

Targeted Treatment of Individuals With Psychosis Carrying a Copy Number Variant Containing a Genomic Triplication of the Glycine Decarboxylase Gene

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ABSTRACT

BACKGROUND: The increased mutational burden for rare structural genomic variants in schizophrenia and other neurodevelopmental disorders has so far not yielded therapies targeting the biological effects of specific mutations. We identified two carriers (mother and son) of a triplication of the gene encoding glycine decarboxylase, *GLDC*, presumably resulting in reduced availability of the *N*-methyl-D-aspartate receptor coagonists glycine and D-serine and *N*-methyl-D-aspartate receptor hypofunction. Both carriers had a diagnosis of a psychotic disorder.

METHODS: We carried out two double-blind, placebo-controlled clinical trials of *N*-methyl-D-aspartate receptor augmentation of psychotropic drug treatment in these two individuals. Glycine was used in the first clinical trial, and D-cycloserine was used in the second one.

RESULTS: Glycine or D-cycloserine augmentation of psychotropic drug treatment each improved psychotic and mood symptoms in placebo-controlled trials.

CONCLUSIONS: These results provide two independent proof-of-principle demonstrations of symptom relief by targeting a specific genotype and explicitly link an individual mutation to the pathophysiology of psychosis and treatment response.

Keywords: Bipolar disorder, Copy number variant, Genetics, Glycine decarboxylase, NMDAR hypofunction, Schizophrenia

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Individually rare structural variants of relatively recent evolutionary origin such as copy number variants (CNVs) collectively account for an increased mutational burden for schizophrenia and other neurodevelopmental disorders (e.g., autism spectrum disorders, intellectual disability, epilepsy, bipolar disorder [to a lesser extent]). The most recurrent include microdeletions and microduplications, with odds ratios for phenotypic expression ranging from 2 to greater than 60, and have effect sizes much larger than those associated with common genetic risk variants (1–21). Structural variants and CNVs can be large and involve many genes, and their underlying structure can be complex (22). Because pleiotropic clinical effects are the norm, discoveries based on any single mutation are potentially relevant to other individuals who carry the same mutation or mutations that affect the same

biological pathways even if the clinical phenotype differs. The fact that shared molecular mechanisms are implicated in a range of neurodevelopmental disorders (23–26) suggests that a genotype-first approach (27–31) may be more instructive about pathophysiology and potential treatments than a disease-oriented approach. CNV loci continue to be linked to cognitive phenotypes (32,33). The challenge is to link mutations in specific genes to the underlying disease biology (34), which in turn can be translated into targeted treatment interventions with positive therapeutic effects in appropriately selected patients (35–39).

We identified several CNVs spanning 9p24.1 in a proband and his mother, who presented with DSM-IV (40) diagnoses of schizoaffective disorder and bipolar disorder with psychotic features, respectively (Figure 1A); this structural rearrangement

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seems to segregate with psychosis in this family (Figure 1B). The rearrangement was confirmed as a de novo event in the mother (Supplemental Figure S1) (41). The complete architecture of this complex rearrangement and the proposed DNA replicative/repair mechanism underlying its formation are described in Grochowski *et al.* (42). See the Supplement for details about the CNV.

Although several of the genes in the CNV may be potentially relevant to the development of neuropsychiatric disorders (see Supplement), *GLDC*, the gene encoding glycine decarboxylase, is particularly compelling because it codes for the enzyme that catabolizes glycine, a precursor of D-serine, both of which are coagonists at the *N*-methyl-D-aspartate receptor (NMDAR) (43). Triplication of *GLDC* would be expected to increase glycine catabolism, resulting in low levels of brain glycine and D-serine. Reduced availability of these two NMDAR coagonists would result in NMDAR hypofunction, which is associated with psychotic disorders (44–47). The *GLDC* triplication seemed fortuitously amenable to a treatment intervention tailored to normalizing the biology hypothesized to be affected by the mutation. Therefore, we undertook a proof-of-principle clinical trial to determine whether augmentation of usual psychotropic drug treatment with glycine, a full agonist at the NMDAR glycine modulatory site (GMS), reduced psychotic and mood symptoms in the two carriers of the *GLDC* triplication. Due to the encouraging results of the glycine augmentation trial, we undertook a clinical trial with D-cycloserine (DCS), a relatively selective partial agonist at the GMS at low doses (48,49).

METHODS AND MATERIALS

Subjects

Two carriers of the 9p24.1 CNV participated in the clinical trials: the proband (subject 3363) and his mother (subject

5459). Demographic information and details of the study designs and methods are included below and in the Supplement.

Study Design

Both studies were approved by institutional review boards at McLean Hospital and Partners Healthcare. Subjects provided written informed consent.

Glycine Augmentation Clinical Trial

Procedures. Symptom severity and treatment side effects were monitored at least weekly; formal clinical ratings were carried out every 2 weeks blinded to drug condition using the following primary instruments: Brief Psychiatric Rating Scale (BPRS), Positive and Negative Syndrome Scale, Young Mania Rating Scale, Hamilton Depression Scale, Columbia–Suicide Severity Rating Scale, and Clinical Global Impression Scale (50–55). Motor abnormalities were assessed at baseline and at the end of each treatment arm blinded to condition (56,57). Plasma concentrations of small and large amino acids, kynurenine (KYN), kynurenic acid (KYNA), quinolinic acid, and homocysteine were obtained at baseline and during week 6 of each arm of the short-term glycine trial (see Supplement). All baseline procedures were carried out in person; some clinical assessments and movement disorder exams were carried out using a secure form of video conferencing because the subjects were not local.

Short-term Glycine Augmentation Trial. Both subjects were maintained on stable doses of psychotropic medications during the short-term trial (see Supplement). The design was a double-blind random-order glycine–placebo crossover followed by open-label glycine. Each of these 3 arms lasted 6 weeks, separated by 2 weeks to wash out treatment effects from the previous arm (58). During each arm, each subject

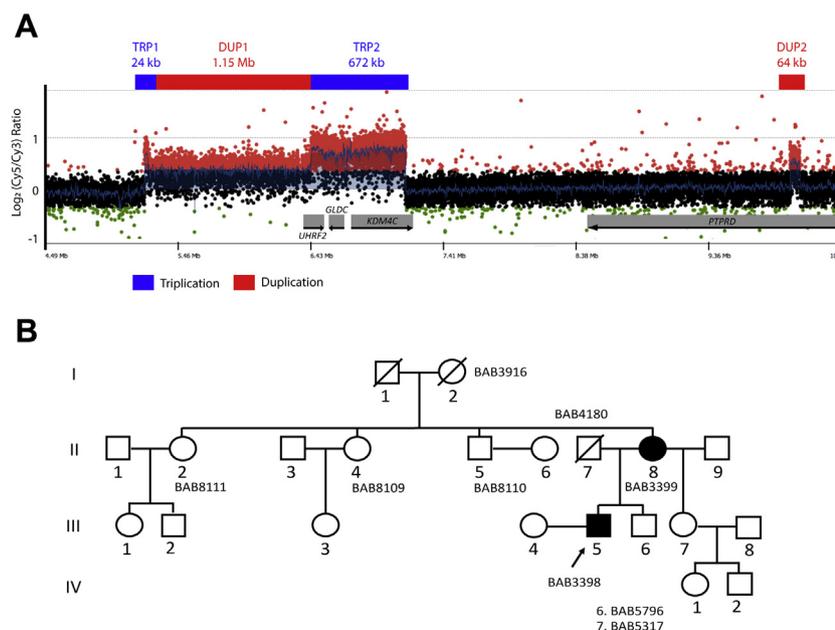


Figure 1. Structure of the 9p24.1 duplication (DUP)–triplication (TRP) (A) and pedigree (B).

Treatment of Carriers of a *GLDC* Triplication

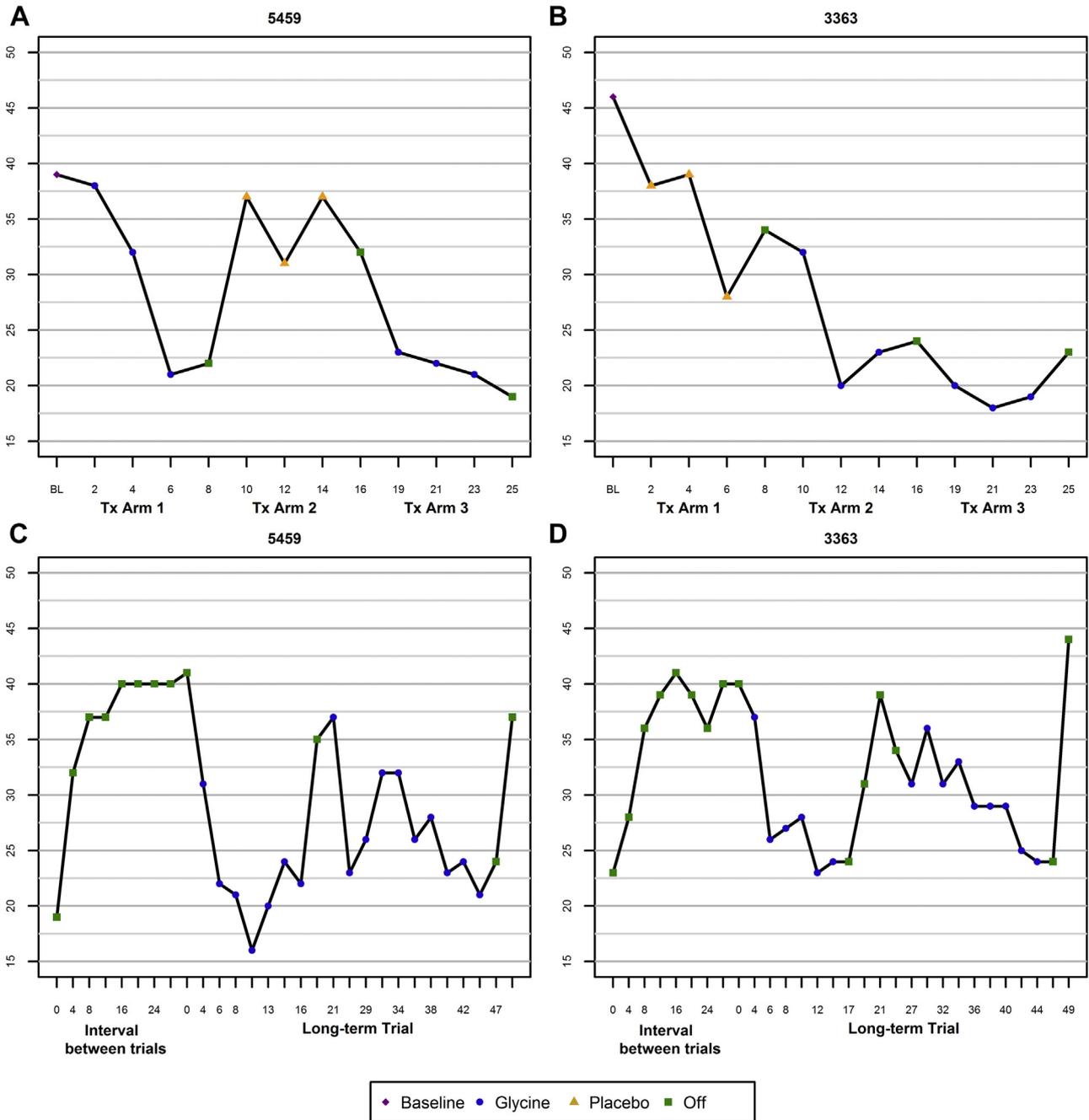


Figure 2. (A, B) Changes in total Brief Psychiatric Rating Scale score as a function of treatment (Tx) with glycine or placebo in subject 5459 (A) and subject 3363 (B) during the short-term trials. (C, D) Changes in total Brief Psychiatric Rating Scale score during the interval between the short-term and long-term glycine trials and during long-term glycine treatment in subject 5459 (C) and subject 3363 (D).

received pharmaceutical-grade glycine (Ajinomoto, Raleigh, NC) or placebo as determined by the research pharmacist's randomization. See the Supplement for details of dose preparation. Starting with a dose of 6 g, the daily dose of glycine or placebo was titrated upward by 3 g/day (3-times-daily dosing)

until the target dose was reached or gastrointestinal (GI) side effects occurred. The target glycine dose was 0.8 g/kg/day based on reports that this dose yielded optimal therapeutic effects with minimal side effects (59,60). Subject 5459 received glycine during arm 1 and placebo during arm 2; subject 3363

Table 1. Changes in Brief Psychiatric Rating Scale as a Function of Glycine or Placebo

Trial	Subject	Condition		% Change
		Off Glycine	On Glycine	
Double-Blind ^a	5459	35.0 (3.5)	30.3 (8.6)	13.4
	3363	35.0 (6.1)	25.0 (6.2)	28.6
Open-Label ^b	5459	35.2 (7.0)	24.7 (5.0)	29.8
	3363	34.5 (6.9)	27.2 (5.4)	21.2

Values are mean (SD).

^aAverage decrease: 7.3 (3.0) (20.0%).

^bAverage decrease: 8.8 (1.5) (25.9%).

received placebo during arm 1 and glycine during arm 2. See the [Supplement](#) for details regarding dosing, titration schedules, side effect management, and de facto sustainable dosing due to GI side effects.

Long-term Glycine Augmentation Clinical Trial.

Following completion of the short-term glycine study, both subjects experienced an exacerbation of clinical symptoms ([Figure 2C, D](#)). An 8-month period elapsed between the end of the short-term trial and the start of the long-term trial. The long-term glycine trial lasted 47 weeks (see [Supplement](#) and [supplemental tables](#)). Although both subjects showed an initial reduction in total BPRS score ([Figure 2C, D](#)), the long-term trial was temporarily suspended at 16 weeks owing to the intolerability of GI side effects with long-term 3-times-daily dosing. Both subjects asked to end the trial during week 47, having experienced chronic GI side effects even at reduced doses. See the [Supplement](#) for details. The maximum sustainably tolerable doses were ~18.8% to 27.5% of the target doses.

DCS Augmentation Trial

The same subjects participated in the DCS study. Age and medication information is contained in the [Supplement](#).

The first arm was an 8-week open-label trial in response to reports of positive symptom exacerbation in two patients with schizophrenia treated with conventional neuroleptics (61) and of a significant mean worsening of negative symptoms in patients treated with clozapine (CLZ) (62) during exposure to 50 mg of DCS. Therefore, it seemed prudent to ascertain that DCS was not negatively affecting symptom severity and that DCS plasma levels were not unusually high [see (61)] before embarking on a double-blind trial. Following a 1-week washout period after the open-label trial, sufficient for the 7- to 15-hour half-life of DCS (63), the double-blind placebo-controlled trial started; each arm lasted 6 weeks with a washout week between arms. The dose of DCS was 50 mg (every morning), a dose at which DCS is a partial agonist of the NMDAR (48,49,64–66), which has been shown to reduce negative symptoms of schizophrenia in non-CLZ-treated patients (61,67) and to augment cognitive behavioral therapy for delusions (68). The double-blind phase was followed by 24 weeks of open-label exposure. Clinical ratings (every 2 weeks) and movement disorder exams (at the end of each arm) were carried out blinded to

condition. Owing to the unexpected death of an immediate family member at the end of the washout between phase 1 and phase 2 of the double-blind trial (week 7), the washout period was extended for an additional 6 weeks until the subjects' clinical states had stabilized sufficiently for the trial to resume.

Statistical Analyses

We fit linear models with fixed effects for treatment and subject for the primary outcome, total BPRS score, from 1) the postbaseline double-blind arms and 2) the open-label periods, which included the open-label arm of the short-term trial, the interval between the short- and long-term trials, and the long-term trial. We also fit a model that included a treatment-by-subject interaction. Owing to the small sample size, *p* values were estimated using permutation tests and standard errors were estimated using the bootstrap method (69). A two-tailed *p* value of .05 was considered statistically significant.

RESULTS

Glycine Augmentation Clinical Trial

Short-term Glycine Trial. Both subjects showed improvement in clinical symptoms during administration of glycine. [Figure 2A, B](#) presents total BPRS scores during the short-term trials in each subject. During the two arms of the double-blind trial (treatment arms 1 and 2), the mean (SD) total BPRS score for subject 5459 was 30.3 (8.6) while on glycine and 35.0 (3.5) while off glycine; for subject 3363, the mean (SD) total BPRS score was 25.0 (6.2) while on glycine and 35.0 (6.1) while off glycine ([Table 1](#)). See [Supplemental Figures S2 and S3](#) and [Supplement](#) for data from other salient Positive and Negative Syndrome Scale symptom domains.

The estimated magnitude of effect, or mean (SE) decrease in total BPRS score while receiving glycine, was 7.3 (3.0) points (20%) lower than while receiving placebo; however, this difference did not reach statistical significance (*p* = .083). The interaction between condition and subject was not statistically significant (*p* = .494) ([Table 1](#)), indicating that the effect of glycine on total BPRS score did not significantly differ between subjects.

During the subsequent 6 weeks of open-label treatment with glycine, both subjects again showed a substantial reduction of symptoms ([Figure 2A, B](#), treatment arm 3). Following completion of that arm, both subjects experienced an exacerbation of clinical symptoms during the 8-month interval between the end of the short-term trial and the start of the open-label long-term glycine trial ([Figure 2C, D](#)).

Long-term Open-Label Glycine Trial. The mean (SE) decrease in total BPRS score while receiving glycine in all open-label periods was 8.8 (1.5) (*p* < .001), a reduction of 26%. The interaction between condition and subject was not statistically significant (*p* = .343) ([Table 1](#)).

Plasma Levels. At baseline, plasma glycine levels were within the normal range (Figure 3A, B). L-serine plasma levels were at the low end of the normal range (Figure 3C, D). During treatment with glycine, plasma levels of glycine and L-serine increased by 121% to 179% and by 146% to 210%, respectively (see Supplement). KYN levels were elevated above the normal range in both subjects independent of glycine–placebo condition and during the long-term glycine trial; the increase was particularly prominent in subject 5459 (Supplemental Figure S4). KYNA levels were markedly elevated in subject 5459 at baseline and were consistently normalized during short-term treatment with glycine and during short periods when glycine was tolerated long term; KYNA levels in subject 3363 were in the upper range of normal at baseline and were also reduced by glycine, but not as consistently as in subject 5459 (Supplemental Figure S5).

DCS Augmentation Trial

Both subjects showed improvement in clinical symptoms during administration of DCS. Figure 4 presents total BPRS scores during all phases of the DCS trial in each subject. During the two arms of the double-blind trial, the mean (SD) total BPRS score for subject 5459 was 28.3 (1.5) while on DCS and 34.3 (1.2) while off DCS; for subject 3363, the mean (SD) total BPRS score was 25.3 (0.6) while on DCS and 42.7 (4.0) while off DCS. Subject 3363 showed improvement in both positive psychotic and negative symptoms. Improvement in subject 5459 was restricted to positive and mood symptoms. The magnitude of the clinical improvement was statistically significant during the open-label and double-blind arms (Table 2). During the double-blind phase, the estimated magnitude of effect, or mean (SE) decrease in total BPRS score while receiving DCS, was 11.7 (1.1) points (30.3%) lower than while receiving placebo ($p = .006$). There was a significant interaction between condition and subject ($p = .002$).¹

During the short-term and long-term open-label periods, the mean (SE) decrease in total BPRS score while receiving DCS in all open-label periods was 3.8 (2.1) ($p < .009$), a mean

¹It is likely that the 40.7% difference in severity of symptoms between the DCS placebo conditions in subject 3363 exaggerates the clinical worsening associated with the placebo condition, which was confounded by the persisting impact of the unexpected death of his father. Although the study was suspended for 6 weeks and subject 3363 had returned to near his original baseline before the study was resumed, further worsening in this context is not surprising. This interpretation is strengthened by the fact that the reduction in severity of clinical symptoms during the initial open-label trial and the first arm of the double-blind trial, prior to this unexpected life event, was in the range of 32% to 37% and was in the range of 27% to 35% for most of the long-term open-label phase, suggesting a notable reduction in symptom severity while on DCS. Similarly, the magnitude of the DCS–placebo difference in subject 5459 may have been attenuated by the effect of the personal loss prior to exposure to DCS during the second arm of the double-blind trial where the reduction in total BPRS score ranged from 12% to 24% compared with the various open-label conditions when symptom reduction generally ranged from 21% to 35%.

reduction of 12.7%. There was no significant interaction between condition and subject ($p = .834$) (Table 2), indicating that the effect of DCS on total BPRS score did not significantly differ between subjects.

Plasma Levels. Figure 5A, B shows increases in plasma DCS levels as a function of exposure to DCS.

In contrast to the dramatic increases in L-serine (96–99% of total serine) observed during exposure to glycine (Figure 3C, D), total serine plasma levels increased only modestly (10.2–54%) with exposure to DCS but remained below or barely within the normal range (Figure 5C, D). Plasma glycine level also did not change substantially as a function of treatment with DCS (Supplemental Figure S6). These findings are consistent with DCS being a weak inhibitor of serine racemase (70).

KYN levels were consistently elevated in both subjects independent of DCS–placebo condition, especially in subject 5459 (Supplemental Figure S4). KYNA levels were markedly elevated in subject 5459 at baseline and were more clearly normalized during treatment with DCS than with glycine, but this normalization was not sustained during long-term exposure; KYNA levels in subject 3363 were at the upper end of the normal range at baseline and were not altered by exposure to DCS (Supplemental Figure S5).

Neurocognition. No consistent changes in neurocognition were observed as a function of glycine or DCS exposure (see Supplement and Supplemental Figure S7).

DISCUSSION

We report the results of two proof-of-principle clinical trials showing that interventions tailored to a specific genetic mutation reduced symptom severity in two individuals with psychotic disorders. Although both subjects were partially remitted at baseline, the additional 20% to 26% reduction in symptom severity on glycine and 13% to 30% reduction on DCS reflected substantial relief beyond that achieved by their usual psychotropic drug regimen, an effect that is considered clinically meaningful in augmentation treatment studies of schizophrenia (71). The pleiotropic clinical effects of the *GLDC* triplication—present in schizoaffective and bipolar disorder—are consistent with the variable expressivity of rare CNVs (see Supplement). The demonstration of tractable symptom relief by targeting a specific genotype explicitly links an individual mutation to disease biology and pathophysiology and to treatment response. Although we do not know conclusively that the *GLDC* triplication or any of the other genetic elements in the CNV region is causally implicated in the psychiatric illnesses of the carriers (72,73),² our data show that the severity of their symptoms was reduced by augmentation with glycine or DCS. This result underscores the importance of molecular

²De novo structural variants have been linked to sporadic psychiatric illness in some studies (110,111) but not in others (41,112). The extended family of these carriers has a history of psychotic disorders in earlier generations (Supplemental Figure S9A). If some variant(s) within the 9p24.1 complex rearrangement was causal in the case of our two carriers, different genetic risk factors were likely present in cases in earlier generations.

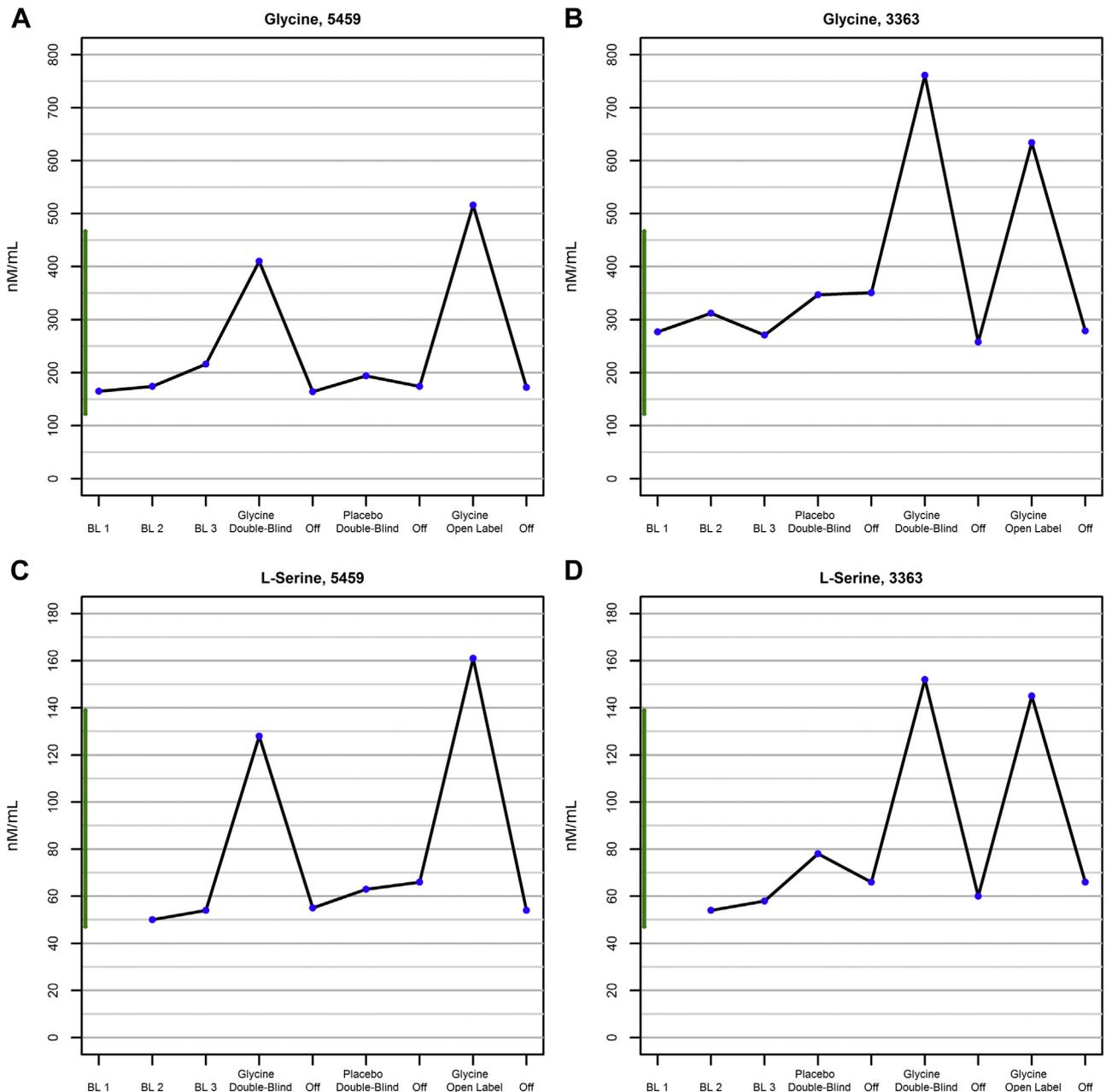


Figure 3. (A, B) Changes in plasma glycine level as a function of short-term treatment with glycine or placebo in subject 5459 (A) and subject 3363 (B). (C, D) Changes in plasma L-serine level as a function of short-term treatment with glycine or placebo in subject 5459 (C) and subject 3363 (D). All data are from the short-term trials. BL, baseline.

diagnosis and targeting specific biological processes rather than clinical diagnoses per se. Indeed, it is not unusual for medications to be efficacious in only a subgroup of individuals treated for a particular clinical condition (74–77). The *GLDC* gene is among the 15.4% of genes least tolerant of functional variation (see Supplement) (78), suggesting that gain and loss of function changes in this gene might not be phenotypically neutral. Indeed, triplication of a disease gene may convey a more severe disease phenotype than does duplication of the same locus (79–81).

The success of this genotype-first (27) approach underscores the utility of targeting specific biological processes in appropriately selected individuals. NMDAR modulators have shown variable efficacy in patients with schizophrenia selected on the basis of refractory negative symptoms (82,83), but not for having an identified disturbance in NMDAR function. Inasmuch as rare and common structural and sequence variants converge on specific biological pathways, including but not limited to the NMDAR (e.g., immune function, calcium channel signaling) (15,25,26,84–89), our results

Treatment of Carriers of a *GLDC* Triplication

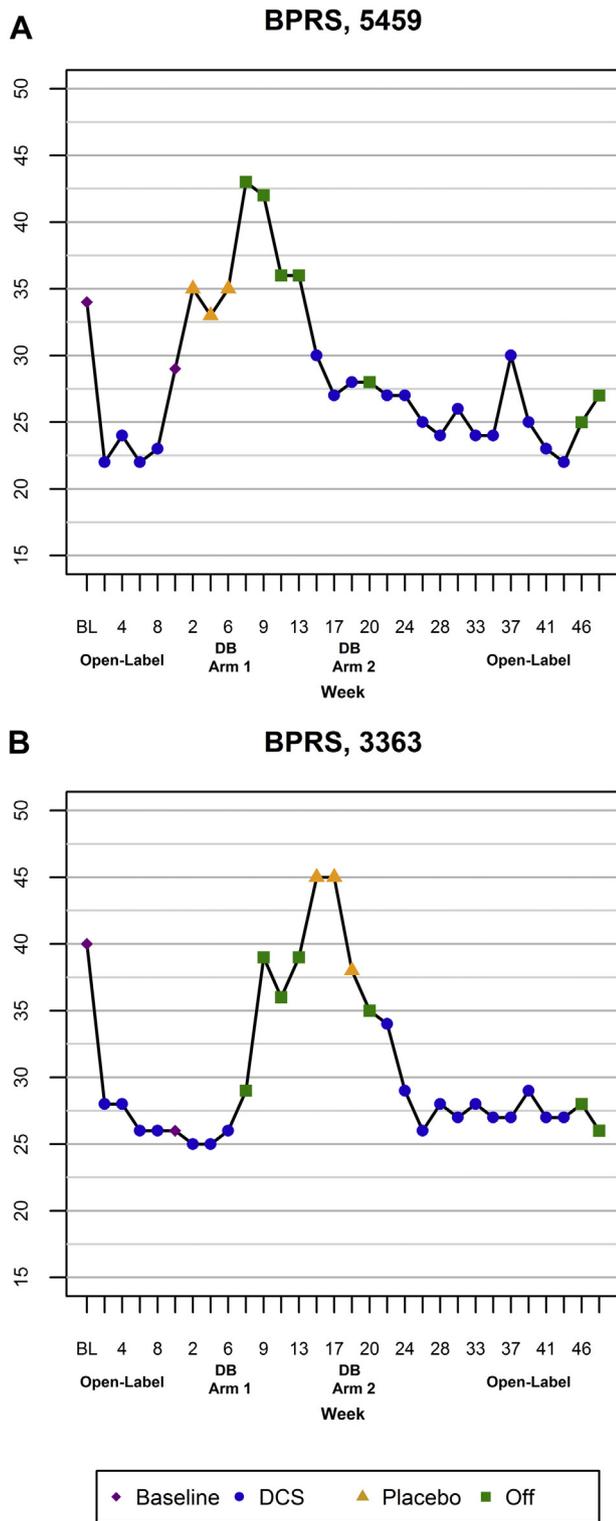


Figure 4. Changes in total Brief Psychiatric Rating Scale (BPRS) score as a function of treatment with D-cycloserine (DCS) or placebo in subject 5459 (A) and subject 3363 (B). BL, baseline; DB, double-blind.

Table 2. Changes in Brief Psychiatric Rating Scale as a Function of DCS or Placebo

Trial	Subject	Condition		% Change
		Off DCS	On DCS	
Double-Blind ^a	5459	34.3 (1.2)	28.3 (1.5)	17.5
	3363	42.7 (4.0)	25.3 (0.6)	40.7
Open Label ^b	5459	28.7 (4.7)	24.5 (2.2)	14.6
	3363	31.3 (7.6)	27.8 (2.0)	11.2

Values are mean (SD).

DCS, D-cycloserine.

^aAverage decrease: 11.7 (1.1) (30.3%).

^bAverage decrease: 3.8 (2.1) (12.7%).

are potentially relevant to a broader group than carriers of increased *GLDC* copy number per se [e.g., see (90)]. Genes involved in glutamate neurotransmission, in particular, are overrepresented among the rare variants associated with schizophrenia, autism spectrum disorders, and nonsyndromic intellectual disability (25,26). Thus, even though the *GLDC* triplication is so far a private mutation, individuals with mutations in genes affecting glutamatergic and NMDAR signaling may constitute a molecular subtype amenable to pathway-defined treatment (27) who would have a high prior probability of responding to treatments that normalize glutamatergic dysregulation. In keeping with the pleiotropy that is characteristic of CNVs, the pool of potential beneficiaries is likely to transcend diagnostic categories. Notably, multiple lines of evidence implicate genetic variants in *KDM4C/JMJD2C*, *GLDC*, and other genes in the 9p24.1 region with schizophrenia, autism spectrum disorders, bipolar disorder, and neurodevelopmental disorders (summarized in Supplement). Our findings are consistent with other data illustrating the value of molecular diagnosis in clarifying the significance of newly emerging clinical symptoms (91) or guiding treatment in the context of atypical psychiatric presentations (92).

At baseline, plasma levels of glycine were within the normal range, and those of L-serine were at the low end of the normal range. Plasma levels poorly reflect brain extracellular levels of glycine and L-serine. In the brain, *GLDC* is expressed exclusively in astrocytes (93). Astrocytic serine hydroxyl methyl transferase converts glycine to L-serine, which is converted to D-serine, the NMDAR coagonist, by neuronal serine racemase. Thus, reduced availability of glycine and L-serine would decrease neuronal synthesis of D-serine (94). Both glycine and L-serine plasma levels showed marked increases during glycine treatment; these increases were much greater for L-serine than for glycine. Inasmuch as glutamatergic signaling through NMDARs requires glycine or D-serine at the GMS, it is possible that exogenous glycine increased the conversion of glycine to L-serine, thereby increasing precursor availability for D-serine synthesis (94). D-serine is a more potent agonist at the GMS than glycine and is the preferential agonist at fore-brain NMDARs. Thus, the mechanism underlying the therapeutic effect of glycine may, at least in part, have been mediated by increased D-serine rather than by glycine per se (Supplemental Figure S8).

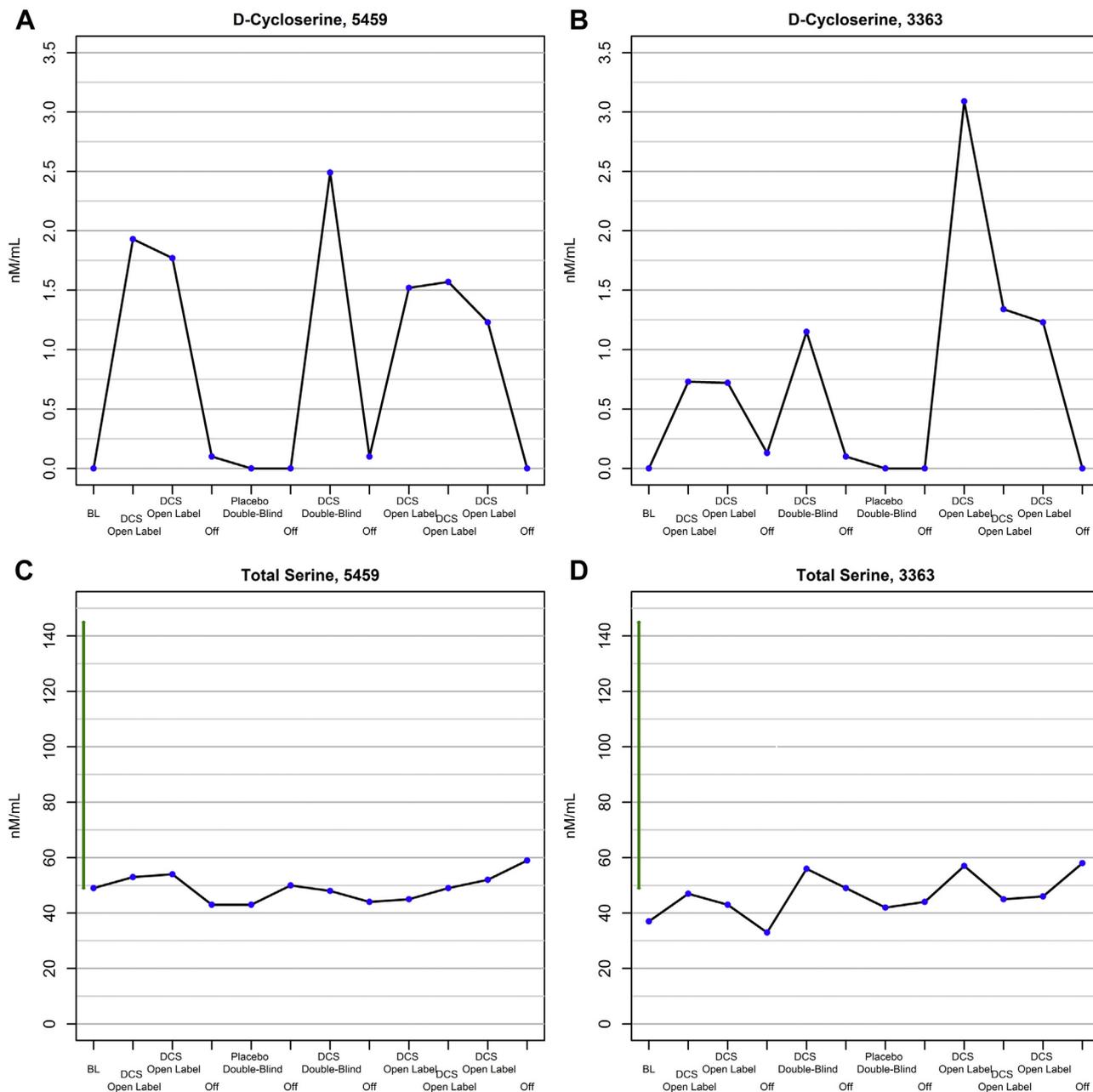


Figure 5. (A, B) Changes in plasma D-cycloserine (DCS) level as a function of treatment with DCS or placebo in subject 5459 (A) and subject 3363 (B). (C, D) Changes in plasma total serine level as a function of short-term treatment with DCS or placebo in subject 5459 (C) and subject 3363 (D). BL, baseline.

We selected the particular subjects in these studies to undergo NMDAR modulation based on the presence of the *GLDC* triplication, which we hypothesized would result in increased catabolism of glycine and D-serine resulting in NMDAR hypofunction. Unbeknownst to us at the start of the glycine clinical trial, both carriers had elevated baseline KYN and KYNA plasma levels (for reasons that are currently unknown). KYNA is a nonselective competitive glutamate receptor antagonist (95) with a particularly strong affinity for the GMS of the NMDAR (96). KYNA was normalized by

augmentation with glycine (although this effect was not sustained in subject 3363), suggesting that glycine may have partially counteracted the antagonism of KYNA *in vivo*, consistent with *in vitro* data on glycine and D-serine (97). KYN remained elevated independent of treatment with glycine (or DCS). Conceivably, the magnitude of the glycine effect on symptom severity may have been attenuated by persistent elevations of KYN and/or KYNA. Thus, elevated KYN and KYNA may have potentiated the deleterious effect of increased glycine and D-serine degradation caused by the

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GLDC triplication (or vice versa). Either the *GLDC* triplication or elevated KYNA would have been sufficient to implicate NMDAR hypofunction. The possibility that two processes impairing NMDAR function were present is consistent with evidence of multiple genetic hits in neuropsychiatric disorders (98,99). Glycine's efficacy in modulating symptom severity may have been an opportunistic effect of different processes that independently contributed to NMDAR dysregulation such that glycine agonism at the GMS or antagonism of KYNA partially normalized NMDAR function irrespective of whether this triplication or other genes in the rearranged region were causal.

DCS is a partial agonist at the GMS, with only 40% to 60% of the potency of glycine, with activity ranging from 65% to 200% compared with glycine at saturating doses depending on NMDAR subunit composition (100), but it crosses the blood-brain barrier more readily (63). In addition, it blocks the formation of KYNA in vitro (101). These dual actions, agonism at the GMS and KYNA inhibition, suggested that DCS might produce an even greater clinical benefit than was achieved with glycine in restoring NMDAR function (Supplemental Figure S8) and guided the rationale for undertaking the second clinical trial. Notably, DCS produced substantially greater normalization of KYNA than glycine (in subject 5459, although it was not sustained). These dual actions of DCS in normalizing NMDAR hypofunction may have contributed to the substantially greater therapeutic benefit of DCS than of glycine.

In addition to its antagonism of the GMS, KYNA is also a potent noncompetitive antagonist of the $\alpha 7$ nicotinic acetylcholine receptor (102–104) [reviewed in (105)]. The *CHRNA7* gene has been linked to schizophrenia (106,107). Both the NMDAR and $\alpha 7$ nicotinic acetylcholine receptor have important roles in cognition and synaptic plasticity (108,109). Notably, we observed no systematic changes in neurocognitive function in the subjects during exposure to glycine or DCS, possibly related to sustained elevations of KYN and incompletely normalized levels of KYNA.³

Other Relevant Considerations

Although side-effect-free up until the threshold for GI side effects is reached, glycine is cumbersome to use, especially over the long term. In our experience, the optimal target dose of 0.8 g/kg not only was much too high to be tolerated without prohibitive side effects but also was above the dose needed to achieve therapeutic benefit. Indeed, we observed clinical improvement at substantially lower doses. Whether our subjects had an unusual sensitivity to glycine or DCS (related to the *GLDC* triplication or to elevated KYN and/or KYNA) is unclear.

³The elevated baseline KYN and KYNA levels implicate increased activity of indoleamine 2,3-dioxygenase/tryptophan 2,3-dioxygenase and kynurenine aminotransferase. Normalizing KYNA through kynurenine aminotransferase inhibition (113,114) has been shown to enhance cognition (115,116). Such a strategy alone or in combination with partial agonism of the $\alpha 7$ nicotinic acetylcholine receptor (117) may have had greater benefit on cognition in our subjects.

Patients taking CLZ are generally excluded from augmentation with NMDAR modulators.⁴ We hypothesize that the magnitude of CLZ's normalization of NMDAR-dependent neurotransmission will depend on the presence and severity of a glutamatergic deficit. In carriers of mutations that compromise NMDAR-dependent glutamatergic function, CLZ may provide only partial neurochemical remission, leaving room for additional normalization with NMDAR modulators. Thus, it seems reasonable to propose that patients with mutations in NMDAR and glutamate-related genes may benefit from NMDAR modulators even when taking CLZ.

Several limitations should be considered. First, the optimal compound to antagonize an excess of *GLDC* is a *GLDC* inhibitor. Because no such compound is available, we used proxies to try to normalize the effects of the *GLDC* triplication. Ideally, Food and Drug Administration–approved compounds found to have *GLDC* inhibitory activity could be repurposed to more directly target increased degradation. Second, the small sample size and limited number of observations during the double-blind arms reduced power to detect a statistically

⁴The reason is that some of CLZ's neurochemical actions affect synaptic glycine levels either by inhibiting glycine transporter type 1 activity (118) or by inhibiting system A–mediated transport of glycine and other amino acids (119). Consistent with these mechanisms for neutralizing the effects of NMDAR modulators, CLZ-induced increase in extracellular D-serine leads to subsequent NMDAR-dependent release of L-glutamate in rats (120). This is likely the reason that augmentation with NMDAR modulators tends to show clinical efficacy primarily in patients with schizophrenia taking antipsychotics other than CLZ (60,61,121,122) and often not in patients taking CLZ (82,119,123–125). However, no clinical benefit (67) or an equivocal benefit (61) of DCS has also been observed in patients not taking CLZ, small groups of CLZ-treated and non-CLZ-treated patients experienced similar clinical benefits from glycine (126), and CLZ-resistant patients experienced significant improvement in negative and overall symptom severity with sodium benzoate augmentation of CLZ (127). Two studies even reported statistically significant (but clinically modest) worsening of negative symptoms with DCS and CLZ (62,119) compared with CLZ alone. Notably, clinical benefit and worsening with DCS and glycine have been almost entirely limited to negative symptoms (82,83,128) (except for enhancing the effect of cognitive behavioral treatment of delusions) (68), whereas the effects in our subjects were primarily on positive symptoms. The most parsimonious explanation for this pattern of findings in nongenotyped individuals is that patients with schizophrenia are heterogeneous with respect to NMDAR hypofunction. In the subgroup of patients with NMDAR hypofunction, augmentation is most likely to provide clinical benefit in those patients whose NMDAR function has not been normalized (i.e., non-CLZ antipsychotic medication); in this same subgroup, CLZ may generally be sufficient to normalize NMDAR function, resulting in no further benefit from NMDAR modulators. When CLZ is not sufficient to normalize NMDAR function, clinical benefit may occur. In the subgroup of patients who do not have NMDAR hypofunction, augmentation with NMDAR modulators would not be expected to have a clinical benefit.

significant effect of glycine in this short-term trial and precluded use of more standard analytic techniques (e.g., modeling subject as a fixed effect rather than a random effect, modeling trajectory of response or baseline to end-point change rather than comparing observations across all postbaseline time points). Notably, the magnitude of the reduction in symptom severity during the double-blind phase of the glycine trial was consistent with the statistically significant treatment effect observed during the open-label phases and with the significant reduction in symptom severity in the double-blind and open-label arms of the DCS study. Third, carryover effects from prior glycine exposure may have reduced the estimate of the magnitude of the treatment effect (Figure 2). Given the small sample size, it is not possible to formally evaluate these potential effects.⁵ Similarly, subject 3363 had a partial response to placebo during the first arm of the double blind in the glycine study, attenuating the magnitude of the difference between placebo and glycine. Fourth, although the blind was not broken until after the short-term glycine study ended, on the basis of side effects, both staff and the subjects correctly guessed when they received glycine. Although this recognition may have favorably influenced the subjects' clinical states, subjects were so much less symptomatic on glycine that they were willing to tolerate the side effects and dosing for extended periods, suggesting that their substantial clinical improvements are unlikely to reflect placebo effects alone. Notably, there were no side effects during exposure to DCS, making it unlikely that the clinical improvements observed reflected placebo effects. Finally, it would have been ideal had the subjects not suffered a personal loss during the DCS trial and had it been feasible to undertake multiple crossovers between drug and placebo conditions in both studies.

In summary, we report two individuals with psychotic disorders in whom identification of a specific genomic variation resulted in improved clinical symptoms during two proof-of-principle trials targeting a similar mechanism implicated by the mutation. These studies have important implications for the treatment of other patients with alterations of the same or overlapping biochemical pathways.

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⁵Two early studies of glycine augmentation in treatment-resistant chronic schizophrenia, one involving 7 patients and the other involving 9 patients, reported that the significant improvement in negative symptoms observed during 6 weeks of treatment with glycine was sustained for the next 8 weeks (2-week washout and 6 weeks of placebo) (59,126). A third study by the same group did not report a significant carryover effect in 7 patients (60). The persisting effects of glycine on positive symptoms that we observed, however, did not last longer than 2 weeks (Figure 2) but may have reduced the magnitude of the difference in symptom severity between on-glycine and off-glycine assessments. Given the half-life of glycine (26–245 minutes depending on dose) (58), a 2-week washout would generally be expected to be long enough to eliminate persisting effects of the drug.

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JTC reports consulting relationships with Concert Pharm and BVF Partners. Baylor College of Medicine and Miraca Holdings have formed a joint venture with shared ownership and governance of Baylor Genetics, which performs clinical genomics studies including chromosomal microarray analysis and clinical exome sequencing. JRL serves on the Scientific Advisory Board of Baylor Genetics. JRL has stock ownership in 23andMe, is a paid consultant for Regeneron Pharmaceuticals, has stock options in Lasergen Inc., and is a coinventor on multiple U.S. and European patents related to molecular diagnostics for inherited neuropathies, eye diseases, and bacterial genomic fingerprinting. The other authors report no biomedical financial interests or potential conflicts of interest.

ClinicalTrials.gov: Neurobiology of a Mutation in Glycine Metabolism in Psychotic Disorders; <https://clinicaltrials.gov/ct2/show/NCT01720316>; NCT01720316; Targeting a Genetic Mutation in Glycine Metabolism With D-cycloserine (DCS); <https://clinicaltrials.gov/ct2/show/NCT02304432>; NCT02304432.

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REFERENCES

- Rees E, Walters JTR, Georgieva L, Isles AR, Chambert KD, Richards AL, *et al.* (2014): Analysis of copy number variations at 15 schizophrenia-associated loci. *Br J Psychiatry* 204:108–114.
- Malhotra D, Sebat J (2012): CNVs: Harbinger of a rare variant revolution in psychiatric genetics. *Cell* 148:1223–1241.
- Marshall CR, Howrigan DP, Merico D, Thiruvahindrapuram B, Wu W, Greer DS, *et al.* (2017): Contribution of copy number variants to schizophrenia from a genome-wide study of 41,321 subjects. *Nat Genet* 49:27–35.
- International Schizophrenia Consortium (2008): Rare chromosomal deletions and duplications increase risk of schizophrenia. *Nature* 455:237–241.
- Stefansson H, Rujescu D, Cichon S, Pietilainen OPH, Ingason A, Steinberg S, *et al.* (2008): Large recurrent microdeletions associated with schizophrenia. *Nature* 455:232–236.
- McCarthy S, Makarov V, Kirov G, Addington A, McClellan J, Yoon S, *et al.* (2009): Microduplications of 16p11.2 are associated with schizophrenia. *Nat Genet* 41:1223–1227.
- Cantor RM, Geschwind DH (2008): Schizophrenia: Genome, interrupted. *Neuron* 58:165–167.
- Weiss LA, Shen Y, Korn JM, Arking DE, Miller DT, Fossdal R, *et al.* (2008): Association between microdeletion and microduplication at 16p11.2 and autism. *N Engl J Med* 358:667–675.
- Mefford HC, Batshaw ML, Hoffman EP (2012): Genomics, intellectual disability, and autism. *N Engl J Med* 366:733–743.
- Heinzen EL, Neale BM, Traynelis SF, Allen AS, Goldstein DB (2015): The genetics of neuropsychiatric diseases: Looking in and beyond the exome. *Ann Rev Neurosci* 38:47–68.
- Kirov G, Rees E, Walters JTR, Escott-Price V, Georgieva L, Richards AL, *et al.* (2014): The penetrance of copy number variations for schizophrenia and developmental delay. *Biol Psychiatry* 75:378–385.
- Kirov G (2015): CNVs in neuropsychiatric disorders. *Hum Mol Genet* 24:R45–R49.
- Sullivan PF, Daly MJ, O'Donovan M (2012): Genetic architectures of psychiatric disorders: The emerging picture and its implications. *Nat Rev Genet* 13:537–551.
- Geschwind DH, Flint J (2015): Genetics and genomics of psychiatric disease. *Science* 349:1489–1494.
- Schizophrenia Working Group of the Psychiatric Genomics Consortium (2014): Biological insights from 108 schizophrenia-associated genetic loci. *Nature* 511:421–427.
- Sanders SJ, He X, Willsey AJ, Ercan-Sencicek AG, Samocha KE, Cicek AE, *et al.* (2015): Insights into autism spectrum disorder genomic architecture and biology from 71 risk loci. *Neuron* 87:1215–1233.
- Vacic V, McCarthy S, Malhotra D, Murray F, Chou H-H, Peoples A, *et al.* (2011): Duplications of the neuropeptide receptor gene *VIPR2* confer significant risk for schizophrenia. *Nature* 471:499–503.
- Brunetti-Pierri N, Berg JS, Scaglia F, Belmont J, Bacino CA, Sahoo T, *et al.* (2008): Recurrent reciprocal 1q21.1 deletions and duplications associated with microcephaly or macrocephaly and developmental and behavioral abnormalities. *Nat Genet* 40:1466–1471.
- Mefford H, Sharp AJ, Baker C, Itsara A, Jiang Z, Buysse K, *et al.* (2008): Recurrent rearrangements of chromosome 1q21.1 and variable pediatric phenotypes. *N Engl J Med* 359:1685–1699.
- Levinson DF, Duan J, Oh S, Wang K, Sanders AR, Shi J, *et al.* (2011): Copy number variants in schizophrenia: Confirmation of five previous findings and new evidence for 3q29 microdeletions and *VIPR2* duplications. *Am J Psychiatry* 168:302–316.
- Shinawi M, Schaaf CP, Bhatt SS, Xia Z, Patel A, Cheung SW, *et al.* (2009): A small recurrent deletion within 15q13.3 is associated with a range of neurodevelopmental phenotypes. *Nat Genet* 41:1269–1271.
- Carvalho CMB, Lupski JR (2016): Mechanisms underlying structural variant formation in genomic disorders. *Nat Rev Genet* 17:224–238.
- Network and Pathway Analysis Subgroup of Psychiatric Genomics Consortium (2015): Psychiatric genome-wide association study analyses implicate neuronal, immune and histone pathways. *Nat Neurosci* 18:199–209.
- Parikshak NN, Gandal MJ, Geschwind DH (2015): Systems biology and gene networks in neurodevelopmental and neurodegenerative disorders. *Nat Rev Genet* 16:441–458.
- Pocklington AJ, Rees E, Walters JTR, Han J, Kavanagh DH, Chambert KD, *et al.* (2015): Novel findings from CNVs implicate inhibitory and excitatory signaling complexes in schizophrenia. *Neuron* 86:1203–1214.
- Harrison PJ (2015): Recent genetic findings in schizophrenia and their therapeutic relevance. *J Psychopharmacol* 29:85–96.
- Stessman HA, Bernier R, Eichler EE (2014): A genotype-first approach to defining the subtypes of a complex disease. *Cell* 156:872–877.
- White J, Beck CR, Harel T, Posey JE, Jhangiani SN, Tang S, *et al.* (2016): POGZ truncating alleles cause syndromic intellectual disability. *Genome Med* 8:3.
- Treadwell-Deering DE, Powell MP, Potocki L (2010): Cognitive and behavioral characterization of the Potocki-Lupski syndrome (duplication 17p11.2). *J Dev Behav Pediatr* 31:137–143.
- Berg JS, Brunetti-Pierri N, Peters SU, Kang S-HL, Fong C, Salamone J, *et al.* (2007): Speech delay and autism spectrum behaviors are frequently associated with duplication of the 7q11.23 Williams-Beuren syndrome region. *Genet Med* 9:427–441.
- Ben-Shachar S, Lanpher B, German JR, Qasaymeh M, Potocki L, Nagamani SCS, *et al.* (2009): Microdeletion 15q13.3: A locus with incomplete penetrance for autism, mental retardation, and psychiatric disorders. *J Med Genet* 46:382–388.
- Männik K, Mägi R, Macé A, Cole B, Guyatt AL, Shihab HA, *et al.* (2015): Copy number variations and cognitive phenotypes in unselected populations. *JAMA* 313:2044–2054.
- Lupski JR (2016): Clinical genomics: From a truly personal genome viewpoint. *Hum Genet* 135:591–601.
- Scolnick EM (2010): 3rd Annual Report: January 2010—Executive Summary Version. Available at: https://www.broadinstitute.org/files/shared/psych/_StanleyCenterAnnualReportExecSum2009.pdf. Accessed August 14, 2016.
- Bainbridge MN, Wiszniewski W, Murdock DR, Friedman J, Gonzaga-Jauregui C, Newsham I, *et al.* (2011): Whole-genome sequencing for optimized patient management. *Sci Transl Med* 3:87re83.
- Worthey EA, Mayer AN, Syverson GD, Helbling D, Bonacci BB, Decker B, *et al.* (2011): Making a definitive diagnosis: Successful clinical application of whole exome sequencing in a child with intractable inflammatory bowel disease. *Genet Med* 13:255–262.
- Daly AK, Donaldson PT, Bhatnagar P, Shen Y, Pe'er I, Floratos A, *et al.* (2009): HLA-B*5701 genotype is a major determinant of drug-induced liver injury due to flucloxacillin. *Nat Genet* 41:816–819.
- Loi S, Haibe-Kains B, Majaj S, Lallemand F, Durbecq V, Larsimont D, *et al.* (2010): PIK3CA mutations associated with gene signature of low mTORC1 signaling and better outcomes in estrogen receptor-positive breast cancer. *Proc Natl Acad Sci U S A* 107:10208–10213.
- Nathwani AC, Tuddenham EGD, Rangarajan S, Rosales C, McIntosh J, Linch DC, *et al.* (2011): Adenovirus-associated virus vector-mediated gene transfer in Hemophilia B. *N Engl J Med* 365:2357–2365.
- American Psychiatric Association (1994): Diagnostic and Statistical Manual of Mental Disorders (DSM-IV). Washington, DC: American Psychiatric Association.
- Malhotra D, McCarthy S, Michaelson JJ, Vacic V, Burdick KE, Yoon S, *et al.* (2011): High frequencies of de novo CNVs in bipolar disorder and schizophrenia. *Neuron* 72:951–963.
- Grochowski CM, Gu S, Yuan B, TCW J, Brennan KJ, Sebat J, *et al.* (2018): Marker chromosome genomic structure and temporal origin implicate a chromoanasythesis event in a family with pleiotropic psychiatric phenotypes. *Hum Mutat* 39:939–946.
- Mothet JP, Le Bail M, Billard JM (2015): Time and space profiling of NMDA receptor co-agonist functions. *J Neurochem* 135:210–225.
- Olney JW, Farber NB (1995): Glutamate receptor dysfunction and schizophrenia. *Arch Gen Psychiatry* 52:998–1007.
- Coyle JT (2006): Glutamate and schizophrenia: Beyond the dopamine hypothesis. *Cell Mol Neurobiol* 26:365–384.

46. Krystal JH, Anand A, Moghaddam B (2002): Effects of NMDA receptor antagonists: Implications for the pathophysiology of schizophrenia. *Arch Gen Psychiatry* 59:663–664.
47. Goff DC, Coyle JT (2001): The emerging role of glutamate in the pathophysiology and treatment of schizophrenia. *Am J Psychiatry* 158:1367–1377.
48. Henderson G, Johnson JW, Ascher P (1990): Competitive antagonists and partial agonists at the glycine modulatory site of the mouse *N*-methyl-D-aspartate receptor. *J Physiol* 430:189–212.
49. Watson GB, Bolanowski MA, Baganoff MP, Deppeler CL, Lanthorn TH (1990): D-cycloserine acts as a partial agonist at the glycine modulatory site of the NMDA receptor expressed in *Xenopus* oocytes. *Brain Res* 510:158–160.
50. Overall J, Gorham D (1962): The Brief Psychiatric Rating Scale. *Psychol Rep* 10:799–812.
51. Kay SR, Fiszbein A, Opler LA (1987): The Positive and Negative Syndrome Scale (PANSS) for schizophrenia. *Schizophr Bull* 13:261–276.
52. Guy W (1976): Clinical global impression (CGI) Scale. In: Guy W, editor. *ECDEU Assessment Manual for Psychopharmacology*. Rockville, MD: Department of Health, Education, and Welfare, 125–126.
53. Hamilton M (1960): A rating scale for depression. *J Neurol Neurosurg Psychiatry* 23:56–62.
54. Young RC, Biggs JT, Ziegler VE, Meyer DA (1978): A rating scale for mania: Reliability, validity and sensitivity. *Br J Psychiatry* 133:429–435.
55. Posner K, Brent D, Lucas C, Gould M, Stanley B, Brown G, et al. (2009): Columbia–Suicide Severity Rating Scale (C-SSRS). New York: Research Foundation for Mental Hygiene.
56. Simpson GM, Angus JW (1970): A rating scale for extrapyramidal side effects. *Acta Psychiatr Scand* 212(suppl 44):11–19.
57. Guy W (1976): Abnormal Involuntary Movement Scale (AIMS). In: Guy W, editor. *ECDEU Assessment Manual for Psychopharmacology*, rev. Rockville, MD: Department of Health, Education, and Welfare, 118–119.
58. Hahn RG (1993): Dose-dependent half-life of glycine. *Urol Res* 21:289–291.
59. Heresco-Levy U, Javitt DC, Ermilov M, Mordel C, Silipo G, Lichtenstein M (1999): Efficacy of high-dose glycine in the treatment of enduring negative symptoms of schizophrenia. *Arch Gen Psychiatry* 56:29–36.
60. Heresco-Levy U, Ermilov M, Lichtenberg P, Bar G, Javitt DC (2004): High-dose glycine added to olanzapine and risperidone for the treatment of schizophrenia. *Biol Psychiatry* 55:165–171.
61. Goff DC, Tsai G, Levitt J, Amico E, Manoch D, Schoenfeld D, et al. (1999): A placebo-controlled trial of D-cycloserine added to conventional neuroleptics in patients with schizophrenia. *Arch Gen Psychiatry* 56:21–27.
62. Goff DC, Tsai G, Manoch DS, Flood J, Darby DG, Coyle JT (1996): D-cycloserine added to clozapine for patients with schizophrenia. *Am J Psychiatry* 153:1628–1630.
63. Hanngren H, Hansson E, Ullberg S (1961): An autoradiographic study of the distribution of tritium-labeled cycloserine in mice. *Antibiot Chemother* 12:46–54.
64. Hood WF, Compton RP, Monahan JB (1989): D-cycloserine: A ligand for the *N*-methyl-D-aspartate coupled glycine receptor has partial agonist characteristics. *Neurosci Lett* 98:91–95.
65. Emmett MR, Mick SJ, Cker JA, Rao TS, Iyengar S, Wood PL (1991): Actions of D-cycloserine at the *N*-methyl-D-aspartate-associated glycine receptor site in vivo. *Neuropharmacology* 30:1167–1171.
66. Chessell IP, Procter AW, Francis PT, Bowen DM (1991): D-cycloserine, a putative cognitive enhancer, facilitates activation of the *N*-methyl-D-aspartate receptor-ionophore complex in Alzheimer brain. *Brain Res* 565:345–348.
67. Goff DC, Herz L, Posever T, Shih V, Tsai G, Henderson D, et al. (2005): A six-month, placebo-controlled trial of D-cycloserine co-administered with conventional antipsychotics in schizophrenia patients. *Psychopharmacology (Berl)* 179:144–150.
68. Gottlieb JD, Cather C, Shanahan M, Creedon T, Macklin EA, Goff DC (2011): D-cycloserine facilitation of cognitive behavioral therapy for delusions in schizophrenia. *Schizophr Res* 131:69–74.
69. Edgington ES, Onghena P (2007): *Randomization Tests*, 4th ed. London: Chapman & Hall.
70. Cook SP, Galve-Roperh I, Martínez del Pozo A, Rodríguez-Crespo I (2002): Direct calcium binding results in activation of brain serine racemase. *J Biol Chem* 277:27782–27792.
71. Freudenreich O, Goff DC (2002): Antipsychotic combination therapy in schizophrenia: A review of efficacy and risks of current combinations. *Acta Psychiatr Scand* 106:323–330.
72. Cooper GM, Shendure J (2011): Needles in stacks of needles: Finding disease-causal variants in a wealth of genomic data. *Nat Rev Genet* 12:628–640.
73. Buchanan JA, Scherer SW (2008): Contemplating effects of genomic structural variation. *Genet Med* 10:639–647.
74. Perakslis ED, Kohane IS (2016): Treating the enigmatic “exceptional responders” as patients with undiagnosed diseases. *Sci Transl Med* 8:340ed348.
75. Goff DC (2014): Bitopertin: The good news and bad news. *JAMA Psychiatry* 71:621–622.
76. Chau NG, Lorch JH (2015): Exceptional responders inspire change: Lessons for drug development from the bedside to the bench and back. *Oncologist* 20:699–701.
77. Iyer G, Hanrahan AJ, Milowsky MI, Al-Ahmadie H, Scott SN, Janakiraman M, et al. (2012): Genome sequencing identifies a basis for everolimus sensitivity. *Science* 338:221.
78. Petrovski S, Wang Q, Heinzen EL, Allen AS, Goldstein DB (2013): Genetic intolerance to functional variation and the interpretation of personal genomes. *PLoS Genet* 9:e1003709.
79. Liu P, Gelowani V, Zhang F, Drory VE, Ben-Shachar S, Roney E, et al. (2014): Mechanism, prevalence, and more severe neuropathy phenotype of the Charcot-Marie-Tooth type 1A triplication. *Am J Hum Genet* 94:462–469.
80. Ramocki MB, Peters SU, Tavyev YJ, Zhang F, Carvalho CMB, Schaaf CP, et al. (2009): Autism and other neuropsychiatric symptoms are prevalent in individuals with MECP2 duplication syndrome. *Ann Neurol* 66:771–782.
81. Carvalho CMB, Ramocki MB, Pehliva D, Franco LM, Gonzaga-Jauregui C, Fang P, et al. (2011): Inverted genomic segments and complex triplication rearrangements are mediated by inverted repeats in the human genome. *Nat Genet* 43:1074–1081.
82. Tsai GE, Lin PY (2010): Strategies to enhance *N*-methyl-D-aspartate receptor-mediated neurotransmission in schizophrenia: A critical review and meta-analysis. *Curr Pharm Des* 16:522–537.
83. Singh SP, Singh V (2011): Meta-analysis of the efficacy of adjunctive NMDA receptor modulators in chronic schizophrenia. *CNS Drugs* 25:859–885.
84. Kirov G, Pocklington AJ, Holmans P, Ivanov D, Ikeda M, Ruderfer D, et al. (2012): De novo CNV analysis implicates specific abnormalities of postsynaptic signalling complexes in the pathogenesis of schizophrenia. *Mol Psychiatry* 17:142–153.
85. Raychaudhuri S, Plenge RM, Rossin EJ, Ng ACY, International Schizophrenia Consortium, Purcell SM, et al. (2009): Identifying relationships among genomic disease regions: Predicting genes at pathogenic SNP associations and rare deletions. *PLoS Genet* 5:e1000534.
86. Hamdan FF, Gauthier J, Araki Y, Lin D-T, Yoshizawa Y, Higashi K, et al. (2011): Excess of de novo deleterious mutations in genes associated with glutamatergic systems in nonsyndromic intellectual disability. *Am J Hum Genet* 88:306–316.
87. Elia J, Gai X, Xie HM, Perin JC, Geiger E, Glessner JT, et al. (2010): Rare structural variants found in attention-deficit hyperactivity disorder are preferentially associated with neurodevelopmental genes. *Mol Psychiatry* 15:637–646.
88. Yu Y, Lin Y, Takasaki Y, Wang C, Kimura H, Xing J, et al. (2018): Rare loss of function mutations in *N*-methyl-D-aspartate glutamate receptors and their contributions to schizophrenia susceptibility. *Transl Psychiatry* 8:12.
89. Chang X, Lima LdA, Liu Y, Li J, Li Q, Sleiman PMA, et al. (2018): Common and rare genetic risk factors converge in protein interaction networks underlying schizophrenia. *Front Genet* 9:434.

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90. Kathiresan S (2015): Developing medicines that mimic the natural successes of the human genome: Lessons from NPC1L1, HMGCR, PCSK9, APOC3, and CETP. *J Am Coll Cardiol* 65:1562–1566.
91. Farrell M, Lichtenstein M, Crowley JJ, Filmyer DM, Lázaro-Muñoz G, Shaughnessy RA, *et al.* (2018): Developmental delay, treatment-resistant psychosis, and early-onset dementia in a man with 22q11 deletion syndrome and Huntington's disease. *Am J Psychiatry* 175:400–407.
92. Alexandre C, Chaumette B, Martinez G, Christa L, Dupont J-M, Kebir O, *et al.* (2016): Paradoxical improvement of schizophrenic symptoms by a dopaminergic agonist: An example of personalized psychiatry in a CNV carrying patient. *Biol Psychiatry* 80:e21–e23.
93. Sakata Y, Owada Y, Sato K, Kojima K, Hisanaga K, Shinka T, *et al.* (2001): Structure and expression of the glycine cleavage system in rat central nervous system. *Brain Res Mol Brain Res* 94:119–130.
94. Wolosker H, Balu DT, Coyle JT (2016): The rise and fall of the D-serine-mediated gliotransmission hypothesis. *Trends Neurosci* 39:712–721.
95. Kessler M, Terramani T, Lynch G, Baudry M (1989): A glycine site associated with *N*-methyl-D-aspartic acid receptors: Characterization and identification of a new class of antagonists. *J Neurochem* 52:1319–1328.
96. Parsons CG, Danysz W, Quack G, Hartmann S, Lorenz B, Wollenburg C, *et al.* (1997): Novel systemically active antagonists of the glycine site of the *N*-methyl-D-aspartate receptor: Electrophysiological, biochemical and behavioral characterization. *J Pharmacol Exp Ther* 283:1264–1275.
97. Birch PJ, Grossman CJ, Hayes AG (1988): Kynurenic acid antagonises responses to NMDA via an action at the strychnine-insensitive glycine receptor. *Eur J Pharmacol* 154:85–87.
98. Girirajan S, Brkanac Z, Coe BP, Baker C, Vives L, Vu TH, *et al.* (2011): Relative burden of large CNVs on a range of neurodevelopmental phenotypes. *PLoS Genet* 7:e1002334.
99. O'Roak BJ, Deriziotis P, Lee C, Vives L, Schwartz JJ, Girirajan S, *et al.* (2011): Exome sequencing in sporadic autism spectrum disorders identifies severe de novo mutations. *Nat Genet* 43:585–589.
100. David SM, Burger PB, Prakash A, Geballe MT, Yadav R, Le P, *et al.* (2010): Structural determinants of D-cycloserine efficacy at the NR1/NR2C NMDA receptors. *J Neurosci* 30:2741–2754.
101. Baran H, Kepplinger B (2014): D-cycloserine lowers kynurenic acid formation—New mechanism of action. *Eur Neuropsychopharmacol* 24:639–644.
102. Lopes C, Pereira EF, Wu HQ, Purushottamachar P, Njar V, Schwarcz R, *et al.* (2007): Competitive antagonism between the nicotinic allosteric potentiating ligand galantamine and kynurenic acid at alpha7* nicotinic receptors. *J Pharmacol Exp Ther* 322:48–58.
103. Hilmas C, Pereira EF, Alkondon M, Rassoulpour A, Schwarcz R, Albuquerque EX (2001): The brain metabolite kynurenic acid inhibits alpha7 nicotinic receptor activity and increases non-alpha7 nicotinic receptor expression: Physiopathological implications. *J Neurosci* 21:7463–7473.
104. Wu HQ, Pereira EF, Bruno JP, Pellicciari R, Albuquerque EX, Schwarcz R (2010): The astrocyte-derived alpha7 nicotinic receptor antagonist kynurenic acid controls extracellular glutamate levels in the prefrontal cortex. *J Mol Neurosci* 40:204–210.
105. Albuquerque EX, Schwarcz R (2013): Kynurenic acid as an antagonist of $\alpha 7$ nicotinic acetylcholine receptors in the brain: Facts and challenges. *Biochem Pharmacol* 85:1027–1032.
106. Freedman R, Coon H, Myles-Worsley M, Orr-Urtreger A, Olincy A, Davis A, *et al.* (1997): Linkage of a neurophysiological deficit in schizophrenia to a chromosome 15 locus. *Proc Natl Acad Sci U S A* 94:587–592.
107. Xu J, Pato M, Torre C, Medeiros H, Carvalho C, Basile V, *et al.* (2001): Evidence of linkage disequilibrium between the alpha 7-nicotinic receptor gene (CHRNA7) locus and schizophrenia in Azorean families. *Am J Med Genet* 105:669–674.
108. Albuquerque EX, Pereira EF, Alkondon M, Rogers SW (2009): Mammalian nicotinic acetylcholine receptors: From structure to function. *Physiol Rev* 89:73–120.
109. MacDonald JF, Jackson MF, Beazely MA (2006): Hippocampal long-term synaptic plasticity and signal amplification of NMDA receptors. *Crit Rev Neurobiol* 18:71–84.
110. Xu B, Roos JW, Levy S, van Rensburg EJ, Gogos JA, Karayiorgou M (2008): Strong association of de novo copy number mutations with sporadic schizophrenia. *Nat Genet* 40:880–885.
111. Xu B, Woodroffe A, Rodriguez-Murillo L, Roos JL, van Rensburg EJ, Abecasis GR, *et al.* (2009): Elucidating the genetic architecture of familial schizophrenia using rare copy number variant and linkage scans. *Proc Natl Acad Sci U S A* 106:16746–16751.
112. Georgieva L, Rees E, Moran JL, Chambert KD, Milanova V, Craddock N, *et al.* (2014): De novo CNVs in bipolar affective disorder and schizophrenia. *Hum Mol Genet* 23:6677–6682.
113. Schwarcz R, Bruno JP, Muchowski PJ, Wu H-Q (2012): Kynurenic acid in the mammalian brain: When physiology meets pathology. *Nat Rev Neurosci* 13:465–477.
114. Linderholm KR, Alm MT, Larsson MK, Olsson SK, Goiny M, Hajos M, *et al.* (2016): Inhibition of kynurenine aminotransferase II reduces activity of midbrain dopamine neurons. *Neuropharmacology* 102:42–47.
115. Potter MC, Elmer GI, Bergeron R, Albuquerque EX, Guidetti P, Wu HQ, *et al.* (2010): Reduction of endogenous kynurenic acid formation enhances extracellular glutamate, hippocampal plasticity, and cognitive behavior. *Neuropsychopharmacology* 35:1734–1742.
116. Kozak R, Campbell BM, Strick CA, Horner W, Hoffmann WE, Kiss T, *et al.* (2014): Reduction of brain kynurenic acid improves cognitive function. *J Neurosci* 34:10592–10602.
117. Alexander KS, Wu H-Q, Schwarcz R, Bruno JP (2012): Acute elevations of brain kynurenic acid impair cognitive flexibility: Normalization by the alpha 7 positive modulator galantamine. *Psychopharmacology (Berl)* 220:627–637.
118. Williams JB, Mallorga PJ, Conn PJ, Pettibone DJ, Sur C (2004): Effects of typical and atypical antipsychotics on human glycine transporters. *Schizophr Res* 71:103–112.
119. Goff DC, Henderson DC, Evins AE, Amico E (1999): A placebo-controlled crossover trial of D-cycloserine added to clozapine in patients with schizophrenia. *Biol Psychiatry* 45:512–514.
120. Tanahashi S, Yamamura S, Nakagawa M, Motomura E, Okada M (2012): Clozapine, but not haloperidol, enhances glial D-serine and L-glutamate release in rat frontal cortex and primary cultured astrocytes. *Br J Pharmacol* 165:1543–1555.
121. Evins AE, Amico E, Posever TA, Toker R, Goff DC (2002): D-cycloserine added to risperidone in patients with primary negative symptoms of schizophrenia. *Schizophr Res* 56:19–23.
122. Heresco-Levy U, Ermilov M, Shimoni J, Shapira B, Silipo G, Javitt DC (2002): Placebo-controlled trial of D-cycloserine added to conventional neuroleptics, olanzapine, or risperidone in schizophrenia. *Am J Psychiatry* 159:480–482.
123. Potkin SG, Jin Y, Bunney BG, Costa J, Gulasekaram B (1999): Effect of clozapine and adjunctive high-dose glycine in treatment-resistant schizophrenia. *Am J Psychiatry* 156:145–147.
124. Evins AE, Fitzgerald SM, Wine L, Rosselli R, Goff DC (2000): Placebo-controlled trial of glycine added to clozapine in schizophrenia. *Am J Psychiatry* 157:826–828.
125. Tsai G, Yang P, Chung LC, Tsai CW, Coyle JT (1999): D-serine added to clozapine for the treatment of schizophrenia. *Am J Psychiatry* 156:1822–1825.
126. Heresco-Levy U, Javitt DC, Ermilov M, Mordel C, Horowitz A, Kelly D (1996): Double-blind, placebo-controlled, crossover trial of glycine adjuvant therapy for treatment-resistant schizophrenia. *Br J Psychiatry* 169:610–617.
127. Lin C-H, Lin C-H, Chang Y-C, Huang Y-J, Chen P-W, Yang H-T, *et al.* (2018): Sodium benzoate, a D-amino acid oxidase inhibitor, added to clozapine for the treatment of schizophrenia: A randomized, double-blind, placebo-controlled trial. *Biol Psychiatry* 84:422–432.
128. Goff DC (2017): D-cycloserine in schizophrenia: New strategies for improving clinical outcomes by enhancing plasticity. *Curr Neuropharmacol* 15:21–34.