

Review Article

The role of vitamin D and P2X7R in multiple sclerosis

Veronica Tsin Fong Voo^{a,b}, Terence O'Brien^{b,c}, Helmut Butzkueven^b, Mastura Monif^{a,b,c,*}^a Department of Physiology, The University of Melbourne, Melbourne, Australia^b Department of Neuroscience, Monash University, Melbourne, Australia^c Department of Neurology, Melbourne Health, Melbourne, Australia

ARTICLE INFO

Keywords:

Multiple sclerosis
Neuroinflammation
P2X7R
Vitamin D
Innate immunity
Monocytes

ABSTRACT

Multiple sclerosis (MS) is characterized by neuroinflammatory infiltrates and central nervous system demyelination. In the neuroinflammatory foci of MS there is increased expression of a purinergic receptor, P2X7R. Although implicated in the neuroinflammation, the exact role of P2X7R in the context of MS is unclear and forms the basis of this review.

In this review, we also introduce the immunopathologies and inflammatory processes in MS, with a focus on P2X7R and the possible immunomodulatory role of vitamin D deficiency in this setting.

1. Introduction

Multiple sclerosis (MS) is a chronic autoimmune, neuroinflammatory disease that is characterized by demyelination and axonal damage in the central nervous system (CNS) (Goldenberg, 2012). MS is the most common cause of neurological disability in young adults between 20–50 years old, with a peak incidence occurring at 30 years of age (Milo and Kahana, 2010). The etiology of the disease remains largely unknown but a combination of genetic (Sadovnick et al., 1993) and environmental factors (Miller et al., 1990) are implicated in its pathogenesis. Moreover, MS has a two-three fold greater incidence in women than men (Orton et al., 2006; Trojano et al., 2012) and incidence and prevalence of MS increase with increasing distance from the equator (Dean, 1967; Milo and Kahana, 2010). Currently, there is no single diagnostic test for MS. A combination of clinical assessments, typical magnetic resonance imaging (MRI) lesion findings and laboratory investigations are used to make a diagnosis for MS, which is then refined to either the relapsing or progressive forms of MS according to disease progression (Goldenberg, 2012; Koriem, 2016). Many increasingly effective therapies have been introduced into the clinic in the last 20 years, these greatly reduce relapses and disability progression. However, the cause of MS remains elusive and the disease is still incurable.

Interestingly, even though vitamin D deficiency has been associated with an increased risk of developing MS (Ascherio et al., 2012; Ascherio et al., 2010), the exact role of vitamin D and its mechanism of action in the disease remain unclear. Additionally, the expression of P2X7 receptor (P2X7R) on cells of the immune and nervous system involved in MS suggest its potential role in the pathogenesis of the disease

(Sperlagh and Illes, 2014; Gu and Wiley, 2018). Hence, this review provides a summary of the current literature available on MS, focusing on the role of vitamin D deficiency and P2X7R as possible contributors to the disease state. Firstly, it will briefly elaborate on the pathophysiology of MS, including the inflammatory aspects of the disease with a focus on innate and adaptive immunity. The roles of P2X7R and vitamin D in MS are further evaluated, notably the effect of vitamin D on P2X7Rs expressed on immune cells involved in MS.

2. Multiple sclerosis

2.1. Overview

Pathologically, MS is characterized by focal destruction of the myelin sheath (Nakahara et al., 2012). According to the course of disease, MS can be categorized into two groups, the relapsing form and the progressive form (Lublin et al., 2014). The first attack of demyelination without disseminating neurological symptoms in both time and space is termed a clinically isolated syndrome (CIS), which is the onset for 85–95% of MS patients (Miller et al., 2005). Recurrent inflammation, demonstrated as either a clinical relapse or as a new MRI-visible lesion defines the onset of relapsing-remitting MS (RRMS), which is the relapsing form of MS (Lublin et al., 2014). The progressive forms of MS include secondary progressive MS (SPMS) and primary progressive MS (PPMS) (Lublin et al., 2014). These are characterized by steady, gradual and progressive neurologic deficits that can occur over months to years (Lublin et al., 2014). SPMS is diagnosed when a patient has had at least one relapse with improvement in the past, while PPMS is diagnosed in

* Corresponding author.

E-mail address: mastura.monif@monash.edu (M. Monif).

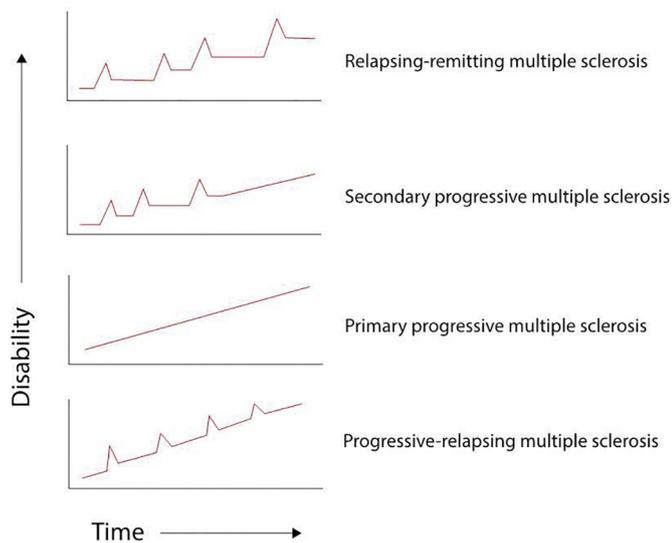


Fig. 1. Illustration of course of disease for different forms of MS. As outlined in figure 1, the relapsing form of MS, RRMS is marked by periods of relapse and remission of symptoms. Two-thirds of RRMS patients progress to SPMS where neurological deficits gradually worsen over time, without periods of remission. Patients with PPMS have persistent and gradually worsening disease from onset with no relapse-remission periods. PPMS progresses to primary-relapsing multiple sclerosis (PRMS) when patients gradually worsen with possible later super-imposed relapses.

the absence of any prior relapse but with progressive CNS disease consistent with MS (Lublin et al., 2014) (See Fig. 1).

2.2. Symptoms

The symptoms of MS are broad and can include physical, neuropsychiatric and cognitive deficits due to the multifocal nature of the lesions formed in the CNS (brain and spinal cord) (Korie, 2016; Savransky, 2018). The variability of MS symptoms depends on the location of lesions formed. MS generally starts with focal demyelination in the CNS then followed by worsening neuronal damage, and consequently in some cases ensuing weakness, paralysis or other neurological deficits (Korie, 2016). One of the first manifestations of MS is optic neuritis, which is an acute visual disturbance that is caused by an autoimmune attack on the optic nerves (Wilhelm and Schabet, 2015). The optic nerves are one of the most classical sites of attack in MS patients; half of all patients diagnosed with optic neuritis are found to develop MS within 15 years from the original episode (Brodsky et al., 2008). MS is a disease that evolves with relapses and remission with multifocal neurological deficits, the most common being optic neuritis and transverse myelitis (Savransky, 2018). Other possible symptoms include motor weakness, paresthesia or numbness, incoordination, problems in speech and hearing, fatigue, acute and chronic neuropathic pain, urinary urgency or retention, bowel function difficulties, sexual dysfunction, cognitive and neuropsychological deficits (Goldenberg, 2012; Korie, 2016).

The variability of MS symptoms can lead to diagnostic uncertainty resulting in delayed diagnosis and treatment (Murphy et al., 2017). Neuropsychiatric symptoms in MS are diverse and are reported to affect 60% of MS patients (Murphy et al., 2017). They can range from mood disorders such as depression, euphoria, anxiety disorders to cognitive deficits (Murphy et al., 2017; Sommerlad et al., 2014). Major depressive disorder (MDD) is the most common neuropsychiatric symptom associated with MS, with almost a 50% lifetime prevalence following disease onset (Murphy et al., 2017; Sommerlad et al., 2014). The etiology of MDD in MS patients is complex, due to the neurological and psychosocial factors implicated in the disease (Murphy et al., 2017; Ghaffar

and Feinstein, 2007). Suicide is also a significant cause of mortality in MS patients, which is found to be 7.5 times higher than the general population (Murphy et al., 2017; Sommerlad et al., 2014). The neuro-pathogenesis of the psychiatric conditions associated with MS remains obscure and it is still uncertain whether they are indicators of MS illness severity (Sommerlad et al., 2014). However, timely detection is essential as these disorders negatively impact quality of life and the ability of patients to adhere to recommended treatment regimens (Sommerlad et al., 2014).

2.3. Diagnosis and treatment

The diagnosis of MS is based on three criteria: (1) the space dissemination criterion (at least two different attacks), (2) the time dissemination criterion (at least two different attacks in the disease course, or evidence of simultaneous contrast enhancement and no contrast enhancement in a single MRI scan to indicate lesions of different age) and (3) the inflammatory criterion (chronic inflammation in the CNS) (McDonald et al., 2001; Garg and Smith, 2015; Thompson et al., 2018). MRI is the most common diagnostic test used to detect MS lesions in the CNS (Korie, 2016). Currently, the pattern of MRI lesions and presence of cerebrospinal fluid (CSF) specific oligoclonal bands (OCBs) are used to confirm MS diagnosis (Fadda et al., 2018).

In 2017, there are twelve treatments approved for RRMS in Australia (Sedal et al., 2017). These include interferons beta, glatiramer acetate, monoclonal antibodies such as natalizumab and ocrelizumab, small molecule oral agents such as fingolimod, cladribine, teriflunomide and dimethyl fumarate (Reich et al., 2018; Hollingworth et al., 2014; Deeks, 2018). The interferons and glatiramer acetate are used as treatment for RRMS as both can effectively reduce MS relapses by 30% and decrease the number of active lesions on brain MRI (Tsang and Macdonell, 2011); whereas infusion therapies such as natalizumab and ocrelizumab can reduce relapse rates by up to 60–75% (Polman et al., 2006; Mulero et al., 2018). All approved treatments in general have shown beneficial effects in the reduction of relapses and accumulation of disability in MS patients (Reich et al., 2018). However, the effects of these treatments on PPMS and SPMS have not been proven and they remain largely ineffective. This is partly due to the lack of literature on the exact mechanism of action of a number of MS disease modifying agents (Korie, 2016; Reich et al., 2018). Despite the recent development of new treatments for MS, there is still no cure with current therapies.

2.4. Glucocorticoids

Glucocorticoids such as intravenous (IV) methylprednisolone, oral prednisolone or dexamethasone are used to treat acute MS exacerbations due to their anti-inflammatory and immunosuppressive properties (Goodin, 2014; Johnson and Bhimji, 2018; Berkovich, 2013). A brief glucocorticoid treatment regimen such as IV methylprednisolone (1g daily for 3 days) is generally used to accelerate recovery from MS relapses with no significant adverse effects for most patients (Berkovich, 2013). However, the response to glucocorticoid treatment differs among patients, suggesting individual differences in sensitivity to glucocorticoid (DeRijk et al., 2004; van Winsen et al., 2005). Some studies have shown that patients with MS, particularly RRMS are less sensitive to glucocorticoids compared to healthy controls and patients with progressive forms of MS (DeRijk et al., 2004; van Winsen et al., 2005). Moreover, despite being effective in reducing relapse severity, glucocorticoids have no impact on MS disease course or the final degree of recovery (Goodin, 2014). Currently, there is no standard dose, route of administration or glucocorticoid types to treat MS relapses (Miller et al., 2000; Frohman et al., 2007). In most centers, 1g IV methylprednisolone for 3 days is utilized. More studies are required to determine the optimal glucocorticoid treatment for MS relapses, in conjunction with other MS therapeutics to optimize treatment during acute

attacks.

3. Immune system

Our present understanding of MS suggests that it is an immune-mediated disease (Yadav et al., 2015) due to the following findings: (1) T and B lymphocytes, and immune cell infiltrations are evident in lesions formed in the CNS (Lassmann, 2013); (2) immunomodulatory therapeutics affect MS disease course (Reich et al., 2018; Hartung and Kieseier, 2014); (3) genome wide association studies (GWAS) have identified numerous immune-related genetic variants (single nucleotide polymorphisms, SNPs) associated with MS (Sawcer et al., 2011; Beecham et al., 2013). Animal models and immunological studies of MS patients have also demonstrated a change in the involvement of the immune system during different phases of disease (Hemmer et al., 2015). Many studies have suggested that the peripheral immune responses targeting the CNS mediate early disease progression, whereas immune reactions within the CNS drive the progressive phase (Hemmer et al., 2015; Yadav et al., 2015; Kebir et al., 2009). The peripheral blood could also provide an accessible 'window' into the immunopathologies of MS. As suggested by Jones et al, changes in the circulating immune cells could reflect the immunological characteristics of MS lesions in the CNS and the immune dysregulation of MS patients (Jones et al., 2017).

3.1. Adaptive immune system

Neuroinflammation is the hallmark of MS, and is shown to consist of innate and adaptive immune cells such as T and B lymphocyte infiltrates as well as microglia, monocyte and macrophage accumulation (Jones et al., 2017; Yadav et al., 2015). The current central hypothesis is that myelin-reactive CD4⁺ T-cells drive the pathogenesis of MS through the migration of autoreactive CD4⁺ T-helper 1 (Th1) and CD4⁺ T-helper 17 (Th17) cells into the CNS (Yadav et al., 2015). The initial activation of autoreactive CD4⁺ T-cells is believed to take place in the systemic lymphoid organs by unknown factors; these T-cells are then subsequently activated upon encountering their target antigen (myelin components) in the CNS, resulting in demyelination, axonal damage and subsequent neurological disability (Hartung et al., 2014). For many years, MS was widely recognized as a Th1-mediated disease. These Th1 cells are differentiated in response to interleukin-12 (IL-12) and secrete the proinflammatory cytokine, interferon- γ (IFN- γ) (Renno et al., 1994; Tzartos et al., 2008). However, human in -vivo studies using peripheral blood and tissues from MS lesion and brain have later shown that along with Th1 cells, Th17 cells (i.e. CD4⁺ T-cells that express IL-17 and IL-22) are also likely to be important co-effectors in MS (Kebir et al., 2007; Tzartos et al., 2008; Li et al., 2017). IL-17 and IL-22 secreted by Th17 cells have been identified to disrupt the blood-brain-barrier (BBB) by binding to IL-17 and IL-22 receptors expressed on BBB-endothelial cells (Kebir et al., 2007). Although current literature does not convincingly determine whether single cytokines such as IL-17 or IFN- γ can sufficiently induce MS pathology, it is likely that any combinations of factors produced by Th1 and Th17 cells will contribute to the inflammatory demyelination processes in the CNS (Kebir et al., 2009). It is also believed that defects in peripheral tolerance due to intrinsic (costimulatory signaling, transcriptional and epigenetic mechanisms) and extrinsic (regulatory T-cells) factors can lead to dysregulation in CD4⁺ T-cell responses (Yadav et al., 2015; Gonsette, 2012). Immune responses are found to be skewed towards proinflammatory Th1/Th17 development and disrupt immune system homeostasis, subsequently leading to MS pathology (Yadav et al., 2015; Gonsette, 2012). Although CD8⁺ T-cells are the predominant T-cells in human MS lesions, their role in MS pathology are still largely unknown (Sinha et al., 2014). Several studies have found that neuroantigen-specific CD8⁺ T-cells exhibit immunoregulatory ability by suppressing the proliferation of self-reactive CD4⁺ T-cells when stimulated by their cognate antigen (Correale and Villa, 2010; Baughman et al., 2011). In addition, IL-17

and IFN- γ secretion are also reduced when CD8⁺ T-cells are added into CD4⁺ T-cell cultures isolated from MS patients (Correale and Villa, 2010; Baughman et al., 2011). However, the immunosuppressive effects of CD8⁺ T-cells are found to be reduced during acute exacerbations in MS, likely due to altered functionality rather than simple quantitative suppression (Cunnusamy et al., 2014; Baughman et al., 2011).

B-cells are also thought to be an important contributor to MS pathogenesis (Rahmanzadeh et al., 2018). Emerging studies have identified the following roles for B-cells in MS: (1) B-cells with hypermutated immunoglobulin (Ig) have been found to accumulate in CSF and lesions of MS patients (Colombo et al., 2000; Obermeier et al., 2008); (2) pronounced effects of B-cell therapies, in particular B-cell depleting agents (e.g. ocrelizumab and rituximab) on MS patients support the role of B-cells in MS pathogenesis (Kappos et al., 2011; Sabatino Jr. et al., 2018); (3) OCBs (produced by B cells) are detectable in 95% of MS patients (Rahmanzadeh et al., 2018) and are shown to be the second sensitive diagnostic marker after MRI (Rahmanzadeh et al., 2018; Arrambide et al., 2018). These findings suggest the potential role of B-cells in MS pathology and their use as therapeutic treatments for MS. It is also important to note that T and B lymphocytes are costimulatory. For instance, B-cell antigen presentation and co-stimulation are found to contribute to the induction of proinflammatory T-cell responses in MS pathology (Fraussen et al., 2016). Hence, it is unlikely that either T and B cell responses are independent from each other in the pathogenesis of MS.

3.2. Innate immune system

The rapid recruitment of monocytes from the periphery and activation of brain resident microglia are among the most consistent histological findings noted from MS autopsy specimens (Bruck et al., 1995; Ford et al., 1995). Despite the significant presence of innate immune cells in MS plaques, their underlying mechanisms contributing to MS pathology are not well understood (Weiner, 2009). It has been suggested that the innate immune system activates myelin-reactive T and B lymphocytes in the CNS (Hernandez-Pedro et al., 2013). The effector cells then in turn express various cytokines and activation markers that further activate the innate immune cells, creating a perpetuating cycle of neuroinflammation with ensuing neuronal damage and demyelination (Monney et al., 2002).

Due to similarities in both morphology and function, it is often difficult to distinguish microglia and monocytes using conventional immunohistochemical techniques (O'Loughlin et al., 2018). However, advancements in identifying specific molecular markers and signatures of these cells are currently being developed, allowing us to distinguish the complex roles of these cells in the neuroinflammatory and degenerative cascade in MS (Butovsky et al., 2014; Rinchai et al., 2016). Both the resident microglia and infiltrating monocytes could give rise to macrophages, but the macrophages derived are found to have different functions in the neuroinflammation of MS (Yamasaki et al., 2014). During both MS and EAE (experimental autoimmune encephalomyelitis; the murine model of MS), monocyte-derived macrophages accumulate in the inflammatory infiltrates associated with active demyelination, while CNS resident microglia-derived macrophages only participate in the later stages of disease (Yamasaki et al., 2014). Distinguishing the roles of microglia from monocytes will point towards new strategies to treat MS. Thus, more research is required to identify unique markers and signatures of these immune cells and also their specific roles in MS pathogenesis.

Microglia. Microglia cells are the most common immune cells in the CNS and are considered as CNS resident macrophages (Benveniste, 1997). They contribute to MS and EAE pathology through antigen presentation and secretion of proinflammatory cytokines (Benveniste, 1997). Persistent activation of microglia have been observed in chronic phases of relapsing-remitting (RR) EAE while increased microglial activation in white matter inflammation is more frequent in progressive

than RR patients (Kutzelnigg et al., 2005; Rasmussen et al., 2007). Microglia activate T and B lymphocytes through antigen presentation via major histocompatibility complex (MHC) class II and co-stimulatory molecules, CD83 and CD40 (Raivich and Banati, 2004; Gandhi et al., 2010). The rapid upregulation of MHC antigens is also one of the first molecular changes observed in MS patients and EAE models using immunohistochemistry techniques (Raivich and Banati, 2004). As the first-line defense in the brain, microglia express Toll-like receptors (TLRs) on their surface which contribute to neuroimmune regulation and CNS inflammation (Aravalli et al., 2007; Lee and Lee, 2002). Studies have shown an enhanced expression and upregulation of TLRs in the brain and spinal cord tissues of MS patients (Bsibsi et al., 2002; Marta et al., 2009). The role of TLRs in EAE pathology is also supported by findings that indicate the resistance of mice that lack MyD88 (the common TLR adaptor molecule) to EAE due to inhibition of monocyte and microglia activation (Marta et al., 2008; Prinz et al., 2006; Lee et al., 2018). Microglia are also involved in demyelination and phagocytosis of degraded myelin (Bauer et al., 1994), which lead to elevation in the expression of myeloperoxidases and reactive oxygen species (ROS), resulting in neuronal damage (Gray et al., 2008). Moreover, the activation patterns of microglia are distinct during different stages of MS lesions (O'Loughlin et al., 2018). Microglia in the early stages of active lesions show a more proinflammatory phenotype, with expression of inflammatory markers involved in antigen presentation, phagocytosis, generation of ROS and T-cell co-stimulation (Zrzavy et al., 2017). In later stages of active lesions, the microglial cells switch to an intermediate between pro- and anti-inflammatory phenotype (O'Loughlin et al., 2018; Zrzavy et al., 2017). In addition, microglial depletion is also shown to inhibit the development of inflammatory CNS lesions in EAE models (Heppner et al., 2005; Ponomarev et al., 2011). Thus, an understanding of the roles of microglia in MS pathology could provide a novel therapeutic target in attenuation of disease severity and demyelination for MS patients.

Monocytes. Human monocytes are derived from hematopoietic stem cells in the bone marrow and constitute around 10% of the total circulating blood leukocytes (Rinchai et al., 2016). They are released into the peripheral blood upon maturation, where they are either recruited to tissues or removed from the circulation after 1–2 days (Rinchai et al., 2016). Monocyte infiltration is evident in active MS lesions even in the early stages of disease and are associated with axonal demyelination (Baufeld et al., 2017; Bruck et al., 1995). Human monocytes are classified into three major subsets depending on the expression of cell surface markers CD14 and CD16, which also represent different inflammatory stages (Rinchai et al., 2016). The non-classical monocytes (CD14+CD16++) from MS patients are found to be significantly increased in the peripheral blood, highlighting the relevance of further focus on CD16 monocyte subsets in MS (Chuluundorj et al., 2014; Gjelstrup et al., 2018). In addition, findings from EAE models also demonstrated a global monocyte profile that is biased towards the proinflammatory 'M1 phenotype' in the blood and CNS (Ajami et al., 2011; Mikita et al., 2011). Administration of ex-vivo activated 'M2 monocytes' by interleukin-10 (IL-10) and interleukin-13 (IL-13) to the peripheral blood has also been found to suppress ongoing EAE and promote immunomodulatory actions limiting the expansion of acute CNS lesions (Mikita et al., 2011). These findings open new perspectives for therapeutic applications in MS. Interesting, one study has suggested the potential pathological role of monocytes in MS disease state by demonstrating high levels of monocyte-secreted proinflammatory factors in serum of MS patients compared to healthy controls (Baufeld et al., 2017). Analysis of the cytokine production in monocytes isolated from MS patients showed a significant increase in interleukin-1 (IL-1), interleukin-6 (IL-6) and tumour necrosis factor alpha (TNF α) compared to normal controls (Rudick and Ransohoff, 1992; Imamura et al., 1993; Fiedler et al., 2017). Although monocytes are thought to have primarily detrimental effects on MS lesion formation, studies have also shown that they play a dual role by promoting tissue repair and secreting anti-

inflammatory factors (Vaknin et al., 2011; Denney et al., 2012). For instance, in one study, monocytes derived from MS patients were found to have increased enzyme tissue transglutaminase (TG2) levels and this increase is correlated with an anti-inflammatory and migratory profile in MS (Sestito et al., 2017).

4. P2X7 receptor

4.1. Overview

P2X7R is one of the functional members of the ionotropic purine P2X receptor family (North, 2002). In general, all recombinant P2X subunits can assemble in vitro to form heteromeric structures, with the exception of P2X7R, which is only present as a homo-trimeric receptor (Di Virgilio et al., 2017). The main physiological agonist of P2X7R is adenosine triphosphate (ATP) but other structurally unrelated agents have been reported to activate it with unknown mechanisms (Di Virgilio et al., 2018a). Additionally, the synthetic analog, 2'-(3')-O-(4-benzoylbenzoyl)-ATP (BzATP) is found to be more potent at P2X7R activation than ATP (Jiang et al., 2013). P2X7Rs are bifunctional (Volonte et al., 2012). The binding of ATP induces a channel formation within milliseconds which is selective for small cations (Ca²⁺, Na⁺ and K⁺) (Di Virgilio et al., 2018b) and within seconds to minutes, a larger pore opens allowing permeation of molecules up to 900 Da (Volonte et al., 2012). In response to prolonged or repetitive stimulation, pore formation is often studied by measuring influx and intracellular accumulation of fluorescent dyes, such as YoPro-1 and ethidium (Jiang et al., 2013). The mechanism of P2X7R channel-to-pore transition remains unclear, and various intermolecular (acquisition of more P2X7R subunits) and intramolecular (dilatation of the channel opening) mechanisms have been proposed (Alves et al., 2014; Jindrichova et al., 2015; Khadra et al., 2013). A non-pore-forming mutant of P2X7R, P2X7RG345Y (where glycine 345 is changed to tyrosine) can be used to identify functional differences between the two states as this point mutation only abolishes pore conductance without altering the function and properties of the channel component (Monif et al., 2009).

4.2. P2X7R in inflammation

P2X7R has low affinity for ATP, in the millimolar range (Di Virgilio et al., 2018b). Extracellular ATP can often increase dramatically and accumulates at inflammatory sites, thus reaching the receptor activation threshold (Wilhelm et al., 2010). In addition, several factors that accumulate at inflammatory sites are shown to either act as positive allosteric modulators or support P2X7R gating mechanisms (Di Virgilio et al., 2018a; Ellsner et al., 2004; Ferrari et al., 2004; Sanz et al., 2009). The binding of ATP at P2X7R alters intracellular ion homeostasis, which can result in initiation and propagation of inflammatory cascades (Di Virgilio et al., 2018b; Monif et al., 2009). In addition, the presence of P2X7Rs on virtually all cells of the innate and adaptive immunity (Di Virgilio et al., 2017) suggests they regulate immune function and inflammatory responses (Aga et al., 2002; Monif et al., 2009). P2X7R also promotes the release of proinflammatory mediators such as IL-6 (Shieh et al., 2014), interleukin-8 (IL-8) (Wei et al., 2008), monocyte-chemoattractant protein-1 (MCP-1) (Panenka et al., 2001) and ROS (Hewinson and Mackenzie, 2007).

4.3. P2X7R and IL-1 β

P2X7R activation triggers a multiplicity of inflammatory responses, the most important being interleukin-1 β (IL-1 β) release via the activation of NLRP3 inflammasome (Munoz-Planillo et al., 2013; Savio et al., 2018; Monif et al., 2016). Initially, during innate immune responses, damage-associated molecular patterns (DAMPs) and pathogen-associated molecular patterns (PAMPs) activate pattern recognition receptors (PRRs) such as TLRs expressed on immune cells (Savio et al.,

2018). TLR activation mediates the NF κ B pathway, which acts as the first signal promoting the transcription of several genes that encode for inflammatory mediators such as inflammasome components and pro-IL-1 β (Savio et al., 2018). P2X7R stimulation represents the second signal through triggering K⁺ efflux, promoting NLRP3 inflammasome assembly and pro-IL-1 β processing and IL-1 release (Quan et al., 2018). The ability of P2X7R to drive IL-1 β release has clinical significance as IL-1 β is involved in various immune processes such as immunogenic apoptosis (Ghiringhelli et al., 2009). IL-1 β is also a key mediator in neurodegeneration, chronic inflammation and chronic pain (Volonte et al., 2012).

In addition, IL-1 β is also found to be able to rescue glucocorticoid-induced apoptosis in monocytes (Schmidt et al., 1999; Achuthan et al., 2018; McColl et al., 2007). One of the apoptotic mechanisms of glucocorticoids is through inhibiting the NF κ B pathways, leading to suppression of NF κ B-mediated inflammatory activities and interference of immune cell survival (van der Burg and van der Saag, 1996; Ottonello et al., 2005). The transcription factor, NF κ B is induced by pro-inflammatory cytokines and plays a role in inflammatory processes by directing the transcription of chemoattractants, cytokines (including NF κ B-inducing cytokines), cytokine receptors and cell adhesion molecules (van der Burg and van der Saag, 1996). Even in a normal physiological setting, IL-1 β is found to be able to prevent apoptosis in monocytes, highlighting its role in homeostatic control of the number of monocytes available for immune responses (Mangan et al., 1991).

4.4. P2X7R in MS

Despite the wide expression of P2X7R on immune cells (Di Virgilio and Vuerich, 2015; Yiangou et al., 2006), astrocytes (Narcisse et al., 2005), oligodendrocytes (Matute, 2008), Schwann cells (Faroni et al., 2014) and neurons (Rodrigues et al., 2015), only incomplete information is available regarding P2X7R-mediated signaling in these cells. Many immune cell types are involved in MS lesion formation, however activated monocytes are shown to be one of the first cell types to reach the brain and possibly initiate neuroinflammation (Amadio et al., 2017). Around 80% of human monocytes are found to express immunoreactive P2X7R, implicating the role of these receptors in mediating monocyte-induced inflammatory responses (Aga et al., 2002; Gudipaty et al., 2001). Moreover, human monocytes are also shown to have four to fivefold greater P2X7R expression compared to B and T lymphocytes (Gu et al., 2000). Glatiramer acetate which is an approved MS therapy has been shown to cause downregulation of P2X7R expression in peripheral blood monocytes and it is unclear if its effectiveness is simply due to this mechanism or otherwise. (Caragnano et al., 2012). In rat hippocampal cultures, we have shown that P2X7R expression drives microglia activation and proliferation (Monif et al., 2009). Another study using post-mortem spinal cord specimen of MS patients, also showed the localization of P2X7R immunoreactivity in activated microglial cells/macrophages in affected regions (Yiangou et al., 2006). These data suggest that by acquiring further knowledge about P2X7R expression and function on innate immune cells such as monocytes and microglia, we can better understand the molecular pathways of MS and possibly identify P2X7R as a potential marker for the disease.

Due to the inflammatory nature of MS, a link between P2X7R activation and MS lesion development has been proposed (Savio et al., 2018). Early post-mortem studies of MS patients have revealed high level of P2X7R expression in astrocytes and microglia in the brain and spinal cord (Narcisse et al., 2005; Yiangou et al., 2006). In studies of EAE murine models, upregulation of P2X7R in the astroglial cells has been observed during the early stages of EAE while increased expression of P2X7R in neurons and oligodendrocytes occurred in the later stages (Grygorowicz et al., 2010; Matute et al., 2007). Interestingly, the total P2X7R expression levels remained high in the brain of EAE rats even after 20 days after immunization and are found to correlate with

increased levels of glial fibrillary acidic protein (GAPF), a marker of astrocyte activation (Holley et al., 2003; Grygorowicz et al., 2011). This could suggest a role for P2X7R activation in the sustained astrocytosis that is observed in the later stages of EAE and MS. A study using brain samples from MS patients has shown upregulation of P2X7R on astrocytes in the parenchyma of the frontal cortex of SPMS patients, validating the findings from EAE models (Amadio et al., 2017). Moreover, the administration of P2X7R antagonist to EAE rats has been found to decrease astrocytosis, reduce demyelination and improve neurological symptoms (Matute et al., 2007; Grygorowicz et al., 2016). These findings reinforce the role of P2X7R in EAE and possibly as a contributing factor in MS pathology.

5. Vitamin D

Vitamin D deficiency has been implicated in the onset, susceptibility, relapse frequency as well as disability and disease progression in MS (Oliveira et al., 2017; Kocovska et al., 2017; Fyfe, 2016; Breuer et al., 2018; Salzer et al., 2014). Epidemiological data supports a correlative link between an insufficiency in vitamin D and/or sunlight exposure in the early stages of life with an increased risk in developing MS during adulthood (Balbuena et al., 2016; Dobson et al., 2013). Prospective epidemiological studies have further established vitamin D as a protective factor associated with MS risk; low serum vitamin D levels have been found to correlate with an increased risk in developing MS (Munger et al., 2006; Salzer et al., 2012).

5.1. Overview

Vitamin D has two major forms: the circulating form, 25(OH) D_3 and its active form, 1,25(OH) $_2D_3$ (Jones, 2008). 25(OH) D_3 circulates at a concentration of 25–200nmol/L in the serum and has a half-life of 15 days, whereas 1,25(OH) $_2D_3$ only has a short half-life of 15 hours (Jones, 2008). The synthesis pathway for vitamin D obtained from both diet and synthesis using ultraviolet (UV) light is shown in Fig. 2.

1,25(OH) $_2D_3$ is a ligand for VDR, a member of the nuclear receptor superfamily (Carlberg, 1996). Because the binding of VDR to vitamin D response element (VDRE) leads to structural changes in the chromatin,

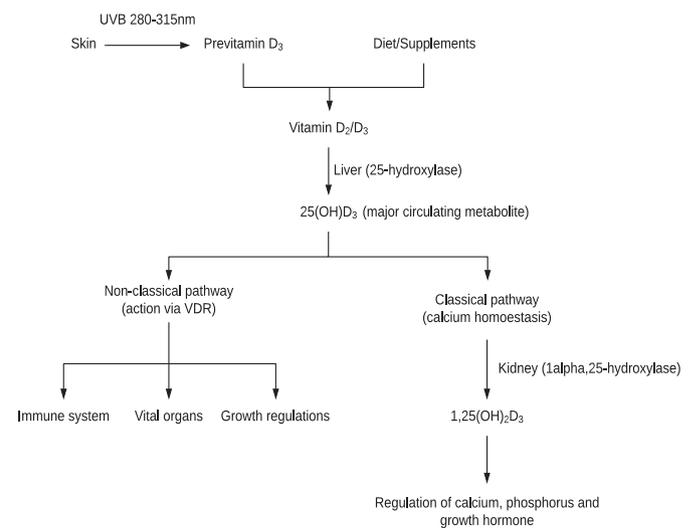


Fig. 2. Synthesis of vitamin D and its main target tissue. Vitamin D $_2$ and D $_3$ have no biological activity without a two-step hydroxylation process. The first step occurs in the liver, which requires 25-hydroxylase to convert the biological precursors, vitamin D $_2$ (ergocalciferol) and D $_3$ (cholecalciferol) to 25(OH)D $_3$. The second step modifies 25(OH)D $_3$ to its active metabolite, 1,25(OH) $_2$ D $_3$ via 1,25-hydroxylase in the kidneys. Physiological actions of 1,25(OH) $_2$ D $_3$ are mediated through its interactions with vitamin D receptors (VDRs) expressed in the nucleus of many different cell types.

vitamin D can regulate gene expression by facilitating gene transcription through interactions with VDR (Zhang et al., 2012). Hence, vitamin D has been found to have pleiotropic physiological functions; it not only modulates calcium and phosphorus homeostasis but is also involved in innate and adaptive immunity (Nurminen et al., 2018).

5.2. Role of vitamin D in immune function/immunity

Vitamin D is well known for its role in calcium and bone homeostasis (Gong et al., 2018; Thacher et al., 2014). However, recent studies have started to focus on the functions of vitamin D in both innate and adaptive immunity (Adams and Hewison, 2008). VDRs are expressed on most immune cell types (Provvedini et al., 1983; Brennan et al., 1987; Veldman et al., 2000) and the level of expression is found to increase with $1,25(\text{OH})_2\text{D}_3$ treatment (Veldman et al., 2000). In addition, a study also observed that African-American individuals (known to have increased susceptibility to tuberculosis) have lower levels of $25(\text{OH})\text{D}_3$ in their sera and were inefficient in the induction of antimicrobial peptide cathelicidin as well as the killing of intracellular *Mycobacterium tuberculosis* (Liu et al., 2006). These data suggest the vitamin D level variabilities might contribute to the differences in microbial infection susceptibility among human populations. This could partly be explained by the role of $1,25(\text{OH})_2\text{D}_3$ in phenotypic differentiation of immature myeloid leukemia cells (Abe et al., 1981; Amento et al., 1984; Mangelsdorf et al., 1984; Miyaura et al., 1981). Thus, a deficiency in vitamin D could lead to ineffective development of these cells, resulting in altered antimicrobial responses to pathogens. Other than immune cell maturation, vitamin D also plays a role in maintaining immune cell responses and function during infections (Djukic et al., 2014). Vitamin D deficiency ($< 15 \text{ ng/ml}$) has been reported to reduce the rate of microglial intracellular killing of pathogens and phagocytosis by decreasing the amount of cytokines such as $\text{TNF}\alpha$ and IL-6 released upon stimulation (Djukic et al., 2014). Moreover, studies in human monocytes have shown that a level of serum vitamin D above 30 ng/ml (in the physiological range) is important in maintaining optimal anti-inflammatory responses (Zhang et al., 2012). Vitamin D deficiency could lead to proinflammatory states due to compromised suppression in cytokine production such as IL-6 and $\text{TNF}\alpha$ by monocytes (Zhang et al., 2012). The function of these inhibitory signals by $1,25(\text{OH})_2\text{D}_3$ is further supported by their role in homeostatic control on dendritic cell development and function (Hewison et al., 2003). It has been suggested that the negative feedback mediated by $1,25(\text{OH})_2\text{D}_3$ could potentially contribute to the maintenance of peripheral tolerance of self-antigens via VDR-dependent mechanisms (Griffin et al., 2001; Hewison et al., 2003). VDRs are expressed in lymphocytes, and vitamin D could therefore regulate adaptive immune system responses (Provvedini et al., 1983; Nunn et al., 1986). Studies have shown that the effects of vitamin D on T-cells depend on the T-cell subtype; for instance, the ability of $1,25(\text{OH})_2\text{D}_3$ to suppress or enhance T-cell proliferation depends on the subtype that is being treated (Hewison, 2012). Although numerous studies have demonstrated the role of vitamin D in immunoregulation, it is important to note that many of these studies are circumstantial or association studies. Moreover, supplementation studies in humans have shown contradictory results; findings from these studies were not consistent in the demonstration of the immune-regulatory effects of vitamin D (Allen et al., 2012; Prietl et al., 2014; Sotirchos et al., 2016; Muris et al., 2016; Berlanga-Taylor et al., 2018). As suggested by Damoiseaux and Smolders, because peripheral blood are frequently used in human studies, the heterogeneity of these cell fractions under homeostatic conditions could be responsible for the discrepancies found across results from different studies (Damoiseaux and Smolders, 2018). Perhaps using a disease-specific cell or biomarker to demonstrate or measure the effects of vitamin D supplementation on immunity may yield more consistent results across different human in vivo studies.

5.3. Role of vitamin D in MS

According to current literature, vitamin D and its interaction with the immune system are implicated in the regulation of clinical disease activity in MS. This is suggested by the lower serum $25(\text{OH})\text{D}_3$ levels during relapse than remission in MS patients (Soilu-Hanninen et al., 2005; Correale et al., 2009). Moreover, higher levels of serum $25(\text{OH})\text{D}_3$ are also found to have a protective effect on MS risk such as a reduction in CNS lesions and relapse rate (Ascherio et al., 2014; Simpson Jr. et al., 2010). The exact mechanism of action of vitamin D in MS remains elusive, but it is likely due to its immunomodulatory effects on CNS inflammation (Lu et al., 2018). Numerous studies have documented the role of vitamin D in immune responses, in particular its association with reduced frequency in differentiation of IL-17/IL-22 producing T-cell subtypes in humans (Allen et al., 2012; Sommer and Fabri, 2015), which are implicated in MS pathology. In addition, the potential benefits of vitamin D supplementation in MS are also shown by studies in human regulatory-like T-cells (da Costa et al., 2016) and B-cells (Haas et al., 2016). The results from these studies suggest that the active form of vitamin D could reduce relapse risk by attenuating B-cell immunoreactivity and limit pathogenic T-cell responses through enhancing classical or non-classical regulatory T-cells (da Costa et al., 2016; Haas et al., 2016). Given previous research implicating vitamin D level and MS predisposition, it is likely that vitamin D deficiency plays a role in several key pathophysiological processes in MS, such as neuroinflammation, demyelination, axonal damage and remyelination (Smolders et al., 2011). Although numerous findings have demonstrated the immunomodulatory actions of vitamin D on the CNS, the exact mechanisms remain inconclusive. Inconsistency across results obtained from human studies has also shed light on the complexity and diversity of the effects of vitamin D on immune regulation. Assessment of MS-specific biomarkers or cells could be more relevant in revealing the effects of vitamin D supplementation on MS. But it is promising that a reduction in anti-Epstein-Barr virus nuclear antigen-1 (anti-EBNA-1) antibody levels in RRMS patients on vitamin D supplementation is a very consistent finding (Damoiseaux and Smolders, 2018; Rolf et al., 2018). It is also important to note that solid evidence on the effects of vitamin D status in clinical course or disability progression in MS is still lacking. There is a growing consensus on the beneficial actions of vitamin D on the inflammatory component of MS, but more studies are still required to assess its effects on the progressive degenerative component in MS pathology (Pierrot-Deseilligny and Souberbielle, 2017).

5.4. Effects of vitamin D on P2X7R

Studies on the modulation of P2X7R expression and function by vitamin D on cells of the immune and nervous systems are lacking. There are some studies on human mononuclear cells that have demonstrated the inhibitory effects of vitamin D on P2X7R expression, thus reducing P2X7R-mediated proinflammatory actions on immune cells (Lajdova et al., 2016). Moreover, because the activation of P2X7R leads to the secretion of pro-inflammatory cytokines and chemokines such as IL-1, IL-6, $\text{TNF}\alpha$ and C-C motif chemokine ligand 2 (CCL2) (Englezou et al., 2015; Shieh et al., 2014), the inhibitory effects of vitamin D on P2X7R reduce the inflammatory responses promoted by these inflammatory factors. Currently, the mechanism of the anti-inflammatory effects of vitamin D on P2X7R is largely undefined, but it has been shown that vitamin D prevents calcium influx through inhibiting P2X7R pore permeation (Lajdova et al., 2008). Due to the role of calcium signaling in activation and proliferation of T lymphocytes as well as innate immune cells such as microglia and monocytes (Brignall et al., 2017; Kotturi et al., 2003; Lewis, 2001; Saul et al., 2016; Tvrdik and Kalani, 2017), the inhibitory effects of vitamin D on P2X7R could reduce the activation of adaptive and innate immune cells through modulating calcium influx (Lajdova et al., 2016). Although the beneficial effects of vitamin D on MS risk have been shown in some studies,

its mechanisms on the predominant immune cells in MS such as T lymphocytes, monocytes and microglial cells remain unclear. Thus further studies are needed to explore the mechanisms and therapeutic benefits of vitamin D in MS through modulation of P2X7R expressed on immune cells.

5.5. Effects of vitamin D and dexamethasone on P2X7R

The active metabolite of vitamin D, 1,25(OH)₂D₃ is considered as a true steroid hormone due to its origin from cholesterol (D hormone) (Cutolo et al., 2011), and therefore like glucocorticoids, it exerts immunomodulatory activities. Because of an overlap in their immunomodulatory actions, vitamin D is found to exert synergistic effects with glucocorticoids (Cutolo et al., 2014). A study has shown that 1,25(OH)₂D₃ has additive effects on the inhibition of lymphocyte proliferation when combined with dexamethasone (Jirapongsananuruk et al., 2000), suggesting that vitamin D supplementation in conjunction with glucocorticoids during MS acute exacerbation could enhance the therapeutic benefits of the glucocorticoid treatment. In addition, 1,25(OH)₂D₃ is also found to have additive effects on dexamethasone-mediated inhibition of Th1 cytokine production, in particular IFN-γ (Jirapongsananuruk et al., 2000), which is implicated in MS pathogenesis. While vitamin D seems to enhance the inhibitory actions of dexamethasone, high doses of dexamethasone are shown to decrease the expression of CYP27B1, which is required for local 1,25(OH)₂D₃ production in the brain (prefrontal cortex and hippocampus) (Jiang et al., 2014). This suggests a possibility that glucocorticoids could influence the local production of 1,25(OH)₂D₃ and disrupt vitamin D activation and signaling in the brain (Jiang et al., 2014). Thus, the effects of vitamin D and glucocorticoids on each other need to be further explored to optimize glucocorticoid treatments during MS acute exacerbation. Currently the literature on the effects of dexamethasone and vitamin D on P2X7R is scarce, but dexamethasone alone is found to inhibit P2X7R expression and function in immune cells, in particular mast cells (Yoshida et al., 2017). Taking into account the synergism between vitamin D and dexamethasone on immunity, these observations raise an intriguing possibility that vitamin D may enhance the anti-inflammatory actions of glucocorticoids on P2X7R in immune cells, providing a potential explanation for the additive effects of 1,25(OH)₂D₃ on dexamethasone-mediated anti-inflammatory responses. In addition, because most in vitro vitamin D studies frequently use the active form of vitamin D, 1,25(OH)₂D₃ and vitamin D supplementation is often in the form of vitamin D₂ or D₃, the exact mechanism of vitamin D supplementation on glucocorticoids remains uncertain.

6. Conclusions

MS is a chronic neuroinflammatory disease of the CNS with unknown etiology. The mechanisms of MS pathology are not well-defined, but it is known that they involve complex interactions between systems and cell types, including T lymphocytes, monocytes, microglial cells and neurons, which lead to focal lesions and neuroinflammatory demyelination in the brain and spinal cord. Interestingly, even though P2X7R is expressed on cells of both innate and adaptive immunity as well as peripheral and centrally derived cells, its exact role in MS is yet to be determined. Furthermore, the mechanisms of vitamin D on P2X7R expressed on immune cells need to be investigated as current findings suggest their possible roles in MS risk and disease activity.

Declarations of interest

None

References

Abe, E., Miyaura, C., Sakagami, H., Takeda, M., Konno, K., Yamazaki, T., Yoshiki, S.,

- Suda, T., 1981. Differentiation of mouse myeloid leukemia cells induced by 1 alpha,25-dihydroxyvitamin D₃. *Proc Natl Acad Sci U S A* 78 (8), 4990–4994.
- Achuthan, A., Aslam, A.S.M., Nguyen, Q., Lam, P.Y., Fleetwood, A.J., Frye, A.T., Louis, C., Lee, M.C., Smith, J.E., Cook, A.D., Olshansky, M., Turner, S.J., Hamilton, J.A., 2018. Glucocorticoids promote apoptosis of proinflammatory monocytes by inhibiting ERK activity. *Cell Death Dis* 9 (3), 267.
- Adams, J.S., Hewison, M., 2008. Unexpected actions of vitamin D: new perspectives on the regulation of innate and adaptive immunity. *Nat Clin Pract Endocrinol Metab* 4 (2), 80–90.
- Aga, M., Johnson, C.J., Hart, A.P., Guadarrama, A.G., Suresh, M., Svaren, J., Bertics, P.J., Darien, B.J., 2002. Modulation of monocyte signaling and pore formation in response to agonists of the nucleotide receptor P2X(7). *J. Leukoc Biol* 72 (1), 222–232.
- Ajami, B., Bennett, J.L., Krieger, C., McNagny, K.M., Rossi, F.M., 2011. Infiltrating monocytes trigger EAE progression, but do not contribute to the resident microglia pool. *Nat Neurosci* 14 (9), 1142–1149.
- Allen, A.C., Kelly, S., Basdeo, S.A., Kinsella, K., Mulready, K.J., Mills, K.H., Tubridy, N., Walsh, C., Brady, J.J., Hutchinson, M., Fletcher, J.M., 2012. A pilot study of the immunological effects of high-dose vitamin D in healthy volunteers. *Mult Scler* 18 (12), 1797–1800.
- Alves, L.A., de Melo Reis, R.A., de Souza, C.A., de Freitas, M.S., Teixeira, P.C., Neto Moreira Ferreira, D., Xavier, R.F., 2014. The P2X7 receptor: shifting from a low- to a high-conductance channel - an enigmatic phenomenon? *Biochim Biophys Acta* 1838 (10), 2578–2587.
- Amadio, S., Parisi, C., Piras, E., Fabbriozzi, P., Apolloni, S., Montilli, C., Luchetti, S., Ruggieri, S., Gasperini, C., Laghi-Pasini, F., Battistini, L., Volonte, C., 2017. Modulation of P2X7 Receptor during Inflammation in Multiple Sclerosis. *Front Immunol* 8 (1529).
- Amento, E.P., Bhalla, A.K., Kurnick, J.T., Kradin, R.L., Clemens, T.L., Holick, S.A., Holick, M.F., Krane, S.M., 1984. 1 alpha,25-dihydroxyvitamin D₃ induces maturation of the human monocyte cell line U937, and, in association with a factor from human T lymphocytes, augments production of the monokine, mononuclear cell factor. *J. Clin Invest* 73 (3), 731–739.
- Aravalli, R.N., Peterson, P.K., Lokensgard, J.R., 2007. Toll-like receptors in defense and damage of the central nervous system. *J. Neuroimmune Pharmacol* 2 (4), 297–312.
- Arrambide, G., Tintore, M., Espejo, C., Auger, C., Castillo, M., Rio, J., Castillo, J., Vidal-Jordana, A., Galan, I., Nos, C., Mitjana, R., Mulero, P., de Barros, A., Rodriguez-Acevedo, B., Midaglia, L., Sastre-Garriga, J., Rovira, A., Comabella, M., Montalban, X., 2018. The value of oligoclonal bands in the multiple sclerosis diagnostic criteria. *Brain*. 141 (4), 1075–1084.
- Ascherio, A., Munger, K.L., Simon, K.C., 2010. Vitamin D and multiple sclerosis. *Lancet Neurol* 9 (6), 599–612.
- Ascherio, A., Munger, K.L., Lunemann, J.D., 2012. The initiation and prevention of multiple sclerosis. *Nat Rev Neurol* 8 (11), 602–612.
- Ascherio, A., Munger, K.L., White, R., Kochert, K., Simon, K.C., Polman, C.H., Freedman, M.S., Hartung, H.P., Miller, D.H., Montalban, X., Edan, G., Barkhof, F., Pleimes, D., Radu, E.W., Sandbrink, R., Kappos, L., Pohl, C., 2014. Vitamin D as an early predictor of multiple sclerosis activity and progression. *JAMA Neurol* 71 (3), 306–314.
- Balbuena, L.D., Middleton, R.M., Tuite-Dalton, K., Pouliou, T., Williams, K.E., Noble, G.J., 2016. Sunshine, Sea, and Season of Birth: MS Incidence in Wales. *PLoS One* 11 (5), e0155181.
- Bauer, J., Sminia, T., Wouterlood, F.G., Dijkstra, C.D., 1994. Phagocytic activity of macrophages and microglial cells during the course of acute and chronic relapsing experimental autoimmune encephalomyelitis. *J. Neurosci Res* 38 (4), 365–375.
- Baufeld, C., O'Loughlin, E., Calcagno, N., Madore, C., Butovsky, O., 2017. Differential contribution of microglia and monocytes in neurodegenerative diseases. *J. Neural Transm (Vienna)*. 125 (5), 809–826.
- Baughman, E.J., Mendoza, J.P., Ortega, S.B., Ayers, C.L., Greenberg, B.M., Frohman, E.M., Karandikar, N.J., 2011. Neuroantigen-specific CD8+ regulatory T-cell function is deficient during acute exacerbation of multiple sclerosis. *J. Autoimmun* 36 (2), 115–124.
- Beecham, A.H., Patsopoulos, N.A., Xifara, D.K., Davis, M.F., Kempainen, A., Cotsapas, C., Shah, T.S., Spencer, C., Booth, D., Goris, A., Oturai, A., Saarela, J., Fontaine, B., Hemmer, B., Martin, C., Zipp, F., D'Alfonso, S., Martinelli-Boneschi, F., Taylor, B., Harbo, H.F., Kockum, I., Hillert, J., Olsson, T., Ban, M., Oksenberg, J.R., Hintzen, R., Barcellos, L.F., Agliardi, C., Alfredsson, L., Alizadeh, M., Anderson, C., Andrews, R., Sondergaard, H.B., Baker, A., Band, G., Baranzini, S.E., Barizzone, N., Barrett, J., Bellenguez, C., Bergamaschi, L., Bernardinelli, L., Berthele, A., Biberacher, V., Binder, T.M., Blackburn, H., Bomfim, I.L., Brambilla, P., Broadley, S., Brochet, B., Brundin, L., Buck, D., Butzkueven, H., Caillier, S.J., Camu, W., Carpentier, W., Cavalla, P., Celius, E.G., Coman, I., Comi, G., Corrado, L., Cosemann, L., Cournu-Rebeix, I., Cree, B.A., Cusi, D., Damotte, V., Defer, G., Delgado, S.R., Deloukas, P., di Sapio, A., Dilthey, A.T., Donnelly, P., Dubois, B., Duddy, M., Edkins, S., Elovaara, I., Esposito, F., Evangelou, N., Fiddes, B., Field, J., Franke, A., Freeman, C., Frohlich, I.Y., Galimberti, D., Gieger, C., Gourraud, P.A., Graetz, C., Graham, A., Grummel, V., Guaschino, C., Hadjixenofontos, A., Hakonarson, H., Halfpenny, C., Hall, G., Hall, P., Hamsten, A., Harley, J., Harrower, T., Hawkins, C., Hellenthal, G., Hillier, C., et al., 2013. Analysis of immune-related loci identifies 48 new susceptibility variants for multiple sclerosis. *Nat Genet* 45 (11), 1353–1360.
- Benveniste, E.N., 1997. Cytokines: influence on glial cell gene expression and function. *Chem Immunol* 69, 31–75.
- Berkovich, R., 2013. Treatment of acute relapses in multiple sclerosis. *Neurotherapeutics* 10 (1), 97–105.
- Berlanga-Taylor, A.J., Plant, K., Dahl, A., Lau, E., Hill, M., Sims, D., Heger, A., Emberson, J., Armitage, J., Clarke, R., Knight, J.C., 2018. Genomic response to vitamin D supplementation in the setting of a randomized, Placebo-Controlled Trial. *EBioMedicine* 31, 133–142.

- Brennan, A., Katz, D.R., Nunn, J.D., Barker, S., Hewison, M., Fraher, L.J., O'Riordan, J.L., 1987. Dendritic cells from human tissues express receptors for the immunoregulatory vitamin D3 metabolite, dihydroxycholecalciferol. *Immunology* 61 (4), 457–461.
- Breuer, J., Loser, K., Mykicik, N., Wiendl, H., Schwab, N., 2018. Does the environment influence multiple sclerosis pathogenesis via UVB light and/or induction of vitamin D? *J. Neuroimmunol* S0165-5728 (17), 30478–30572.
- Brignall, R., Cauchy, P., Bevington, S.L., Gorman, B., Pisco, A.O., Bagnall, J., Boddington, C., Rowe, W., England, H., Rich, K., Schmidt, L., Dyer, N.P., Travis, M.A., Ott, S., Jackson, D.A., Cockerill, P.N., Paszek, P., 2017. Integration of kinase and calcium signaling at the level of chromatin underlies inducible gene activation in T cells. *J. Immunol* 199 (8), 2652–2667.
- Brodsky, M., Nazarian, S., Orengo-Nania, S., Hutton, G.J., Buckley, E.G., Massey, E.W., Bhatti, M.T., Greer, M., Goodwin, J., Wall, M., Savino, P.J., Leist, T., Miller, N.R., Irani, D., Trobe, J.D., Cornblath, W., Kaufman, D.I., Eggenberger, E., Kupersmith, M.J., Shults, W.T., McAllister, L., Hamilton, S., Beck, R.W., Dontchev, M., Gal, R.L., Kollman, C., Keltner, J.L., Smith, C.H., 2008. Multiple sclerosis risk after optic neuritis: final optic neuritis treatment trial follow-up. *Arch Neurol* 65 (6), 727–732.
- Bruck, W., Porada, P., Poser, S., Rieckmann, P., Hanefeld, F., Kretschmar, H.A., Lassmann, H., 1995. Monocyte/macrophage differentiation in early multiple sclerosis lesions. *Ann Neurol* 38 (5), 788–796.
- Bsibsi, M., Ravid, R., Gveric, D., van Noort, J.M., 2002. Broad expression of Toll-like receptors in the human central nervous system. *J. Neuropathol Exp Neurol* 61 (11), 1013–1021.
- Butovsky, O., Jedrychowski, M.P., Moore, C.S., Cialic, R., Lanser, A.J., Gabrieli, G., Koelsperger, T., Dake, B., Wu, P.M., Doykan, C.E., Fanek, Z., Liu, L., Chen, Z., Rothstein, J.D., Ransohoff, R.M., Gygi, S.P., Antel, J.P., Weiner, H.L., 2014. Identification of a unique TGF-beta-dependent molecular and functional signature in microglia. *Nat Neurosci* 17 (1), 131–143.
- Caragnano, M., Tortorella, P., Bergami, A., Ruggieri, M., Livrea, P., Specchio, L.M., Martino, G., Trojano, M., Furlan, R., Avolio, C., 2012. Monocytes P2X7 purinergic receptor is modulated by glatiramer acetate in multiple sclerosis. *J. Neuroimmunol* 245 (1–2), 93–97.
- Carlberg, C., 1996. The vitamin D(3) receptor in the context of the nuclear receptor superfamily: The central role of the retinoid X receptor. *Endocrine* 4 (2), 91–105.
- Chuluundorj, D., Harding, S.A., Abernethy, D., La Flamme, A.C., 2014. Expansion and preferential activation of the CD14(+)CD16(+) monocyte subset during multiple sclerosis. *Immunol Cell Biol* 92 (6), 509–517.
- Colombo, M., Dono, M., Gazzola, P., Roncella, S., Valetto, A., Chiorazzi, N., Mancardi, G.L., Ferrarini, M., 2000. Accumulation of clonally related B lymphocytes in the cerebrospinal fluid of multiple sclerosis patients. *J. Immunol* 164 (5), 2782–2789.
- Correa, J., Villa, A., 2010. Role of CD8+ CD25+ Foxp3+ regulatory T cells in multiple sclerosis. *Ann Neurol* 67 (5), 625–638.
- Correale, J., Ysraelit, M.C., Gaitan, M.I., 2009. Immunomodulatory effects of Vitamin D in multiple sclerosis. *Brain* 132 (Pt 5), 1146–1160.
- Cunnusamy, K., Baughman, E.J., Franco, J., Ortega, S.B., Sinha, S., Chaudhary, P., Greenberg, B.M., Frohman, E.M., Karandikar, N.J., 2014. Disease exacerbation of multiple sclerosis is characterized by loss of terminally differentiated autoregulatory CD8+ T cells. *Clin Immunol* 152 (1–2), 115–126.
- Cutolo, M., Pizzorni, C., Sulli, A., 2011. Vitamin D endocrine system involvement in autoimmune rheumatic diseases. *Autoimmun Rev* 11 (2), 84–87.
- Cutolo, M., Paolino, S., Sulli, A., Smith, V., Pizzorni, C., Seriolo, B., 2014. Vitamin D, steroid hormones, and autoimmunity. *Ann N Y Acad Sci* 1317, 39–46.
- da Costa, D.S., Hygino, J., Ferreira, T.B., Kasahara, T.M., Barros, P.O., Monteiro, C., Oliveira, A., Tavares, F., Vasconcelos, C.C., Alvarenga, R., Bento, C.A., 2016. Vitamin D modulates different IL-17-secreting T cell subsets in multiple sclerosis patients. *J. Neuroimmunol* 299, 8–18.
- Damoiseaux, J., Smolders, J., 2018. The engagement between vitamin D and the immune system: is consolidation by a marriage to be expected? *EBioMedicine* 31, 9–10.
- Dean, G., 1967. Annual incidence, prevalence, and mortality of multiple sclerosis in white South-African-born and in white immigrants to South Africa. *British Medical Journal* 2 (5554), 724–730.
- Deeks, E.D., 2018. Cladribine tablets: a review in relapsing MS. *CNS Drugs* 32 (8), 785–796.
- Denney, L., Kok, W.L., Cole, S.L., Sanderson, S., McMichael, A.J., Ho, L.P., 2012. Activation of invariant NKT cells in early phase of experimental autoimmune encephalomyelitis results in differentiation of Ly6Chi inflammatory monocyte to M2 macrophages and improved outcome. *J. Immunol* 189 (2), 551–557.
- DeRijk, R.H., Eskandari, F., Sternberg, E.M., 2004. Corticosteroid resistance in a sub-population of multiple sclerosis patients as measured by ex vivo dexamethasone inhibition of LPS induced IL-6 production. *J. Neuroimmunol* 151 (1–2), 180–188.
- Di Virgilio, F., Vuerich, M., 2015. Purinergic signaling in the immune system. *Auton Neurosci* 191, 117–123.
- Di Virgilio, F., Dal Ben, D., Sarti, A.C., Giuliani, A.L., Falzoni, S., 2017. The P2X7 receptor in infection and inflammation. *Immunity* 47 (1), 15–31.
- Di Virgilio, F., Giuliani, A.L., Vultaggio-Poma, V., Falzoni, S., Sarti, A.C., 2018a. Non-nucleotide agonists triggering P2X7 receptor activation and pore formation. *Front Pharmacol* 9 (39).
- Di Virgilio, F., Sarti, A.C., Grassi, F., 2018b. Modulation of innate and adaptive immunity by P2X ion channels. *Curr Opin Immunol* 52, 51–59.
- Djukic, M., Onken, M.L., Schutze, S., Redlich, S., Gotz, A., Hanisch, U.K., Bertsch, T., Ribes, S., Hanenberg, A., Schneider, S., Bollheimer, C., Sieber, C., Nau, R., 2014. Vitamin D deficiency reduces the immune response, phagocytosis rate, and intracellular killing rate of microglial cells. *Infect Immun* 82 (6), 2585–2594.
- Dobson, R., Giovannoni, G., Ramagopalan, S., 2013. The month of birth effect in multiple sclerosis: systematic review, meta-analysis and effect of latitude. *J. Neurol Neurosurg Psychiatry* 84 (4), 427–432.
- Elsner, A., Duncan, M., Gavrilin, M., Wewers, M.D., 2004. A novel P2X7 receptor activator, the human cathelicidin-derived peptide LL37, induces IL-1 beta processing and release. *J. Immunol* 172 (8), 4987–4994.
- Englezou, P.C., Rothwell, S.W., Ainscough, J.S., Brough, D., Landsiedel, R., Verkhratsky, A., Kimber, I., Dearman, R.J., 2015. P2X7R activation drives distinct IL-1 responses in dendritic cells compared to macrophages. *Cytokine* 74 (2), 293–304.
- Fadda, G., Brown, R.A., Longoni, G., Castro, D.A., O'Mahony, J., Verhey, L.H., Branson, H.M., Waters, P., Bar-Or, A., Marrie, R.A., Yeh, E.A., Narayanan, S., Arnold, D.L., Banwell, B., Wambara, K., Connolly, M.B., Yager, J., Mah, J.K., Sebire, G., Callen, D., Meaney, B., Dilenge, M.E., Lortie, A., Pohl, D., Doja, A., Venkateswaran, S., Levin, S., MacDonald, E.A., Meek, D., Wood, E., Buckley, D., Awuku, M., Baird, J.B., Bhan, V., Arnaoutelis, R., Nandamalavan, D., 2018. MRI and laboratory features and the performance of international criteria in the diagnosis of multiple sclerosis in children and adolescents: a prospective cohort study. *Lancet Child Adolesc Health* 2 (3), 191–204.
- Faroni, A., Smith, R.J., Procacci, P., Castelnuovo, L.F., Puccianti, E., Reid, A.J., Magnaghi, V., Verkhratsky, A., 2014. Purinergic signaling mediated by P2X7 receptors controls myelination in sciatic nerves. *J. Neurosci Res* 92 (10), 1259–1269.
- Ferrari, D., Pizzirani, C., Adinolfi, E., Forchap, S., Sitta, B., Turchet, L., Falzoni, S., Minelli, M., Baricordi, R., Di Virgilio, F., 2004. The antibiotic polymyxin B modulates P2X7 receptor function. *J. Immunol* 173 (7), 4652–4660.
- Fiedler, S.E., George, J.D., Love, H.N., Kim, E., Spain, R., Bourdette, D., Salinithone, S., 2017. Analysis of IL-6, IL-1beta and TNF-alpha production in monocytes isolated from multiple sclerosis patients treated with disease modifying drugs. *J. Syst Integr Neurosci* 3 (3).
- Ford, A.L., Goodsall, A.L., Hickey, W.F., Sedgwick, J.D., 1995. Normal adult ramified microglia separated from other central nervous system macrophages by flow cytometric sorting. Phenotypic differences defined and direct ex vivo antigen presentation to myelin basic protein-reactive CD4+ T cells compared. *J. Immunol* 154 (9), 4309–4321.
- Fraussen, J., Claes, N., Van Wijmeersch, B., van Horsen, J., Stinissen, P., Hupperts, R., Somers, V., 2016. B cells of multiple sclerosis patients induce autoreactive proinflammatory T cell responses. *Clin Immunol* 173, 124–132.
- Frohman, E.M., Shah, A., Eggenberger, E., Metz, L., Zivadinov, R., Stuve, O., 2007. Corticosteroids for multiple sclerosis: I. Application for treating exacerbations. *Neurotherapeutics* 4 (4), 618–626.
- Fyfe, I., 2016. Multiple sclerosis: Vitamin D deficiency leads to excessive B-cell responses in multiple sclerosis. *Nat. Rev Neurol* 12 (5), 252.
- Gandhi, R., Laroni, A., Weiner, H.L., 2010. Role of the innate immune system in the pathogenesis of multiple sclerosis. *J. Neuroimmunol* 221 (1–2), 7–14.
- Garg, N., Smith, T.W., 2015. An update on immunopathogenesis, diagnosis, and treatment of multiple sclerosis. *Brain Behav* 5 (9), e00362.
- Ghaffar, O., Feinstein, A., 2007. The neuropsychiatry of multiple sclerosis: A review of recent developments. *Current Opinion in Psychiatry* 20 (3), 278–285.
- Ghiringhelli, F., Apetoh, L., Tesnière, A., Ayméric, L., Ma, Y., Ortiz, C., Vermaelen, K., Panaretakis, T., Mignot, G., Ullrich, E., Perfettini, J.L., Schlemmer, F., Tasdemir, E., Uhl, M., Genin, P., Civas, A., Ryffel, B., Kanellopoulos, J., Tschopp, J., Andre, F., Lidereau, R., McLaughlin, N.M., Haynes, N.M., Smyth, M.J., Kroemer, G., Zitvogel, L., 2009. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1beta-dependent adaptive immunity against tumors. *Nat Med* 15 (10), 1170–1178.
- Gjelstrup, M.C., Stilund, M., Petersen, T., Møller, H.J., Petersen, E.L., Christensen, T., 2018. Subsets of activated monocytes and markers of inflammation in incipient and progressed multiple sclerosis. *Immunol Cell Biol* 96 (2), 160–174.
- Goldenberg, M.M., 2012. Multiple Sclerosis Review. *Pharmacy and Therapeutics* 37 (3), 175–184.
- Gong, A., Chen, J., Wu, J., Li, J., Wang, L., Goltzman, D., Miao, D., 2018. 1,25-Dihydroxyvitamin D deficiency accelerates alveolar bone loss independent of aging and extracellular calcium and phosphorus. *J. Periodontol* 89 (8), 983–994.
- Gonsette, R.E., 2012. Self-tolerance in multiple sclerosis. *Acta Neurol Belg* 112 (2), 133–140.
- Goodin, D.S., 2014. Glucocorticoid treatment of multiple sclerosis. *Handb Clin Neurol* 122, 455–464.
- Gray, E., Thomas, T.L., Betmouni, S., Scolding, N., Love, S., 2008. Elevated activity and microglial expression of myeloperoxidase in demyelinated cerebral cortex in multiple sclerosis. *Brain Pathol* 18 (1), 86–95.
- Griffin, M.D., Lutz, W., Phan, V.A., Bachman, L.A., McKean, D.J., Kumar, R., 2001. Dendritic cell modulation by 1alpha,25 dihydroxyvitamin D3 and its analogs: a vitamin D receptor-dependent pathway that promotes a persistent state of immaturity in vitro and in vivo. *Proc Natl Acad Sci U S A* 98 (12), 6800–6805.
- Grygorowicz, T., Struzynska, L., Sulkowski, G., Chalimoniuk, M., Sulejczak, D., 2010. Temporal expression of P2X7 purinergic receptor during the course of experimental autoimmune encephalomyelitis. *Neurochem Int* 57 (7), 823–829.
- Grygorowicz, T., Sulejczak, D., Struzynska, L., 2011. Expression of purinergic P2X7 receptor in rat brain during the symptomatic phase of experimental autoimmune encephalomyelitis and after recovery of neurological deficits. *Acta Neurobiol Exp (Wars)* 71 (1), 65–73.
- Grygorowicz, T., Welniak-Kaminska, M., Struzynska, L., 2016. Early P2X7R-related astrogliosis in autoimmune encephalomyelitis. *Mol Cell Neurosci* 74, 1–9.
- Gu, B.J., Wiley, J.S., 2018. P2X7 as a scavenger receptor for innate phagocytosis in the brain. *Br J Pharmacol* 175 (22), 4195–4208.
- Gu, B.J., Zhang, W.Y., Bendall, L.J., Chessell, I.P., Buell, G.N., Wiley, J.S., 2000. Expression of P2X7(7) purinergic receptors on human lymphocytes and monocytes: evidence for nonfunctional P2X7(7) receptors. *Am J Physiol Cell Physiol* 279 (4), C1189–C1197.
- Gudipaty, L., Humphreys, B.D., Buell, G., DUBYAK, G.R., 2001. Regulation of P2X7(7) nucleotide receptor function in human monocytes by extracellular ions and receptor

- density. *Am J Physiol Cell Physiol* 280 (4), C943–C953.
- Haas, J., Schwarz, A., Korporal-Kuhnke, M., Fallner, S., Jarius, S., Wildemann, B., 2016. Hypovitaminosis D upscales B-cell immunoreactivity in multiple sclerosis. *J. Neuroimmunol* 294, 18–26.
- Hartung, H.P., Kieseier, B.C., 2014. The new therapeutic landscape in multiple sclerosis: exciting times and new perspectives. *Curr Opin Neurol* 27 (3), 243–245.
- Hartung, H.P., Aktas, O., Menge, T., Kieseier, B.C., 2014. Immune regulation of multiple sclerosis. *Handb Clin Neurol* 122, 3–14.
- Hemmer, B., Kerschensteiner, M., Korn, T., 2015. Role of the innate and adaptive immune responses in the course of multiple sclerosis. *Lancet Neurol* 14 (4), 406–419.
- Heppner, F.L., Greter, M., Marino, D., Falsig, J., Raivich, G., Hovelmeyer, N., Waisman, A., Rulicke, T., Prinz, M., Priller, J., Becher, B., Aguzzi, A., 2005. Experimental autoimmune encephalomyelitis repressed by microglial paralysis. *Nat Med* 11 (2), 146–152.
- Hernandez-Pedro, N.Y., Espinosa-Ramirez, G., de la Cruz, V.P., Pineda, B., Sotelo, J., 2013. Initial immunopathogenesis of multiple sclerosis: innate immune response. *Clin Dev Immunol* 2013 (413465).
- Hewinson, J., Mackenzie, A.B., 2007. P2X(7) receptor-mediated reactive oxygen and nitrogen species formation: from receptor to generators. *Biochem Soc Trans* 35 (Pt 5), 1168–1170.
- Hewison, M., 2012. Vitamin D and immune function: an overview. *Proc Nutr Soc* 71 (1), 50–61.
- Hewison, M., Freeman, L., Hughes, S.V., Evans, K.N., Bland, R., Eliopoulos, A.G., Kilby, M.D., Moss, P.A., Chakraverty, R., 2003. Differential regulation of vitamin D receptor and its ligand in human monocyte-derived dendritic cells. *J. Immunol* 170 (11), 5382–5390.
- Holley, J.E., Gveric, D., Newcombe, J., Cuzner, M.L., Gutowski, N.J., 2003. Astrocyte characterization in the multiple sclerosis glial scar. *Neuropathol Appl Neurobiol* 29 (5), 434–444.
- Hollingworth, S., Walker, K., Page, A., Eadie, M., 2014. Multiple sclerosis disease modifying medicine utilisation in Australia. *J. Clinical Neurosciences* 21 (12), 2083–2087.
- Imamura, K., Suzumura, A., Hayashi, F., Marunouchi, T., 1993. Cytokine production by peripheral blood monocytes/macrophages in multiple sclerosis patients. *Acta Neurol Scand* 87 (4), 281–285.
- Jiang, L.H., Baldwin, J.M., Roger, S., Baldwin, S.A., 2013. Insights into the molecular mechanisms underlying mammalian P2X7 receptor functions and contributions in diseases, revealed by structural modeling and single nucleotide polymorphisms. *Front Pharmacol* 4 (55).
- Jiang, P., Xue, Y., Li, H.D., Liu, Y.P., Cai, H.L., Tang, M.M., Zhang, L.H., 2014. Dysregulation of vitamin D metabolism in the brain and myocardium of rats following prolonged exposure to dexamethasone. *Psychopharmacology (Berl)* 231 (17), 3445–3451.
- Jindrichova, M., Bhattacharya, A., Rupert, M., Skopek, P., Obsil, T., Zemkova, H., 2015. Functional characterization of mutants in the transmembrane domains of the rat P2X7 receptor that regulate pore conductivity and agonist sensitivity. *J. Neurochem* 133 (6), 815–827.
- Jirapongsananuruk, O., Melamed, I., Leung, D.Y., 2000. Additive immunosuppressive effects of 1,25-dihydroxyvitamin D3 and corticosteroids on TH1, but not TH2, responses. *J. Allergy Clin Immunol* 106 (5), 981–985.
- Johnson, D.B., Bhimji, S.S., 2018. Dexamethasone. In: StatPearls. StatPearls Publishing StatPearls Publishing LLC., Treasure Island (FL).
- Jones, G., 2008. Pharmacokinetics of vitamin D toxicity. *Am J Clin Nutr* 88 (2), 582s–586s.
- Jones, A.P., Kermod, A.G., Lucas, R.M., Carroll, W.M., Nolan, D., Hart, P.H., 2017. Circulating immune cells in multiple sclerosis. *Clin Exp Immunol* 187 (2), 193–203.
- Kappos, L., Li, D., Calabresi, P.A., O'Connor, P., Bar-Or, A., Barkhof, F., Yin, M., Leppert, D., Glanzman, R., Tinbergen, J., Hauser, S.L., 2011. Ocrelizumab in relapsing-remitting multiple sclerosis: a phase 2, randomised, placebo-controlled, multicentre trial. *Lancet* 378 (9805), 1779–1787.
- Kebir, H., Kreymborg, K., Ifergan, I., Dodelet-Devillers, A., Cayrol, R., Bernard, M., Giuliani, F., Arbour, N., Becher, B., Prat, A., 2007. Human TH17 lymphocytes promote blood-brain barrier disruption and central nervous system inflammation. *Nat Med* 13 (10), 1173–1175.
- Kebir, H., Ifergan, I., Alvarez, J.I., Bernard, M., Poirier, J., Arbour, N., Duquette, P., Prat, A., 2009. Preferential recruitment of interferon-gamma-expressing TH17 cells in multiple sclerosis. *Ann Neurol* 66 (3), 390–402.
- Khadra, A., Tomic, M., Yan, Z., Zemkova, H., Sherman, A., Stojilkovic, S.S., 2013. Dual gating mechanism and function of P2X7 receptor channels. *Biophys Journal* 104 (12), 2612–2621.
- Kocovska, E., Gaughan, F., Krivoy, A., Meier, U.C., 2017. Vitamin-D deficiency as a potential environmental risk factor in multiple sclerosis, schizophrenia, and autism. *Front Psychiatry* 8 (47).
- Koriem, K.M.M., 2016. Multiple sclerosis: New insights and trends. *Asian Pacific J Trop Biomed* 6 (5), 429–440.
- Kotturi, M.F., Carlow, D.A., Lee, J.C., Ziltener, H.J., Jefferies, W.A., 2003. Identification and functional characterization of voltage-dependent calcium channels in T lymphocytes. *J Biol Chem* 278 (47), 46949–46960.
- Kutzelnigg, A., Lucchinetti, C.F., Stadelmann, C., Bruck, W., Rauschka, H., Bergmann, M., Schmidbauer, M., Parisi, J.E., Lassmann, H., 2005. Cortical demyelination and diffuse white matter injury in multiple sclerosis. *Brain* 128 (Pt 11), 2705–2712.
- Lajdova, I., Chorvat Jr., D., Chorvatova, A., 2008. Rapid effects of 1 α ,25(OH) $_2$ D $_3$ in resting human peripheral blood mononuclear cells. *Eur J Pharmacol* 586 (1–3), 14–23.
- Lajdova, I., Spustova, V., Oksa, A., Chorvat, D., Marec Chorvatova, A., 2016. MP015 the effect of vitamin D3 supplementation on P2X7 receptor function in early stages of chronic kidney disease. *Nephrol Dial Transplant* 31, i347–i348.
- Lassmann, H., 2013. Pathology and disease mechanisms in different stages of multiple sclerosis. *J. Neurol Sci* 333 (1–2), 1–4.
- Lee, S.J., Lee, S., 2002. Toll-like receptors and inflammation in the CNS. *Curr Drug Targets Inflamm Allergy* 1 (2), 181–191.
- Lee, A.H., Ledderose, C., Li, X., Slubowski, C.J., Sueyoshi, K., Staudenmaier, L., Bao, Y., Zhang, J., Junger, W.G., 2018. Adenosine triphosphate release is required for toll-like receptor-induced monocyte/macrophage activation, inflammasome signaling, interleukin-1 β production, and the host immune response to infection. *Crit Care Med* 46 (12), e1183–e1189.
- Lewis, R.S., 2001. Calcium signaling mechanisms in T lymphocytes. *Annu Rev Immunol* 19, 497–521.
- Li, Y.F., Zhang, S.X., Ma, X.W., Xue, Y.L., Gao, C., Li, X.Y., 2017. Levels of peripheral Th17 cells and serum Th17-related cytokines in patients with multiple sclerosis: A meta-analysis. *Mult Scler Relat Disord* 18, 20–25.
- Liu, P.T., Stenger, S., Li, H., Wenzel, L., Tan, B.H., Krutzik, S.R., Ochoa, M.T., Schauber, J., Wu, K., Meinken, C., Kamen, D.L., Wagner, M., Bals, R., Steinmeyer, A., Zugel, U., Gallo, R.L., Eisenberg, D., Hewison, M., Hollis, B.W., Adams, J.S., Bloom, B.R., Modlin, R.L., 2006. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* 311 (5768), 1770–1773.
- Lu, M., Taylor, B.V., Korner, H., 2018. Genomic effects of the vitamin D receptor: potentially the link between vitamin D, immune cells, and multiple sclerosis. *Front Immunol* 9 (477).
- Lublin, F.D., Reingold, S.C., Cohen, J.A., Cutter, G.R., Sorensen, P.S., Thompson, A.J., Wolinsky, J.S., Balcer, L.J., Banwell, B., Barkhof, F., Bebo Jr., B., Calabresi, P.A., Clanet, M., Comi, G., Fox, R.J., Freedman, M.S., Goodman, A.D., Ingles, M., Kappos, L., Kieseier, B.C., Lincoln, J.A., Lubetzki, C., Miller, A.E., Montalban, X., O'Connor, P.W., Petkau, J., Pozzilli, C., Rudick, R.A., Sormani, M.P., Stuve, O., Waubant, E., Polman, C.H., 2014. Defining the clinical course of multiple sclerosis: the 2013 revisions. *Neurology* 83 (3), 278–286.
- Mangan, D.F., Welch, G.R., Wahl, S.M., 1991. Lipopolysaccharide, tumor necrosis factor- α , and IL-1 β prevent programmed cell death (apoptosis) in human peripheral blood monocytes. *J. Immunol* 146 (5), 1541–1546.
- Mangelsdorf, D.J., Koefler, H.P., Donaldson, C.A., Pike, J.W., Haussler, M.R., 1984. 1,25-Dihydroxyvitamin D $_3$ -induced differentiation in a human promyelocytic leukemia cell line (HL-60): receptor-mediated maturation to macrophage-like cells. *J. Cell Biol* 98 (2), 391–398.
- Marta, M., Andersson, A., Isaksson, M., Kampe, O., Lobell, A., 2008. Unexpected regulatory roles of TLR4 and TLR9 in experimental autoimmune encephalomyelitis. *Eur J. Immunol* 38 (2), 565–575.
- Marta, M., Meier, U.C., Lobell, A., 2009. Regulation of autoimmune encephalomyelitis by toll-like receptors. *Autoimmun Rev* 8 (6), 506–509.
- Matute, C., 2008. P2X7 receptors in oligodendrocytes: a novel target for neuroprotection. *Mol Neurobiol* 38 (2), 123–128.
- Matute, C., Torre, I., Perez-Cerda, F., Perez-Samartin, A., Alberdi, E., Etxebarria, E., Arranz, A.M., Ravid, R., Rodriguez-Antiguedad, A., Sanchez-Gomez, M., Domercq, M., 2007. P2X(7) receptor blockade prevents ATP excitotoxicity in oligodendrocytes and ameliorates experimental autoimmune encephalomyelitis. *J. Neurosci* 27 (35), 9525–9533.
- McColl, A., Michlewska, S., Dransfield, I., Rossi, A.G., 2007. Effects of glucocorticoids on apoptosis and clearance of apoptotic cells. *Sci World J* 7, 1165–1181.
- McDonald, W.I., Compston, A., Edan, G., Goodkin, D., Hartung, H.P., Lublin, F.D., McFarland, H.F., Paty, D.W., Polman, C.H., Reingold, S.C., Sandberg-Wollheim, M., Sibley, W., Thompson, A., van den Noort, S., Weinshenker, B.Y., Wolinsky, J.S., 2001. Recommended diagnostic criteria for multiple sclerosis: guidelines from the International Panel on the diagnosis of multiple sclerosis. *Ann Neurol* 50 (1), 121–127.
- Mikita, J., Dubourdieu-Cassagno, N., Deloire, M.S., Vekris, A., Biran, M., Raffard, G., Brochet, B., Canon, M.H., Franconi, J.M., Boiziau, C., Petry, K.G., 2011. Altered M1/M2 activation patterns of monocytes in severe relapsing experimental rat model of multiple sclerosis. Amelioration of clinical status by M2 activated monocyte administration. *Mult Scler* 17 (1), 2–15.
- Miller, D.H., Hammond, S.R., McLeod, J.G., Purdie, G., Skegg, D.C., 1990. Multiple sclerosis in Australia and New Zealand: are the determinants genetic or environmental? *J. Neurology, Neurosurgery, and Psychiatry* 53 (10), 903–905.
- Miller, D.M., Weinstock-Guttman, B., Bethoux, F., Lee, J.C., Beck, G., Block, V., Durelli, L., LaMantia, L., Barnes, D., Sellebjerg, F., Rudick, R.A., 2000. A meta-analysis of methylprednisolone in recovery from multiple sclerosis exacerbations. *Mult Scler* 6 (4), 267–273.
- Miller, D., Barkhof, F., Montalban, X., Thompson, A., Filippi, M., 2005. Clinically isolated syndromes suggestive of multiple sclerosis, part I: natural history, pathogenesis, diagnosis, and prognosis. *Lancet Neurol* 4 (5), 281–288.
- Milo, R., Kahana, E., 2010. Multiple sclerosis: geoepidemiology, genetics and the environment. *Autoimmunity Rev* 9 (5), A387–A394.
- Miyaura, C., Abe, E., Kuribayashi, T., Tanaka, H., Konno, K., Nishii, Y., Suda, T., 1981. 1 α ,25-Dihydroxyvitamin D $_3$ induces differentiation of human myeloid leukemia cells. *Biochem Biophys Res Commun* 102 (3), 937–943.
- Monif, M., Reid, C.A., Powell, K.L., Smart, M.L., Williams, D.A., 2009. The P2X7 receptor drives microglial activation and proliferation: a trophic role for P2X7R pore. *J. Neurosci* 29 (12), 3781–3791.
- Monif, M., Reid, C.A., Powell, K.L., Drummond, K.J., O'Brien, T.J., Williams, D.A., 2016. Interleukin-1 β has trophic effects in microglia and its release is mediated by P2X7R pore. *J. Neuroinflammation* 13 (1), 173.
- Monney, L., Sabatos, C.A., Gaglia, J.L., Ryu, A., Waldner, H., Chernova, T., Manning, S., Greenfield, E.A., Coyle, A.J., Sobel, R.A., Freeman, G.J., Kuchroo, V.K., 2002. Th1-specific cell surface protein Tim-3 regulates macrophage activation and severity of an autoimmune disease. *Nature* 415 (6871), 536–541.

- Mulero, P., Midaglia, L., Montalban, X., 2018. Ocrelizumab: a new milestone in multiple sclerosis therapy. *Ther Adv Neurol Disord* 11.
- Munger, K.L., Levin, L.I., Hollis, B.W., Howard, N.S., Ascherio, A., 2006. Serum 25-hydroxyvitamin D levels and risk of multiple sclerosis. *Jama* 296 (23), 2832–2838.
- Munoz-Planillo, R., Kuffa, P., Martinez-Colon, G., Smith, B.L., Rajendiran, T.M., Nunez, G., 2013. K(+) efflux is the common trigger of NLRP3 inflammasome activation by bacterial toxins and particulate matter. *Immunity* 38 (6), 1142–1153.
- Muris, A.H., Smolders, J., Rolf, L., Thewissen, M., Hupperts, R., Damoiseaux, J., 2016. Immune regulatory effects of high dose vitamin D3 supplementation in a randomized controlled trial in relapsing remitting multiple sclerosis patients receiving IFNbeta; the SOLARIUM study. *J. Neuroimmunol* 300, 47–56.
- Murphy, R., O'Donoghue, S., Counihan, T., McDonald, C., Calabresi, P.A., Ahmed, M.A.S., Kaplin, A., Hallahan, B., 2017. Neuropsychiatric syndromes of multiple sclerosis. *J. Neurol Neurosurg Psychiatry* 88 (8), 697–708.
- Nakahara, J., Maeda, M., Aiso, S., Suzuki, N., 2012. Current concepts in multiple sclerosis: autoimmunity versus oligodendroglialopathy. *Clin Rev Allergy Immunol* 42 (1), 26–34.
- Narcisse, L., Scemes, E., Zhao, Y., Lee, S.C., Brosnan, C.F., 2005. The cytokine IL-1beta transiently enhances P2X7 receptor expression and function in human astrocytes. *Glia* 49 (2), 245–258.
- North, R.A., 2002. Molecular physiology of P2X receptors. *Physiol Rev* 82 (4), 1013–1067.
- Nunn, J.D., Katz, D.R., Barker, S., Fraher, L.J., Hewison, M., Hendy, G.N., O'Riordan, J.L., 1986. Regulation of human tonsillar T-cell proliferation by the active metabolite of vitamin D3. *Immunology* 59 (4), 479–484.
- Nurminen, V., Neme, A., Seuter, S., Carlberg, C., 2018. The impact of the vitamin D-modulated epigenome on VDR target gene regulation. *Biochim Biophys Acta* 1861 (8), 697–705.
- Obermeier, B., Mentele, R., Malotka, J., Kellermann, J., Kumpfel, T., Wekerle, H., Lottspeich, F., Hohlfeld, R., Dormair, G., 2008. Matching of oligoclonal immunoglobulin transcriptomes and proteomes of cerebrospinal fluid in multiple sclerosis. *Nat Med* 14 (6), 688–693.
- Oliveira, S.R., Simao, A.N.C., Alfieri, D.F., Flauzino, T., Kallaur, A.P., Mezzaroba, L., Lozovoy, M.A.B., Sabino, B.S., Ferreira, K.P.Z., Pereira, W., Kaimen-Maciel, D.R., Dichi, I., Reiche, E.M.V., 2017. Vitamin D deficiency is associated with disability and disease progression in multiple sclerosis patients independently of oxidative and nitrosative stress. *J Neurol Sci* 381, 213–219.
- O'Loughlin, E., Madore, C., Lassmann, H., Butovsky, O., 2018. Microglial phenotypes and functions in multiple sclerosis. *Cold Spring Harb Perspect Med* 8 (2).
- Orton, S.M., Herrera, B.M., Yee, I.M., Valdar, W., Ramagopalan, S.V., Sadovnick, A.D., Ebers, G.C., 2006. Sex ratio of multiple sclerosis in Canada: a longitudinal study. *Lancet Neurol* 5 (11), 932–936.
- Ottonello, L., Bertolotto, M., Montecucco, F., Dapino, P., Dallegrì, F., 2005. Dexamethasone-induced apoptosis of human monocytes exposed to immune complexes. Intervention of CD95- and XIAP-dependent pathways. *Int J Immunopathol Pharmacol* 18 (3), 403–415.
- Paneka, W., Jijon, H., Herx, L.M., Armstrong, J.N., Feighan, D., Wei, T., Yong, V.W., Ransohoff, R.M., MacVicar, B.A., 2001. P2X7-like receptor activation in astrocytes increases chemokine monocyte chemoattractant protein-1 expression via mitogen-activated protein kinase. *J Neurosci* 21 (18), 7135–7142.
- Pierrot-Deseilligny, C., Souberbielle, J.C., 2017. Vitamin D and multiple sclerosis: An update. *Mult Scler Relat Disord* 14, 35–45.
- Polman, C.H., O'Connor, P.W., Havrdova, E., Hutchinson, M., Kappos, L., Miller, D.H., Phillips, J.T., Lublin, F.D., Giovannoni, G., Wajgt, A., Toal, M., Lynn, F., Panzara, M.A., Sandrock, A.W., 2006. A randomized, placebo-controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl J Med* 354 (9), 899–910.
- Ponomarev, E.D., Veremeyko, T., Barteneva, N., Krichevsky, A.M., Weiner, H.L., 2011. MicroRNA-124 promotes microglia quiescence and suppresses EAE by deactivating macrophages via the C/EBP-alpha-PU.1 pathway. *Nat Med* 17 (1), 64–70.
- Prieti, B., Treiber, G., Mader, J.K., Hoeller, E., Wolf, M., Pilz, S., Graninger, W.B., Obermayer-Pietsch, B.M., Pieber, T.R., 2014. High-dose cholecalciferol supplementation significantly increases peripheral CD4(+) Tregs in healthy adults without negatively affecting the frequency of other immune cells. *Eur J Nutr* 53 (3), 751–759.
- Prinz, M., Garbe, F., Schmidt, H., Mildner, A., Gutcher, I., Wolter, K., Piesche, M., Schroers, R., Weiss, E., Kirschning, C.J., Rochford, C.D., Bruck, W., Becher, B., 2006. Innate immunity mediated by TLR9 modulates pathogenicity in an animal model of multiple sclerosis. *J. Clin Invest* 116 (2), 456–464.
- Provvedini, D.M., Tsoukas, C.D., Defetos, L.J., Manolagas, S.C., 1983. 1,25-dihydroxyvitamin D3 receptors in human leukocytes. *Science* 221 (4616), 1181–1183.
- Quan, J.H., Huang, R., Wang, Z., Huang, S., Choi, I.W., Zhou, Y., Lee, Y.H., Chu, J.Q., 2018. P2X7 receptor mediates NLRP3-dependent IL-1beta secretion and parasite proliferation in *Toxoplasma gondii*-infected human small intestinal epithelial cells. *Parasit Vectors* 11 (1), 1.
- Rahmanzadeh, R., Weber, M.S., Bruck, W., Navardi, S., Sahraian, M.A., 2018. B cells in multiple sclerosis therapy-A comprehensive review. *Acta Neurol Scand.* 137 (6), 544–556.
- Raivich, G., Banati, R., 2004. Brain microglia and blood-derived macrophages: molecular profiles and functional roles in multiple sclerosis and animal models of autoimmune demyelinating disease. *Brain Res Brain Res Rev* 46 (3), 261–281.
- Rasmussen, S., Wang, Y., Kivisakk, P., Bronson, R.T., Meyer, M., Imitola, J., Khoury, S.J., 2007. Persistent activation of microglia is associated with neuronal dysfunction of callosal projecting pathways and multiple sclerosis-like lesions in relapsing-remitting experimental autoimmune encephalomyelitis. *Brain* 130 (Pt 11), 2816–2829.
- Reich, D.S., Lucchinetti, C.F., Calabresi, P.A., 2018. Multiple sclerosis. *New Engl J Med* 378 (2), 169–180.
- Renno, T., Zeine, R., Girard, J.M., Gillani, S., Dodelet, V., Owens, T., 1994. Selective enrichment of Th1 CD45RbLow CD4+ T cells in autoimmune infiltrates in experimental allergic encephalomyelitis. *Int Immunol* 6 (3), 347–354.
- Rinchai, D., Boughorbel, S., Presnell, S., Quinn, C., Chaussabel, D., 2016. A curated compendium of monocyte transcriptome datasets of relevance to human monocyte immunobiology research. *F1000Res* 5 (291).
- Rodrigues, R.J., Tome, A.R., Cunha, R.A., 2015. ATP as a multi-target danger signal in the brain. *Front Neurosci* 9 (148).
- Rolf, L., Muris, A.H., Mathias, A., Du Pasquier, R., Konecny, I., Disanto, G., Kuhle, J., Ramagopalan, S., Damoiseaux, J., Smolders, J., Hupperts, R., 2018. Exploring the effect of vitamin D3 supplementation on the anti-EBV antibody response in relapsing-remitting multiple sclerosis. *Mult Scler* 24 (10), 1280–1287.
- Rudick, R.A., Ransohoff, R.M., 1992. Cytokine secretion by multiple sclerosis monocytes. Relationship to disease activity. *Arch Neurol* 49 (3), 265–270.
- Sabatino Jr., J.J., Zamvil, S.S., Hauser, S.L., 2018. B-cell therapies in multiple sclerosis. *Cold Spring Harb Perspect Med* 9 (2) pii: a032037.
- Sadovnick, A.D., Armstrong, H., Rice, G.P., Bulman, D., Hashimoto, L., Paty, D.W., Hashimoto, S.A., Warren, S., Hader, W., Murray, T.J., et al., 1993. A population-based study of multiple sclerosis in twins: update. *Ann Neurol* 33 (3), 281–285.
- Salzer, J., Hallmans, G., Nystrom, M., Stenlund, H., Wadell, G., Sundstrom, P., 2012. Vitamin D as a protective factor in multiple sclerosis. *Neurology* 79 (21), 2140–2145.
- Salzer, J., Bistrom, M., Sundstrom, P., 2014. Vitamin D and multiple sclerosis: where do we go from here? *Expert Rev Neurother* 14 (1), 9–18.
- Sanz, J.M., Chiozzi, P., Ferrari, D., Colaianna, M., Idzko, M., Falzoni, S., Fellin, R., Trabace, L., Di Virgilio, F., 2009. Activation of microglia by amyloid beta requires P2X7 receptor expression. *J. Immunol* 182 (7), 4378–4385.
- Saul, S., Gihardt, C.S., Schmidt, B., Lis, A., Pasieka, B., Conrad, D., Jung, P., Gaupp, R., Wonnemberg, B., Diler, E., Stanisz, H., Vogt, T., Schwarz, E.C., Bischoff, M., Herrmann, M., Tschernig, T., Kappl, R., Rieger, H., Niemeier, B.A., Bogeski, I., 2016. A calcium-redox feedback loop controls human monocyte immune responses: The role of ORAI Ca2+ channels. *Sci Signal* 9 (418), ra26.
- Savio, L.E.B., de Andrade Mello, P., da Silva, C.G., Coutinho-Silva, R., 2018. The P2X7 receptor in inflammatory diseases: angel or demon? *Front Pharmacol* 9 (52).
- Savransky, A., 2018. Demyelinating disorders. *Medicina (B Aires)* 78 (Suppl. 2), 75–81.
- Sawcer, S., Hellenthal, G., Pirinen, M., Spencer, C.C., Patsopoulos, N.A., Moutsianas, L., Dilthey, A., Su, Z., Freeman, C., Hunt, S.E., Edkins, S., Gray, E., Booth, D.R., Potter, S.C., Goris, A., Band, G., Oturai, A.B., Strange, A., Saarela, J., Bellenguez, C., Fontaine, B., Gillman, M., Hemmer, B., Gwilliam, R., Zipp, F., Jayakumar, A., Martin, R., Leslie, S., Hawkins, S., Giannoulatos, E., D'Alfonso, S., Blackburn, H., Martinelli Boneschi, F., Liddle, J., Harbo, H.F., Perez, M.L., Spurkland, A., Waller, M.J., Mycko, M.P., Ricketts, M., Comabella, M., Hammond, N., Kockum, I., McCann, O.T., Ban, M., Whittaker, P., Kempainen, A., Weston, P., Hawkins, C., Widaa, S., Zajicek, J., Drnov, S., Robertson, N., Bumpstead, S.J., Barcellos, L.F., Ravindrarajah, R., Abraham, R., Alfredsson, L., Ardlie, K., Aubin, C., Baker, A., Baker, K., Baranzini, S.E., Bergamaschi, L., Bergamaschi, R., Bernstein, A., Berthele, A., Boggild, M., Bradfield, J.P., Brassat, D., Broadley, S.A., Buck, D., Butzkueven, H., Capra, R., Carroll, W.M., Cavalla, P., Celius, E.G., Cepok, S., Chiavacci, R., Clerget-Darpoux, F., Cysters, K., Comi, G., Cossburn, M., Courmu-Rebeix, I., Cox, M.B., Cozen, W., Cree, B.A., Cross, A.H., Cusi, D., Daly, M.J., Davis, E., de Bakker, P.I., Debouverie, M., D'Hooghe, M., Dixon, K., Dobosi, R., Dubois, B., Ellinghaus, D., Elovaara, I., Esposito, F., et al., 2011. Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. *Nature* 476 (7359), 214–219.
- Schmidt, M., Pauels, H.G., Luger, N., Luger, A., Domschke, W., Kucharzik, T., 1999. Glucocorticoids induce apoptosis in human monocytes: potential role of IL-1 beta. *J. Immunol* 163 (6), 3484–3490.
- Sedal, L., Winkel, A., Laing, J., Law, L.Y., McDonald, E., 2017. Current concepts in multiple sclerosis therapy. *Degener Neurol Neuromuscul Dis* 7, 109–125.
- Sestito, C., Breve, J.J.P., van Eggermond, M., Killestein, J., Teunissen, C.E., van Rossum, J., Wilhelmus, M.M.M., Drukarch, B., van den Elsen, P.J., van Dam, A.M., 2017. Monocyte-derived tissue transglutaminase in multiple sclerosis patients: reflecting an anti-inflammatory status and function of the cells? *J. Neuroinflammation* 14 (1), 257.
- Shieh, C.H., Heinrich, A., Serchov, T., van Calker, D., Biber, K., 2014. P2X7-dependent, but differentially regulated release of IL-6, CCL2, and TNF-alpha in cultured mouse microglia. *Glia* 62 (4), 592–607.
- Simpson Jr., S., Taylor, B., Blizzard, L., Ponsonby, A.L., Pittas, F., Tremlett, H., Dwyer, T., Gies, P., van der Mei, I., 2010. Higher 25-hydroxyvitamin D is associated with lower relapse risk in multiple sclerosis. *Ann Neurol* 68 (2), 193–203.
- Sinha, S., Itani, F.R., Karandikar, N.J., 2014. Immune regulation of multiple sclerosis by CD8+ T cells. *Immunol Res* 59 (1-3), 254–265.
- Smolders, J., Moen, S.M., Damoiseaux, J., Huitinga, I., Holmoy, T., 2011. Vitamin D in the healthy and inflamed central nervous system: access and function. *J. Neurol Sci* 311 (1-2), 37–43.
- Soilu-Hanninen, M., Airas, L., Mononen, I., Heikkilä, A., Viljanen, M., Hanninen, A., 2005. 25-Hydroxyvitamin D levels in serum at the onset of multiple sclerosis. *Mult Scler* 11 (3), 266–271.
- Sommer, A., Fabri, M., 2015. Vitamin D regulates cytokine patterns secreted by dendritic cells to promote differentiation of IL-22-producing T cells. *PLoS One* 10 (6), e0130395.
- Sommerlad, A., Price, G., Trip, A., 2014. Management of neuropsychiatric symptoms in multiple sclerosis. *Prog Neurol Psychiatr* 18 (2), 14–19.
- Sotirchos, E.S., Bhargava, P., Eckstein, C., Van Haren, K., Baynes, M., Ntranos, A., Gocke, A., Steinman, L., Mowry, E.M., Calabresi, P.A., 2016. Safety and immunologic effects of high- vs low-dose cholecalciferol in multiple sclerosis. *Neurology* 86 (4), 382–390.
- Sperlagh, B., Illes, P., 2014. P2X7 receptor: an emerging target in central nervous system diseases. *Trends Pharmacol Sci* 35 (10), 537–547.
- Thacher, T.D., Fischer, P.R., Pettifor, J.M., 2014. Vitamin D treatment in calcium-deficiency rickets: a randomised controlled trial. *Arch Dis Child* 99 (9), 807–811.

- Thompson, A.J., Banwell, B.L., Barkhof, F., Carroll, W.M., Coetzee, T., Comi, G., Correale, J., Fazekas, F., Filippi, M., Freedman, M.S., Fujihara, K., Galetta, S.L., Hartung, H.P., Kappos, L., Lublin, F.D., Marrie, R.A., Miller, A.E., Miller, D.H., Montalban, X., Mowry, E.M., Sorensen, P.S., Tintore, M., Traboulsee, A.L., Trojano, M., Uitdehaag, B.M.J., Vukusic, S., Waubant, E., Weinshenker, B.G., Reingold, S.C., Cohen, J.A., 2018. Diagnosis of multiple sclerosis: 2017 revisions of the McDonald criteria. *Lancet Neurol* 17 (2), 162–173.
- Trojano, M., Lucchese, G., Graziano, G., Taylor, B.V., Simpson Jr., S., Lepore, V., Grand'maison, F., Duquette, P., Izquierdo, G., Grammond, P., Amato, M.P., Bergamaschi, R., Giuliani, G., Boz, C., Hupperts, R., Van Pesch, V., Lechner-Scott, J., Cristiano, E., Fiol, M., Oreja-Guevara, C., Saladino, M.L., Verheul, F., Slee, M., Paolicelli, D., Tortorella, C., D'Onghia, M., Iaffaldano, P., Drenzo, V., Butzkueven, H., 2012. Geographical variations in sex ratio trends over time in multiple sclerosis. *PLoS One* 7 (10), e48078.
- Tsang, B.K.-T., Macdonell, R., 2011. Multiple sclerosis diagnosis, management and prognosis. *Austral Family Phys* 40 (12), 948–955.
- Tvrdek, P., Kalani, M.Y.S., 2017. In vivo imaging of microglial calcium signaling in brain inflammation and injury. *Int J Mol Sci* 18 (11).
- Tzartos, J.S., Friese, M.A., Craner, M.J., Palace, J., Newcombe, J., Esiri, M.M., Fugger, L., 2008. Interleukin-17 production in central nervous system-infiltrating T cells and glial cells is associated with active disease in multiple sclerosis. *Am J Pathol* 172 (1), 146–155.
- Vaknin, I., Kunis, G., Miller, O., Butovsky, O., Bukshpan, S., Beers, D.R., Henkel, J.S., Yoels, E., Appel, S.H., Schwartz, M., 2011. Excess circulating alternatively activated myeloid (M2) cells accelerate ALS progression while inhibiting experimental autoimmune encephalomyelitis. *PLoS One* 6 (11), e26921.
- van der Burg, B., van der Saag, P.T., 1996. Nuclear factor-kappa-B/steroid hormone receptor interactions as a functional basis of anti-inflammatory action of steroids in reproductive organs. *Mol Hum Reprod* 2 (6), 433–438.
- van Winsen, L.M., Muris, D.F., Polman, C.H., Dijkstra, C.D., van den Berg, T.K., Uitdehaag, B.M., 2005. Sensitivity to glucocorticoids is decreased in relapsing remitting multiple sclerosis. *J. Clin Endocrinol Metab* 90 (2), 734–740.
- Veldman, C.M., Cantorna, M.T., DeLuca, H.F., 2000. Expression of 1,25-dihydroxyvitamin D(3) receptor in the immune system. *Arch Biochem Biophys* 374 (2), 334–338.
- Volonte, C., Apolloni, S., Skaper, S.D., Burnstock, G., 2012. P2X7 receptors: channels, pores and more. *CNS Neurol Disord Drug Targets* 11 (6), 705–721.
- Wei, W., Ryu, J.K., Choi, H.B., McLarnon, J.G., 2008. Expression and function of the P2X (7) receptor in rat C6 glioma cells. *Cancer Lett* 260 (1–2), 79–87.
- Weiner, H.L., 2009. The challenge of multiple sclerosis: how do we cure a chronic heterogeneous disease? *Ann Neurol* 65 (3), 239–248.
- Wilhelm, H., Schabet, M., 2015. The diagnosis and treatment of optic neuritis. *Deutsches Ärzteblatt Int* 112 (37), 616–626.
- Wilhelm, K., Ganesan, J., Muller, T., Durr, C., Grimm, M., Beilhack, A., Krempl, C.D., Sorichter, S., Gerlach, U.V., Juttner, E., Zerweck, A., Gartner, F., Pellegatti, P., Di Virgilio, F., Ferrari, D., Kambham, N., Fisch, P., Finke, J., Idzko, M., Zeiser, R., 2010. Graft-versus-host disease is enhanced by extracellular ATP activating P2X7R. *Nat Med* 16 (12), 1434–1438.
- Yadav, S.K., Mindur, J.E., Ito, K., Dhib-Jalbut, S., 2015. Advances in the immunopathogenesis of multiple sclerosis. *Curr Opin Neurol* 28 (3), 206–219.
- Yamasaki, R., Lu, H., Butovsky, O., Ohno, N., Rietsch, A.M., Cialic, R., Wu, P.M., Doykan, C.E., Lin, J., Coteleur, A.C., Kidd, G., Zorlu, M.M., Sun, N., Hu, W., Liu, L., Lee, J.C., Taylor, S.E., Uehlein, L., Dixon, D., Gu, J., Floruta, C.M., Zhu, M., Charo, I.F., Weiner, H.L., Ransohoff, R.M., 2014. Differential roles of microglia and monocytes in the inflamed central nervous system. *J. Exp Med* 211 (8), 1533–1549.
- Yiangou, Y., Facer, P., Durrenberger, P., Chessell, I.P., Naylor, A., Bountra, C., Banati, R.R., Anand, P., 2006. COX-2, CB2 and P2X7-immunoreactivities are increased in activated microglial cells/macrophages of multiple sclerosis and amyotrophic lateral sclerosis spinal cord. *BMC Neurol* 6 (12).
- Yoshida, K., Ito, M., Hoshino, Y., Matsuoka, I., 2017. Effects of dexamethasone on purinergic signaling in murine mast cells: Selective suppression of P2X7 receptor expression. *Biochem Biophys Res Commun* 493 (4), 1587–1593.
- Zhang, Y., Leung, D.Y., Richers, B.N., Liu, Y., Remigio, L.K., Riches, D.W., Goleva, E., 2012. Vitamin D inhibits monocyte/macrophage proinflammatory cytokine production by targeting MAPK phosphatase-1. *J. Immunol* 188 (5), 2127–2135.
- Zrzavy, T., Hametner, S., Wimmer, I., Butovsky, O., Weiner, H.L., Lassmann, H., 2017. Loss of 'homeostatic' microglia and patterns of their activation in active multiple sclerosis. *Brain* 140 (7), 1900–1913.