



Tetra-linoleoyl cardiolipin depletion plays a major role in the pathogenesis of sarcopenia



Richard D. Semba^{a,*}, Ruin Moaddel^b, Pingbo Zhang^a, Christopher E. Ramsden^{b,c}, Luigi Ferrucci^b

^a Wilmer Eye Institute, Johns Hopkins University School of Medicine, Baltimore, MD, United States

^b National Institute on Aging, National Institutes of Health, Baltimore, MD, United States

^c National Institute on Alcohol Abuse and Alcoholism, National Institutes of Health, Bethesda, MD, United States

ARTICLE INFO

Keywords:

Aging
Cardiolipin
Lipids
Mitochondria
Sarcopenia

ABSTRACT

Sarcopenia, the progressive loss of muscle mass, strength, and physical performance that occurs during aging, is highly prevalent among the elderly. Sarcopenia increases the risk of falls, disability, and death. The biological basis for sarcopenia is not well understood. There are no specific preventive or therapeutic strategies for sarcopenia except exercise. The elucidation of biological pathways and identification of therapeutic targets for treating or preventing sarcopenia remain a high priority in aging research. Mitochondria play a critical role in skeletal muscle by providing energy in the form of ATP, regulation of signaling, calcium homeostasis, autophagy, and other functions. Cardiolipin, a unique dimeric phospholipid specific to mitochondria and an essential component of mitochondrial membranes, is involved in mitochondrial protein transport, maintaining structural organization of mitochondrial membranes, cellular signaling, regulating enzymes involved in β -oxidation of fatty acids, and facilitating normal electron transport chain (ETC) function and generation of ATP. The fatty acid species composition of cardiolipin is critical to mitochondrial bioenergetics, as cardiolipin affects membrane biophysical properties, binds and stabilizes ETC protein complexes, and shapes the curvature of the mitochondrial cristae. Tetra-linoleoyl cardiolipin (18:2)₄ comprises ~80% of cardiolipin in mitochondria in normal human skeletal and cardiac muscle and is optimal for effective ETC function and ATP generation. Aging is associated with a decrease in cardiolipin content, decrease in tetra-linoleoyl cardiolipin (18:2)₄ and replacement of linoleic acid (18:2) with other fatty acids in cardiolipin composition, decline of ETC function, and increased generation of reactive oxygen species in muscle. Together, these findings from the literature prompt the hypothesis that depletion of the cardiolipin (18:2)₄ species may be at the root of mitochondrial dysfunction with aging, in turn leading to sarcopenia. Corroboration of the tetra-linoleoyl cardiolipin depletion hypothesis suggests new leads for the prevention and treatment of sarcopenia by enhancing the biosynthesis, accretion, and integrity of tetra-linoleoyl cardiolipin.

Introduction

Sarcopenia is the progressive loss of muscle mass, strength, and physical performance that occurs during aging [1,2]. Sarcopenia is estimated to affect 10% of community-dwelling adults ≥ 60 years worldwide [3] and ~20 to 70% of chronically ill older adults [4]. From 20 to 80 years of age, humans generally lose ~20 to 40% of both skeletal muscle mass and strength [5,6]. Aging is accompanied by loss of muscle fibers and a reduction in the size of muscle fibers [7]. Low muscle mass is associated with low strength [8], decreased lower extremity performance [9], functional impairment [10], falls [11,12], and physical disability [10,13,14]. The decline of muscle strength with aging is greater than would be predicted by loss of muscle mass alone

[15]. Other factors such as muscle quality [16], neuromuscular junction and motor unit size may contribute to the age-related loss of muscle strength [17,18]. Longitudinal studies show that by around 75 years of age, muscle strength is lost at a rate of 3–4% per year in men and 2.5–3% per year in women [19]. Low muscle strength predicts disability [20,21] and mortality [22,23]. Major risk factors for sarcopenia are age, physical inactivity, inadequate nutrition, alteration in sex hormones, and the age-related pro-inflammatory state [1,2,24,25].

Biological pathways that have been implicated in sarcopenia include hormonal changes [26], anorexia and malnutrition [27,28], reduced satellite cell function [29], altered proteostasis [30], impairment of neuromuscular function [17], increased inflammation [31], and mitochondrial dysfunction [32]. Mitochondria are essential for fatty

* Corresponding author.

E-mail address: rdsemba@jhmi.edu (R.D. Semba).

acid and glucose metabolism [33], energy production [32], regulation of intracellular calcium homeostasis [34], reactive oxygen species (ROS) signaling [35], mitophagy [36,37], and apoptosis in skeletal muscle [32,38] and are a nexus in the pathway of some of these proposed pathways to sarcopenia.

Aging and sarcopenia are associated with a loss of bioenergetic capacity

Progressive mitochondrial dysfunction is a central hallmark of aging [39–44]. Skeletal muscle mitochondrial protein synthesis, biogenesis, respiratory capacity, coupling control, ATP production, and calcium-handling capacity declines with age [44–48]. The relationship of aging with mitochondrial energetics has been studied invasively using direct measurement of oxygen consumption by respirometry in isolated mitochondria or permeabilized muscle fibers from skeletal muscle biopsies [48] or non-invasively in humans using phosphorus magnetic resonance spectroscopy (^{31}P -MRS). ^{31}P -MRS measures the post exercise recovery rate of phosphocreatine (PCr), k_{PCr} , which reflects the capacity of muscle mitochondria to synthesize ATP [48]. ^{31}P -MRS correlates well with *in vitro* measurements of mitochondrial oxidative capacity [49,50]. Low k_{PCr} is independently associated with slower gait speed in adults [51]. Impaired mitochondrial oxidative function, as assessed by ^{31}P -MRS is associated with insulin resistance in older adults without diabetes [52].

Hypothesis: Depletion of tetra-linoleoyl cardiolipin plays a major role in the pathogenesis of sarcopenia

Cardiolipin is a unique dimeric phospholipid specific to mitochondria and an essential component of mitochondrial membranes [53]. Cardiolipin consists of a glycerol headgroup, two phosphatidyl moieties, and four fatty acid chains (Fig. 1). The four fatty acid chains can differ in length and saturation. Cardiolipin is distinct from other phospholipids in that it contains two phosphatidyl moieties bound to a single glycerol group, a feature which results in a small, relatively rigid head group and a large rigid hydrophobic tail [54]. The functions of cardiolipin include mitochondrial protein transport, maintaining structural organization of the inner mitochondrial membrane, cellular signaling, regulating enzymes involved in β -oxidation of fatty acids, and facilitating electron transport chain (ETC) function and generation of ATP [55–58]. Alterations in cardiolipin could play a major role in the impairment of mitochondrial function and decline of muscle mass, strength, and physical performance with aging. A conceptual model for the hypothesis that depletion of tetra-linoleoyl cardiolipin plays a

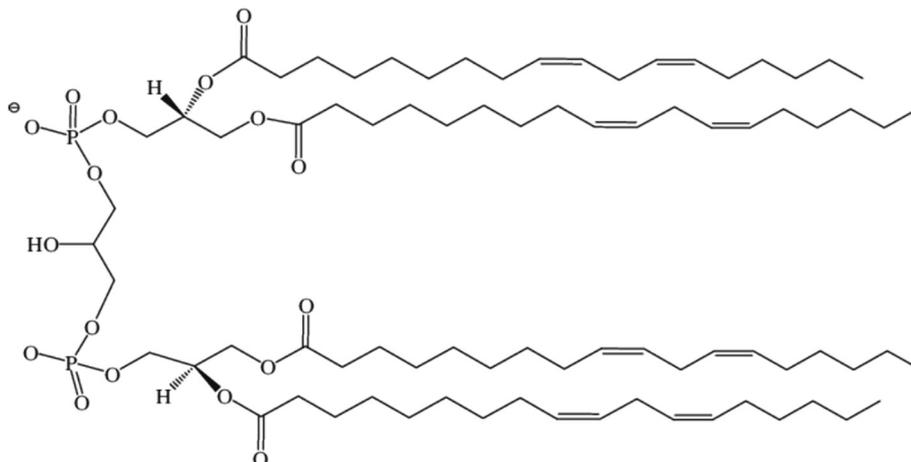


Fig. 1. The structure of cardiolipin includes a glycerol headgroup, two phosphatidyl moieties, and four fatty acid chains. The example shown is tetra-linoleoyl cardiolipin (18:2)₄ with the four fatty acid chains consisting of linoleic acid.

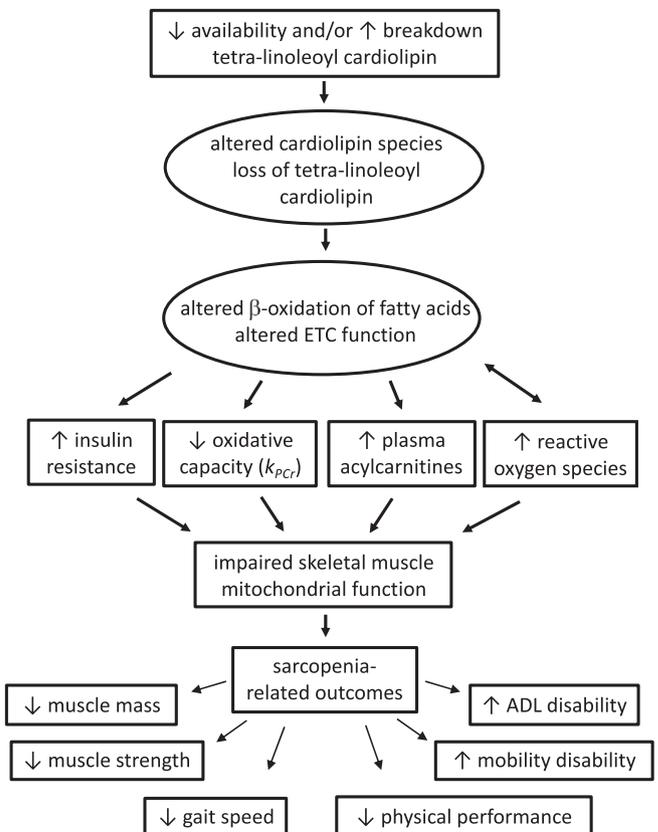


Fig. 2. A conceptual model for the hypothesis that depletion of tetra-linoleoyl cardiolipin plays a central role in sarcopenia.

central role in sarcopenia is shown in Fig. 2.

Synthesis and remodeling of cardiolipin

Cardiolipin is synthesized in a pathway involving lysophosphatidic acid (LPA), phosphatidic acid (PA), and other intermediates (Fig. 3). LPA can be generated from the addition of an acyl group from acyl-CoA to the *sn*-1 position of glycerol-3-phosphate (G3P) by glycerol-3-phosphate acyltransferase (GPAT) [59]. There are four isoforms of GPAT, coded by separate genes, with localization to the outer mitochondrial membrane (GPAT1, GPAT2) and endoplasmic reticulum (ER) (GPAT3, GPAT4) [60]. LPA can also be generated through the hydrolysis of LPC

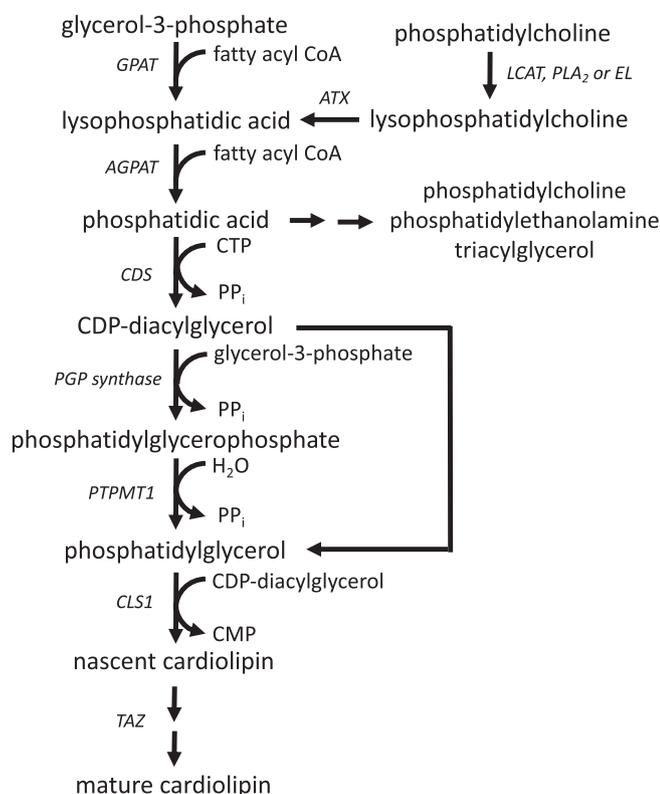


Fig. 3. Synthesis pathway of cardiolipin from lysophosphatidic acid and lysophosphatidylcholine. Abbreviations: acylglycerol-3-phosphate acyltransferase (AGPAT), autotaxin (ATX), cardiolipin synthase (CLS1), cytidine diphosphate (CDP), CDP-diacylglycerol synthase (CDS), cytidine monophosphate (CMP), cytidine triphosphate (CTP), endothelial lipase (EL), glycerol-3-phosphate acyltransferase (GPAT), phosphatidylcholine-sterol acyltransferase (LCAT), phosphatidylglycerophosphate and protein-tyrosine phosphatase 1 (PTPMT1), phosphatidylglycerophosphate synthase (PGP synthase), phospholipase A₂ (PLA₂), pyrophosphate (PP_i), tafazzin (TAZ).

by ectonucleotide pyrophosphatase/phosphodiesterase family member 2 (ENPP2) (also known as autotaxin), a glycoprotein widely expressed in tissues and present in ER, cytoplasm, and plasma [61–63]. LPC is present in mitochondria and ER membranes [64–67] and intracellular lipid droplets, which are located adjacent to mitochondria in skeletal muscle [68–70]. The role of lipid droplets in skeletal muscle lipid metabolism is not well understood. Lipid droplets consist of neutral lipids surrounded by a phospholipid monolayer containing proteins involved in lipid synthesis and remodeling, lipid droplet clustering and fusion, and scaffold proteins [68–71]. LPC can be generated from membrane phospholipids by phospholipase A₂ enzymes, a step in the Lands' cycle of lipid remodeling [72]. Peroxiredoxin 6 (PRDX6) is localized in the cytoplasm and has PLA₂ activity to generate LPC from phosphatidylcholine (PC) [73,74].

Most of the enzymes involved in cardiolipin synthesis have been localized to ER or a specialized ER region known as the mitochondrial-associated membrane (MAM) where the ER membrane and outer mitochondrial membrane are in close proximity [59]. The precise locations of the different enzymes involved in phospholipid synthesis, the transfer of lipid species between ER and mitochondria, and the potential role of autophagosomes are not completely known [59,75]. The MAM is enriched with terminal enzymes of different lipid synthesis pathways, suggesting MAM is a hub for lipid synthesis [59]. PA, the central precursor for cardiolipin synthesis, can be generated via pathways within ER or in the outer mitochondrial membrane, but the relative contribution of each source for cardiolipin synthesis has not been established [54]. Acylglycerol-3-phosphate acyltransferase (AGPAT)

catalyzes the acylation of LPA at the *sn*-2 position to form PA. AGPAT3 is located in the ER membrane [76]. AGPAT4 and AGPAT5 are located on the outer mitochondrial membrane [77,78]. Subsequently, PA is converted in the ER to cytidinediphosphate-1,2,-diacyl-*sn*-glycerol (CDP-DAG) by CDP-diacylglycerol synthase, also known as Tam41 [79]. An example of the intersection and overlap of different lipid pathways in the MAM is CDP-DAG, which can be converted to phosphatidylglycerol phosphate (PGP) by PGP synthase as the committed step in the cardiolipin synthesis pathway, or can enter synthesis pathways for PC (Kennedy pathway), phosphatidylethanolamine (PE), monoacylglycerol, or triacylglycerol [80]. PGP synthase transfers the activated phosphatidyl group from CDP-DAG to the *sn*-1 position of G3P to yield PGP. PGP is then hydrolyzed by protein tyrosine phosphatase mitochondrion 1 (PTPMT1) to form phosphatidylglycerol (PG) [81]. In the final step of the cardiolipin synthesis pathway, which occurs in the inner mitochondrial membrane, cardiolipin synthase (CLS) transfers a phosphatidyl group from another CDP-DAG molecule to PG to form nascent cardiolipin [80].

Nascent cardiolipin contains four fatty acid chains but then undergoes structural remodeling in which cardiolipin acquires a new set of fatty acids. The final fatty acid composition of the four acyl chains of mature cardiolipin is significantly different from nascent cardiolipin [53]. The remodeling of cardiolipin is catalyzed by tafazzin (TAZ), an enzyme that transfers fatty acids between phospholipids and lysophospholipids [53]. After cycles of deacylation and reacylation, remodeled cardiolipin contains predominantly unsaturated fatty acids. TAZ is generally considered to have no substrate preference and is not specific for fatty acid chains; substrate availability, physical properties of lipids, and thermodynamics are considered to influence the final form of cardiolipin in mitochondrial inner membranes [57,82]. Cardiolipin is relatively long-lived and has a half-life that is several times longer than the half-life of other phospholipids [53,83–85].

The fatty species composition of cardiolipin is critical to mitochondrial bioenergetics, as cardiolipin affects membrane biophysical properties and shapes the curvature of the mitochondrial cristae [57,86]. Cardiolipin is cone-shaped and generates negative curvature elastic stress in lipid bilayers [54,87]. PE is also cone-shaped with similar membrane properties. Together cardiolipin and PE comprise ~50% of phospholipid mass in the inner mitochondrial membrane [Ikou, 2017]. The conical shape of these lipids gives stability to the tight curvature of the crista membranes, with the small glycerol head groups of cardiolipin and PE facing the cristae lumen. The curvature and properties of the membrane affect the assembly and function of the ETC complexes [88] and mitochondrial metabolic signaling [58]. Cardiolipin directly binds to complexes I [89,90], III [89,91–93], IV [94,95], and V [96] and the ADP/ATP carrier (ATC) [97,98] of the ETC. Cardiolipin is an integral component to ETC proteins and is critical to their folding, configuration, and function [88]. Disruption of cardiolipin biosynthesis results in impaired ETC complex formation [99,100], increased proton leak [101], and altered cristae morphology [102]. Reduction of cardiolipin impairs the activity of cytochrome *c* oxidase (COX), or ETC complex IV [103,104].

Tetra-linoleoyl cardiolipin (18:2)₄ is the dominant species in skeletal muscle and heart

The cardiolipin that comprises ~80% of cardiolipin in mitochondria from normal human skeletal and cardiac muscle is tetra-linoleoyl cardiolipin (18:2)₄ (Fig. 1) [57]. There are many different possible combinations of fatty acids that could comprise the four fatty acid chains found in cardiolipin. With fourteen major fatty acids, eukaryotes could potentially produce 38,416 distinct cardiolipin species [105], but it is important to note that most cardiolipin in skeletal muscle and heart consists of tetra-linoleoyl cardiolipin (18:2)₄ [57,106]. This highly symmetrical conical configuration of cardiolipin and cardiolipin binding to membrane proteins optimizes function of the ETC in the

inner mitochondrial membrane [88].

The evidence that cardiolipin (18:2)₄ is critical for mitochondrial function comes from studies in which cardiolipin fatty acid composition was modified by dietary intervention and observations from human heart failure. Reduction of cardiolipin (18:2)₄ content in mitochondria is associated with impairment of mitochondrial function. Feeding rats with a diet extremely low or deficient in linoleic acid (18:2) resulted in decreased cardiolipin (18:2)₄ in cardiac mitochondria and a large reduction in COX activity [107–109]. In a rat model of heart failure, a diet rich in linoleic acid increased cardiolipin (18:2)₄ in cardiac mitochondria, improved mitochondrial function, and reduced mortality [110,111]. Rats fed a diet rich in oleic acid (18:1) showed increased cardiolipin (18:1)₂, (18:2)₂ and decreased cardiolipin (18:2)₄ in liver mitochondria, with comitant decrease in mitochondrial state 3 respiration [112]. In a rat model, lower cardiolipin (18:2)₄ content of skeletal muscle mitochondria was significantly associated with decreased COX activity [113]. In a mouse model, a diet rich in docosahexaenoic acid (DHA) (22:6) led to replacement of cardiolipin (18:2)₄ with tetra-docosahexaenoyl (22:6)₄ cardiolipin in the heart, with impaired activity of ETC complexes I, IV, V, and I + III [114]. Mitochondrial respiratory activity was recovered with reintroduction of linoleic acid via fusion of phospholipid vesicles to mitochondria isolated from DHA-fed mice [114]. In humans, the relationship of aging with cardiolipin species in tissues remains a major gap in knowledge. In patients with heart failure, the tetra-linoleoyl cardiolipin (18:2)₄ content of mitochondria in left ventricle is significantly lower than those without heart failure [115,116].

Additional evidence for the importance of tetra-linoleoyl cardiolipin (18:2)₄ in mitochondrial function comes from studies of aging in animal models and Barth syndrome, as discussed in further detail below.

The cardiolipin content of mitochondria decreases with aging

Aging is associated with a decrease in the cardiolipin content of mitochondria and changes in the fatty acid composition of cardiolipin in a variety of tissues. The cardiolipin content of mitochondria decreases with age in rat heart [115,117–125], mouse heart [126], fish heart [127], rat brain [128–130], monkey brain [131]; rat liver [132–136], rat kidney [132], mouse skeletal muscle [137], and fish skeletal muscle [138]. The cardiolipin content of mitochondria decreases with age in human skin [139]. Animal studies show that the age-related decrease in mitochondrial cardiolipin content is accompanied by a decrease in tetra-linoleoyl cardiolipin (18:2)₄ accompanied by substitution of other fatty acids in place of linoleic acid (18:2) in the heart, such as arachidonic and docosahexaenoic acids [115,126]. Other aging changes that have been described in the mitochondria of rat heart include a decline in complex I activity, state 3 respiration [140], and complex IV activity (cytochrome *c* oxidase) [115,121], and decreased carnitine-acyltranslocase (CAT) activity [120,141,142]. Whether the loss of tetra-linoleoyl cardiolipin (18:2)₄ with age is due to decreased availability of linoleic acid or increased degradation of tetra-linoleoyl cardiolipin is not clear.

Tetra-linoleoyl cardiolipin as a substrate for peroxidation

Cardiolipin is susceptible to peroxidation due to its high content of polyunsaturated fatty acid chains containing one or more 1,4 *cis,cis* pentadiene structures in close proximity to sites where ROS are produced. Peroxidation of cardiolipin leads to a conformation change in both the glycerol headgroup and fatty acid chains, inducing a tilt to the molecule and loss of symmetry and change in thickness of the lipid bilayer [143]. Aging is associated with an increase in oxidized cardiolipin species in the rat heart [140,144] and fish heart [127]. Higher oxidized cardiolipin content is associated with impaired ETC function [140].

Linoleic acid in cardiolipin is a substrate for cytochrome *c*-mediated

peroxidation [145], generating multiple esterified oxidized linoleic acid species including hydroperoxy-, hydroxy- and keto-octadecadienoic acids and epoxy-octadecenoic acids [146,147]. Peroxidation of linoleic acid moieties in cardiolipin is reported to release cytochrome *c* to the cytoplasm where it plays a key role in triggering apoptosis [148–150]. Oxidized linoleic acid species in cardiolipin are also substrates for enzyme-catalysed hydrolysis, releasing bioactive unesterified acids [145]. While specific actions and relevance of these oxidized lipids in skeletal muscle have not been identified, these products are reported to evoke inflammation and nociceptive responses in other tissues [151,152]. Thus, cardiolipin linoleic acid peroxidation could potentially impact sarcopenia by depleting tetra-linoleoyl cardiolipin with accompanying alterations in inflammatory signaling and apoptosis.

Implications of Barth syndrome for sarcopenia

Barth syndrome is a rare, X-linked recessive disorder due to mutations in the tafazzin (*TAZ*) gene locus. Barth syndrome is characterized by impaired cardiolipin synthesis and remodeling, altered cardiolipin species, decreased cardiolipin content of mitochondria, and abnormal mitochondrial structure with a clinical phenotype of cardiomyopathy, skeletal muscle weakness, neutropenia, and organic aciduria [153–155]. The cardiolipin alterations are associated with abnormalities of the mitochondrial inner membrane, impaired ETC function, decreased ATP production, inhibition of the tricarboxylic acid (TCA) cycle, disrupted β -oxidation of fatty acids, decreased plasma LPC, elevated plasma acylcarnitines, and accumulation of organic acids [155–157]. Skeletal muscle from Barth syndrome patients shows extremely low content of cardiolipin, primarily due to deficiency of tetra-linoleoyl cardiolipin (18:2)₄ [158]. In mouse models of Barth syndrome that involve *TAZ* knockdown or depletion, there is a large decrease in tetra-linoleoyl cardiolipin (18:2)₄ from cardiac and skeletal muscle, accumulation of cardiolipin species with aberrant fatty acid chains, and disrupted cristae in mitochondria [159,160]. Under normal conditions, cardiolipin has slow turnover [53,83,84,161], however, in Barth syndrome, cardiolipin loses its association with membrane proteins and supercomplexes and is rapidly degraded [161]. Patients with Barth syndrome have severe exercise intolerance and greatly reduced skeletal muscle O₂ utilization [162]. Studies using ³¹P-MRS show that children, adolescents, and young adults with Barth syndrome have impaired skeletal muscle mitochondrial oxidative capacity [163].

Barth syndrome has some salient features in common with sarcopenia and aging, such as a reduction in cardiolipin content, decrease in tetra-linoleoyl cardiolipin (18:2)₄, impaired β -oxidation of fatty acids, reduced exercise tolerance, reduced skeletal muscle O₂ utilization, impaired skeletal muscle mitochondrial oxidative capacity, and a plasma metabolomic profile characterized by increased plasma acylcarnitines and decreased plasma LPC.

The tetra-linoleoyl cardiolipin depletion hypothesis and other pathways implicated in sarcopenia

Since cardiolipin plays a fundamental role in mitochondrial function, alterations in cardiolipin could affect other hypothesized pathways in the pathogenesis of sarcopenia such as anorexia of aging, altered proteostasis, and inflammation. The insufficient dietary intake that occurs with anorexia of aging [28,164] could involve qualitative changes in diet and an inadequate intake of essential fatty acids and other factors required for cardiolipin and optimal mitochondrial function. Alterations in cardiolipin could impair the role that cardiolipin plays in protein folding and stabilization and impair proteostasis [165,166]. Inadequate cardiolipin content and composition are associated with impairment of ETC function and contribute to excessive generation of ROS and inflammation [31,32].

Cardiolipin as a potential therapeutic target in sarcopenia

The cardiolipin content and composition of skeletal muscle could potentially be targeted through changes in lifestyle such as exercise, changes in diet or through weight loss, and by pharmacological intervention. In the rat model, exercise increased cardiolipin content in heart and enhanced cytochrome oxidase activity in mitochondria [167,168]. Endurance training increased cardiolipin content in red gastrocnemius muscle in the rat [169]. Twelve weeks of exercise training increased the cardiolipin content of vastus lateralis muscle in older sedentary adults [170]. In older overweight to obese individuals, 16 weeks of moderate exercise increased cardiolipin content in vastus lateralis muscle, while both 16 weeks of either caloric restriction or exercise induced cardiolipin remodeling in muscle with a significant increase in tetra-linoleoyl cardiolipin (18:2)₄ [171]. Twelve weeks of daily moderate-intensity exercise combined with weight loss increased mitochondrial cardiolipin content and mitochondrial oxidative enzymes in vastus lateralis muscle in patients with type 2 diabetes [172]. In patients who underwent bariatric surgery, six months of moderate exercise induced cardiolipin remodeling with an increase of tetra-linoleoyl cardiolipin (18:2)₄ content in vastus lateralis muscle [173].

In rodent models, the tetra-linoleoyl cardiolipin (18:2)₄ content of heart can be decreased by restricting linoleic acid and increasing oleic acid [112] or docosahexaenoic acid in the diet [114]. Conversely the tetra-linoleoyl cardiolipin (18:2)₄ content of heart can be increased by a diet rich in linoleic acid [110,111]. Humans are unable to synthesize linoleic acid (18:2n-6), thus, this essential omega-6 fatty acid must be obtained from the diet. Linoleic acid is widely available in a concentrated form from plant seed oils such as soybean, corn, and cottonseed oils, and is the primary polyunsaturated fatty acid in the US diet [174]. Among US adults, mean linoleic acid intake tends to decrease with age and is lower among those aged ≥ 70 years compared with those aged 20–70 years [174]. However, current U.S. LA intakes are several-fold higher than can be achieved from pre-industrial diets without added liquid vegetable oils [175].

Recently, a family of small, synthetic cell-penetrating peptides, also known as Szeto-Schiller (SS) peptides, were described that selectively target cardiolipin in the inner mitochondrial membrane [176,177]. These peptides bind to cardiolipin and increase coupling efficiency while reducing generation of ROS [177]. The SS peptide, SS-31 (elamipretide) has shown promise in rejuvenating mitochondrial bioenergetics in phase I-II trials for heart failure [178] and primary mitochondrial myopathy [179]. There are over a dozen enzymes involved in the lipid pathways leading to cardiolipin synthesis and remodeling; these enzymes are potential therapeutic targets for modulating cardiolipin content in skeletal muscle and other tissues.

Gaps in knowledge and unanswered questions

While we have hypothesized that cardiolipin plays a major role in the pathogenesis of sarcopenia, there are many unanswered questions. Do the content and fatty acid composition of cardiolipin change in human skeletal muscle with aging? Does the susceptibility to cardiolipin to peroxidation increase in human skeletal muscle with sarcopenia? What is the pathway by which cardiolipin fatty acid composition is modulated in humans by diet and physical activity? What is the optimal range of linoleic acid intake to maintain tetra-linoleoyl cardiolipin across the lifespan? Is the content and fatty acid composition of cardiolipin in human skeletal muscle related to muscle strength or physical performance? What is the relationship between cardiolipin fatty acid content and composition in human skeletal muscle with mitochondrial oxidative capacity? Addressing these questions may reveal novel therapeutic targets for the prevention of age-related sarcopenia.

Conclusion

In summary, we propose the hypothesis that the depletion of tetra-linoleoyl cardiolipin plays a major role in the pathogenesis of sarcopenia. This is a testable hypothesis, that if corroborated, will provide new leads for the prevention and treatment of sarcopenia.

Support

The National Institutes of Health R01 AG027012, R01 EY024596, R56 AG052973, R01 AG057723 and the Intramural Branch of the National Institute on Aging, Baltimore, Maryland.

Acknowledgement

The authors declare no conflicts of interest.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.mehy.2019.04.015>.

References

- [1] Evans WJ. Skeletal muscle loss: cachexia, sarcopenia, and inactivity. *Am J Clin Nutr* 2010;91(suppl):1123–7.
- [2] Walston JD. Sarcopenia in older adults. *Curr Opin Rheumatol* 2012;24:623–7.
- [3] Shafiee G, Keshkar A, Soltani A, Ahadi Z, Larjani B, Heshmat R. Prevalence of sarcopenia in the world: a systematic review and meta-analysis of general population studies. *J Diabetes Metab Disord* 2017;16:21.
- [4] Peterson SJ, Braunschweig CA. Prevalence of sarcopenia and associated outcomes in the clinical setting. *Nutr Clin Pract* 2016;31:40–8.
- [5] Carmeli E, Coleman R, Reznick AZ. The biochemistry of aging muscle. *Exp Gerontol* 2002;37:477–89.
- [6] Doherty TJ. Aging and sarcopenia. *J Appl Physiol* 2003;95:1717–27.
- [7] Lexell J, Taylor CC, Sjöström M. What is the cause of the ageing atrophy? Total number, size and proportion of different fiber types studied in whole vastus lateralis muscle from 15- to 83-year-old men. *J Neurol Sci* 1988;84:275–94.
- [8] Frontera WR, Hughes VA, Lutz KJ, Evans WJ. A cross-sectional study of muscle strength and mass in 45- to 78-yr-old men and women. *J Appl Physiol* 1991;71:644–50.
- [9] Visser M, Kritchevsky SB, Goodpaster BH, Newman AB, Nevitt M, Stamm E, Harris TB. Leg muscle mass and composition in relation to lower extremity performance in men and women aged 70 to 79: the Health, Aging and Body Composition Study. *J Am Geriatr Soc* 2002;50:897–904.
- [10] Janssen I, Heymsfield SB, Ross R. Low relative skeletal muscle mass (sarcopenia) in older persons is associated with functional impairment and physical disability. *J Am Geriatr Soc* 2002;50:889–96.
- [11] Wolfson L, Judge J, Whipple R, King M. Strength is a major factor in balance, gait, and the occurrence of falls. *J Gerontol A Biol Sci Med Sci* 1995;50:64–7.
- [12] Tinetti ME, Williams CS. Falls, injuries due to falls, and the risk of admission to a nursing home. *N Engl J Med* 1997;337:1279–84.
- [13] Baumgartner RN, Koehler KM, Gallagher D, Romero L, Heymsfield SB, Ross RR, Garry PJ, Lindeman RD. Epidemiology of sarcopenia among the elderly in New Mexico. *Am J Epidemiol* 1998;147:755–63.
- [14] Melton III LJ, Khosla S, Crowson CS, O'Connor MK, O'Fallon WM, Riggs BL. Epidemiology of sarcopenia. *J Am Geriatr Soc* 2000;48:625–30.
- [15] Goodpaster BH, Park SW, Harris TB, et al. The loss of skeletal muscle strength, mass, and quality in older adults: the health, aging and body composition study. *J Gerontol A Biol Sci Med Sci* 2006;61:1059–64.
- [16] Moore AZ, Caturegli G, Metter EJ, et al. Difference in muscle quality over the adult life span and biological correlates in the Baltimore Longitudinal Study of Aging. *J Am Geriatr Soc* 2014;62:230–6.
- [17] Gonzalez-Freire M, de Cabo R, Studenski SA, Ferrucci L. The neuromuscular junction: aging at the crossroad between nerves and muscle. *Front Aging Neurosci* 2014;6:208.
- [18] Piasecki M, Ireland A, Piasecki J, et al. Failure to expand the motor unit size to compensate for declining motor unit numbers distinguishes sarcopenic from non-sarcopenic older men. *J Physiol* 2018;596:1627–37.
- [19] Mitchell WK, Williams J, Atherton P, Larvin M, Lund J, Narici M. Sarcopenia, dynapenia, and the impact of advancing age on human skeletal muscle size and strength; a quantitative review. *Front Physiol* 2012;3:260.
- [20] Rantanen T, Guralnik JM, Foley D, Masaki K, Leveille S, Curb JD, White L. Midlife hand grip strength as a predictor of old age disability. *JAMA* 1999;281:558–60.
- [21] Giampaoli S, Ferrucci L, Cecchi F, et al. Hand-grip strength predicts incident disability in non-disabled older men. *Age Ageing* 1999;28:283–8.
- [22] Rantanen T, Volpato S, Ferrucci L, Heikkinen E, Fried LP, Guralnik JM. Handgrip strength and cause specific and total mortality in older disabled women: exploring

- the mechanism. *J Am Geriatr Soc* 2003;51:636–41.
- [23] Tang TC, Hwang AC, Liu LK, et al. FNIIH-defined sarcopenia predicts adverse outcomes among community-dwelling older people in Taiwan: results from I-Lan Longitudinal Aging Study. *J Gerontol A Biol Sci Med Sci* 2018;73:828–34.
- [24] Janssen I. The epidemiology of sarcopenia. *Clin Geriatr Med* 2011;27:355–63.
- [25] Robinson SM, Reginster JY, Rizzoli R, et al. Does nutrition play a role in the prevention and management of sarcopenia? *Clin Nutr* 2018;37:1121–32.
- [26] Vitale G, Cesari M, Mari D. Aging of the endocrine system and its potential impact on sarcopenia. *Eur J Intern Med* 2016;35:10–5.
- [27] Landi F, Picca A, Calvani R, Marzetti E. Anorexia of aging: assessment and management. *Clin Geriatr Med* 2017;33:315–23.
- [28] Morley JE. Anorexia of ageing: a key component in the pathogenesis of both sarcopenia and cachexia. *J Cachexia Sarcopenia Muscle* 2017;8:523–6.
- [29] Snijders T, Parise G. Role of muscle stem cells in sarcopenia. *Curr Opin Clin Nutr Metab Care* 2017;20:186–90.
- [30] Musci RV, Hamilton KL, Miller BF. Targeting mitochondrial function and proteostasis to mitigate dynapenia. *Eur J Appl Physiol* 2018;118:1–9.
- [31] Ferrucci L, Fabbri E. Inflammaging: chronic inflammation in ageing, cardiovascular disease, and frailty. *Nat Rev Cardiol* 2018;15:505–22.
- [32] Picca A, Calvani R, Bossola M, et al. Update on mitochondria and muscle aging: all wrong roads lead to sarcopenia. *Biol Chem* 2018;399:421–36.
- [33] Wajner M, Amaral AU. Mitochondrial dysfunction in fatty acid oxidation disorders: insights from human and animal studies. *Biosci Rep* 2015;36.
- [34] De Stefani D, Rizzuto R, Pozzan T. Enjoy the trip: calcium in mitochondria back and forth. *Annu Rev Biochem* 2016;85:161–92.
- [35] Dan Dunn J, Alvarez LA, Zhang X, Soldati T. Reactive oxygen species and mitochondria: a nexus of cellular homeostasis. *Redox Biol* 2015;6:472–85.
- [36] Chu CT, Ji J, Dagda RK, et al. Cardiolipin externalization to the outer mitochondrial membrane acts as an elimination signal for mitophagy in neuronal cells. *Nat Cell Biol* 2013;15:1197–205.
- [37] Shen Z, Li Y, Gasparski AN, Abeliovich H, Greenberg ML. Cardiolipin regulates mitophagy through the protein kinase C pathway. *J Biol Chem*. 2017 Feb 17;292(7):2916–23.
- [38] Gonzalez F, Schug ZT, Houtkooper RH, et al. Cardiolipin provides an essential activating platform for caspase-8 on mitochondria. *J Cell Biol* 2008;183:681–96.
- [39] López-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell* 2013;153:1194–217.
- [40] Peterson CM, Johansson DL, Ravussin E. Skeletal muscle mitochondria and aging: a review. *J Aging Res* 2012;2012.
- [41] Johnson ML, Robinson MM, Nair KS. Skeletal muscle aging and the mitochondrion. *Trends Endocrinol Metab* 2013;24:247–56.
- [42] Marzetti E, Calvani R, Cesari M, et al. Mitochondrial dysfunction and sarcopenia of aging: from signaling pathways to clinical trials. *Int J Biochem Cell Biol* 2013;45:2288–301.
- [43] Newgard CB, Pessin JE. Recent progress in metabolic signaling pathways regulating aging and life span. *J Gerontol A Biol Sci Med Sci* 2014;69(Suppl 1):S21–7.
- [44] Gonzalez-Freire M, de Cabo R, Bernier M, et al. Reconsidering the role of mitochondria in aging. *J Gerontol A Biol Sci Med Sci* 2015;70:1334–42.
- [45] Rooyackers OE, Adey DB, Ades PA, Nair KS. Effect of age on in vivo rates of mitochondrial protein synthesis in human skeletal muscle. *Proc Natl Acad Sci USA* 1996;93:15364–9.
- [46] Short KR, Bigelow ML, Kahl J, et al. Decline in skeletal muscle mitochondrial function with aging in humans. *Proc Natl Acad Sci USA* 2005;102:5618–23.
- [47] Hepple RT. Mitochondrial involvement and impact in aging skeletal muscle. *Front Aging Neurosci* 2014;6:211.
- [48] Kent JA, Fitzgerald LF. In vivo mitochondrial function in aging skeletal muscle: capacity, flux, and patterns of use. *J Appl Physiol* 1985;2016(121):996–1003.
- [49] Lanza IR, Bhagra S, Nair KS, Port JD. Measurement of human skeletal muscle oxidative capacity by 31P-MR spectroscopy: a cross-validation with in vitro measurements. *J Magn Reson Imag* 2011;34:1143–50.
- [50] Gonzalez-Freire M, Scalzo P, D'Agostino J, et al. Skeletal muscle ex vivo mitochondrial respiration parallels decline in vivo oxidative capacity, cardiorespiratory fitness, and muscle strength: the Baltimore Longitudinal Study of Aging. *Aging Cell* 2018;17(2).
- [51] Choi S, Reiter DA, Shardell M, et al. 31P magnetic resonance spectroscopy assessment of muscle bioenergetics as a predictor of gait speed in the Baltimore Longitudinal Study of Aging. *J Gerontol A Biol Sci Med Sci* 2016;71:1638–45.
- [52] Fabbri E, Chia CW, Spencer RG, et al. Insulin resistance is associated with reduced mitochondrial oxidative capacity measured by 31P-magnetic resonance spectroscopy in participants without diabetes from the Baltimore Longitudinal Study of Aging. *Diabetes* 2017;66:170–6.
- [53] Schlame M, Greenberg ML. Biosynthesis, remodeling and turnover of mitochondrial cardiolipin. *Biochim Biophys Acta* 2017;1862:3–7.
- [54] Ren M, Phoon CK, Schlame M. Metabolism and function of mitochondrial cardiolipin. *Prog Lipid Res* 2014;55:1–16.
- [55] Noël H, Pande SV. An essential requirement of cardiolipin for mitochondrial carnitine acylcarnitine translocase activity. Lipid requirement of carnitine acylcarnitine translocase. *Eur J Biochem* 1986;155:99–102.
- [56] Kashfi K, Mynatt RL, Park EA, Cook GA. Membrane microenvironment regulation of carnitine palmitoyltransferases I and II. *Biochem Soc Trans* 2011;39:833–7.
- [57] Ikon N, Ryan RO. Cardiolipin and mitochondrial cristae organization. *Biochim Biophys Acta* 2017;1859:1156–63.
- [58] Dudek J. Role of cardiolipin in mitochondrial signaling pathways. *Front Cell Dev Biol* 2017;5:90.
- [59] Gonzalez-Baro MR, Coleman RA. Mitochondrial acyltransferases and glycerophospholipid metabolism. *Biochim Biophys Acta* 2017;1862:49–55.
- [60] Coleman RA, Mashek DG. Mammalian triacylglycerol metabolism: synthesis, lipolysis, and signaling. *Chem Rev* 2011;111:6359–86.
- [61] Perrakis A, Moolenaar WH. Autotaxin: structure-function and signaling. *J Lipid Res* 2014;55:1010–8.
- [62] Xu A, Ahsanul Kabir Khan M, Chen F, Zhong Z, Chen HC, Song Y. Overexpression of autotaxin is associated with human renal cell carcinoma and bladder carcinoma and their progression. *Med Oncol* 2016;33:131.
- [63] Lyu L, Wang B, Xiong C, Zhang X, Zhang X, Zhang J. Selective export of autotaxin from the endoplasmic reticulum. *J Biol Chem* 2017;292:7011–22.
- [64] Rustenbeck I, Münster W, Lenzen S. Relation between accumulation of phospholipase A2 reaction products and Ca²⁺ release in isolated liver mitochondria. *Biochim Biophys Acta* 1996;1304:129–38.
- [65] Tsalouhidou S, Argyrou C, Theofilidis G, et al. Mitochondrial phospholipids of rat skeletal muscle are less polyunsaturated than whole tissue phospholipids: implications for protection against oxidative stress. *J Anim Sci* 2006;84:2818–25.
- [66] Pollard AK, Ortori CA, Stöger R, Barrett DA, Chakrabarti L. Mouse mitochondrial lipid composition is defined by age in brain and muscle. *Aging (Albany NY)* 2017;9:986–98.
- [67] Veyrat-Durebex C, Bocca C, Chupin S, et al. Metabolomics and lipidomics profiling of a combined mitochondrial plus endoplasmic reticulum fraction of human fibroblasts: a robust tool for clinical studies. *J Proteome Res* 2018;17:745–50.
- [68] Moessinger C, Kuerschner L, Spandl J, Shevchenko A, Thiele C. Human lysophosphatidylcholine acyltransferases 1 and 2 are located in lipid droplets where they catalyze the formation of phosphatidylcholine. *J Biol Chem* 2011;286:21330–9.
- [69] Guijas C, Rodríguez JP, Rubio JM, Balboa MA, Balsinde J. Phospholipase A2 regulation of lipid droplet formation. *Biochim Biophys Acta* 2014;1841:1661–71.
- [70] Bosma M. Lipid droplet dynamics in skeletal muscle. *Exp Cell Res* 2016;340:180–6.
- [71] Bersuker K, Olzmann JA. Establishing the lipid droplet proteome: mechanisms of lipid droplet protein targeting and degradation. *Biochim Biophys Acta Mol Cell Biol Lipids* 2017;1862:1166–77.
- [72] Leslie CC. Cytosolic phospholipase A₂: physiological function and role in disease. *J Lipid Res* 2015;56:1386–402.
- [73] Eismann T, Huber N, Shin T, et al. Peroxiredoxin-6 protects against mitochondrial dysfunction and liver injury during ischemia-reperfusion in mice. *Am J Physiol Gastrointest Liver Physiol* 2009;296:G266–74.
- [74] Fisher AB, Dodia C, Sorokina EM, Li H, Zhou S, Raabe T, Feinstein SI. A novel lysophosphatidylcholine acyl transferase activity is expressed by peroxiredoxin 6. *J Lipid Res* 2016;57:587–96.
- [75] Janikiewicz J, Szymański J, Malinska D, et al. Mitochondria-associated membranes in aging and senescence: structure, function, and dynamics. *Cell Death Dis* 2018;9:332.
- [76] Schmidt JA, Yvone GM, Brown WJ. Membrane topology of human AGPAT3 (LPAAT3). *Biochem Biophys Res Commun* 2010;397:661–7.
- [77] Bradley RM, Marvyn PM, Aristizabal Henao JJ, et al. Acylglycerophosphate acyltransferase 4 (AGPAT4) is a mitochondrial lysophosphatidic acid acyltransferase that regulates brain phosphatidylcholine, phosphatidylethanolamine, and phosphatidylinositol levels. *Biochim Biophys Acta* 2015;1851:1566–76.
- [78] Prasad SS, Garg A, Agarwal AK. Enzymatic activities of the human AGPAT isoform 3 and isoform 5: localization of AGPAT5 to mitochondria. *J Lipid Res* 2011;52:451–62.
- [79] Tamura Y, Harada Y, Nishikawa S, et al. Tam41 is a CDP-diacylglycerol synthase required for cardiolipin biosynthesis in mitochondria. *Cell Metab* 2013;17:709–18.
- [80] Mejia EM, Nguyen H, Hatch GM. Mammalian cardiolipin biosynthesis. *Chem Phys Lipids* 2014;179:11–6.
- [81] Zhang J, Guan Z, Murphy AN, et al. Mitochondrial phosphatase PTPMT1 is essential for cardiolipin biosynthesis. *Cell Metab* 2011;13:690–700.
- [82] Schlame M. Cardiolipin remodeling and the function of tafazzin. *Biochim Biophys Acta* 2013;1831:582–8.
- [83] Landriscina C, Megli FM, Quagliariello E. Turnover of fatty acids in rat liver cardiolipin: comparison with other mitochondrial phospholipids. *Lipids* 1976;11:61–6.
- [84] Wahjudi PN, Yee KJ, Martinez SR, et al. Turnover of nonessential fatty acids in cardiolipin from the rat heart. *J Lipid Res* 2011;52:2226–33.
- [85] Xu Y, Schlame M. The turnover of glycerol and acyl moieties of cardiolipin. *Chem Phys Lipids* 2014;179:17–24.
- [86] Pennington ER, Fix A, Sullivan EM, Brown DA, Kennedy A, Shaikh SR. Distinct membrane properties are differentially influenced by cardiolipin content and acyl chain composition in biomimetic membranes. *Biochim Biophys Acta* 2017;1859:257–67.
- [87] Renner LD, Weibel DB. Cardiolipin microdomains localize to negatively curved regions of *Escherichia coli* membranes. *Proc Natl Acad Sci USA* 2011;108:6264–9.
- [88] Paradies G, Paradies V, De Benedictis V, Ruggiero FM, Petrosillo G. Functional role of cardiolipin in mitochondrial bioenergetics. *Biochim Biophys Acta* 2014;1837:408–17.
- [89] Fry M, Green DE. Cardiolipin requirement for electron transfer in complex I and III of the mitochondrial respiratory chain. *J Biol Chem* 1981;256:1874–80.
- [90] Paradies G, Petrosillo G, Pistolesse M, Ruggiero FM. Reactive oxygen species affect mitochondrial electron transport complex I activity through oxidative cardiolipin damage. *Gene* 2002;286:135–41.
- [91] Gomez Jr B, Robinson NC. Phospholipase digestion of bound cardiolipin reversibly inactivates bovine cytochrome bc1. *Biochemistry* 1999;38:9031–8.
- [92] Lange C, Nett JH, Trumpower BL, Hunte C. Specific roles of protein-phospholipid interactions in the yeast cytochrome bc1 complex structure. *EMBO J* 2001;20:6591–600.
- [93] Zhang M, Mileykovskaya E, Dowhan W. Cardiolipin is essential for organization of complexes III and IV into a supercomplex in intact yeast mitochondria. *J Biol*

- Chem 2005;280:29403–8.
- [94] Sedláč E, Robinson NC. Phospholipase A(2) digestion of cardiolipin bound to bovine cytochrome c oxidase alters both activity and quaternary structure. *Biochemistry* 1999;38:14966–72.
- [95] Musatov A, Robinson NC. Bound cardiolipin is essential for cytochrome c oxidase proton translocation. *Biochimie* 2014;105:159–64.
- [96] Eble KS, Coleman WB, Hantgan RR, Cunningham CC. Tightly associated cardiolipin in the bovine heart mitochondrial ATP synthase as analyzed by 31P nuclear magnetic resonance spectroscopy. *J Biol Chem* 1990;265:19434–40.
- [97] Pebay-Peyroula E, Dahout-Gonzalez C, Kahn R, Trézéguet V, Lauquin GJ, Brandolin G. Structure of mitochondrial ADP/ATP carrier in complex with carboxyatractyloside. *Nature* 2003;426:39–44.
- [98] Duncan AL, Ruprecht JJ, Kunji ERS, Robinson AJ. Cardiolipin dynamics and binding to conserved residues in the mitochondrial ADP/ATP carrier. *Biochim Biophys Acta Biomembr* 2018;1860:1035–45.
- [99] Pfeiffer K, Gohil V, Stuart RA, Hunte C, Brandt U, Greenberg ML, Schägger H. Cardiolipin stabilizes respiratory chain supercomplexes. *J Biol Chem* 2003;278:52873–80.
- [100] Gohil VM, Hayes P, Matsuyama S, Schägger H, Schlame M, Greenberg ML. Cardiolipin biosynthesis and mitochondrial respiratory chain function are interdependent. *J Biol Chem* 2004;279:42612–8.
- [101] Koshkin V, Greenberg ML. Cardiolipin prevents rate-dependent uncoupling and provides osmotic stability in yeast mitochondria. *Biochem J* 2002;364:317–22.
- [102] Xu Y, Sutachan JJ, Plesken H, Kelley RI, Schlame M. Characterization of lymphoblast mitochondria from patients with Barth syndrome. *Lab Invest* 2005;85:823–30.
- [103] Fry M, Green DE. Cardiolipin requirement by cytochrome oxidase and the catalytic role of phospholipid. *Biochem Biophys Res Commun* 1980;93:1238–46.
- [104] Vik SB, Georgevich G, Capaldi RA. Diphosphatidylglycerol is required for optimal activity of beef heart cytochrome c oxidase. *Proc Natl Acad Sci USA* 1981;78:1456–60.
- [105] Maguire JJ, Tyurina YY, Mohammadyani D, et al. Known unknowns of cardiolipin signaling: the best is yet to come. *Biochim Biophys Acta Mol Cell Biol Lipids* 2017;1862:8–24.
- [106] Schlame M, Shanske S, Doty S, König T, Sculco T, DiMauro S, Blanck TJ. Microanalysis of cardiolipin in small biopsies including skeletal muscle from patients with mitochondrial disease. *J Lipid Res* 1999;40:1585–92.
- [107] Yamaoka S, Urade R, Kito M. Mitochondrial function in rats is affected by modification of membrane phospholipids with dietary sardine oil. *J Nutr* 1988;118:290–6.
- [108] Yamaoka S, Urade R, Kito M. Cardiolipin molecular species in rat heart mitochondria are sensitive to essential fatty acid-deficient dietary lipids. *J Nutr* 1990;120:415–21.
- [109] Yamaoka-Koseki S, Urade R, Kito M. Cardiolipins from rats fed different dietary lipids affect bovine heart cytochrome c oxidase activity. *J Nutr* 1991;121:956–8.
- [110] Chicco AJ, Sparagna GC, McCune SA, et al. Linoleate-rich high-fat diet decreases mortality in hypertensive heart failure rats compared with lard and low-fat diets. *Hypertension* 2008;52:549–55.
- [111] Mulligan CM, Sparagna GC, Le CH, et al. Dietary linoleate preserves cardiolipin and attenuates mitochondrial dysfunction in the failing rat heart. *Cardiovasc Res* 2012;94:460–8.
- [112] Monteiro JP, Pereira CV, Silva AM, et al. Rapeseed oil-rich diet alters hepatic mitochondrial membrane lipid composition and disrupts bioenergetics. *Arch Toxicol* 2013;87:2151–63.
- [113] Fajardo VA, McMeekin L, Saint C, LeBlanc PJ. Cardiolipin linoleic acid content and mitochondrial cytochrome c oxidase activity are associated in rat skeletal muscle. *Chem Phys Lipids* 2015;187:50–5.
- [114] Sullivan EM, Pennington ER, Sparagna GC, et al. Docosahexaenoic acid lowers cardiac mitochondrial enzyme activity by replacing linoleic acid in the phospholipidome. *J Biol Chem* 2018;293:466–83.
- [115] Sparagna GC, Chicco AJ, Murphy RC, et al. Loss of cardiac tetralinoleoyl cardiolipin in human and experimental heart failure. *J Lipid Res* 2007;48:1559–70.
- [116] Chatfield KC, Sparagna GC, Sucharov CC, et al. Dysregulation of cardiolipin biosynthesis in pediatric heart failure. *J Mol Cell Cardiol* 2014;74:251–9.
- [117] Nohl H, Krämer R. Molecular basis of age-dependent changes in the activity of adenine nucleotide translocase. *Mech Ageing Dev* 1980;14:137–44.
- [118] Lewin MB, Timiras PS. Lipid changes with aging in cardiac mitochondrial membranes. *Mech Ageing Dev* 1984;24:343–51.
- [119] McMillin JB, Taffet GE, Taegtmeier H, Hudson EK, Tate CA. Mitochondrial metabolism and substrate competition in the aging Fischer rat heart. *Cardiovasc Res* 1993;27:2222–8.
- [120] Paradies G, Ruggiero FM, Petrosillo G, Gadaleta MN, Quagliariello E. Carnitine-acylcarnitine translocase activity in cardiac mitochondria from aged rats: the effect of acetyl-L-carnitine. *Mech Ageing Dev* 1995;84:103–12.
- [121] Paradies G, Ruggiero FM, Petrosillo G, Quagliariello E. Age-dependent decline in the cytochrome c oxidase activity in rat heart mitochondria: role of cardiolipin. *FEBS Lett* 1997;406:136–8.
- [122] Paradies G, Petrosillo G, Gadaleta MN, Ruggiero FM. The effect of aging and acetyl-L-carnitine on the pyruvate transport and oxidation in rat heart mitochondria. *FEBS Lett* 1999;454:207–9.
- [123] Pepe S, Tsuchiya N, Lakatta EG, Hansford RG. PUFA and aging modulate cardiac mitochondrial membrane lipid composition and Ca²⁺ activation of PDH. *Am J Physiol* 1999;276:H149–58.
- [124] Savitha S, Panneerselvam C. Mitochondrial membrane damage during aging process in rat heart: potential efficacy of L-carnitine and DL alpha lipoic acid. *Mech Ageing Dev* 2006;127:349–55.
- [125] Lee HJ, Mayette J, Rapoport SI, Bazinet RP. Selective remodeling of cardiolipin fatty acids in the aged rat heart. *Lipids Health Dis* 2006;5:2.
- [126] Mulligan CM, Le CH, deMooy AB, Nelson CB, Chicco AJ. Inhibition of delta-6 desaturase reverses cardiolipin remodeling and prevents contractile dysfunction in the aged mouse heart without altering mitochondrial respiratory function. *J Gerontol A Biol Sci Med Sci* 2014;69:799–809.
- [127] Almáida-Pagán PF, de Costa J, Mendiola P, Tocher DR. Age-related changes in mitochondrial membrane composition of rainbow trout (*Oncorhynchus mykiss*) heart and brain. *Comp Biochem Physiol B Biochem Mol Biol* 2012;163:129–37.
- [128] Ruggiero FM, Cafagna F, Petruzella V, Gadaleta MN, Quagliariello E. Lipid composition in synaptic and nonsynaptic mitochondria from rat brains and effect of aging. *J Neurochem* 1992;59:487–91.
- [129] Sen T, Sen N, Jana S, Khan FH, Chatterjee U, Chakrabarti S. Depolarization and cardiolipin depletion in aged rat brain mitochondria: relationship with oxidative stress and electron transport chain activity. *Neurochem Int* 2007;50:719–25.
- [130] Petrosillo G, Matera M, Casanova G, Ruggiero FM, Paradies G. Mitochondrial dysfunction in rat brain with aging: Involvement of complex I, reactive oxygen species and cardiolipin. *Neurochem Int* 2008;53:126–31.
- [131] Lam SM, Chua GH, Li XJ, Su B, Shui G. Biological relevance of fatty acyl heterogeneity to the synaptic membrane dynamics of rhesus macaques during normative aging. *Oncotarget* 2016;7:55970–89.
- [132] Grinna LS. Age related changes in the lipids of the microsomal and the mitochondrial membranes of rat liver and kidney. *Mech Ageing Dev* 1977;6:197–205.
- [133] Vorbeck ML, Martin AP, Long Jr JW, Smith JM, Orr Jr. RR. Aging-dependent modification of lipid composition and lipid structural order parameter of hepatic mitochondria. *Arch Biochem Biophys* 1982;217:351–61.
- [134] Paradies G, Ruggiero FM. Effect of aging on the activity of the phosphate carrier and on the lipid composition in rat liver mitochondria. *Arch Biochem Biophys* 1991;284:332–7.
- [135] Hagen TM, Ingersoll RT, Wehr CM, et al. Acetyl-L-carnitine fed to old rats partially restores mitochondrial function and ambulatory activity. *Proc Natl Acad Sci USA* 1998;95:9562–6.
- [136] Hagen TM, Wehr CM, Ames BN. Mitochondrial decay in aging. Reversal through supplementation of acetyl-L-carnitine and N-tert-butyl-alpha-phenyl-nitron. *Ann NY Acad Sci* 1998;854:214–23.
- [137] Zhang X, Trevino MB, Wang M, et al. Impaired mitochondrial energetics characterize poor early recovery of muscle mass following hind limb unloading in old mice. *J Gerontol A Biol Sci Med Sci* 2018;73:1313–22.
- [138] Suleimanova RR, Hudz EA, Melnychuk DO, Kalachniuk LH. Age-related changes phospholipids of sterlet in liver and dorsal muscles. *Ukr Biochem J* 2017;89:71–5.
- [139] Maftah A, Ratinaud MH, Dumas M, Bonté F, Meybeck A, Julien R. Human epidermal cells progressively lose their cardiolipins during ageing without change in mitochondrial transmembrane potential. *Mech Ageing Dev* 1994;77:83–96.
- [140] Petrosillo G, Matera M, Moro N, Ruggiero FM, Paradies G. Mitochondrial complex I dysfunction in rat heart with aging: critical role of reactive oxygen species and cardiolipin. *Free Radic Biol Med* 2009;46:88–94.
- [141] Paradies G, Ruggiero FM, Gadaleta MN, Quagliariello E. The effect of aging and acetyl-L-carnitine on the activity of the phosphate carrier and on the phospholipid composition in rat heart mitochondria. *Biochim Biophys Acta* 1992;1103:324–6.
- [142] Paradies G, Ruggiero FM, Petrosillo G, Gadaleta MN, Quagliariello E. Effect of aging and acetyl-L-carnitine on the activity of cytochrome oxidase and adenine nucleotide translocase in rat heart mitochondria. *FEBS Lett* 1994;350:213–5.
- [143] Vähäheikkilä M, Peltomaa T, Róg T, Vazdar M, Pöyry S, Vattulainen I. How cardiolipin peroxidation alters the properties of the inner mitochondrial membrane? *Chem Phys Lipids* 2018;214:15–23.
- [144] Lesnefsky EJ, Hoppel CL. Cardiolipin as an oxidative target in cardiac mitochondria in the aged rat. *Biochim Biophys Acta* 2008;1777:1020–7.
- [145] Tyurina YY, Poloyac SM, Tyurin VA, et al. A mitochondrial pathway for biosynthesis of lipid mediators. *Nat Chem* 2014;6:542–52.
- [146] Tyurina YY, Dominguez RM, Tyurin VA, et al. Characterization of cardiolipins and their oxidation products by LC-MS analysis. *Chem Phys Lipids* 2014;179:3–10.
- [147] Yin H, Zhu M. Free radical oxidation of cardiolipin: chemical mechanisms, detection and implication in apoptosis, mitochondrial dysfunction and human diseases. *Free Radic Res* 2012;46:959–74.
- [148] Kagan VE, Tyurin VA, Jiang J, et al. Cytochrome c acts as a cardiolipin oxygenase required for release of proapoptotic factors. *Nat Chem Biol* 2005;1:223–32.
- [149] Kagan VE, Tyurina YY, Bayir H, et al. The “pro-apoptotic genes” get out of mitochondria: oxidative lipidomics and redox activity of cytochrome c/cardiolipin complexes. *Chem Biol Interact* 2006;163:15–28.
- [150] Kagan VE, Chu CT, Tyurina YY, Cheikhi A, Bayir H. Cardiolipin asymmetry, oxidation and signaling. *Chem Phys Lipids* 2014;179:64–9.
- [151] Alsalem M, Wong A, Mills P, et al. The contribution of the endogenous TRPV1 ligands 9-HODE and 13-HODE to nociceptive processing and their role in peripheral inflammatory pain mechanisms. *Br J Pharmacol* 2013;168:1961–74.
- [152] Hattori T, Obinata H, Ogawa A, et al. G2A plays proinflammatory roles in human keratinocytes under oxidative stress as a receptor for 9-hydroxyoctadecadienoic acid. *J Invest Dermatol* 2008;128:1123–33.
- [153] Barth PG, Scholte HR, Berden JA, et al. An X-linked mitochondrial disease affecting cardiac muscle, skeletal muscle and neutrophil leucocytes. *J Neurol Sci* 1983;62:327–55.
- [154] Kelley RI, Cheatham JP, Clark BJ, et al. X-linked dilated cardiomyopathy with neutropenia, growth retardation, and 3-methylglutaconic aciduria. *J Pediatr* 1991;119:738–47.
- [155] Ikon N, Ryan RO. Barth syndrome: connecting cardiolipin to cardiomyopathy. *Lipids* 2017;52:99–108.

- [156] McKenzie M, Lazarou M, Thorburn DR, Ryan MT. Mitochondrial respiratory chain supercomplexes are destabilized in Barth syndrome patients. *J Mol Biol* 2006;361:462–9.
- [157] Sandlers Y, Mercier K, Pathmasiri W, et al. Metabolomics reveals new mechanisms for pathogenesis in Barth syndrome and introduces novel roles for cardiolipin in cellular function. *PLoS One* 2016;11.
- [158] Schlame M, Kelley RI, Feigenbaum A, et al. Phospholipid abnormalities in children with Barth syndrome. *J Am Coll Cardiol* 2003;42:1994–9.
- [159] Acehan D, Vaz F, Houtkooper RH, et al. Cardiac and skeletal muscle defects in a mouse model of human Barth syndrome. *J Biol Chem* 2011;286:899–908.
- [160] Soustek MS, Falk DJ, Mah CS, et al. Characterization of a transgenic short hairpin RNA-induced murine model of tafazzin deficiency. *Hum Gene Ther* 2011;22:865–71.
- [161] Xu Y, Phoon CK, Berno B, et al. Loss of protein association causes cardiolipin degradation in Barth syndrome. *Nat Chem Biol* 2016;12:641–7.
- [162] Spencer CT, Byrne BJ, Bryant RM, et al. Impaired cardiac reserve and severely diminished skeletal muscle O₂ utilization mediate exercise intolerance in Barth syndrome. *Am J Physiol Heart Circ Physiol* 2011;301:H2122–9.
- [163] Bashir A, Bohnert KL, Reeds DN, et al. Impaired cardiac and skeletal muscle bioenergetics in children, adolescents, and young adults with Barth syndrome. *Physiol Rep* 2017;5(3). pii: e13130.
- [164] Landi F, Calvani R, Tosato M, et al. Anorexia of aging: risk factors, consequences, and potential treatments. *Nutrients* 2016;8:69.
- [165] Laganowsky A, Reading E, Allison TM, et al. Membrane proteins bind lipids selectively to modulate their structure and function. *Nature* 2014;510:172–5.
- [166] Ryan T, Bamm VV, Stykel MG, et al. Cardiolipin exposure on the outer mitochondrial membrane modulates α -synuclein. *Nat Commun* 2018;9:817.
- [167] Kalofoutis A, Lekakis J, Miras C. Heart mitochondrial and microsomal phospholipid fluctuations induced by chronic exercise in rats. *Int J Biochem* 1981;13:195–9.
- [168] Chicco AJ, McCune SA, Emter CA, et al. Low-intensity exercise training delays heart failure and improves survival in female hypertensive heart failure rats. *Hypertension* 2008;51:1096–102.
- [169] Górski J, Zendzian-Piotrowska M, de Jong YF, Niklińska W, Glatz JF. Effect of endurance training on the phospholipid content of skeletal muscles in the rat. *Eur J Appl Physiol Occup Physiol* 1999;79:421–5.
- [170] Menshikova EV, Ritov VB, Fairfull L, Ferrell RE, Kelley DE, Goodpaster BH. Effects of exercise on mitochondrial content and function in aging human skeletal muscle. *J Gerontol A Biol Sci Med Sci* 2006;61:534–40.
- [171] Menshikova EV, Ritov VB, Dube JJ, et al. Calorie restriction-induced weight loss and exercise have differential effects on skeletal muscle mitochondria despite similar effects on insulin sensitivity. *J Gerontol A Biol Sci Med Sci* 2017;73:81–7.
- [172] Toledo FG, Menshikova EV, Ritov VB, et al. Effects of physical activity and weight loss on skeletal muscle mitochondria and relationship with glucose control in type 2 diabetes. *Diabetes* 2007;56:2142–7.
- [173] Coen PM, Menshikova EV, Distefano G, et al. Exercise and weight loss improve muscle mitochondrial respiration, lipid partitioning, and insulin sensitivity after gastric bypass surgery. *Diabetes* 2015;64:3737–50.
- [174] Raatz SK, Conrad Z, Jahns L. Trends in linoleic acid intake in the United States adult population: NHANES 1999–2014. *Prostaglandins Leukot Essent Fatty Acids* 2018;133:23–8.
- [175] Ramsden CE, Zamora D, Majchrzak-Hong S, et al. Re-evaluation of the traditional diet-heart hypothesis: analysis of recovered data from Minnesota Coronary Experiment (1968–73). *BMJ* 2016;353.
- [176] Birk AV, Liu S, Soong Y, et al. The mitochondrial-targeted compound SS-31 re-energizes ischemic mitochondria by interacting with cardiolipin. *J Am Soc Nephrol* 2013;24:1250–61.
- [177] Szeto HH, Liu S. Cardiolipin-targeted peptides rejuvenate mitochondrial function, remodel mitochondria, and promote tissue regeneration during aging. *Arch Biochem Biophys* 2018;660:137–48.
- [178] Daubert MA, Yow E, Dunn G, et al. Novel mitochondria-targeting peptide in heart failure treatment: a randomized, placebo-controlled trial of elamipretide. *Circ Heart Fail* 2017;10. pii: e004389.
- [179] Karaa A, Haas R, Goldstein A, Vockley J, Weaver WD, Cohen BH. Randomized dose-escalation trial of elamipretide in adults with primary mitochondrial myopathy. *Neurology* 2018;90:e1212–21.