



Review

Role of tumor-derived exosomes in cancer metastasis

Ian Wee^{a,b}, Nicholas Syn^{a,b}, Gautam Sethi^{b,***}, Boon Cher Goh^{a,b,c,**}, Lingzhi Wang^{a,b,*}^a Cancer Science Institute of Singapore, National University of Singapore, Singapore^b Department of Pharmacology, Yong Loo Lin School of Medicine, National University of Singapore, 117600, Singapore^c Department of Haematology-Oncology, National University Cancer Institute, Singapore, Singapore

ARTICLE INFO

Keywords:

Tumor-derived exosomes

Cancer metastasis

EMT

Angiogenesis

ABSTRACT

The highlights of cancer research include the discovery of exosomes, which are small (30-100 nm) sized vesicular nanoparticles released virtually by all cells. Tumor-derived exosomes (TDEs) are notoriously known for orchestrating the invasion-metastasis cascade via systemic pathways that we have previously proposed (1), resulting in a paradigm shift of our understanding about the pathobiology of metastases. In principle, exosomes serve as transport medium for proteins, mRNAs and miRNAs to transmit targeted cues from the primary cell to distant sites via horizontal transfer or cell-receptor interaction. In this chapter, we seek to explore in-depth the mechanisms engendering TDE in the metastatic cascade, along with experimental models to augment our understanding. The aforementioned has also paved way for parallel advancements in the therapeutic armamentarium, as evident from pronounced efforts to exploit the metastatic process for therapeutic targeting. In this light, we aim to examine potential anti-metastatic therapeutic opportunities derived from exosomal research. Lastly, exosomes may play a crucial role in the contemporary era of “liquid biopsies”, given the array of molecular information with diagnostic and predictive indications. We thus intend to end this chapter off by exploring future applications of exosomes that could illuminate shortcomings and propel advancements in biomarker research.

1. Introduction

Our understanding of cancerous metastasis has traditionally hinged upon tumor-host interaction. This field of research has garnered renewed interest ever since the discovery of vesicle-based cell-cell communication mediated by exosomes. This concept was first propounded by Trams and colleagues in 1981 [1] as endocytic microvesicles exhibiting 5'-nucleotidase activity, which are released by virtually all cell lines including neoplastic variants. These nanoparticles are distinct in size between 30 and 100 nm, morphologically round or cup-shaped, with a density of between 1.13 and 1.19 g/ml [2]. The inception event to its secretion into the extracellular environs is the fusion of multivesicular endosomes with the cell surface or by reverse budding from the plasma membrane [2,3], giving rise to its distinct protein-lipid composition. Secretion of exosomes is pronounced in cancer cells, best attributed to a constellation of Rab3D overexpression [4]; other signaling cascades such as the Wnt pathway [5]; and a nurturing acidic

microenvironment that further promotes cell fusion events [6].

Whilst exosomes have traditionally been understood to be cellular “trash bags”, their enigmatic nature has warranted persistent research into their structure and functions as molecular Palkis. The highlights of recent research revealed the crucial role that tumor-derived exosomes (TDEs) may have in cancer inception, propagation, and metastasis [7,8]. TDEs are regarded as functional molecular cargos, carrying a myriad of more than 40,000 proteins reported to date, such as tubulin, actin, actin-binding molecules, cytosolic proteins, and those derived from plasma or endosomal membranes [9]. MHC class I [10] and heat shock protein (HSP 70 and 90) [3,9] are also key features present that are involved in antigen presentation. Tetraspanins are characteristically carried at high concentrations in exosomes; CD9 and CD82 in particular inhibit invasion and migration of cancer cells as well as interaction with integrins as reported in both in vitro and in vivo studies respectively [11]. Besides proteins, exosomes carry lipids, mRNA and DNA, all of which are adroit to ignite malignant changes and field cancerization.

* Corresponding author at: Cancer Science Institute of Singapore, National University of Singapore, 14 Medical Drive, #12-01, Centre for Translational Medicine, 117599, Singapore.

** Corresponding author at: Cancer Science Institute of Singapore, National University of Singapore, 14 Medical Drive, #12-01, Centre for Translational Medicine, 117599, Singapore.

*** Corresponding author at: Department of Pharmacology, Yong Loo Lin School of Medicine, National University of Singapore, 117600, Singapore.

E-mail addresses: phcgs@nus.edu.sg (G. Sethi), phgbc@nus.edu.sg (B.C. Goh), csiwl@nus.edu.sg (L. Wang).

<https://doi.org/10.1016/j.bbcan.2018.10.004>

Received 13 September 2018; Received in revised form 22 October 2018; Accepted 22 October 2018

Available online 09 November 2018

0304-419X/ © 2018 Elsevier B.V. All rights reserved.

MiRNA constitutes the major component of RNA in exosomes [12,13]; it exhibits the dysregulated profile of cancer cells and serve as genetic messengers allowing for ubiquitous influence on gene expression of recipient target cells [14]. Lastly, long non-coding RNAs (LnRNAs) form an integral part of the molecular cargo, disrupting gene expression via histone complex modification or annealing with genomic DNA [15]. The actions of these oncogenic molecules potentially reflect the ever-changing pathognomonic pathways that cancer cells have adopted to ensure their own continuance.

It is of belief that TDEs are well involved in the invasion-metastasis cascade [8]. First, TDEs are capable of initiating the preliminary events of metastatic dissemination in the epithelial-mesenchymal transition (EMT) program by secreting autocrine and paracrine signals within the neoplastic environment. Penultimately once disseminated into the systemic circulation, TDEs may reside in distal organ tissues to attain a pre-metastatic niche where they either remain dormant (dormant tumor cells) or colonize to form micro-metastases and eventually macro-metastases [8]. Lastly, TDEs exhibit immune-modulating effects to escape immune surveillance and allow for unquenchable metastatic progression. A summary of the invasion-metastasis cascade is summarized in Fig. 1. In the following sections, we elucidate these mechanisms with contemporary evidence, which could form the basis for novel therapeutic tactics in the bid to outwit cancer.

2. Key players in EMT

The hallmark of the invasion-metastasis cascade is formation of macro-metastases in distant organs, and epithelial to mesenchymal transition (EMT) is a paramount process in the initiation phase. EMT is characterized by trans-differentiation from epithelial neoplastic cells into motile mesenchymal cells under the influence of cancer-associated fibroblasts (CAFs) [16] to create dramatic cell transpositions. The pronounced invasion and migratory properties of epithelial neoplastic cells occurs as a loss of polarized organization and cell-cell junctions [17]. Under EMT, there is diminished expression of E-cadherin as epithelial marker, heightened expression of vimentin as mesenchymal marker [18,19], and proteases such as matrix metalloproteinases (MMP) [20–26]. These enable epithelial cells to emanate cytoplasmic extensions and intrude the basement membrane. These protrusions then form new connections with the interstitial matrix and thereafter cytoskeleton contraction allows for cells to translocate into the stroma.

The locomotion of invading cells depends heavily on repeated bouts of well-coordinated contractile and protrusive events.

A plethora of signaling pathways have been reported as crucial during the EMT program. The process depends on various putative signal transduction cascades that are mediated by SNAIL, zinc-finger E-box binding (ZEB), and TWIST factors [16,20,21,22,26]. Other novel development drivers include GATA family [27,28], SRY, and forkhead box (FOX), which modulate epithelial genes or polarity complexes, and to harmonize with SNAIL/SNAIL 2 to instigate EMT [29,30]. Given the hypoxic state that TDEs tend to shed in, there is increased propensity for TDEs to instigate metastasis due to enriched potency of EMT-transducing signaling molecules such as MMPs, tumor necrosis factor alpha, transforming growth factor beta (TGF β), protein kinase B (AKT), interleukin-6 (IL-6), integrin-linked kinase, caveolin-1, platelet-derived growth factors (PDGFs), and β -catenin [31,32].

Recipient cells that have internalized TDEs undergo proteomic alterations consistent with EMT-like features [8]. Urothelial cells treated with exosomes isolated from muscle-invasive bladder urothelial cells displayed EMT-typical features with conspicuous migratory and invasive characteristics. Chief of these was an increased expression of vimentin, SNAIL and TWIST coupled with downregulation of E-cadherin and β -catenin expression via the TGF- β 1 pathway [33]. These findings corroborated with another in-vivo study, where a nasopharyngeal carcinoma (NPC) cell line was co-cultured with exosomes isolated from a similar cell line, resulting in heightened expression of mesenchymal markers (vimentin and N-cadherin) and downregulation of E-cadherin [34]. Furthermore, functional assays of EBV-negative cells treated with LMP1 exosomes demonstrated EMT phenotypical features including tendency to invade and migrate [35]. These findings were further extrapolated in other neoplastic cell lines including lung, gastric and glioblastoma [36–38]. Pro-oncogenic mi-RNAs transported in exosomes appears to be key players orchestrating these processes. Inhibition of E-cadherin synthesis in lung cancer and melanoma cells is modulated by exosomal miR-23a, which perpetuates the EMT process via TGF- β pathway [39,40]. Other notable modulators of EMT include MiR-191 and let7a, which are found in melanoma, gastric and colorectal cancer [41–43].

Aside from EMT associated pathways, tight and adherens junctions serve as potential targets for TDEs [8]. They may exert direct influence on the invasive process into blood vessels by secreting miR-105. As manifested in breast cancer models, miR-105 has been shown to

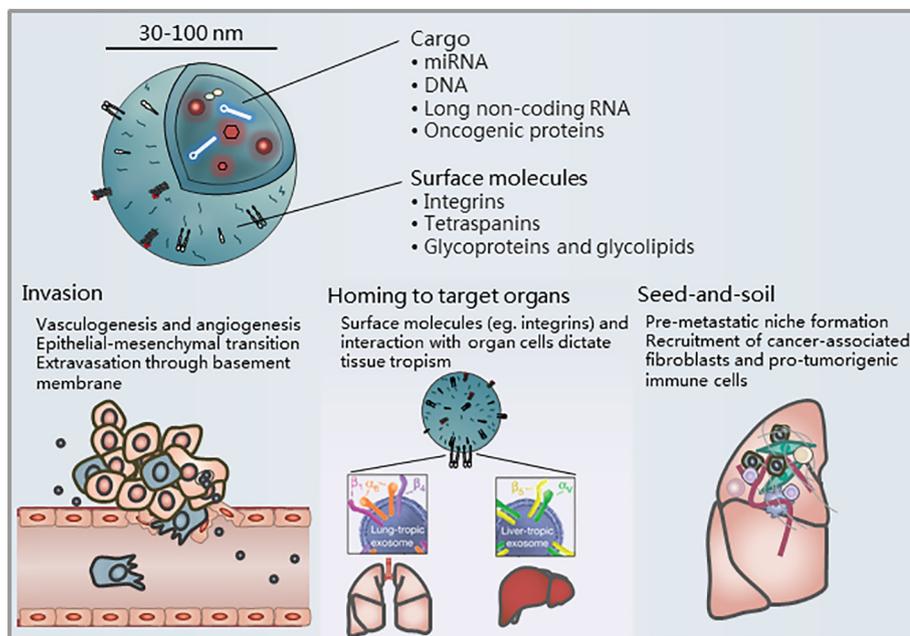


Fig. 1. – Contents of tumor derived exosomes (TDE) and invasion-metastasis cascade. TDEs are molecular cargos containing miRNA, DNA, long non-coding RNA, and oncogenic proteins. It is also bounded by surface molecules including integrins, tetraspanins, glycoproteins and glycolipids. TDEs are primarily responsible for initiating the epithelial-mesenchymal transition. Invasion first occurs through vasculogenesis and angiogenesis, and once disseminated into the systemic circulation, TDEs may reside in distal organ tissues to attain a pre-metastatic niche where they colonize to form micro-metastases and eventually macro-metastases. Immune-modulating effects occur due to recruitment of cancer-associated fibroblasts and pro-tumorigenic immune cells, allowing TDEs to escape immune surveillance.

interact with and downregulate the tight junction protein ZO-1 in endothelial monolayer cells, magnifying vascular permeability and accelerating formation of lung and brain metastases [14]. Implicitly, exosomal miR-105 serves as a prognostic marker for patients with breast carcinoma and serum levels reflect the level in the primary tumor itself [14]. However, it appears that exosomes secreted by non-neoplastic cells may have a role in driving cancer progression. In a salient study by Wrana and colleagues, fibroblast-secreted exosomes were reported to enhance the invasive activity and motility of breast cancer cell (BCC) via Wnt-planar cell polarity (PCP) signaling. Upon internalization of fibroblast-derived exosomes, BCCs repackaged the exosome cargo with Wnt11 and initiated activation of Wnt-PCP signaling in an autocrine fashion. Furthermore, co-culture of both BCC and fibroblasts promoted metastasis that was driven by both fibroblastic exosome tetraspanins CD81, and PCP signaling in BCC [44]. These fascinating findings provide mechanistic insights into the crosstalk between stromal cells and cancer cells to instigate metastasis.

Vice versa, TDEs have been shown to modulate stromal fibroblasts. In a comprehensive study by Webber and colleagues [45], they showed that TGF β positive TDE were capable of elevating α -smooth muscle actin expression, converting mesenchymal stem cells into cancer-associated fibroblasts (CAFs). Just recently, the same group demonstrated that TDEs secreted by prostate cancer cells activated TGF β 1 dependent fibroblast differentiation to a unique myofibroblast phenotype, propagating tumor growth in vivo and reinforcing angiogenesis [46]. The aforementioned processes are specific to TDEs, as myofibroblasts induced using soluble TGF β 1 were not oncogenic. This exemplifies the indispensable role of cancer exosomes as key players in tumor-associated stromal differentiation.

3. Angiogenesis

Angiogenesis is a requisite process for tumor growth and metastasis [47]. Particularly in hypoxic conditions, exosomes function as key messengers between neoplastic and vascular cells during hypoxia-driven pro-angiogenic tumor responses [32]. Furthermore, TDEs secreted from melanoma cells can incite a proangiogenic signaling cascade characterized by endothelial spheroid formation in a dose-dependent fashion [48]. Elevated expression of Wnt5a in melanoma has also been correlated with the angiogenic marker ESAM during gene analysis, affecting extent of endothelial cell branching [5]. Just recently, a novel study by Asada and colleagues [49] reported that Delta-like 4 (DII4) containing exosomes resulted in loss of filopodia in capillary endothelial tip cells and blunted sprout formation. These findings suggest that DII4 containing exosomes elevate endothelial cell motility whilst suppressing their growth. Evidence is accumulating supporting the role of exosomal miRNA in regulating angiogenesis and its consequent impacts on tumor progression [50]. Grange and colleagues, for instance, reported that proangiogenic mRNAs and miRNAs released by CD105-positive renal cancer stem cells potentiated lung metastases [51]. In another instance, miR-214 – which controls endothelial cell function and angiogenesis – is responsible for exosome-mediated signaling between endothelial cells, resulting in ataxia telangiectasia mutated silencing in recipient cells [52].

4. Organotropic metastasis

The penchant of primary tumors to disseminate to specific secondary sites is understood to be a result of intricate, well-coordinate tumor-stroma interaction in the host organ. This formed the crux of the “soil and seed” hypothesis that was first theorized by Paget in 1889, where primary tumor cells tended to form metastasis only at compatible secondary organ sites [53]. Fidler and colleagues later bolstered these findings by showing that metastasis was site-specific regardless of the vasculature or lymphatic vessels that drain the primary tumor site [54]. This set forth the concept that a conducive microenvironment, or niche,

is paramount for circulating tumor cells to engraft at distant organ sites. Bone marrow derived cells, endothelial progenitor cells and mesenchymal cells play key roles in establishing the pre-metastatic niche [55]. Robust evidence to date comes from melanoma-derived exosomes, which are capable of “educating” bone marrow progenitors – through the receptor tyrosine kinase MET – to become pro-vasculogenic [56]. This is perpetrated by enhancement of vascular leakiness and upregulation of pro-inflammatory molecules at metastatic sites [56].

As astutely described by Rak as “organ-seeking vesicles” [57], there is mounting evidence accumulated across cancer types suggesting that the biodistribution of exosomes commensurate with pattern of organotropic metastatic spread, including breast, melanomas, and pancreatic carcinomas [8,48,58,59]. Mouse model studies for pancreatic ductal adenocarcinoma (PDAC) demonstrated that TDEs are important in promoting engraftment of metastatic pancreatic cells in the liver by interaction with Kupffer-cells [60]. Through migration inhibitory factor (MIF) signaling, hepatic stellate cells were found to upregulate fibronectin production and TGF β secretion upon internalization of PDAC-derived exosomes [60]. Similar observations were made in malignant melanomas, where exosomes had greater propensity to harbor in the interstitial space of the kidney, liver, bone marrows and lungs, all of which are organotropic sites for melanoma metastasis. This consequently leads to an upregulation of genes like S100A8 and S100A9 involved in inflammation and extracellular matrix remodeling (ECM); annexins A6 and CD44v6 potentiating metastasis; and HSPs (HSP 90 and HSP 70) [61–63].

The integrin repertoire is thought to be – at least in part – responsible for the biased distribution of exosomes. For instance, Integrin beta 5 enhances exosome adhesion in the liver whilst integrin beta 4 directs metastasis to the lungs [59]. However, it is probably not the only means to the end – the uptake of TDEs into recipient cells could trigger a myriad of other endocytic processes including lipid raft, clathrin, and caveolin-mediated internalization [64–67].

5. Homing of neoplastic cells to their target tissue

After “seeding” has taken place in the new foreign soil, exosomes are able to foment tumor-supportive processes including proliferation, angiogenesis, and ECM remodeling. Uptake of TDEs by myofibroblast progenitors can enhance recruitment [64,68] and augment differentiation into myofibroblast-like cells via TGF β /Smad signaling [45,46,69]. Intriguingly, it appears that the homing process occurs in a bidirectional fashion, where exosomes secreted by non-neoplastic cells may also promote adaptation of circulating tumor cells. The remarkable plasticity of the tumor microenvironment was further demonstrated by Zhang and colleagues, who reported a reduction of PTEN expression after metastasis to the brain [70]. This was epigenetically mediated by microRNAs from brain astrocytes, which release exosomes to orchestrate an intercellular transfer of PTEN-specific microRNAs to metastatic tumor cells. The consequent loss of PTEN expression resulted in increased secretion of chemokine CCL2, consequently recruiting IBA1-expressing myeloid cells that promoted the development of brain metastatic tumor cells [70].

6. Oncogenic transformation of non-neoplastic cells

Non-neoplastic cells are at risk of undergoing oncogenic transformation in the presence of TDEs. For instance, an in vivo study involving patient adipose-derived stem cells (pASCs) acquired cytogenetic aberrations and mesenchymal-to-epithelial (MET) transition when primed with prostate cancer cells. Aside from the downregulation of large tumor suppressor homolog2 and the programmed cell death protein 4, the oncogenic transformation potential was attributed to a host of exosomes carrying oncogenic factors, such as miR-125B, miR-130b, miR-155, H-ras, K-ras, Rab1a, Rab1b and Rab11a [71]. This phenomenon is also evident in breast cancer cells, where human mammary

epithelial MCF10A cells underwent neoplastic changes when co-cultured with breast cancer MDA-MB-231 derived exosomes. This was further characterized by downregulation of target transcripts PTEN and homeobox protein (HOXD1) and reciprocal upregulation of miR-21 and miR-10b [72].

Oncogenic viruses are also capable of conferring pro-oncogenic signaling in recipient cells by exploiting the host exosomal apparatus [66,70,73,74]. Epstein-Barr virus positive nasopharyngeal cancer cells (EBV positive NPC) secrete exosomes containing EBV oncoproteins LMP1 and viral miRNAs, which activates AKT, extracellular signal-regulated kinases (ERK), and epidermal growth factor receptor (EGFR) expression in normal epithelial cells [35,73,74]. However, this field is still in its relative infancy, and much remains to be explored regarding mechanisms exploited by viral cancers to amplify their progression.

7. Immune modulation

Immunosurveillance is perhaps the host's main defense mechanism against proliferation of circulating clones; unfortunately, metastasis remains a common phenomenon reflecting the success of neoplastic cells in evading the immune system. We seek to uncover the various mechanisms in which cancer cells have employed to ensure their longevity.

Since some cancers spread via hematogenous routes, it is crucial for these cells to be able to survive in the blood circulation and not succumb to immune surveillance. In this regard, platelets appear to be a key player ever since the first link between platelets and cancer was inaugurated by Trousseau in 1865 [75]. Exosomes secreted by platelets consist of P-selectin and GP IIb-IIIa, which interact with cancer cells, endothelial cells and leukocytes [76,77]. The relationship between platelets and cancer is akin to deadly allies; platelets appear to be chaperons to cancer cells, accompanying them in the blood stream and facilitating adhesion at the vessel endothelium via P-selectin, resulting in extravasation of neoplastic cells into the premetastatic niche [78–80]. Mice model studies reinstated the crucial role of platelets, where platelet depletion inhibited metastasis whilst recovery of these platelets was associated with metastatic activity [81]. Besides vessel adhesion, platelets function as “guardians” of these neoplastic cells against immune cell activity, such as natural killer (NK) cell mediated antibody-dependent cytotoxicity [82,83]. Coating cancer cells with fibrin appears to be the key mechanism responsible for repudiating NK cell activity due to impaired recognition of coated cells [79]. A second plausible mechanism hinges on the secretion of TGFβ1 by hypoxic tumor cells, blunting expression of NK cell activating receptor (NKG2D) [84,85]. This further activates the EMT-associated TGFβ1 pathway, which is a key component of the metastatic cascade [86].

Upon arrival the secondary organ site, TDEs activate immunosuppressive pathways to combat the foreign microenvironment in order to ensure tumor cell proliferation. This is achieved by inhibiting effector cells and stimulation of regulatory T cells [87]. An in vitro culture model study of acute myeloid leukemia showcased the reduction of CD8+ T cells by TDEs via activation of Fas/Fas ligand mediated apoptosis [88,89]. This was coupled with proliferation and conversion of CD4+ T cells into regulatory T cells (CD4+, CD25+, FOXP3+) characterized by expression of interleukin-10, TGFβ1, cytotoxic lymphocyte antigen – 4 (CTLA-4) and granzyme, all of which curtail the cytotoxic effect of NK cells [90,91]. Furthermore, TDEs were shown to activate human myeloid derived suppressor cells – immature myeloid cells that suppress T cell activation – through HSP72/TLR-2 via IL-6 expression and STAT3 signaling pathway [92,93]. The immunosuppressive effects of TDEs could possibly stem from miRNAs as evident from an in vitro model study of NPC, where immunosuppressive miRNAs (hsa-miR-24-3p, hsa-miR-891a, hsa-miR-106a-5p, hsa-miR-20a-5p, and hsa-miR-1908) were present in TDEs [94,95]. TDEs were also capable of converting CD4+ CD25- T cells into inhibitory CD4+ CD25^{high} T cells [94]. One of the many proposed mechanisms of

T cell suppression include extracellular adenosine production by TDEs positive for CD39 and CD73 [96], which may occur due to epigenetic changes conferred by TDEs to T cells. For instance, TDEs can down-regulate regulatory T cell genes modulating the adenosine pathway leading to pronounced expression of CD39 and adenosine production [97]. This portends the possibility that TDEs exuberate a myriad of immunosuppressive mechanisms to ensure neoplastic survival.

Besides modulating cellular immune responses, the humoral response – specifically antibody-dependent cytotoxicity (ADCC) [98] – falls victim to TDEs as well, which in fact is the major of the three response mechanisms including complement-mediated cytotoxicity. For instance, in breast cancer cells, TDEs can express human epidermal growth factor receptor 2 (Her2) and epithelial cell adhesion molecule (EPCAM), therefore binding and sequestering antibodies exerting ADCC on neoplastic cells positive for these antigens. Hence, therapeutic response against Trastuzumab (mammalian Her2-antibody) is diminished [99]. A similar observation was made in B cell lymphomas, where exosomes carrying CD20 intercepted the anti-CD20 antibody (Rituximab), diminishing the membrane attack complex mediated by binding of Rituximab at the B cell surface. Paradoxically, administration of Rituximab increased secretion of CD20 positive exosomes whereas depletion of these exosomes improved therapeutic responses of Rituximab [100]. These findings hint at the impending need for future research to investigate potential immunotherapy pitfalls associated with TDEs [87].

Inflammation is a paramount process necessary for tumor growth and adaption to the “foreign soil” [55]. Tumor-associated macrophages (TAM) promote invasion, ECM remodeling and angiogenesis [87,101]. Histological evidence of macrophages in high density has been correlated with poorer prognosis in breast cancer specimens [102]. It has been established now that TDEs activate TAMs via nuclear factor-κB and interaction with toll-like receptors (TLRs) found on the macrophage cell surface [103]. Alternatively, this could be achieved by binding of exosomal miRNAs (mi-R21, mi-R29a) as ligands to receptors of TLRs (TLR-7, TLR-8) in a paracrine fashion [104], resulting in a pro-metastatic inflammatory response involving cytokines TNFα and IL-6. The relationship between TDEs and inflammation is also evident in pancreatic ductal adenocarcinoma, where the uptake of TDEs by liver Kupffer cells lead to heightened fibronectin production by hepatic stellate cells. This in part is mediated by macrophage migratory inhibition factor (MIF) potentiating the release of TGFβ1 by Kupffer cells [60]. Deposition of fibronectin in the liver subsequently fosters arrest of bone marrow-derived macrophages hence creating a pre-metastatic niche.

8. Pharmacological developments

Alluding to the aforementioned discussions, it is now discernible that tumor derived exosomes exert a spectrum of oncogenic mechanisms that instigate tumor growth, progression and metastasis. Every stage along this multi-level process are potential therapeutic targets to be exploited. It is the aim of the next few paragraphs to comprehensively review the development of the therapeutic armamentarium that could target these pathways.

The biogenesis of exosomes constitutes several components, some of which are regarded as fundamental regulators of metastasis. Heparanase, for instance, has been identified as one potential therapeutic target. The role of heparanase in exosome biogenesis remains to be elucidated, but it functions primarily as an endoglucuronidase that cleaves heparan sulfate chains on syndecan-1 propagating formation of syndecan-syntenin-ALIX complex [105]. Syndecan-syntenin-ALIX plays a key role in membrane transport and signaling processes, hence altering exosome composition; promoting exosome production; and driving exosome secretion [106,107]. In this light, investigations in animal models are now underway to explore the potential of heparanase inhibitors, heparin mimetics and modified heparins as anticancer

therapeutics [105].

Other potential targets include RAB genes, which are highly expressed in metastatic tumors and are responsible for secreting innumerable amounts of exosomes [56]. Blockade of GTPase RAB27a, for example, has been demonstrated in mouse models to downregulate exosomal secretion and metastatic dissemination, hence suggesting a role for RAB27a in exosome-dependent mechanisms that promote oncogenic progression and modify the tumor microenvironment [56,108]. In the same vein, a synergistic interaction has been discovered between invadopodia biogenesis and exosome secretion. The invasive actin structural nature of invadopodia allows for docking and secretion of CD63- and RAB27a positive exosomes [109]. Hence targeting of key proteins in these structures (N-WASP, TKS4, TKS5, MT1MMP) could spur therapeutic strategies aimed at bridling exosome biogenesis and secretion [110]. One other key target that could be exploited to inhibit TDE release is neutral sphingomyelinase 2 (nSMase2), a key regulator of ceramide biosynthesis. *In vitro* inhibition of nSMase2 with GW4869 has been shown to inhibit TDE release, hence suggesting that ceramide plays a weighty role in the dissemination of exosomes [111].

A complementary strategy to inhibiting exosome release is blockage of exosome internalization pathways [8]. Through competitive inhibition, persistent treatment with heparin and heparan sulfate chains likewise are capable of inhibiting TDE uptake and consequent oncogenic progression in oral squamous cell carcinoma and glioblastoma [33,37,112], hence lending further support to the pharmacological development of heparanase-targeting agents.

In addition, the functionality of TDEs are amenable to alterations, as evident from dietary studies of polyphenol curcumin. Curcumin was shown to reverse the TDE-mediated suppression of NK cell lysis in breast cancer models [113]. In chronic myelogenous leukemia (CML), the addition of curcumin as an adjunct resulted in a dose-dependent increase of PTEN, a target of miR-21. A consequent reduction of miR-21 in CML cells and exosomal packaging was observed. This was accompanied by downregulation of VEGF expression and release, as well as AKT phosphorylation [114].

A novel therapeutic paradigm involving the use of exosomes as immunologic stimulants has also been propounded [115]. Exosomes derived from antigen-presenting cells (APCs) are functional cargos of MHC/tumor antigens complexes, which can subsequently be presented to dendritic cells *in vivo* to prime cytotoxic T cells for T cell-dependent response against tumor cells. Hence, exosome-based cell-free vaccines may be an appealing anti-cancer therapeutic alternative [115,116]. However, there are shortcomings that might impede therapeutic advancement in this field. First and foremost, APCs are technically difficult to culture along with the need for specific tumor antigens to be transferred to APCs. Secondly, such an approach cannot be generalized to all patients as MHC-1 haplotype matching is warranted given the MHC-1 dependent antitumor response [117]. In addition, APC-exosomes pale in comparison to TDEs as the latter is able to carry shared tumor antigens hence conferring protection against different cancer types. TDEs also abolish the need for MHC-1 haplotype matching since they exert antitumor immunogenicity without expressing MHC-1 molecules [118]. Human Mucin 1 (hMUC1), for example, is a shared tumor antigen which can be incorporated into exosomes to be employed as anti-cancer vaccination against MUC1-expressing cancers [119]. In another instance, *in vitro* and *in vivo* murine studies reported that HSP70-enriched TDEs released by heat-treated tumors potentiated stronger Th1 immune response in comparison to normal TDEs [117]. This was accompanied by direct activation of NK cell mediated apoptosis of tumor cells [120]. Dendritic cells too, have shown to be primed by exosomes isolated from ascites in cancer patients, resulting in T cell mediated antitumor response [121]. Considerable progress has been made, as success from these experimental models have now translated into phase I clinical trials, two of which have demonstrated encouraging results in patients with non-small cell lung cancer [122] and late-stage malignant melanoma [123].

The lipid bilayer membrane-encapsulated structure of exosomes prohibits degradation and RNase damage, potentially serving as delivery systems for traditional chemotherapeutic agents, which reduces chemotherapy associated toxicity [115,124,125]. Intriguingly, cytotoxicity increased more than 50-fold in multi-drug resistant cancer upon amalgamation of paclitaxel with exosomes [126]. In another instance, the exosome-based delivery of certain miRNAs into lung cancer cells can increase the sensitivity to taxanes. This has been shown in the delivery of miR-181a into lung adenocarcinoma cells to increase its sensitivity to paclitaxel and platinum-based chemotherapy [127]. Aside from its unique structure, the small size of exosomes allows for easy passage through different biological barriers, such as the blood-brain barrier [128]. Therefore, exosomes may have the potential to bolster classical chemotherapeutic regimens, by improving its efficacy and safety profile [115].

The use of exosomes in vaccine development has heightened interest as well. Together with dendritic cell-derived exosomes (Dex), TDEs can migrate to lymph nodes to activate CD4⁺ and CD8⁺ T cells to initiate immunological reactions [129]. In a Phase IIB trial, the administration of TG4010 vaccine enhanced the effect of chemotherapy in patients with advanced non-small cell lung cancer (NSCLC), with a superior 6-month progression free survival (PFS) of 43.2% compared to just 35.1% in the chemotherapy only group [130]. In another Phase II trial, the generation of Dex increased the PFS of NSCLC patients with defective expression of Nkp30 [129].

However, as discussed in Section 7, exosome-mediated immunosuppression may be the Achilles' heel of pharmaceutical developments. In this regard, the use of TDEs as a therapeutic tool may serve as a double-edged sword. Strategies are still needed to overcome TDE-mediated apoptosis of CD8⁺ T cells, as well as inhibition of effector T cell proliferation. Aside, TDEs may alter the immunologic receptors and ligands of receptor cells similar to that of oncoytes, disrupt the immune function of these cells, or even sequester therapeutic antibodies. This has been shown in a TDE-mediated transfer of epidermal growth factor receptor to macrophages, resulting in compromised innate immunity [131,132].

9. Biomarkers

It is understood that the blood concentration of exosomes in cancer patients is twice that of a normal health adult, as cancer cells have a higher propensity to generate more exosomes [13,133,134]. Unfortunately, technical limitations related to isolation cost and technique have impeded adoption of these biomarkers in clinical practice. This warrants the need to develop a robust method capable of minimize isolating protein aggregates and membranous particles.

Within the cargo that exosomes carry, miRNA serves as a useful source for predictive biomarkers, given that tumors tend to alter their miRNA expression profiles making it distinctive from normal cells [13,135]. Biomarkers have been studied heavily in prostatic carcinoma patients, where exosomal miR-141 and miR-375 were present in superior concentrations after radical prostatectomy [136–138]. Furthermore, varying levels of miRNAs may also help to distinguish high grade from low grade tumors, allowing for early risk identification amongst patients [138].

Similar observations were made with exosomal protein markers [13]. The most robust evidence to date comes from pancreatic patients, where Glypican-1 carrying exosomes were detected with 100% sensitivity and 100% specificity in comparison to health donors or patients with benign pancreatic lesions. Furthermore, the concentration of Glypican-1 correlated with tumor load hence facilitating prompt diagnosis of relapse post-surgery or metastatic spread. Whilst impressive, the study's sample size ($n = 31$ in total) warrants the need for larger studies to establish these conclusions, particularly for early detection of pancreatic cancer (pancreatic cancer precursor lesions, PCPL) in which only 5 cases were included in the discovery cohort but none in the

validation cohort. Aside from pancreatic cancer, the same study also showed elevated Glypican-1 carrying exosomes in patients with breast cancer (24/32, 75%) compared to healthy donors. Hence, the 100% specificity of Glypican-1 as a pancreatic diagnostic biomarker could be greatly compromised [134]. Exosomal transmembrane256 has also been implicated in patients with prostate cancer, with relative success in terms of sensitivity and specificity [139]. Evidence for other cancer types including gastric and colorectal cancer [140] are also emerging in support of exosomes functioning as novel diagnostic biomarkers [141]. In the surge of the “liquid biopsy” era, exosomes may prove to be promising biomarkers for diagnostic, prognostic and predictive purposes in the near future [142].

10. Conclusion and future directions

Whilst TDEs hold significant potential in offering advanced therapeutic options, it may serve as a double-edged sword. On the one hand, TDEs hold strong potential for pharmaceutical development, be it as a vector or biological target. On the other hand, they are key players in the oncogenic-metastatic cascade, further impeding pharmaceutical developments through immune-modulation rendering these therapies ineffective. Nevertheless, several questions remain unanswered. The integration of exosomes and CRISPR-Cas genome-editing systems, for instance, is an unexplored area for future research and advancements in precision oncology. Similarly, can exosomes be integrated and loaded with multiple tumoricidal compounds? Next, it is unclear whether the route of administration (subcutaneous, intravenous etc), affects the therapeutic efficacy of exosome-based treatments.

From a legislative standpoint, there is yet to be an established TDE-specific guideline for therapeutic development. Nonetheless, it is expected that aspects of regulatory and safety requirements for pharmaceutical development and clinical application will be derived from current legislature on cell-based guidelines [129,143]. In the same vein, logistical issues regarding processing and manufacturing of exosomes may pose a challenge given the large amount exosomes needed to overcome the poor therapeutic efficacy of nanoparticle administration [144]. Ultimately, in the bid to offer patients with newer, advanced, and more effective treatment modalities, a multidisciplinary team effort consisting of scientists, clinicians, and engineers, is first needed to collectively address these challenges.

Acknowledgements

The study was sponsored by the National Research Foundation of Singapore (Experimental Therapeutics Program) and the National Medical Research Council of Singapore (NMRC-CSA/SI/0006/2016).

Conflicts of interest

The authors declare no conflict of interest.

References

- [1] E.G. Trams, C.J. Lauter, N. Salem Jr., U. Heine, Exfoliation of membrane ectoenzymes in the form of micro-vesicles, *Biochim. Biophys. Acta* 645 (1981) 63–70.
- [2] M. Simons, G. Raposo, Exosomes—vesicular carriers for intercellular communication, *Curr. Opin. Cell Biol.* 21 (2009) 575–581.
- [3] C. Thery, L. Zitvogel, S. Amigorena, Exosomes: composition, biogenesis and function, *Nat. Rev. Immunol.* 2 (2002) 569–579.
- [4] J. Yang, W. Liu, X. Lu, et al., High expression of small GTPase Rab3D promotes cancer progression and metastasis, *Oncotarget* 6 (2015) 11125–11138.
- [5] E.J. Ekstrom, C. Bergenfelz, V. von Bulow, et al., WNT5A induces release of exosomes containing pro-angiogenic and immunosuppressive factors from malignant melanoma cells, *Mol. Cancer* 13 (2014) 88.
- [6] I. Parolini, C. Federici, C. Raggi, et al., Microenvironmental pH is a key factor for exosome traffic in tumor cells, *J. Biol. Chem.* 284 (2009) 34211–34222.
- [7] A. Subramanian, V. Gupta, S. Sarkar, et al., Exosomes in carcinogenesis: molecular palkis carry signals for the regulation of cancer progression and metastasis, *J. Cell Commun. Signal* 10 (2016) 241–249.
- [8] N. Syn, L. Wang, G. Sethi, et al., Exosome-mediated metastasis: from epithelial-mesenchymal transition to escape from immunosurveillance, *Trends Pharmacol. Sci.* 37 (2016) 606–617.
- [9] C. Thery, M. Boussac, P. Veron, et al., Proteomic analysis of dendritic cell-derived exosomes: a secreted subcellular compartment distinct from apoptotic vesicles, *J. Immunol.* 166 (2001) 7309–7318.
- [10] N. Blanchard, D. Lankar, F. Faure, et al., TCR activation of human T cells induces the production of exosomes bearing the TCR/CD3/zeta complex, *J. Immunol.* 168 (2002) 3235–3241.
- [11] M. Zoller, Tetraspanins: push and pull in suppressing and promoting metastasis, *Nat. Rev. Cancer* 9 (2009) 40–55.
- [12] H. Valadi, K. Ekstrom, A. Bossios, et al., Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells, *Nat. Cell Biol.* 9 (2007) 654–659.
- [13] N.L. Syn, W.P. Yong, B.C. Goh, S.C. Lee, Evolving landscape of tumor molecular profiling for personalized cancer therapy: a comprehensive review, *Expert Opin. Drug Metab. Toxicol.* 12 (2016) 911–922.
- [14] W. Zhou, M.Y. Fong, Y. Min, et al., Cancer-secreted miR-105 destroys vascular endothelial barriers to promote metastasis, *Cancer Cell* 25 (2014) 501–515.
- [15] J.E. Wilusz, H. Sunwoo, D.L. Spector, Long noncoding RNAs: functional surprises from the RNA world, *Genes Dev.* 23 (2009) 1494–1504.
- [16] S. Lamouille, J. Xu, R. Derynck, Molecular mechanisms of epithelial-mesenchymal transition, *Nat. Rev. Mol. Cell Biol.* 15 (2014) 178–196.
- [17] M. Diepenbruck, G. Christofori, Epithelial-mesenchymal transition (EMT) and metastasis: yes, no, maybe? *Curr. Opin. Cell Biol.* 43 (2016) 7–13.
- [18] J. Dudas, Supportive and repressive functions of tumor stroma on tumor cell growth, survival, and invasivity: the cancer evolution, *Front. Oncol.* 5 (2015) 44.
- [19] T.B. Steinbichler, V. Metzler, C. Pritz, et al., Tumor-associated fibroblast-conditioned medium induces CDDP resistance in HNSCC cells, *Oncotarget* 7 (2016) 2508–2518.
- [20] J.P. Thiery, Epithelial-mesenchymal transitions in tumour progression, *Nat. Rev. Cancer* 2 (2002) 442–454.
- [21] J.P. Thiery, J.P. Sleeman, Complex networks orchestrate epithelial-mesenchymal transitions, *Nat. Rev. Mol. Cell Biol.* 7 (2006) 131–142.
- [22] E.D. Hay, An overview of epithelial-mesenchymal transformation, *Acta Anat. (Basel)* 154 (1995) 8–20.
- [23] H. Hugo, M.L. Ackland, T. Blick, et al., Epithelial-mesenchymal and mesenchymal-epithelial transitions in carcinoma progression, *J. Cell. Physiol.* 213 (2007) 374–383.
- [24] R. Kalluri, R.A. Weinberg, The basics of epithelial-mesenchymal transition, *J. Clin. Invest.* 119 (2009) 1420–1428.
- [25] M.A. Nieto, The ins and outs of the epithelial to mesenchymal transition in health and disease, *Annu. Rev. Cell Dev. Biol.* 27 (2011) 347–376.
- [26] M. Guarino, Epithelial-mesenchymal transition and tumour invasion, *Int. J. Biochem. Cell Biol.* 39 (2007) 2153–2160.
- [27] E.H. Bresnack, H.Y. Lee, T. Fujiwara, et al., GATA switches as developmental drivers, *J. Biol. Chem.* 285 (2010) 31087–31093.
- [28] K. Campbell, G. Whissell, X. Franch-Marro, et al., Specific GATA factors act as conserved inducers of an endodermal-EMT, *Dev. Cell* 21 (2011) 1051–1061.
- [29] H. Kondoh, Y. Kamachi, SOX-partner code for cell specification: regulatory target selection and underlying molecular mechanisms, *Int. J. Biochem. Cell Biol.* 42 (2010) 391–399.
- [30] G. Moreno-Bueno, F. Portillo, A. Cano, Transcriptional regulation of cell polarity in EMT and cancer, *Oncogene* 27 (2008) 6958–6969.
- [31] A. Ramteke, H. Ting, C. Agarwal, et al., Exosomes secreted under hypoxia enhance invasiveness and stemness of prostate cancer cells by targeting adherens junction molecules, *Mol. Carcinog.* 54 (2015) 554–565.
- [32] P. Kucharczyk, H.C. Christianson, J.E. Welch, et al., Exosomes reflect the hypoxic status of glioma cells and mediate hypoxia-dependent activation of vascular cells during tumor development, *Proc. Natl. Acad. Sci. U. S. A.* 110 (2013) 7312–7317.
- [33] C.A. Franzen, R.H. Blackwell, V. Todorovic, et al., Urothelial cells undergo epithelial-to-mesenchymal transition after exposure to muscle invasive bladder cancer exosomes, *Oncogene* 4 (2015) e163.
- [34] Y. You, Y. Shan, J. Chen, et al., Matrix metalloproteinase 13-containing exosomes promote nasopharyngeal carcinoma metastasis, *Cancer Sci.* 106 (2015) 1669–1677.
- [35] M. Aga, G.L. Bentz, S. Raffa, et al., Exosomal HIF1alpha supports invasive potential of nasopharyngeal carcinoma-associated LMP1-positive exosomes, *Oncogene* 33 (2014) 4613–4622.
- [36] M.A. Rahman, J.F. Barger, F. Lovat, et al., Lung cancer exosomes as drivers of epithelial mesenchymal transition, *Oncotarget* 7 (2016) 54852–54866.
- [37] H.C. Christianson, K.J. Svensson, T.H. van Kuppevelt, et al., Cancer cell exosomes depend on cell-surface heparan sulfate proteoglycans for their internalization and functional activity, *Proc. Natl. Acad. Sci. U. S. A.* 110 (2013) 17380–17385.
- [38] D. Liu, C. Li, B. Trojanowicz, et al., CD97 promotion of gastric carcinoma lymphatic metastasis is exosome dependent, *Gastric Cancer* 19 (2016) 754–766.
- [39] M. Cao, M. Seike, C. Soeno, et al., MiR-23a regulates TGF-beta-induced epithelial-mesenchymal transition by targeting E-cadherin in lung cancer cells, *Int. J. Oncol.* 41 (2012) 869–875.
- [40] J. Kim, T.Y. Kim, M.S. Lee, et al., Exosome cargo reflects TGF-beta1-mediated epithelial-to-mesenchymal transition (EMT) status in A549 human lung adenocarcinoma cells, *Biochem. Biophys. Res. Commun.* 478 (2016) 643–648.
- [41] K. Ohshima, K. Inoue, A. Fujiwara, et al., Let-7 microRNA family is selectively secreted into the extracellular environment via exosomes in a metastatic gastric cancer cell line, *PLoS One* 5 (2010) e13247.

- [42] S. Tanaka, M. Hosokawa, K. Ueda, S. Iwakawa, Effects of decitabine on invasion and exosomal expression of miR-200c and miR-141 in oxaliplatin-resistant colorectal cancer cells, *Biol. Pharm. Bull.* 38 (2015) 1272–1279.
- [43] D. Xiao, S. Barry, D. Kmetz, et al., Melanoma cell-derived exosomes promote epithelial-mesenchymal transition in primary melanocytes through paracrine/autocrine signaling in the tumor microenvironment, *Cancer Lett.* 376 (2016) 318–327.
- [44] V. Luga, L. Zhang, A.M. Vilorio-Petit, et al., Exosomes mediate stromal mobilization of autocrine Wnt-PCP signaling in breast cancer cell migration, *Cell* 151 (2012) 1542–1556.
- [45] J. Webber, R. Steadman, M.D. Mason, et al., Cancer exosomes trigger fibroblast to myofibroblast differentiation, *Cancer Res.* 70 (2010) 9621–9630.
- [46] J.P. Webber, L.K. Spary, A.J. Sanders, et al., Differentiation of tumour-promoting stromal myofibroblasts by cancer exosomes, *Oncogene* 34 (2015) 290–302.
- [47] S.M. Weis, D.A. Cheresh, Tumor angiogenesis: molecular pathways and therapeutic targets, *Nat. Med.* 17 (2011) 1359–1370.
- [48] J.L. Hood, R.S. San, S.A. Wickline, Exosomes released by melanoma cells prepare sentinel lymph nodes for tumor metastasis, *Cancer Res.* 71 (2011) 3792–3801.
- [49] S. Sharghi-Namini, E. Tan, L.L. Ong, et al., Dll4-containing exosomes induce capillary sprout retraction in a 3D microenvironment, *Sci. Rep.* 4 (2014) 4031.
- [50] G. Bergers, L.E. Benjamin, Tumorigenesis and the angiogenic switch, *Nat. Rev. Cancer* 3 (2003) 401–410.
- [51] C. Grange, M. Tapparo, F. Collino, et al., Microvesicles released from human renal cancer stem cells stimulate angiogenesis and formation of lung premetastatic niche, *Cancer Res.* 71 (2011) 5346–5356.
- [52] B.W. van Balkom, O.G. de Jong, M. Smits, et al., Endothelial cells require miR-214 to secrete exosomes that suppress senescence and induce angiogenesis in human and mouse endothelial cells, *Blood* 121 (2013) 3997–4006 (s3991–3915).
- [53] S. Paget, The distribution of secondary growths in cancer of the breast. 1889, *Cancer Metastasis Rev.* 8 (1989) 98–101.
- [54] I.J. Fidler, Selection of successive tumour lines for metastasis, *Nat. New Biol.* 242 (1973) 148–149.
- [55] B. Psaila, D. Lyden, The metastatic niche: adapting the foreign soil, *Nat. Rev. Cancer* 9 (2009) 285–293.
- [56] H. Peinado, M. Aleckovic, S. Lavotshkin, et al., Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET, *Nat. Med.* 18 (2012) 883–891.
- [57] J. Rak, Cancer: organ-seeking vesicles, *Nature* 527 (2015) 312–314.
- [58] A. Suetsugu, K. Honma, S. Saji, et al., Imaging exosome transfer from breast cancer cells to stroma at metastatic sites in orthotopic nude-mouse models, *Adv. Drug Deliv. Rev.* 65 (2013) 383–390.
- [59] A. Hoshino, B. Costa-Silva, T.L. Shen, et al., Tumour exosome integrins determine organotropic metastasis, *Nature* 527 (2015) 329–335.
- [60] B. Costa-Silva, N.M. Aiello, A.J. Ocean, et al., Pancreatic cancer exosomes initiate pre-metastatic niche formation in the liver, *Nat. Cell Biol.* 17 (2015) 816–826.
- [61] Y.K. Chan, H. Zhang, P. Liu, et al., Proteomic analysis of exosomes from nasopharyngeal carcinoma cell identifies intercellular transfer of angiogenic proteins, *Int. J. Cancer* 137 (2015) 1830–1841.
- [62] T. Jung, D. Castellana, P. Klingbeil, et al., CD44v6 dependence of premetastatic niche preparation by exosomes, *Neoplasia* 11 (2009) 1093–1105.
- [63] Z. Wang, A. von Au, M. Schnolzer, et al., CD44v6-competent tumor exosomes promote motility, invasion and cancer-initiating cell marker expression in pancreatic and colorectal cancer cells, *Oncotarget* 7 (2016) 55409–55436.
- [64] C. Escrevente, S. Keller, P. Altevogt, J. Costa, Interaction and uptake of exosomes by ovarian cancer cells, *BMC Cancer* 11 (2011) 108.
- [65] R.B. Koumangoye, A.M. Sakwe, J.S. Goodwin, et al., Detachment of breast tumor cells induces rapid secretion of exosomes which subsequently mediate cellular adhesion and spreading, *PLoS One* 6 (2011) e24234.
- [66] A. Nanbo, E. Kawanishi, R. Yoshida, H. Yoshiyama, Exosomes derived from Epstein-Barr virus-infected cells are internalized via caveola-dependent endocytosis and promote phenotypic modulation in target cells, *J. Virol.* 87 (2013) 10334–10347.
- [67] L.A. Mulcahy, R.C. Pink, D.R. Carter, Routes and mechanisms of extracellular vesicle uptake, *J. Extracell. Vesic.* 3 (2014).
- [68] C.A. Sanchez, E.I. Andahur, R. Valenzuela, et al., Exosomes from bulk and stem cells from human prostate cancer have a differential microRNA content that contributes cooperatively over local and pre-metastatic niche, *Oncotarget* 7 (2016) 3993–4008.
- [69] R. Chowdhury, J.P. Webber, M. Gurney, et al., Cancer exosomes trigger mesenchymal stem cell differentiation into pro-angiogenic and pro-invasive myofibroblasts, *Oncotarget* 6 (2015) 715–731.
- [70] L. Zhang, S. Zhang, J. Yao, et al., Microenvironment-induced PTEN loss by exosomal microRNA primes brain metastasis outgrowth, *Nature* 527 (2015) 100–104.
- [71] Z.Y. Abd Elmageed, Y. Yang, R. Thomas, et al., Neoplastic reprogramming of patient-derived adipose stem cells by prostate cancer cell-associated exosomes, *Stem Cells* 32 (2014) 983–997.
- [72] S.A. Melo, H. Sugimoto, J.T. O'Connell, et al., Cancer exosomes perform cell-independent microRNA biogenesis and promote tumorigenesis, *Cancer Cell* 26 (2014) 707–721.
- [73] D.G. Meckes Jr., H.P. Gunawardena, R.M. Dekroon, et al., Modulation of B-cell exosome proteins by gamma herpesvirus infection, *Proc. Natl. Acad. Sci. U. S. A.* 110 (2013) E2925–E2933.
- [74] T. Yoshizaki, S. Kondo, N. Wakisaka, et al., Pathogenic role of Epstein-Barr virus latent membrane protein-1 in the development of nasopharyngeal carcinoma, *Cancer Lett.* 337 (2013) 1–7.
- [75] dolens ATPa, Plegmasia alba dolens, *Lect. Clin. Med.* 18655 (2016) 281–332.
- [76] W.L. Dean, M.J. Lee, T.D. Cummins, et al., Proteomic and functional characterization of platelet microparticle size classes, *Thromb. Haemost.* 102 (2009) 711–718.
- [77] M. Drovizio, S. Alberti, A. Sacco, et al., Novel insights into the regulation of cyclooxygenase-2 expression by platelet-cancer cell cross-talk, *Biochem. Soc. Trans.* 43 (2015) 707–714.
- [78] L. Erpenbeck, M.P. Schon, Deadly allies: the fatal interplay between platelets and metastasizing cancer cells, *Blood* 115 (2010) 3427–3436.
- [79] L.J. Gay, B. Felding-Habermann, Contribution of platelets to tumour metastasis, *Nat. Rev. Cancer* 11 (2011) 123–134.
- [80] E. Sierko, M.Z. Wojtukiewicz, Inhibition of platelet function: does it offer a chance of better cancer progression control? *Semin. Thromb. Hemost.* 33 (2007) 712–721.
- [81] S. Karpatkin, E. Pearlstein, C. Ambrogio, B.S. Collier, Role of adhesive proteins in platelet tumor interaction in vitro and metastasis formation in vivo, *J. Clin. Invest.* 81 (1988) 1012–1019.
- [82] B. Nieswandt, M. Hafner, B. Echtenacher, D.N. Mannel, Lysis of tumor cells by natural killer cells in mice is impeded by platelets, *Cancer Res.* 59 (1999) 1295–1300.
- [83] L. Borsig, R. Wong, J. Feramisco, et al., Heparin and cancer revisited: mechanistic connections involving platelets, P-selectin, carcinoma mucins, and tumor metastasis, *Proc. Natl. Acad. Sci. U. S. A.* 98 (2001) 3352–3357.
- [84] G. Berchem, M.Z. Noman, M. Bosseler, et al., Hypoxic tumor-derived microvesicles negatively regulate NK cell function by a mechanism involving TGF- β and miR23a transfer, *Oncoimmunology* 5 (2016) e1062968.
- [85] L.E.I. Rong, R. Li, S. Li, R. Luo, Immunosuppression of breast cancer cells mediated by transforming growth factor- β in exosomes from cancer cells, *Oncol. Lett.* 11 (2016) 500–504.
- [86] M. Labelle, S. Begum, R.O. Hynes, Direct signaling between platelets and cancer cells induces an epithelial-mesenchymal-like transition and promotes metastasis, *Cancer Cell* 20 (2011) 576–590.
- [87] N.L. Szn, M.W.L. Teng, T.S.K. Mok, R.A. Soo, De-novo and acquired resistance to immune checkpoint targeting, *Lancet Oncol* 18 (2017) e731–e741.
- [88] E.U. Wieckowski, C. Visus, M. Szajnik, et al., Tumor-derived microvesicles promote regulatory T cell expansion and induce apoptosis in tumor-reactive activated CD8+ T lymphocytes, *J. Immunol.* 183 (2009) 3720–3730.
- [89] A.J. Abusamra, Z. Zhong, X. Zheng, et al., Tumor exosomes expressing Fas ligand mediate CD8+ T-cell apoptosis, *Blood Cells Mol. Dis.* 35 (2005) 169–173.
- [90] T.L. Whiteside, Immune modulation of T-cell and NK (natural killer) cell activities by TEXs (tumour-derived exosomes), *Biochem. Soc. Trans.* 41 (2013) 245–251.
- [91] M. Szajnik, M. Czystowska, M.J. Szczepanski, et al., Tumor-derived microvesicles induce, expand and up-regulate biological activities of human regulatory T cells (Treg), *PLoS One* 5 (2010) e11469.
- [92] S. Nagaraj, D.I. Gabrilovich, Regulation of suppressive function of myeloid-derived suppressor cells by CD4+ T cells, *Semin. Cancer Biol.* 22 (2012) 282–288.
- [93] F. Chalmin, S. Ladoire, G. Mignot, et al., Membrane-associated Hsp72 from tumor-derived exosomes mediates STAT3-dependent immunosuppressive function of mouse and human myeloid-derived suppressor cells, *J. Clin. Invest.* 120 (2010) 457–471.
- [94] D. Mrizak, N. Martin, C. Barjon, et al., Effect of nasopharyngeal carcinoma-derived exosomes on human regulatory T cells, *J. Natl. Cancer Inst.* 107 (2015) 363.
- [95] S.B. Ye, Z.L. Li, D.H. Luo, et al., Tumor-derived exosomes promote tumor progression and T-cell dysfunction through the regulation of enriched exosomal microRNAs in human nasopharyngeal carcinoma, *Oncotarget* 5 (2014) 5439–5452.
- [96] A. Clayton, S. Al-Taei, J. Webber, et al., Cancer exosomes express CD39 and CD73, which suppress T cells through adenosine production, *J. Immunol.* 187 (2011) 676–683.
- [97] L. Muller, M. Mitsuhashi, P. Simms, et al., Tumor-derived exosomes regulate expression of immune function-related genes in human T cell subsets, *Sci. Rep.* 6 (2016) 20254.
- [98] R.A. Clynes, T.L. Towers, L.G. Presta, J.V. Ravetch, Inhibitory Fc receptors modulate in vivo cytotoxicity against tumor targets, *Nat. Med.* 6 (2000) 443–446.
- [99] C. Battke, R. Ruiss, U. Welsch, et al., Tumour exosomes inhibit binding of tumour-reactive antibodies to tumour cells and reduce ADCC, *Cancer Immunol. Immunother.* 60 (2011) 639–648.
- [100] T. Aung, B. Chapuy, D. Vogel, et al., Exosomal evasion of humoral immunotherapy in aggressive B-cell lymphoma modulated by ATP-binding cassette transporter A3, *Proc. Natl. Acad. Sci. U. S. A.* 108 (2011) 15336–15341.
- [101] B.Z. Qian, J.W. Pollard, Macrophage diversity enhances tumor progression and metastasis, *Cell* 141 (2010) 39–51.
- [102] L.S. Ojalvo, C.A. Whittaker, J.S. Condeelis, J.W. Pollard, Gene expression analysis of macrophages that facilitate tumor invasion supports a role for Wnt-signaling in mediating their activity in primary mammary tumors, *J. Immunol.* 184 (2010) 702–712.
- [103] A. Chow, W. Zhou, L. Liu, et al., Macrophage immunomodulation by breast cancer-derived exosomes requires Toll-like receptor 2-mediated activation of NF-kappaB, *Sci. Rep.* 4 (2014) 5750.
- [104] M. Fabbri, A. Paone, F. Calore, et al., MicroRNAs bind to Toll-like receptors to induce prometastatic inflammatory response, *Proc. Natl. Acad. Sci. U. S. A.* 109 (2012) E2110–E2116.
- [105] V.C. Ramani, A. Purushothaman, M.D. Stewart, et al., The heparanase/syndecan-1 axis in cancer: mechanisms and therapies, *FEBS J.* 280 (2013) 2294–2306.
- [106] C.A. Thompson, A. Purushothaman, V.C. Ramani, et al., Heparanase regulates secretion, composition, and function of tumor cell-derived exosomes, *J. Biol. Chem.* 288 (2013) 10093–10099.
- [107] M.F. Baietti, Z. Zhang, E. Mortier, et al., Syndecan-syntenin-ALIX regulates the

- biogenesis of exosomes, *Nat. Cell Biol.* 14 (2012) 677–685.
- [108] A. Bobrie, S. Krumeich, F. Rey, et al., Rab27a supports exosome-dependent and -independent mechanisms that modify the tumor microenvironment and can promote tumor progression, *Cancer Res.* 72 (2012) 4920–4930.
- [109] D. Hoshino, K.C. Kirkbride, K. Costello, et al., Exosome secretion is enhanced by invadopodia and drives invasive behavior, *Cell Rep.* 5 (2013) 1159–1168.
- [110] D.A. Murphy, S.A. Courtneidge, The 'ins' and 'outs' of podosomes and invadopodia: characteristics, formation and function, *Nat. Rev. Mol. Cell Biol.* 12 (2011) 413–426.
- [111] N. Kosaka, H. Iguchi, Y. Yoshioka, et al., Secretory mechanisms and intercellular transfer of microRNAs in living cells, *J. Biol. Chem.* 285 (2010) 17442–17452.
- [112] S. Sento, E. Sasabe, T. Yamamoto, Application of a persistent heparin treatment inhibits the malignant potential of oral squamous carcinoma cells induced by tumor cell-derived exosomes, *PLoS One* 11 (2016) e0148454.
- [113] H.G. Zhang, H. Kim, C. Liu, et al., Curcumin reverses breast tumor exosomes mediated immune suppression of NK cell tumor cytotoxicity, *Biochim. Biophys. Acta* 2007 (1773) 1116–1123.
- [114] S. Taverna, M. Giallombardo, M. Pucci, et al., Curcumin inhibits in vitro and in vivo chronic myelogenous leukemia cells growth: a possible role for exosomal disposal of miR-21, *Oncotarget* 6 (2015) 21918–21933.
- [115] N.L. Syn, L. Wang, E.K. Chow, et al., Exosomes in cancer nanomedicine and immunotherapy: prospects and challenges, *Trends Biotechnol.* 35 (2017) 665–676.
- [116] L. Zitvogel, A. Regnault, A. Lozier, et al., Eradication of established murine tumors using a novel cell-free vaccine: dendritic cell-derived exosomes, *Nat. Med.* 4 (1998) 594–600.
- [117] J.A. Cho, Y.S. Lee, S.H. Kim, et al., MHC independent anti-tumor immune responses induced by Hsp70-enriched exosomes generate tumor regression in murine models, *Cancer Lett.* 275 (2009) 256–265.
- [118] J. Wolfers, A. Lozier, G. Raposo, et al., Tumor-derived exosomes are a source of shared tumor rejection antigens for CTL cross-priming, *Nat. Med.* 7 (2001) 297–303.
- [119] J.A. Cho, D.J. Yeo, H.Y. Son, et al., Exosomes: a new delivery system for tumor antigens in cancer immunotherapy, *Int. J. Cancer* 114 (2005) 613–622.
- [120] C. Kahlert, R. Kalluri, Exosomes in tumor microenvironment influence cancer progression and metastasis, *J. Mol. Med. (Berl)* 91 (2013) 431–437.
- [121] F. Andre, N.E. Scharz, M. Movassagh, et al., Malignant effusions and immunogenic tumour-derived exosomes, *Lancet* 360 (2002) 295–305.
- [122] M.A. Morse, J. Garst, T. Osada, et al., A phase I study of dexosome immunotherapy in patients with advanced non-small cell lung cancer, *J. Transl. Med.* 3 (2005) 9.
- [123] B. Escudier, T. Dorval, N. Chaput, et al., Vaccination of metastatic melanoma patients with autologous dendritic cell (DC) derived-exosomes: results of the first phase I clinical trial, *J. Transl. Med.* 3 (2005) 10.
- [124] T. Boulikas, Clinical overview on lipoplatin: a successful liposomal formulation of cisplatin, *Expert Opin. Investig. Drugs* 18 (2009) 1197–1218.
- [125] M. Airolidi, D. Amadori, S. Barni, et al., Clinical activity and cardiac tolerability of non-pegylated liposomal doxorubicin in breast cancer: a synthetic review, *Tumori* 97 (2011) 690–692.
- [126] M.S. Kim, M.J. Haney, Y. Zhao, et al., Development of exosome-encapsulated paclitaxel to overcome MDR in cancer cells, *Nanomedicine* 12 (2016) 655–664.
- [127] H. Li, P. Zhang, X. Sun, et al., MicroRNA-181a regulates epithelial-mesenchymal transition by targeting PTEN in drug-resistant lung adenocarcinoma cells, *Int. J. Oncol.* 47 (2015) 1379–1392.
- [128] L. Alvarez-Erviti, Y. Seow, H. Yin, et al., Delivery of siRNA to the mouse brain by systemic injection of targeted exosomes, *Nat. Biotechnol.* 29 (2011) 341–345.
- [129] B. Besse, M. Charrier, V. Lapiere, et al., Dendritic cell-derived exosomes as maintenance immunotherapy after first line chemotherapy in NSCLC, *Oncoimmunology* 5 (2016) e1071008.
- [130] E. Quoix, R. Ramlau, V. Westeel, et al., Therapeutic vaccination with TG4010 and first-line chemotherapy in advanced non-small-cell lung cancer: a controlled phase 2B trial, *Lancet Oncol.* 12 (2011) 1125–1133.
- [131] P. Peng, Y. Yan, S. Keng, Exosomes in the ascites of ovarian cancer patients: origin and effects on anti-tumor immunity, *Oncol. Rep.* 25 (2011) 749–762.
- [132] I.V. Miller, T.G. Grunewald, Tumour-derived exosomes: tiny envelopes for big stories, *Biol. Cell.* 107 (2015) 287–305.
- [133] R. Kalluri, The biology and function of exosomes in cancer, *J. Clin. Invest.* 126 (2016) 1208–1215.
- [134] S.A. Melo, L.B. Luecke, C. Kahlert, et al., Glypican-1 identifies cancer exosomes and detects early pancreatic cancer, *Nature* 523 (2015) 177–182.
- [135] J. Lu, G. Getz, E.A. Miska, et al., MicroRNA expression profiles classify human cancers, *Nature* 435 (2005) 834–838.
- [136] R.J. Bryant, T. Pawlowski, J.W. Catto, et al., Changes in circulating microRNA levels associated with prostate cancer, *Br. J. Cancer* 106 (2012) 768–774.
- [137] Z.H. Chen, G.L. Zhang, H.R. Li, et al., A panel of five circulating microRNAs as potential biomarkers for prostate cancer, *Prostate* 72 (2012) 1443–1452.
- [138] J. Shen, G.W. Hruby, J.M. McKiernan, et al., Dysregulation of circulating microRNAs and prediction of aggressive prostate cancer, *Prostate* 72 (2012) 1469–1477.
- [139] C. Zijlstra, W. Stoorvogel, Prostatomes as a source of diagnostic biomarkers for prostate cancer, *J. Clin. Invest.* 126 (2016) 1144–1151.
- [140] H. Ogata-Kawata, M. Izumiya, D. Kurioka, et al., Circulating exosomal microRNAs as biomarkers of colon cancer, *PLoS One* 9 (2014) e92921.
- [141] J. Wang, Y. Liu, W. Sun, et al., Plasma exosomes as novel biomarker for the early diagnosis of gastric cancer, *Cancer Biomark* 21 (2018) 805–812.
- [142] S. Cui, Z. Cheng, W. Qin, L. Jiang, Exosomes as a liquid biopsy for lung cancer, *Lung Cancer* 116 (2018) 46–54.
- [143] T. Lener, M. Gimona, L. Aigner, et al., Applying extracellular vesicles based therapeutics in clinical trials - an ISEV position paper, *J. Extracell. Vesic.* 4 (2015) 30087.
- [144] S. Wilhelm, A.J. Tavares, Q. Dai, et al., Analysis of nanoparticle delivery to tumours, *Nat. Rev. Mater.* 1 (2016) 16014.