

- 1 Rylander E, Ruusuvaara L, Almstromer MW, Evander M, Wadell G. The absence of vaginal human papillomavirus 16 DNA in women who have not experienced sexual intercourse. *Obstet Gynecol* 1994; **83**: 735–37.
- 2 Dillner J, Kallings I, Brihmer C, et al. Seropositivities to human papillomavirus types 16, 18, or 33 capsids and to Chlamydia trachomatis are markers of sexual behavior. *J Infect Dis* 1996; **173**: 1394–98.
- 3 Widdice L, Ma Y, Jonte J, et al. Concordance and transmission of human papillomavirus within heterosexual couples observed over short intervals. *J Infect Dis* 2013; **207**: 1286–94.
- 4 Winer RL, Hughes JP, Feng Q, et al. Detection of genital HPV types in fingertip samples from newly sexually active female university students. *Cancer Epidemiol Biomarkers Prev* 2010; **19**: 1682–85.
- 5 Roden RB, Lowy DR, Schiller JT. Papillomavirus is resistant to desiccation. *J Infect Dis* 1997; **176**: 1076–79.
- 6 Malagón T, Louvanto K, Wissing M, et al. Hand-to-genital and genital-to-genital transmission of human papillomaviruses between male and female sexual partners (HITCH): a prospective cohort study. *Lancet Infect Dis* 2019; published online Feb 8. [http://dx.doi.org/10.1016/S1473-3099\(18\)30655-8](http://dx.doi.org/10.1016/S1473-3099(18)30655-8).
- 7 Forslund O, Nordin P, Andersson K, Stenquist B, Hansson BG. DNA analysis indicates patient-specific human papillomavirus type 16 strains in Bowen's disease on fingers and in archival samples from genital dysplasia. *Br J Dermatol* 1997; **136**: 678–82.
- 8 Egawa N, Egawa K, Griffin H, Doorbar J. Human papillomaviruses, epithelial tropisms, and the development of neoplasia. *Viruses* 2015; **7**: 3863–90.
- 9 Wools K, Bryan JT, Katz BP, Rodriguez M, Davis T, Brown DR. Detection of human papillomavirus L1 protein in condylomata acuminata from various anatomical sites. *Sex Transm Dis* 1994; **21**: 103–06.



Long-term effectiveness of MenAfriVac

Published Online
February 8, 2019
[http://dx.doi.org/10.1016/S1473-3099\(18\)30725-4](http://dx.doi.org/10.1016/S1473-3099(18)30725-4)

See [Articles](#) page 327

A conjugate vaccine for *Neisseria meningitidis* serogroup A (MenAfriVac) was introduced through mass campaigns in people aged 1–29 years in the African meningitis belt and is now being added to national infant immunisation programmes.¹ Following the mass campaigns in 2010–17, incidence of *N meningitidis* serogroup A meningitis has declined from a hyperendemic pattern with regular epidemics to a few sporadic cases.² The hope is that control of the disease will be long lasting. The mechanism for such control would be the initial interruption of circulation of *N meningitidis* serogroup A in the population through the effect of the vaccine against asymptomatic nasopharyngeal infection, followed by sustained direct protection against disease, including in new birth cohorts. The duration of persistence of antibodies is a major variable in this scheme.

In *The Lancet Infectious Diseases*, Michael White and colleagues report results from a mathematical model,³ originally developed for natural malaria infection but used here to estimate the kinetics of *N meningitidis* serogroup A-specific serum bactericidal antibodies (SBAs) in the African meningitis belt. The model uses serological data obtained in clinical trials^{4,5} before and up to 2 years after vaccination with MenAfriVac. These trials tested a single dose of MenAfriVac in children and adults aged 12 months to 29 years.^{4,5} The model predicts that vaccine efficacy after 20 years will be 52% (95 credible interval 29–73) among 12–23 month-old children and 70% (60–79) among older children and adults. These results suggest that no booster vaccination would be needed for people vaccinated with MenAfriVac at age 2 years or older.

This research is an important complement to two other approaches: first, the two clinical trials^{4,5} followed SBA persistence up to 5 years after vaccination and found

that after a substantial decline of 40–80% during the first year after vaccination, antibody titres were stable in all age groups between 1 and 29 years. One limitation of these estimates is that, unlike the current situation of quasi-elimination, *N meningitidis* serogroup A continued to circulate in trial populations and natural boosting might have maintained higher antibody levels. White and colleagues overcome this limitation in their model by simply not assuming any such boosting effect and also provide a long-term estimate up to 20 years after vaccination. Second, from a series of population-level cross-sectional studies in Burkina Faso and exponential extrapolation of the decline, we estimated that SBA antibody titres would return to pre-vaccination levels after 12 years at the earliest.⁶ Although our work's strength was that it reflects the real-life situation of immigrating unvaccinated individuals, its ecological design and lack of document-based vaccination status for most participants in subsequent surveys was a limitation.

The results we obtained, however, shed light on an important limitation of all current work: there is no confirmed correlate of protection against *N meningitidis* serogroup A in the meningitis belt, neither for meningitis nor for asymptomatic infection. We have previously shown that the seroprevalence of putatively protective SBA titres against the *N meningitidis* serogroup A reference strain ($\geq 1/8$ or $\geq 1/128$) does not correlate with meningitis incidence.⁷ This finding was not surprising, given that the seroprevalence of these titres was more than 70% among children and adults already in the pre-vaccination era. As an alternative serological measure, we proposed the SBA against strain 3125, which should be specific for vaccine-induced protection.⁸ Among children vaccinated before 5 years of age, the return to pre-vaccination levels

of these 3125-SBA titres would be expected after 8 years.⁶ These results suggest that a MenAfriVac booster vaccination would currently be required for this group to maintain immune protection.

This uncertainty around the correlate of protection raises some concern. It is worrying that the pathophysiology of serogroup A meningitis found in the African belt is not well understood. One hypothesis is that under the extreme climatic conditions during the dry season (high aerosol load and low air humidity), meningococci can invade the meninges directly from the nasopharynx through the lymphatic tissue along the olfactory nerve.⁹ In this scenario, conjugate vaccines are effective primarily, or perhaps only, through their capacity to protect against nasopharyngeal infection. In any case, it is widely accepted that the interrupted pathogen circulation is the main determinant of strong and persistent impact of high-coverage conjugate vaccine programmes.¹ The key factor to evaluate, then, is the mucosal immunity induced by conjugate vaccines. Unfortunately, there is even less understanding of how this mucosal immunity works. No validated measures are available, although salivary antibodies might be candidates.^{10,11} For now, the sole way to evaluate the vaccine-induced protection against asymptomatic infection is through large and costly carriage studies.

The pragmatic approach is to acknowledge that conjugate vaccines work well, particularly because of indirect protection, and that further evaluation is necessary only if *N meningitidis* serogroup A epidemics re-emerge. Furthermore, a vaccine against five meningococcal serogroups (A, C, W, X, and Y) is being developed¹ and its introduction through mass campaigns will provide a booster against *N meningitidis* serogroup A.

To move forwards, my wish would be to see the development of a simple, specific test of mucosal immunity against meningococci, which would allow for the follow-up assessment of protection against

asymptomatic nasopharyngeal infection among trial participants and vaccinated populations. Mathematical modelling based on such data will allow estimation of the duration of both direct and indirect protection after vaccination and thus offer a powerful tool to develop strategies for long-term control of meningococcal meningitis.

Judith E Mueller

EHESP French School of Public Health, Paris, France and Institut Pasteur, 75724 Paris cedex 15, France
judith.mueller@ehesp.fr

I declare no competing interests.

- 1 LaForce FM, Djingarey M, Viviani S, Preziosi MP. Successful African introduction of a new group A meningococcal conjugate vaccine: future challenges and next steps. *Hum Vaccin Immunother* 2018; **14**: 1098–102.
- 2 Diallo AO, Soeters HM, Yameogo I, et al. MenAfriNet Consortium. Bacterial meningitis epidemiology and return of *Neisseria meningitidis* serogroup A cases in Burkina Faso in the five years following MenAfriVac mass vaccination campaign. *PLoS One* 2017; **12**: e0187466.
- 3 White M, Idoko O, Sox S, et al. Antibody kinetics following vaccination with MenAfriVac: an analysis of serological data from randomised trials. *Lancet Infect Dis* 2019; published online Feb 5. [http://dx.doi.org/10.1016/S1473-3099\(18\)30674-1](http://dx.doi.org/10.1016/S1473-3099(18)30674-1).
- 4 Diallo A, Sow SO, Idoko OT, et al. Antibody persistence at 1 and 4 years following a single dose of MenAfriVac or quadrivalent polysaccharide vaccine in healthy subjects aged 2–29 years. *Clin Infect Dis* 2015; **61**: S521–30.
- 5 Tapia MD, Findlow H, Idoko OT, et al. Antibody persistence 1–5 years following vaccination with MenAfriVac in African children vaccinated at 12–23 months of age. *Clin Infect Dis* 2015; **61**: S514–20.
- 6 Yaro S, Njanpop Lafourcade BM, Ouangraoua S, et al. Antibody persistence at the population level 5 years after mass vaccination with meningococcal serogroup A conjugate vaccine (PsA-TT) in Burkina Faso: need for a booster campaign? *Clin Infect Dis* 2019; **68**: 435–43.
- 7 Trotter CL, Yaro S, Njanpop-Lafourcade BM, et al. Seroprevalence of bactericidal, specific IgG antibodies and incidence of meningitis due to group A *Neisseria meningitidis* by age in Burkina Faso 2008. *PLoS One* 2013; **8**: e55486.
- 8 Poolman JT, De Vleeschauwer I, Durant N, et al. Measurement of functional anti-meningococcal serogroup A activity using strain 3125 as the target strain for serum bactericidal assay. *Clin Vaccine Immunol* 2011; **18**: 1108–17.
- 9 Sjölander H, Jonsson AB. Olfactory nerve—a novel invasion route of *Neisseria meningitidis* to reach the meninges. *PLoS One* 2010; **5**: e14034.
- 10 Bärnes GK, Workalemahu B, Kristiansen PA, et al. Salivary and serum antibody response against *Neisseria meningitidis* after vaccination with conjugate polysaccharide vaccines in Ethiopian volunteers. *Scand J Immunol* 2016; **84**: 118–29.
- 11 van Ravenhorst MB, den Hartog G, van der Klis FRM, van Rooijen DM, Sanders EAM, Berbers GAM. Induction of salivary antibody levels in Dutch adolescents after immunization with monovalent meningococcal serogroup C or quadrivalent meningococcal serogroup A, C, W and Y conjugate vaccine. *PLoS One* 2018; **13**: e0191261.

Twitter to engage, educate, and advocate for global antibiotic stewardship and antimicrobial resistance



The WHO Global Action Plan on Antimicrobial Resistance recommends countries work together to improve awareness and understanding of antimicrobial

resistance, including through social media.¹² Twitter disseminates news in seconds around the world in real-time to anyone with an internet-connected