, T., Ligthart, L., Terwindt, G.M., Freilinger, T., Ran, C., Gordon, S. D., Borck, G., Adams, H.H.H., Lehtimäki, T., Wedenoja, J., Buring, J.E., Schürks, M., Hrafnóttir, M., Hottenga, J.J., Penninx, B., Artto, V., Kaunisto, M., Vepsäläinen, S., Martin, N.G., Montgomery, G.W., Kurki, M.I., Hämäläinen, E., Huang, H., Huang, J., Sandor, C., Webber, C., Müller-Miyhsok, B., Schreiber, S., Salomaa, V., Loehrer, E., Göbel, H., Macaya, A., Pozo-Rosich, P., Hansen, T., Werge, T., Kaprio, J., Metspalu, A., Kubisch, C., Ferrari, M.D., Belin, A.C., van den Maagdenberg, A.M.J.M., Zwart, J. A., Boomsma, D., Eriksson, N., Olesen, J., Chasman, D.I., Nyholt, D.R., Avbersek, A., Baum, L., Berkovic, S., Bradfield, J., Buono, R., Catarino, C.B., Cossette, P., De Jonghe, P., Depondt, C., Dlugos, D., Ferraro, T.N., French, J., Hjalgrim, H., Jamnadas-Khoda, J., Kälviäinen, R., Kunz, W.S., Lerche, H., Leu, C., Lindhout, D., Lo, W., Lowenstein, D., McCormack, M., Möller, R.S., Molloy, A., Ng, P.W., Oliver, K., Privitera, M., Radtke, R., Ruppert, A.K., Sander, T., Schachter, S., Schankin, C., Scheffer, I., Schoch, S., Sisodiya, S.M., Smith, P., Sperling, M., Striano, P., Surges, R., Thomas, G.N., Visscher, F., Whelan, C.D., Zera, F., Heinzen, E.L., Marson, A., Becker, F., Stroink, H., Zimprich, F., Gasser, T., Gibbs, R., Heutink, P., Martinez, M., Morris, H.R., Sharma, M., Rytén, M., Mok, K.Y., Pulit, S., Bevan, S., Holliday, E., Attia, J., Battey, T., Boncoraglio, G., Thijs, V., Chen, W.M., Mitchell, B., Rothwell, P., Sharma, P., Sudlow, C., Vicente, A., Markus, H., Kourkoulis, C., Pera, J., Raffeld, M., Silliman, S., Boraska Perica, V., Thornton, L.M., Huckins, L.M., William Rayner, N., Lewis, C. M., Gratacos, M., Rybakowski, F., Keski-Rahkonen, A., Raevuori, A., Hudson, J.I., Reichborn-Kjennerud, T., Monteleone, P., Karwautz, A., Mannik, K., Baker, J.H., O'Toole, J.K., Trace, S.E., Davis, O.S.P., Helder, S.G., Ehrlich, S., Herpertz-Dahlmann, B., Dannen, U.N., van Elburg, A.A., Clementi, M., Forzan, M., Docampo, E., Lissowska, J., Hauser, J., Tortorella, A., Maj, M., Gonidakis, F., Tziouvas, K., Papezova, H., Yilmaz, Z., Wagner, G., Cohen-Woods, S., Herms, S., Julià, A., Rabioner, R., Dick, D. M., Ripatti, S., Andreassen, O.A., Espeseth, T., Lundervold, A.J., Steen, V.M., Pinto, D., Scherer, S.W., Aschauer, H., Schosser, A., Alfredsson, L., Padyukov, L., Halmi, K. A., Mitchell, J., Strober, M., Bergen, A.W., Kaye, W., Szatkiewicz, J.P., Cormand, B., Ramos-Quiroga, J.A., Sánchez-Mora, C., Ribasés, M., Casas, M., Hervás, A., Arranz, M.J., Haavik, J., Zayats, T., Johansson, S., Williams, N., Dempfle, A., Rothenberger, A., Kuntsi, J., Oades, R.D., Banaschewski, T., Franke, B., Buitelaar, J.K., Arias Vasquez, A., Doyle, A.E., Reif, A., Lesch, K.P., Freitag, C., Rivero, O., Palmason, H., Romanos, M., Langley, K., Rietschel, M., Witt, S.H., Dalsgaard, S., Børglum, A.D., Waldman, I., Wilmot, B., Molly, N., Bau, C.H.D., Crosbie, J., Schachar, R., Loo, S.K., McGough, J.J., Grevet, E.H., Medland, S.E., Robinson, E., Weiss, L.A., Bacchelli, E., Bailey, A., Bal, V., Battaglia, A., Betancur, C., Bolton, P., Cantor, R., Celestino-Soper, P., Dawson, G., De Rubeis, S., Duque, F., Green, A., Klauk, S.M., Leboyer, M., Levitt, P., Maestrini, E., Mane, S., DeLuca, D.M., Parr, J., Regan, R., Reichenberg, A., Sandin, S., Vorstman, J., Wassink, T., Wijsman, E., Cook, E., Santangelo, S., Delorme, R., Rogé, B., Magalhaes, T., Arking, D., Schulze, T.G., Thompson, R.C., Strohmaier, J., Matthews, K., Melle, I., Morris, D., Blackwood, D., McIntosh, A., Bergen, S.E., Schalling, M., Jamain, S., Maaser, A., Fischer, S.B., Reinbold, C.S., Fullerton, J.M., Guzman-Parra, J., Mayoral, F., Schofield, P.R., Cichon, S., Mühleisen, T.W., Degenhardt, F., Schumacher, J., Bauer, M., Mitchell, P.B., Gershon, E.S., Rice, J., Potash, J.B., Zandi, P.P., Craddock, N., Ferrier, I.N., Alda, M., Rouleau, G.A., Turecki, G., Ophoff, R., Pato, C., Anjorian, A., Stahl, E., Leber, M., Czerski, P.M., Cruceanu, C., Jones, I.R., Posthuma, D., Andlauer, T.F.M., Forstner, A.J., Streit, F., Baune, B.T., Air, T., Sinnamón, G., Wray, N.R., MacIntyre, D.J., Porteous, D., Homuth, G., Rivera, M., Grove, J., Middeldorp, C.M., Hickie, I., Pergadia, M., Mehta, D., Smit, J.H., Jansen, R., de Geus, E., Dunn, E., Li, Q.S., Nauck, M., Schoevers, R.A., Beekman, A.T., Knowles, J.A., Viktorin, A., Arnold, P., Barr, C.L., Bedoya-Berrio, G., Bienvenu, O.J., Brentani, H., Burton, C., Camarena, B., Capi, C., Cath, D., Cavallini, M., Cusi, D., Darrow, S., Denys, D., Derks, E.M., Dietrich, A., Fernandez, T., Figeo, M., Freimer, N., Gerber, G., Grados, M., Greenberg, E., Hanna, G.L., Hartmann, A., Hirschtritt, M.E., Hoekstra, P.J., Huang, A., Huyser, C., Illmann, C., Jenike, M., Kuperman, S., Lenthal, B., Lochner, C., Lyon, G.J., Macciardi, F., Madruga-Garrido, M., Malaty, I. A., Maras, A., McGrath, L., Miguel, E.C., Mir, P., Nestadt, G., Nicolini, H., Okun, M.S., Pakstis, A., Paschou, P., Piacentini, J., Pittenger, C., Plessen, K., Ramensky, V., Ramos, E.M., Reus, V., Richter, M.A., Riddle, M.A., Robertson, M.M., Roessner, V., Rosario, M., Samuels, J.F., Sandor, P., Stein, D.J., Tsetos, F., Van Nieuwerburgh, F., Weatherall, S., Wendland, J.R., Wolanczyk, T., Worbe, Y., Zai, G., Goes, F.S., McLaughlin, N., Nestadt, P.S., Grabe, H.J., Depienne, C., Konkashbaev, A., Lanzagorta, N., Valencia-Duarte, A., Bramon, E., Buccola, N., Cahn, W., Cairns, M., Chong, S.A., Cohen, D., Crespo-Facorro, B., Crowley, J., Davidson, M., DeLisi, L., Dinan, T., Donohoe, G., Drapeau, E., Duan, J., Haan, L., Hougaard, D., Karachanak
- Yankova, S., Khrunin, A., Klovins, J., Kučinskās, V., Lee Chee Keong, J., Limborska, S., Loughland, C., Lönnqvist, J., Maher, B., Mattheisen, M., McDonald, C., Murphy, K. C., Nenadic, I., van Os, J., Pantelis, C., Pato, M., Petryshen, T., Quedsted, D., Roussos, P., Sanders, A.R., Schall, U., Schwab, S.G., Sim, K., So, H.C., Stögmann, E., Subramaniam, M., Toncheva, D., Waddington, J., Walters, J., Weiser, M., Cheng, W., Cloninger, R., Curtis, D., Gejman, P.V., Henskens, F., Mattingsdal, M., Oh, S.Y., Scott, R., Webb, B., Breen, G., Churchhouse, C., Bulik, C.M., Daly, M., Dichgans, M., Faraone, S.V., Guerreiro, R., Holmans, P., Kendler, K.S., Koeleman, B., Mathews, C.A., Price, A., Scharf, J., Sklar, P., Williams, J., Wood, N.W., Cotsapas, C., Palotie, A., Smoller, J.W., Sullivan, P., Rosand, J., Corvin, A., Neale, B.M., Schott, J.M., Anney, R., Elia, J., Grigoriou-Serbanescu, M., Edenberg, H.J., Murray, R., Consortium, B., 2018. Analysis of shared heritability in common disorders of the brain. *Science* 360(6395).
- Aoki, Y., Yoncheva, Y.N., Chen, B., Nath, T., Sharp, D., Lazar, M., Velasco, P., Milham, M.P., Di Martino, A., 2017. Association of white matter structure with autism spectrum disorder and attention-deficit/hyperactivity disorder. *JAMA Psychiatry* 74 (11), 1120–1128.
- APA, 2013. *Diagnostic and Statistical Manual of Mental Disorders*, 5th ed. American Psychiatric Publishing, Arlington, VA.
- Arnold, L.E., Hurt, E., Lofthouse, N., 2013. Attention-deficit/hyperactivity disorder: dietary and nutritional treatments. *Child Adolesc. Psychiatr. Clin. N. Am.* 22 (3), 381–402.
- Arnsten, A.F., Rubia, K., 2012. Neurobiological circuits regulating attention, cognitive control, motivation, and emotion: disruptions in neurodevelopmental psychiatric disorders. *J. Am. Acad. Child Adolesc. Psychiatry* 51 (4), 356–367.
- Arsenault, D., St-Amour, I., Cisbani, G., Rousseau, L.S., Cicchetti, F., 2014. The different effects of LPS and poly I:C prenatal immune challenges on the behavior, development and inflammatory responses in pregnant mice and their offspring. *Brain Behav. Immun.* 38, 77–90.
- Ayala, R., Shu, T., Tsai, L.H., 2007. Trekking across the brain: the journey of neuronal migration. *Cell* 128 (1), 29–43.
- Baharooni, M., Bhardwaj, S.K., Srivastava, L.K., 2013. Effect of maternal lipopolysaccharide administration on the development of dopaminergic receptors and transporter in the rat offspring. *PLoS One* 8 (1), e54439.
- Banerjee, E., Nandagopal, K., 2015. Does serotonin deficit mediate susceptibility to ADHD? *Neurochem. Int.* 82, 56–68.
- Barkley, R.A., 1997. Behavioral inhibition, sustained attention, and executive functions: constructing a unifying theory of ADHD. *Psychol. Bull.* 121 (1), 65–94.
- Barry, D.S., Pakan, J.M., McDermott, K.W., 2014. Radial glial cells: key organisers in CNS development. *Int. J. Biochem. Cell Biol.* 46, 76–79.
- Bates, V., Maharjan, A., Millar, J., Bilkey, D.K., Ward, R.D., 2018. Spared motivational modulation of cognitive effort in a maternal immune activation model of schizophrenia risk. *Behav. Neurosci.* 132 (1), 66–74.
- Bayer, T.A., Buslei, R., Havas, L., Falkai, P., 1999. Evidence for activation of microglia in patients with psychiatric illnesses. *Neurosci. Lett.* 271 (2), 126–128.
- Becker, K., El-Faddagh, M., Schmidt, M.H., Esser, G., Laucht, M., 2008. Interaction of dopamine transporter genotype with prenatal smoke exposure on ADHD symptoms. *J. Pediatr.* 152 (2), 263–269.
- Belmadani, A., Tran, P.B., Ren, D., Miller, R.J., 2006. Chemokines regulate the migration of neural progenitors to sites of neuroinflammation. *J. Neurosci.* 26 (12), 3182–3191.
- Bergeron, J.D., Deslauriers, J., Grignon, S., Fortier, L.C., Lepage, M., Stroth, T., Poyart, C., Sebire, G., 2013. White matter injury and autistic-like behavior predominantly affecting male rat offspring exposed to group B streptococcal maternal inflammation. *Dev. Neurosci.* 35 (6), 504–515.
- Bhattacharya, A., Derecki, N.C., Lovenberg, T.W., Drevets, W.C., 2016. Role of neuro-immunological factors in the pathophysiology of mood disorders. *Psychopharmacology* 233 (9), 1623–1636.
- Biederman, J., 2005. Attention-deficit/hyperactivity disorder: a selective overview. *Biol. Psychiatry* 57 (11), 1215–1220.
- Bilbo, S.D., Schwarz, J.M., 2009. Early-life programming of later-life brain and behavior: a critical role for the immune system. *Front. Behav. Neurosci.* 3, 14.
- Bitanihirwe, B.K., Peleg-Raibstein, D., Mouttet, F., Feldon, J., Meyer, U., 2010. Late prenatal immune activation in mice leads to behavioral and neurochemical abnormalities relevant to the negative symptoms of schizophrenia. *Neuropsychopharmacology* 35 (12), 2462–2478.
- Boerwinkle, A., Ances, B.M., 2018. Molecular imaging of neuroinflammation in HIV. *J. NeuroImmune Pharmacol.* 14 (1), 9–15.
- Bollmann, S., Ghisleni, C., Poil, S.S., Martin, E., Ball, J., Eich-Hochli, D., Edden, R.A., Klaver, P., Michels, L., Brandeis, D., O'Gorman, R.L., 2015. Developmental changes in gamma-aminobutyric acid levels in attention-deficit/hyperactivity disorder. *Transl. Psychiatry* 5, e589.
- Boy, F., Evans, C.J., Edden, R.A., Lawrence, A.D., Singh, K.D., Husain, M., Sumner, P., 2011. Dorsolateral prefrontal gamma-aminobutyric acid in men predicts individual differences in rash impulsivity. *Biol. Psychiatry* 70 (9), 866–872.
- Bronson, S.L., Bale, T.L., 2014. Prenatal stress-induced increases in placental inflammation and offspring hyperactivity are male-specific and ameliorated by maternal anti-inflammatory treatment. *Endocrinology* 155 (7), 2635–2646.
- Buske-Kirschbaum, A., Schmitt, J., Plessow, F., Romanos, M., Weidinger, S., Roessner, V., 2013. Psychoendocrine and psychoneuroimmunological mechanisms in the comorbidity of atopic eczema and attention deficit/hyperactivity disorder. *Psychoneuroendocrinology* 38 (1), 12–23.
- Buss, C., Entringer, S., Davis, E.P., Hobel, C.J., Swanson, J.M., Wadhwa, P.D., Sandman, C.A., 2012. Impaired executive function mediates the association between maternal pre-pregnancy body mass index and child ADHD symptoms. *PLoS One* 7 (6), e37758.
- Carrey, N.J., MacMaster, F.P., Gaudet, L., Schmidt, M.H., 2007. Striatal creatine and glutamate/glutamine in attention-deficit/hyperactivity disorder. *J Child Adolesc*

- Psychopharmacol 17 (1), 11–17.
- Castellanos, F.X., Lee, P.P., Sharp, W., Jeffries, N.O., Greenstein, D.K., Clasen, L.S., Blumenthal, J.D., James, R.S., Ebens, C.L., Walter, J.M., Zijdenbos, A., Evans, A.C., Giedd, J.N., Rapoport, J.L., 2002. Developmental trajectories of brain volume abnormalities in children and adolescents with attention-deficit/hyperactivity disorder. *JAMA* 288 (14), 1740–1748.
- Chang, J.P., Su, K.P., Mondelli, V., Pariante, C.M., 2018. Omega-3 polyunsaturated fatty acids in youths with attention deficit hyperactivity disorder: a systematic review and meta-analysis of clinical trials and biological studies. *Neuropsychopharmacology* 43 (3), 534–545.
- Chen, Q., Sjolander, A., Langstrom, N., Rodriguez, A., Serlachius, E., D'Onofrio, B.M., Lichtenstein, P., Larsson, H., 2014. Maternal pre-pregnancy body mass index and offspring attention deficit hyperactivity disorder: a population-based cohort study using a sibling-comparison design. *Int. J. Epidemiol.* 43 (1), 83–90.
- Chen, L., Yang, X., Zhou, X., Wang, C., Gong, X., Chen, B., Chen, Y., 2015. Hyperactivity and impaired attention in Gamma aminobutyric acid transporter subtype 1 gene knockout mice. *Acta Neuropsychiatr* 27 (6), 368–374.
- Clarke, H.F., Walker, S.C., Dalley, J.W., Robbins, T.W., Roberts, A.C., 2007. Cognitive inflexibility after prefrontal serotonin depletion is behaviorally and neurochemically specific. *Cereb. Cortex* 17 (1), 18–27.
- Coleman, M., 1971. Serotonin concentrations in whole blood of hyperactive children. *J. Pediatr.* 78 (6), 985–990.
- Colombo, J., Gustafson, K.M., Gajewski, B.J., Shaddy, D.J., Kerling, E.H., Thodosoff, J.M., Doty, T., Brez, C.C., Carlson, S.E., 2016. Prenatal DHA supplementation and infant attention. *Pediatr. Res.* 80 (5), 656–662.
- Coppen, A.J., Doogan, D.P., 1988. Serotonin and its place in the pathogenesis of depression. *J. Clin Psychiatry* 49, 4–11 Suppl.
- Cortese, S., 2012. The neurobiology and genetics of attention-deficit/hyperactivity disorder (ADHD): what every clinician should know. *European Journal of Paediatric Neurology: EJPN: Official Journal of the European Paediatric Neurology Society* 16 (5), 422–433.
- Costenbader, K.H., Karlson, E.W., 2006. Cigarette smoking and autoimmune disease: what can we learn from epidemiology? *Lupus* 15 (11), 737–745.
- Crockett, M.J., Clark, L., Robbins, T.W., 2009. Reconciling the role of serotonin in behavioral inhibition and aversion: acute tryptophan depletion abolishes punishment-induced inhibition in humans. *J. Neurosci.* 29 (38), 11993–11999.
- Crum, W.R., Sawiak, S.J., Chege, W., Cooper, J.D., Williams, S.C.R., Vernon, A.C., 2017. Evolution of structural abnormalities in the rat brain following in utero exposure to maternal immune activation: a longitudinal in vivo MRI study. *Brain Behav. Immun.* 63, 50–59.
- Cunningham, C.L., Martinez-Cerdeno, V., Noctor, S.C., 2013. Microglia regulate the number of neural precursor cells in the developing cerebral cortex. *J. Neurosci.* 33 (10), 4216–4233.
- da Silveira, V.T., Medeiros, D.C., Ropke, J., Guidine, P.A., Rezende, G.H., Moraes, M.F., Mendes, E.M., Macedo, D., Moreira, F.A., de Oliveira, A.C., 2017. Effects of early or late prenatal immune activation in mice on behavioral and neuroanatomical abnormalities relevant to schizophrenia in the adulthood. *Int. J. Dev. Neurosci.* 58, 1–8.
- Dan, O., Raz, S., 2012. The relationships among ADHD, self-esteem, and test anxiety in young adults. *J. Atten. Disord.* 19 (3), 231–239.
- Danielson, M.L., Bitsko, R.H., Ghandour, R.M., Holbrook, J.R., Kogan, M.D., Blumberg, S.J., 2018. Prevalence of parent-reported ADHD diagnosis and associated treatment among U.S. children and adolescents, 2016. *J. Clin Child Adolesc Psychol* 47 (2), 199–212.
- Darwish, A.H., Elgohary, T.M., Nosair, N.A., 2018. Serum interleukin-6 level in children with attention-deficit hyperactivity disorder (ADHD). *J. Child Neurol.* 883073818809831.
- Daws, L.C., Gould, G.G., 2011. Ontogeny and regulation of the serotonin transporter: providing insights into human disorders. *Pharmacol. Ther.* 131 (1), 61–79.
- Dayan, P., Huys, Q.J., 2009. Serotonin in affective control. *Annu. Rev. Neurosci.* 32, 95–126.
- De Picker, L.J., Morrens, M., Chance, S.A., Boche, D., 2017. Microglia and brain plasticity in acute psychosis and schizophrenia illness course: a meta-review. *Front Psychiatry* 8, 238.
- de Wit, S., Watson, P., Harsay, H.A., Cohen, M.X., van de Vijver, I., Ridderinkhof, K.R., 2012. Corticostriatal connectivity underlies individual differences in the balance between habitual and goal-directed action control. *J. Neurosci.* 32 (35), 12066–12075.
- Del Campo, N., Chamberlain, S.R., Sahakian, B.J., Robbins, T.W., 2011. The roles of dopamine and noradrenaline in the pathophysiology and treatment of attention-deficit/hyperactivity disorder. *Biol. Psychiatry* 69 (12), e145–e157.
- Depino, A.M., 2015. Early prenatal exposure to LPS results in anxiety- and depression-related behaviors in adulthood. *Neuroscience* 299, 56–65.
- Donfrancesco, R., Nativio, P., Borrelli, E., Guia, E., Andriola, E., Villa, M.P., M, D.I.T., 2016a. Serum cytokines in paediatric neuropsychiatric syndromes: focus on Attention Deficit Hyperactivity Disorder. *Minerva Pediatr.*
- Donfrancesco, R., Nativio, P., Di Benedetto, A., Villa, M.P., Andriola, E., Melegari, M.G., Cipriano, E., Di Trani, M., 2016b. Anti-Yo antibodies in children with ADHD: first results about serum cytokines. *J. Atten. Disord pii: 1087054716643387*. [Epub ahead of print].
- Doorduyn, J., de Vries, E.F., Willemsen, A.T., de Groot, J.C., Dierckx, R.A., Klein, H.C., 2009. Neuroinflammation in schizophrenia-related psychosis: a PET study. *J. Nucl. Med.* 50 (11), 1801–1807.
- Doshi, J.A., Hodgkins, P., Kahle, J., Sikirica, V., Cangelosi, M.J., Setyawan, J., Erder, M.H., Neumann, P.J., 2012. Economic impact of childhood and adult attention-deficit/hyperactivity disorder in the United States. *J. Am. Acad. Child Adolesc. Psychiatry* 51 (10), 990–1002 (e1002).
- Dougherty, C.C., Evans, D.W., Myers, S.M., Moore, G.J., Michael, A.M., 2016. A comparison of structural brain imaging findings in autism spectrum disorder and attention-deficit hyperactivity disorder. *Neuropsychol. Rev.* 26 (1), 25–43.
- Dramsahl, M., Erland, L., Plessen, K.J., Haavik, J., Hugdahl, K., Specht, K., 2011. Adults with attention-deficit/hyperactivity disorder - a brain magnetic resonance spectroscopy study. *Front Psychiatry* 2, 65.
- Edden, R.A., Crocetti, D., Zhu, H., Gilbert, D.L., Mostofsky, S.H., 2012. Reduced GABA concentration in attention-deficit/hyperactivity disorder. *Arch. Gen. Psychiatry* 69 (7), 750–753.
- Eison, M.S., 1990. Serotonin: a common neurobiologic substrate in anxiety and depression. *J. Clin. Psychopharmacol.* 10 (3 Suppl), 26S–30S.
- Elmore, A.L., Nigg, J.T., Friderici, K.H., Jernigan, K., Nikolas, M.A., 2016. Does 5HTTLPR genotype moderate the association of family environment with child attention-deficit hyperactivity disorder symptomatology? *J. Clin Child Adolesc Psychol* 45 (3), 348–360.
- Evers, E.A., van der Veen, F.M., van Deursen, J.A., Schmitt, J.A., Deutz, N.E., Jolles, J., 2006. The effect of acute tryptophan depletion on the BOLD response during performance monitoring and response inhibition in healthy male volunteers. *Psychopharmacology* 187 (2), 200–208.
- Eyo, U.B., Dailey, M.E., 2013. Microglia: key elements in neural development, plasticity, and pathology. *J. NeuroImmune Pharmacol.* 8 (3), 494–509.
- Faraone, S.V., Larsson, H., 2019. Genetics of attention deficit hyperactivity disorder. *Mol. Psychiatry* 24 (4), 562–575.
- Faraone, S.V., Mick, E., 2010. Molecular genetics of attention deficit hyperactivity disorder. *Psychiatr Clin North Am* 33 (1), 159–180.
- Fasmer, O.B., Halmoy, A., Egan, T.M., Oedegaard, K.J., Haavik, J., 2011. Adult attention deficit hyperactivity disorder is associated with asthma. *BMC Psychiatry* 11, 128.
- Fatemi, S.H., Reutiman, T.J., Folsom, T.D., Huang, H., Oishi, K., Mori, S., Smees, D.F., Pearce, D.A., Winter, C., Sohr, R., Juckel, G., 2008. Maternal infection leads to abnormal gene regulation and brain atrophy in mouse offspring: implications for genesis of neurodevelopmental disorders. *Schizophr. Res.* 99 (1–3), 56–70.
- Ferreira, P.E., Palmieri, A., Bau, C.H., Grevet, E.H., Hoefel, J.R., Rohde, L.A., Anes, M., Ferreira, E.E., Belmonte-de-Abreu, P., 2009. Differentiating attention-deficit/hyperactivity disorder inattentive and combined types: a (1)H-magnetic resonance spectroscopy study of fronto-striato-thalamic regions. *J. Neural Transm. (Vienna)* 116 (5), 623–629.
- Giana, G., Romano, E., Porfirio, M.C., D'Ambrosio, R., Giovinnazzo, S., Troianiello, M., Barlocchi, E., Travaglini, D., Granstrem, O., Pascale, E., Tarani, L., Curatolo, P., Laviola, G., Adriani, W., 2015. Detection of auto-antibodies to DAT in the serum: interactions with DAT genotype and psycho-stimulant therapy for ADHD. *J. Neuroimmunol.* 278, 212–222.
- Giedd, J.N., Blumenthal, J., Jeffries, N.O., Castellanos, F.X., Liu, H., Zijdenbos, A., Paus, T., Evans, A.C., Rapoport, J.L., 1999. Brain development during childhood and adolescence: a longitudinal MRI study. *Brain Neurosci.* 2 (10), 861–863.
- Gilmore, J.H., Shi, F., Woolson, S.L., Knickmeyer, R.C., Short, S.J., Lin, W., Zhu, H., Hamer, R.M., Styner, M., Shen, D., 2012. Longitudinal development of cortical and subcortical gray matter from birth to 2 years. *Cereb. Cortex* 22 (11), 2478–2485.
- Giovanoli, S., Notter, T., Richetto, J., Labouesse, M.A., Vuilleumot, S., Riva, M.A., Meyer, U., 2015. Late prenatal immune activation causes hippocampal deficits in the absence of persistent inflammation across aging. *J. Neuroinflammation* 12, 221.
- Golan, H.M., Lev, V., Hallak, M., Sorokin, Y., Huleihel, M., 2005. Specific neurodevelopmental damage in mice offspring following maternal inflammation during pregnancy. *Neuropharmacology* 48 (6), 903–917.
- Golan, H., Stilman, M., Lev, V., Huleihel, M., 2006. Normal aging of offspring mice of mothers with induced inflammation during pregnancy. *Neuroscience* 141 (4), 1909–1918.
- Grabli, D., McCairn, K., Hirsch, E.C., Agid, Y., Feger, J., Francois, C., Tremblay, L., 2004. Behavioural disorders induced by external globus pallidus dysfunction in primates: I. Behavioural study. *Brain* 127, 2039–2054 Pt 9.
- Graciarena, M., Depino, A.M., Pitossi, F.J., 2010. Prenatal inflammation impairs adult neurogenesis and memory related behavior through persistent hippocampal TGFbeta1 downregulation. *Brain Behav. Immun.* 24 (8), 1301–1309.
- Hanamsagar, R., Alter, M.D., Block, C.S., Sullivan, H., Bolton, J.L., Bilbo, S.D., 2018. Generation of a microglial developmental index in mice and in humans reveals a sex difference in maturation and immune reactivity. *Glia* 66 (2), 460.
- Hart, H., Radua, J., Nakao, T., Mataix-Cols, D., Rubia, K., 2013. Meta-analysis of functional magnetic resonance imaging studies of inhibition and attention in attention-deficit/hyperactivity disorder: exploring task-specific, stimulant medication, and age effects. *JAMA Psychiatry* 70 (2), 185–198.
- Hassan, W., Noreen, H., Castro-Gomes, V., Mohammadzai, I., da Rocha, J.B., Landeira-Fernandez, J., 2016. Association of oxidative stress with psychiatric disorders. *Curr. Pharm. Des.* 22 (20), 2960–2974.
- Hawkey, E., Nigg, J.T., 2014. Omega-3 fatty acid and ADHD: blood level analysis and meta-analytic extension of supplementation trials. *Clin. Psychol. Rev.* 34 (6), 496–505.
- Holloway, T., Moreno, J.L., Umali, A., Rayannavar, V., Hodes, G.E., Russo, S.J., Gonzalez-Maeso, J., 2013. Prenatal stress induces schizophrenia-like alterations of serotonin 2A and metabotropic glutamate 2 receptors in the adult offspring: role of maternal immune system. *J. Neurosci.* 33 (3), 1088–1098.
- Hoogman, M., Bralten, J., Hibar, D.P., Mennes, M., Zwiers, M.P., Schweren, L.S.J., van Hulzen, K.J.E., Medland, S.E., Shumskaya, E., Jahanshad, N., Zeeuw, P., Szekely, E., Sudre, G., Wolfers, T., Onnink, A.M.H., Dammers, J.T., Mostert, J.C., Vives-Gilabert, Y., Kohls, G., Oberwilleand, E., Seitz, J., Schulte-Rüther, M., Ambrosino, S., Doyle, A.E., Hovik, M.F., Dramsahl, M., Tamm, L., van Erp, T.G.M., Dale, A., Schork, A., Conzelmann, A., Zierhut, K., Baur, R., McCarthy, H., Yoncheva, Y.N., Cubillo, A., Chantiluke, K., Mehta, M.A., Paloyelis, Y., Hohmann, S., Baumeister, S., Bramati, I.,

- Mattos, P., Tovar-Moll, F., Douglas, P., Banaschewski, T., Brandeis, D., Kuntsi, J., Asherson, P., Rubia, K., Kelly, C., Martino, A.D., Milham, M.P., Castellanos, F.X., Frodl, T., Zentis, M., Lesch, K.P., Reif, A., Pauli, P., Jernigan, T.L., Haavik, J., Plessen, K.J., Lundervold, A.J., Hugdahl, K., Seidman, L.J., Biederman, J., Rommelse, N., Helsenfeld, D.J., Hartman, C.A., Hoeksma, P.J., Oosterlaan, J., Polier, G.V., Konrad, K., Vilarroya, O., Ramos-Quiroga, J.A., Soliva, J.C., Durston, S., Buitelaar, J.K., Faraone, S.V., Shaw, P., Thompson, P.M., Franke, B., 2017. Subcortical brain volume differences in participants with attention deficit hyperactivity disorder in children and adults: a cross-sectional mega-analysis. *Lancet Psychiatry* 4 (4), 310–319.
- Hsueh, P.T., Wang, H.H., Liu, C.L., Ni, W.F., Chen, Y.L., Liu, J.K., 2017. Expression of cerebral serotonin related to anxiety-like behaviors in C57BL/6 offspring induced by repeated subcutaneous prenatal exposure to low-dose lipopolysaccharide. *PLoS One* 12 (6), e0179970.
- Instones, J.T., Halmoy, A., Engeland, A., Haavik, J., Furu, K., Klungsoyr, K., 2017. Attention-deficit/hyperactivity disorder in offspring of mothers with inflammatory and immune system diseases. *Biol. Psychiatry* 81 (5), 452–459.
- Kannan, S., Saadani-Makki, F., Balakrishnan, B., Dai, H., Chakraborty, P.K., Janisse, J., Muzik, O., Romero, R., Chugani, D.C., 2011. Decreased cortical serotonin in neonatal rabbits exposed to endotoxin in utero. *Journal of Cerebral Blood Flow and Metabolism: Official Journal of the International Society of Cerebral Blood Flow and Metabolism* 31 (2), 738–749.
- Karg, K., Burmeister, M., Shedden, K., Sen, S., 2011. The serotonin transporter promoter variant (5-HTTLPR), stress, and depression meta-analysis revisited: evidence of genetic moderation. *Arch. Gen. Psychiatry* 68 (5), 444–454.
- Khalil, O.S., Forrest, C.M., Pizar, M., Smith, R.A., Darlington, L.G., Stone, T.W., 2013. Prenatal activation of maternal TLR3 receptors by viral-mimetic poly(I:C) modifies GluN2B expression in embryos and sonic hedgehog in offspring in the absence of kynurenine pathway activation. *Immunopharmacol. Immunotoxicol.* 35 (5), 581–593.
- Kim, Y.S., Leventhal, B.L., 2015. Genetic epidemiology and insights into interactive genetic and environmental effects in autism spectrum disorders. *Biol. Psychiatry* 77 (1), 66–74.
- Kim, Y.K., Na, K.S., Myint, A.M., Leonard, B.E., 2016. The role of pro-inflammatory cytokines in neuroinflammation, neurogenesis and the neuroendocrine system in major depression. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 64, 277–284.
- Kronfol, Z., Remick, D.G., 2000. Cytokines and the brain: implications for clinical psychiatry. *Am. J. Psychiatry* 157 (5), 683–694.
- Landaas, E.T., Johansson, S., Jacobsen, K.K., Ribases, M., Bosch, R., Sanchez-Mora, C., Jacob, C.P., Boreatti-Hummer, A., Kreiker, S., Lesch, K.P., Kiemeny, L.A., Kooij, J.J., Kan, C., Buitelaar, J.K., Faraone, S.V., Halmoy, A., Ramos-Quiroga, J.A., Cormand, B., Reif, A., Franke, B., Mick, E., Knappskog, P.M., Haavik, J., 2010. An international multicenter association study of the serotonin transporter gene in persistent ADHD. *Genes Brain Behav* 9 (5), 449–458.
- Levy, F., 1991. The dopamine theory of attention deficit hyperactivity disorder (ADHD). *Aust N Z J Psychiatry* 25 (2), 277–283.
- Li, X., Cao, Q., Pu, F., Li, D., Fan, Y., An, L., Wang, P., Wu, Z., Sun, L., Li, S., Wang, Y., 2015. Abnormalities of structural covariance networks in drug-naïve boys with attention deficit hyperactivity disorder. *Psychiatry Res.* 231 (3), 273–278.
- Li, X.W., Cao, L., Wang, F., Yang, Q.G., Tong, J.J., Li, X.Y., Chen, G.H., 2016. Maternal inflammation linearly exacerbates offspring age-related changes of spatial learning and memory, and neurobiology until senescence. *Behav. Brain Res.* 306, 178–196.
- Liao, T.C., Lien, Y.T., Wang, S., Huang, S.L., Chen, C.Y., 2016. Comorbidity of atopic disorders with autism spectrum disorder and attention deficit/hyperactivity disorder. *J. Pediatr.* 171, 248–255.
- Lim, L., Chantiluke, K., Cubillo, A.I., Smith, A.B., Simmons, A., Mehta, M.A., Rubia, K., 2015. Disorder-specific grey matter deficits in attention deficit hyperactivity disorder relative to autism spectrum disorder. *Psychol. Med.* 45 (5), 965–976.
- Lin, Y.T., Chen, Y.C., Gau, S.S., Yeh, T.H., Fan, H.Y., Hwang, Y.Y., Lee, Y.L., 2016. Associations between allergic diseases and attention deficit hyperactivity/oppositional defiant disorders in children. *Pediatr. Res.* 80 (4), 480–485.
- Luchicchi, A., Lecca, S., Melis, M., De Felice, M., Cadeddu, F., Frau, R., Muntoni, A.L., Fadda, P., Devoto, P., Pistis, M., 2016. Maternal immune activation disrupts dopamine system in the offspring. *Int. J. Neuropsychopharmacol.* 19 (7).
- Machado, C.J., Whitaker, A.M., Smith, S.E., Patterson, P.H., Bauman, M.D., 2015. Maternal immune activation in nonhuman primates alters social attention in juvenile offspring. *Biol. Psychiatry* 77 (9), 823–832.
- MacMaster, F.P., Carrey, N., Sparkes, S., Kusumakar, V., 2003. Proton spectroscopy in medication-free pediatric attention-deficit/hyperactivity disorder. *Biol. Psychiatry* 53 (2), 184–187.
- Makinson, R., Lloyd, K., Rayasam, A., McKee, S., Brown, A., Barila, G., Grissom, N., George, R., Marini, M., Fabry, Z., Elovitz, M., Reyes, T.M., 2017. Intrauterine inflammation induces sex-specific effects on neuroinflammation, white matter, and behavior. *Brain Behav. Immun.* 66, 277–288.
- Makris, N., Biederman, J., Valera, E.M., Bush, G., Kaiser, J., Kennedy, D.N., Caviness, V.S., Faraone, S.V., Seidman, L.J., 2007. Cortical thinning of the attention and executive function networks in adults with attention-deficit/hyperactivity disorder. *Cereb. Cortex* 17 (6), 1364–1375.
- Malkova, N.V., Yu, C.Z., Hsiao, E.Y., Moore, M.J., Patterson, P.H., 2012. Maternal immune activation yields offspring displaying mouse versions of the three core symptoms of autism. *Brain Behav. Immun.* 26 (4), 607–616.
- Maltezos, S., Horder, J., Coghlan, S., Skirrow, C., O’Gorman, R., Lavender, T.J., Mendez, M.A., Mehta, M., Daly, E., Xenitidis, K., Paliokosta, E., Spain, D., Pitts, M., Asherson, P., Lythgoe, D.J., Barker, G.J., Murphy, D.G., 2014. Glutamate/glutamine and neuronal integrity in adults with ADHD: a proton MRS study. *Transl. Psychiatry* 4, e373.
- Marsh, R., Gerber, A.J., Peterson, B.S., 2008. Neuroimaging studies of normal brain development and their relevance for understanding childhood neuropsychiatric disorders. *J. Am. Acad. Child Adolesc. Psychiatry* 47 (11), 1233–1251.
- Martin, J., Cooper, M., Hamshere, M.L., Pocklington, A., Scherer, S.W., Kent, L., Gill, M., Owen, M.J., Williams, N., O’Donovan, M.C., Thapar, A., Holmans, P., 2014. Biological overlap of attention-deficit/hyperactivity disorder and autism spectrum disorder: evidence from copy number variants. *J. Am. Acad. Child Adolesc. Psychiatry* 53 (7), 761–770 (e726).
- Mattei, D., Ivanov, A., Ferrai, C., Jordan, P., Guneykaya, D., Buonfiglioli, A., Schaafsma, W., Przanowski, P., Deuther-Conrad, W., Brust, P., Hesse, S., Patt, M., Sabri, O., Ross, T.L., Eggen, B.J.L., Boddeke, E., Kaminska, B., Beule, D., Pombo, A., Kettenmann, H., Wolf, S.A., 2017. Maternal immune activation results in complex microglial transcriptome signature in the adult offspring that is reversed by minocycline treatment. *Transl. Psychiatry* 7 (5), e1120.
- Meinzer, M.C., Pettit, J.W., Viswesvaran, C., 2014. The co-occurrence of attention-deficit/hyperactivity disorder and unipolar depression in children and adolescents: a meta-analytic review. *Clin. Psychol. Rev.* 34 (8), 595–607.
- Metin, C., Vallee, R.B., Rakic, P., Bhide, P.G., 2008. Modes and mishaps of neuronal migration in the mammalian brain. *J. Neurosci.* 28 (46), 11746–11752.
- Meyer, U., Feldon, J., 2009. Neural basis of psychosis-related behaviour in the infection model of schizophrenia. *Behav. Brain Res.* 204 (2), 322–334.
- Mick, E., Biederman, J., Faraone, S.V., Sayer, J., Kleinman, S., 2002. Case-control study of attention-deficit hyperactivity disorder and maternal smoking, alcohol use, and drug use during pregnancy. *J. Am. Acad. Child Adolesc. Psychiatry* 41 (4), 378–385.
- Miller, V.M., Zhu, Y., Bucher, C., McGinnis, W., Ryan, L.K., Siegel, A., Zalcman, S., 2013. Gestational flu exposure induces changes in neurochemicals, affiliative hormones and brainstem inflammation, in addition to autism-like behaviors in mice. *Brain Behav. Immun.* 33, 153–163.
- Miller, E.M., Pomerleau, F., Huettl, P., Gerhardt, G.A., Glaser, P.E., 2014. Aberrant glutamate signaling in the prefrontal cortex and striatum of the spontaneously hypertensive rat model of attention-deficit/hyperactivity disorder. *Psychopharmacology* 231 (15), 3019–3029.
- Missault, S., Van den Eynde, K., Vanden Bergh, W., Franssen, E., Weeren, A., Timmermans, J.P., Kumar-Singh, S., Dedeurwaerdere, S., 2014. The risk for behavioural deficits is determined by the maternal immune response to prenatal immune challenge in a neurodevelopmental model. *Brain Behav. Immun.* 42, 138–146.
- Missig, G., Mokler, E.L., Robbins, J.O., Alexander, A.J., McDougle, C.J., Carlezon, W.A., Jr., 2018. Perinatal immune activation produces persistent sleep alterations and epileptiform activity in male mice. *Neuropsychopharmacology* 43(3), 482–491.
- Mittleman, B.B., Castellanos, F.X., Jacobsen, L.K., Rapoport, J.L., Swedo, S.E., Shearer, G.M., 1997. Cerebrospinal fluid cytokines in pediatric neuropsychiatric disease. *J. Immunol.* 159 (6), 2994–2999.
- Miyazaki, C., Koyama, M., Ota, E., Swa, T., Munde, L.B., Amiya, R.M., Tachibana, Y., Yamamoto-Hanada, K., Mori, R., 2017. Allergic diseases in children with attention deficit hyperactivity disorder: a systematic review and meta-analysis. *BMC Psychiatry* 17 (1), 120.
- Monji, A., Kato, T.A., Mizoguchi, Y., Horikawa, H., Seki, Y., Kasai, M., Yamauchi, Y., Yamada, S., Kanba, S., 2013. Neuroinflammation in schizophrenia especially focused on the role of microglia. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 42, 115–121.
- Moore, C.M., Biederman, J., Wozniak, J., Mick, E., Aleardi, M., Wardrop, M., Dougherty, M., Harpold, T., Hammerness, P., Randall, E., Renshaw, P.F., 2006. Differences in brain chemistry in children and adolescents with attention deficit hyperactivity disorder with and without comorbid bipolar disorder: a proton magnetic resonance spectroscopy study. *Am. J. Psychiatry* 163 (2), 316–318.
- Morales, M., Root, D.H., 2014. Glutamate neurons within the midbrain dopamine regions. *Neuroscience* 282, 60–68.
- Nakao, T., Radua, J., Rubia, K., Mataix-Cols, D., 2011. Gray matter volume abnormalities in ADHD: voxel-based meta-analysis exploring the effects of age and stimulant medication. *Am. J. Psychiatry* 168 (11), 1154–1163.
- Neuman, R.J., Lobos, E., Reich, W., Henderson, C.A., Sun, L.W., Todd, R.D., 2007. Prenatal smoking exposure and dopaminergic genotypes interact to cause a severe ADHD subtype. *Biol. Psychiatry* 61 (12), 1320–1328.
- Nigg, J.T., 2005. Neuropsychologic theory and findings in attention-deficit/hyperactivity disorder: the state of the field and salient challenges for the coming decade. *Biol. Psychiatry* 57 (11), 1424–1435.
- Nigg, J., Nikolas, M., Burt, S.A., 2010. Measured gene-by-environment interaction in relation to attention-deficit/hyperactivity disorder. *J. Am. Acad. Child Adolesc. Psychiatry* 49 (9), 863–873.
- Nigg, J.T., Elmore, A.L., Natarajan, N., Friderici, K.H., Nikolas, M.A., 2016. Variation in an iron metabolism gene moderates the association between blood lead levels and attention-deficit/hyperactivity disorder in children. *Psychol. Sci.* 27 (2), 257–269.
- Nigg, J.T., Gustafsson, H.C., Karalunas, S.L., Ryabinin, P., McWeeny, S.K., Faraone, S.V., Mooney, M.A., Fair, D.A., Wilmot, B., 2018. Working memory and vigilance as multivariate endophenotypes related to common genetic risk for attention-deficit/hyperactivity disorder. *J. Am. Acad. Child Adolesc. Psychiatry* 57 (3), 175–182.
- Novotny, E., Ashwal, S., Shevell, M., 1998. Proton magnetic resonance spectroscopy: an emerging technology in pediatric neurology research. *Pediatr. Res.* 44 (1), 1–10.
- Nyffeler, M., Meyer, U., Yee, B.K., Feldon, J., Knuesel, I., 2006. Maternal immune activation during pregnancy increases limbic GABA<sub>A</sub> receptor immunoreactivity in the adult offspring: implications for schizophrenia. *Neuroscience* 143 (1), 51–62.
- Oades, R.D., Dauvermann, M.R., Schimmelmann, B.G., Schwarz, M.J., Myint, A.M., 2010a. Attention-deficit hyperactivity disorder (ADHD) and glial integrity: S100B, cytokines and kynurenine metabolism—effects of medication. *Behav. Brain Funct.* 6, 29.
- Oades, R.D., Myint, A.M., Dauvermann, M.R., Schimmelmann, B.G., Schwarz, M.J., 2010b. Attention-deficit hyperactivity disorder (ADHD) and glial integrity: an exploration of associations of cytokines and kynurenine metabolites with symptoms and

- attention. *Behav. Brain Funct.* 6, 32.
- O'Shea, T.M., Joseph, R.M., Kuban, K.C., Allred, E.N., Ware, J., Coster, T., Fichorova, R.N., Dammann, O., Leviton, A., Investigators, E.S., 2014. Elevated blood levels of inflammation-related proteins are associated with an attention problem at age 24 mo in extremely preterm infants. *Pediatr. Res.* 75 (6), 781–787.
- Ozawa, K., Hashimoto, K., Kishimoto, T., Shimizu, E., Ishikura, H., Iyo, M., 2006. Immune activation during pregnancy in mice leads to dopaminergic hyperfunction and cognitive impairment in the offspring: a neurodevelopmental animal model of schizophrenia. *Biol. Psychiatry* 59 (6), 546–554.
- Park, P., Caballero, J., Omidian, H., 2014. Use of serotonin norepinephrine reuptake inhibitors in the treatment of attention-deficit hyperactivity disorder in pediatrics. *Ann. Pharmacother.* 48 (1), 86–92.
- Pennington, B.F., McGrath, L.M., Rosenberg, J., Barnard, H., Smith, S.D., Willcutt, E.G., Friend, A., Defries, J.C., Olson, R.K., 2009. Gene X environment interactions in reading disability and attention-deficit/hyperactivity disorder. *Dev. Psychol.* 45 (1), 77–89.
- Penteado, S.H., Teodorov, E., Kirsten, T.B., Eluf, B.P., Reis-Silva, T.M., Acenjo, M.K., de Melo, R.C., Suffredini, I.B., Bernardi, M.M., 2014. Prenatal lipopolysaccharide disrupts maternal behavior, reduces nest odor preference in pups, and induces anxiety: studies of F1 and F2 generations. *Eur. J. Pharmacol.* 738, 342–351.
- Perlov, E., Philippen, A., Hesslinger, B., Buechert, M., Ahrendts, J., Feige, B., Bubl, E., Hennig, J., Ebert, D., Tebartz van Elst, L., 2007. Reduced cingulate glutamate/glutamine-to-creatine ratios in adult patients with attention deficit/hyperactivity disorder — a magnet resonance spectroscopy study. *J. Psychiatr. Res.* 41 (11), 934–941.
- Perlov, E., Philippen, A., Matthies, S., Drieling, T., Maier, S., Bubl, E., Hesslinger, B., Buechert, M., Hennig, J., Ebert, D., Tebartz Van Elst, L., 2009. Spectroscopic findings in attention-deficit/hyperactivity disorder: review and meta-analysis. *World J Biol Psychiatry* 10 (4), 355–365 Pt 2.
- Perlov, E., Tebartz van Elst, L., Buechert, M., Maier, S., Matthies, S., Ebert, D., Hesslinger, B., Philippen, A., 2010. H(1)-MR-spectroscopy of cerebellum in adult attention deficit/hyperactivity disorder. *J. Psychiatr. Res.* 44 (14), 938–943.
- Pfaff, A.W., Mousli, M., Senegas, A., Marcellin, L., Takikawa, O., Klein, J.P., Candolfi, E., 2008. Impact of foetus and mother on IFN-gamma-induced indoleamine 2,3-dioxygenase and inducible nitric oxide synthase expression in murine placenta following *Toxoplasma gondii* infection. *Int. J. Parasitol.* 38 (2), 249–258.
- Pietropaolo, S., Crusio, W.E., Feldon, J., 2017. Gene-environment interactions in neurodevelopmental disorders. *Neural Plast* 2017, 9272804.
- Pinto, R.Q., Soares, I., Carvalho-Correia, E., Mesquita, A.R., 2015. Gene-environment interactions in psychopathology throughout early childhood: a systematic review. *Psychiatr. Genet.* 25 (6), 223–233.
- Piontkewitz, Y., Arad, M., Weiner, I., 2012. Tracing the development of psychosis and its prevention: what can be learned from animal models. *Neuropharmacology* 62 (3), 1273–1289.
- Quist, J.F., Kennedy, J.L., 2001. Genetics of childhood disorders: XXIII. ADHD, part 7: the serotonin system. *J. Am. Acad. Child Adolesc. Psychiatry* 40 (2), 253–256.
- Rahman, T., Zavitsanou, K., Purves-Tyson, T., Harms, L.R., Meehan, C., Schall, U., Todd, J., Hodgson, D.M., Michie, P.T., Weickert, C.S., 2017. Effects of immune activation during early or late gestation on N-methyl-D-aspartate receptor measures in adult rat offspring. *Front Psychiatry* 8, 77.
- Reisinger, S.N., Kong, E., Khan, D., Schulz, S., Ronovsky, M., Berger, S., Horvath, O., Cabatic, M., Berger, A., Pollak, D.D., 2016. Maternal immune activation epigenetically regulates hippocampal serotonin transporter levels. *Neurobiol Stress* 4, 34–43.
- Reus, G.Z., Fries, G.R., Stertz, L., Badawy, M., Passos, I.C., Barichello, T., Kapczynski, F., Quevedo, J., 2015. The role of inflammation and microglial activation in the pathophysiology of psychiatric disorders. *Neuroscience* 300, 141–154.
- Rijlaarsdam, J., Cecil, C.A., Walton, E., Mesirov, M.S., Relton, C.L., Gaunt, T.R., McArdle, W., Barker, E.D., 2017. Prenatal unhealthy diet, insulin-like growth factor 2 gene (IGF2) methylation, and attention deficit hyperactivity disorder symptoms in youth with early-onset conduct problems. *J. Child Psychol. Psychiatry* 58 (1), 19–27.
- Risch, N., Herrell, R., Lehner, T., Liang, K.Y., Eaves, L., Hoh, J., Griem, A., Kovacs, M., Ott, J., Merikangas, K.R., 2009. Interaction between the serotonin transporter gene (5-HTTLPR), stressful life events, and risk of depression: a meta-analysis. *JAMA* 301 (23), 2462–2471.
- Robbins, T.W., Sahakian, B.J., 1979. “Paradoxical” effects of psychomotor stimulant drugs in hyperactive children from the standpoint of behavioural pharmacology. *Neuropharmacology* 18 (12), 931–950.
- Rodriguez, A., 2010. Maternal pre-pregnancy obesity and risk for inattention and negative emotionality in children. *J. Child Psychol. Psychiatry* 51 (2), 134–143.
- Rodriguez, A., Miettunen, J., Henriksen, T.B., Olsen, J., Obel, C., Taanila, A., Ebeling, H., Linnet, K.M., Moilanen, I., Jarvelin, M.R., 2008. Maternal adiposity prior to pregnancy is associated with ADHD symptoms in offspring: evidence from three prospective pregnancy cohorts. *Int. J. Obes.* 32 (3), 550–557.
- Rommelse, N., Buitelaar, J.K., Hartman, C.A., 2017. Structural brain imaging correlates of ASD and ADHD across the lifespan: a hypothesis-generating review on developmental ASD-ADHD subtypes. *J. Neural Transm. (Vienna)* 124 (2), 259–271.
- Roumier, A., Pascual, O., Bechade, C., Wakselman, S., Poncer, J.C., Real, E., Triller, A., Bessis, A., 2008. Prenatal activation of microglia induces delayed impairment of glutamatergic synaptic function. *PLoS One* 3 (7), e2595.
- Sagvolden, T., Sergeant, J.A., 1998. Attention deficit/hyperactivity disorder—from brain dysfunctions to behaviour. *Behav. Brain Res.* 94 (1), 1–10.
- Salter, M.W., Beggs, S., 2014. Sublime microglia: expanding roles for the guardians of the CNS. *Cell* 158 (1), 15–24.
- Sanchez, C.E., Barry, C., Sabhlok, A., Russell, K., Majors, A., Kollins, S.H., Fuemmeler, B.F., 2018. Maternal pre-pregnancy obesity and child neurodevelopmental outcomes: a meta-analysis. *Obes. Rev.* 19 (4), 464–484.
- Schaafsma, W., Basterra, L.B., Jacobs, S., Brouwer, N., Meerlo, P., Schaafsma, A., Boddeke, E., Eggen, B.J.L., 2017a. Maternal inflammation induces immune activation of fetal microglia and leads to disrupted microglia immune responses, behavior, and learning performance in adulthood. *Neurobiol. Dis.* 106, 291–300.
- Schaafsma, S.M., Gagnidze, K., Reyes, A., Norstedt, N., Mansson, K., Francis, K., Pfaff, D.W., 2017b. Sex-specific gene-environment interactions underlying ASD-like behaviors. *Proc. Natl. Acad. Sci. U. S. A.* 114 (6), 1383–1388.
- Schans, J.V., Çiçek, R., de Vries, T.W., Hak, E., Hoekstra, P.J., 2017. Association of atopic diseases and attention-deficit/hyperactivity disorder: a systematic review and meta-analyses. *Neurosci Biobehav Rev* 74(Pt A) 139–148.
- Schmitt, J., Romanos, M., Schmitt, N.M., Meurer, M., Kirch, W., 2009. Atopic eczema and attention-deficit/hyperactivity disorder in a population-based sample of children and adolescents. *JAMA* 301 (7), 724–726.
- Schmitt, J., Buske-Kirschbaum, A., Roessner, V., 2010. Is atopic disease a risk factor for attention-deficit/hyperactivity disorder? A systematic review. *Allergy* 65 (12), 1506–1524.
- Schur, R.R., Draisma, L.W., Wijnen, J.P., Boks, M.P., Koevoets, M.G., Joels, M., Klomp, D.W., Kahn, R.S., Vinkers, C.H., 2016. Brain GABA levels across psychiatric disorders: a systematic literature review and meta-analysis of (1) H-MRS studies. *Hum. Brain Mapp.* 37 (9), 3337–3352.
- Segman, R.H., Meltzer, A., Gross-Tsur, V., Kosov, A., Frisch, A., Inbar, E., Darvasi, A., Levy, S., Goltser, T., Weizman, A., Galili-Weisstub, E., 2002. Preferential transmission of interleukin-1 receptor antagonist alleles in attention deficit hyperactivity disorder. *Mol. Psychiatry* 7 (1), 72–74.
- Sen, S., Duman, R., Sanacora, G., 2008. Serum brain-derived neurotrophic factor, depression, and antidepressant medications: meta-analyses and implications. *Biol. Psychiatry* 64 (6), 527–532.
- Shankar, K., Zhong, Y., Kang, P., Lau, F., Blackburn, M.L., Chen, J.R., Borengasser, S.J., Ronis, M.J., Badger, T.M., 2011. Maternal obesity promotes a proinflammatory signature in rat uterus and blastocyst. *Endocrinology* 152 (11), 4158–4170.
- Shaw, P., Eckstrand, K., Sharp, W., Blumenthal, J., Lerch, J.P., Greenstein, D., Clasen, L., Evans, A., Giedd, J., Rapoport, J.L., 2007. Attention-deficit/hyperactivity disorder is characterized by a delay in cortical maturation. *Proc. Natl. Acad. Sci. U. S. A.* 104 (49), 19649–19654.
- Shaw, P., Stringaris, A., Nigg, J., Leibenluft, E., 2014. Emotion dysregulation in attention deficit hyperactivity disorder. *Am. J. Psychiatry* 171 (3), 276–293.
- Short, S.J., Lubach, G.R., Karasin, A.I., Olsen, C.W., Styner, M., Knickmeyer, R.C., Gilmore, J.H., Coe, C.L., 2010. Maternal influenza infection during pregnancy impacts postnatal brain development in the rhesus monkey. *Biol. Psychiatry* 67 (10), 965–973.
- Silva, D., Colvin, L., Hagemann, E., Bower, C., 2014. Environmental risk factors by gender associated with attention-deficit/hyperactivity disorder. *Pediatrics* 133 (1), e14–e22.
- Sleator, E.K., Ullman, R.K., 1981. Can a physician diagnose hyperactivity in the office? *Pediatrics* 67, 13–17.
- Smith, K.M., 2018. Hyperactivity in mice lacking one allele of the glutamic acid decarboxylase 67 gene. *Atten Defic Hyperact Disord* 10 (4), 267–271.
- Smith, S.E., Li, J., Garbett, K., Mirnics, K., Patterson, P.H., 2007. Maternal immune activation alters fetal brain development through interleukin-6. *J. Neurosci.* 27 (40), 10695–10702.
- Soiza-Reilly, M., Commons, K.G., 2011. Glutamatergic drive of the dorsal raphe nucleus. *J. Chem. Neuroanat.* 41 (4), 247–255.
- Spivak, B., Vered, Y., Yoran-Hegesh, R., Averbuch, E., Mester, R., Graf, E., Weizman, A., 1999. Circulatory levels of catecholamines, serotonin and lipids in attention deficit hyperactivity disorder. *Acta Psychiatr. Scand.* 99 (4), 300–304.
- Stanley, B.J., Yamamoto, B.K., 2013. L-dopa-induced dopamine synthesis and oxidative stress in serotonergic cells. *Neuropharmacology* 67, 243–251.
- Sterley, T.L., Howells, F.M., Russell, V.A., 2013. Evidence for reduced tonic levels of GABA in the hippocampus of an animal model of ADHD, the spontaneously hypertensive rat. *Brain Res.* 1541, 52–60.
- Stevenson, J., Sonuga-Barke, E., McCann, D., Grimshaw, K., Parker, K.M., Rose-Zerilli, M.J., Holloway, J.W., Warner, J.O., 2010. The role of histamine degradation gene polymorphisms in moderating the effects of food additives on children's ADHD symptoms. *Am. J. Psychiatry* 167 (9), 1108–1115.
- Stoff, D.M., Pollock, L., Vitiello, B., Behar, D., Bridger, W.H., 1987. Reduction of (3H)-imipramine binding sites on platelets of conduct-disordered children. *Neuropsychopharmacology* 1 (1), 55–62.
- Stolp, H.B., Dziegielewska, K.M., 2009. Review: role of developmental inflammation and blood-brain barrier dysfunction in neurodevelopmental and neurodegenerative diseases. *Neuropathol. Appl. Neurobiol.* 35 (2), 132–146.
- Stoodley, C.J., 2014. Distinct regions of the cerebellum show gray matter decreases in autism, ADHD, and developmental dyslexia. *Front. Syst. Neurosci.* 8, 92.
- Stoodley, C.J., 2016. The cerebellum and neurodevelopmental disorders. *Cerebellum* 15 (1), 34–37.
- Straley, M.E., Van Oeffelen, W., Theze, S., Sullivan, A.M., O'Mahony, S.M., Cryan, J.F., O'Keefe, G.W., 2017. Distinct alterations in motor & reward seeking behavior are dependent on the gestational age of exposure to LPS-induced maternal immune activation. *Brain Behav. Immun.* 63, 21–34.
- Sullivan, G.M., Mann, J.J., Oquendo, M.A., Lo, E.S., Cooper, T.B., Gorman, J.M., 2006. Low cerebrospinal fluid transthyretin levels in depression: correlations with suicidal ideation and low serotonin function. *Biol. Psychiatry* 60 (5), 500–506.
- Suzuki, K., Sugihara, G., Ouchi, Y., Nakamura, K., Futatsubashi, M., Takebayashi, K., Yoshihara, Y., Omata, K., Matsumoto, K., Tsuchiya, K.J., Iwata, Y., Tsujii, M., Sugiyama, T., Mori, N., 2013. Microglial activation in young adults with autism spectrum disorder. *JAMA Psychiatry* 70 (1), 49–58.
- Tang, B., Jia, H., Kast, R.J., Thomas, E.A., 2013. Epigenetic changes at gene promoters in response to immune activation in utero. *Brain Behav. Immun.* 30, 168–175.
- Terasaki, L.S., Schwarz, J.M., 2016. Effects of moderate prenatal alcohol exposure during

- early gestation in rats on inflammation across the maternal-fetal-immune interface and later-life immune function in the offspring. *J. NeuroImmune Pharmacol.* 11 (4), 680–692.
- Thanellou, A., Green, J.T., 2013. Cerebellar structure and function in male Wistar-Kyoto hyperactive rats. *Behav. Neurosci.* 127 (2), 311–324.
- Thapar, A., Cooper, M., Eyre, O., Langley, K., 2013. What have we learnt about the causes of ADHD? *J. Child Psychol. Psychiatry* 54 (1), 3–16.
- Toto, M., Margari, F., Simone, M., Craig, F., Petruzzelli, M.G., Tafuri, S., Margari, L., 2015. Antibasal ganglia antibodies and antistreptolysin O in noncomorbid ADHD. *J. Atten. Disord.* 19 (11), 965–970.
- Tritsch, N.X., Ding, J.B., Sabatini, B.L., 2012. Dopaminergic neurons inhibit striatal output through non-canonical release of GABA. *Nature* 490 (7419), 262–266.
- Uher, R., McGuffin, P., 2008. The moderation by the serotonin transporter gene of environmental adversity in the aetiology of mental illness: review and methodological analysis. *Mol. Psychiatry* 13 (2), 131–146.
- Uher, R., McGuffin, P., 2010. The moderation by the serotonin transporter gene of environmental adversity in the etiology of depression: 2009 update. *Mol. Psychiatry* 15 (1), 18–22.
- Valentini, V., Piras, G., De Luca, M.A., Perra, V., Bordini, F., Borsini, F., Frau, R., Di Chiara, G., 2013. Evidence for a role of a dopamine/5-HT6 receptor interaction in cocaine reinforcement. *Neuropharmacology* 65, 58–64.
- van Rooij, D., Hoekstra, P.J., Mennes, M., von Rhein, D., Thissen, A.J., Heslenfeld, D., Zwiers, M.P., Faraone, S.V., Oosterlaan, J., Franke, B., Rommelse, N., Buitelaar, J.K., Hartman, C.A., 2015. Distinguishing adolescents with ADHD from their unaffected siblings and healthy comparison subjects by neural activation patterns during response inhibition. *Am. J. Psychiatry* 172 (7), 674–683.
- Vargas, D.L., Nascimbene, C., Krishnan, C., Zimmerman, A.W., Pardo, C.A., 2005. Neuroglial activation and neuroinflammation in the brain of patients with autism. *Ann. Neurol.* 57 (1), 67–81.
- Velasquez, J.C., Goeden, N., Bonnin, A., 2013. Placental serotonin: implications for the developmental effects of SSRIs and maternal depression. *Front. Cell. Neurosci.* 7, 47.
- Vermiglio, F., Lo Presti, V.P., Moleti, M., Sidoti, M., Tortorella, G., Scaffidi, G., Castagna, M.G., Mattina, F., Violi, M.A., Crisa, A., Artemisia, A., Trimarchi, F., 2004. Attention deficit and hyperactivity disorders in the offspring of mothers exposed to mild-moderate iodine deficiency: a possible novel iodine deficiency disorder in developed countries. *J. Clin. Endocrinol. Metab.* 89 (12), 6054–6060.
- Villa, A., Gelosa, P., Castiglioni, L., Cimino, M., Rizzi, N., Pepe, G., Lolli, F., Marcello, E., Sironi, L., Vegeto, E., Maggi, A., 2018. Sex-specific features of microglia from adult mice. *Cell Rep.* 23 (12), 3501–3511.
- Volkow, N.D., Wang, G.J., Fowler, J.S., Ding, Y.S., 2005. Imaging the effects of methylphenidate on brain dopamine: new model on its therapeutic actions for attention-deficit/hyperactivity disorder. *Biol. Psychiatry* 57 (11), 1410–1415.
- Volkow, N.D., Wang, G.J., Kollins, S.H., Wigal, T.L., Newcorn, J.H., Telang, F., Fowler, J.S., Zhu, W., Logan, J., Ma, Y., Pradhan, K., Wong, C., Swanson, J.M., 2009. Evaluating dopamine reward pathway in ADHD: clinical implications. *JAMA* 302 (10), 1084–1091.
- Vorhees, C.V., Graham, D.L., Braun, A.A., Schaefer, T.L., Skelton, M.R., Richtand, N.M., Williams, M.T., 2012. Prenatal immune challenge in rats: altered responses to dopaminergic and glutamatergic agents, prepulse inhibition of acoustic startle, and reduced route-based learning as a function of maternal body weight gain after prenatal exposure to poly IC. *Synapse* 66 (8), 725–737.
- Vuillermot, S., Joodmardi, E., Perlmann, T., Ogren, S.O., Feldon, J., Meyer, U., 2012. Prenatal immune activation interacts with genetic Nurr1 deficiency in the development of attentional impairments. *J. Neurosci.* 32 (2), 436–451.
- Wang, M.Z., Jin, P., Bumcrot, D.A., Marigo, V., McMahon, A.P., Wang, E.A., Woolf, T., Pang, K., 1995. Induction of dopaminergic neuron phenotype in the midbrain by sonic hedgehog protein. *Nat. Med.* 1 (11), 1184–1188.
- Weaver-Mikaere, L., Gunn, A.J., Mitchell, M.D., Bennet, L., Fraser, M., 2013. LPS and TNF alpha modulate AMPA/NMDA receptor subunit expression and induce PGE2 and glutamate release in preterm fetal ovine mixed glial cultures. *J. Neuroinflammation* 10, 153.
- Werenberg Dreier, J., Nybo Andersen, A.M., Hvolby, A., Garne, E., Kragh Andersen, P., Berg-Beckhoff, G., 2016. Fever and infections in pregnancy and risk of attention deficit/hyperactivity disorder in the offspring. *J. Child Psychol. Psychiatry* 57 (4), 540–548.
- Wiggs, K., Elmore, A.L., Nigg, J.T., Nikolas, M.A., 2016. Pre- and perinatal risk for attention-deficit hyperactivity disorder: does neuropsychological weakness explain the link? *J. Abnorm. Child Psychol.* 44 (8), 1473–1485.
- Willcutt, E.G., Doyle, A.E., Nigg, J.T., Faraone, S.V., Pennington, B.F., 2005. Validity of the executive function theory of attention-deficit/hyperactivity disorder: a meta-analytic review. *Biol. Psychiatry* 57 (11), 1336–1346.
- Willcutt, E.G., Nigg, J.T., Pennington, B.F., Solanto, M.V., Rohde, L.A., Tannock, R., Loo, S.K., Carlson, C.L., McBurnett, K., Lahey, B.B., 2012. Validity of DSM-IV attention deficit/hyperactivity disorder symptom dimensions and subtypes. *J. Abnorm. Psychol.* 121 (4), 991–1010.
- Winter, C., Reutiman, T.J., Folsom, T.D., Sohr, R., Wolf, R.J., Juckel, G., Fatemi, S.H., 2008. Dopamine and serotonin levels following prenatal viral infection in mouse—implications for psychiatric disorders such as schizophrenia and autism. *Eur. Neuropsychopharmacol.* 18 (10), 712–716.
- Winter, C., Djodari-Irani, A., Sohr, R., Morgenstern, R., Feldon, J., Juckel, G., Meyer, U., 2009. Prenatal immune activation leads to multiple changes in basal neurotransmitter levels in the adult brain: implications for brain disorders of neurodevelopmental origin such as schizophrenia. *Int. J. Neuropsychopharmacol.* 12 (4), 513–524.
- Wohleb, E.S., McKim, D.B., Sheridan, J.F., Godbout, J.P., 2014. Monocyte trafficking to the brain with stress and inflammation: a novel axis of immune-to-brain communication that influences mood and behavior. *Front. Neurosci.* 8, 447.
- Yan, Q.S., Yan, S.E., 2001. Activation of 5-HT(1B/1D) receptors in the mesolimbic dopamine system increases dopamine release from the nucleus accumbens: a microdialysis study. *Eur. J. Pharmacol.* 418 (1–2), 55–64.
- Yates, C.M., Calder, P.C., Ed Rainger, G., 2014. Pharmacology and therapeutics of omega-3 polyunsaturated fatty acids in chronic inflammatory disease. *Pharmacol. Ther.* 141 (3), 272–282.
- Zavitsanou, K., Lim, C.K., Purves-Tyson, T., Karl, T., Kassiou, M., Banister, S.D., Guillemin, G.J., Weickert, C.S., 2014. Effect of maternal immune activation on the kynurenine pathway in preadolescent rat offspring and on MK801-induced hyperlocomotion in adulthood: amelioration by COX-2 inhibition. *Brain Behav. Immun.* 41, 173–181.
- Zayats, T., Athanasiu, L., Sonderby, I., Djurovic, S., Westlye, L.T., Tamnes, C.K., Fladby, T., Aase, H., Zeiner, P., Reichborn-Kjennerud, T., Knappskog, P.M., Knudsen, G.P., Andreassen, O.A., Johansson, S., Haavik, J., 2015. Genome-wide analysis of attention deficit hyperactivity disorder in Norway. *PLoS One* 10 (4), e0122501.
- Zhang, Z., Bassam, B., Thomas, A.G., Williams, M., Liu, J., Nance, E., Rojas, C., Slusher, B.S., Kannan, S., 2016. Maternal inflammation leads to impaired glutamate homeostasis and up-regulation of glutamate carboxypeptidase II in activated microglia in the fetal/newborn rabbit brain. *Neurobiol. Dis.* 94, 116–128.