



Peptidase neurolysin is an endogenous cerebroprotective mechanism in acute neurodegenerative disorders

Vardan T. Karamyan*

Department of Pharmaceutical Sciences and Center for Blood Brain Barrier Research, School of Pharmacy, TTUHSC, Amarillo, TX, United States



ARTICLE INFO

Keywords:

Neuroprotection
Neuropeptide
Neurogenic inflammation
Brain edema
Drug target

ABSTRACT

Stroke and traumatic brain injury (TBI) are significant clinical problems characterized by high rate of mortality and long-lasting disabilities, and an unmet need for new treatments. Current experimental stroke and TBI research are evolving to focus more on understanding the brain's self-protective mechanisms to meet the critical need of developing new therapies for these disorders. In this hypothesis-based manuscript, I provide several lines of evidence that peptidase neurolysin (Nln) is one of the brain's potent, self-protective mechanisms promoting preservation and recovery of the brain after acute injury. Based on published experimental observations and ongoing studies in our laboratory, I posit that Nln is a compensatory and cerebroprotective mechanism in the post-stroke/TBI brain that functions to process a diverse group of extracellular neuropeptides and by that to reduce excitotoxicity, oxidative stress, edema formation, blood brain barrier hyper-permeability, and neuroinflammation. If this hypothesis is correct, Nln could potentially serve as a single therapeutic target to modulate the function of multiple targets, the involved neuropeptide systems, critically involved in various mechanisms of brain injury and cerebroprotection/restoration. Such multi-pathway target would be highly desired for pharmacotherapy of stroke and TBI, because targeting one pathophysiological pathway has proven to be ineffective for such complex disorders.

Introduction

Stroke and traumatic brain injury (TBI) are two major, acute neurodegenerative disorders which constitute the leading causes of death and disability worldwide affecting both old and young individuals [1]. Current experimental stroke and traumatic brain injury research focuses more on understanding the brain's self-protective/repair mechanisms to meet the critical need of developing new stroke therapies [2,3]. The underlying premise of this approach is that the brain has well-developed, complex and highly conserved endogenous mechanisms for self-preservation and repair in adverse conditions and following injury. It is believed that detailed understanding of these mechanisms is important as such knowledge could lead to development of pharmacological or other therapeutic interventions to mimic or engage brain's self-protective/repair mechanisms for successful therapy [2].

While etiologies of stroke and TBI are different, the main secondary brain injury mechanisms are shared in these disorders and involve excitotoxicity, oxidative stress, edema formation, blood brain barrier (BBB) hyperpermeability, and neuroinflammation. All of these brain injury cascades have been studied thoroughly over the last several decades [4,5], however, there has been much less emphasis on the role

of neuropeptides in these pathophysiological processes. This is despite the fact that neuropeptides are the most diverse signaling molecules in mammals and primarily come into play when the nervous system is adapting/responding to various challenges, thus representing the "language of the stressed nervous system" [6]. It is well known that the actions of neuropeptides are tightly linked to the function of peptidases, which are hydrolytic enzymes involved in processing of bioactive peptides [7–9]. Altered expression and/or activity of several brain peptidases have been reported in experimental stroke and TBI studies linking the function of peptidases and related neuropeptide systems to the pathophysiology of these disease [10,11]. Here, I summarize the results of our published studies which provide evidence about the role of peptidase neurolysin (Nln) in the post-stroke brain. In addition, based on our unpublished observations and published studies from other laboratories, I make a case that Nln is one of the brain's self-protective mechanisms directed towards preservation and recovery of brain in acute neurodegenerative disorders.

Association of neurolysin with neuronal cell death

Our interest in Nln originates from studies related to the non-AT₁,

* Address: 1300 Coulter St., Amarillo, TX 79106, United States.

E-mail address: vardan.karamyan@ttuhsc.edu.

non-AT₂ angiotensin binding site, which was discovered, characterized and identified by us as membrane Nln [12–16]. Nln (EC 3.4.24.16) is a zinc endopeptidase belonging to M3 family which make the most important group of peptidases responsible for hydrolytic processing of bioactive peptides in the extracellular environment [9]. It was by serendipity that in experiments conducted several years ago we observed increased expression of membrane-bound variant of neurolysin (Nln) in primary mouse cortical neurons that were maintained in culture for about 2 weeks without replenishment of the culture medium [17]. This was followed by more systematic set of experiments which revealed upregulation of Nln in primary neurons challenged in four *in vitro* models of cell death (excitotoxicity by NMDA, oxygen-glucose deprivation followed by re-oxygenation, hypoxia by sodium azide, and oxidative stress by hydrogen peroxide), and provided evidence about the potential role of this peptidase in neuronal cell death for the first time. However, these original observations did not answer two critical questions, a) whether upregulation of Nln occurs only *in vitro* or it has *in vivo* relevance, and b) what the function of Nln is in neuronal death.

Upregulation of Nln in mouse brain after stroke

We sought to answer the first question in a mouse model of ischemic stroke which confirmed our *in vitro* observations. Experiments carried out in transient, middle cerebral artery occlusion model of stroke (MCAO, 1 h occlusion followed by 24 h re-perfusion) revealed significant increase (\geq twofold) in quantity and activity of membrane Nln in ischemia-affected parts of the mouse brain [18]. Importantly, this study also documented sustained functional upregulation of Nln in ischemia-affected cerebral cortical membranes for at least 7 days after stroke. Upregulation of Nln did not appear to be transcriptionally or translationally regulated, but rather depended on translocation of cytosolic Nln to the membranes and mitochondria [18]. Collectively these findings confirmed our *in vitro* observations and suggested that Nln may play a role in processes modulating the brain's response to stroke.

Additional *in vivo* evidence linking Nln to cell death

The potential role of Nln in neuronal cell death is additionally supported by published studies which evaluated the expression of Nln in developing rodent brains [17,19]. Expression of membrane Nln was shown to increase gradually in mouse forebrain membranes from E14 to P10, followed by a dramatic drop in P21 animals and similar levels in 9- to 12-week-old animals [17]. It is noteworthy that the pattern of developmental changes in the expression of Nln is very similar to the occurrence of neuronal death in developing brain [20]. The number of dead neurons in rat cerebral cortex is low at birth, which increases from P2, peaking at the end of the first week, and decreases during the second week followed by low numbers at the end of the first month [20]. These *in vivo* observations complement the above discussed data and support the association between neuronal cell death and Nln. Moreover, these results clarify earlier observations in adult rat brain where distribution of Nln was studied by *in vitro* receptor autoradiography [15]. In the latter study, among brain regions with the highest expression of Nln were the olfactory bulb (highest compared with other brain regions), walls of ventricles (throughout brain), and dentate gyrus. Complementary to this, it was shown that the frequency of apoptosis in adult rat brain is up to 100 times higher in olfactory bulb, ventricular wall, and dentate gyrus (in decreasing order) compared with other brain areas [21]. Lastly, another line of evidence comes from a list of functional proteins which have ontogenic and post-stroke expression profile very similar to Nln (i.e. overexpression), and are viewed as key molecular players in response of brain to ischemia and acute injury [22].

It is important to note that in the brain, Nln is expressed not only in neurons but also in astrocytes [23] and likely in endothelial cells of the brain vasculature [24]. Cellular localization (plasma membrane vs.

cytosol and mitochondria) of Nln differs in these cell types and changes during brain development [19,23]. Currently, it is unknown which transcription factor(s) control expression of Nln and how the latter is affected in dying astrocytes and endothelial cells.

Nln substrates and their role in acute neurodegeneration

To understand the potential significance of Nln upregulation in the post-stroke brain it is important to recognize the peptide substrates that are hydrolyzed by Nln and their function in the setting of acute brain injury. Endogenous substrates of Nln are neurotensin, bradykinin, substance P, angiotensins I and II, hemopressin, dynorphin A(1–8), and metorphamide [9,18,25]. Nln cleaves and inactivates all of these peptides except angiotensin I, dynorphin A(1–8) and metorphamide, which are converted into angiotensin-(1–7) and enkephalins, respectively. Importantly, all of these peptide systems are directly involved in pathogenesis of stroke and/or TBI [18]. Integral role of bradykinin in these disorders was demonstrated by *in vivo* studies indicating that both B1 and B2 receptors are involved in development of neuroinflammation, brain edema and cell death [26,27]. Pathological role of neurotensin in ischemic brain is supported by *in vitro* studies demonstrating decreased survival of primary neurons after ischemia in the presence of neurotensin, and blockade of its effects by NT1 receptor antagonists [28,29]. This effect involves potentiation of glutamate release and amplification of NMDA-R-mediated glutamate signaling in neurons [28,30]. Substance P is another member of the kinin family with critical role in neurogenic inflammation, oxidative stress, genesis of edema and cell death after stroke and TBI [31,32]. Angiotensin I is an inactive peptide, serving as a precursor for bioactive angiotensins. Cleavage of angiotensin I by Nln results in angiotensin-(1–7), which was shown to have neuroprotective and anti-inflammatory effects in *in vivo* stroke and TBI models [33,34]. Complementary to this, Nln inactivates angiotensin II, which is the most studied peptide in the renin-angiotensin system with mounting evidence supporting its pathological role in stroke and TBI [35,36]. Hemopressin is a potent CB1 cannabinoid receptor antagonist (also acts as inverse agonist) which is inactivated by Nln. Inactivation of hemopressin facilitates CB1 receptor activation, which is known to mediate neuro- and angiogenesis, and survival of new neurons [37]. Dynorphin A(1–8), a kappa-opioid receptor agonist, and metorphamide, a kappa- and mu-opioid receptor agonist, are converted into potent delta-opioid receptor agonists Leu- and Met-enkephalins, respectively, by Nln. The role of delta-opioid receptors in resistance to ischemia, neuroprotection and post-stroke brain repair was demonstrated by our and other groups [38,39]. It is important to note that findings of a small number of studies do not support the cerebrotoxic effects of some Nln substrates in the setting of acute brain injury [27,40,41]. Unfortunately, it would be impossible to discuss all experimental studies on the pivotal function of Nln substrate peptides in pathophysiology of stroke and TBI here, given their vast number and page limitations of this article. Importantly though, a number of recent clinical studies support this preclinical evidence and strongly relate the severity of stroke injury and TBI, and subsequent mortality to elevated serum and/or cerebrospinal levels of bradykinin, substance P and neurotensin [42–45], in some cases documenting 5 to 10-fold increase of these peptides following injury.

Hypothesis and future directions

Considering diversity of endogenous Nln substrates and the well-documented role of these peptidergic systems in pathogenesis of stroke and TBI, I hypothesize that upregulation of Nln following neuronal cell death is a compensatory and self-protective mechanism which functions to preserve expansion of damage and restore the brain after injury. To test this hypothesis, our current studies focus on ischemic stroke using mouse models, where we are applying two independent approaches, inhibition of Nln by a specific inhibitor and brain overexpression by an

AAV vector, to verify functional significance of Nln in the post-stroke brain. The outcome measures in this study include brain infarction and cell death, formation of edema, disruption of the blood brain-barrier (BBB), functional impairment and overall survival. Our preliminary results support the hypothesis and it is believed that cerebroprotective function of Nln is, at least in part, linked to hydrolysis of its extracellular peptide substrates and by that prevention of cellular and vasogenic edema, oxidative stress, preservation of BBB, reduction of neuroinflammation and cell death following stroke. These ongoing studies will be followed by more mechanistic and cell type-specific investigations to reveal the role of different brain cell types and substrate peptides in Nln-mediated cerebroprotection. If this hypothesis is confirmed in stroke settings, it will be critical to evaluate the potential of Nln function in other neurological disorders including TBI and vascular dementia.

It is important to recognize that the ability of Nln to process several neuropeptides suggests that it could potentially serve as a single therapeutic target to modulate the function of multiple targets, the noted neuropeptide systems, critically involved in various mechanisms of brain injury or cerebroprotection/restoration. Such multi-pathway target would be highly desired for pharmacotherapy of stroke and TBI, because in recent years it has been recognized that targeting one pathophysiological pathway is unlikely to be therapeutically effective in such complex disorders. If this idea is confirmed by systematic studies, then brain-penetrating variants of Nln or small molecule activators of Nln, which are being developed in our laboratory, could become a new class of drugs to be used for cerebroprotection and restoration of the brain after acute injuries.

Declaration of Competing Interest

VTK is the senior inventor on a provisional patent, filed by Texas Tech University Systems, focusing on discovery of small molecule activators of Nln.

Acknowledgements

This research was supported by research grants from the American Heart Association (14BGIA20380826) and NIH (1R01NS106879). I apologize that the scope of this manuscript prevented citation of many important studies.

References

- [1] Sorby-Adams AJ, Marcoianni AM, Dempsey ER, Woenig JA, Turner RJ. The role of neurogenic inflammation in blood-brain barrier disruption and development of cerebral oedema following acute central nervous system (CNS) injury. *Int J Mol Sci* 2017;18.
- [2] Iadecola C, Anrather J. Stroke research at a crossroad: asking the brain for directions. *Nat Neurosci* 2011;14:1363–8.
- [3] Tovar-y-Romo LB, Penagos-Puig A, Ramirez-Jarquín JO. Endogenous recovery after brain damage: molecular mechanisms that balance neuronal life/death fate. *J Neurochem* 2016;136:13–27.
- [4] Kunz A, Dirnagl U, Mergenthaler P. Acute pathophysiological processes after ischaemic and traumatic brain injury. *Best Pract Res Clin Anaesthesiol* 2010;24:495–509.
- [5] Bramlett HM, Dietrich WD. Pathophysiology of cerebral ischemia and brain trauma: similarities and differences. *J Cereb Blood Flow Metab* 2004;24:133–50.
- [6] Hokfelt T, Bartfai T, Bloom F. Neuropeptides: opportunities for drug discovery. *Lancet Neurol* 2003;2:463–72.
- [7] Karamyan VT, Speth RC. Enzymatic pathways of the brain renin-angiotensin system: unsolved problems and continuing challenges. *Regul Pept* 2007;143:15–27.
- [8] Speth RC, Karamyan VT. The significance of brain aminopeptidases in the regulation of the actions of angiotensin peptides in the brain. *Heart Fail Rev* 2008;13:299–309.
- [9] Shrimpton CN, Smith AI, Lew RA. Soluble metalloendopeptidases and neuroendocrine signaling. *Endocr Rev* 2002;23:647–64.
- [10] Rashid M, Karamyan VT. Peptidase neurolysin: Its function related to the brain renin-angiotensin system and pathophysiology of stroke. Letter to the Editor. *J Clin Neurosci* 2018;48:245.
- [11] Abdul-Muneer PM, Pfister BJ, Haorah J, Chandra N. Role of matrix metalloproteinases in the pathogenesis of traumatic brain injury. *Mol Neurobiol* 2016;53:6106–23.
- [12] Karamyan VT, Gembardt F, Rabey FM, Walther T, Speth RC. Characterization of the brain-specific non-AT(1), non-AT(2) angiotensin binding site in the mouse. *Eur J Pharmacol* 2008;590:87–92.
- [13] Karamyan VT, Speth RC. Identification of a novel non-AT1, non-AT2 angiotensin binding site in the rat brain. *Brain Res* 2007;1143:83–91.
- [14] Karamyan VT, Stockmeier CA, Speth RC. Human brain contains a novel non-AT1, non-AT2 binding site for active angiotensin peptides. *Life Sci* 2008;83:421–5.
- [15] Karamyan VT, Speth RC. Distribution of the non-AT1, non-AT2 angiotensin-binding site in the rat brain: preliminary characterization. *Neuroendocrinology* 2008;88:256–65.
- [16] Wangler NJ, Santos KL, Schadock I, et al. Identification of membrane-bound variant of metalloendopeptidase neurolysin (EC 3.4.24.16) as the non-angiotensin type 1 (Non-AT1), Non-AT2 angiotensin binding site. *J Biol Chem* 2012;287:114–22.
- [17] Rashid M, Arumugam TV, Karamyan VT. Association of the novel non-AT1, non-AT2 angiotensin binding site with neuronal cell death. *J Pharmacol Exp Ther* 2010;335:754–61.
- [18] Rashid M, Wangler NJ, Yang L, et al. Functional upregulation of endopeptidase neurolysin during post-acute and early recovery phases of experimental stroke in mouse brain. *J Neurochem* 2014;129:179–89.
- [19] Dauch P, Masuo Y, Vincent JP, Checler F. Endopeptidase 24–16 in murines: tissue distribution, cerebral regionalization, and ontogeny. *J Neurochem* 1992;59:1862–7.
- [20] Naruse I, Keino H. Apoptosis in the developing CNS. *Prog Neurobiol* 1995;47:135–55.
- [21] Biebl M, Cooper CM, Winkler J, Kuhn HG. Analysis of neurogenesis and programmed cell death reveals a self-renewing capacity in the adult rat brain. *Neurosci Lett* 2000;291:17–20.
- [22] Cramer SC, Chopp M. Recovery recapitulates ontogeny. *Trends Neurosci* 2000;23:265–71.
- [23] Vincent B, Beaudet A, Dauch P, Vincent JP, Checler F. Distinct properties of neuronal and astrocytic endopeptidase 3.4.24.16: a study on differentiation, subcellular distribution, and secretion processes. *J Neurosci* 1996;16:5049–59.
- [24] Norman MU, Lew RA, Smith AI, Hickey MJ. Metalloendopeptidases EC 3.4.24.15/16 regulate bradykinin activity in the cerebral microvasculature. *Am J Physiol Heart Circ Physiol* 2003;284:H1942–8.
- [25] Checler F, Ferro ES. Neurolysin: from initial detection to latest advances. *Neurochem Res* 2018;43:2017–24.
- [26] Sobey CG. Bradykinin B2 receptor antagonism: a new direction for acute stroke therapy? *Br J Pharmacol* 2003;139:1369–71.
- [27] Albert-Weissenberger C, Siren AL, Kleinschnitz C. Ischemic stroke and traumatic brain injury: the role of the kallikrein-kinin system. *Prog Neurobiol* 2013;101–102:65–82.
- [28] Antonelli T, Ferraro L, Fuxe K, et al. Neurotensin enhances endogenous extracellular glutamate levels in primary cultures of rat cortical neurons: involvement of neurotensin receptor in NMDA induced excitotoxicity. *Cereb Cortex* 2004;14:466–73.
- [29] Antonelli T, Tomasini MC, Fournier J, et al. Neurotensin receptor involvement in the rise of extracellular glutamate levels and apoptotic nerve cell death in primary cortical cultures after oxygen and glucose deprivation. *Cereb Cortex* 2008;18:1748–57.
- [30] Ferraro L, Tomasini MC, Beggiato S, et al. Emerging evidence for neurotensin receptor 1 antagonists as novel pharmaceuticals in neurodegenerative disorders. *Mini Rev Med Chem* 2009;9:1429–38.
- [31] Richter F, Eitner A, Leuchtweis J, et al. The potential of substance P to initiate and perpetuate cortical spreading depression (CSD) in rat in vivo. *Sci Rep* 2018;8:17656.
- [32] Corrigan F, Vink R, Turner RJ. Inflammation in acute CNS injury: a focus on the role of substance P. *Br J Pharmacol* 2016;173:703–15.
- [33] Mecca AP, Regenhardt RW, O'Connor TE, et al. Cerebroprotection by angiotensin-(1–7) in endothelin-1-induced ischaemic stroke. *Exp Physiol* 2011;96:1084–96.
- [34] Janatpour ZC, Korotcov A, Bosomtwi A, Dardzinski B, Symes A. Subcutaneous administration of angiotensin-(1–7) improves recovery after traumatic brain injury in mice. *J Neurotrauma* 2019.
- [35] Iadecola C, Gorelick PB. Hypertension, angiotensin, and stroke: beyond blood pressure. *Stroke* 2004;35:348–50.
- [36] Villapol S, Saavedra JM. Neuroprotective effects of angiotensin receptor blockers. *Am J Hypertens* 2015;28:289–99.
- [37] van der Stelt M, Di Marzo V. Cannabinoid receptors and their role in neuroprotection. *Neuromol Med* 2005;7:37–50.
- [38] Crowley MG, Liska MG, Lippert T, Corey S, Borlongan CV. Utilizing delta opioid receptors and peptides for cytoprotection: implications in stroke and other neurological disorders. *CNS Neurol Disord Targets* 2017;16:414–24.
- [39] Vaidya B, Sifat AE, Karamyan VT, Abbruscato TJ. The neuroprotective role of the brain opioid system in stroke injury. *Drug Discov Today* 2018;23:1385–95.
- [40] Choi KE, Hall CL, Sun JM, et al. A novel stroke therapy of pharmacologically induced hypothermia after focal cerebral ischemia in mice. *Faseb J* 2012;26:2799–810.
- [41] Amadoro G, Pieri M, Ciotti MT, et al. Substance P provides neuroprotection in cerebellar granule cells through Akt and MAPK/Erk activation: evidence for the involvement of the delayed rectifier potassium current. *Neuropharmacology* 2007;52:1366–77.
- [42] Lorente L, Martín MM, Almeida T, et al. Serum levels of substance P and mortality in patients with a severe acute ischemic stroke. *Int J Mol Sci* 2016;17.
- [43] Januzzi Jr. JL, Lyass A, Liu Y, et al. Circulating proneurotensin concentrations and cardiovascular disease events in the community: the framingham heart study. *Arterioscler Thromb Vasc Biol* 2016;36:1692–7.
- [44] Kunz M, Nussberger J, Holtmannspotter M, Bitterling H, Plesnila N, Zausinger S. Bradykinin in blood and cerebrospinal fluid after acute cerebral lesions: correlations with cerebral edema and intracranial pressure. *J Neurotrauma* 2013;30:1638–44.
- [45] Zacest AC, Vink R, Manavis J, Sarvestani GT, Blumbergs PC. Substance P immunoreactivity increases following human traumatic brain injury. *Acta Neurochirurgica* 2010;106(Supplement):211–6.