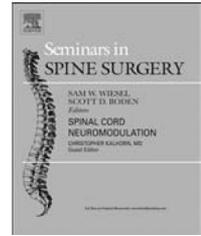


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Postoperative spine infections

John Attenello^a, and R.Todd Allen^{b,*}

^aDepartment of Orthopaedic Surgery, University of Hawaii, Honolulu, HI, United States

^bDepartment of Orthopaedic Surgery, UC San Diego Health System, University of California, San Diego, California, United States

ABSTRACT

Postoperative spine infections can be a devastating complication with significant consequences for a patient. Focus should be placed on prevention, early diagnosis, and successful treatment strategies. Surgeons should maintain a high index of suspicion and initiate proper diagnostic workup and evaluation when concerned for a possible infection. This article will provide a global review of postoperative spine infection including incidence by surgical approach, specific risk factors, common presentations, diagnostic workup, prevention strategies, and both nonsurgical and surgical management.

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1. Introduction

Postoperative infection of the spine is a feared complication that can be devastating to patients. In addition to the increased morbidity and poor patient outcomes, infections account for several billion dollars in U.S. health care expenditures.¹ Deep infections after spinal fusion have been shown to result in a greater than 4 times increase in total costs, in excess of \$100,000 in some cases.² Prevention, early diagnosis, and successful treatment strategies are critical to reducing morbidity and health care costs. Surgeons should maintain a high index of suspicion for infection and initiate proper diagnostic workup and evaluation when concerned.

1.1. Epidemiology and incidence

The incidence of surgical site infection (SSI) following spine surgery has historically been reported in the literature between 1% to 6%, with some reports up to 20%.^{3–9} This wide variation is in part due to variation in the definition of SSI, the length of follow up, and small heterogeneous patient populations. A recent report from the Scoliosis Research Society

Morbidity and Mortality Committee reported a 2.1% infection rate (0.8% superficial and 1.3% deep) from their database of 108,419 spine surgery cases.¹⁰ However, incidence varies depending on spine region (cervical, thoracic, or lumbar), surgical technique (MIS or open), approach (anterior, posterior or lateral), and presence of instrumentation.

Non-instrumented procedures typically have lower rates of infection. Lumbar discectomy have reported rates of less than 1%,^{11,12} and one series of 262 endoscopic discectomies reported no infections.¹³ Higher rates have been reported between 1.25% to 2% with laminectomy without fusion^{14–15} and even slightly higher between 2.1% to 3.5% when non-instrumented fusion is performed.^{15,16} In general, longer, more invasive procedures with greater blood loss and soft tissue dissection have been shown to increase the risk for infection.¹⁷

A higher incidence of infection is seen with the use of instrumentation in posterior spine procedures at approximately 5%.^{18–22} The SRS M&M database reported a 28% higher rate of infection in cases with implants than those without (2.3% vs 1.8%).¹⁰ The implants do not introduce the source of infection but rather serve as a site for secondary inoculation.²³ Colonization of implants may also lead to delayed and resistant infections.²⁴

From UC San Diego Health System, University of California, San Diego

*Corresponding author.

E-mail address: rtallen@health.ucsd.edu (R.T. Allen).

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Anterior spinal procedures have a very low incidence of infection which is likely in part due to improved vascularity of the anterior spinal column and less soft tissue trauma. Reported rates are as low as 0 to 1%.^{6,25–27} Combined anterior-posterior have the highest risk of infection,^{10,27} but are typically more complex. Minimally invasive techniques are designed to minimize soft tissue disruption, blood loss, and thereby theoretically decrease infection risk. However, several studies have reported inconsistent results. Some studies favor a lower infection risk with MIS,^{28–29} whereas others show no difference,^{30,31} but there does appear to be a benefit of MIS in multilevel fusions.³²

Spine trauma patients are a particularly high-risk population due to significant soft tissue injury and subsequent tissue necrosis and hematoma formation. Surgically treated traumatic spine injuries have postoperative infection rates reported between 9% to 15%.³³ Blam et al. reported a 9.4% postoperative wound infection rate in 256 spinal trauma patients compared to a 3.7% infection rate in 112 patients undergoing elective spine surgery.³⁴ Another study demonstrated similar rates in both trauma and elective patients.³⁵

1.2. Risk factors

Causes of postoperative infections are multifactorial and risk factors are both patient-specific and surgery-specific. Patient specific risk factors are important to recognize in the preoperative work-up because many can be modifiable and if addressed early, improve surgical outcomes. These modifiable risk factors include tobacco and alcohol use, obesity, diabetes, and malnutrition.^{8,28,36–38}

Obesity is frequently encountered and can lead to poor outcomes.^{28,39–40} Obese patients typically require longer operative times, more extensive tissue dissection, and greater retraction force resulting in tissue necrosis in already poorly vascularized adipose tissue. The distribution of adipose tissue may serve as a better predictor for SSI than actual BMI.^{41,42} Mehta et al. showed a significant association between SSI and the thickness of subcutaneous fat and lamina-to-skin distance.⁴¹ Meng et al. reported an odds ratio of 2.13 for patients with a BMI > 30 kg/m².⁴³ Obesity is also typically associated with diabetes, malnutrition, and other medical comorbidities which further increase the risk of infection.

Poorly controlled diabetes significantly impairs wound healing and increases risk for infection. Postoperative wound infections have been reported up to 24%.^{28,44} Schuster et al. performed a large meta-analysis where the majority of studies found diabetes and elevated glucose levels to have a statistically significant association.⁴⁵ Chen et al. performed a retrospective level II study which showed that diabetic patients undergoing spinal fusion are greater than 4 times more likely to develop SSI.⁴⁶ Olsen et al. found DM patients to be 3.5 times as likely to develop an infection in their cohort of 2316 patients.⁴⁰ Meng et al. reported a 2 times higher risk of infection in diabetic patients.⁴³ In regards to hemoglobin A1C, Hikata et al. showed that patients with values greater than 7 had a 35% infection rate compared to no infections seen in patients with a values less than 7.⁴⁷ Diabetes can also lead to cardiovascular and renal disease which further

increases infection risk. Strict perioperative control of blood glucose levels should be maintained.

Malnutrition is commonly undetected but has been shown to impair the immune response and delay wound healing. Common indices used are serum albumin less than 3.5 mg/dL or a total lymphocyte count less than 1500 cell/mm.⁴⁸ Klein et al. reviewed 114 patients undergoing elective spine surgery and 1 out of every 4 were considered malnourished.³⁷ Of these 13 malnourished patients, 11 (85%) developed a postoperative wound infection. Malnutrition can also develop during postoperative hospitalizations or between staged procedures.⁴⁹ Malnutrition can also be a sign of malignancy which has infection rates reported up to 20%.^{6,50}

Tobacco use significantly increases the risk of postoperative wound infections.⁵¹ Meng et al. found a relative risk of 1.17⁴³ and Thomsen et al. showed that patients who stopped smoking had half as many complications.⁵² Proposed mechanisms include poor tissue oxygenation and impaired neutrophil function.⁵³ Routine preoperative counseling on smoking cessation should be performed.

Non modifiable risk factors include immunocompromised states, rheumatoid arthritis medications, steroid use, and age.

Radiation therapy for the management of tumors poses a higher risk of infection.⁵⁴ Spine surgery following neoadjuvant irradiation to the surgical site increases the risk of infection and wound healing issues.^{8,50,55} Therefore, elective spine surgery should typically be delayed for 6 to 12 weeks after preoperative radiation therapy. For postoperative radiation therapy, a delay of 3 weeks after surgery is typically recommended.

Surgery specific risk factors include increased number of levels, surgery greater than 5 h, blood loss greater than 1 liter, blood transfusion, and high complexity of procedure are associated with increased risk of infection.^{56–57} Revision surgery, use of allograft, and procedures involving the sacrum or pelvis are also surgical risk factors.^{18,58}

2. Patient evaluation

2.1. Presentation

The presentation of a patient with postoperative spine infection is different for superficial and deep infections, but the most common presenting symptom for both is pain. A superficial infection occurs above the fascia and typically presents within the first 2 postoperative weeks with local tenderness, erythema, warmth and drainage. If presenting early and without the presence of systemic symptoms or increasing surgical site pain, superficial infections can often be adequately treated with local wound care and 2 weeks of oral antibiotics. However, there should be a high index of suspicion for a deep infection if the wound continues to drain, surgical site pain worsens, or systemic symptoms such as fever, chills, night sweats, lethargy, or malaise develop.

The presentation of deep infections is variable and can occur anytime postoperatively from weeks to months to years.^{59,60} Pain, fever and night sweats may be seen, but many have no systemic symptoms. New onset of pain is particularly worrisome. There may be purulent drainage from a sinus tract to the skin or a lack of any impressive external

evidence. Delayed presentations typically have back pain, drainage and erythema but without fever. Neurologic deficits can occur, usually due to a spinal epidural abscess.

2.2. Evaluation

Diagnostic work up should include a thorough history and physical exam, labs, imaging and further studies as indicated. Laboratory tests include WBC, ESR, and CRP. Results can be difficult to interpret in the immediate postoperative period. WBC can be elevated from surgical stress. ESR elevates immediately postoperatively to a peak at day 4 before normalizing two to four weeks later.⁶¹ Higher peak ESR values have been shown to correlate with the more extensive or invasive the procedures is.⁶¹ Persistently elevated ESR, particularly over 2 standard deviations, are concerning for infection.⁶² CRP is also elevated in the immediate postoperative period with a peak at day 3, but with a more rapid decline than ESR, with complete normalization occurring at 10–14 days. The rapid decline makes CRP a more sensitive and useful diagnostic tool and CRP is nearly always elevated in SSI.⁶³ Variations in pathogen virulence as well as host immunocompetence can mitigate results, but when taken together and viewed as a trend, WBC, ESR and CRP can be helpful in recognizing a postoperative infection as well as monitor treatment efficacy and prognosis.

Identification of the offending organism requires reliable cultures. Blood cultures should be taken prior to the initiation of antibiotics and if positive for a specific organisms, it can be presumed to be the causative organism. Wound cultures can be obtained easily but are often contaminated with skin flora and unreliable. CT guided aspiration can obtain more precise deep cultures. The most precise cultures come directly from the source during surgical debridement when intervention is feasible and necessary.

2.3. Imaging

Imaging can also aid in the diagnosis. Evidence on plain radiographs often takes up to 4 weeks to show.⁶⁴ Early findings can include lucency around instrumentation, soft tissue gas, bony destruction, and disc space narrowing which eventually transition to vertebral body collapse, endplate sclerosis, and osteolysis months later.^{65–66} Distinguishing early changes from coexisting degenerative disc disease can be difficult. CT can provide better detail than plain radiographs on bony changes in early infection or as a substitute when MRI is contraindicated or difficult to interpret due to artifact. CT can also be used to assist in biopsy and delineate an abscess with contrast enhancement.

MRI is the most sensitive (93%) and specific (96%) for diagnosing spinal infections.^{67–68} MRI findings of discitis or osteomyelitis are marrow and disc space hypointensity on T1-weighted sequence and hyperintensity on T2-weighted or STIR sequences (Fig. 1).⁶⁸ Additional findings of endplate destruction and disc space narrowing further contribute to the diagnosis of infection. A spinal epidural abscess appears isointense on T1-weighted sequence and hyperintense on T2-weighted sequence, and typically is compressing neural elements (Fig. 2). Addition of gadolinium contrast delineates an abscess with rim enhancement. MRI can also identifies adjacent soft tissue masses.

3. Prevention techniques

3.1. Preoperative options

Prevention is the first step to reducing postoperative infections and begins with preoperative optimization of modifiable

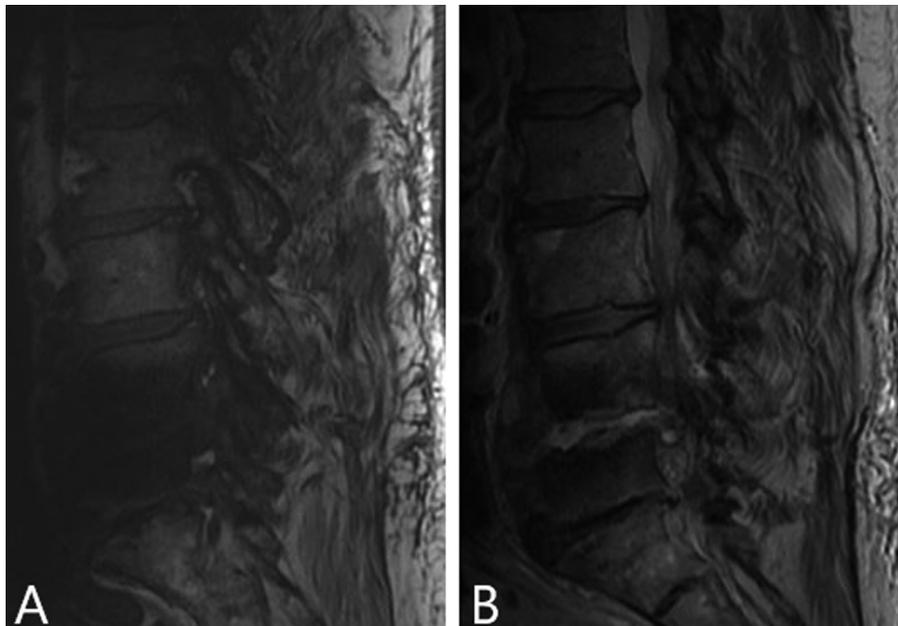


Fig. 1 – 65 year old male with severe lumbar osteomyelitis and spondylodiskitis. (A) Sagittal T1 weighted MRI demonstrating L4-L5 disc space and adjacent marrow hypointensity. (B) Sagittal T2 weighted MRI demonstrating relative hyperintensity in the same region.



Fig. 2—75 year old diabetic female with end stage renal disease who presented with severe progressive back pain and lower extremity weakness with epidural and retroperitoneal abscess. (A) Sagittal T2 MRI image showing an anterior epidural abscess extending from L4-S1. (B) Sagittal CT reconstruction demonstrating severe end plate erosions of L5 and S1. (C and D) Postoperative AP and lateral radiographs following anterior debridement, L4 and L5 corpectomy, L4-S1 interbody cage reconstruction and L3 to pelvis posterior instrumentation with PMMA cement augmentation.

risk factors including smoking cessation, nutritional status, and glycemic control. Smoking cessation for a minimum of 4 weeks prior to surgery has been shown to reduce SSI.⁶⁹ Nutrition should be optimized prior to surgery with a focus on serum albumin and total lymphocyte count.³⁴ Obesity patients may require dietary or exercise counseling, or referral for bariatric surgery. Tight glycemic control should be maintained in the perioperative period. Any infection including urinary tract, gastrointestinal or respiratory should be fully resolved prior to elective surgery. Hair over the surgical site should be removed with clippers rather than a razor to avoid inadvertent microabrasions. MSSA and MRSA screening and eradication with intranasal 2% mupirocin ointment and 2% chlorhexidine gluconate (CHG) showers for 5 days lowered the rate of SSI after spine surgery by half.^{70–71}

3.2. Intraoperative options

Prophylactic antibiotics against common skin flora has shown a trend toward decreasing infection rates^{62,72–73} and is recommended by the North American Spine Society.⁷⁴ Recently cephalosporin administration 30 min prior to skin incision was shown to significantly decrease SSI risk compared to the previous recommendation of 31 to 60 min prior to incision.⁷⁵ Cephalosporins should be readministered every 4 h or after 1500 mL of blood loss and vancomycin and gentamycin should be redosed every 8 h.^{76–78} Cefazolin is a first generation cephalosporin that is most commonly used because it is effective against gram-positive bacteria such as *Staphylococcus Aureus*. Clindamycin can be used as a substitute for patients with beta lactam allergies, and vancomycin for patients with a history of MRSA. Postoperative antibiotics are typically discontinued after 24 h and the use of drains should not alter the duration.

Proper sterile technique and skin antisepsis preparation with iodine or chlorhexidine solutions should be performed in all cases. Surgeon double gloving to prevent perforation,⁷⁹ and intermittently changing outer gloves, particularly prior to handling instrumentation has also been shown to reduce SSI during spine surgery.⁸⁰ Intraoperatively, surgeons should periodically release retractors, maintain hemostasis and remove of any necrotic tissue that could become infected.^{81–82} Surgical site irrigation with saline or various solutions have been used without much significant clinical data to support one solution over another. However, Cheng et al. performed a prospective, blinded and randomized study using 3.5% betadine irrigation for 3 min after saline versus saline alone in 414 spine surgeries and reported a significant reduction in deep SSI without any adverse events.⁸³

The use of topical vancomycin powder applied directly to the wound provides a locally high concentration of antibiotic without systemic implications. Several meta-analyses and systematic reviews support its use in significantly reducing postoperative SSI.^{84–87} Khan et al. reported a relative risk of 0.34 with the use of vancomycin powder.⁸⁸ Sweet et al. reviewed 1732 consecutive posterior instrumented thoracic and lumbar fusions and reported an SSI rate of 2.6% in 821 patients receiving IV cephalexin prophylaxis alone compared to 0.2% in 911 patients when adjunctive vancomycin powder was added.⁸⁹ Vancomycin powder is cost effective,⁹⁰ safe in adults^{89,91} and children,^{92–93} easy to deliver and provides broad coverage

The use of postsurgical drains is controversial. The goal is to prevent hematoma formation and therefore the risk of subsequent seeding. Some studies report decreased risk of SSI while others report no difference or an increased risk of SSI.^{94–95} Use of drains have been associated with higher rates of blood transfusions.⁹⁶ Prolonged use of antibiotics with the use of a drain have not been supported.⁹⁷

4. Management of infection

4.1. Nonsurgical

The goal of nonsurgical and surgical debridement is resolution of the infection while maintaining spinal stability. Determining the timing, location and extent of the infection will help guide the choice of management. Infections presenting early in the first few weeks postoperatively are typically due to direct inoculation of more virulent bacteria such as MRSA.^{28,98–99} In contrast, delayed infections present several months after the index surgery and are commonly associated with less virulent bacteria that may have colonized instrumentation with a biofilm, such as *Propionibacterium acnes*.^{100–101} Preoperative imaging may identify the region of infection. Minor superficial infections without wound breakdown, purulence, or fluctuance may only require local wound care and antibiotics. If no organism is identified, antibiotics targeting skin flora, such as *Staphylococcus*, may be adequate. For anything beyond a simple stitch abscess, a typical course is 6 weeks IV antibiotics followed by 6 weeks of oral antibiotics.¹⁰² An orthotic brace can help mitigate symptoms.

Surgeons should remain cautious and maintain close follow up with serial inflammatory markers because these nonsurgical interventions may not completely resolve the infection. Failure to respond to antibiotics or presence of neurologic deterioration, spinal instability or severe persistent pain may necessitate surgical debridement in addition to IV antibiotics.

4.2. Surgical

Surgical intervention includes meticulous and systematic debridement of nonviable tissue, obtaining multiple deep and superficial tissue cultures and exploration of the wound for sinus tracts and integrity of instrumentation. Preoperative antibiotics should be withheld until culture specimens are obtained. The wound bed should be copiously irrigated and the need for repeat debridement should be determined based on the severity of the infection and surgeon discretion. A scoring system was developed to assist surgeons with this decision. The Postoperative Infection Treatment Score for the Spine (PITSS) predicts which patients may need multiple irrigation and debridements based on 6 categories – location, comorbidity, microbiology, distant site infection, instrumentation and bone graft.¹⁰³ Low risk, indeterminate and high risk correspond to scores of 7–14, 15–20, and 21–33, respectively. Repeat debridements typically take place 48 to 72 h after initial debridement.

No consensus exists on whether to remove well-fixed instrumentation. No level I or II evidence is available on retention versus removal. Some authors support routine removal of all instrumentation regardless of fusion status in order to adequately eradicate the infection.^{59,60,104–107} Others authors advocate retaining well-fixed instrumentation after a thorough debridement to prevent loss of correction or catastrophic instability leading to neurologic compromise or deformity.^{4,18,57,108–116} These authors have also demonstrated adequate removal of infection with retained instrumentation.

Retaining instrumentation is favored in early infection whereas removal of instrumentation is favored in delayed infection.¹¹⁷ Another technique is to rely on suppressive IV antibiotics until a solid fusion is achieved and then remove instrumentation. If loose instrumentation is encountered and must be reimplanted into an infected site, titanium implants are favored over stainless steel implants because they are more resistant to glycoalyx biofilm formation.¹¹⁸ Pseudoarthrosis is encountered more frequently after infection and close follow-up should be maintained, particularly if instrumentation is removed and there is a risk for instability.^{57,119}

Primary layered wound closure after adequate debridement and irrigation is typically preferred, if feasible. Significant infection and myonecrosis often results in large soft tissue defects that require multiple returns to the operating room and delayed closure via secondary intention or potentially reconstructive soft tissue coverage such as a trapezius rotation flap¹²⁰ or paraspinous muscle flap.¹²¹ The use of a drain to eliminate dead space is typically recommended. The use of vacuum-assisted closure (VAC) devices safely and effectively has been described^{56,98,122–124} and some authors recommend using it on all postoperative spine infections.¹²⁵ The wound VAC is thought to eliminate dead space, improve vascularity and decrease bacteria levels.⁶ Lee et al. reported 42 patients with deep postoperative spinal infections treated with wound VACs that all fully healed. 30 patients also had exposed dura which were covered with a layer of Jelonet or Mepitel, a white sponge, and negative pressure limited to 50 mmHg and no complications occurred from the wound VAC. Wound VACs have also shown efficacy in MRSA or multiple bacteria infections.¹²⁶

Patients should be followed for symptomatic improvement of back pain and downtrending ESR, CRP, and WBC. Serial Imaging can be used to evaluate for maintenance of instrumentation, alignment, fusion. Evidence of resolution of infection on imaging may lag behind treatment for several weeks to months.

5. Conclusions

Postoperative spinal infections can have devastating outcomes for patients. Prevention is the critical first step to protecting patients. Patients should be medically optimized and modifiable risk factors should be addressed preoperatively. Intraoperative measures include routine administration of preoperative antibiotics, maintenance of proper sterile technique, and application of vancomycin powder prior to closure. Despite these precautions, postoperative infections inevitably occur and therefore require a high index of suspicion. Patients with increasing surgical site pain or pain out of proportion to what is expected, particularly with concomitant systemic symptoms, should undergo a diagnostic infectious work up. MRI with and without contrast should be obtained along with ESR, CRP, WBC, blood cultures, and possibly CT guided or open biopsy to tailor antibiotics. Most patients may be adequately treated with antibiotics alone, but surgical debridement is often necessary. Single or multiple thorough debridements may be required with primary or delayed

closure. There is no consensus on whether to remove or retain instrumentation. Patients should be followed closely for improvement and be wary of the development of pseudoarthrosis or instability.

Disclosures

The authors report no proprietary or commercial interest in any product mentioned or concept discussed in this article.

REFERENCES

- Kirkland KB, Briggs JP, Trivette SL, et al. The impact of surgical site infections in the 1990s: attributable mortality, excess length of hospitalization, and extra costs. *Infect Control Hosp Epidemiol.* 1999;20:725–730.
- Calderone RR, Garland DE, Capen DA, et al. Cost of medical care for postoperative spinal infections. *Orthop Clin North Am.* 1996;27:171–182.
- Abbey DM, Turner DM, Warson JS, et al. Treatment of postoperative wound infections following spinal fusion with instrumentation. *J Spinal Disord.* 1995;8:278–283.
- Levi AD, Dickman CA, Sonntag VK. Management of postoperative infections after spinal instrumentation. *J Neurosurg.* 1997;86:975–980.
- Roberts FJ, Walsh A, Wing P, et al. The influence of surveillance methods on surgical wound infection rates in a tertiary care spinal surgery service. *Spine.* 1998;23:366–370.
- Weinstein MA, McCabe JP, Cammisa Jr FP. Postoperative spinal wound infection: a review of 2,391 consecutive index procedures. *J Spinal Disord.* 2000;13:422–426.
- Banco SP, Vaccaro AR, Blam O, et al. Spine infections: variations in incidence during the academic year. *Spine.* 2002;27:962–965.
- Olsen MA, Mayfield J, Laurysen C, et al. Risk factors for surgical site infection in spinal surgery. *J Neurosurg.* 2003;98:149–155.
- Wimmer C, Gluch H. Management of postoperative wound infection in posterior spinal fusion with instrumentation. *J Spinal Disord.* 1996;9:505–508.
- Smith JS, Shaffrey CI, Sansur CA, et al. Rates of infection after spine surgery based on 108,419 procedures: a report from the Scoliosis Research Society Morbidity and Mortality Committee. *Spine.* 2011;36(7):556–563.
- Bongartz EB, Ulrich P, Fidler M, et al. Reoperation in the management of post-operative disc space infection. *Zentralbl Neurochir.* 1994;55:120–124.
- Mastrorardi L, Rychlicki F, Tatta C, et al. Spondylodiscitis after lumbar microdiscectomy: effectiveness of two protocols of intraoperative antibiotic prophylaxis in 1167 cases. *Neurosurg Rev.* 2005;28:303–307.
- Hoogland T, van den Brekel-Dijkstra K, Schubert M, et al. Endoscopic transforaminal discectomy for recurrent lumbar disc herniation: a prospective, cohort evaluation of 262 consecutive cases. *Spine.* 2008;33:973–978.
- Hansraj KK, Cammisa Jr FP, O'Leary PF, et al. Decompressive surgery for typical lumbar spinal stenosis. *Clin Orthop Relat Res.* 2001;384:10–17.
- 15 National Nosocomial Infections Surveillance (NNIS). System report, data summary from January 1992 through June 2004, issued October 2004. *Am J Infect Control.* 2004;32(8):470–485.
- Horwitz NH, Curtin JA. Prophylactic antibiotics and wound infections following laminectomy for lumbar disc herniation. *J Neurosurg.* 1975;43:727–731.
- Cizik AM, Lee MJ, Martin BI, et al. Using the spine surgical invasiveness index to identify risk of surgical site infection: a multivariate analysis. *J Bone Joint Surg Am.* 2012;94(4):335–342.
- Perry JW, Montgomerie JZ, Swank S, Gilmore DS, Maeder K. Wound infections following spinal fusion with posterior segmental spinal instrumentation. *Clin Infect Dis.* 1997;24(4):558–561.
- Richards BR, Emara KM. Delayed infections after posterior TSRH spinal instrumentation for idiopathic scoliosis: revisited. *Spine.* 2001;26(18):1990–1996.
- Richards BS. Delayed infections following posterior spinal instrumentation for the treatment of idiopathic scoliosis. *J Bone Joint Surg Am.* 1995;77(4):524–529.
- Massie JB, Heller JG, Abitbol JJ, McPherson D, Garfin SR. Postoperative posterior spinal wound infections. *Clin Orthop Relat Res.* 1992;284:99–108.
- Kasliwal MK, Tan LA, Traynelis VC. Infection with spinal instrumentation: review of pathogenesis, diagnosis, prevention, and management. *Surg Neurol Int.* 2013;4(suppl):S392–S403.
- Singh K, Rechtine G, Heller J, et al. Postoperative spine infections. In: Herkowitz, Garfin, Eismont, et al., eds. *Rothman-Simeone The Spine.* 5th ed Philadelphia: Saunders; 2006:1496–1510.
- Kasliwal MK, Tan LA, Traynelis VC. Infection with spinal instrumentation: review of pathogenesis, diagnosis, prevention, and management. *Surg Neurol Int.* 2013;4:S392–S403.
- Zeidman SM, Ducker TB, Raycroft J. Trends and complications in cervical spine surgery: 1989–1993. *J Spinal Disord.* 1997;10(6):523–526.
- Fang A, Hu SS, Endres N, et al. Risk factors for infection after spinal surgery. *Spine.* 2005;30:1460–1465.
- O'Toole JE, Eichholz KM, Fessler RG. Surgical site infection rates after minimally invasive spinal surgery. *J Neurosurg Spine.* 2009;11(4):471–476.
- Parker SL, Adogwa O, Witham TF, et al. Post-operative infection after minimally invasive versus open transforaminal lumbar interbody fusion (TLIF): literature review and cost analysis. *Minim Invasive Neurosurg.* 2011;54(1):33–37.
- McGirt MJ, Parker SL, Lerner J, et al. Comparative analysis of perioperative surgical site infection after minimally invasive versus open posterior/transforaminal lumbar interbody fusion: analysis of hospital billing and discharge data from 5170 patients. *J Neurosurg Spine.* 2011;14(6):771–778.
- Park Y, Ha JW. Comparison of one-level posterior lumbar interbody fusion performed with a minimally invasive approach or a traditional open approach. *Spine.* 2007;32(5):537–543.
- Kepler CK, Yu AL, Gruskay JA, et al. Comparison of open and minimally invasive techniques for posterior lumbar instrumentation and fusion after open anterior lumbar interbody fusion. *Spine J.* 2013;13(5):489–497.
- Radcliff KE, Neusner AD, Millhouse PW, et al. What is new in the diagnosis and prevention of spine surgical site infections. *Spine J.* 2015;15(2):336–347.
- Rechtine GR, Bono PL, Cahill D, Bolesta MJ, Chrin AM. Postoperative wound infection after instrumentation of thoracic and lumbar fractures. *J Orthop Trauma.* 2001;15(8):566–569.
- Blam OG, Vaccaro AR, Vanichkachorn JS, et al. Risk factors for surgical site infection in the patient with spinal injury. *Spine.* 2003;28(13):1475–1480.
- Dubory A, Giorgi H, Walter A, et al. Surgical-site infection in spinal injury: incidence and risk factors in a prospective cohort of 518 patients. *Eur Spine J.* 2015;24(3):543–554.
- Klein JD, Hey LA, Yu CS, et al. Perioperative nutrition and postoperative complications in patients undergoing spinal surgery. *Spine.* 1996;21:2676–2682.
- Klein JD, Garfin SR. Nutritional status in the patient with spinal infection. *Orthop Clin North Am.* 1996;27:33–36.

38. Carreon LY, Puno RM, Dimar 2nd JR, et al. Perioperative complications of posterior lumbar decompression and arthrodesis in older adults. *J Bone Joint Surg Am.* 2003;85:2089–2092.
39. Patel N, Bagan B, Vadera S, et al. Obesity and spine surgery: relation to perioperative complications. *J Neurosurg Spine.* 2007;6:291–297.
40. Olsen MA, Nepple JJ, Riew KD, et al. Risk factors for surgical site infection following orthopaedic spinal operations. *J Bone Joint Surg Am.* 2008;90:62–69.
41. Mehta AI, Babu R, Karikari IO, et al. Young investigator award winner: the distribution of body mass as a significant risk factor for lumbar spinal fusion postoperative infections. *Spine.* 2012;37(19):1652–1656.
42. Mehta AI, Babu R, Sharma R, et al. Thickness of subcutaneous fat as a risk factor for infection in cervical spine fusion surgery. *J Bone Joint Surg Am.* 2013;95(4):323–328.
43. Meng F, Cao J, Meng X. Risk factors for surgical site infections following spinal surgery. *J Clin Neurosci.* 2015;22(12):1862–1866.
44. Simpson JM, Silveri CP, Balderston RA, et al. The results of operations on the lumbar spine in patients who have diabetes mellitus. *J Bone Joint Surg Am.* 1993;75:1823–1829.
45. Schuster JM, Rehtine G, Norvell DC, Dettori JR. The influence of perioperative risk factors and therapeutic interventions on infection rates after spine surgery: a systematic review. *Spine.* 2010;35(9 suppl):S125–S137.
46. Chen S, Anderson MV, Cheng WK, Wongworawat MD. Diabetes associated with increased surgical site infections in spinal arthrodesis. *Clin Orthop Relat Res.* 2009;467(7):1670–1673.
47. Hikata T, Iwanami A, Hosogane N, et al. High preoperative hemoglobin A1c is a risk factor for surgical site infection after posterior thoracic and lumbar spinal instrumentation surgery. *J Orthop Sci.* 2014;19(2):223–228.
48. Dickhaut SC, DeLee JC, Page CP. Nutritional status: importance in predicting wound-healing after amputation. *J Bone Joint Surg Am.* 1984;66:71–75.
49. Mandelbaum BR, Tolo VT, McAfee PC, et al. Nutritional deficiencies after staged anterior and posterior spinal reconstructive surgery. *Clin Orthop Relat Res.* 1988;234:5–11.
50. McPhee IB, Williams RP, Swanson CE. Factors influencing wound healing after surgery for metastatic disease of the spine. *Spine.* 1998;23:726–732.
51. Gruskay J, Kepler C, Smith J, Radcliff K, Vaccaro A. Is surgical case order associated with increased infection rate after spine surgery? *Spine.* 2012;37(13):1170–1174.
52. Thomsen T, Tonnesen H, Moller AM. Effect of preoperative smoking cessation interventions on postoperative complications and smoking cessation. *Br J Surg.* 2009;96(5):451–461.
53. Sorensen LT, Karlsmark T, Gottrup F. Abstinence from smoking reduces incisional wound infection: a randomized controlled trial. *Ann Surg.* 2003;238:1–5.
54. Omeis IA, Dhir M, Sciubba DM, et al. Postoperative surgical site infections in patients undergoing spinal tumor surgery: incidence and risk factors. *Spine.* 2011;36(17):1410–1419.
55. Ghogawala Z, Mansfield FL, Borges LF. Spinal radiation before surgical decompression adversely affects outcomes of surgery for symptomatic metastatic spinal cord compression. *Spine.* 2001;26:818–824.
56. Wimmer C, Gluch H, Franzreb M, et al. Predisposing factors for infection in spine surgery: a survey of 850 spinal procedures. *J Spinal Disord.* 1998;11:124–128.
57. Glassman SD, Dimar JR, Puno RM, et al. Salvage of instrumental lumbar fusions complicated by surgical wound infection. *Spine.* 1996;21:2163–2169.
58. Sponseller PD, LaPorte DM, Hungerford MW, et al. Deep wound infections after neuromuscular scoliosis surgery: a multicenter study of risk factors and treatment outcomes. *Spine.* 2000;25:2461–2466.
59. Clark CE, Shufflebarger HL. Late-developing infection in instrumented idiopathic scoliosis. *Spine.* 1999;24:1909–1912.
60. Muschik M, Luck W, Schlenzka D. Implant removal for late developing infection after instrumented posterior spinal fusion for scoliosis: reinstrumentation reduces loss of correction. A retrospective analysis of 45 cases. *Eur Spine J.* 2004;13:645–651.
61. Jonsson B, Soderholm R, Stromqvist B. Erythrocyte sedimentation rate after lumbar spine surgery. *Spine.* 1991;16:1049–1050.
62. Rubinstein E, Findler G, Amit P, Shaked I. Perioperative prophylactic cephazolin in spinal surgery. A double-blind placebo-controlled trial. *J Bone Joint Surg Br.* 1994;76(1):99–102.
63. PullterGunne AF, Hosman AJ, Cohen DB, Schuetz M, Habi LD, van Laarhoven CJ, et al. A methodological systematic review on surgical site infections following spinal surgery: part1: risk factors. *Spine.* 2012;37(24):2017–2033.
64. Silber JS, Anderson DG, Vaccaro AR, Anderson PA, McCormick P. Management of post procedural discitis. *Spine J.* 2002;2(4):279–287.
65. Chaudhary SB, Vives MJ, Basra SK, et al. Postoperative spinal wound infections and post procedural diskitis. *J Spinal Cord Med.* 2007;30:441–451.
66. Djukic S, Lang P, Morris J, Hoaglund F, Genant HK. The post-operative spine. Magnetic resonance imaging. *Orthop Clin North Am.* 1990;21(3):603–624.
67. Vaccaro AR, Shah SH, Schweitzer ME, Rosenfeld JF, Cotler JM. MRI description of vertebral osteomyelitis, neoplasm, and compression fracture. *Orthopedics.* 1999;22(1):67–73.
68. Dagirmanjian A, Schils J, McHenry M, et al. MR imaging of vertebral osteomyelitis revisited. *AJR Am J Roentgenol.* 1996;167:1539–1543.
69. Thomsen T, Villebro N, Moller AM. Interventions for preoperative smoking cessation. *Cochrane Database Syst Rev.* 2014;3:1–42.
70. Bode LG, Kluytmans JA, Wertheim HF, et al. Preventing surgical-site infections in nasal carriers of staphylococcus aureus. *N Engl J Med.* 2010;362(1):9–17.
71. Kim DH, Spencer M, Davidson SM, et al. Institutional pre-screening for detection and eradication of methicillin-resistant staphylococcus aureus in patients undergoing elective orthopaedic surgery. *J Bone Joint Surg Am.* 2010;92(9):1820–1826.
72. Djindjian M, Lepresle E, Homs J-B. Antibiotic prophylaxis during prolonged clean neurosurgery. *J Neurosurg.* 1990;73:383–386.
73. Bullock R, vanDellen JR, Ketelbey W, et al. A double-blind placebo-controlled trial of perioperative prophylactic antibiotics for elective neurosurgery. *J Neurosurg.* 1988;69:687–691.
74. Shaffer WO, Baisden JL, Fernand R, Matz PG. An evidence-based clinical guideline for antibiotic prophylaxis in spine surgery. *Spine J.* 2013;13(10):1387–1392.
75. Steinberg JP, Braun BI, Hellinger WC, et al. Timing of antimicrobial prophylaxis and the risk of surgical site infections: results from the trial to reduce antimicrobial prophylaxis errors. *Ann Surg.* 2009;250(1):10–16.
76. Brown EM, Pople IK, de Louvois J, et al. Spine update: prevention of postoperative infection in patients undergoing spinal surgery. *Spine.* 2004;29:938–945.
77. Mangram A, Horan T, Pearson M, et al. Guideline for prevention of surgical site infection, 1999. centers for disease control and prevention (CDC) hospital infection control practices advisory committee. *Am J Infect Control.* 1999;27:97–132.
78. Classen DC, Evans RS, Pestotnik SL, et al. The timing of prophylactic administration of antibiotics and the risk of surgical-wound infection. *N Engl J Med.* 1992;326:281–286.

79. Tanner J, Parkinson H. Double gloving to reduce surgical cross-infection. *Cochrane Database Syst Rev*. 2006;(3):1–43.
80. Rehman A, Rehman AU, Rehman TU, Freeman C. Removing outer gloves as a method to reduce spinal surgery infection. *J Spinal Disord Tech*. 2015;28(6):E343–E346.
81. Kawaguchi Y, Matsui H, Gejo R, et al. Preventive measures of back muscle injury after posterior lumbar spine surgery in rats. *Spine*. 1998;23:2282–2287.
82. Datta G, Gnanalingham KK, Peterson D, et al. Back pain and disability after lumbar laminectomy: is there a relationship to muscle retraction? *Neurosurgery*. 2004;54:1413–1420.
83. Cheng MT, Chang MC, Wang ST, Yu WK, Liu CL, Chen TH. Efficacy of dilute betadine solution irrigation in the prevention of postoperative infection of spinal surgery. *Spine*. 2005;30(15):1689–1693.
84. Chiang HY, Herwaldt LA, Blevins AE, Cho E, Schweizer ML. Effectiveness of local vancomycin powder to decrease surgical site infections: a meta-analysis. *Spine J*. 2014;14(3):397–407.
85. Bakhsheshian J, Dahdaleh NS, Lam SK, Savage JW, Smith ZA. The use of vancomycin powder in modern spine surgery: systematic review and meta-analysis of the clinical evidence. *World Neurosurg*. 2015;83(5):816–823.
86. Evaniew N, Khan M, Drew B, Peterson D, Bhandari M, Ghert M. Intrawound vancomycin to prevent infections after spine surgery: a systematic review and metaanalysis. *Eur Spine J*. 2015;24(3):533–542.
87. Kang DG, Holekamp TF, Wagner SC, Lehman RA. Jr intrasite vancomycin powder for the prevention of surgical site infection in spine surgery: a systematic literature review. *Spine J*. 2015;15(4):762–770.
88. Khan NR, Thompson CJ, DeCuypere M, et al. A meta-analysis of spinal surgical site infection and vancomycin powder. *J Neurosurg Spine*. 2014;21:974–983.
89. Sweet FA, Roh M, Sliva C. Intrawound application of vancomycin for prophylaxis in instrumented thoracolumbar fusions: efficacy, drug levels, and patient outcomes. *Spine (Phila Pa 1976)*. 2011;36:2084–2088.
90. Emohare O, Ledonio CG, Hill BW, et al. Cost savings analysis of intrawound vancomycin powder in posterior spinal surgery. *Spine J*. 2014;14(11):2710–2715.
91. Strom RG, Pacione D, Kalthorn SP, Frempong-Boadu AK. Lumbar laminectomy and fusion with routine local application of vancomycin powder: decreased infection rate in instrumented and non-instrumented cases. *Clin Neurol Neurosurg*. 2013;115(9):1766–1769.
92. Armaghani SJ, Menge TJ, Lovejoy SA, Mencio GA, Martus JE. Safety of topical vancomycin for pediatric spinal deformity: nontoxic serum levels with supratherapeutic drain levels. *Spine*. 2014;39(20):1683–1687.
93. Gans I, Dormans JP, Spiegel DA, et al. Adjunctive vancomycin powder in pediatric spine surgery is safe. *Spine*. 2013;38(19):1703–1707.
94. Brown MD, Brookfield KF. A randomized study of closed wound suction drainage for extensive lumbar spine surgery. *Spine*. 2004;29(10):1066–1068.
95. Scuderi GJ, Brusovanik GV, Fitzhenry LN, Vaccaro AR. Is wound drainage necessary after lumbar spinal fusion surgery? *Med Sci Monit*. 2005;11(2):CR64–CR66.
96. Diab M, Smucny M, Dormans JP, et al. Use and outcomes of wound drain in spinal fusion for adolescent idiopathic scoliosis. *Spine*. 2012;37(11):966–973.
97. Kanayama M, Hashimoto T, Shigenobu K, et al. Effective prevention of surgical site infection using a centers for disease control and prevention guideline–based antimicrobial prophylaxis in lumbar spine surgery. *J Neurosurg Spine*. 2007;6:327–329.
98. Cahill PJ, Warnick DE, Lee MJ, et al. Infection after spinal fusion for pediatric spinal deformity: thirty years of experience at a single institution. *Spine (Phila Pa 1976)*. 2010;35:1211–1217.
99. Beiner JM, Grauer J, Kwon BK, et al. Postoperative wound infections of the spine. *Neurosurg Focus*. 2003;15:E14.
100. Maruo K, Berven SH. Outcome and treatment of postoperative spine surgical site infections: predictors of treatment success and failure. *J Orthop Sci*. 2014;19:398–404.
101. Hahn F, Zbinden R, Min K. Late implant infections caused by propionibacterium acnes in scoliosis surgery. *Eur Spine J*. 2005;14:783–788.
102. Kowalski TJ, Berbari EF, Huddleston PM, et al. The management and outcome of spinal implant infections: contemporary retrospective cohort study. *Clin Infect Dis*. 2007;44:913–920.
103. Dipaola CP, Saravanja DD, Boriani L, et al. Postoperative infection treatment score for the spine (PITSS): construction and validation of a predictive model to define need for single versus multiple irrigation and debridement for spinal surgical site infection. *Spine J*. 2012;12(3):218–230.
104. Collins I, Wilson-MacDonald J, Chami G, et al. The diagnosis and management of infection following instrumented spinal fusion. *Eur Spine J*. 2008;17:445–450.
105. de Jonge T, Stullitel H, Dubouset J, et al. Late-onset spinal deformities in children treated by laminectomy and radiation therapy for malignant tumours. *Eur Spine J*. 2005;14(8):765–771.
106. Ha KY, Kim YH. Postoperative spondylitis after posterior lumbar interbody fusion using cages. *Eur Spine J*. 2004;13:419–424.
107. Ho C, Skaggs DL, Weiss JM, et al. Management of infection after instrumented posterior spine fusion in pediatric scoliosis. *Spine*. 2007;32:2739–2744.
108. Bose B. Delayed infection after instrumented spine surgery: case reports and review of the literature. *Spine J*. 2003;3(5):394–399.
109. Wenger DR, Mubarak SJ, Leach J. Managing complications of posterior spinal instrumentation and fusion. *Clin Orthop Relat Res*. 1992;284:24–33.
110. Benli IT, Acaroglu E, Akalin S, et al. Anterior radical debridement and anterior instrumentation in tuberculosis spondylitis. *Eur Spine J*. 2003;12(2):224–234.
111. Lonstein J, Winter R, Moe J, Gaines D. Wound infection with Harrington instrumentation and spine fusion for scoliosis. *Clin Orthop Relat Res*. 1973;96:222–233.
112. Ahmed R, Greenlee JDW, Traynelis VC. Preservation of spinal instrumentation after development of postoperative bacterial infections in patients undergoing spinal arthrodesis. *J Spinal Disord Tech*. 2012;25(6):299–302.
113. Keller RB, Pappas AM. Infection after spinal fusion using internal fixation instrumentation. *Orthop Clin North Am*. 1972;3(1):99–111.
114. Picada R, Winter RB, Lonstein JE, et al. Postoperative deep wound infection in adults after posterior lumbosacral spine fusion with instrumentation: incidence and management. *J Spinal Disord*. 2000;13(1):42–45.
115. Mirovsky Y, Floman Y, Smorgick Y, et al. Management of deep wound infection after posterior lumbar interbody fusion with cages. *J Spinal Disord Tech*. 2007;20:127–131.
116. Lee MC, Wang MY, Fessler RG, et al. Instrumentation in patients with spinal infection. *Neurosurg Focus*. 2004;17(6):E7.
117. Lall RR, Wong AP, Lall RR, Lawton CD, Smith ZA, Dahdaleh NS. Evidence-based management of deep wound infection after spinal instrumentation. *J Clin Neurosci*. 2015 Feb;22(2):238–242.
118. Sheehan E, McKenna J, Mulhall KJ, Marks P, McCormack D. Adhesion of staphylococcus to orthopaedic metals, an in vivo study. *J Orthop Res*. 2004;22(1):39–43.

119. Weiss L, Vaccaro AR, Scuderi G, et al. Pseudoarthrosis after postoperative wound infection in the lumbar spine. *J Spinal Disord*. 1997;10:482–487.
120. Mericli AF, Tarola NA, Moore Jr JH, Copit SE, Fox JW, Tuma GA. Paraspinous muscle flap reconstruction of complex mid-line back wounds: risk factors and post reconstruction complications. *Ann PlastSurg*. 2010;65(2):219–224.
121. Mericli AF, Mirzabeigi MN, Moore Jr JH, Fox JW, Copit SE, Tuma GA. Reconstruction of complex posterior cervical spine wounds using the paraspinous muscle flap. *Plast Reconstr Surg*. 2011;128(1):148–153.
122. Ogilvie AA, Pinto MR, et al. Postoperative deep wound infections in adults after spinal fusion: management with vacuum assisted wound closure. *J Spinal Disord Tech*. 2005;18:14–17.
123. Aydinli U, Karaeminogullari O, Tiskaya K. Postoperative deep wound infection in instrumented spinal surgery. *Acta Orthop Belg*. 1999;65:182–187.
124. Rihn JA, Lee JY, Ward WT. Infection after the surgical treatment of adolescent idiopathic scoliosis: evaluation of the diagnosis, treatment, and impact on clinical outcomes. *Spine (Phila Pa 1976)*. 2008;33:289–294.
125. Lee R, Beder D, Street J, et al. The use of vacuum-assisted closure in spinal wound infections with or without exposed dura. *Eur Spine J*. 2018 Apr 25.
126. Ploumis A, Mehbod AA, Dressel TD, et al. Therapy of spinal wound infections using vacuum-assisted wound closure: risk factors leading to resistance to treatment. *J Spinal Disord Tech*. 2008;21(5):320–323.