



Transient immunosuppression during short interruption of HAART: Another key to HIV cure in the “Berlin patient”?



Konstantin V. Suslov

Institute of Immunology, 24/2 Kashirskoe Sh., Moscow 115 478, Russia

ARTICLE INFO

Keywords:

HIV
Transient immunosuppression
HAART
Neutralization-enhancing RF antibodies
NeRFa
“Berlin patient”
HIV cure

ABSTRACT

Transient immunosuppression in lentiviral infections leads to an auto-vaccination followed by the rise of serum neutralizing activity and a significant decrease in a set-point viral load, which becomes undetectable in some cases. Arguably, in the “Berlin patient” (Hütter G, et al., *N Engl J Med*, 2009) an induction chemotherapy-mediated transient immunosuppression episode during short interruption of HAART might have led to at least a “functional cure” before allogeneic stem cell transplantation. Neutralization-enhancing RF antibodies (NeRFa) induced as a part of secondary immune response after transient immunosuppression may have played a key role in neutralization of infectious HIV-IgG complexes in extracellular reservoirs. Transient immunosuppression during short non-structured treatment interruption (TI-SNSTI/HAART) regimen would be promising for the achievement of HIV cure on a large scale.

Introduction

Human immunodeficiency virus (HIV) in patients can form both extracellular [1–3] and intracellular reservoirs [4,5]. Nowadays significant efforts are being made towards a “sterilizing cure” by targeting latent virus in intracellular reservoirs [4–7]. It automatically implies that extracellular HIV has already been eliminated. In the reality, follicular dendritic cells (FDCs) in HIV-infected patients represent another obstacle to HIV cure [8,9] by forming the largest extracellular viral reservoir, which is not eradicated by highly active antiretroviral therapy (HAART) [3,10,11]. By the beginning of the asymptomatic period more than 98% of HIV particles are located on the surface of FDCs [12].

Follicular dendritic cells retain the long-term infectiousness of HIV particles covered by antibodies [3,13] and effectively support an infection of CD4+ T cells migrating through the secondary lymphoid organs [1,3]. An autopsy data showed that FDCs on their surface contain a monophyletic group of HIV variants with highly progressing diversity at the *env* gene by comparison with other compartments such as T cells [2]. This fact along with *in vitro* data [14] further support the hypothesis that at the end of asymptomatic period HIV *env* gene mutates via AID-mediated somatic hypermutation mechanism [15] like immunoglobulin genes encoding antibodies. Such mutations not only allow the virus to be ahead of the adaptive immune response but also may lead to uncontrolled R5-X4 HIV-1 switching towards acquired immunodeficiency syndrome (AIDS) [15].

IgM NeRFa for neutralization of infectious HIV-IgG complexes

Eradication of the extracellular HIV reservoirs always represented a problem probably due to the absence of the methodologies how to neutralize infectious HIV-IgG complexes especially in the light of the role of antibody-dependent enhancement of infection (ADE) in progression to AIDS [16,17]. At present rheumatoid factor-mediated neutralization of the virus in the presence of complement [18] is the only known way to neutralize extracellular viral particles covered by infection-enhancing antibodies. The fundamental defensive role of IgM rheumatoid factor (RF) in the protection of mother and the offspring from infection [19,20] is further supported by the ability of such antibodies to enhance neutralization of different pathogens [21], including HIV [22]. Physiological IgM RFs appearing during secondary immune response [23,24] should be discerned from pathological self-associating IgG RFs in rheumatoid arthritis [25].

Neutralization-enhancing RF antibodies (NeRFa) [26–28] might potentially prevent ADE and neutralize HIV in the presence of complement. Induction of NeRFa is promising for an elimination of infectious HIV-IgG complexes covered by complement molecules from the FDCs reservoir [27,29]. Transient immunosuppression with dexamethasone in ponies infected with equine infectious anemia virus with deleted principal neutralizing domain (EIAV_{ΔPND}) led to a 100-fold drop in a set-point viral load with a highly specific rise of the neutralizing activity in the serum [30]. This is consistent with the induction of NeRFa against EIAV_{ΔPND} [28]. In other cases of EIAV infection such an

E-mail address: suslov_kv@mail.ru.

<https://doi.org/10.1016/j.mehy.2018.12.002>

Received 2 October 2018; Accepted 6 December 2018

0306-9877/ © 2018 Elsevier Ltd. All rights reserved.

immunosuppressive treatment led even to undetectable values of set-point viral load [31]. Similar auto-vaccination effect leading to the rise of neutralizing activity coinciding with a sharp drop in viral load has been observed in HIV-infected patient after treatment with rituximab during HAART interruption [32].

A Key from the “Berlin patient”

The “Berlin patient” [33] up to date is the only patient in which a “sterilizing cure” for HIV has presumably been achieved [34]. Despite multiple thorough investigations of the case the following two questions arise:

- (i) Why the success of the “Berlin patient” still cannot be reproduced?
- (ii) Could the “functional” or even a “sterilizing” cure states in the “Berlin patient” have occurred before allogeneic stem cell transplantation (alloSCT)?

Indeed, several attempts to achieve HIV cure by means of homozygous CCR5delta32 (CC chemokine receptor 5 gene with a deletion of 32 base pairs) alloSCT have failed [4,35,36]. Also the viral load has been undetectable for the whole period starting from several days before alloSCT onwards indefinitely [33,34,37]. Therefore it is not excluded that even without alloSCT the result would be same.

In the article [37] authors compared one unique case, i.e. the “Berlin patient”, with other settings where HIV rebounded after HAART discontinuation following alloSCT. But this comparison cannot be the “evidence” that homozygous CCR5delta32 alloSCT has cured the “Berlin patient”. Indeed, one of the induction chemotherapy courses before alloSCT in the “Berlin patient” took place during a short interruption of HAART thus leading to a viral rebound up to 6.9×10^6 (HIV RNA/ml) [33]. This is a key aspect that distinguished the “Berlin patient” from other patients. Induction chemotherapy course for acute myeloid leukemia is inevitably immunosuppressive [33]. Unfortunately, for unknown reasons the authors avoided mentioning this key episode of the short interruption of HAART both in the article with an “evidence” for a cure [37] and even in the figure depicting timeline of clinical events in the “Berlin patient” [34].

It can be hypothesized that at least a “functional cure” in the “Berlin patient” has been achieved before alloSCT due to immunosuppressive episode on the induction chemotherapy course during short interruption of HAART by taking into account the following facts:

- (i) Post-transplantation therapy is severely immunosuppressive [33]. Therefore after transplantation while being on the regimen off HAART the rapid viral rebound within 3–4 weeks is expected like in similar post-transplantation settings [36,38,39]. This time is almost twice as little as the time (61 day after alloSCT) [33] of achievement of complete chimerism in the “Berlin patient”.
- (ii) Extracellular viral reservoirs are not affected by alloSCT. Viral load after alloSCT was undetectable despite the fact that intestinal macrophages expressed CCR5 receptor even 159 days after alloSCT [33].
- (iii) Preexisting X4/R5X4 HIV-1 variants (2,9% of the total viral population) “disappeared” after alloSCT [33]. This is in contrast to the quick rebound of X4 variants in a similar setting of homozygous CCR5delta32 alloSCT [36].
- (iv) Antibodies against HIV gp120 and gp41 envelope proteins, in contrast to other viral proteins, remain in significant amounts 625 days after alloSCT [33]. The same trend continues to be at 40 months [37] and even 60 months [34] after alloSCT despite constantly undetectable viral load.

All these facts are consistent with a model according to which NeRFa might have been induced after the short immunosuppressive episode off HAART at the second course of induction chemotherapy

before alloSCT. Neutralization-enhancing RF antibodies may neutralize the extracellular virus via complement-dependent mechanism [18,22,40] with subsequent clearance of immune complexes [41]. It is not excluded that cells expressing HIV antigens, like cancer cells expressing viral antigens on their surfaces, might be recognized by NeRFa with subsequent destruction by complement-mediated cytotoxicity [27,42]. Latently infected cells either have a limited life span or might be destructed anyway after spontaneous HIV reactivation by the mechanism mentioned above. Continuous NeRFa-mediated neutralization of all extracellular viral particles even on its own might be able to eliminate all HIV from the organism due to the limited life span of infected cells. To sum up, the successful induction of NeRFa in the “Berlin patient” might have led to the gradual eradication of both extracellular and intracellular HIV reservoirs thus approaching the “sterilizing cure” state in the long term.

The last “feast” for HIV

Transient immunosuppression (TI) regimen may lead to a significant decrease of the set-point viral load (SPVL) [30]. On the contrary, HAART does not change SPVL [32,43]. It seems that the decrease of SPVL might prolong the length of the asymptomatic period in HIV-1 infection [44]. Both short non-structured treatment interruption (≤ 4 weeks) [45] and transient immunosuppression off HAART [32,33,38] regimens seem to be safe for patients. Combination of two regimens into one would lead to the transient immunosuppression during short non-structured treatment interruption (TI-SNSTI/HAART) setting, in which HAART plays a very important role to control the maximal viral load during immunosuppressive episode.

Transient immunosuppression off HAART regimen is characterized by at least 10 to 100-fold rises in viral load [32,38] during immune suppression in comparison to the initial SPVL. It seems that maximal value of viral load and the speed with which it rises during immunosuppression episode might be critical for the successful induction of NeRFa at secondary immune response upon “burst” of viral replication. Maximal viral load upon TI-SNSTI/HAART regimen in the “Berlin patient” before alloSCT [33] and in the “Frankfurt patient” after alloSCT [38] reach almost the same value. Therefore future analytical treatment interruption in the “Frankfurt patient” might say whether the “feast” for HIV upon its replication just after alloSCT was the last one. Also analytical treatment interruption in the “Oxford patient” [32] may be informative.

A view outlined above points to the new possibilities to be investigated. Reproduction of TI-SNSTI/HAART regimen in HIV-infected individuals exactly as in the “Berlin patient” might be particularly promising. Will it lead to the absence of viral rebound after analytical treatment interruption or even to a “sterilizing cure”? By that time the beautiful molecular mechanisms behind such a success will probably require even more attention from many physicians working urgently to reproduce HIV eradication on a massive scale.

Conflict of interest statement

None

References

- [1] Spiegel H, Herbst H, Niedobitek G, Foss HD, Stein H. Follicular dendritic cells are a major reservoir for human immunodeficiency virus type 1 in lymphoid tissues facilitating infection of CD4+ T-helper cells. *Am J Pathol* 1992;140:15–22.
- [2] Keele BF, Tazi L, Gartner S, et al. Characterization of the follicular dendritic cell reservoir of human immunodeficiency virus type 1. *J Virol* 2008;82:5548–61.
- [3] Burton GF, Keele BF, Estes JD, Thacker TC, Gartner S. Follicular dendritic cell contributions to HIV pathogenesis. *Semin Immunol* 2002;14:275–84.
- [4] Pino M, Paiardini M, Marconi VC. Progress in achieving long-term HIV remission. *Curr Opin HIV AIDS* 2018;13:435–45.
- [5] Passaes CP, Sáez-Cirión A. HIV cure research: advances and prospects. *Virology* 2014;454–455:340–52.

- [6] Johnston R, Barré-Sinoussi F. Controversies in HIV cure research. *J Int AIDS Soc* 2012;15:16.
- [7] Deleage C, Wietgreffe SW, Del Prete G, et al. Defining HIV and SIV reservoirs in lymphoid tissues. *Pathog Immun* 2016;1:68–106.
- [8] Bronnimann MP, Skinner PJ, Connick E. The B-cell follicle in HIV infection: barrier to a cure. *Front Immunol* 2018;9:20.
- [9] Dave RS, Jain P, Byrareddy SN. Follicular dendritic cells of lymph nodes as human immunodeficiency virus/simian immunodeficiency virus reservoirs and insights on cervical lymph node. *Front Immunol* 2018;9:805.
- [10] Alòs L, Navarrete P, Morente V, et al. Immunoarchitecture of lymphoid tissue in HIV-infection during antiretroviral therapy correlates with viral persistence. *Mod Pathol* 2005;18:127–36.
- [11] Dutartre H. HIV-1 is stored by follicular dendritic cells in lymph nodes even under antiviral treatments. *Med Sci (Paris)* 2016;32:803–5.
- [12] Pantaleo G, Cohen OJ, Schacker T, et al. Evolutionary pattern of human immunodeficiency virus (HIV) replication and distribution in lymph nodes following primary infection: implications for antiviral therapy. *Nat Med* 1998;4:341–5.
- [13] Heath SL, Tew JG, Tew JG, Szakal AK, Burton GF. Follicular dendritic cells and human immunodeficiency virus infectivity. *Nature* 1995;377:740–4.
- [14] Balin SJ, Ross TM, Platt JL, Cascalho M. HIV genes diversify in B cells. *Curr HIV Res* 2008;6:10–8.
- [15] Suslov KV. Does AID aid AIDS? *Immunol Lett* 2004;91:1–2.
- [16] Homsy J, Meyer M, Levy JA. Serum enhancement of human immunodeficiency virus (HIV) infection correlates with disease in HIV-infected individuals. *J Virol* 1990;64:1437–40.
- [17] Szabó J, Prohászka Z, Tóth FD, et al. Strong correlation between the complement-mediated antibody-dependent enhancement of HIV-1 infection and plasma viral load. *AIDS* 1999;13:1841–9.
- [18] Ashe WK, Daniels CA, Scott GS, Notkins AL. Interaction of rheumatoid factor with infectious herpes simplex virus-antibody complexes. *Science* 1971;172:176–7.
- [19] Clarkson Jr AB, Mellow GH. Rheumatoid factor-like immunoglobulin M protects previously uninfected rat pups and dams from *Trypanosoma lewisi*. *Science* 1981;214:186–8.
- [20] Reimer CB, Black CM, Phillips DJ, et al. The specificity of fetal IgM: antibody or anti-antibody? *Ann N Y Acad Sci* 1975;254:77–93.
- [21] Green TJ, Packer BJ. A role for rheumatoid factor enhancement of *Plasmodium falciparum* schizont inhibition in vitro. *Infect Immun* 1984;46:668–72.
- [22] Douvas A, Takehana Y, Ehresmann G, Chernyovskiy T, Daar ES. Neutralization of HIV type 1 infectivity by serum antibodies from a subset of autoimmune patients with mixed connective tissue disease. *AIDS Res Hum Retroviruses* 1996;12:1509–17.
- [23] Tarkowski A, Czerkinsky C, Nilsson LA. Simultaneous induction of rheumatoid factor- and antigen-specific antibody-secreting cells during the secondary immune response in man. *Clin Exp Immunol* 1985;61:379–87.
- [24] Coulie P, Van Snick J. Rheumatoid factors and secondary immune responses in the mouse. II. Incidence, kinetics and induction mechanisms. *Eur J Immunol* 1983;13:895–9.
- [25] Almeida JD, Griffith AH. Viral infections and rheumatic factor. *Lancet* 1980;2:1361–2.
- [26] Suslov KV. Neutralization-enhancing RF antibodies for HIV vaccines. *Front Immunol* 2014;5:634.
- [27] Suslov KV. Neutralization-enhancing RF antibodies. Berhardt LV, editor. *Advances in Medicine and Biology*, Vol. 87. New York: Nova Science Publishers; 2015. p. 81–106.
- [28] Suslov KV. Orchestrating HIV neutralization by secondary immune response-mediated induction of RF antibodies. *Curr Immunol Rev* 2016;12:118–24.
- [29] Suslov KV. HIV neutralization in extracellular reservoirs. 2017. Available: <http://konstantin.suslov.googlepages.com> (Published: 17 July 2017; accessed 17.09.18).
- [30] Craigo JK, Leroux C, Howe L, et al. Transient immune suppression of inapparent carriers infected with a principal neutralizing domain-deficient equine infectious anaemia virus induces neutralizing antibodies and lowers steady-state virus replication. *J Gen Virol* 2002;83:1353–9.
- [31] Kono Y, Hirasawa K, Fukunaga Y, Taniguchi T. Recrudescence of equine infectious anemia by treatment with immunosuppressive drugs. *Natl Inst Anim Health Q (Tokyo)* 1976;16:8–15.
- [32] Huang KH, Bonsall D, Katzourakis A, et al. B-cell depletion reveals a role for antibodies in the control of chronic HIV-1 infection. *Nat Commun* 2010;1:102.
- [33] Hütter G, Nowak D, Mossner M, et al. Long-term control of HIV by CCR5 Delta32/Delta32 stem-cell transplantation. *N Engl J Med* 2009;360:692–8.
- [34] Yukl SA, Boritz E, Busch M, et al. Challenges in detecting HIV persistence during potentially curative interventions: a study of the Berlin patient. *PLoS Pathog* 2013;9. e1003347.
- [35] Rothenberger M, Wagner JE, Haase A, et al. Transplantation of CCR5Δ32 homozygous umbilical cord blood in a child with acute lymphoblastic leukemia and perinatally acquired HIV infection. *Open forum. Infect Dis* 2018;5:ofy090.
- [36] Verheyen J, Thielen A, Lübke N, et al. Rapid rebound of a preexisting CXCR4-tropic HIV variant after allogeneic transplantation with CCR5 delta32 homozygous stem cells. *Clin Infect Dis* 2018. <https://doi.org/10.1093/cid/ciy565>.
- [37] Allers K, Hütter G, Hofmann J, et al. Evidence for the cure of HIV infection by CCR5Δ32/Δ32 stem cell transplantation. *Blood* 2011;117:2791–9.
- [38] Wolf T, Rickerts V, Staszewski S, et al. First case of successful allogeneic stem cell transplantation in an HIV-patient who acquired severe aplastic anemia. *Haematologica* 2007;92:e56–8.
- [39] Kordelas L, Verheyen J, Beelen DW, et al. Shift of HIV tropism in stem-cell transplantation with CCR5 Delta32 mutation. *N Engl J Med* 2014;371:880–2.
- [40] Gipson TG, Daniels CA, Notkins AL. Interaction of rheumatoid factor with infectious vaccinia virus-antibody complexes. *J Immunol* 1974;112:2087–93.
- [41] Van Snick JL, Van Roost E, Markowitz B, Cambiaso CL, Masson PL. Enhancement by IgM rheumatoid factor of in vitro ingestion by macrophages and in vivo clearance of aggregated IgG or antigen-antibody complexes. *Eur J Immunol* 1978;8:279–85.
- [42] Hayashi K, Lodmell D, Rosenthal J, Notkins AL. Binding of ¹²⁵I-labeled anti-IgG, rheumatoid factor and anti-C3 to immune complexes on the surface of virus-infected cells. *J Immunol* 1973;110:316–9.
- [43] Koegl C, Wolf E, Hanhoff N, et al. Treatment during primary HIV infection does not lower viral set point but improves CD4 lymphocytes in an observational cohort. *Eur J Med Res* 2009;14:277–83.
- [44] Fraser C, Hollingsworth TD, Chapman R, de Wolf F, Hanage WP. Variation in HIV-1 set-point viral load: epidemiological analysis and an evolutionary hypothesis. *Proc Natl Acad Sci USA* 2007;104:17441–6.
- [45] Sanchez R, Portilla J, Gimeno A, et al. Immunovirologic consequences and safety of short, non-structured interruptions of successful antiretroviral treatment. *J Infect* 2007;54:159–66.