



## Topical Review

## Pediatric West Nile Virus-Associated Neuroinvasive Disease: A Review of the Literature

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## ABSTRACT

Over the past two decades, West Nile virus has become the most common arbovirus in North America, leading to several outbreaks and infecting thousands of people. Mosquitos help transmit the virus in the majority of cases, but transmission occurs via blood transfusions, organ transplantation, and possibly pregnancy and breastfeeding. While most infected patients experience mild to no symptoms, thousands of West Nile virus-associated neuroinvasive cases have been reported in the United States, with over 700 cases occurring in children from 2003 to 2016. Neuroinvasive disease presents as meningitis, encephalitis, or acute flaccid paralysis, and carries a high likelihood of poor outcome, including severe neurological disability or death. To date, no pharmacologic treatment has proven effective. Therapeutic clinical trials have not been successfully completed due to the sporadic nature of viral outbreaks and resultant poor study enrollment. Although older age and chronic disease are risk factors for neuroinvasive West Nile virus disease in adults, the specific factors that influence the risk in pediatric populations have not been fully elucidated. This review summarizes the most recent literature regarding West Nile virus-associated neuroinvasive disease, especially as it pertains to the pediatric population. Moreover, the review describes the epidemiology, clinical, laboratory, and radiographic findings, and outlines the various therapies that have been trialed and potential future research directions.

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## Introduction

West Nile virus (WNV), a mosquito-borne flavivirus, is the most common arbovirus in North America. The virus was first isolated in 1937 in Uganda and has been well studied since the 1950s when several outbreaks occurred in the Mediterranean basin.<sup>1</sup> The first WNV infection in North America was diagnosed in New York City in 1999 and was followed by rapid spread across the continent. Within four years, patients were reported from 45 states and multiple Canadian provinces.<sup>2,3</sup> Since then, sporadic outbreaks have occurred, the largest of which was in 2012 with over 5000 patients reported in the United States.<sup>4</sup> While most infections result in mild to no symptoms, nervous system involvement can result in

severe outcomes, and some cases are fatal.<sup>5</sup> From 1999 to 2016, over 2000 WNV-associated fatalities were reported to the Centers for Disease Control and Prevention (CDC), with an estimated mortality rate of 4% among all reported WNV infections and 9% of neuroinvasive cases.<sup>6</sup> WNV-associated neuroinvasive disease (WNND) primarily affects adults, but children are also susceptible, accounting for 4% of neuroinvasive cases annually.<sup>7</sup> This review summarizes the current literature regarding WNND, especially as it pertains to the pediatric population.

## Epidemiology

*Incidence and geographic distribution*

WNV causes the majority of arboviral-related neuroinvasive disease overall in North America, primarily in adults.<sup>8</sup> Pediatric cases account for 4% of all WNND, with an average annual incidence of 0.68 per million children.<sup>7,8</sup> From 1999 to 2016, 2397 cases in individuals under 19 years of age have been reported to the CDC, 34% of which were neuroinvasive (Fig 1, CDC, personal

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communication, May 3, 2018). It should be noted that this reflects the incidence of neuroinvasive disease of all reported cases; the true risk of neuroinvasion following infection remains sparingly low, since 80% of infections are asymptomatic and typically go unreported.<sup>9</sup>

In addition to WNV, children in North America are at risk for other arboviral infections, including Eastern Equine encephalitis virus, La Crosse virus (LACV), Powassan virus, and St. Louis encephalitis virus.<sup>8</sup> Of these, LACV, rarely reported in adults, is the most common cause of neuroinvasive disease in children and adolescents. LACV is found in the Midwest, Appalachian, and Atlantic states, while WNV infections occur throughout the United States, having been reported in every state with the exceptions of Hawaii and Alaska.<sup>6,10</sup> Presumably, much of the difference in geographic distribution is due to habitats of various mosquito species and the activity of the vertebrate host associated with each virus. The wide variety of mosquito and bird species that carry WNV have likely aided in its widespread distribution.<sup>11</sup> While the *Culex* mosquito most often associated with WNV is able to survive relatively cold climates, its population growth is accelerated by warm weather, so as might be expected, the highest frequency of WNV occurs in the highly populated, warm states of Texas and California.<sup>6,11</sup> Consequently, these states are also associated with the highest incidence of WNND, each with over 100 patients in most years since 2002 per CDC data.<sup>6,11</sup> However, sporadic outbreaks have occurred throughout most of the United States and Canada, including regions with typically cooler climates. Many factors are involved in priming a region for an outbreak, including climate of the current season and preceding winter, extent of viral activity the previous transmission season, mosquito control, and human behavior such as dress and outdoor activity.<sup>11</sup> It is therefore quite difficult to predict where outbreaks might occur, although predictive models have recently been developed with some accuracy.<sup>12</sup>

#### Risk factors

Literature examining WNND risk factors specific to children is lacking, presumably because of its rare occurrence in this population. The few published pediatric epidemiologic studies have found

a slight predilection for males and adolescents for both West Nile fever and WNND.<sup>7,13,14</sup> Several studies have evaluated potential host risk factors in adults; from these, there is consensus that advanced age correlates with risk of WNND and death.<sup>15,16</sup> The literature is conflicting regarding other potential risk factors in adults, but may include other demographic factors (male sex and minority race), cancer, diabetes, hypertension, alcohol abuse, renal disease, immunosuppression, and chronic obstructive pulmonary disease.<sup>15–18</sup> Some of the predilection for older adults is likely due to the lack of these comorbidities in most young patients. The high prevalence of chronic disease in older patients is thought to play a role in altering the cerebral endothelium, facilitating viral entry into the central nervous system (CNS).<sup>8</sup> Interestingly, this pattern is similar to that of St. Louis encephalitis but contrasts with LACV, possibly due to an immature adaptive immune response as demonstrated in mouse models.<sup>19,20</sup>

#### Transmission

##### Mosquitos

Nearly all cases of WNV are transmitted via a mosquito bite.<sup>21</sup> The virus is maintained in a mosquito-bird transmission cycle, with members of the *Culex* mosquito genus serving as the most common vectors. Several bird species act as viral hosts, transmitting the virus back to the mosquito after infection.<sup>22</sup> After the virus is injected intradermally by the mosquito, it infects targets such as the Langerhans cells that spread via the lymphatic system to the bloodstream, resulting in viremia.<sup>23,24</sup> The virus then further replicates, causing a secondary viremia, and can potentially gain access to the CNS, presumably through hematogenous and trans-neuronal mechanisms.<sup>25</sup>

##### Blood transfusion

Transmission of WNV through blood transfusion was first reported in 2002. Four of the 23 confirmed cases occurred in children.<sup>26</sup> In 2003, routine measures were put in place to screen all blood donations in the United States for WNV RNA. Since then, 14

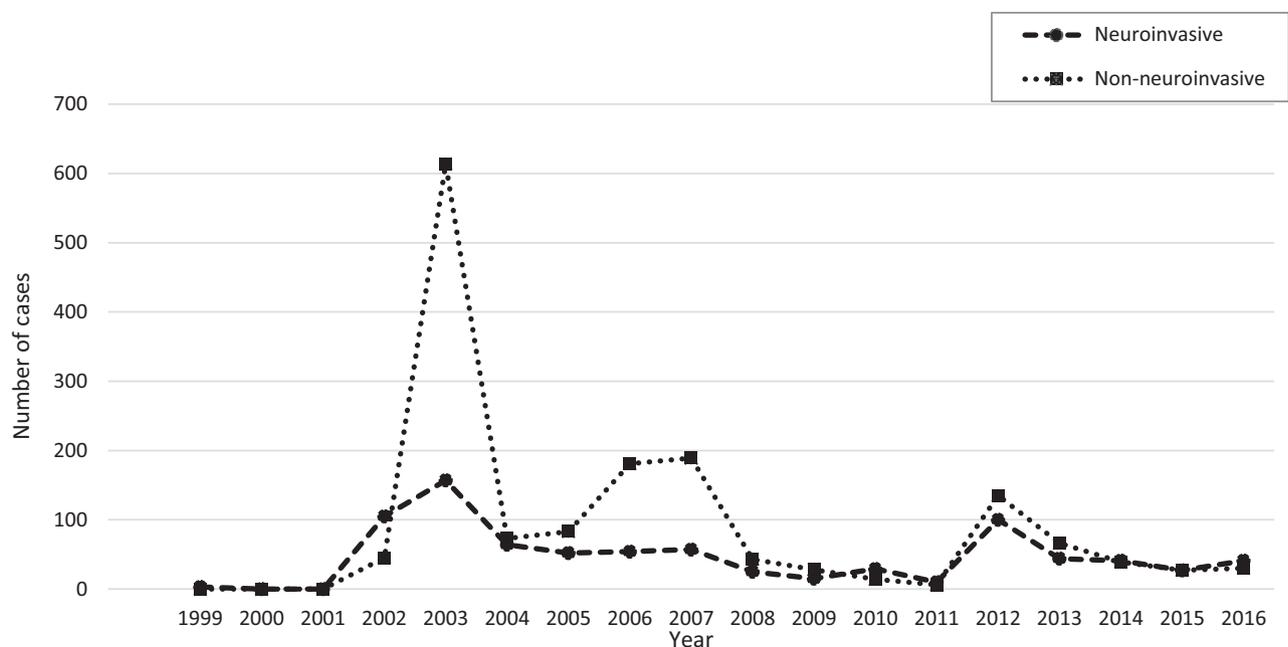


FIGURE 1. Annual number of WNV infections reported in individuals less than 19 years old in the United States. WNV, West Nile virus.

transfusion-related transmissions have been reported to the CDC.<sup>27,28</sup> In 13 of these patients, the donor samples were screened with false-negative results.<sup>28</sup> This typically occurs because blood is often initially screened through minipool-nucleic acid testing (NAT) consisting of six or 16 individual donors, which is less sensitive than individual-nucleic acid testing (ID-NAT).<sup>28</sup> The Food and Drug Administration therefore recommends using ID-NAT instead of minipool-NAT for initial screening during periods of high WNV activity.<sup>29</sup> Selective use of ID-NAT as of 2004 has resulted in greater detection of WNV by blood banks; however, whether this has translated into fewer clinical infections is unclear due the brief interval since this change, and that not all transfusions of infected samples result in clinical infection.<sup>30</sup>

#### *Organ transplant*

WNV transmission via solid organ transplant has been documented in 20 adults, whereas cases of community-acquired WNNND in the setting of immunosuppression *after* transplant have been reported among both pediatric and adult recipients.<sup>16,31–35</sup> Among the published reports of donor-derived infection, 70% of recipients developed WNNND (as compared with the estimated 1% of the general population who contract infection via mosquito bite), and 30% resulted in persistent coma or death.<sup>36</sup> Some patients remain asymptomatic, and those who do not develop coma tend to recover to variable degrees.<sup>36–41</sup> The proper management of immunosuppressants in these cases is uncertain.<sup>42</sup> Of the six published cases documenting an immunosuppressant reduction following WNV infections, two of the patients recovered, one remained asymptomatic, and three died.<sup>36,37,39,40</sup> It should be noted that these patients received other therapies as well, as listed in [Table 3](#).

#### *Pregnancy and breastfeeding*

While cases of transmission related to pregnancy and breastfeeding have been reported, such cases are rare and often unconfirmed. There are no confirmed cases of clinical illness secondary to breast milk ingestion. There has been a single clearly documented case of seropositivity in an infant following ingestion of infected breast milk, but the infant remained healthy.<sup>43</sup> Two other breastfed infants whose mothers contracted WNV before delivery tested positive for WNV IgM antibodies; therefore, it is unclear whether this was transmitted through breastmilk, placenta, or through mosquito exposure. One of these infants developed a rash, and the other was asymptomatic.<sup>44</sup>

Transplacental transmission has been reported, but the risk appears to be quite low. A 2004 study determined that of 72 infants born to mothers who contracted WNV during pregnancy, only three became infected, and two developed clinical neuroinvasive disease. However, transplacental transmission could not be verified due to the unavailability of appropriate specimens (umbilical cord, cord blood, or placenta) for confirmatory testing.<sup>45</sup> There does not appear to be a significant risk of congenital malformations, developmental delay, or pregnancy complications among pregnancies of WNV-infected mothers, but larger studies are needed to confirm these findings.<sup>46</sup>

#### **Clinical features**

West Nile fever is the most common symptomatic presentation of WNV infection, characterized by fever, headache, fatigue, and myalgia. Vomiting, diarrhea, rash, and lymphadenopathy can also be present. These nonspecific symptoms typically resolve within a week.<sup>47</sup> The three most common presentations of WNNND are meningitis, encephalitis, and acute flaccid paralysis (AFP). In adults,

involvement of the spinal nerve root, brachial plexus, and peripheral nerve has also been described.<sup>48,49</sup> Other uncommon presentations of WNV infection in children have included cerebellitis, cranial nerve palsies, and hepatic and cardiac involvement.<sup>50–53</sup>

#### *Meningitis and encephalitis*

Similar to adults, WNNND represents 30% of all reported pediatric WNV infections. In contrast to older adults, however, children with WNNND are more likely to present with meningitis rather than encephalitis.<sup>7</sup> This differentiation is of important prognostic significance as patients with encephalitis have more severe outcomes.<sup>7</sup> The initial presenting symptoms of both meningitis and encephalitis may be quite vague, most often consisting of fever (which can be low-grade), headache, and vomiting or abdominal pain. Rash may occur in WNNND but is more frequently reported in children with WNV fever; it has been postulated as a positive prognostic sign.<sup>14,54</sup> Meningismus may or may not occur in children with WNV meningitis, and it can develop later in the course if not initially present.<sup>55</sup> Children with WNV encephalitis (WNE) may first present with the above nonspecific symptoms with progression to altered mental status or obtundation within one to seven days, while others initially present to the emergency room with unresponsiveness or seizures.<sup>16,34,56</sup> In addition to altered mental status, WNE can manifest as movement disorders such as myoclonus, tremor, and parkinsonism, which may be secondary to the frequent involvement of the basal ganglia.<sup>57</sup> Patients with suspected meningoencephalitis should undergo fundoscopy, given the high incidence of associated chorioretinitis, papilledema, and optic neuritis.<sup>34,55,58</sup> In one child the characteristic pattern of WNV chorioretinitis, described as scattered or linear lesions, guided the treating team toward the diagnosis of WNV.<sup>56</sup>

#### *Acute flaccid paralysis*

AFP has been reported in around 1% of children with WNNND, and can occur with or without encephalitis, with the former being more common.<sup>59</sup> Similar to poliomyelitis, the syndrome is caused by WNV invasion of the anterior horn cells. This results in a rapidly progressive, asymmetric paralysis, however, a slow progression may also be observed in children.<sup>52,59</sup> Paralysis may be preceded by pain of the affected extremity.<sup>52,59</sup> The neurological examination in these patients reveals asymmetric flaccid paralysis with loss of deep tendon reflexes in the affected limbs.<sup>52,59,60</sup> Nerve conduction studies and cerebrospinal fluid (CSF) characteristics can be helpful in differentiating viral AFP from other treatable causes of paralysis, such as Guillain-Barré syndrome ([Table 1](#)).

#### **Neuroimaging**

##### *Brain*

Although brain magnetic resonance imaging (MRI) and computed tomography are often normal in patients with WNV meningoencephalitis, MRI should be performed in the initial evaluation of patients with suspected CNS disease given its usefulness in including or excluding differential entities. Computed tomography is often insensitive to manifestations of WNV, but offers an important supportive screening role during primary evaluation of the patient with a neurological deficit who will undergo lumbar puncture when MRI is unavailable.<sup>61</sup> A significant number of patients with WNV meningoencephalitis may have normal brain imaging findings even with significant symptoms such as alteration in mental status and dysphasia.<sup>62</sup> Several reports, including one pediatric, have documented that imaging may be initially normal

**TABLE 1.**  
Ancillary Studies in Viral AFP and Guillain-Barré Syndrome<sup>48</sup>

	Viral AFP, Including WNV	Guillain-Barré Syndrome
CSF	Elevated protein with pleocytosis	Elevated protein with no pleocytosis
NCS	Decreased motor action potentials	Slow conduction velocities
	Normal sensory action potentials	Decreased sensory action potentials

Abbreviation:

NCS = nerve conduction studies

with follow-up imaging proving abnormal; therefore, an initial normal imaging pattern may be a function of time bias.<sup>56,63,64</sup>

On conventional sequences, T2 signal (with corresponding low T1 signal) abnormality with or without diffusion signal abnormality may be seen in the lobar gray and white matter, basal ganglia, thalami, brainstem, and even cerebellum. MRI findings among pediatric patients are similar to those of adults, with abnormalities of the thalami and basal ganglia most often described, and midbrain and brainstem involvement also reported.<sup>55,56,65</sup> While such findings may be focal, patchy, and asymmetric, strikingly symmetric signal abnormality may also occur especially at the level of the central gray matter (Fig 2).<sup>62,66-68</sup> In fact, certain flaviviruses, such as WNV, eastern equine encephalitis virus, Japanese encephalitis virus, and Murray Valley encephalitis virus, have a predilection for bilateral symmetric central gray matter. In contrast, such a pattern is less common with postnatal Zika.<sup>62,66-68</sup> Ultimately, imaging findings are nonspecific to WNV and overlap with a wide variety of viral infections including many arboviruses, as well as nonarboviral infections, such as Epstein-Barr virus and *Mycoplasma pneumoniae*.<sup>69-79</sup> Hemorrhagic necrosis within the central gray matter may be seen with Japanese encephalitis, dengue fever, and *M. pneumoniae*.<sup>70,74,79</sup> WNV meningoencephalitis imaging findings may also be mimicked by many parainfectious and noninfectious processes, such as acute disseminated encephalomyelitis, acute necrotizing encephalitis, toxic-metabolic or mitochondrial disorders, and vasculitides, emphasizing the importance of clinico-radiologic interpretation (Fig. 3 and 4). Focal diffusion restriction with T2 prolongation may mimic lacunar or thromboembolic stroke.

Bilateral or unilateral mesial temporal involvement T2 hyperintensity is an additional imaging pattern that has been reported with WNV meningoencephalitis. When diffusion restriction coexists with such mesial temporal involvement, findings may mimic herpes simplex meningoencephalitis although the

aggressive necrotizing appearance of this entity is usually absent.<sup>66</sup> Many other viral agents may produce a limbic encephalitis including most classically HHV-6.<sup>78</sup> Histopathologically, imaging findings correlate with perivascular lymphocytic infiltration, microglial nodules, and neuronal loss.<sup>80</sup>

In some patients, diffusion-weighted imaging may independently show signal abnormality (i.e., restricted diffusion) without corresponding findings on conventional T1 and T2 sequences. This is likely a time bias effect in that diffusion imaging may be positive in the earliest phases with a more mature inflammatory cascade resulting later in signal abnormality on conventional sequences. Diffusion restriction is presumed a function of inflammatory infiltration rather than true ischemia, although this remains incompletely discovered in the literature.<sup>66</sup> Vasculitis has not been reported as a primary feature in WNV meningoencephalitis.<sup>80</sup>

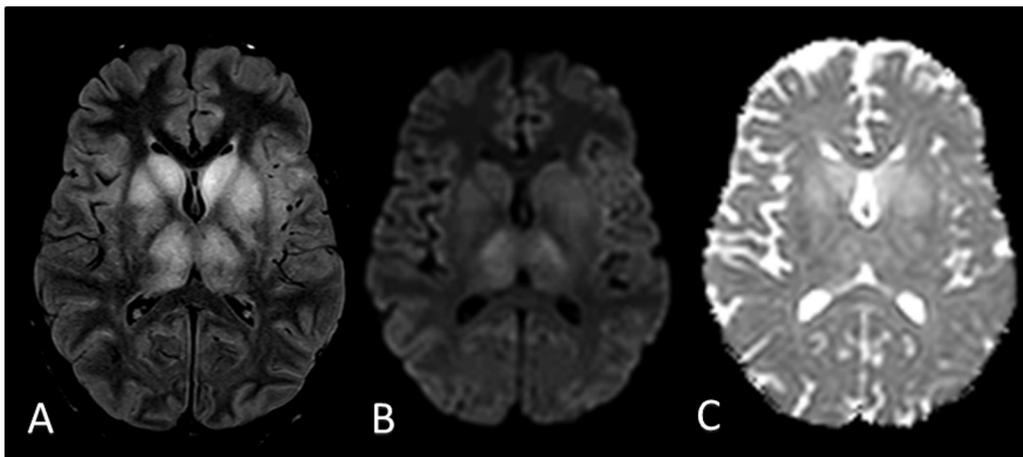
Postcontrast imaging is often normal, distinctly different from typical bacterial, atypical bacterial, and fungal entities. However, leptomeningeal and pachymeningeal enhancement have been reported.<sup>62</sup> T2 fluid-attenuated inversion recovery precontrast imaging may show lack of suppression of T2 signal within the sulci due to alterations in protein and cellular content of the CSF.<sup>62</sup> Subarachnoid imaging findings may be secondary to reactive lymphocytic and plasma cell infiltration.<sup>81</sup>

### Spine

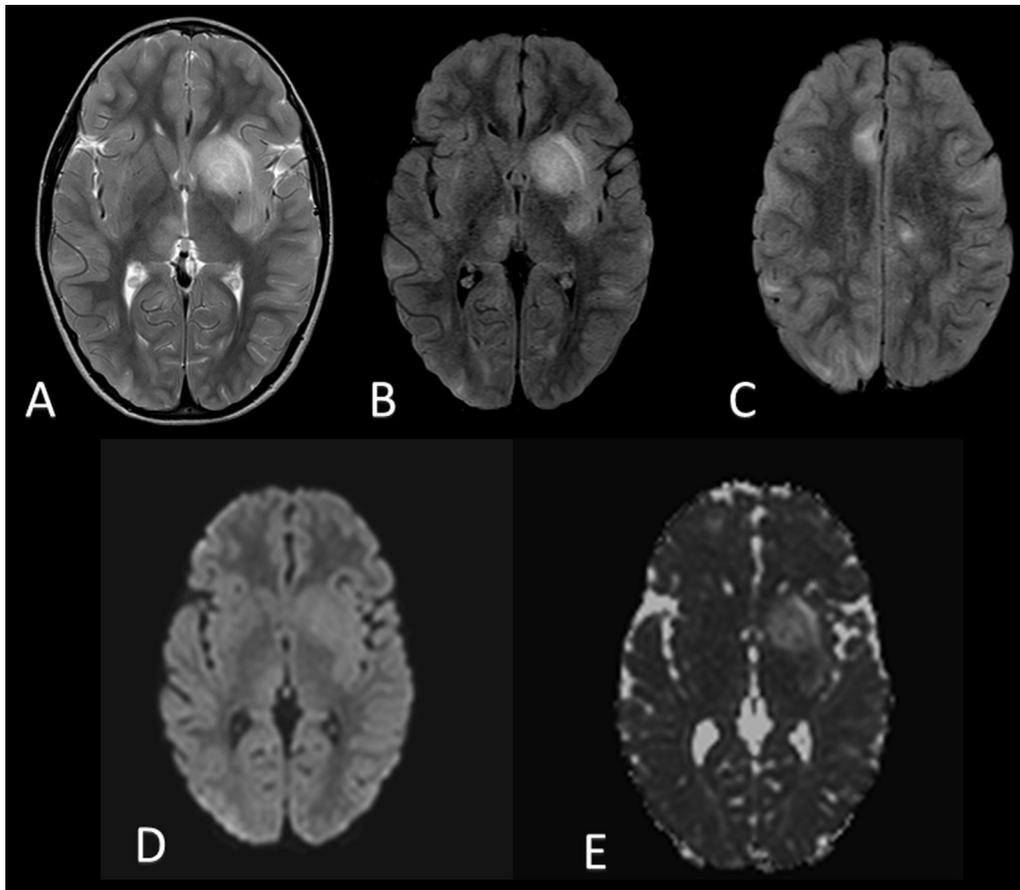
Progressive extremity weakness with sensory sparing has been well reported in the literature. As with brain imaging, spinal MRI may be normal enforcing the known understanding that negative imaging does not exclude myeloradiculitis. When positive, imaging will identify myelitis, polyradiculitis or combination thereof.

Postcontrast imaging may demonstrate diffuse or ventral enhancement of the cauda equina and lumbosacral nerve roots supporting the presence of polyradiculitis.<sup>62,82</sup> While Guillain-Barré syndrome was initially cited as the causative process, clinical, laboratory, and electrodiagnostic features typically do not support such a diagnosis despite imaging similarity.

Primary myeloradiculitis with anterior horn cell involvement and extension into the spinal nerve roots consistent with poliomyelitis has been accepted as the primary manner in which paralysis occurs.<sup>83,84</sup> Many authors have described imaging predilection for the anterior horns of the central gray matter which may demonstrate long segments of contiguous T2 signal



**FIGURE 2.** West Nile encephalitis. (A) T2/FLAIR demonstrating isolated symmetric signal abnormality throughout the basal ganglia and thalami. (B) DWI and ADC sequences demonstrate a lack of corresponding diffusion restriction. Enhancement is not present (not shown). Such findings are typical but not specific for West Nile encephalitis. A wide variety of viral encephalitides may present with similar imaging findings. ADC, apparent diffusion coefficient; DWI, diffusion-weighted imaging; T2/FLAIR, T2 fluid-attenuated inversion recovery.



**FIGURE 3.** ADEM. (A) T2, (B) T2/FLAIR, and (C) T2/FLAIR sequences show patchy regions of signal abnormality asymmetrically involving the left basal ganglia, right thalamus, and cortical-subcortical supratentorium. (D) DWI and (E) ADC sequences demonstrate lack of diffusion restriction. While restricted diffusion and enhancement is not present (not shown) in this case, such findings may occur. Note that viral encephalitis and ADEM may be impossible to differentiate by imaging alone. ADC, apparent diffusion coefficient; ADEM, acute disseminated encephalomyelitis; DWI, diffusion-weighted imaging; T2/FLAIR, T2 fluid-attenuated inversion recovery.

abnormality with or without enhancement consistent with this pathophysiology.<sup>48,85</sup> In cross-section, this has been sometimes referred to as the “owl-eye sign.” Pathologically, these imaging findings correlate with perivascular lymphocytic infiltration, microglial nodules, and neuronal loss.<sup>86</sup> Such a pattern has been described with a variety of viral myelitides presenting with motor weakness to include polio and nonpolio enterovirus (enterovirus D68, enterovirus 71), dengue, St. Louis encephalitis, Powassan, eastern equine encephalitis, Murray Valley encephalitis, and rabies myelitis, among others.<sup>68,87-90</sup> Bilateral signal abnormality of the anterior horns of the spinal cord may be seen in noninfectious causes such as spinal cord ischemia and spinal muscular atrophy.

### Laboratory testing

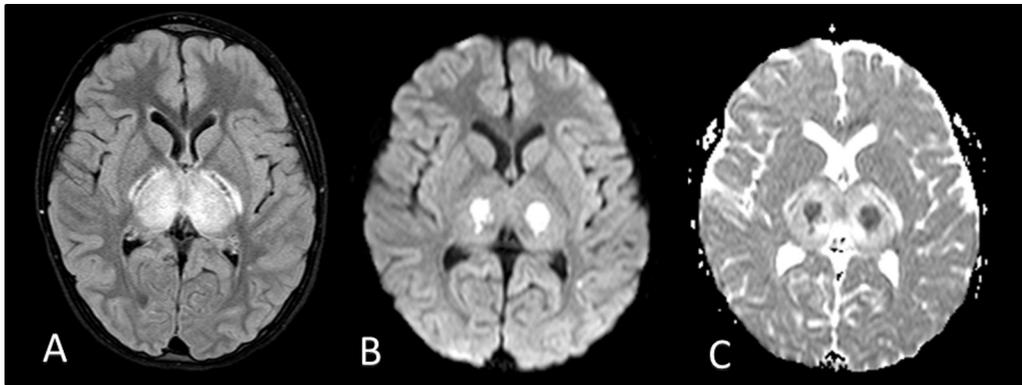
Laboratory diagnosis should be initiated by testing the CSF and serum with WNV-specific IgM immunoassays. IgM antibodies can first be identified as early as three to eight days after disease onset; thus the absence of WNV-specific IgM does not rule out infection if the sample is tested early in the clinical course. Additionally, the presence of IgM antibodies is not sufficient for confirmation of disease. [Table 2](#) lists the criteria for probable and confirmed diagnoses of arboviruses. While these assays serve as readily available screening tools, additional steps must be taken to rule out past WNV infection or infection with another flavivirus. IgM antibodies can persist for up to 90 days after infection, and longer in rare cases. Previous infection can therefore be ruled out by examining acute

and convalescent phase serum antibody titers, with a fourfold increase in titers indicating acute infection. Because antibody immunoassays can cross-react with other flaviviruses, a sample with a positive IgM result should be sent to the CDC or state health department for confirmatory testing. The plaque reduction neutralizing test, performed at the CDC and some state health departments, can differentiate among flavivirus infections. Nucleic acid detection methods and virus isolation techniques may be useful in certain cases, such as in immunocompromised patients in whom antibody production may be delayed or absent.<sup>91</sup>

### Management

#### Prevention

Because there is currently no effective treatment for WNV infection, prevention by limiting mosquito exposure is paramount. Breeding can be reduced by eliminating bodies of free-standing water or treating with larvicide.<sup>92</sup> Adult mosquito abatement occurs at the community level through the distribution of insecticide via ground or aerial spraying. These measures can be quite effective in reducing the mosquito population; aerial spraying, for instance, leads to a 90% reduction in the number of mosquitoes, and was associated with a significant drop in the number of WNNND cases in the Dallas-area outbreak of 2012.<sup>93,94</sup> At the personal level, prevention measures include reducing skin exposure and applying insect repellent when in areas of high mosquito activity.<sup>91</sup> The



**FIGURE 4.** Acute Necrotizing encephalitis (ANE). (A) T2/FLAIR demonstrating signal abnormality within both thalami extending into the diencephalon (not shown) with regions of hemorrhagic necrosis. (B) DWI and (C) ADC show diffusion restriction in both thalami. Findings are characteristic of infectious/ parainfectious ANE in this influenza A positive patient. ADC, apparent diffusion coefficient; DWI, diffusion-weighted imaging; T2/FLAIR, T2 fluid-attenuated inversion recovery.

efficacy and duration of repellent depends on several factors, including the type and concentration of the repellent, as well as the type of mosquito. In general, *N,N*-diethyl-meta-toluamide provides 95% to 100% protection against all mosquito species. Of note, *Culex* mosquitoes tend to be sensitive to all commonly used repellents. Per the CDC, insect repellents can be used in infants as young as two months; however, the natural repellent oil of lemon eucalyptus should not be used in children under three years old.<sup>95</sup>

**Treatment**

Most patients with WNV infection do not require treatment since illness is typically mild and self-limiting. Among patients with severe disease, supportive care is the mainstay of management. Several disease-targeted therapies have been used with variable results (Table 3). Our literature search identified four pediatric cases describing treatment outcome. Two of these patients presented with AFP and were treated with intravenous immune globulin (IVIG); one recovered fully, while the other recovered with only a residual foot drop.<sup>59,96</sup> Two other children presented with encephalitis following chemotherapy. One was treated with ribavirin and had full recovery, while the fourth patient died despite treatment with IVIG, corticosteroids, and Omr-IgG-am, which is an IVIG containing WNV antibodies.<sup>65,97</sup>

Omr-IgG-am was produced after a 2001 report of an adult patient with WNE who had a dramatic response to IVIG produced in Israel, incidentally found to contain high titers of WNV-specific antibodies, while the IVIG from other geographic locations

assessed had none.<sup>98</sup> The drug has subsequently been used in other cases (Table 3) and in a clinical trial comparing it to Polygam, manufactured in the United States, and to placebo (Table 4). The trial, however, was terminated early due to low enrollment and expiration of the Omr-IgG-am product. The preliminary data showed no difference in severity of outcomes among the treatment and control groups.<sup>57</sup> Similarly, transfusion with fresh frozen plasma with WNV IgG has been postulated to ameliorate or prevent clinical disease in two cases, but there have not been any controlled trials.<sup>36,40</sup>

A monoclonal antibody known as MGAWN1 was developed to target the viral E protein, which is vital for fusion with the host cell. The antibody was found to be safe in a phase I trial, but the phase II trial to determine efficacy was terminated early due to poor enrollment.<sup>99,100</sup> Other pharmacologic treatments that have been used include nonspecific antiviral agents and immunomodulators such as ribavirin, interferon alfa-2b, and corticosteroids.

**Prognosis**

Children with WNNND have a better prognosis than older adults, with a fatality rate of 1% compared with 14% in older adults.<sup>7</sup> Some of this difference may be due to a lower rate of comorbidities in the pediatric population, and a lower incidence of encephalitis among children, given that WNE generally has a poor outcome, even when controlling for age.<sup>116</sup> That being stated, children who do develop WNE remain at risk of death or severe neurological sequelae.<sup>7</sup> While data specific to the pediatric

**TABLE 2.**

Diagnostic Criteria for Neuroinvasive Arboviral Disease, Adapted From the Council of State and Territorial Epidemiologists Position Statements. Both Clinical and Laboratory Criteria Must Be Present<sup>91</sup>

Clinical criteria	<ul style="list-style-type: none"> <li>• Fever AND</li> <li>• Meningitis, encephalitis, acute flaccid paralysis, or other acute signs of central or peripheral neurological dysfunction, AND</li> <li>• Absence of a more likely clinical explanation</li> </ul>	
	Confirmed	Probable
Laboratory criteria	<ul style="list-style-type: none"> <li>• Isolation of virus or identification of viral antigen or nucleic acid from tissue or bodily fluid OR</li> <li>• Four-fold or greater increase in virus-specific quantitative antibody titers in paired sera OR</li> <li>• Virus-specific IgM antibodies in serum with confirmatory virus-specific neutralizing antibodies OR</li> <li>• Virus-specific IgM antibodies in CSF and a negative result for other endemic arbovirus IgM antibodies in CSF</li> </ul>	<ul style="list-style-type: none"> <li>• Virus-specific IgM antibodies in CSF or serum with no other testing</li> </ul>

Abbreviations:

CSF = cerebrospinal fluid

CTX = chemotherapy

**TABLE 3.**  
Treatment and Outcome of WNV Infections in Children and Adults. Partially Adapted From the CDC Literature Review for Providers<sup>101</sup>

Therapy	Clinical Presentation	Outcome
<i>Pediatric cases</i>		
IVIG <sup>59,96</sup>	2 AFP	1 full recovery, 1 residual foot drop
RBV <sup>97</sup>	1 WNE post-CTX	Full recovery
IVIG+CS+WNIG <sup>65</sup>	1 WNE post-CTX	Death
<i>Adult cases</i>		
IVIG <sup>33,39,102</sup>	2 AFP	1 no change, 1 partial recovery
WNIG <sup>103-108</sup>	2 WNE postorgan transplant	2 full recovery
	10 WNE	7 full recovery, 3 deaths
	2 postorgan transplant	1 full recovery, 1 death
	1 post-CTX	Death
	1 CLL, not receiving CTX	Full recovery
	6 no pertinent medical history	5 full recovery, 1 death
IFN <sup>3,109-111</sup>	2 AFP	2 full recovery
	1 myalgia after organ transplant	Death
	1 WNE	Full recovery
	2 WNE + unspecified weakness	2 partial recovery
	1 WNE + AFP	Death
CS <sup>112,113</sup>	3 WNE	1 full recovery, 2 partial recovery
	1 WNE	Partial recovery
WNP+WNIG <sup>40</sup>	1 WNE + AFP	Partial recovery
	1 asymptomatic postorgan transplant	Remained asymptomatic
IVIG+IFN <sup>36</sup>	2 organ donor-derived WNE	2 deaths
IVIG+WNP+IFN <sup>36</sup>	1 organ donor-derived WNE	Full recovery
IVIG+RBV <sup>36</sup>	1 organ donor-derived, fever with urinary tract infection, no neurological symptoms	Did not develop neurological symptoms

## Abbreviations:

AFP = acute flaccid paralysis  
 CLL = chronic lymphocytic leukemia  
 CS = corticosteroids  
 CTX = chemotherapy  
 IFN = interferon alfa-2b  
 IVIG = intravenous immunoglobulin  
 RBV = ribavirin  
 WNE = West Nile encephalitis  
 WNIG = high titer WNV-IVIG  
 WNP = plasma with WNV IgG

population regarding the likelihood of full recovery after WNE are unavailable, long-term outcomes have been studied in patients of all ages. Following the WNV outbreak in New York City in 1999, only 37% of patients with WNE had full recovery one year later.<sup>117</sup> The largest study to date examining long-term outcomes found that the majority of patients with West Nile fever or meningitis without encephalitis did not have any neurological deficits one to three years after infection. However, 86% of patients with WNE had an abnormal neurological examination, most frequently involving abnormal tandem gait, motor weakness, and hearing loss. At 10 years postinfection, 20% of patients had died, all but one of whom had been diagnosed with encephalitis.<sup>116</sup> The clinical course of WNNND generally seems to be monophasic, although cases of recurrent weakness in AFP have been reported.<sup>118</sup>

**Future directions***Vaccination*

Much of the current research into WNV therapy is focused on the development of vaccines. While equine vaccines are commercially available, there is currently no equivalent for humans. A number of vaccines have been evaluated in various phases of clinical trials. One recombinant vaccine in particular, ChimeriVax-WN02, has been found safe and effective in two phase II clinical trials among healthy adults.<sup>119</sup> However, due to the relatively low incidence and sporadic nature of WNV outbreaks, completing later phase clinical trials can prove difficult.<sup>120</sup>

**TABLE 4.**  
Therapeutic Trials for Human WNV Infections

Trial	Number Per Treatment Group	Clinical Presentation	Outcome
IVIG vs. WNIG vs. placebo <sup>114</sup>	11 IVIG, 33 WNIG, 11 placebo	WNE or AFP	No significant difference among treatment groups (early termination, incomplete data published)
MGAWN1 vs. placebo <sup>100</sup>	4 MGAWN1, 6 placebo	Unavailable data	MGAWN1: improvement (2), death (2) Placebo: improvement (3), death (1)
IFN vs. placebo <sup>115</sup> (Abstract poster)	15 IFN, 8 placebo	WNE	Significantly greater improvement in treated group than control (Incomplete data)

## Abbreviations:

AFP = acute flaccid paralysis  
 IFN = interferon alfa-2b  
 IVIG = intravenous immunoglobulin  
 MGAWN1 = recombinant humanized monoclonal antibody  
 WNE = West Nile encephalitis  
 WNIG = high titer WNV-IVIG

## Targeted therapies

Regarding therapeutic options for those infected with the virus, several viral components have been identified as potential targets for drug development. A number of these approaches have proven successful in other viruses and show promise in treating WNV. Nucleoside inhibitors targeting RNA-dependent RNA polymerase have proven efficacious in human immunodeficiency virus, herpesvirus, and hepatitis B and C.<sup>121</sup> Favipiravir is a nucleoside inhibitor already licensed in Japan to treat influenza and has demonstrated efficacy in mouse models infected with lethal doses of WNV, but has not yet been trialed in humans with WNV.<sup>122</sup> The development status of this and numerous other compounds are detailed elsewhere and provide promise for the future pharmacological treatment and prevention of severe disease.<sup>121</sup>

## Conclusions

While pediatric WNNND is an uncommon manifestation of a relatively common viral infection, further research is needed given its potential poor prognosis with tendency toward permanent neurological deficits or even death. Larger studies evaluating risk factors for the development of neuroinvasive disease in children and the potential transmission via pregnancy, breast milk, and organ transplantation are needed. Moreover, the development of effective vaccine and treatment may help prevent fatalities and neurological disability among thousands of individuals affected by WNNND worldwide. Practitioners should consider WNNND in the differential diagnosis of meningoencephalitis or AFP, especially during periods of high mosquito activity.

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## References

- Sejvar JJ. West Nile virus: an historical overview. *Ochsner J*. 2003;5:6–10.
- Centers for Disease Control and Prevention. West Nile virus disease cases and presumptive viremic blood donors reported to ArboNET, United States, 2003. <https://www.cdc.gov/westnile/resources/pdfs/data/2003WNVHumanInfectionsbyState.pdf>; 2003. accessed December 11, 2017.
- Sayao AL, Suchowersky O, Al-Khathaami A, et al. Calgary experience with West Nile virus neurological syndrome during the late summer of 2003. *Can J Neurol Sci*. 2004;31:194–203. <https://doi.org/10.1017/S031716710005383X>.
- Centers for Disease Control and Prevention. West Nile virus disease cases and presumptive viremic blood donors reported to ArboNET, United States, 2012. <https://www.cdc.gov/westnile/resources/pdfs/data/2012WNVHumanInfectionsbyState.pdf>; 2012. accessed December 11, 2017.
- Sejvar JJ. The long-term outcomes of human West Nile virus infection. *Emerg Infect Dis*. 2007;44:1617–1624. <https://doi.org/10.1086/518281>.
- Centers for Disease Control and Prevention. West Nile virus—final cumulative maps & data for 1999–2016. <https://www.cdc.gov/westnile/statsmaps/cumMapsData.html>; 2016.
- Lindsey NP, Hayes EB, Staples JE, Fischer M. West Nile virus disease in children, United States, 1999–2007. *Pediatrics*. 2009;123:1084–1089. <https://doi.org/10.1542/peds.2008-3278>.
- Gaensbauer JT, Lindsey NP, Messacar K, Staples JE, Fischer M. Neuroinvasive arboviral disease in the United States: 2003 to 2012. *Pediatrics*. 2014;134:e642–e650. <https://doi.org/10.1542/peds.2014-0498>.
- Centers for Disease Control and Prevention. Symptoms, diagnosis, and treatment 2017. <https://www.cdc.gov/westnile/symptoms/index.html> (accessed October 4, 2018).
- Bewick S, Augusto F, Calabrese JM, Muturi EJ, Fagan WF. Epidemiology of la crosse virus emergence, Appalachia Region, United States. *Emerg Infect Dis*. 2016;22:1921–1929. <https://doi.org/10.3201/eid2211.160308>.
- Reisen WK. Ecology of West Nile virus in North America. *Viruses*. 2013;5:2079–2105. <https://doi.org/10.3390/v5092079>.
- Defelice NB, Little E, Campbell SR, Shaman J. Ensemble forecast of human West Nile virus cases and mosquito infection rates. *Nat Commun*. 2017;8:1–6. <https://doi.org/10.1038/ncomms14592>.
- LaBeaud A, Lisgaris M, King C, Mandalakas A. Pediatric West Nile virus infection: Neurologic disease presentations during the 2002 epidemic in Cuyahoga County, Ohio. *Pediatr Infect Dis J*. 2006;25:751–753. <https://doi.org/10.1097/01.inf.0000226840.84196.0b>.
- Civen R, Villacorte F, Robles DT, et al. West Nile virus infection in the pediatric population. *Pediatr Infect Dis J*. 2006;25:75–78.
- Jean CM, Honarmand S, Louie JK, Glaser CA. Risk factors for West Nile virus neuroinvasive disease, California, 2005. *Emerg Infect Dis*. 2007;13:1918–1920. <https://doi.org/10.3201/eid1312.061265>.
- Levi ME, Curtis DJ. West Nile infections in pediatric solid organ transplant recipients. *Pediatr Transplant*. 2016;20:744–746. <https://doi.org/10.1111/ptr.12710>.
- Lindsey NP, Staples JE, Lehman JA, Fischer M. Medical risk factors for severe West Nile Virus disease, United States, 2008–2010. *Am J Trop Med Hyg*. 2012;87:179–184. <https://doi.org/10.4269/ajtmh.2012.12-0113>.
- Patel H, Sander B, Nelder MP. Long-term sequelae of West Nile virus-related illness: a systematic review. *Lancet Infect Dis*. 2015;15:951–959. [https://doi.org/10.1016/S1473-3099\(15\)00134-6](https://doi.org/10.1016/S1473-3099(15)00134-6).
- Bennett RS, Cress CM, Ward JM, Firestone CY, Murphy BR, Whitehead SS. La Crosse virus infectivity, pathogenesis, and immunogenicity in mice and monkeys. *Virology*. 2008;5:1–15. <https://doi.org/10.1186/1743-422X-5-25>.
- Winkler CW, Myers LM, Woods TA, Carmody AB, Taylor KG, Peterson KE. Lymphocytes have a role in protection, but not in pathogenesis, during La Crosse Virus infection in mice. *J Neuroinflammation*. 2017;14:1–14. <https://doi.org/10.1186/s12974-017-0836-3>.
- Kleinschmidt-DeMasters B. West Nile virus encephalitis 16 years later. *Brain Pathol*. 2015;25:625–633.
- Petersen LR, Brault AC, Nasci RS. West Nile virus: review of the literature. *Jama*. 2013;310:308–315. <https://doi.org/10.1001/jama.2013.8042>.
- Davis LE, DeBiasi R, Goade DE, et al. West Nile virus neuroinvasive disease. *Ann Neurol*. 2006;60:286–300. <https://doi.org/10.1002/ana.20959>.
- Wilson CA, Bale JF. West Nile virus infections in children. *Curr Infect Dis Rep*. 2014;16. <https://doi.org/10.1007/s11908-014-0391-3>.
- Suen WW, Prow NA, Hall RA, Bielefeldt-Ohmann H. Mechanism of west nile virus neuroinvasion: a critical appraisal. *Viruses*. 2014;6:2796–2825. <https://doi.org/10.3390/v6072796>.
- Pealer LN, Marfin AA, Petersen LR, et al. Transmission of West Nile virus through blood transfusion in the United States in 2002. *N Engl J Med*. 2003;349:1236–1245. <https://doi.org/10.1056/NEJMoa030969>.
- Dodd RY, Foster GA, Stramer SL. Keeping blood transfusion safe from West Nile virus: American Red Cross experience, 2003 to 2012. *Transfus Med Rev*. 2015;29:153–161. <https://doi.org/10.1016/j.tmr.2015.03.001>.
- Groves JA, Shafi H, Nomura JH, et al. A probable case of West Nile virus transfusion transmission. *Transfusion*. 2017;57:850–856. <https://doi.org/10.1111/trf.14018>.
- Food and Drug Administration Center for Biologics Evaluation and Research. Guidance for industry: Use use of nucleic acid tests to reduce the risk of transmission of West Nile Virus from donors of whole blood and blood components intended for transfusion. 2009.
- Kleinman SH, Williams JD, Robertson G, et al. Criteria for triggering individual-donation nucleic acid testing. *Transfusion*. 2009;49:1160–1170. <https://doi.org/10.1111/j.1537-2995.2009.02127.x>.
- Kumar D, Drebot MA, Wong SJ, et al. A seroprevalence study of West Nile virus infection in solid organ transplant recipients. *Am J Transplant*. 2004;4:1883–1888. <https://doi.org/10.1111/j.1600-6143.2004.00592.x>.
- Cushing MM, Brat DJ, Mosunjac MI, et al. Fatal West Nile Virus encephalitis in a renal transplant recipient. *Am J Clin Pathol*. 2004;121:26–31. <https://doi.org/10.1309/G23C-P54D-AR1B-CY8L>.
- Saqui R, Randall H, Chandrakantan A, Spak CW, Barri YM. West Nile virus encephalitis in a renal transplant recipient: the role of intravenous immunoglobulin. *Am J Kidney Dis*. 2008;52:19–21. <https://doi.org/10.1053/j.ajkd.2008.03.042>.
- Lambert SL, Aviles D, Vehaskari VM, Ashoor IF. Severe West Nile virus meningoencephalitis in a pediatric renal transplant recipient: successful recovery and long-term neuropsychological outcome. *Pediatr Transplant*. 2016;20:836–839. <https://doi.org/10.1111/ptr.12768>.
- Wilson MR, Zimmermann LL, Crawford ED, et al. Acute West Nile virus meningoencephalitis diagnosed via metagenomic deep sequencing of cerebrospinal fluid in a renal transplant patient. *Am J Transplant*. 2017;17:803–808. <https://doi.org/10.1111/ajt.14058>.
- Winston DJ, Vikram HR, Rabe IB, et al. Donor-derived West Nile virus infection in solid organ transplant recipients. *Transplantation*. 2014;97:881–889. <https://doi.org/10.1097/TP.000000000000024>.
- Rabe IB, Schwartz BS, Farnon EC, et al. Fatal transplant-associated West Nile virus encephalitis and public health investigation—California, 2010. *Transplantation*. 2013;96:463–468. <https://doi.org/10.1097/TP.0b013e31829b4142.Fatal>.
- Inojosa WO, Scottton PG, Fuser R, et al. West Nile virus transmission through organ transplantation in north-eastern Italy: a case report and implications for pre-procurement screening. *Infection*. 2012;40:557–562. <https://doi.org/10.1007/s15101-012-0263-4>.
- Rhee C, Eaton EF, Concepcion W, Blackburn BG. West Nile virus encephalitis acquired via liver transplantation and clinical response to intravenous immunoglobulin: Case report and review of the literature. *Transp Infect Dis*. 2011;13:312–317. <https://doi.org/10.1111/j.1399-3062.2010.00595.x>.

40. Morelli MC, Sambri V, Grazi GL, et al. Absence of neuroinvasive disease in a liver transplant recipient who acquired West Nile virus (WNV) infection from the organ donor and who received WNV antibodies prophylactically. *Clin Infect Dis*. 2010;51:e34–e37. <https://doi.org/10.1086/655146>.
41. Iwamoto M, Jernigan D, Guasch A, et al. Transmission of West Nile virus from an organ donor to four transplant recipients. *N Engl J Med*. 2003;348:2196–2203.
42. Razonable RR. Management of viral infections in solid organ transplant recipients. *Expert Rev Anti Infect Ther*. 2011;9:685–700. <https://doi.org/10.1586/eri.11.43>.
43. Centers for Disease Control and Prevention. Possible West Nile virus transmission to an infant through breast-feeding—Michigan, 2002. *MMWR Morb Mortal Wkly Rep*. 2002;51:877–878.
44. Hinckley AF, O'Leary DR, Hayes EB. Transmission of West Nile virus through human breast milk seems to be rare. *Pediatrics*. 2007;119:e666–e671. <https://doi.org/10.1542/peds.2006-2107>.
45. O'Leary DR. Birth outcomes following West Nile virus infection of pregnant women in the United States: 2003–2004. *Pediatrics*. 2006;117:e537–e545. <https://doi.org/10.1542/peds.2005-2024>.
46. Pridjian G, Sirois PA, McRae S, et al. Prospective study of pregnancy and newborn outcomes in mothers with West Nile illness during pregnancy. *Birth Defects Res A Clin Mol Teratol*. 2016;106:716–723. <https://doi.org/10.1002/bdra.23523>.
47. Smith JC, Mailman T, MacDonald NE. West Nile virus: should pediatricians care? *J Infect*. 2014;69:S70–S76. <https://doi.org/10.1016/j.jinf.2014.07.019>.
48. Leis AA, Stokic DS. Neuromuscular manifestations of West Nile virus infection. *Front Neurol*. 2012;3:37. <https://doi.org/10.3389/fneur.2012.00037>.
49. Chahil M, Nguyen TP. West Nile virus-associated brachial plexopathy. *BMJ Case Rep*. 2016. <https://doi.org/10.1136/bcr-2016-214428>.
50. Natarajan N, Varman M. West Nile virus cerebellitis in a healthy 10-year-old child. *Pediatr Infect Dis J*. 2007;26:767.
51. Braun LRE, Tsuchida T, Spiegel H. Meningoencephalitis in a child complicated by myocarditis, quadripareisis and respiratory failure. *Pediatr Infect Dis J*. 2006;25. <https://doi.org/10.1097/01.inf.0000234058.31683.70>.
52. Hainline ML, Kincaid JC, Carpenter DL, Golomb MR. West Nile poliomyelitis in a 7-Year-old child. *Pediatr Neurol*. 2008;39:350–354. <https://doi.org/10.1016/j.pediatrneurol.2008.07.027>.
53. Yim R, Posfay-Barbe KM, Nolt D, Fatula G, Wald ER. Spectrum of clinical manifestations of West Nile virus infection in children. *Pediatrics*. 2004;114:1673–1675. <https://doi.org/10.1542/peds.2004-0491>.
54. Huhn GD, Dworkin MS. Rash as a prognostic factor in West Nile virus disease. *Clin Infect Dis*. 2006;43:388–389.
55. Arnold JC, Revivo GA, Senac MO, Leake JAD. West Nile virus encephalitis with thalamic involvement in an immunocompromised child. *Pediatr Infect Dis J*. 2005;24:932–934. <https://doi.org/10.1097/01.inf.0000180972.63966.35>.
56. Messacar K, Cree-Green M, Lovell M, Anderson MS, Dominguez SR. Severe neuroinvasive West Nile virus infection in a child with undiagnosed Addison's disease. *IDCases*. 2014;1:29–31. <https://doi.org/10.1016/j.idcr.2014.04.001>.
57. Hart J, Tillman G, Kraut MA, et al. West Nile virus neuroinvasive disease: neurological manifestations and prospective longitudinal outcomes. *BMC Infect Dis*. 2014;14:248. <https://doi.org/10.1186/1471-2334-14-248>.
58. Hayes EB, O'Leary DR. West Nile virus infection: a pediatric perspective. *Pediatrics*. 2004;113:1375–1381. <https://doi.org/10.1542/peds.113.5.1375>.
59. Thabet FI, Servinsky SE, Naz F, Kovas TE, Raghbi TO. Unusual case of West Nile Virus flaccid paralysis in a 10-year-old child. *Pediatr Neurol*. 2013;48:393–396. <https://doi.org/10.1016/j.pediatrneurol.2012.12.017>.
60. Heresi G, Mancias P, Mazar L, Butler I, Murphy J, Cleary T. Poliomyelitis-like syndrome in a child with West Nile virus infection. *Pediatr Infect Dis J*. 2004;23:788–789.
61. Nash D, Mostashari F, Fine A, et al. The outbreak of West Nile virus infection in the New York City area in 1999. *N Engl J Med*. 2001;344:1807–1814. doi: 14;344(24):1807-14.
62. Ali M, Safriel Y, Sohi J, Llave A, Weathers S. West Nile virus infection: MR imaging findings in the nervous system. *AJNR Am J Neuroradiol*. 2005;26:289–297.
63. Lyons JL, Schaefer PW, Cho TA, Azar MM. Case 34–2017. A 76-year-old man with fever, weight loss, and weakness. *N Engl J Med*. 2017;377:1878–1886. <https://doi.org/10.1056/NEJMcpc1707557>.
64. Mandel JJ, Tummala S, Woodman KH, Tremont-Lukats I. Delayed imaging abnormalities of neuro-invasive West Nile virus in cancer patients. *J Neurol Sci*. 2015;350:115–117. <https://doi.org/10.1016/j.jns.2015.02.014>.
65. Hindo H, Buescher ES, Frank LM, Pettit D, Dory C, Byrd R. West Nile virus infection in a teenage boy with acute lymphocytic leukemia in remission. *J Pediatr Hematol Oncol*. 2005;27:659–662.
66. Petropoulou KA, Gordon SM, Prayson RA, Ruggieri PM. West Nile virus meningoencephalitis: MR imaging findings. *AJNR Am J Neuroradiol*. 2005;26:1986–1995.
67. Rosas H, 2nd Wippold FJ. West Nile virus: case report with MR imaging findings. *AJNR Am J Neuroradiol*. 2003;24:1376–1378.
68. Kraushaar G, Patel R, Stoneham GW. West Nile Virus: a case report with flaccid paralysis and cervical spinal cord: MR imaging findings. *AJNR Am J Neuroradiol*. 2005;26:26–29.
69. Cerna F, Mehrad B, Luby JP, Burns D, Fleckenstein JL. St. Louis encephalitis and the substantia nigra: MR imaging evaluation. *AJNR Am J Neuroradiol*. 1999;20:1281–1283.
70. Handique SK. Viral infections of the central nervous system. *Neuroimaging Clin N Am*. 2011;21:777–794. <https://doi.org/10.1016/j.nic.2011.07.012>.
71. Piantadosi A, Rubin DB, McQuillen DP, et al. Emerging cases of Powassan Virus encephalitis in New England: clinical presentation, imaging, and review of the literature. *Clin Infect Dis*. 2016;62:707–713. <https://doi.org/10.1093/cid/civ1005>.
72. Einsiedel L, Kat E, Ravindran J, Slavotinek J, Gordon DL. MR findings in Murray Valley encephalitis. *Am J Neuroradiol*. 2003;24:1379–1382.
73. Deresiewicz RL, Thaler SJ, Hsu L, Zamani AA. Clinical and neuroradiographic manifestations of Eastern Equine encephalitis. *N Engl J Med*. 1997;336:1867–1874. <https://doi.org/10.1056/NEJM199706263362604>.
74. Jugpal TS, Dixit R, Garg A, et al. Spectrum of findings on magnetic resonance imaging of the brain in patients with neurological manifestations of dengue fever. *Radiol Bras*. 2017;50:285–290. <https://doi.org/10.1590/0100-3984.2016.0048>.
75. Ganesan K, Diwan A, Shankar SK, Desai SB, Sainani GS, Katrak SM. Chikungunya encephalomyelioradiculitis: report of 2 cases with neuroimaging and 1 case with autopsy findings. *AJNR Am J Neuroradiol*. 2008;29:1636–1637. <https://doi.org/10.3174/ajnr.A1133>.
76. Chusri S, Siripaitoon P, Hirunpat S, Silpapojakul K, Siripaitoon P, Chusri S. Case reports of neuro-chikungunya in Southern Thailand. *Am J Trop Med Hyg*. 2011;85:386–389. <https://doi.org/10.4269/ajtmh.2011.10-0725>.
77. Zare Mehrjardi M, Carteaux G, Poretti A, et al. Neuroimaging findings of postnatally acquired Zika virus infection: a pictorial essay. *Jpn J Radiol*. 2017;35:341–349. <https://doi.org/10.1007/s11604-017-0641-z>.
78. Soares BP, Provenzale JM. Imaging of herpesvirus infections of the CNS. *AJR Am J Roentgenol*. 2016;206:39–48. <https://doi.org/10.2214/AJR.15.15314>.
79. Daxboeck F, Blacky A, Seidl R, Krause R, Assadian O. Diagnosis, treatment, and prognosis of Mycoplasma pneumoniae childhood encephalitis: systematic review of 58 cases. *J Child Neurol*. 2004;19:865–871. <https://doi.org/10.1177/08830738040190110401>.
80. Sampson BA, Ambrosi C, Charlot A, Reiber K, Veress JF, Armbrustmacher V. The pathology of human West Nile virus infection. *Hum Pathol*. 2000;31:527–531.
81. Cushing MM, Brat DJ, Mosunjac MI, et al. Fatal West Nile virus encephalitis in a renal transplant recipient. *Am J Clin Pathol*. 2004;121:26–31. <https://doi.org/10.1309/G23C-P54D-AR1B-CY8L>.
82. Patel CB, Trikambji B, Mathisen G, Yim C, Zipser B, Mishra S. MRI ventral nerve root enhancement in five patients presenting with extremity weakness secondary to neuroinvasive West Nile virus. *J Clin Neuromuscul Dis*. 2016;18:41–43. <https://doi.org/10.1097/01.cnd.0000496973.95654.fcd>.
83. Jeha LE, Sila CA, Lederman RJ, Prayson RA, Isada CM, Gordon SM. West Nile virus infection: A new acute paralytic illness. *Neurology*. 2003;61:55–59.
84. Sejvar JJ, Haddad MB, Tierney BC, Campbell GL, Van Gerpen JA, Petersen LR. Neurologic manifestations and outcome of West Nile virus infection. *JAMA*. 2003;290:511–515.
85. Doron SI, Dashe JF, Adelman LS, Brown WF, Werner BG, Hadley S. Histopathologically proven poliomyelitis with quadriplegia and loss of brainstem function due to West Nile virus infection. *Clin Infect Dis*. 2003;37:e74–e77. <https://doi.org/10.1086/37177>.
86. Guarner J, Shieh W-J, Hunter S, et al. Clinicopathologic study and laboratory diagnosis of 23 cases with West Nile virus encephalomyelitis. *Hum Pathol*. 2004;35:983–990.
87. Davis LE, Beckham JD, Tyler KL. North American encephalitic arboviruses. *Neurol Clin*. 2008;26:727–757. <https://doi.org/10.1016/j.ncl.2008.03.012>.
88. Maloney JA, Mirsky DM, Messacar K, Dominguez SR, Schreiner T, Stence N V. MRI findings in children with acute flaccid paralysis and cranial nerve dysfunction occurring during the 2014 enterovirus D68 outbreak. *AJNR Am J Neuroradiol*. 2015;36:245–250. <https://doi.org/10.3174/ajnr.A4188>.
89. Choudhary A, Sharma S, Sankhyan N, et al. Midbrain and spinal cord magnetic resonance imaging (MRI) changes in poliomyelitis. *J Child Neurol*. 2010;25:497–499. <https://doi.org/10.1177/0883073809340918>.
90. Chen CY, Chang YC, Huang CC, Lui CC, Lee KW, Huang SC. Acute flaccid paralysis in infants and young children with enterovirus 71 infection: MR imaging findings and clinical correlates. *AJNR Am J Neuroradiol*. 2001;22:200–205.
91. Centers for Disease Control and Prevention. West Nile Virus in the United States: Guidelines for Surveillance, Prevention, and Control. 2013.
92. Davis LE, Beckham JD, Tyler KL. Progress on the development of therapeutics against West Nile virus. *Neurol Clin*. 2008;26:1–26. <https://doi.org/10.1016/j.ncl.2008.03.012.North>.
93. Breidenbaugh MS, Haagsma KA, Walker WW, Sanders DM. Post-hurricane Rita mosquito surveillance and the efficacy of Air Force aerial applications for mosquito control in East Texas. *J Am Mosq Control Assoc*. 2008;24:327–330. <https://doi.org/10.2987/5731.1>.
94. Ruktanonchai DJ, Stonecipher S, Lindsey N, et al. Effect of aerial insecticide spraying on West Nile virus disease—north-central Texas, 2012. *Am J Trop Med Hyg*. 2014;91:240–245. <https://doi.org/10.4269/ajtmh.14-0072>.
95. Centers for Disease Control and Prevention. Prevent mosquito bites 2018. <https://www.cdc.gov/zika/prevention/prevent-mosquito-bites.html>.
96. Soldatou A, Vartzelis G, Vorre S, Papa A, Voudris K, Garoufi A. A toddler with acute flaccid paralysis due to West Nile virus infection. *Pediatr Infect Dis J*. 2013;32:1023–1024. <https://doi.org/10.1097/INF.0b013e318292bf72>.
97. Spiegel R, Miron D, Gaviel H, Horovitz Y. West Nile virus meningoencephalitis complicated by motor aphasia in Hodgkin's lymphoma. *Arch Dis Child*. 2002;86:441–442.
98. Shimoni Z, Niven MJ, Pitlick S, Bulvik S. Treatment of West Nile virus encephalitis with intravenous immunoglobulin. *Emerg Infect Dis*. 2001;7:759. <https://doi.org/10.3201/eid0704.010432>.

99. Beigel JH, Nordstrom JL, Pillemer SR, et al. Safety and pharmacokinetics of single intravenous dose of MGAWN1, a novel monoclonal antibody to West Nile virus. *Antimicrob Agents Chemother*. 2010;54:2431–2436. <https://doi.org/10.1128/AAC.01178-09>.
100. MacroGenics. Treatment of West Nile virus with MGAWN1 (PARADIGM). <https://clinicaltrials.gov/ct2/show/NCT00927953>; 2009 (accessed December 1, 2017).
101. Centers for Disease Control and Prevention. Fact sheet: West Nile virus. <https://www.cdc.gov/media/pressrel/GeneralFacts.pdf>; 2000 (accessed December 1, 2017).
102. Li J, Loeb JA, Shy ME, et al. Asymmetric flaccid paralysis: a neuromuscular presentation of West Nile virus infection. *Ann Neurol*. 2003;53:703–710. <https://doi.org/10.1002/ana.10575>.
103. Haley M, Retter AS, Fowler D, Gea-Banacloche J, O'grady NP. The role of intravenous immunoglobulin in the treatment of West Nile virus encephalitis. *Clin Infect Dis*. 2003;37:88–90.
104. Hamdan a, Green P, Mendelson E, Kramer MR, Pitlik S, Weinberger M. Possible benefit of intravenous immunoglobulin therapy in a lung transplant recipient with West Nile virus encephalitis. *Transpl Infect Dis*. 2002;4:160–162. <https://doi.org/10.1034/j.1399-3062.2002.01014.x>.
105. Makhoul B, Braun E, Herskovitz M, Ramadan R, Hadad S, Krivoy N. Hyper-immune gammaglobulin for the treatment of West Nile virus encephalitis. *Isr Med Assoc J*. 2009;11:151–153.
106. Shimoni Z, Bin H, Bulvik S, et al. The clinical response of West Nile virus neuroinvasive disease to intravenous immunoglobulin therapy. *Clin Pract*. 2012;2:18. <https://doi.org/10.4081/cp.2012.e18>.
107. Walid MS, Mahmoud FA. Successful treatment with intravenous immunoglobulin of acute flaccid paralysis caused by west Nile virus. *Perm J*. 2009;13:43–46.
108. Levi ME, Quan D, Ho JT, Kleinschmidt-DeMasters BK, Tyler KL, Grazia TJ. Impact of rituximab-associated B-cell defects on West Nile virus meningo-encephalitis in solid organ transplant recipients. *Clin Transplant*. 2010;24:223–228. <https://doi.org/10.1111/j.1399-0012.2009.01044.x>.
109. Chan-Tack KM, Forrest G. Failure of interferon alpha-2b in a patient with West Nile virus meningoencephalitis and acute flaccid paralysis. *Scand J Infect Dis*. 2005;37:944–946. <https://doi.org/10.1080/00365540500262690>.
110. Kalil AC, Devetten MP, Singh S, et al. Use of interferon-alpha in patients with West Nile encephalitis: report of 2 cases. *Clin Infect Dis*. 2005;40:764–766. <https://doi.org/10.1086/427945>.
111. Lewis M, Amsden JR. Successful treatment of West Nile virus infection after approximately 3 weeks into the disease course. *Pharmacotherapy*. 2007;27:455–458. <https://doi.org/10.1592/phco.27.3.455>.
112. Pyrgos V, Younus F. High-dose steroids in the management of acute flaccid paralysis due to West Nile virus infection. *Scand J Infect Dis*. 2004;36:509–512.
113. Alker A. West Nile virus-associated acute flaccid paralysis. *BMJ Case Rep*. 2015;2015. <https://doi.org/10.1136/bcr-2014-206480>. [bcr2014206480-bcr2014206480](https://doi.org/10.1136/bcr-2014-206480).
114. National Institute of Allergy and Infectious Diseases. IVIG—West Nile encephalitis: Safety and efficacy 2003. <https://clinicaltrials.gov/ct2/show/NCT00068055> (accessed December 1, 2017).
115. Wehbeh W. Treatment of West Nile virus central nervous system infections with interferon alpha-2b. In: 44th ICAAC Meet. *Am. Soc. Microbiol*. 2004.
116. Weatherhead JE, Miller VE, Garcia MN, et al. Long-term neurological outcomes in West Nile virus-infected patients: an observational study. *Am J Trop Med Hyg*. 2015;92:1006–1012. <https://doi.org/10.4269/ajtmh.14-0616>.
117. Klee AL, Maidin B, Edwin B, et al. Long-term prognosis for clinical West Nile virus infection. *Emerg Infect Dis*. 2004;10:1405–1411. <https://doi.org/10.3201/eid1008.030879>.
118. Sejvar J, Davis L, Szabados E, Jackson A. Delayed-onset and recurrent limb weakness associated with West Nile virus infection. *J Neurovirol*. 2010;16:93–100. <https://doi.org/10.3109/13550280903586378>.
119. Biedenbender R, Bevilacqua J, Gregg AM, Watson M, Dayan G. Phase II, randomized, double-blind, placebo-controlled, multicenter study to investigate the immunogenicity and safety of a West Nile virus vaccine in healthy adults. *J Infect Dis*. 2011;203:75–84. <https://doi.org/10.1093/infdis/jiq003>.
120. Dayan GH, Pugachev K, Bevilacqua J, Lang J, Monath TP. Preclinical and clinical development of a YFV 17 D-based chimeric vaccine against West Nile virus. *Viruses*. 2013;5:3048–3070. <https://doi.org/10.3390/v5123048>.
121. Acharya D, Bai F. An overview of current approaches toward the treatment and prevention of West Nile virus infection editor. In: Colpitts TM, ed. *West Nile Virus Methods Protoc.*, 1435 New York, NY: Springer New York; 2016: 249–291. [https://doi.org/10.1007/978-1-4939-3670-0\\_19](https://doi.org/10.1007/978-1-4939-3670-0_19).
122. Escribano-Romero E, De Oya NJ, Domingo E, Saiza JC. Extinction of West Nile virus by favipiravir through lethal mutagenesis. *Antimicrob Agents Chemother*. 2017;61:1–9. <https://doi.org/10.1128/AAC.01400-17>.