

Possible involvement of PI3K/AKT/mTOR signaling pathway in the protective effect of selegiline (deprenyl) against memory impairment following ischemia reperfusion in rat



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ABSTRACT

Short-term cerebral ischemia led to memory dysfunction. There is a pressing need to introduce effective agents to reduce complications of the ischemia. Involvement of PI3K/AKT/mTOR signaling pathway has been determined in the neuroprotective effect of various agents. Selegiline (deprenyl) possessed neuroprotective properties. In this study global ischemia/reperfusion was established in rats. Selegiline (5 mg/kg for 7 consecutive days) administered via intraperitoneal route. Possible involvement of PI3K/AKT/mTOR signaling pathway was evaluated using qRT-PCR, immunohistochemistry and histopathologic evaluations in the hippocampus. Spatial memory was evaluated by morris water maze (MWM). Results showed that ischemia impaired the memory and ischemic rats spent more time to find hidden platform in the MWM. Ischemia significantly decreased levels of PI3K, AKT and mTOR in the hippocampus. Histopathologic assessment revealed that the percent of dark neurons significantly increased in the CA1 area of the hippocampus of ischemic rats. Selegiline improved the memory as ischemic rats spent fewer time to find hidden platform in the MWM. Findings showed that selegiline increased the level and expression of PI3K, AKT and mTOR as well as decreased the proportion of dark neurons in the CA1 area of the pyramidal layer of the hippocampus. We concluded that selegiline, partially at least, through increases the expression of PI3K, AKT and mTOR as well as decreases the percent of dark neurons in the hippocampus could improve the memory impairment following the ischemia in rats.

1. Introduction

Stroke is one of the most common causes of disability with high economic burden and increasing incidence in the world (Mozaffarian et al., 2016; Hirt et al., 2017; Schuhmann et al., 2017). Cut of blood flow to the brain in the ischemic stroke (IS) is associated with the brain injury (Bi et al., 2017; Jia et al., 2008). Short-term cerebral ischemia led to neuronal necrosis, apoptotic cell death, silent infarcts and cognitive decline (Ünal et al., 2001). Several clinical and preclinical studies have demonstrated that ischemic stroke led to memory dysfunction, neurodegeneration and cognition impairment (Schaapsmeeders et al., 2015; Eve et al., 2016; Silva et al., 2015; Sadelli et al., 2017). Although several agents have been introduced for treatment of stroke, little have effectiveness in this disorder (O'collins et al., 2006; Sacco et al., 2007). Today, thrombolysis is only acute treatment available apply to restore

blood flow to the ischemic area. In this regards, tissue plasminogen activator (tPA) approved for acute treatment. Unfortunately, tPA has some adverse effects including hemorrhage and also has short therapeutic time-window (Siket, 2016; Karatas et al., 2018). Indeed, evaluation and development of novel agents with high therapeutic index and protective effects on memory impairment consequence of ischemia warranted further studies.

Selegiline (deprenyl) is a selective and irreversible inhibitor of the monoamine oxidase (MAO)-B broadly administered for Parkinsonism patients (Mizuno et al., 2017; Cereda et al., 2017). It has been showed that selegiline at higher doses acts as non-selective inhibitor of MAO-A and MAO-B enzymes so is effective for treatment of major and atypical depression (Finberg and Tenne, 1982; Youdim and Weinstock, 2004; Youdim and Bakhle, 2006). In case of preclinical studies, literature demonstrated that selegiline improves motivational dysfunctions and

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also exerts antidepressant effect (Yohn et al., 2017; Contreras-Mora et al., 2018; Amiri et al., 2016). Selegiline enhances striatal dopamine concentrations and has amphetamine-like action in the brain (Lamensdorf et al., 1996; Reynolds et al., 1978; Kalász et al., 2014). It has been well-known that levels of dopamine significantly decreased in Alzheimer's disease (AD). In this concept, studies have clarified that augmentation of dopaminergic activity improve memory and learning deficit in animal model of AD (Golani et al., 2014; Okada et al., 2015; Kemppainen et al., 2015; Martorana and Koch, 2014). It has been determined that acute and chronic administration of selegiline possessed anti-apoptotic and neuroprotective effects and reduce the size of infarct area in experimental ischemia (Semkova et al., 1996; Ünal et al., 2001). An explorative study showed that L-deprenyl significantly improves cognitive tests and functional recovery in stroke patients (Bartolo et al., 2015). However, the exact mechanisms of protective effect of selegiline in IS are still unknown.

The phosphatidylinositol 3-kinase/protein kinase-B/mammalian target of rapamycin (PI3K/AKT/mTOR) signaling pathway is an important intracellular cascade controls cell proliferation, differentiation, cellular metabolism, apoptosis, cell survival and cytoskeletal restructuring (Janku et al., 2012; Polivka and Janku, 2014; Porta et al., 2014; Peltier et al., 2007). Previous studies showed that mTOR/cadherin signaling is involved in cell growth and adhesion (Wei and Wang, 2018; Jiang et al., 2018; Yin et al., 2018). It has been demonstrated that PI3K/AKT has a pivotal role in the proliferation of hippocampal neural progenitor cells (Peltier et al., 2007). Activation of PI3K/AKT cascade triggers neural stem cells proliferation consequently induced neurogenesis (Le Belle et al., 2011). The PI3K/AKT/mTOR pathway exerted neuroprotective activity in traumatic brain injury. In this regard, it has been determined that this pathway via suppression of neuronal autophagy in the hippocampus, possessed neuroprotective effects (Zhang et al., 2017). Researchers showed that activation of AKT/mTOR pathway possessed neuroprotective effects in ischemic brain injury (Huang et al., 2014). Recently, it has been well-known that activation of PI3K/AKT/mTOR pathway exerted the neuroprotective effect via decrease of oxidative stress, improvement of neurotransmission and neurogenesis in the AD induced by Amyloid- β in rat (Singh et al., 2017). However, there is currently almost no data about involvement of this signaling pathway in the protective effect of selegiline.

Since ischemia accounts for majority of strokes, it is crucial to evaluate the underlying mechanisms of cerebral ischemia. Therefore, introducing effective therapeutic targets has high importance to prevent neural damage in ischemic injuries of the brain. Considering neuroprotective effect of PI3K/AKT/mTOR signaling pathway and also above-mentioned beneficial effects of selegiline in ischemia, in the current study we aimed to evaluate the possible involvement of PI3K/AKT/mTOR pathway in advantageous effect of selegiline (L-deprenyl) in rat model of stroke.

2. Materials and methods

2.1. Animals

Forty male, two months old Sprague Dawley rats (Pasteur institute, Tehran, Iran) weighing 250–300 g were used. Animals were kept in Plexiglas boxes under standard laboratory conditions (temperature: $22 \pm 2^\circ\text{C}$, humidity: $50 \pm 10\%$, 12-h light–dark cycle and free access to food and water ad libitum). All procedures were performed according to the National Institutes of Health (NIH) Guide for the Care and Use of Laboratory Animals (NIH publication # 80–23) and institutional guidelines for animal care and use (Shahrekord University of Medical Sciences, Shahrekord, Iran). Each experimental group contained 10 animals.

2.2. Study design

Selegiline HCl (Sigma, St Louis, MO, USA) was dissolved in saline and injected subcutaneously (s.c.) at dose of 5 mg/kg for 7 consecutive days. Dose and duration of selegiline's administration was selected according to previous published studies (Amiri et al., 2016; Tsunekawa et al., 2018; Shimazu et al., 2005) and our pilot studies.

Of the forty rats used in this study, twenty rats were subjected to global ischemia model and twenty rats were remained intact. Rats were divided into four groups as follows: 1) Control group without surgery received saline 2) rats which were underwent ischemia reperfusion model and received saline 3) ischemic reperfusion rats received selegiline and 4) rats which were underwent ischemia reperfusion model and received selegiline.

Rats were treated with saline or selegiline for 7 days (days 0–7) and then were subjected to water maze test for evaluation of memory. After memory assessment, rats were euthanized under anesthesia using pentobarbital (60 mg/kg, i.p.) and hippocampi were dissected out and histopathological changes in the CA1 area as well as expression of PI3K, AKT and mTOR genes were evaluated in the hippocampus using RT-PCR method. In addition, the level of PI3K, AKT, mTOR and p-mTOR (phosphorylated mTOR) was evaluated by immunohistochemistry method.

2.3. Global ischemia/reperfusion model establishment

Transient global ischemia was induced according to the previously described method (Li et al., 2006; Cao et al., 2011). For short, anesthesia was induced by intraperitoneal administration of ketamine (60 mg/kg) and xylazine (6 mg/kg). The bilateral common carotid arteries were exposed through a 2 cm ventral midline cervical opening and carefully detached from the vagus nerves, then obstructed bilaterally for 5 min using clip. Five minutes later, the clips were removed to restore cerebral blood flow and reperfusion. Animals were recovered on a heating pad for 2 h to protect from hypothermia. Also, full efforts were made to minimize the use of animals and to optimize their comfort.

2.4. Morris water maze test (MWM)

MWM is a valid device to evaluate spatial memory in rodents. The apparatus is a round black-painted tank (150 cm diameter and 60 cm deep) which filled with water ($20 \pm 2^\circ\text{C}$) to a depth of 30 cm. Several distal visual objects were placed on the walls of the MWM room and their location stayed unchanged during the tests. The maze was divided into four quadrants with four starting locations called north (N), east (E), south (S), and west (W) at same distances to the border. A Plexiglas escape circular platform (10 cm in diameter) was kept 1 cm beneath the surface of the water in the center of the north-west quadrant (target quadrant). Throughout the tests, the animal motion was recorded by a camera located above the maze which was connected to a computer. A videotracking system (Etho-Vision XT[®] v 8.5; Noldus Information Technology, Wageningen, the Netherlands) was used to record the time spent to find the hidden platform (escape latency) and also path length to reach the hidden platform (traveled distance). To do this experiment, rats were trained in the MWM. For this purpose, each rat was allowed to swim during 60 s to discover the hidden platform directed by distal spatial indications.

Subsequently finding the platform, animals were permitted to stay there for 20 s, and were then placed in a cage for 20 s till the start of the next trial. If an animal did not find the platform within this period, it was manually guided to the platform by the experimenter and allowed to rest for 20 s. Escape latency was recorded in each trial for evaluation of spatial memory. Probe trial (retrieval test session) was performed 24 h afterward training. The probe trial was involved a 60-s free swimming period without a platform and escape latency as well as

Table 1
Primer sequences for qRT-PCR.

Primer name	Forward sequence	Reverse sequence
AKT	TAGCCATTGTGAAGGAGGGC	CCTGAGGCCGTTCTTGTAG
mTOR	GCTCCAGCACTATGTACCA	CGTCTGAGCTGGAAACCAGT
PI3K	GCAACTCCTGGACTGCAACT	CAGCGCACTGTCATGGTATG
B2m	CGTGATCTTTCTGGTGCTTGTG	GGAAAGTTGGGCTTCCCATCT

traveled distance were recorded (Amiri et al., 2016; Vorhees and Williams, 2006).

2.5. Quantitative reverse transcription-PCR (qRT-PCR)

Total RNA was extracted using Tripure isolation reagent (Roche) according to the manufacturer's instructions and quantified by a ND-100 spectrophotometer (Nanodrop Technologies). Variations in mRNA expression of looked-for genes were assessed by qRT-PCR after reverse transcription of 1 µg RNA from each sample with PrimeScript RT reagent kit (Takara) according to the manufacturer's order. The qRT-PCR was done on a light cycler apparatus (Roche Diagnostics) using SYBR Premix Ex Taq technology (Takara). Thermal cycling environment involved an initial activation phase for 30 s at 95 °C followed by 45 cycles including a denaturation step for 5 s at 95 °C and a combined annealing/extension step for 20 s at 60 °C. Beta 2-Microglobulin was considered as a normalizer and fold changes in expression of each target mRNA relative to beta 2-Microglobulin (B2m) was calculated based on $2^{-\Delta\Delta Ct}$ relative expression formula as described earlier (Haj-Mirzaian et al., 2017; Amini-Khoei et al., 2017; Lorigooini et al., 2019). The primer sequences are listed in Table 1.

2.6. Immunohistochemistry

Immunohistochemical staining was applied using the streptavidin biotin peroxidase-complex method according to our previous protocol (Sabzevary-Ghahfarokhi et al., 2018). AKT, PI3K, mTOR and p-mTOR antibodies were purchased from the Cell signaling company (Cell signaling technology, USA). In brief, hippocampi were cut into 4-µm thick sections and stuck on poly-L-lysine slides. The slides were deparaffinized and rehydrated using xylene and a series of ethanols (100%, 100%, 80% and 70%). In order to do antigen retrieving stage, sections were wrapped up in citrate buffer solution (10 mM Sodium Citrate, 0.05% Tween 20, pH 6.0) and were exposed to pressure for 20 min. To avoid nonspecific staining, slides were incubated for 2 h with protein block (Abcam, England) containing albumin. Primary antibodies were incubated overnight at 4 °C which was followed by adding 0.3% H₂O₂ solved in TBS to inhibit endogenous peroxidase activity. Following incubating with biotinylated IgG antibody and Streptavidin-Peroxidase Plus at room temperature, 3-diaminobenzidine tetrahydrochloride DAB was used to visualize specific antigen. Finally, Sections were counterstained with hematoxylin and washed with cool water. Intensity of immunoreactivities against primary antibodies were inspected on all sections using a light microscope (Olympus BX41) by a pathologist blind to the study using a 6-score system (0 = negative), 0.5 = 0–5% positive, 1 = 5–15% positive, 2 = 16–40% positive, 3 = 41–90% positive, and 5 > 90% positive.

2.7. Microscopy

After euthanasia under anesthesia using pentobarbital (60 mg/kg, i.p.), trans-cardiac perfusion was performed via 0.9% normal saline first and then continued with ice-cold 4% paraformaldehyde in 0.1M phosphate buffer (pH 7.5). Then, the hippocampi were isolated and after fixation samples were immersed in 10% formalin. Formalin-fixed brains were paraffin-embedded and 5 µm sections were obtained.

Five sections obtained from each brain and were deparaffinized using xylene and stained with H&E. Histological analysis was performed under light microscopy (400; Olympus microscope) after preparing images under objective lens using a digital camera (Olympus, Japan) and exhibited on a computer monitor. Three fields from each slide were selected and the compactness of dark neurons and normal neurons within the pyramidal cell layer of CA1 area was estimated in each field. In histological studies dark neurons are recognized by hyperbasophilia property as a type of cell degeneration. The percent of dark (dead) neurons (the relation of dark neurons to normal neurons + dark neurons (total number of neurons)) was evaluated in each group. The fields were randomly selected. All measurements were performed using Image J software by a blinded pathologist (Zsombok et al., 2005; Amini-Khoei et al., 2017).

2.8. Statistical analysis

Comparison between the groups was analyzed using two-way ANOVA followed by tukey's post test. Graph-pad prism software (version 6) was used for data analysis. $P < 0.05$ was considered statistically significant.

3. Results

3.1. Selegiline improved the memory function in the Morris water maze swimming test

Two-way ANOVA analysis showed that ischemic (IS) rats significantly spent fewer time in the zone1 of the apparatus in compared with control (CO) rats in the probe trail (on fifth day of test) ($P < 0.001$, Fig. 1A). Results demonstrated that following treatment with selegiline, time spent in the correct quadrant (zone1) significantly increased in the IS rats ($P < 0.001$). In case of spatial memory assessments (Fig. 1B), ischemic rats spent more time to find the escape platform in training days in comparison with control rats (training days 1 and 3 $P < 0.01$, training days 2 and 4 $P < 0.05$). Our findings showed that treatment of ischemic rats with selegiline significantly reduced the latency time to find the hidden platform in compared with saline-treated IS rats ($P < 0.05$ in training day 1 and 4).

3.2. Selegiline decreased the dead neurons (%) of the CA1 region

The percentage of dead neurons (damaged cells with sparsely arrange and fuzzy shape) were calculated in the CA1 region of the hippocampus (Fig. 2A). The mean percentage of dead neurons in the ischemic (IS) rats was significantly higher than those in the control (CO) rats ($P < 0.001$, Fig. 2B). A significant decrease was recorded in the mean percentage of dead neurons in the selegiline-treated IS (IS+SE) rats ($P < 0.01$) in compared with the IS group.

3.3. Selegiline increased the level of AKT, PI3K, mTOR and p-mTOR in the hippocampus

As summarized in Table 2 and showed in Fig. 3 (AKT), Fig. 4 (PI3K), Fig. 5 (mTOR) and Fig. 6 (p-mTOR) ischemia hypoperfusion significantly decreased the expression of AKT, PI3K, mTOR and p-mTOR (phosphorylated mTOR) in the hippocampus in compared to the control group. Treatment with selegiline in the IS rats significantly increased the expression of AKT, PI3K, mTOR and pmTOR when compared with the saline-treated IS animals.

3.4. Selegiline increased the gene expression of AKT, PI3K and mTOR in the hippocampus

As shown in Fig. 7, expression of AKT (A), PI3K (B) and mTOR (C) was significantly decreased in the IS group in comparison with the

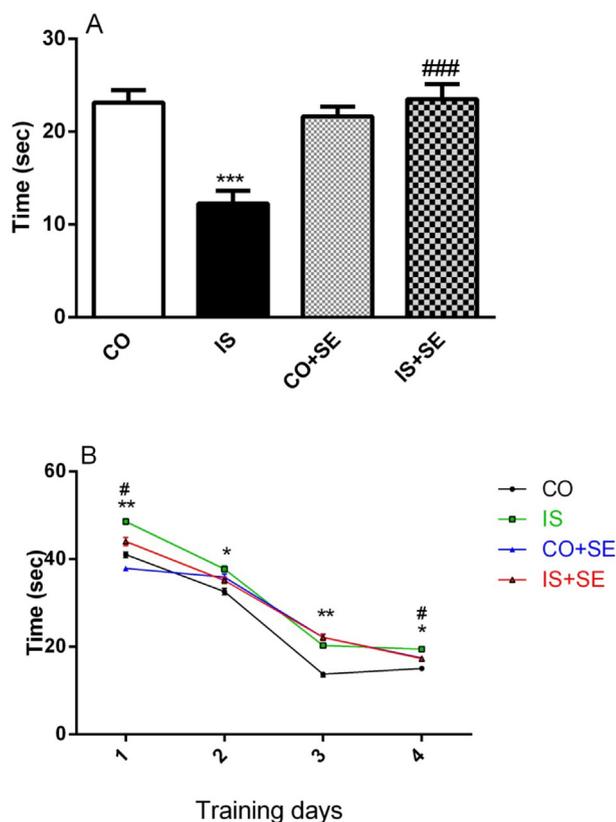


Fig. 1. (A): Spent time in zone1 through the probe trial in experimental groups. Data are presented as mean \pm SD ($n = 7$). *** $p < 0.001$ compared with control group, ### $p < 0.001$ compared with saline-treated ischemic rats. (B): Spatial learning in hidden platform in the MWM through four training days. Data are presented as mean \pm SD ($n = 10$). * $p < 0.05$ and ** $p < 0.01$ compared with control group, # $p < 0.05$ compared with saline-treated ischemic rats. CO (saline-treated control rat), IS (saline-treated ischemic rat), CO + SE (selegiline-treated control rat) and IS + SE (selegiline-treated ischemic rat).

control group ($P < 0.05$, $P < 0.05$ and $P < 0.001$, respectively). Furthermore, treatment with selegiline in IS rats significantly increased expression of AKT ($P < 0.01$), PI3K ($P < 0.05$) and mTOR ($P < 0.05$) in the hippocampus when compared with saline-treated IS rats.

4. Discussion

Results of the present study showed that global ischemia-reperfusion led to memory impairment status in the morris water maze test. We found that this status is accompanied with low expression of PI3K/AKT/mTOR signaling pathway at gene and protein levels as well as histopathological alterations in the hippocampus. Our findings demonstrated that treatment with selegiline reversed memory impairment following global ischemia-reperfusion model. Interestingly, this constructive behavioral effect was relevant with over expression of PI3K, AKT and mTOR as well as modification in histology of the hippocampus.

Stroke is a disabling disease with high incidence through the world which accounts for > 150 million deaths annually (Tang et al., 2012; Li et al., 2015). It has been demonstrated that cerebral ischemia is the major cause of strokes (Urban et al., 2010). Ischemic stroke is developed when cerebral blood vessel (s) is (are) occluded. Following ischemia neurons enter the apoptotic stage, initiate inflammatory responses, cell death and finally loss of brain's function is formed (Urnuksaikhani et al., 2017; Chen et al., 2008). Following obstruction of vessels neurons encounter with an oxidative stress which result in mitochondrial dysfunction, activation of caspase family and then DNA

fragmentation lastly lead to ischemic infarction in the brain (Uzar et al., 2012). Previous evaluations have been determined that subsequent of ischemia, during a period of hours or days neurons have potential to recover from injury (Moskowitz et al., 2010; Ginsberg, 2008). While there are accessible drugs for treatment of ischemic stroke but the lack of an effective treatment is felt. In addition there is no specific agent to expand functional recovery of neurons in ischemic zone. Hence, introducing a neuroprotective agent with ability for prevention of neuronal death and accelerating of recovery is needed (Ginsberg Ginsberg, 2009).

It has been demonstrated that activation of the PI3K/AKT/mTOR is important for cell proliferation and apoptosis (Annovazzi et al., 2009). Ample evidences have showed that the PI3K/AKT signaling pathway has critical role in intermediating survival signals in neurons. In this regards, it has been well-known that AKT has an anti-apoptotic role (Datta et al., 1999; Zhao et al., 2006). Beneficial effects of activation of the PI3K/AKT in neuroprotection consequence of ischemic stroke have been determined. In this concept literature said that this pathway through suppression of inflammatory response, decrease of vascular permeability and improve vascular function possessed protective effects. mTOR optimized cytotrophy, energy resource, stimulates protein synthesis and angiogenesis (Xu et al., 2008; Schabbaauer et al., 2004). Considering the neuroprotective role of the PI3K/AKT/mTOR pathway in ischemic stroke we showed that this survival pathway inactivated subsequent of cerebral ischemia injury. In line with previous studies we showed that expression of PI3K/AKT/mTOR signals decreased in the hippocampus specimens of rats were subjected to ischemic stroke (Li et al., 2015). Evidences showed that activation of AKT pathway lead to activation of NF- κ B transactivation resulting in initiation of transcription of survival genes such as Bcl-xL and also stimulation of trophic factors (Hussain et al., 2012; Wu et al., 2015).

Evidences demonstrated that activation of the PI3K/AKT pathway via suppression of JNK prevent neuronal cell death in cerebellar granule neurons (Choi et al., 2018; Shimoke et al., 1999). It has been determined that MTOR stimulates angiogenesis, neuronal regeneration, synaptic plasticity and removes neurotoxic substances which are linked with the recovery and survival of injured neurons in ischemic zone (Zhang et al., 2007; Chen et al., 2012a). In this concept it has been shown that inhibition of mTOR using rapamycin increased neuronal apoptosis following brain injury (Chen et al., 2012b). Our results showed that the AKT, PI3K, mTOR and p-mTOR level were significantly decreased in the hippocampus of ischemic rats. However, interestingly treatment with selegiline significantly increased the expression of aforementioned gene and proteins in the hippocampus of ischemic rats.

Selegiline [(-)-deprenyl] is an irreversible monoamine oxidase (MAO) type B inhibitor which increase level of dopamine in the striatum (Lamensdorf et al., 1996; Amiri et al., 2016). It has been resolute that selegiline metabolize to (-)-methamphetamine and (-)-amphetamine and in this way affects the brain functions (Reynolds et al., 1978). Furthermore, selegiline through upregulation of dopaminergic activity exerts beneficial effects in brain's functions such as memory and learning (Kesby et al., 2016; Kumar et al., 2018). It is well-known that agents which enhance dopaminergic neurotransmission increase activity of the PI3K/AKT/mTOR pathway (Emamian, 2012). Previous studies have demonstrated that neuroprotective properties of rasagiline in experimental model of focal ischemia were mediated through MAO independent inhibition (Speiser et al., 1999). In this regards, evidences showed that selegiline possessed neuroprotective effects and increased brain's resistance in response to ischemia (Kwon et al., 2004; Ünal et al., 2001).

CA1 pyramidal neurons are sensitive to ischemia and relatively high percentages of these neurons die following the hypoxia (Duszczuk et al., 2009). In case of learning and memory deficits, literature revealed that selegiline attenuated memory impairment following ischemic brain damage (Puurunen et al., 2001; Kesby et al., 2016). Our results showed that selegiline significantly improved memory impairment in ischemic

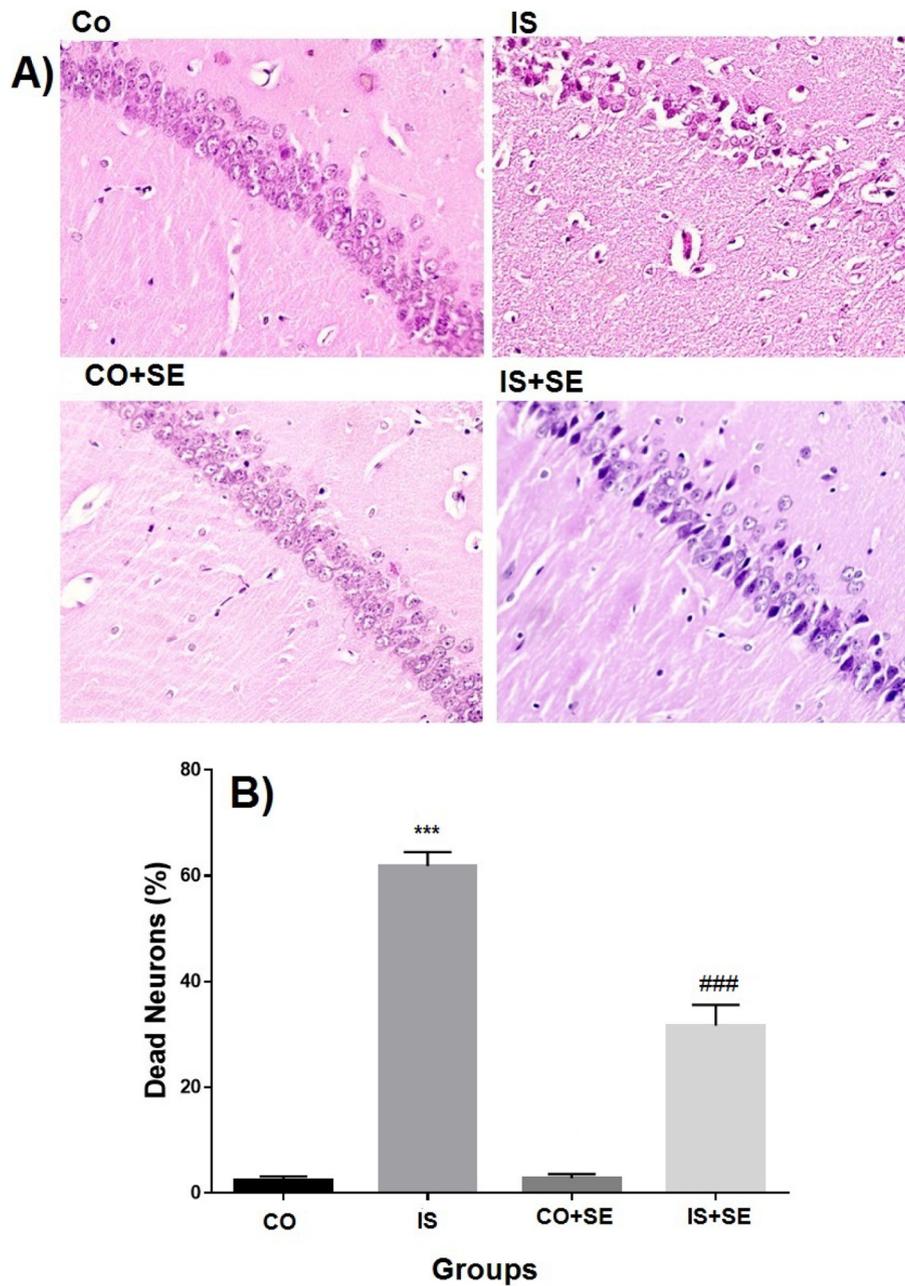


Fig. 2. The effects of treatment with selegiline on hippocampal CA1 area in hypoperfused rat, (A): Representative hematoxylin and eosin (H&E) stained slides from CA1 area ($\times 400$). (B): the percent of dead (dark) neurons in the CA1 area of the hippocampus. *** $P < 0.001$ compared with saline-treated control rats, ### $P < 0.001$ compared with the saline- treated ischemic rats.CO (saline-treated control rat), IS (saline-treated ischemic rat), CO + SE (selegiline-treated control rat) and IS + SE (selegiline-treated ischemic rat).

Table 2

Immunohistochemistry expression of AKT, PI3K, mTOR and p-mTOR in the hippocampus. The expression of AKT, PI3K, mTOR and p-mTOR were scored. Data are expressed as percent of positive cells ($n = 8$). * $P < 0.05$ compared with saline-treated control rats, # $P < 0.005$ compared with the saline- treated ischemic rats. CO (saline-treated control rat), IS (saline-treated ischemic rat), CO + SE (selegiline-treated control rat) and IS + SE (selegiline-treated ischemic rat).

P-mTOR	mTOR	PIK3	AKT	Groups
14%	24%	15%	42%	CO
10%*	7%*	11%*	5% > *	IS
16%	16%	19%	38%	CO + SE
14.3%#	18.75%#	18%#	23%#	IS + SE

rats. Furthermore, following treatment with selegiline the percent of dark neurons in the CA1 area of the hippocampus significantly decreased in ischemic rats. Clinical investigations have been clarified that selegiline has therapeutic effects in Neurological diseases including Alzheimer's disease and improves cognitive impairment (Sano et al., 1997; Ebadi et al., 2006). According to experimental studies administration of selegiline enhanced the survival and density of pyramidal neurons of the hippocampus including CA1 and CA3 cells as well as decreased the proportion of dark neurons in pyramidal area (Paterson et al., 1997; Lahtinen et al., 1997).

There are evidences revealed that hippocampus is a critical area for processing of memory (Danielson et al., 2016; Garthe et al., 2016). In this regards it has been determined that expansion of connectivity and plasticity of pyramidal cell especially CA1 cells improved memory and

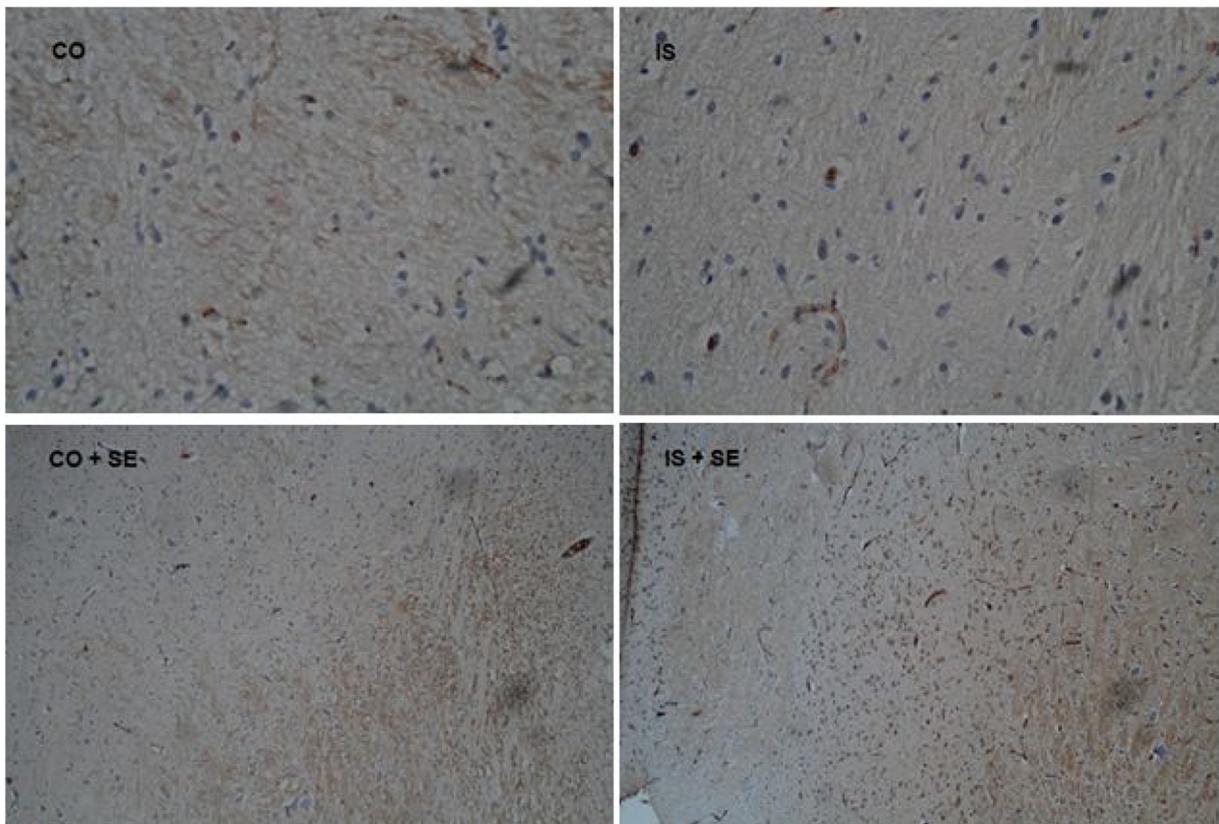


Fig. 3. The immunohistochemical features of AKT expression in the hippocampus ($\times 400$). CO (saline-treated control rat), IS (saline-treated ischemic rat), CO + SE (selegiline-treated control rat) and IS + SE (selegiline-treated ischemic rat).

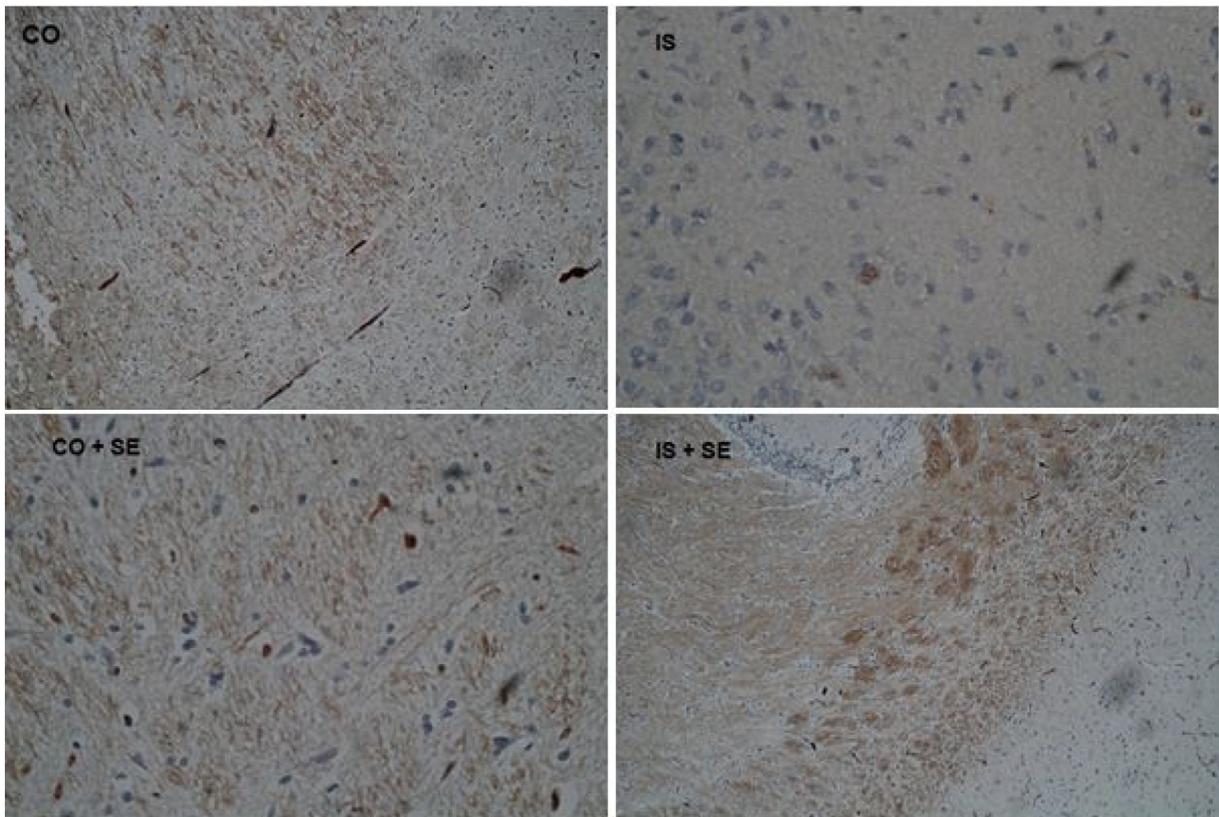


Fig. 4. The immunohistochemical features of PI3K expression in the hippocampus ($\times 400$). CO (saline-treated control rat), IS (saline-treated ischemic rat), CO + SE (selegiline-treated control rat) and IS + SE (selegiline-treated ischemic rat).

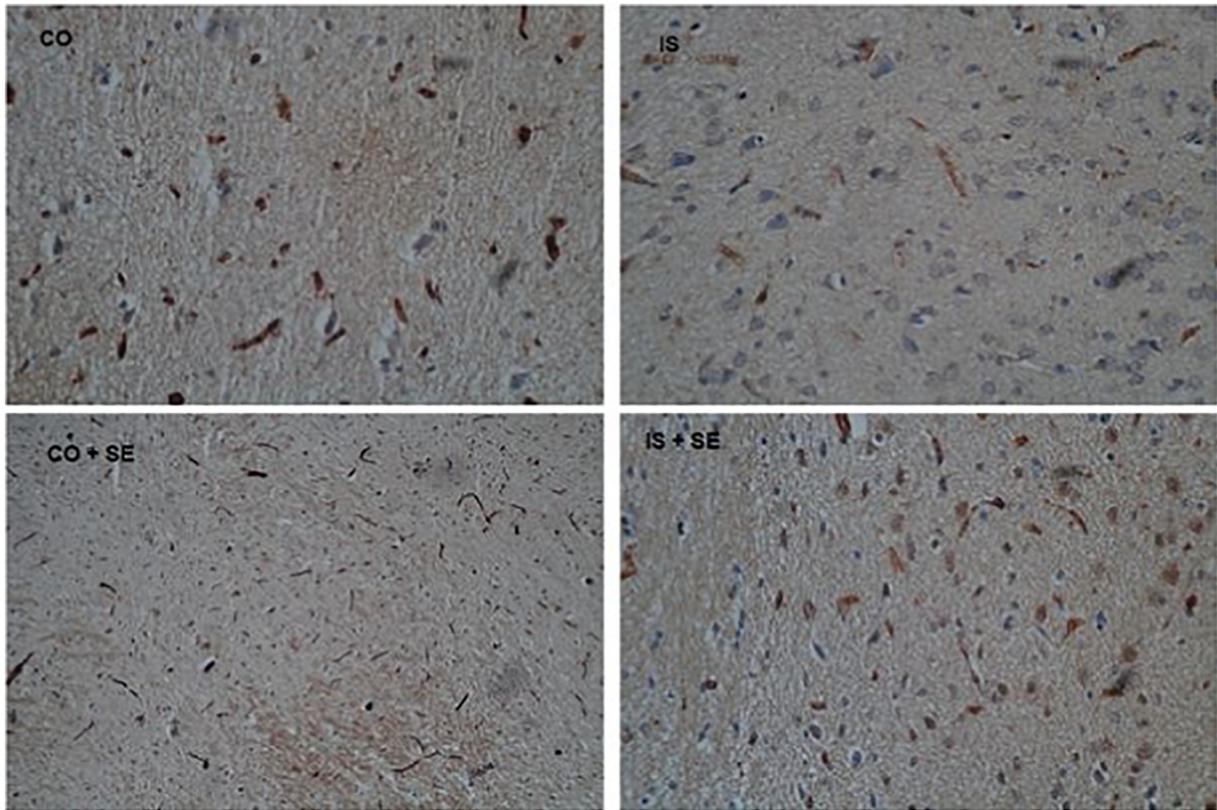


Fig. 5. The immunohistochemical features of mTOR expression in the hippocampus ($\times 400$). CO (saline-treated control rat), IS (saline-treated ischemic rat), CO + SE (selegiline-treated control rat) and IS + SE (selegiline-treated ischemic rat).

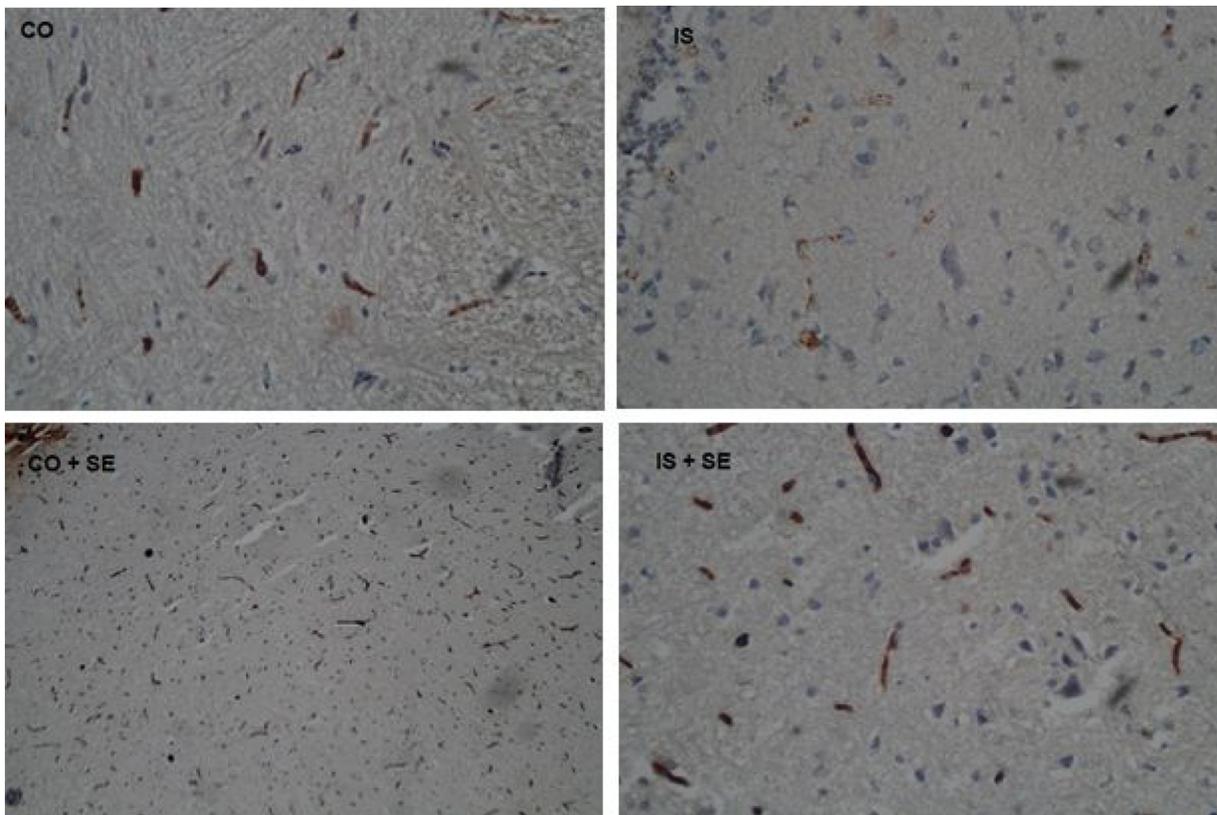


Fig. 6. The immunohistochemical features of p-mTOR expression in the hippocampus ($\times 400$). CO (saline-treated control rat), IS (saline-treated ischemic rat), CO + SE (selegiline-treated control rat) and IS + SE (selegiline-treated ischemic rat).

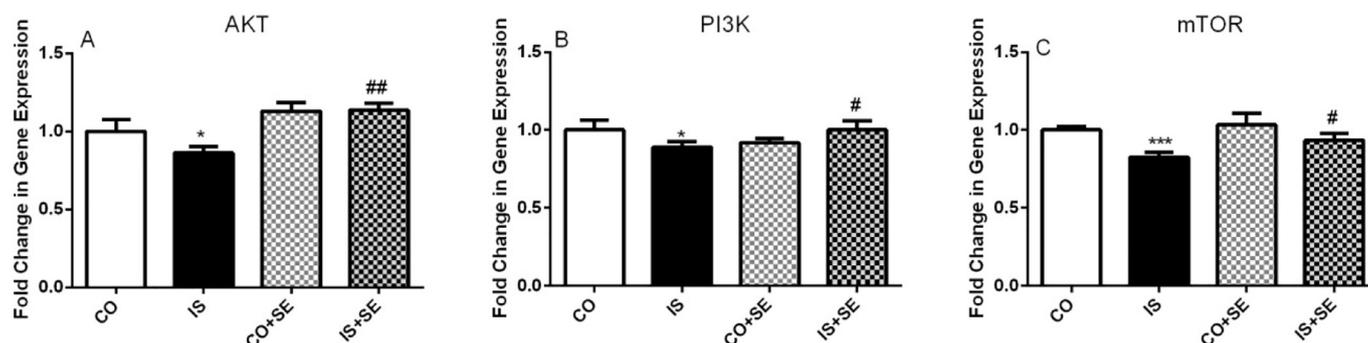


Fig. 7. The expression of AKT (A), PI3K (B) and mTOR (C) in the hippocampus was determined by qRT-PCR. Data are shown as mean \pm SEM from triplicate tests and were analyzed using two-way ANOVA followed by Tukey's post-hoc test. * $P < 0.05$ and *** $P < 0.001$ compared with saline-treated control rats, # $P < 0.05$ and ## $P < 0.01$ compared with the saline-treated ischemic rats. CO (saline-treated control rat), IS (saline-treated ischemic rat), CO + SE (selegiline-treated control rat) and IS + SE (selegiline-treated ischemic rat).

learning deficits following hippocampal injury (Danielson et al., 2016; Stackman Jr et al., 2016; Havekes et al., 2016; Hansen et al., 2015).

Morris water maze as a valid behavioral test performed for evaluation of memory and learning in rodents (Wang et al., 2017). In line with previous studies we found that rats were subjected to ischemic stroke model showed memory impairment in this hippocampal-related behavioral test (Wang et al., 2017; Fan et al., 2015). Our results showed that treatment with selegiline significantly improved memory deficit in ischemic rats.

5. Conclusion

Findings of this *in vivo* ischemia study showed that activation of the PI3K/AKT/mTOR pathway partially, at least, has critical role in reversing the adverse impacts of ischemic model of stroke in rat. Interestingly our results showed that selegiline probably, at part, through upregulation of PI3K/AKT/mTOR in the hippocampus improves memory following ischemia in rat.

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Conflict of interests

The authors declare that there is no conflict of interest.

Compliance with ethical standards

All applicable international and institutional guidelines for the care and use of animals were followed.

References

Amini-Khoei, H., Amiri, S., Mohammadi-Asl, A., Alijanpour, S., Poursaman, S., Haj-Mirzaian, A., Rastegar, M., Mesdaghinia, A., Banafshe, H.R., Sadeghi, E., 2017. Experiencing neonatal maternal separation increased pain sensitivity in adult male mice: involvement of oxytocinergic system. *Neuropeptides* 61, 77–85.

Amiri, S., Amini-Khoei, H., Mohammadi-Asl, A., Alijanpour, S., Haj-Mirzaian, A., Rahimi-Balaei, M., Razmi, A., Olson, C.O., Rastegar, M., Mehdizadeh, M., 2016. Involvement of D1 and D2 dopamine receptors in the antidepressant-like effects of selegiline in maternal separation model of mouse. *Physiol. Behav.* 163, 107–114.

Annovazzi, L., Mellai, M., Caldera, V., Valente, G., Tessitore, L., Schiffer, D., 2009. mTOR, S6 and AKT expression in relation to proliferation and apoptosis/autophagy in glioma. *Anticancer Res.* 29, 3087–3094.

Bartolo, M., Zucchella, C., Capone, A., Sandrini, G., Pierelli, F., 2015. An explorative study regarding the effect of l-deprenyl on cognitive and functional recovery in patients after stroke. *J. Neurol. Sci.* 349, 117–123.

Bi, M., Gladbach, A., Eersel, J., Ittner, A., Przybyla, M., Hummel, A., Chua, S.W., Van Der Hoven, J., Lee, W.S., Muller, J., 2017. Tau exacerbates excitotoxic brain damage in an animal model of stroke. *Nat. Commun.* 8, 473.

Cao, Y., MAO, X., SUN, C., ZHENG, P., GAO, J., WANG, X., MIN, D., SUN, H., XIE, N., CAI, J., 2011. Baicalin attenuates global cerebral ischemia/reperfusion injury in gerbils via anti-oxidative and anti-apoptotic pathways. *Brain Res. Bull.* 85, 396–402.

Cereda, E., Cilia, R., Canesi, M., Tesi, S., Mariani, C.B., Zecchinelli, A.L., Pezzoli, G., 2017. Efficacy of rasagiline and selegiline in Parkinson's disease: a head-to-head 3-year retrospective case-control study. *J. Neurol.* 1–10.

Chen, G., Frøkiær, J., Pedersen, M., Nielsen, S., Si, Z., Pang, Q., Stødkilde-Jørgensen, H., 2008. Reduction of ischemic stroke in rat brain by alpha melanocyte stimulating hormone. *Neuropeptides* 42, 331–338.

Chen, H., Qu, Y., Tang, B., Xiong, T., Mu, D., 2012a. Role of Mammalian Target of Rapamycin in Hypoxic or Ischemic Brain Injury: Potential Neuroprotection and Limitations.

Chen, H., Xiong, T., Qu, Y., Zhao, F., Ferrero, D., Mu, D., 2012b. mTOR activates hypoxia-inducible factor-1 α and inhibits neuronal apoptosis in the developing rat brain during the early phase after hypoxia-ischemia. *Neurosci. Lett.* 507, 118–123.

Choi, H.-W., Shin, P.-G., Lee, J.-H., Choi, W.-S., Kang, M.-J., Kong, W.-S., Oh, M.-J., Seo, Y.-B., Kim, G.-D., 2018. Anti-inflammatory effect of lovastatin is mediated via the modulation of NF- κ B and inhibition of HDAC1 and the PI3K/Akt/mTOR pathway in RAW264.7 macrophages. *Int. J. Mol. Med.* 41, 1103–1109.

Contreras-Mora, H., Rowland, M.A., Yohn, S.E., Correa, M., Salamone, J.D., 2018. Partial reversal of the effort-related motivational effects of tetrabenazine with the MAO-B inhibitor deprenyl (selegiline): implications for treating motivational dysfunctions. *Pharmacol. Biochem. Behav.* 166, 13–20.

Danielson, N.B., Zaremba, J.D., Kaifosh, P., Bowler, J., Ladow, M., Losonczy, A., 2016. Sublayer-specific coding dynamics during spatial navigation and learning in hippocampal area CA1. *Neuron* 91, 652–665.

Datta, S.R., Brunet, A., Greenberg, M.E., 1999. Cellular survival: a play in three Akts. *Genes Dev.* 13, 2905–2927.

Duszczyk, M., Ziembowicz, A., Gadamski, R., Wieronska, J.M., Smialowska, M., Lazarewicz, J.W., 2009. Changes in the NPY immunoreactivity in gerbil hippocampus after hypoxic and ischemic preconditioning. *Neuropeptides* 43, 31–39.

Ebadi, M., Brown-Borg, H., Ren, J., Sharma, S., Shavali, S., El Refaey, H., Carlson, E.C., 2006. Therapeutic efficacy of selegiline in neurodegenerative disorders and neurological diseases. *Curr. Drug Targets* 7, 1513–1529.

Emamian, E., 2012. AKT/GSK3 signaling pathway and schizophrenia. *Front. Mol. Neurosci.* 5, 33.

Eve, M., O'keefe, F., Jhuty, S., Ganesan, V., Brown, G., Murphy, T., 2016. Computerized working-memory training for children following arterial ischemic stroke: A pilot study with long-term follow-up. *Appl. Neuropsychol. Child* 5, 273–282.

Fan, M., Jin, W., Zhao, H., Xiao, Y., Jia, Y., Yin, Y., Jiang, X., Xu, J., Meng, N., Lv, P., 2015. Lithium chloride administration prevents spatial learning and memory impairment in repeated cerebral ischemia-reperfusion mice by depressing apoptosis and increasing BDNF expression in hippocampus. *Behav. Brain Res.* 291, 399–406.

Finberg, J., Tenne, M., 1982. Relationship between tyramine potentiation and selective inhibition of monoamine oxidase types A and B in the rat vas deferens. *Br. J. Pharmacol.* 77, 13–21.

Garthe, A., Roeder, I., Kempermann, G., 2016. Mice in an enriched environment learn more flexibly because of adult hippocampal neurogenesis. *Hippocampus* 26, 261–271.

Ginsberg, M.D., 2008. Neuroprotection for ischemic stroke: past, present and future. *Neuropharmacology* 55, 363–389.

Ginsberg, M.D., 2009. Current status of neuroprotection for cerebral ischemia: synaptic overview. *Stroke* 40, S111–S114.

Golani, I., Tadmor, H., Buonanno, A., Kremer, I., Shamir, A., 2014. Disruption of the ErbB signaling in adolescence increases striatal dopamine levels and affects learning and hedonic-like behavior in the adult mouse. *Eur. Neuropsychopharmacol.* 24, 1808–1818.

Haj-Mirzaian, A., Amiri, S., Amini-Khoei, H., Hosseini, M.-J., Haj-Mirzaian, A., Momeny, M., Rahimi-Balaei, M., Dehpour, A.R., 2017. Anxiety-and depressive-like behaviors are associated with altered hippocampal energy and inflammatory status in a mouse

- model of Crohn's disease. *Neuroscience* 366, 124–137.
- Hansen, H.H., Fabricius, K., Barkholt, P., Niehoff, M.L., Morley, J.E., Jelsing, J., Pyke, C., Knudsen, L.B., Farr, S.A., Vrang, N., 2015. The GLP-1 receptor agonist liraglutide improves memory function and increases hippocampal CA1 neuronal numbers in a senescence-accelerated mouse model of Alzheimer's disease. *J. Alzheimers Dis.* 46, 877–888.
- Havékes, R., Park, A.J., Tudor, J.C., Luczak, V.G., Hansen, R.T., Ferri, S.L., Bruinenberg, V.M., Poplawski, S.G., Day, J.P., Aton, S.J., 2016. Sleep deprivation causes memory deficits by negatively impacting neuronal connectivity in hippocampal area CA1. *Elife* 5, e13424.
- Hirt, L., Fukuda, A.M., Ambadipudi, K., Rashid, F., Binder, D., Verkman, A., Ashwal, S., Obenaus, A., Badaut, J., 2017. Improved long-term outcome after transient cerebral ischemia in aquaporin-4 knockout mice. *J. Cereb. Blood Flow Metab.* 37, 277–290.
- Huang, H., Zhong, R., Xia, Z., Song, J., Feng, L., 2014. Neuroprotective effects of rhynchophylline against ischemic brain injury via regulation of the Akt/mTOR and TLRs signaling pathways. *Molecules* 19, 11196–11210.
- Hussain, A.R., Ahmed, S.O., Ahmed, M., Khan, O.S., Al Abdulmohsen, S., Platanias, L.C., Al-Kuraya, K.S., Uddin, S., 2012. Cross-talk between NF κ B and the PI3-kinase/AKT pathway can be targeted in primary effusion lymphoma (PEL) cell lines for efficient apoptosis. *PLoS ONE* 7, e39945.
- Janku, F., Wheler, J.J., Westin, S.N., Moulder, S.L., Naing, A., Tsimberidou, A.M., Fu, S., Falchook, G.S., Hong, D.S., Garrido-Laguna, I., 2012. PI3K/AKT/mTOR inhibitors in patients with breast and gynecologic malignancies harboring PIK3CA mutations. *J. Clin. Oncol.* 30, 777–782.
- Jia, J., Chen, X., Zhu, W., Luo, Y., Hua, Z., Xu, Y., 2008. CART protects brain from damage through ERK activation in ischemic stroke. *Neuropeptides* 42, 653–661.
- Jiang, H., Chen, Y., Chen, G., Tian, X., Tang, J., Luo, L., Huang, M., Yan, B., Ao, X., Zhou, W., Wang, L., 2018. Leptin accelerates the pathogenesis of heterotopic ossification in rat tendon tissues via mTORC1 signaling. *J. Cell. Physiol.* 233, 1017–1028.
- Kalász, H., Magyar, K., Szoke, É., Adeghate, E., Adem, A., Hasan, M., Nurulain, S., Takes, K., 2014. Metabolism of selegiline [(–)-(deprenyl)]. *Curr. Med. Chem.* 21, 1522–1530.
- Karatas, H., Jung, J.E., Lo, E., Van Leyen, K., 2018. Inhibiting 12/15-lipoxygenase to treat acute stroke in permanent and tPA induced thrombolysis models. *Brain Res.* 1678, 123–128.
- Kemppainen, S., Lindholm, P., Galli, E., Lahtinen, H.-M., Koivisto, H., Hämäläinen, E., Saarna, M., Tanila, H., 2015. Cerebral dopamine neurotrophic factor improves long-term memory in APP/PS1 transgenic mice modeling Alzheimer's disease as well as in wild-type mice. *Behav. Brain Res.* 291, 1–11.
- Kesby, J.P., Markou, A., Semenova, S., Group, T., 2016. Effects of HIV/TAT protein expression and chronic selegiline treatment on spatial memory, reversal learning and neurotransmitter levels in mice. *Behav. Brain Res.* 311, 131–140.
- Kumar, S., Dang, S., Nigam, K., Ali, J., Baboota, S., 2018. Selegiline nanoformulation in attenuation of oxidative stress and upregulation of dopamine in the brain for the treatment of Parkinson's disease. *Rejuvenation Res.* 21 (5), 464–476.
- Kwon, Y., Ann, H., Nabeshima, T., Shin, E., Kim, W., Jhoo, J., Jhoo, W., Wie, M., Kim, Y., Jang, K., 2004. Selegiline potentiates the effects of EGB 761 in response to ischemic brain injury. *Neurochem. Int.* 45, 157–170.
- Lahtinen, H., Koistinaho, J., Kauppinen, R., Haapalinna, A., Keinänen, R., Sivenius, J., 1997. Selegiline treatment after transient global ischemia in gerbils enhances the survival of CA1 pyramidal cells in the hippocampus. *Brain Res.* 757, 260–267.
- Lamensdorf, I., Youdim, M.B., Finberg, J.P., 1996. Effect of long-term treatment with selective monoamine oxidase A and B inhibitors on dopamine release from rat striatum in vivo. *J. Neurochem.* 67, 1532–1539.
- LE Belle, J.E., Orozco, N.M., Paucar, A.A., Saxe, J.P., Mottahedeh, J., Pyle, A.D., Wu, H., Kornblum, H.I., 2011. Proliferative neural stem cells have high endogenous ROS levels that regulate self-renewal and neurogenesis in a PI3K/Akt-dependant manner. *Cell Stem Cell* 8, 59–71.
- Li, D.-Q., Bao, Y.-M., Li, Y., Wang, C.-F., Liu, Y., An, L.-J., 2006. Catalpol modulates the expressions of Bcl-2 and Bax and attenuates apoptosis in gerbils after ischemic injury. *Brain Res.* 1115, 179–185.
- Li, W., Yang, Y., Hu, Z., Ling, S., Fang, M., 2015. Neuroprotective effects of DAHP and Triptolide in focal cerebral ischemia via apoptosis inhibition and PI3K/Akt/mTOR pathway activation. *Front. Neuroanat.* 9, 48.
- Lorigooini, Z., Salimi, N., Soltani, A., Amini-Khoei, H., 2019. Implication of NMDA-NO pathway in the antidepressant-like effect of ellagic acid in male mice. *Neuropeptides* 3, 25–33.
- Martorana, A., Koch, G., 2014. Is Dopamine Involved in Alzheimer's Disease? *Frontiers in aging neuroscience*. pp. 6.
- Mizuno, Y., Hattori, N., Kondo, T., Nomoto, M., Origasa, H., Takahashi, R., Yamamoto, M., Yanagisawa, N., 2017. A randomized double-blind placebo-controlled phase III trial of Selegiline Monotherapy for early Parkinson disease. *Clin. Neuropharmacol.* 40, 201.
- Moskowitz, M.A., Lo, E.H., Iadecola, C., 2010. The science of stroke: mechanisms in current of treatments. *Neuron* 67, 181–198.
- Mozaffarian, D., Benjamin, E.J., Go, A.S., Arnett, D.K., Blaha, M.J., Cushman, M., Das, S.R., De Ferranti, S., Despres, J.-P., Fullerton, H.J., 2016. Heart disease and stroke statistics—2016 update: a report from the American Heart Association. *Circulation* 133, e38–e360.
- O'collins, V.E., Macleod, M.R., Donnan, G.A., Horkey, L.L., Van Der Worp, B.H., Howells, D.W., 2006. 1,026 experimental treatments in acute stroke. *Ann. Neurol.* 59, 467–477.
- Okada, R., Fujiwara, H., Mizuki, D., Araki, R., Yabe, T., Matsumoto, K., 2015. Involvement of dopaminergic and cholinergic systems in social isolation-induced deficits in social affiliation and conditional fear memory in mice. *Neuroscience* 299, 134–145.
- Paterson, I., Barber, A., Gelowitz, D., Voll, C., 1997. (–) Deprenyl reduces delayed neuronal death of hippocampal pyramidal cells. *Neurosci. Biobehav. Rev.* 21, 181–186.
- Peltier, J., O'Neill, A., Schaffer, D.V., 2007. PI3K/Akt and CREB regulate adult neural hippocampal progenitor proliferation and differentiation. *Dev. Neurobiol.* 67, 1348–1361.
- Polivka, J., Janku, F., 2014. Molecular targets for cancer therapy in the PI3K/AKT/mTOR pathway. *Pharmacol. Ther.* 142, 164–175.
- Porta, C., Paglino, C., Mosca, A., 2014. Targeting PI3K/Akt/mTOR signaling in cancer. *Front. Oncol.* 4.
- Puurunen, K., Jolkkonen, J., Sirviö, J., Haapalinna, A., Sivenius, J., 2001. Selegiline combined with enriched-environment housing attenuates spatial learning deficits following focal cerebral ischemia in rats. *Exp. Neurol.* 167, 348–355.
- Reynolds, G., Elsworth, J., Blau, K., Sandler, M., Lees, A., Stern, G., 1978. Deprenyl is metabolized to methamphetamine and amphetamine in man. *Br. J. Clin. Pharmacol.* 6, 542–544.
- Sabzevary-Ghahfarokhi, M., Shohan, M., Shirzad, H., Rahimian, G., Soltani, A., Ghatreh-Samani, M., Deris, F., Bagheri, N., Shafiq, M., Tahmasbi, K., 2018. The regulatory role of Nrf2 in antioxidants phase2 enzymes and IL-17A expression in patients with ulcerative colitis. *Pathol. Res. Pract.* 214 (8), 1149–1155.
- Sacco, R.L., Chong, J.Y., Prabhakaran, S., Elkind, M.S., 2007. Experimental treatments for acute ischaemic stroke. *Lancet* 369, 331–341.
- Sadelli, K., Stamegna, J.-C., Girard, S.D., Baril, N., Escoffier, G., Brus, M., Veron, A.D., Khrestchatsky, M., Roman, F.S., 2017. Global cerebral ischemia in rats leads to amnesia due to selective neuronal death followed by astroglial scar formation in the CA1 layer. *Neurobiol. Learn. Mem.* 141, 168–178.
- Sano, M., Ernesto, C., Thomas, R.G., Klauber, M.R., Schafer, K., Grundman, M., Woodbury, P., Growdon, J., Cotman, C.W., Pfeiffer, E., 1997. A controlled trial of selegiline, alpha-tocopherol, or both as treatment for Alzheimer's disease. *N. Engl. J. Med.* 336, 1216–1222.
- Schaapsmeeders, P., Van Uden, I.W., Tuladhar, A.M., Maaijwee, N.A., Van Dijk, E.J., Rutten-Jacobs, L.C., Arntz, R.M., Schoonderwaldt, H.C., Dorstestijn, L.D., De Leeuw, F.E., 2015. Ipsilateral hippocampal atrophy is associated with long-term memory dysfunction after ischemic stroke in young adults. *Hum. Brain Mapp.* 36, 2432–2442.
- Schabbauer, G., Tencati, M., Pedersen, B., Pawlinski, R., Mackman, N., 2004. PI3K-Akt pathway suppresses coagulation and inflammation in endotoxemic mice. *Arterioscler. Thromb. Vasc. Biol.* 24, 1963–1969.
- Schuhmann, M.K., Guthmann, J., Stoll, G., Nieswandt, B., Kraft, P., Kleinschnitz, C., 2017. Blocking of platelet glycoprotein receptor 1b reduces “thrombo-inflammation” in mice with acute ischemic stroke. *J. Neuroinflammation* 14, 18.
- Semkova, I., Wolz, P., Schilling, M., Krieglstein, J., 1996. Selegiline enhances NGF synthesis and protects central nervous system neurons from excitotoxic and ischemic damage. *Eur. J. Pharmacol.* 315, 19–30.
- Shimazu, S., Minami, A., Kusumoto, H., Yoneda, F., 2005. Antidepressant-like effects of selegiline in the forced swim test. *Eur. Neuropsychopharmacol.* 15, 563–571.
- Shimoke, K., Yamagishi, S., Yamada, M., Ikeuchi, T., Hatanaka, H., 1999. Inhibition of phosphatidylinositol 3-kinase activity elevates c-Jun N-terminal kinase activity in apoptosis of cultured cerebellar granule neurons. *Dev. Brain Res.* 112, 245–253.
- Siket, M.S., 2016. Treatment of acute ischemic stroke. *Emerg. Med. Clin.* 34, 861–882.
- Silva, B., Sousa, L., Miranda, A., Vasconcelos, A., Reis, H., Barcelos, L., Arantes, R., Teixeira, A., Rachid, M.A., 2015. Memory deficit associated with increased brain proinflammatory cytokine levels and neurodegeneration in acute ischemic stroke. *Arq. Neuropsiquiatr.* 73, 655–659.
- Singh, A.K., Kashyap, M.P., Tripathi, V.K., Singh, S., Garg, G., Rizvi, S.I., 2017. Neuroprotection through rapamycin-induced activation of autophagy and PI3K/Akt1/mTOR/CREB signaling against amyloid- β -induced oxidative stress, synaptic/neurotransmission dysfunction, and neurodegeneration in adult rats. *Mol. Neurobiol.* 54, 5815–5828.
- Speiser, Z., Mayk, A., Eliash, S., Cohen, S., 1999. Studies with rasagiline, a MAO-B inhibitor, in experimental focal ischemia in the rat. *J. Neural Transm.* 106, 593–606.
- Stackman Jr., R.W., Cohen, S.J., Lora, J.C., Rios, L.M., 2016. Temporary inactivation reveals that the CA1 region of the mouse dorsal hippocampus plays an equivalent role in the retrieval of long-term object memory and spatial memory. *Neurobiol. Learn. Mem.* 133, 118–128.
- Tang, Q., Han, R., Xiao, H., Shen, J., Luo, Q., Li, J., 2012. Neuroprotective effects of tanshinone IIA and/or tetramethylpyrazine in cerebral ischemic injury in vivo and in vitro. *Brain Res.* 1488, 81–91.
- Tsunekawa, H., Takahata, K., Okano, M., Ishikawa, T., Satoyoshi, H., Nishimura, T., Hoshino, N., Muraoka, S., 2018. Selegiline increases on time without exacerbation of dyskinesia in 6-hydroxydopamine-lesioned rats displaying l-Dopa-induced wearing-off and abnormal involuntary movements. *Behav. Brain Res.* 347, 350–359.
- Ünal, I., Gursöy-Özdemir, Y., Bolay, H., Söylemezoğlu, F., Sarıbaş, O., Dalkara, T., 2001. Chronic daily administration of selegiline and EGB 761 increases brain's resistance to ischemia in mice. *Brain Res.* 917, 174–181.
- Urban, P.P., Wolf, T., Uebele, M., Marx, J.R.J., Vogt, T., Stoeter, P., Bauermann, T., Weibrich, C., Vucurevic, G.D., Schneider, A., 2010. Occurrence and clinical predictors of spasticity after ischemic stroke. *Stroke* 41, 2016–2020.
- Urnuksaikhan, E., Mishig-Ochir, T., Kim, S.-C., Park, J.-K., Seo, Y.-K., 2017. Neuroprotective effect of low frequency-pulsed electromagnetic fields in ischemic stroke. *Appl. Biochem. Biotechnol.* 181, 1360–1371.
- Uzar, E., Alp, H., Cevik, M.U., Firat, U., Evliyaoglu, O., Tufek, A., Altun, Y., 2012. Ellagic acid attenuates oxidative stress on brain and sciatic nerve and improves histopathology of brain in streptozotocin-induced diabetic rats. *Neurol. Sci.* 33, 567–574.
- Vorhees, C.V., Williams, M.T., 2006. Morris water maze: procedures for assessing spatial and related forms of learning and memory. *Nat. Protoc.* 1, 848.
- Wang, W., Liu, L., Jiang, P., Chen, C., Zhang, T., 2017. Levodopa improves learning and

- memory ability on global cerebral ischemia-reperfusion injured rats in the Morris water maze test. *Neurosci. Lett.* 636, 233–240.
- Wei, G., Wang, L., 2018. Promotion of cell growth and adhesion of a peptide hydrogel scaffold via mTOR/cadherin signaling. *J. Cell. Physiol.* 233, 822–829.
- Wu, H., Ye, M., Yang, J., Ding, J., Yang, J., Dong, W., Wang, X., 2015. Nicorandil protects the heart from ischemia/reperfusion injury by attenuating endoplasmic reticulum response-induced apoptosis through PI3K/Akt signaling pathway. *Cell. Physiol. Biochem.* 35, 2320–2332.
- Xu, X., Chua, C.C., Gao, J., Chua, K.-W., Wang, H., Hamdy, R.C., Chua, B.H., 2008. Neuroprotective effect of humanin on cerebral ischemia/reperfusion injury is mediated by a PI3K/Akt pathway. *Brain Res.* 1227, 12–18.
- Yin, C., Zhang, Y., Hu, L., Tian, Y., Chen, Z., Li, D., Zhao, F., Su, P., Ma, X., Zhang, G., Miao, Z., Wang, L., 2018. Mechanical unloading reduces microtubule actin cross-linking factor 1 expression to inhibit beta-catenin signaling and osteoblast proliferation. *J. Cell. Physiol.* 233, 5405–5419.
- Yohn, S.E., Reynolds, S., Tripodi, G., Correa, M., Salamone, J.D., 2017. The monoamine oxidase B inhibitor deprenyl increases selection of high-effort activity in rats tested on a progressive ratio/chow feeding choice procedure: implications for treating motivational dysfunctions. *Behav. Brain Res.* 342, 27–44.
- Youdim, M.B., Bakhle, Y., 2006. Monoamine oxidase: isoforms and inhibitors in Parkinson's disease and depressive illness. *Br. J. Pharmacol.* 147.
- Youdim, M.B., Weinstock, M., 2004. Therapeutic applications of selective and non-selective inhibitors of monoamine oxidase A and B that do not cause significant tyramine potentiation. *Neurotoxicology* 25, 243–250.
- Zhang, Q.-G., Wu, D.-N., Han, D., Zhang, G.-Y., 2007. Critical role of PTEN in the coupling between PI3K/Akt and JNK1/2 signaling in ischemic brain injury. *FEBS Lett.* 581, 495–505.
- Zhang, M.-H., Zhou, X.-M., Gao, J.-L., Wang, K.-J., Cui, J.-Z., 2017. PI3K/Akt/mTOR pathway participates in neuroprotection by dexmedetomidine inhibits neuronal autophagy following traumatic brain injury in rats. *Int. J. Res. Med. Sci.* 2, 1569–1575.
- Zhao, H., Sapolsky, R.M., Steinberg, G.K., 2006. Phosphoinositide-3-kinase/akt survival signal pathways are implicated in neuronal survival after stroke. *Mol. Neurobiol.* 34, 249–269.
- Zsombok, A., Tóth, Z., Gallyas, F., 2005. Basophilia, acidophilia and argyrophilia of “dark”(compact) neurons during their formation, recovery or death in an otherwise undamaged environment. *J. Neurosci. Methods* 142, 145–152.