



Paranoid schizophrenia and methamphetamine-induced paranoia are both characterized by a similar LINE-1 partial methylation profile, which is more pronounced in paranoid schizophrenia

Rasmon Kalayasiri^{a,b,c}, Korakot Kraijak^d, Apiwat Mutirangura^c, Michael Maes^{a,e,f,*}

^a Department of Psychiatry, Faculty of Medicine, Chulalongkorn University, Bangkok, Thailand

^b Department of Psychiatry, King Chulalongkorn Memorial Hospital, Bangkok, Thailand

^c Center for Excellence in Molecular Genetics of Cancer and Human Diseases, Department of Anatomy, Faculty of Medicine, Chulalongkorn University, Bangkok, Thailand

^d Master of Science Program in Medical Science, Faculty of Medicine, Chulalongkorn University, Bangkok, Thailand

^e IMPACT Strategic Research Center, Barwon Health, Geelong, Australia

^f Department of Psychiatry, Medical University of Plovdiv, Plovdiv, Bulgaria

ARTICLE INFO

Article history:

Received 26 September 2018

Received in revised form 3 February 2019

Accepted 20 February 2019

Available online 27 February 2019

Keywords:

Methamphetamine

DNA methylation

Schizophrenia

Paranoia

Immune, inflammation

ABSTRACT

Background: There is evidence that schizophrenia is a neuro-immune disorder. Genes linked to intragenic LINE-1 methylation show a strong association with immune-associated disorders including psychosis. The aim of this study was to examine LINE-1 methylation patterns in paranoid schizophrenia and methamphetamine-induced paranoia, a model for schizophrenia.

Methods: This study recruited 31 patients with paranoid schizophrenia, 94 with methamphetamine-induced paranoia (MIP) and 163 normal controls. LINE-1 methylation patterns were assayed in peripheral blood mononuclear cells and a combined bisulphite restriction analysis and COBRA were used to estimate LINE1 methylation (mC) and CpG dinucleotide methylation patterns, namely 2 methylated (mCmC) and 2 unmethylated (uCuC) CpGs and the partially methylated loci mCuC (5'm with 3'u) and uCmC (5'u with 3'm).

Results: Patients with paranoid schizophrenia show highly significant changes in LINE-1 partial methylation patterns, namely a higher percentage of mCuC and lower percentage of uCmC as compared with controls and MIP patients, while the latter show a higher percentage of mCuC but lower percentage of uCmC as compared with controls. Higher mCuC significantly predicts paranoid schizophrenia with a sensitivity of 51.6%, specificity of 97.5% and an area under the ROC curve of 0.895.

Conclusions: The results indicate that a common dysfunction in LINE-1 partial methylation may underpin both paranoid schizophrenia and MIP and that this methylation pattern is significantly more expressed in paranoid schizophrenia than MIP. Reciprocal links between impairments in LINE-1 methylation and neuro-immune and neuro-oxidative pathways may underpin the pathophysiology of both MIP and paranoid schizophrenia.

© 2019 Elsevier B.V. All rights reserved.

1. Introduction

There is now evidence that schizophrenia is characterized by immune-inflammatory alterations, including activated immune-inflammatory responses, a subchronic mild inflammatory response and activated nitro-oxidative processes (Anderson et al., 2013; Davis et al., 2016; Smith and Maes, 1995). Major findings indicate signs of an acute phase response (e.g. increased haptoglobin and complement

factors), M1 macrophagic activation (e.g. increased production of interleukin (IL)-1 β , IL-6, and tumor necrosis factor- α), T helper (Th)-1, Th-2 and Th-17 responses (e.g. increased levels of IL-2, IL-17, IL-4, IL-5, and interferon- γ), a T regulatory (Treg) response (e.g. increased IL-10 levels) and indicators of oxidative stress including lowered lipid-associated antioxidant defenses and increased lipid peroxidation (Brinholi et al., 2015; Maes et al., 1994; Noto et al., 2015a; Noto et al., 2015b; Noto et al., 2014; Noto et al., 2016; Zeni-Graiff et al., 2016). M1 macrophagic and Th-1 products may have detrimental effects on the brain (Leonard and Maes, 2012) collectively named neuroprogression, which includes aberrations in neuronal plasticity, dendrite growth neurogenesis, apoptosis, receptor expression and functioning, including NMDA receptors (Anderson and Maes, 2013; Smith and Maes, 1995). Schizophrenia is also accompanied by changes in leukocyte telomere length with for example a lowered telomere length in individuals at

* Corresponding author at: IMPACT Strategic Research Center, Barwon Health, Deakin University, Geelong, Vic, Australia.

E-mail addresses: rasmon.k@chula.ac.th (R. Kalayasiri), apiwat.m@chula.ac.th (A. Mutirangura), dr.michaelmaes@hotmail.com, (M. Maes).

URL: <https://scholar.google.co.th/citations?user=1wzMZ7UAAAAJ&hl=th&oi=ao> (M. Maes).

high risk for psychosis (Maurya et al., 2018; Maurya et al., 2017). There are also data that schizophrenia is accompanied by changes in DNA methylation patterns although a recent systematic review showed inconsistent findings, including hypermethylation, hypomethylation or no changes in global methylation patterns (Teroganova et al., 2016).

Most of the DNA methylation occurs in regions with a high percentage of CpG sites that are concentrated in promoter regions, namely the 5'-CpG-3' islands and this methylation process may be accompanied by gene silencing (Bird, 2002; Kitkumthorn and Mutirangura, 2011). Many studies measured cytosine methylation of LINE-1 (long interspersed element-1s), which are interspersed repetitive sequences and retrotransposons (Kitkumthorn and Mutirangura, 2011). LINE-1 methylation patterns play a key role in maintaining genomic integrity whilst intragenic LINE-1 mediates gene expression in cis (Kemp and Longworth, 2015; Kitkumthorn and Mutirangura, 2011; Wangsri et al., 2012). LINE-1 hypomethylation may cause repression of gene expression, genomic instability, and aberrations in DNA repair genes and DNA double-strand break repair (Aporntewan et al., 2011; Pornthanakasem et al., 2008; Wangsri et al., 2012). As such, hypomethylation of promoter LINE-1 retrotransposable elements is frequently associated with human disorders including cancer (Chalitchagorn et al., 2004; Dammann et al., 2005). Most importantly, genes associated with the cis-regulatory activities of intragenic LINE-1 show an association with immune and oxidative pathways, apoptosis and cell differentiation and many (auto)immune and neuroprogressive diseases, possibly including schizophrenia (Wanichopparat et al., 2013).

Recently, we have shown that the assay of LINE-1 methylation patterns with the combined bisulphite restriction analysis of LINE-1s (COBRALINE-1) methylation is superior as compared with measurements of overall LINE1 methylation (Kitkumthorn et al., 2012a; Kitkumthorn et al., 2012b). This method allows to measure LINE-1 overall methylation levels as well as to classify LINE-1 alleles into 4 patterns based on the methylation status of two CpG dinucleotides on each strand from 5' to 3', namely 2 methylated (mCmC) and 2 unmethylated (uCuC) CpGs and two types of partially methylated loci (mCuC that is 5' m with 3'u and uCmC that is 5'u with 3'm CpGs) (Wangsri et al., 2012). For example, in peripheral blood mononuclear cells (PBMCs), the uCuC LINE-1 hypomethylated loci show a significantly better diagnostic performance for oral cancer than overall methylation (Kitkumthorn et al., 2012b). Nevertheless, there are no data on COBRALINE-1 assays in schizophrenia.

Psychosis including paranoia and schizophrenia-like cognitive deficits are frequently induced by use of methamphetamine (MA) (streetnames: meth, ice, crystal, yaba), a psychostimulant substance (Hsieh et al., 2014; Kalayasiri et al., 2009; Kalayasiri et al., 2014). Such effects are ascribed to enhanced release and turnover of dopamine and glutamate in the brain with ensuing damage to cortical GABAergic neurons (Hsieh et al., 2014; Vasan and Olango, 2018). Nevertheless, MA may activate immune-inflammatory and oxidative pathways, for example by effects on DNA oxidation, the anti-cholinergic anti-inflammatory pathway and gut microbiota causing increased gut permeability (Prakash et al., 2017; Ramkissoon and Wells, 2015). Such effects may at least in part determine MA-induced neuroprogressive effects including glutamate neurotoxicity, damage to neuronal dendrites, neuronal death in frontal, prefrontal and temporal lobes, white matter gliosis and hypertrophy (Prakash et al., 2017; Thompson et al., 2004), which in turn, may cause MA-induced paranoia (MIP) and deficits in information processing, episodic memory and executive functions (Hsieh et al., 2014; Prakash et al., 2017; Thompson et al., 2004). Interestingly, MIP is considered to be a model for schizophrenia (Hsieh et al., 2014; Shelly et al., 2016). Recently, we detected that MA use increases changes in LINE-1 partial methylation, namely an increased percentage of mCuC and mCuC plus uCmC, and that MIP is associated with a lowered percentage of uCmC (Kalayasiri et al., in press). Nevertheless, no studies have directly compared LINE-1 methylation profiles between paranoid schizophrenia and MIP.

Hence, the aim of the present study was to examine COBRALINE-1-derived methylation profiles between patients with paranoid schizophrenia and MIP and normal controls. Similar changes in partial LINE-1 methylation patterns in both MIP and paranoid schizophrenia may point towards a common pathophysiology related to aberrations in LINE-1 methylation.

2. Subjects and methods

2.1. Participants

We recruited subject with MIP who were admitted to the Princess Mother National Institute on Drug Abuse Treatment (PMNIDAT) and patients with paranoid schizophrenia who were treated at the Department of Psychiatry, King Chulalongkorn Memorial Hospital, Bangkok, Thailand. Normal controls were recruited by word of mouth at the blood donation center, Thai Red Cross Society, Bangkok, Thailand. All participants were Thai nationals of both genders and 18–65 years old. The present LINE-1 study included controls and MIP patients as published in our previous paper (Kalayasiri et al., in press), however, we added 66 new controls and 2 new MIP patients, while this is a first publication on LINE-1 patterns in paranoid schizophrenia patients. The diagnostic assessments of substance use were made employing the Thai version of the Semi-Structured Assessment for Drug Dependence and Alcoholism (SSADDA), while the diagnosis of schizophrenia, paranoid subtype, and MIP were made using the Diagnostic and Statistical Manual of Mental Disorders (DSM-IV) (American Psychiatric Association, 2000) and Methamphetamine Experience Questionnaire (MEQ) – Thai version (Kalayasiri et al., 2014) by exploring paranoid experiences during MA use. We have excluded paranoid schizophrenia patients who were diagnosed with substance use disorder, while we excluded MIP subjects who were diagnosed with primary psychotic disorders or schizophrenia. Moreover, we excluded patients and controls with a lifetime history of bipolar disorder, major depressive disorder, psychorganic disorders; neurodegenerative / neuroinflammatory disorders (e.g. Parkinson's disease, multiple sclerosis and stroke), neurologic disorders (e.g. epilepsy and brain trauma) and (auto)immune disorders (e.g. systemic lupus erythematosus). The study was approved by the Human Ethics Committee of the Faculty of Medicine, Chulalongkorn University (Med Chula IRB #417/57).

2.2. Measurements

Socio-demographic data and substance use variables and related diagnoses were obtained by trained clinical psychologists certified for SSADDA interview employing the SSADDA. The principal investigator (R.K.) made the diagnoses of MIP and paranoid schizophrenia using the SSADDA, MEQ and DSM-IV-TR criteria for paranoid schizophrenia. The MEQ shows a good inter-instrument reliability ($K = 0.87$) (Kalayasiri et al., 2014). The body mass index (BMI) was computed as body weight (kg)/length (meter)². Current smoking (last 6 months) and use of psychotropic medications was registered as dummy variables (yes/no).

2.3. DNA extraction, bisulfite modification, and COBRALINE-1s

DNA extraction and COBRALINE-1 assays were carried out at the Center for Excellence in Molecular Genetics of Cancer and Human Diseases, Faculty of Medicine, Chulalongkorn University, Bangkok, Thailand. Blood samples were centrifuged at 1000g for 10 min to collect PBMCs and stored at -80°C until assayed for COBRALINE-1 assays. Whole blood was extracted by 1) adding lysis buffer with 10% sodium dodecyl sulfate and proteinase K and then incubated overnight at 50°C , 2) purifying using phenol-chloroform and centrifuging at 4°C with 14,000 g for 15 min, 3) precipitating the DNA pellet using 10 Molar

ammonium acetate and absolute ethanol, 4) washing DNA pellet by 70% ethanol, and 5) drying DNA pellet and dissolving it by Tris-EDTA.

The combined bisulphite restriction analysis of LINE-1s (COBRALINE-1) was used for determining pattern of LINE-1s methylation. A total of 1 microgram (µg) of DNA was used in the bisulphite treatments that converted unmethylated cytosine to uracil, while methylated cytosine was not changed. The bisulphite DNA modification was performed using the EZ-DNA methylation kit and specific primers is LINE-1s-F (5'GTAAAGAAAGGGTGA YGGT-3') and LINE-1s-R (5' AATACRCRTTTTCTTAAACC RATCTA-3') at 95 °C denature for 15 min, 50 °C annealing for 35 cycles and 72 °C final extension. LINE-1s were digested with *TaqI* and *TasI* at 65 °C overnight. The digested products of bisulphite-treated LINE-1s were separated to strands with different length including 92 (^mC^uC), 60 (^uC^uC), 50 (^mC^mC), 42 (^mC^mC and ^uC^mC), and 32 (^uC^uC and ^uC^mC) base pairs (bp) that were measured by using polyacrylamide gel electrophoresis and stained with SYBR. We used deionized water as a negative control and HeLa, Daudu, and Jurkat as positive control. Fig. 1 shows the pattern of LINE-1s methylation and the separated strands with different lengths including 92 (^mC^uC), 60 (^uC^uC), 50 (^mC^mC), 42 (^mC^mC and ^uC^mC) and 32 (^uC^uC and ^uC^mC) base pairs (bp). The intensity of each band was assigned into A, B, C, D, E (e.g., A = %92/92, B = %60/56, C = %50/48, D = %42/40, E = %32/28). The intensity of 18 bp was calculated and assigned to F = ((D + E) - (B + C))/2). Percentages of each patterns of DNA methylation

were calculated by using the following formula:

$$\%mC \text{ methylation} = ((A + 2C + F) * 100)/(2A + 2B + 2C + 2F)$$

$$\%mCmC \text{ hypermethylation} = ((C/2) * 100)/((C/2) + A + B + F)$$

$$\%uCmC \text{ partial methylation} = (F * 100)/((C/2) + A + B + F)$$

$$\%mCuC \text{ partial methylation} = (A * 100)/((C/2) + A + B + F)$$

$$\%uCuC \text{ hypomethylation} = (B * 100)/((C/2) + A + B + F)$$

2.4. Statistical analyses

We used analysis of contingency tables (χ^2 -tests) to check associations between categorical variables and used analysis of variance (ANOVA) to check differences in scale variables between diagnostic groups. We used multivariate general linear model (GLM) analysis to examine the associations between diagnosis (three groups, namely paranoid schizophrenia, MIP and controls) and LINE-1 methylation patterns while adjusting for sex, age and BMI. Consequently, we used tests for between-subject effects to assess the associations between diagnosis and the separate LINE-1 methylation data. Model-generated estimated

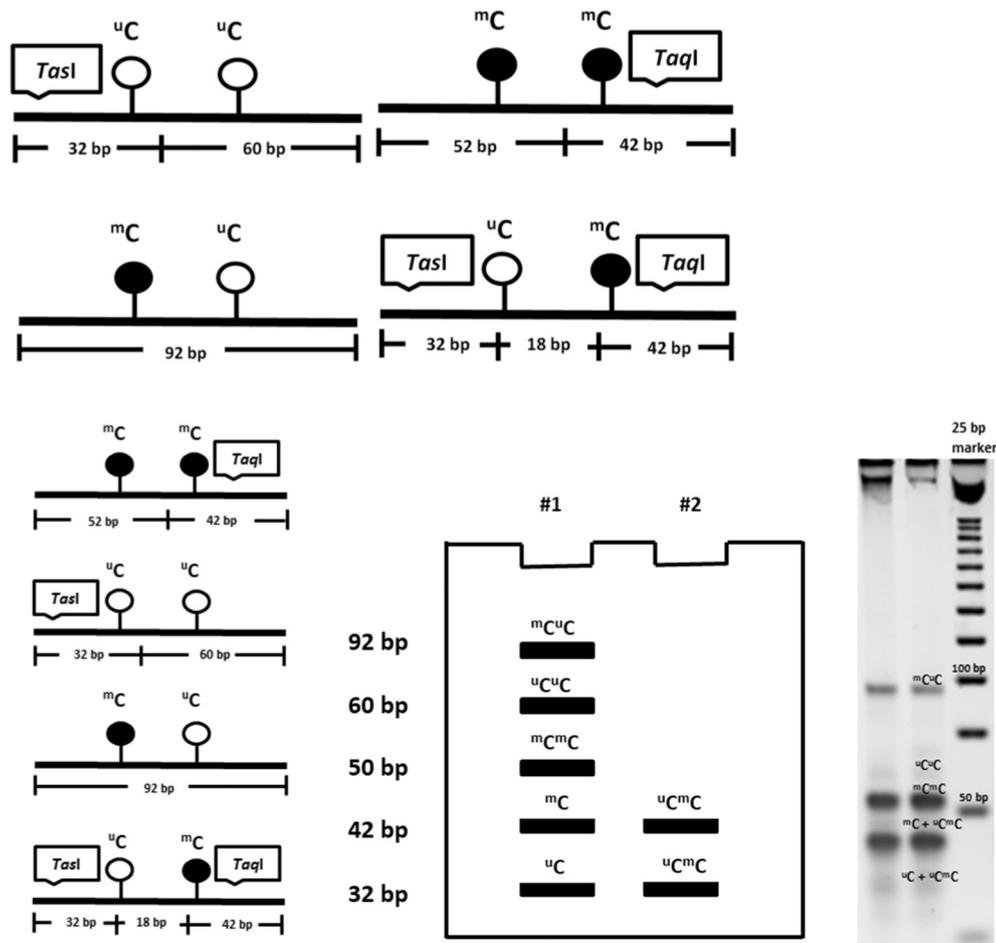


Fig. 1. Pattern of LINE-1s methylation. This figure shows the combined bisulphite restriction analysis of LINE-1s (COBRALINE-1). The digested products of bisulphite-treated LINE-1s were separated to strands with different length including 92 (^mC^uC), 60 (^uC^uC), 50 (^mC^mC), 42 (^mC^mC and ^uC^mC), and 32 (^uC^uC and ^uC^mC) base pairs (bp).

marginal mean values were computed and protected post-hoc analyses were used to assess pairwise differences between categories. We used binary logistic regression analysis to define the most important predictors of paranoid schizophrenia versus controls, paranoid schizophrenia versus MIP and paranoid schizophrenia versus MIP + controls. Odds ratios and 95% confidence intervals were computed, while Nagelkerke values were used to estimate effect size. We used the area under the Receiver Operating Curve (ROC) coupled with sensitivity and specificity to estimate the overall diagnostic performance. In addition, we computed 2 different z weighted composite scores based on the partial LINE-1 methylation data, namely a) sum of the two partial methylation data as z transformation of % mCuC (z mCuC) + z uCmC (reflecting overall partial LINE-1 methylation), and b) z mCuC – z uCmC (reflecting the shift among these partial methylation profiles). All statistical analyses were performed using IBM SPSS windows version 24. Tests were 2-tailed and an alpha level of 0.05 indicated statistically significant results.

3. Results

Table 1 shows the socio-demographic and biomarker data in paranoid schizophrenia, MIP and controls. There were no significant differences in age and BMI between the study groups, while there were more males in the MIP group as compared with controls and paranoid schizophrenia. This table also shows the estimated-marginal mean values of the LINE-1 methylation data in the three diagnostic groups after adjusting for age, sex and BMI. Fig. 2 shows the LINE-1 methylation profile displayed as z values (mean = 0 and standard deviation = 1) in the three diagnostic groups. Table 2 shows the results of multivariate GLM analysis with 7 DNA methylation data as dependent variables and diagnosis (three groups) as primary explanatory variable while adjusting for age, sex and BMI. There was a highly significant effect of diagnosis with a partial eta squared = 0.151, after considering the effects of age, sex and BMI, which were not significant. Table 2 shows the results of the tests for between-subject effects and protected post-hoc analysis. There was a strong association between diagnosis and % mCuC (partial eta squared = 0.217). % mCuC was significantly different between the three groups and increased from controls → MIP → paranoid schizophrenia. There were also significant differences in % uCmC between the groups, although the effect size was smaller (partial eta squared = 0.056). % uCmC decreased from controls → MIP → paranoid schizophrenia. There were also significant differences in the z composite scores z mCuC + z uCmC (effects size = 0.051) and z mCuC – z uCmC (effect size = 0.165). The z mCuC + z uCmC score was significantly higher in both patient groups as compared with controls, while the z

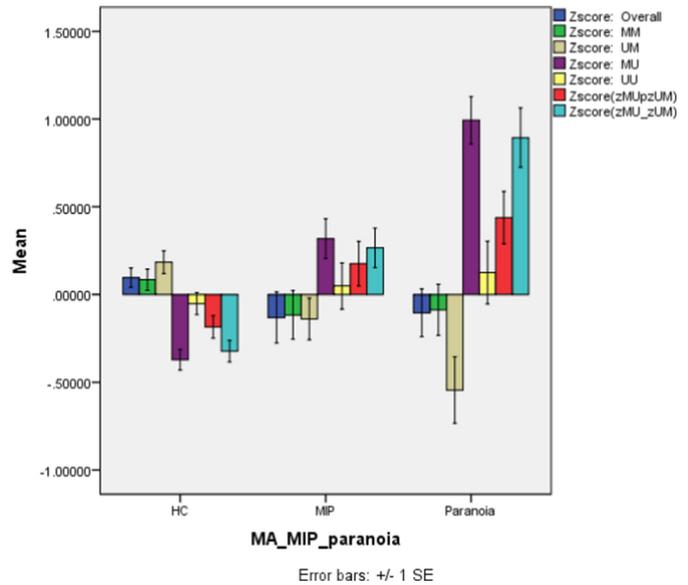


Fig. 2. LINE-1 methylation profile in paranoid schizophrenia. LINE-1 methylation profile is displayed as z values (mean = 0 and standard deviation = 1) in healthy controls (HC), methamphetamine-induced paranoia (MIP) and paranoid schizophrenia (paranoia). MM/UU: 2 methylated (mCmC; MM) and 2 unmethylated (uCmC; UU) CpGs. UM/MU: partially methylated loci mCuC (5'm with 3'u; MU) and uCmC (5'u with 3'm; UM). zMU+zUM: computed as z transformations of % mCuC (z mCuC) + z uCmC (reflecting overall partial LINE-1 methylation). zMU-zUM: computed as z mCuC – z uCmC (reflecting the shift among these partial methylation profiles).

mCuC – z uCmC score was significantly different between the three groups and increased from controls → MIP → paranoid schizophrenia.

We have also examined whether there were any effects of smoking and drug use on the LINE-1 methylation data by entering these data in GLM analysis. Table 2 shows the results of these GLM analyses. There were no significant effects of smoking (versus not smoking), and use of neuroleptics, atypical antipsychotics, antidepressants and benzodiazepines on the results.

Table 3 shows the results of binary logistic regression analyses with paranoid schizophrenia or paranoid schizophrenia + MIP (any psychosis) as dependent variables and the LINE-1 methylation data (alone or combined with sex, age and BMI) as independent variables. We found

Table 1
Demographic data and model-generated estimated marginal mean values of LINE-1 methylation data in healthy controls (HC), patients with methamphetamine-induced paranoia (MIP) and patients with schizophrenia, paranoid subtype (SCZ).

Variables	HC ^A	MIP ^B	SCZ ^C	F/X ²	df	p
Age (years)	33.8 (11.6)	33.0 (6.2)	37.2 (9.6)	2.11	2/285	0.122
Sex (Female/male)	82/81 ^B	31/63 ^{A,C}	18/13 ^B	9.44	2	0.009
BMI (kg/m ²)	23.9 (4.3)	25.0 (2.2)	23.8 (4.3)	2.88	2/285	0.058
% mC	67.1 (0.2)	66.6 (0.3)	66.5 (0.5)	1.25	1/282	0.288
% mCmC	33.9 (0.3)	33.3 (0.4)	33.3 (0.6)	1.06	1/282	0.349
% uCmC	32.3 (0.1) _{B,C}	31.8 (0.2) _{A,C}	31.0 (0.3) _{A,B}	8.30	1/282	<0.001
% mCuC	18.6 (0.2) _{B,C}	20.3 (0.2) _{A,C}	21.9 (0.4) _{A,B}	39.01	1/282	<0.001
% uCuC	21.9 (0.2)	22.0 (0.3)	22.4 (0.2)	0.63	1/282	0.631
z mCuC + z uCmC	-0.193 (0.077) _{B,C}	0.186 (0.104) ^A	0.419 (0.177) ^A	7.65	1/282	0.001
z mCuC – z uCmC	-0.322 (0.072) _{B,C}	0.244 (0.098) _{A,C}	0.895 (0.166) _{A,B}	27.96	1/282	<0.001

Results are shown as mean (SD).
^{A,B,C}: Significant differences between group means.
BMI: body mass index.

Table 2
Associations between diagnostic groups, healthy controls (HC), paranoid schizophrenia (SCZ) and methamphetamine-induced paranoia (MIP) and LINE-1 methylation, while adjusting for background variables.

Multivariate tests	Dependent variables	Explanatory variables	F	df	p
#1	All 7 methylation data	HC/MIP/SCZ	10.12	10/558	<0.001
		Age	0.26	5/278	0.936
		Female/Male	1.33	5/278	0.253
		BMI	1.43	5/278	0.213
#2	All 7 methylation data	Smoking	1.68	5/215	0.141
#3	All 7 methylation data	Neuroleptics	0.07	5/110	0.997
#4	All 7 methylation data	Atypical antipsychotics	0.31	5/110	0.905
#5	All 7 methylation data	Antidepressants	0.51	5/110	0.765
#6	All 7 methylation data	Benzodiazepines	0.47	5/110	0.799

HC/MIP/SCZ: healthy controls, methamphetamine-induced paranoia and schizophrenia, paranoia subtype.
BMI: body mass index.
7 methylation data: entered are % mC, % mCmC, % uCmC, % mCuC, % uCuC, z uCmC + z mCuC, and z mCuC – z uCmC.

Table 3

Results of binary logistic regression analyses with healthy controls (HC), paranoid schizophrenia (SCZ), and methamphetamine-induced paranoia (MIP) as dependent variables and LINE-1 methylation patterns, demographic and clinical data as explanatory variables.

Dichotomy	Explanatory variables	Wald	df	p	OR	95% CI	Nagelkerke ROC curve (SE)
#1. SCZ versus HC	% mCuC	33.53	1	<0.001	12.28	5.26–28.71	Nagelkerke: 0.513 ROC: 0.895 (0.030)
#2. SCZ + MIP versus controls	% mCuC	43.65	1	<0.001	2.93	2.13–4.04	Nagelkerke: 0.246 ROC: 0.740 (0.030)
#3. SCZ versus MIP	% mCuC	8.67	1	0.003	2.03	1.26–3.26	Nagelkerke: 0.314 ROC: 0.720 (0.049)
	Sex	10.03	1	0.002	0.19	0.07–0.53	
	Age	4.96	1	0.026	1.07	1.01–1.14	
	BMI	6.72	1	0.010	0.81	0.70–0.95	
#4. SCZ versus HC + MIP	% mCuC	27.03	1	<0.001	3.39	2.14–5.37	Nagelkerke: 0.274 ROC: 0.831 (0.034)
	Sex	5.41	1	0.020	0.36	0.15–0.85	
	Age	4.41	1	0.036	1.05	1.03–1.09	

OR: Odd's ratio.

95% CI: 95% confidence interval, lower and upper limit.

(regression #1) that increased % mCuC significantly predicted paranoid schizophrenia versus controls with a Nagelkerke value of 0.513. The area under the ROC was 0.895 and 90.2% of all cases were correctly classified with a sensitivity = 51.6% and specificity = 97.5%. Regression #2 shows that any psychosis was best predicted by % mCuC with an effect size = 0.246. The area under the ROC was 0.740 and 70.5% of all cases were correctly classified with a sensitivity = 59.3% and specificity = 79.1%. % mCuC was also significant in discriminating paranoid schizophrenia from MIP and from MIP + controls.

4. Discussion

The first major finding of this study is that PBMCs of paranoid schizophrenia patients show a higher percentage of mCuC and lowered percentage of uCmC DNA partial methylation as compared with controls and that the increase in percentage of mCuC partial methylation was predictive for paranoid schizophrenia with an area under the ROC of 0.895. These findings show that paranoid schizophrenia is accompanied by aberrations in LINE-1 partial methylation, while there are no significant changes in overall LINE1 methylation and no signs of LINE-1 hypermethylation or hypomethylation. These data show that classifying LINE-1 alleles into four classes based on the methylation status of 2 CpG dinucleotides, including partial methylation profiles, provides more specific information than the assessment of LINE-1 methylation alone. Previously, it was reported that the COBRALINE-1 patterns (namely uCuC hypomethylated LINE-1 loci) assayed in PBMCs better discriminate patients with oral cancer from normal controls as compared with LINE1 overall methylation data (Kitkumthorn et al., 2012a). Phrased differently, if we had assayed the latter only we would have obtained negative results, while our COBRALINE-1 analyses show a highly significant association with partial methylation patterns. Our results are difficult to compare with previous DNA and LINE-1 methylation data in schizophrenia because we assayed a more complete LINE-1 methylation pattern, while previous studies in schizophrenia measured global methylation. Likewise, a systematic review on DNA methylation in schizophrenia (Teroganova et al., 2016) reported inconsistent results pointing towards hypo- or hypermethylation or no changes at all.

The second major finding of this study is that both paranoid schizophrenia and MIP show a similar LINE-1 methylation profile, namely an increased percentage of mCuC and lowered percentage of uCmC partial LINE-1 methylation compared with normal controls, while these changes are significantly more pronounced in paranoid schizophrenia as compared with MIP. This is important as MIP is considered to be a model of schizophrenia (Hsieh et al., 2014). Indeed, MA is frequently accompanied by paranoid symptoms and cognitive impairments, including episodic memory and executive functions, which are quite similar with those observed in paranoid schizophrenia (Glasner-Edwards and Mooney, 2014; Hsieh et al., 2014; Prakash et al., 2017; Thompson et al., 2004). The latter authors ascertain that the differential diagnosis

of primary schizophrenia versus MIP may sometimes be challenging although MIP is frequently a transient condition and defined by a temporal relationship between MA use and the consequent psychosis, while “schizophrenia is not related to drug intoxication or withdrawal” (Glasner-Edwards and Mooney, 2014; World Health Organization, 1992). In another study, we established that use of MA increases mCuC LINE-1 methylation percentage but does not change uCmC LINE-1 methylation percentage and that MIP is accompanied by significantly a lowered percentage of uCmC (Kalayasiri et al., in press). Phrased differently, aberrations in both partial methylation patterns and a shift from a lowered percentage of uCmC towards an increased percentage of mCuC is a characteristic of paranoia, either drug-induced or primary paranoia. If aberrations in LINE-1 methylation patterns are causally linked to psychosis, we may hypothesize that a) MA may increase the vulnerability to paranoia by increasing mCuC partial methylation; and b) the greater dysfunctions in LINE-1 methylation in paranoid schizophrenia versus MIP could reflect differences in severity of psychosis with a worse phenomenology in paranoid schizophrenia than MIP.

Previously, it was proposed that an increased release of central dopamine and glutamate and damage to GABA-ergic neurons in the cortex could underpin both MIP and paranoid schizophrenia (Hsieh et al., 2014; Vasan and Olango, 2018). Nevertheless, here we show that an epigenetic process may constitute a common pathophysiology underpinning both types of paranoid psychoses. There is now some evidence that reactive oxygen species may induce changes in LINE-1 methylation (Narvaez et al., 2017). LINE-1 methylation is reduced in cells treated with peroxides (Kloypan et al., 2015), while antioxidants may restore LINE-1 hypomethylation. In UM-UC-3 bladder cell (carcinoma) lines, peroxides induce LINE-1 hypomethylation which could be restored by alpha-tocopherol, while treatment with peroxides increased methylation of the runt-related transcription factor 3 (RUNX3) promoter, a tumor suppressor gene, which was attenuated by alpha-tocopherol (Wongpaiboonwattana et al., 2013). Interestingly, in biliary atresia patients, LINE-1 hypomethylation is associated with signs of oxidative stress, while LINE-1 methylation is positively associated with relative telomere length (Udomsinprasert et al., 2016). In this respect, it is known that DNA methylation processes in subtelomeric DNA repeats are an important factor in the regulation of telomere length (Yehezkel et al., 2008). Moreover, inflammatory biomarkers (including ICAM-1) are negatively associated with leukocyte telomere length and global DNA methylation, which may increase genome instability (Dong et al., 2017). By inference, the neuro-immune and neuro-oxidative pathways that characterize schizophrenia and are inducible by MA use could have affected LINE-1 methylation and consequently telomere length (see Introduction). Importantly, genes linked with intragenic LINE-1 show a strong association with oxidative, inflammatory, immune and apoptotic processes as well as cell differentiation and with many autoimmune, immune, and neurodegenerative disorders, possibly also with

schizophrenia (Wanichnopparat et al., 2013). These new epigenomic findings do not rule out that activated glutamate release and damage to GABA-ergic neurons in the brain may underpin MIP and paranoid schizophrenia (Hsieh et al., 2014). It was explained previously that activation of neuro-immune and neuro-oxidative pathways may cause increased glutamate toxicity leading to a number of neuronal dysfunctions including in GABA-ergic neurons (Morris et al., 2018; Morris et al., 2016). Interestingly, LINE-1 methylation was reduced in individuals with autism spectrum disorder with severe language impairment (Tangsuwansri et al., 2018).

In conclusion, the results indicate that a shift in LINE-1 partial methylation patterns from lower uCmC to increased mCuC may underpin paranoid schizophrenia and MIP and that this pattern is significantly more pronounced in paranoid schizophrenia than MIP. Schizophrenia is now conceptualized as a neuro-immune and neuro-oxidative disorder, while MA use may induce these pathways. Induction of these paths may lead to alterations in LINE-1 methylation patterns, while genes linked with intragenic LINE-1 show an association with many neuro-immune disorders, including schizophrenia. Future research should examine the reciprocal links between neuro-immune and neuro-oxidative pathways and impairments in LINE-1 partial methylation in relation to the pathophysiology of both MIP and paranoid schizophrenia.

Acknowledgement

We thank Mr. Prakasit Rattanatyong and Dr. Nakarin Kitkumthorn for excellent technical support in laboratory. We appreciate Ms. Maturada Phetsung and Ms. Sirapat Setayanon for the helps on laboratory work. We thank Drs Dolnapha Rattanakorn and Thitiporn Supasitthumrong for DNA samplings, Mr. Wuthichai Hasook for healthy controls recruitment, and staffs at the Princess Mother National Institute on Drug Abuse Treatment for facilitating data collection.

Conflict of interest

The authors have no conflict of interest with any commercial or other association in connection with the submitted article.

Contributors

All authors contributed to interpretation of the data and writing of the manuscript.

Role of funding source

This research has been supported by National Science and Technology Development Agency (NSTDA), Thailand (A.M.). RK is supported by the Center for Alcohol Studies, Thailand and by the Fogarty International Center of the National Institutes of Health (NIH) under the subaward of D43TW009087 (Yale University School of Medicine) (Joel Gelernter, M.D. and Robert T. Malison, M.D.). This funding source had no role in the design and conduct of the study, collection, management, analysis and interpretation of the data, preparation, review, or approval of the manuscript and decision to submit the manuscript for publication.

References

American Psychiatric Association (Ed.), 2000. *Diagnostic and Statistical Manual of Mental Disorders*, 4th ed American Psychiatric Association, Washington, D.C.

Anderson, G., Maes, M., 2013. Schizophrenia: linking prenatal infection to cytokines, the tryptophan catabolite (TRYCAT) pathway, NMDA receptor hypofunction, neurodevelopment and neuroprogression. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 42, 5–19.

Anderson, G., Berk, M., Dodd, S., Bechter, K., Altamura, A.C., Dell'osso, B., Kanba, S., Monji, A., Fatemi, S.H., Buckley, P., Debnath, M., Das, U.N., Meyer, U., Muller, N., Kanchanatawan, B., Maes, M., 2013. Immuno-inflammatory, oxidative and nitrosative stress, and neuroprogressive pathways in the etiology, course and treatment of schizophrenia. *Prog. Neuro-Psychopharmacol. Biol. Psychiatry* 42, 1–4.

Aporntewan, C., Phokaew, C., Piriyaopongsa, J., Ngamphiw, C., Ittiwut, C., Tongsimma, S., Mutirangura, A., 2011. Hypomethylation of intragenic LINE-1 represses transcription in cancer cells through AGO2. *PLoS One* 6 (3), e17934.

Bird, A., 2002. DNA methylation patterns and epigenetic memory. *Genes Dev.* 16 (1), 6–21.

Brinholi, F.F., Noto, C., Maes, M., Bonifácio, K.L., Brietzke, E., Ota, V.K., Gadelha, A., Cordeiro, Q., Belangero, S.I., Bressan, R.A., Vargas, H.O., Higachi, L., de Farias, C.C., Moreira, E.G., Barbosa, D.S., 2015. Lowered paraoxonase 1 (PON1) activity is associated with increased cytokine levels in drug naive first episode psychosis. *Schizophr. Res.* 166 (1–3), 225–230.

Chalitthagorn, K., Shuangshoti, S., Hourpai, N., Kongruttanachok, N., Tangkijvanich, P., Thong-ngam, D., Voravud, N., Sriuranpong, V., Mutirangura, A., 2004. Distinctive pattern of LINE-1 methylation level in normal tissues and the association with carcinogenesis. *Oncogene* 23 (54), 8841–8846.

Dammann, R., Strunnikova, M., Schagdarsurengin, U., Rastetter, M., Papritz, M., Hattenhorst, U.E., Hofmann, H.S., Silber, R.E., Burdach, S., Hansen, G., 2005. CpG island methylation and expression of tumour-associated genes in lung carcinoma. *Eur. J. Cancer* 41 (8), 1223–1236.

Davis, J., Eyre, H., Jacka, F.N., Dodd, S., Dean, O., McEwen, S., Debnath, M., McGrath, J., Maes, M., Amminger, P., McGorry, P.D., Pantelis, C., Berk, M., 2016. A review of vulnerability and risks for schizophrenia: beyond the two hit hypothesis. *Neurosci. Biobehav. Rev.* 65, 185–194.

Dong, Y., Huang, Y., Gutin, B., Raed, A., Dong, Y., Zhu, H., 2017. Associations between global DNA methylation and telomere length in healthy adolescents. *Sci. Rep.* 7 (1), 4210.

Glasner-Edwards, S., Mooney, L.J., 2014. Methamphetamine psychosis: epidemiology and management. *CNS Drugs* 28 (12), 1115–1126.

Hsieh, J.H., Stein, D.J., Howells, F.M., 2014. The neurobiology of methamphetamine induced psychosis. *Front. Hum. Neurosci.* 8, 537.

Kalayasiri, R., Mutirangura, A., Verachai, V., Gelernter, J., Malison, R.T., 2009. Risk factors for methamphetamine-induced paranoia and latency of symptom onset in a Thai drug treatment cohort. *Asian Biomedicine* 3 (6), 635–643.

Kalayasiri, R., Verachai, V., Gelernter, J., Mutirangura, A., Malison, R.T., 2014. Clinical features of methamphetamine-induced paranoia and preliminary genetic association with DBH-1021C→T in a Thai treatment cohort. *Addiction* 109 (6), 965–976.

Kalayasiri, R., Kraijak, K., Maes, M., Mutirangura, A., 2019. Methamphetamine (MA) use induces specific changes in LINE-1 partial methylation patterns, which are associated with MA-induced paranoia: a multivariate and neuronal network study. *Mol. Neurobiol.* in press.

Kemp, J.R., Longworth, M.S., 2015. Crossing the LINE toward genomic instability: LINE-1 retrotransposition in cancer. *Front Chem* 3, 68.

Kitkumthorn, N., Mutirangura, A., 2011. Long interspersed nuclear element-1 hypomethylation in cancer: biology and clinical applications. *Clin. Epigenetics* 2 (2), 315–330.

Kitkumthorn, N., Keelawat, S., Rattanatyong, P., Mutirangura, A., 2012a. LINE-1 and Alu methylation patterns in lymph node metastases of head and neck cancers. *Asian Pac. J. Cancer Prev.* 13 (9), 4469–4475.

Kitkumthorn, N., Tuangsintanakul, T., Rattanatyong, P., Tiwawech, D., Mutirangura, A., 2012b. LINE-1 methylation in the peripheral blood mononuclear cells of cancer patients. *Clin. Chim. Acta* 413 (9–10), 869–874.

Kloypan, C., Srisa-art, M., Mutirangura, A., Boonla, C., 2015. LINE-1 hypomethylation induced by reactive oxygen species is mediated via depletion of S-adenosylmethionine. *Cell Biochem. Funct.* 33 (6), 375–385.

Leonard, B., Maes, M., 2012. Mechanistic explanations how cell-mediated immune activation, inflammation and oxidative and nitrosative stress pathways and their sequels and concomitants play a role in the pathophysiology of unipolar depression. *Neurosci. Biobehav. Rev.* 36 (2), 764–785.

Maes, M., Meltzer, H.Y., Bosmans, E., 1994. Immune-inflammatory markers in schizophrenia: association to normal controls and effects of clozapine. *Acta Psychiatr. Scand.* 89 (5), 346–351.

Maurya, P.K., Rizzo, L.B., Xavier, G., Tempaku, P.F., Zeni-Graiff, M., Santoro, M.L., Mazzotti, D.R., Zugman, A., Pan, P., Noto, C., Maes, M., Asevedo, E., Mansur, R.B., Cunha, G.R., Gadelha, A., Bressan, R.A., Belangero, S.I., Brietzke, E., 2017. Shorter leukocyte telomere length in patients at ultra high risk for psychosis. *Eur. Neuropsychopharmacol.* 27 (5), 538–542.

Maurya, P.K., Rizzo, L.B., Xavier, G., Tempaku, P.F., Ota, V.K., Santoro, M.L., Spindola, L.M., Moretti, P.S., Mazzotti, D.R., Gadelha, A., Gouvea, E.S., Noto, C., Maes, M., Cordeiro, Q., Bressan, R.A., Brietzke, E., Belangero, S.I., 2018. Leukocyte telomere length variation in different stages of schizophrenia. *J. Psychiatr. Res.* 96, 218–223.

Morris, G., Carvalho, A.F., Anderson, G., Galecki, P., Maes, M., 2016. The many neuroprogressive actions of tryptophan catabolites (TRYCATs) that may be associated with the pathophysiology of neuro-immune disorders. *Curr. Pharm. Des.* 22 (8), 963–977.

Morris, G., Berk, M., Carvalho, A.F., Maes, M., Walker, A.J., Puri, B.K., 2018. Why should neuroscientists worry about iron? The emerging role of ferroptosis in the pathophysiology of neuroprogressive diseases. *Behav. Brain Res.* 341, 154–175.

Narvaez, D.M., Groot, H., Diaz, S.M., Palma, R.M., Munoz, N., Cros, M.P., Hernandez-Vargas, H., 2017. Oxidative stress and repetitive element methylation changes in artisanal gold miners occupationally exposed to mercury. *Heliyon* 3 (9), e00400.

Noto, C., Ota, V.K., Gouvea, E.S., Rizzo, L.B., Spindola, L.M., Honda, P.H., Cordeiro, Q., Belangero, S.I., Bressan, R.A., Gadelha, A., Maes, M., Brietzke, E., 2014. Effects of risperidone on cytokine profile in drug-naïve first-episode psychosis. *Int. J. Neuropsychopharmacol.* 18 (4).

Noto, C., Maes, M., Ota, V.K., Teixeira, A.L., Bressan, R.A., Gadelha, A., Brietzke, E., 2015a. High predictive value of immune-inflammatory biomarkers for schizophrenia diagnosis and association with treatment resistance. *World J Biol Psychiatry* 1–8.

Noto, C., Ota, V.K., Gadelha, A., Noto, M.N., Barbosa, D.S., Bonifacio, K.L., Nunes, S.O., Cordeiro, Q., Belangero, S.I., Bressan, R.A., Maes, M., Brietzke, E., 2015b. Oxidative stress in drug naive first episode psychosis and antioxidant effects of risperidone. *J. Psychiatr. Res.* 68, 210–216.

Noto, C., Ota, V.K., Santoro, M.L., Gouvea, E.S., Silva, P.N., Spindola, L.M., Cordeiro, Q., Bressan, R.A., Gadelha, A., Brietzke, E., Belangero, S.I., Maes, M., 2016. Depression, cytokine, and cytokine by treatment interactions modulate gene expression in antipsychotic naive first episode psychosis. *Mol. Neurobiol.* 53 (8), 5701–5709.

Pornthanakasem, W., Kongruttanachok, N., Phuangphairoj, C., Suvarnsastakorn, C., Sanghathum, T., Oonsiri, S., Poneyam, W., Thanasupawat, T., Matangkasombut, O., Mutirangura, A., 2008. LINE-1 methylation status of endogenous DNA double-strand breaks. *Nucleic Acids Res.* 36 (11), 3667–3675.

Prakash, M.D., Tangalakis, K., Antonipillai, S., Stojanovska, L., Nurgali, K., Apostolopoulos, V., 2017. Methamphetamine: effects on the brain, gut and immune system. *Pharmacol. Res.* 120, 60–67.

Ramkissoon, A., Wells, P.G., 2015. Methamphetamine oxidative stress, neurotoxicity, and functional deficits are modulated by nuclear factor-E2-related factor 2. *Free Radic. Biol. Med.* 89, 358–368.

- Shelly, J., Uhlmann, A., Sinclair, H., Howells, F.M., Sibeko, G., Wilson, D., Stein, D.J., Temmingh, H., 2016. First-rank symptoms in methamphetamine psychosis and schizophrenia. *Psychopathology* 49 (6), 429–435.
- Smith, R.S., Maes, M., 1995. The macrophage-T-lymphocyte theory of schizophrenia: additional evidence. *Med. Hypotheses* 45 (2), 135–141.
- Tangsuwansri, C., Saeliw, T., Thongkorn, S., Chonchaiya, W., Suphapeetiporn, K., Mutirangura, A., Tencomnao, T., Hu, V.W., Sarachana, T., 2018. Investigation of epigenetic regulatory networks associated with autism spectrum disorder (ASD) by integrated global LINE-1 methylation and gene expression profiling analyses. *PLoS One* 13 (7), e0201071.
- Teroganova, N., Girshkin, L., Suter, C.M., Green, M.J., 2016. DNA methylation in peripheral tissue of schizophrenia and bipolar disorder: a systematic review. *BMC Genet.* 17, 27.
- Thompson, P.M., Hayashi, K.M., Simon, S.L., Geaga, J.A., Hong, M.S., Sui, Y., Lee, J.Y., Toga, A.W., Ling, W., London, E.D., 2004. Structural abnormalities in the brains of human subjects who use methamphetamine. *J. Neurosci.* 24 (26), 6028–6036.
- Udomsinprasert, W., Kitkumthorn, N., Mutirangura, A., Chongsrisawat, V., Poovorawan, Y., Honsawek, S., 2016. Global methylation, oxidative stress, and relative telomere length in biliary atresia patients. *Sci. Rep.* 6, 26969.
- Vasan, S., Olango, G.J., 2018. Toxicity, Amphetamine. *StatPearls, Treasure Island (FL)*.
- Wangsri, S., Subbalekha, K., Kitkumthorn, N., Mutirangura, A., 2012. Patterns and possible roles of LINE-1 methylation changes in smoke-exposed epithelia. *PLoS One* 7 (9), e45292.
- Wanichopparat, W., Suwanwongse, K., Pin-On, P., Aporntewan, C., Mutirangura, A., 2013. Genes associated with the cis-regulatory functions of intragenic LINE-1 elements. *BMC Genomics* 14, 205.
- Wongpaiboonwattana, W., Tosukhowong, P., Dissayabutra, T., Mutirangura, A., Boonla, C., 2013. Oxidative stress induces hypomethylation of LINE-1 and hypermethylation of the RUNX3 promoter in a bladder cancer cell line. *Asian Pac. J. Cancer Prev.* 14 (6), 3773–3778.
- World Health Organization, 1992. *The ICD-10 Classification of Mental and Behavioural Disorders: Clinical Description and Diagnostic Guidelines*. World Health Organization, Geneva.
- Yehezkel, S., Segev, Y., Viegas-Pequignot, E., Skorecki, K., Selig, S., 2008. Hypomethylation of subtelomeric regions in ICF syndrome is associated with abnormally short telomeres and enhanced transcription from telomeric regions. *Hum. Mol. Genet.* 17 (18), 2776–2789.
- Zeni-Graiff, M., Rizzo, L.B., Mansur, R.B., Maurya, P.K., Sethi, S., Cunha, G.R., Asevedo, E., Pan, P., Zugman, A., Yamagata, A.S., Higuchi, C., Bressan, R.A., Gadelha, A., Brietzke, E., 2016. Peripheral immuno-inflammatory abnormalities in ultra-high risk of developing psychosis. *Schizophr. Res.* 176 (2–3), 191–195.