



EVII expression is associated with aggressive behavior in intrahepatic cholangiocarcinoma

Mariko Tanaka¹ · Junji Shibahara² · Shumpei Ishikawa³ · Tetsuo Ushiku¹ · Tepei Morikawa⁴ · Aya Shinozaki-Ushiku¹ · Akimasa Hayashi¹ · Kento Misumi¹ · Atsushi Tanaka¹ · Hiroto Katoh³ · Kei Sakuma¹ · Takashi Kokudo⁵ · Yoshinori Inagaki⁵ · Junichi Arita⁵ · Yoshihiro Sakamoto⁶ · Kiyoshi Hasegawa⁵ · Masashi Fukayama¹

Received: 26 May 2018 / Revised: 10 September 2018 / Accepted: 11 October 2018 / Published online: 23 October 2018
© Springer-Verlag GmbH Germany, part of Springer Nature 2018

Abstract

Ecotropic virus integration site 1 protein homolog (EVII), a well-known oncogenic transcriptional factor of hematopoietic cells, contributes to pancreatic cancer oncogenicity through increased expression of KRAS. Because EVII was upregulated in cholangiocarcinoma by referring The Cancer Genome Atlas, we investigated the importance of EVII in intrahepatic cholangiocarcinoma (ICC) which has been regarded as a heterogeneous group of cancers. Immunohistochemical analysis results demonstrated that EVII was overexpressed in about half of ICC (53/101, 52.5%). Moreover, all intraductal papillary neoplasms of the bile duct cases expressed EVII regardless of histological grading and subtypes such as gastric, intestinal, pancreatobiliary, or oncoytic (20/20, 100%). EVII-positive ICC showed higher frequencies of aggressive pathological indicators such as periductal infiltrative growth ($p = 0.022$), hilar invasion ($p = 0.041$), advanced UICC stage ($p = 0.026$), major vascular invasion ($p = 0.026$), and perineural invasion ($p = 0.007$) than EVII-negative ICC. Patients with EVII-positive ICC showed worse overall survival and recurrence-free survival in all resected cases and in curative resected cases. Recently, we proposed type 1/2 (large/small duct types) classification of ICC based on mucin productivity and immunophenotypes (S100P, N-cadherin, and NCAM). Type 1 predominantly consisted of EVII-positive ICC (33/42 cases, 79%), and the frequency was significantly higher than type 2 (18/55 cases, 32.7%) ($p < 0.0001$). EVII-positive ICC was likely to express stomach-specific claudin CLDN18 (correlation coefficient $r = 0.55373$) and mucin MUC5AC ($r = 0.42718$). EVII-positive ICC is an aggressive ICC showing both large-duct and/or gastric phenotypes. Consequently, a transcriptional factor EVII is associated with aggressive behavior in ICC and can be a therapeutic target molecule, while EVII might be a key molecule for the development of intraductal papillary neoplasms of the bile duct.

Keywords EVII · Intrahepatic cholangiocarcinoma · Type 1 · Type 2 · Gastric phenotype

Introduction

Prognoses of intrahepatic cholangiocarcinoma (ICC), the second most common liver cancer next to hepatocellular carcinoma, are poor because of its typical early invasion

and widespread metastasis, and because of a lack of effective therapeutic modalities [1]. In contrast to hepatocellular carcinoma, no approved targeted molecular therapy exists for ICC. Biliary tract and pancreas progenitors develop from endoderm cells in the embryonic foregut [2].

Electronic supplementary material The online version of this article (<https://doi.org/10.1007/s00428-018-2476-0>) contains supplementary material, which is available to authorized users.

✉ Masashi Fukayama
mfukayama-ky@umin.org

¹ Department of Pathology, Graduate School of Medicine, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo 113-0033, Japan

² Department of Pathology, Kyorin University Hospital, Tokyo, Japan

³ Department of Genomic Pathology, Medical Research Institute Tokyo Medical and Dental University, Tokyo, Japan

⁴ Pathology Division, NTT Medical Center Tokyo, Tokyo, Japan

⁵ Department of Surgery, Hepato-Biliary-Pancreatic Surgery, Graduate School of Medicine, The University of Tokyo, Tokyo, Japan

⁶ Department of Surgery, Kyorin University Hospital, Tokyo, Japan

Pathological conditions are sometimes similar in these organs, especially in hilar, extrahepatic bile duct, and pancreas [3–5]. For example, several inflammatory biliary diseases exist, such as IgG4-related disease and primary sclerosing cholangitis, and several biliary neoplastic lesions such as biliary intraepithelial neoplasia (BilIN) and intraductal papillary neoplasm of the bile duct (IPN-B), with pancreatic counterparts showing similar features [3]. Regarding the relevant genetic background, *KRAS* mutations apparently occur as an early molecular event during the progression of pancreatic intraepithelial neoplasia (PanIN) to pancreatic ductal adenocarcinoma and BilIN to cholangiocarcinoma, whereas p53 overexpression was identified as a late molecular event in pancreatic and biliary carcinogenesis [6].

Cholangiocarcinomas arise from different anatomical locations. They are classified as intrahepatic, perihilar, or distal type according to the staging of the American Joint Committee on Cancer (AJCC) and the Union for International Cancer Control (UICC) system [7]. Recently, several reports have described ICC as heterogeneous. Precise classification has been proposed, with ICC divided into large-duct (type 1) and small-duct (type 2) types [8, 9]. Moreover, recent studies have extracted the aggressive characteristic group of ICC. For example, the ICC cases expressing foregut or gastric epithelium-like molecules such as gastric-specific tight junction molecule, claudin-18 (CLDN18), or gastric mucin such as MUC5AC or MUC6 showed high incidence of metastasis and worse prognosis [10–14].

Recently, we reported that ecotropic virus integration site 1 protein homolog (EVII), a well-known oncogenic transcriptional factor of hematopoietic cells, plays an important oncogenic role for pancreatic cancer through increased expression level of *KRAS* [15]. Actually, we confirmed that EVII was upregulated in cholangiocarcinoma by referring The Cancer Genome Atlas (<http://firebrowse.org/viewGene.html?gene=MECOM>, Supplementary Fig. 1). Therefore, we investigated the importance of EVII in ICC. Furthermore, EVII is expressed immunohistochemically by gastric epithelium and is associated with gastric-related genes in the mRNA expression pattern [15]. We also investigated the correlation of EVII expression with bile duct and gastric phenotypes.

Material and methods

Cases

From The University of Tokyo Hospital pathology archives of 1995–2013, we reviewed 101 consecutive treated cases of ICC and 20 cases of intraductal papillary neoplasms of the bile duct (IPN-Bs). We retrieved records for

101 ICC of the non-papillary and mass-forming type with or without periductal infiltrative growth, and we followed the Union for International Cancer Control (UICC) definition which differentiated ICC from perihilar cholangiocarcinoma at the second-order bile duct. No cases were received preoperative treatment. All cases were examined in our previous study [8]. The intrahepatic IPN-Bs included 3 low-grade dysplasia, 11 high-grade dysplasia, and 6 carcinomas with invasion. The histological diagnosis of each lesion was based on the World Health Organization classification [1].

All aspects of this study were approved by The University of Tokyo Ethics Committee. Pathology reports and tissue slides were reviewed along with medical charts when necessary. Recurrent cases and cases with neoadjuvant treatment that significantly affected tumor morphology were excluded.

As a control material, we retrieved normal and inflamed mucosa from surgical specimens of hepatolithiasis or primary sclerosing cholangitis. All specimens were reviewed independently by two pathologists (MT and JS). Consensus between these pathologists was reached in all cases.

Histopathological examination

For each case, all tissue slides were reviewed. The entire tumor of the surgically resected specimen was fixed in 10% formalin at room temperature and was sectioned at intervals of 0.5–1.0 cm, with all tumor-containing sections processed routinely and embedded in paraffin. Serial sections of each tumor were cut and were stained with hematoxylin and eosin (H&E).

Immunohistochemical staining was performed for surgically resected specimens. The following primary antibodies were used: EVII C50E12 (1:1000; Cell Signaling Technology Inc., Danvers, MA, USA), CLDN18 (ZMD395, 1:1000; Zymed Laboratories Inc. Thermo Fisher Scientific Inc., Waltham, MA, USA), MUC5AC (CLH2, 1:100; Novocastra Laboratories Ltd., Newcastle upon Tyne, UK), and MUC6 (CLH5, 1:100; Novocastra Laboratories Ltd., Newcastle upon Tyne, UK). Normal gastric epithelial cells were used as a control for MUC5AC, MUC6, CLDN18, and EVII. For analyses, 4- μ m-thick, paraffin-embedded tissue sections from the representative area of each case were used. Immunohistochemical staining in surgically resected specimens was conducted according to standard techniques for a Ventana Benchmark® XT Autostainer (Roche Diagnostics Corp, Basel, Switzerland).

EVII was expressed in the nucleus. EVII expression was evaluated according to the staining pattern and intensity. The immunostaining of EVII was evaluated as negative, weakly positive, moderately positive, or strongly positive. Labeling of EVII was regarded as (3+) if more than 50% of the neoplastic cells were labeled at a moderate to strong intensity; (2+) if 25–

50% of the neoplastic cells were labeled at moderate to strong intensity, (1+) if 10–25% of the neoplastic cells were labeled at moderate to strong intensity, or if 25–100% of the neoplastic cells were labeled at weak intensity; and (0) if fewer than 10% of the neoplastic cells were labeled at any intensity. Two pathologists blindly and independently evaluated the immunohistochemical specimens.

To determine types 1 and 2, the methods were reported earlier [8], but they are briefly described as follows. We classified ICC into type 1 and type 2 based on an assessment of the following three factors: mucin productivity, S100P immunoreactivity score, and combined scores of N-cadherin and NCAM immunoreactivity. Typically, type 1 cases showed mucin production and diffuse immunoreactivity to S100P. Type 2 cases showed little mucin production and immunoreactivity to N-cadherin and/or NCAM.

Statistical analysis

Quantitative variables were compared using the Student *t* test. Categorical variables were compared using chi-square test or Fisher's exact test with software (JMP Pro 13.0; SAS Institute Inc., Cary, NC, USA). $p < 0.05$ was inferred as significant ($*p < 0.05$). Overall and recurrence-free survival curves were constructed using the Kaplan–Meier method and compared using the log-rank test with software (JMP Pro 13.0). Univariate and multivariate Cox regression analyses were performed to ascertain the prognostic factors using EZR software (<http://www.jichi.ac.jp/saitama-sct/SaitamaHP.files/statmed.html> Saitama Medical Center, Jichi Medical University). The factors of multivariate Cox regression analyses were generated using stepwise method using Bayesian information criterion. We analyzed the correlation coefficient using excel CORREL Function.

Results

EVII expression in non-neoplastic and neoplastic biliary tract lesions

Non-neoplastic intrahepatic biliary epithelium of any size was negative or only very weakly positive for EVII (Fig. 1a, b). Hepatocytes were also negative or only very weakly positive for EVII. No vascular endothelial cells were immunoreactive for EVII.

By contrast, many intrahepatic neoplasms exhibited frequent immunoreactivity for EVII in the nucleus. In about half of ICCs, EVII was expressed (53/101, 52.5%) (Fig. 1c, d). In intrahepatic IPN-Bs, all tumors were positive for EVII (20/20, 100%) (Fig. 1e, f).

Clinicopathological features of EVII-positive intrahepatic cholangiocarcinoma

EVII expression was observed in 53 cases (53%) of ICC. To assess the clinical significance of EVII expression, we compared the clinicopathologic features of EVII-positive and EVII-negative cases (Table 1). No significant difference was found among patients' clinical backgrounds. Regarding macroscopic features, comparing EVII-negative cases, EVII-positive cases were often associated with periductal infiltrative growth ($p = 0.022$) and hilar invasion ($p = 0.041$). Microscopically, major vascular invasion, perineural invasion, and advanced UICC stage (stage III to IV) were observed more frequently in EVII-positive cases ($p = 0.026$, $p = 0.007$, and $p = 0.026$, respectively). Results of survival analyses using Kaplan–Meier curves are depicted in Fig. 2. EVII-positive cases were associated with poorer recurrence-free survival and overall survival than EVII-negative cases in all resected cases ($p = 0.0287$ and $p = 0.0036$, respectively) and in curative resected cases ($p = 0.0327$ and $p = 0.0019$, respectively).

Univariate and multivariate Cox regression analyses were conducted to identify prognostic factors of ICC (Table 2). The prognostic factors for recurrence-free survival were EVII expression, HBV infection, tumor size, advanced stage, periductal infiltration on macroscopic appearance, hepatic hilar connective tissue invasion, major vessel invasion, perineural invasion, intrahepatic metastasis, lymph node metastasis, and surgical margin on univariate analysis. Regarding overall survival, EVII expression, advanced stage, periductal infiltration on macroscopic appearance, hepatic hilar connective tissue invasion, co-existing dysplasia, poorly differentiated histology, major vessel invasion, perineural invasion, intrahepatic metastasis, lymph node metastasis, and surgical margin were important for univariate analysis. We applied the multivariate analysis using stepwise method. Finally, the multivariate analysis revealed that HBV infection, advanced stage, major vascular invasion, and intrahepatic metastasis were important for recurrence-free survival, and that EVII expression, intrahepatic metastasis, and surgical margin were prognostic factors for overall survival.

Relation between EVII expression and specific subtype/phenotype of intrahepatic cholangiocarcinoma

Recently, several reports have presented some classification or some extraction of a specific phenotype in ICC [8, 9] and ICC is divided into large duct type (type 1) and small duct type (type 2). Furthermore, some aggressive cases can be extracted using immunohistochemical analysis. For example, ICC expressing foregut or gastric epithelium-like molecules behaved

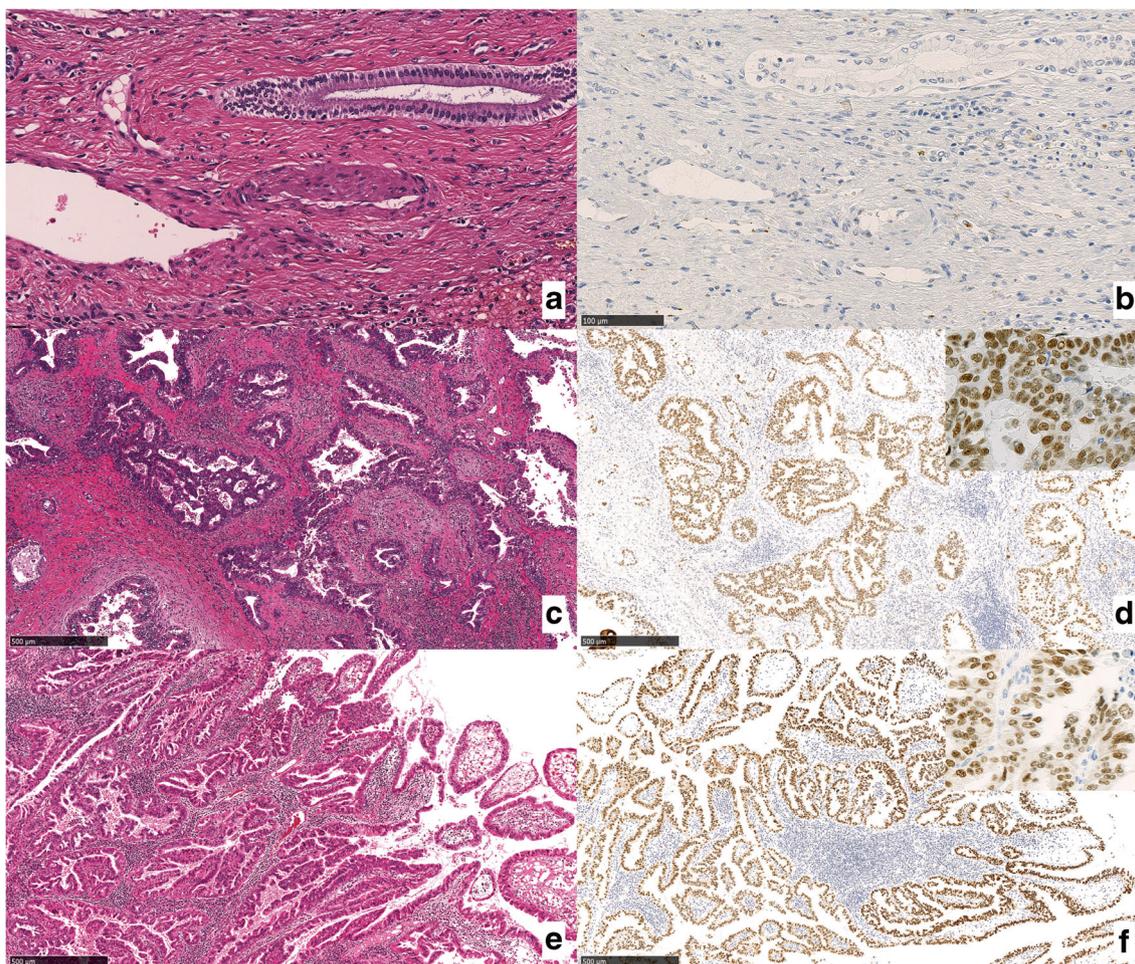


Fig. 1 EVI1 expression in non-neoplastic biliary tract epithelium and intrahepatic bile duct tumors. **a, b** Non-neoplastic biliary tract epithelium. Normal small bile ducts do not express EVI1 (**a** hematoxylin and eosin (H&E) staining; **b** EVI1 immunostaining, $\times 200$). **c, d** Intrahepatic

cholangiocarcinoma expressed EVI1 in the nucleus with strong intensity (**c** H&E; **d** EVI1, $\times 50$ (inset, $\times 400$)). **e, f** Intraductal papillary neoplasm of the bile duct expressed EVI1 in the nucleus with strong intensity (**e** H&E; **f** EVI1, $\times 50$ (inset, $\times 400$))

aggressively. Then, we examined the correlation between EVI1 positive cases and subtype or phenotype (Fig. 3).

We divided these 101 cases of ICC into type 1 and type 2 based on mucin productivity and immunophenotype (S100P, *N*-cadherin, and NCAM) as reported previously [8]. Type 1 ICC included more EVI1-positive cases (33/42, 78.6%) than type 2 ICC (18/55, 32.7%) ($p < 0.0001$, Fig. 3a).

Regarding gastric phenotypes, EVI1 expression was correlated with gastric marker expression such as CLDN18 (correlation coefficient $r = 0.55373$) and MUC5AC ($r = 0.42718$) in ICC (Fig. 3).

Discussion

In this study, EVI1 was expressed in intrahepatic biliary neoplasms; 100% of intraductal papillary neoplasm of bile duct (IPN-B) and 53% of ICC cases, in contrast to negative

expression in normal intrahepatic biliary tract epithelium. Importantly, EVI1-positive cases were associated with higher frequency of periductal infiltrative growth, major vascular invasion, and perineural invasion, and showing unfavorable prognosis. For them, EVI1 expression was an independent prognostic factor.

Recently, two subtypes of ICC were proposed. One type is thought to derive from small bile ducts or ductules consisting of tubular adenocarcinoma with scant mucus. The other type is thought to derive from the large bile ducts and is thought to be located at the perihilar location [8, 9, 16]. We proposed type 1 (almost all corresponding to perihilar, large duct type) and type 2 (almost all corresponding to peripheral and small duct type) as one classification [8]. The use of next-generation sequencing for ICC revealed recurrent actionable molecular alterations such as *TP53*, *IDH1*, and *KRAS* mutation [17–19] or amplification [20]. *KRAS* mutation was enriched in type 1 subtype, but *IDH1* mutation was enriched in type 2 subtype

Table 1 EVI1 expression and clinicopathologic features of intrahepatic cholangiocarcinomas

	EVI1-positive	EVI1-negative	<i>p</i> value
Age [> 65 years, ≤ 65 years]	30/23	24/24	0.506
Sex [M/F]	33/20	22/26	0.098
Hepatitis B virus infection [positive/negative]	5/48	4/43	1
Hepatitis C virus infection [positive/negative]	14/39	10/38	0.51
Tumor size [> 50 mm, ≤ 50 mm]	20/33	21/27	0.539
Periductal infiltrative growth [present/absent]	21/32	9/39	0.022*
Hilar invasion [present/absent]	26/27	14/34	0.041*
Co-existing dysplasia [present/absent]	6/47	3/45	0.493
Histology [poor/well to moderate]	6/47	3/45	0.493
UICC stage [III to IV/I to II]	22/31	10/38	0.026*
Major vascular invasion [present/absent]	12/41	3/45	0.026*
Perineural invasion [present/absent]	33 /20	17/31	0.007*
Intrahepatic metastasis [present/absent]	9/44	13/35	0.219
Lymph node metastasis [present/absent]	14/39	9/39	0.359

**p* < 0.05

[8, 9]. Our results demonstrated that EVI1-positive cases were dominant in the type 1 subtype. Above all, it is possible that the type 1 subtype depends on the KRAS pathway and that some relation exists between EVI1 and KRAS. We reported recently that EVI1 overexpression and KRAS mutation converge on activation of the KRAS pathway in pancreatic carcinogenesis [15]. EVI1 might upregulate KRAS pathway and might contribute to carcinogenesis of type 1 subtype in ICC.

EVI1 was also expressed in IPN-B regardless of histological grading and subtypes such as gastric, intestinal,

pancreatobiliary, or oncocytic. The finding is in parallel with that in EVI1 expression in pancreatic neoplasms, which is at high frequency in both pancreatic duct adenocarcinoma (PDAC) and precursor lesions, such as PanIN and pancreatic intraductal papillary neoplasm (IPMN). Since EVI1 was closely associated with aggressive behavior of ICC as described above, the role of EVI1 in precursor lesions may be different from that in overt carcinomas. KRAS mutation is a good example of the context-dependent role of oncogene. Additional mutations such as *CDKN2A*, *TP53*, or *SMAD4*

Fig. 2 Survival analysis of intrahepatic cholangiocarcinoma. **a** Overall survival of EVI1 +/- cases. **b** Overall survival of EVI1 +/- cases with curative resection. **c** Recurrence-free survival of EVI1 +/- cases. **d** Recurrence survival of EVI1 +/- cases with curative resection

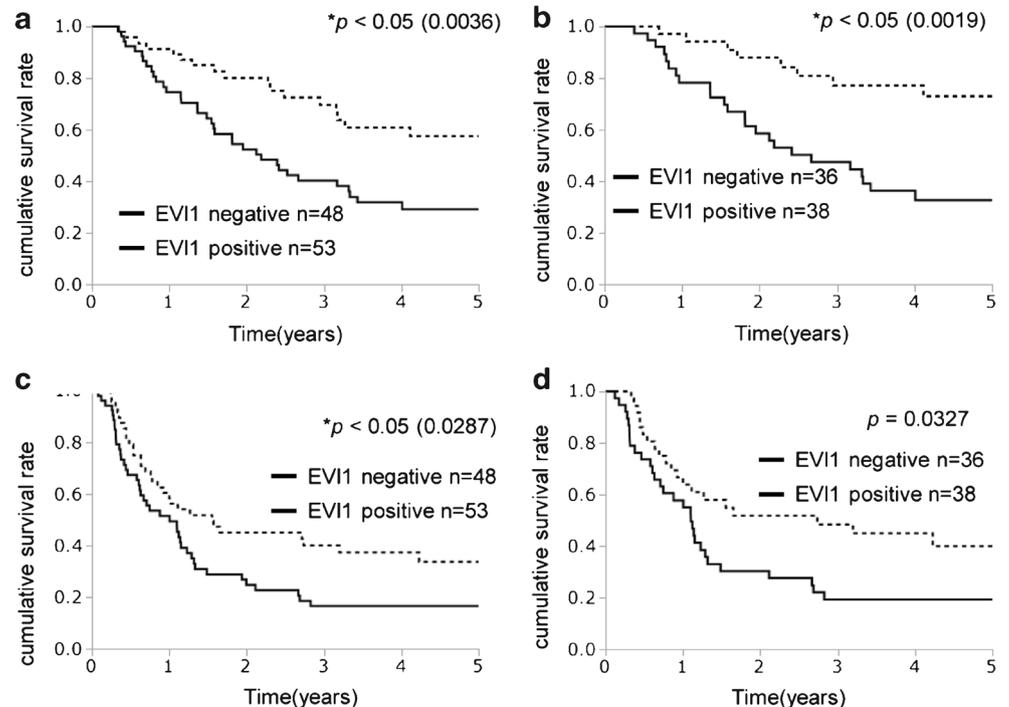


Table 2 Univariate and multivariate analysis of intrahepatic cholangiocarcinoma

Recurrence-free survival	Univariate analysis		Multivariate analysis ^a	
	HR (95% IC)	<i>p</i> value	HR (95% IC)	<i>p</i> value
Factors				
EVI1 expression	1.670 (1.049–2.657)	0.031*		
Age	0.991 (0.626–1.568)	0.968		
Sex	1.088 (0.687–1.722)	0.721		
Hepatitis B virus infection	2.352 (1.122–4.928)	0.023*	3.465 [1.521–7.893]	0.003*
Hepatitis C virus infection	0.583 (0.320–1.062)	0.078		
Tumor size [> 50 mm]	1.738 (1.089–2.773)	0.020*		
Stage [III-IV]	2.926 (1.780–4.810)	< 0.001*	2.376 [1.370–4.121]	0.002*
Periductal infiltrative growth	1.638 (1.009–2.661)	0.046*		
Hilar invasion	1.794 (1.130–2.848)	0.013*		
Co-existing dysplasia	1.836 (0.875–3.855)	0.108		
Histology [poor]	1.836 (0.875–3.855)	0.108		
Major vascular invasion	3.890 (2.147–7.049)	< 0.001*		
Perineural invasion	2.355 (1.467–3.780)	< 0.001*	3.073 [1.569–6.015]	0.001*
Intrahepatic metastasis	2.818 (1.679–4.730)	< 0.001*	2.583 [1.456–4.582]	0.001*
Lymph node metastasis	2.755 (1.637–4.637)	< 0.001*		
Surgical margin	2.244 (1.283–3.926)	0.005*		
Overall survival				
Factors				
EVI1 expression	2.228 (1.280–3.878)	0.005*	3.153 [1.659–5.992]	< 0.001*
Age	0.972 (0.575–1.645)	0.917		
Sex	1.115 (0.656–1.894)	0.689		
Hepatitis B virus infection	1.966 (0.837–4.614)	0.121		
Hepatitis C virus infection	0.865 (0.457–1.638)	0.656		
Tumor size [> 50 mm]	1.510 (0.890–2.561)	0.126		
Stage [III-IV]	2.879 (1.673–4.954)	< 0.001*		
Periductal infiltrative growth	2.225 (1.303–3.800)	0.003*		
Hilar invasion	1.728 (1.016–2.938)	0.043*		
Co-existing dysplasia	2.775 (1.246–6.180)	0.012*		
Histology [Poor]	2.775 (1.246–6.180)	0.012*		
Major vascular invasion	3.073 (1.594–5.924)	< 0.001*		
Perineural invasion	2.777 (1.600–4.818)	< 0.001*		
Intrahepatic metastasis	2.964 (1.692–5.190)	< 0.001*	3.066 [1.625–5.788]	< 0.001*
Lymph node metastasis	2.949 (1.641–5.301)	< 0.001*		
Surgical margin	3.281 (1.778–6.055)	< 0.001*	2.513 [1.326–4.764]	0.005*

**p* < 0.05^a Only significant factors are presented in the table, since all significant factors by univariate analysis were evaluated by multivariate Cox regression analyses using stepwise method with Bayesian information criterion

are needed for the progression to adenocarcinoma in the *KRAS* G12D transgenic mouse model [21]. On the other hand, different mutations, such as *TGF alpha*, *Smarca4/Brg1*, *GNAS*, *Avcrla*, or *PTEN* loss, drive ductal cells with the same *KRAS* mutation to IPMN [22, 23]. We speculate that EVI drives different sets of oncogene in precursor lesions and overt carcinomas of intrahepatic biliary epithelium.

Acquisition phenotypes of gastric epithelium are apparently important in several carcinogenic processes in pancreas

and bile ducts. Some researchers have specifically examined gastric mucin such as MUC5AC or MUC6 [24, 25]. We have also taken notice of the gastric epithelial tight-junction molecules CLDN18 in biliary carcinogenesis [11]. Our results demonstrate that EVI1 was expressed in all cases of IPN-Bs and in about half cases of ICC. Regarding IPN-Bs, all cases were positive for CLDN18 as reported previously [11]. Moreover, as for ICC, correlation between the expression of EVI1 and the expression of gastric markers was confirmed.

- genetic features of 2 histologic subtypes of intrahepatic cholangiocarcinoma. *Am J Surg Pathol* 40(8):1021–1030
9. Aishima S, Oda Y (2015) Pathogenesis and classification of intrahepatic cholangiocarcinoma: different characters of perihilar large duct type versus peripheral small duct type. *J hepato-bil-pan sci* 22(2):94–100
 10. Aishima S, Kuroda Y, Nishihara Y, Taguchi K, Taketomi A, Maehara Y, Tsuneyoshi M (2006) Gastric mucin phenotype defines tumour progression and prognosis of intrahepatic cholangiocarcinoma: gastric foveolar type is associated with aggressive tumour behaviour. *Histopathology* 49(1):35–44
 11. Shinozaki A, Shibahara J, Noda N, Tanaka M, Aoki T, Kokudo N, Fukayama M (2011) Claudin-18 in biliary neoplasms. Its significance in the classification of intrahepatic cholangiocarcinoma. *Virchows Arch* 459(1):73–80
 12. Tian F, Li D, Chen J, Liu W, Cai L, Li J, Jiang P, Liu Z, Zhao X, Guo F, Li X, Wang S (2013) Aberrant expression of GATA binding protein 6 correlates with poor prognosis and promotes metastasis in cholangiocarcinoma. *Eur J Cancer* 49(7):1771–1780
 13. Mall AS, Tyler MG, Ho SB et al (2010) The expression of MUC mucin in cholangiocarcinoma. *Pathol Res Pract* 206(12):805–809
 14. Abe T, Amano H, Shimamoto F, Hattori M, Kuroda S, Kobayashi T, Tashiro H, Ohdan H (2015) Prognostic evaluation of mucin-5AC expression in intrahepatic cholangiocarcinoma, mass-forming type, following hepatectomy. *Eur J Surg Oncol* 41(11):1515–1521
 15. Tanaka M, Suzuki HI, Shibahara J, Kunita A, Isagawa T, Yoshimi A, Kurokawa M, Miyazono K, Aburatani H, Ishikawa S, Fukayama M (2014) EVI1 oncogene promotes KRAS pathway through suppression of microRNA-96 in pancreatic carcinogenesis. *Oncogene* 33(19):2454–2463
 16. Akita M, Fujikura K, Ajiki T, Fukumoto T, Otani K, Azuma T, Itoh T, Ku Y, Zen Y (2017) Dichotomy in intrahepatic cholangiocarcinomas based on histologic similarities to hilar cholangiocarcinomas. *Mod Pathol* 30(7):986–997
 17. Sohal DP, Shrotriya S, Abazeed M et al (2016) Molecular characteristics of biliary tract cancer. *Crit Rev Oncol Hematol* 107:111–118
 18. Chong DQ, Zhu AX (2016) The landscape of targeted therapies for cholangiocarcinoma: current status and emerging targets. *Oncotarget* 7(29):46750–46767
 19. Moeini A, Sia D, Bardeesy N et al (2016) Molecular pathogenesis and targeted therapies for intrahepatic cholangiocarcinoma. *Clin Cancer Res* 22(2):291–300
 20. Nakamura H, Arai Y, Totoki Y, Shiota T, Elzawahry A, Kato M, Hama N, Hosoda F, Urushidate T, Ohashi S, Hiraoka N, Ojima H, Shimada K, Okusaka T, Kosuge T, Miyagawa S, Shibata T (2015) Genomic spectra of biliary tract cancer. *Nat Genet* 47(9):1003–1010
 21. Liu J, Ji S, Liang C, Qin Y et al (2016) Critical role of oncogenic KRAS in pancreatic cancer (review). *Mol Med Rep* 13(6):4943–4949
 22. Patra KC, Bardeesy N, Mizukami Y (2017) Diversity of precursor lesions for pancreatic cancer: the genetics and biology of intraductal papillary mucinous neoplasm. *Clin Transl Gastroen* 8(4):e86
 23. Kopp JL, Dubois CL, Schaeffer DF et al (2018) Loss of Pten and activation of Kras synergistically induce formation of intraductal papillary mucinous neoplasia from pancreatic ductal cells in mice. *Gastroenterology* 154(5):1509–1523.e5
 24. Kim GE, Bae HI, Park HU et al (2002) Aberrant expression of MUC5AC and MUC6 gastric mucins and sialyl Tn antigen in intraepithelial neoplasms of the pancreas. *Gastroenterology* 123(4):1052–1060
 25. Moschovis D, Bamias G, Delladetsima I (2016) Mucins in neoplasms of pancreas, ampulla of Vater and biliary system. *World J Gastrointest Oncol* 8(10):725–734
 26. Syed J, Pandian GN, Sato S et al (2014) Targeted suppression of EVI1 oncogene expression by sequence-specific pyrrole-imidazole polyamide. *Chem Biol* 21(10):1370–1380