

Pathophysiology of Lacunar Stroke: History's Mysteries and Modern Interpretations

Robert W. Regenhardt, MD, PhD,*† Alvin S. Das, MD,*† Ryo Ohtomo, MD,*†
Eng H. Lo, PhD,*† Cenk Ayata, MD,* and Mahmut Edip Gurol, MD*

Since the term “lacune” was adopted in the 1800s to describe infarctions from cerebral small vessels, their underlying pathophysiological basis remained obscure until the 1960s when Charles Miller Fisher performed several autopsy studies of stroke patients. He observed that the vessels displayed segmental arteriolar disorganization that was associated with vessel enlargement, hemorrhage, and fibrinoid deposition. He coined the term “lipohyalinosis” to describe the microvascular mechanism that engenders small subcortical infarcts in the absence of a compelling embolic source. Since Fisher’s early descriptions of lipohyalinosis and lacunar stroke (LS), there have been many advancements in the understanding of this disease process. Herein, we review lipohyalinosis as it relates to modern concepts of cerebral small vessel disease (cSVD). We discuss clinical classifications of LS as well as radiographic definitions based on modern neuroimaging techniques. We provide a broad and comprehensive overview of LS pathophysiology both at the vessel and parenchymal levels. We also comment on the role of biomarkers, the possibility of systemic disease processes, and advancements in the genetics of cSVD. Lastly, we assess preclinical models that can aid in studying LS disease pathogenesis. Enhanced understanding of this highly prevalent disease will allow for the identification of novel therapeutic targets capable of mitigating disease sequelae.

Key Words: Stroke—lacunar stroke—cerebral small vessel disease—genetics
© 2019 Elsevier Inc. All rights reserved.

Introduction and Historical Perspectives

Since the 1800s, lacunes were recognized as small infarcts involving the distal small penetrating branches of larger cerebral arteries. Their cavitated end-products originally seen on autopsy were referred to as “lacunes,” a French word derived from the Latin, *lacuna*, meaning cavity. The first to characterize these lacunes was Amédée

Dechambre, an intern at the Salpêtrière Hospital in Paris.¹ In 1838, he described the pathological findings of a male patient with hemiplegia and stated the following: “a number of small lacunes, of variable size and form, more or less filled with a milky fluid . . . resulting from liquefaction and partial reabsorption in the centre of the softening.

From the *Department of Neurology, Massachusetts General Hospital, Harvard Medical School, Boston, Massachusetts; †Department of Neurology, Brigham and Women’s Hospital, Harvard Medical School, Boston, Massachusetts; and ‡Department of Radiology, Massachusetts General Hospital, Harvard Medical School, Boston, Massachusetts.

Received January 23, 2019; revision received April 13, 2019; accepted May 4, 2019.

Funding for this study was provided by grants from the [National Institutes of Health](#) (R.W.R., NS065743; E.H.L., NS055104, NS099620, NS103786, AG055559, and NS093415; C.A., NS102969; M.E.G., NS083711).

Address correspondence to M. Edip Gurol, MD, Hemorrhagic Stroke Research Program, Department of Neurology, Massachusetts General Hospital, 175 Cambridge Street, Suite 300, Boston, MA 02114. E-mail: edip@mail.harvard.edu.

1052-3057/\$ - see front matter

© 2019 Elsevier Inc. All rights reserved.

<https://doi.org/10.1016/j.jstrokecerebrovasdis.2019.05.006>

ACRONYM DEFINITION

ACE	angiotensin-converting enzyme
ApoE	apolipoprotein E
ASCO	Atherosclerosis, Small-vessel disease, Cardiac pathology, Other causes
ASCOD	Atherosclerosis, Small-vessel disease, Cardiac pathology, Other causes, Dissection
CAA	cerebral amyloid angiopathy
CBF	cerebral blood flow
CCS	Causative Classification of Stroke
CMB	cerebral microbleed
CMI	cerebral microinfarct
cSVD	cerebral small vessel disease
CT	computed tomography
DM	diabetes mellitus
DWI	diffusion-weighted imaging
eNOS	endothelial nitric oxide synthase
ET-1	endothelin-1
GWAS	genome wide association studies
HTN	hypertension
LIMITS	Levels of Inflammatory Markers in the Treatment of Stroke
L-NIO	N5-(1-iminoethyl)-L-ornithine
LS	lacunar stroke
MCAO	middle cerebral artery occlusion
MRI	magnetic resonance imaging
Nogo	neurite outgrowth inhibitor
OPC	oligodendrocyte precursor cell
SHR-SP	stroke-prone Spontaneously Hypertensive Rat
STRIVE	STandards for ReportIng Vascular changes on nEuroimaging
TOAST	Trial of Org 10172 in Acute Stroke Treatment
WMH	white matter hyperintensity

According to the degree of hardening and reabsorption of the liquefied pulp these lacunes appear empty or stay more or less filled ... It is, no doubt, with small foci of partial softening, that these round cavities without membranes should be correlated."^{2,3} Five years later, another French neurologist Maxime Durand-Fardel independently arrived at similar findings.⁴ In his treatise, he noted that "the striatum ... showed a certain number of small lacunes with no associated alteration of color or consistency from whose surface there extended a fine meshwork containing very small vessels."⁵ In later works, he used the term *état criblé* to

describe enlarged spaces in the white matter, which were in a cribriform pattern.^{3,6}

It was not until 1901, when Pierre Marie correctly made a distinction between *état lacunaire* and *état criblé*.^{6,7} Using the collective term *état lacunaire* to describe numerous basal ganglia infarctions, he noted that these lesions were responsible for sudden motor deficits and the characteristic gait, *marche à petits pas de Déjérine*. He astutely observed that the underlying lacunes were created by a "local arteriosclerotic process"⁸ as well as "destructive vaginitis," in which perivascular spaces demolished adjacent areas of brain tissues.⁶ These disease paradigms persisted until the mid-20th century when Charles Miller Fisher performed numerous pathological studies of lacunes.⁹ He used the term lipohyalinosis to describe a unique mechanism that gave rise to such cerebral small vessel disease (cSVD)-related infarcts.^{10,11}

Today, lacunar strokes (LS) comprise up to 20%-30% of all acute ischemic strokes.^{12,13} Although these infarcts are smaller in size compared to those caused by large-vessel occlusions, they were found to be the most common type of acute strokes (37%) to worsen after onset in the Harvard Stroke Registry.¹⁴⁻¹⁶ The term "stuttering" has been used to describe this phenomenon. Stuttering LS are particularly challenging for the clinician as there are limited interventions he or she can implement as patients deteriorate. Furthermore, LS are burdensome given their recurrence rate of up to 20% over 3 years and their resultant functional impairment and disability.¹⁷ Because of their association with cardiovascular disease, one fourth of patients who experience these strokes will die after 5 years.¹⁸ Moreover, there is an increased incidence of LS in younger patients.¹⁹ Estimates suggest that annually 11 million people suffer from silent strokes in the United States alone, the majority of which are due to cSVD,²⁰ showing that the pathology is highly prevalent even without overt symptoms. These asymptomatic infarcts contribute to a large proportion of neurological morbidities including vascular cognitive impairment,²¹ which has been increasing in prevalence.^{18,221} The care of stroke patients is complex²² and while there have been several revolutionary advances in the treatment of large vessel occlusions,²³⁻²⁸ little progress has been made in the targeted treatment of LS.²⁹ However, continued advancements in understanding LS pathogenesis may make targeted cSVD therapeutics a realistic goal.

Clinical and Radiographic Classification Criteria

Clinical LS in the classic sense are caused by infarcts between 5 mm and 15 mm in greatest diameter and involve the deep gray and subcortical white matter (Fig 1). Infarcts above that size in diameter (15-20 mm)

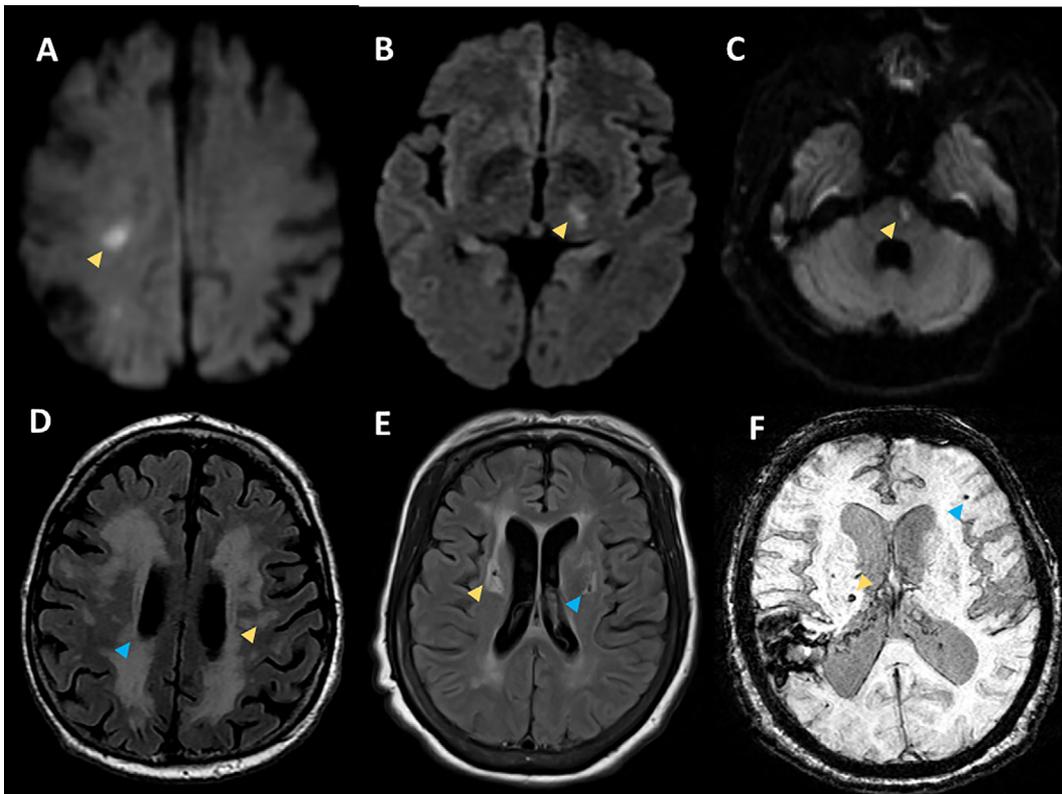


Figure 1. Small vessel disease neuroimaging markers. Small infarcts on DWI sequences within the right subcortical white matter (A), left thalamus (B), and left pons (C) consistent with acute lacunar strokes. (D) T2 FLAIR sequence with white matter hyperintensities in the subcortical white matter (yellow arrow) and periventricular white matter (blue arrow). (E) T2 FLAIR sequence with white matter hyperintensities in the basal ganglia (yellow arrow) and deep chronic lacune with a rim of hyperintensity (blue arrow). (F) Susceptibility-weighted imaging with superficial microbleed (blue arrow) and deep microbleed (yellow arrow).

may be due to involvement of several perforating vessels in the putamino-capsulo-caudate region, and have been referred to by some as “giant” lacunes,³⁰ “lagunes” (a portmanteau for large lacune), and “macunes” (for major lacunes).^{31,32} Not surprisingly, these large LS are more likely to have severe symptoms and disability and may reflect more heterogeneous underlying etiologies including cardioembolism³² or atherosclerotic disease in a major penetrator artery at its origin.³³

There are many classification schemes for LS (Table 1), but perhaps the most widely accepted classification of stroke subtypes was described in the Trial of Org 10172 in Acute Stroke Treatment (TOAST) conducted in 1993.³⁴ In this stroke classification schema, the third subtype of stroke was typified by cSVD. Per TOAST classification, cSVD-related infarcts are generally accompanied by traditional clinical LS syndromes; Fisher described 21 different syndromes.¹¹ A hallmark clinical feature of these syndromes is the absence of cortical signs or symptoms such as aphasia, apraxia, agnosia, visual field cut, memory impairment, stupor, coma, loss of consciousness, or seizures. Perhaps the 5 most notable syndromes are pure motor strokes, pure sensory stroke, ataxia-hemiparesis, dysarthria-clumsy hand, and sensorimotor strokes. In

addition to a LS syndrome, neuroimaging with computed tomography or magnetic resonance imaging (MRI) should be normal or reveal a relevant brain stem or subcortical lesion with a diameter of less than 15 mm. Potential cardiac sources for embolism should be exonerated, and vessel imaging should not demonstrate a stenosis of greater than 50% in an ipsilateral proximal artery. While these criteria have been adopted by many practitioners, some have argued that their strict interpretation underrepresents the total cases of LS.³⁵

The Causative Classification of Stroke was the first computerized, evidence-based algorithm that provided both causative and phenotypic stroke subtypes in a rule-based manner. Through answering a series of questions, the algorithm displays the most likely diagnosis. It was shown to have a high inter-rater reliability and be useful for international multicenter studies.³⁶ Within the algorithm, an LS is defined as being located in the territory of a penetrating artery and should be less than 20 mm in its greatest diameter. There should be no focal pathology in the parent artery at the site of the origin of the penetrating artery.

Another recent classification, the ASCO (A: atherosclerosis; S: small-vessel disease; C: cardiac pathology;

Table 1. Classification schemes for lacunar stroke and cerebral small-vessel disease

LS classification	Assessment	Definition	Reference
TOAST	Clinical	CT/MRI unremarkable or with subcortical lesion <15 mm that explains acute symptoms, no cardioembolism, no ipsilateral large-vessel stenosis >50%	34
CCS	Clinical	CT/MRI with lesion in territory of penetrating vessel and <20 mm that explains acute symptoms, no focal pathology in parent artery	36
ASCOD	Clinical	CT/MRI with small/deep lesion <15 mm that explains acute symptoms, stuttering symptoms, CT/MRI evidence of chronic LS/WMH/CMB/PVS	37
STRIVE	Radiographic	Recent small subcortical infarcts are DWI hyperintensities <20 mm, chronic lacunes are T2 hypointensities with hyperintense rim <15 mm	38
Poirier and Derouesné	Pathologic	Type Ia old infarcts <20 mm with pan-necrotic cavitation and scattered macrophages, Type Ib incomplete infarcts with selective loss of vulnerable elements	92
cSVD disease process	Vessel size (μm)	Definition	Reference
Arteriosclerosis/atherosclerosis	200-800	Microatheromatous plaques with foam cells	11
Arteriolosclerosis	40-150	Concentric hyaline thickening of vessel wall	59
Lipohyalinosis	40-300	Noninflammatory lipid and protein aggregation	60

Abbreviations: TOAST, Trial of Org 10172 in Acute Stroke Treatment; CCS, Causative Classification of Stroke; ASCOD, Atherosclerosis, Small vessel disease, Cardiac pathology, Other Causes, Dissection; STRIVE, STAndards for ReportIng Vascular changes on nEuroimaging; LS, lacunar stroke; WMH, white matter hyperintensity; CMB, cerebral microbleed; PVS, perivascular space.

O: other causes) criteria, and the subsequent ASCOD³⁷ (including dissection) also incorporated cSVD as a stroke subtype. This classification schema incorporated disease causality into the stroke classification using 5 categories: (1) potentially causal, (2) causal link uncertain, (3) causal link likely but the disease is not present, (4) small-vessel disease is not detected, and (5) incomplete workup. Small-vessel strokes are deemed potentially causal if the patient has a small, deep infarct less than 15 mm in diameter on neuroimaging in the territory corresponding to symptoms. Additional criteria that need to be met include the presence of one of the following: (1) several old or silent lacunes or small subcortical strokes in other territories; (2) leukoaraiosis (white matter hyperintensities [WMH]), cerebral microbleeds (CMB), or dilated perivascular spaces; or (3) recurrent transient ischemic attacks that are attributable to the same territory as the LS. The causality is classified as uncertain in cases without accompanying neuroimaging evidence of stroke but with a clinical LS syndrome.

Because of recent advances in the understanding of cSVD, the STAndards for ReportIng Vascular changes on nEuroimaging (STRIVE) consortium developed additional criteria that more accurately reflects neuroimaging findings of cSVD (Fig 2).³⁸ These criteria separate the imaging findings of LS into recent small subcortical infarcts and lacunes of presumed vascular origin. The term recent small subcortical infarcts (which can be up to

20 mm in maximum diameter) describes infarcts that are seen on diffusion-weighted imaging (DWI) sequences in the acute setting. The chronic products of such infarctions include lacunes of presumed vascular origin and white matter hyperintensities.^{29,39} Chronic lacunes of presumed vascular origin are typically subcortical fluid-filled spaces (approximately 3-15 mm in diameter) and are best seen on T2-weighted MRI sequences. Furthermore, WMH have a diverse and heterogenous etiology of their own, in addition to resulting from cSVD.^{40,41} Although not formally adopted in STRIVE definitions, smaller lesions have been termed cerebral microinfarcts (CMI), which are .2-3 mm in size.⁴² These lesions are also likely related to cSVD (discussed below).

For this review, we focus on LS either determined in the acute setting (recent small subcortical infarct) or the chronic cavitated end-product, acknowledging that recent subcortical infarcts can also lead to WMH. For the vessel pathology, we focus on sporadic cSVD (sometimes referred to as hypertension [HTN]-related) as opposed to cerebral amyloid angiopathy (CAA)-cSVD.⁴³ We also acknowledge that while LS may appear "lacunar" clinically and radiographically, some result from etiologies other than diseased small vessels.⁴⁴⁻⁴⁶ These non-cSVD etiologies may account for around 10% of imaging and clinically defined LS.⁴⁷ These include branch atherosclerotic disease involving large vessel disease that obstructs a perforator,⁴⁵ artery-to-artery embolism,⁴⁸ which can arise

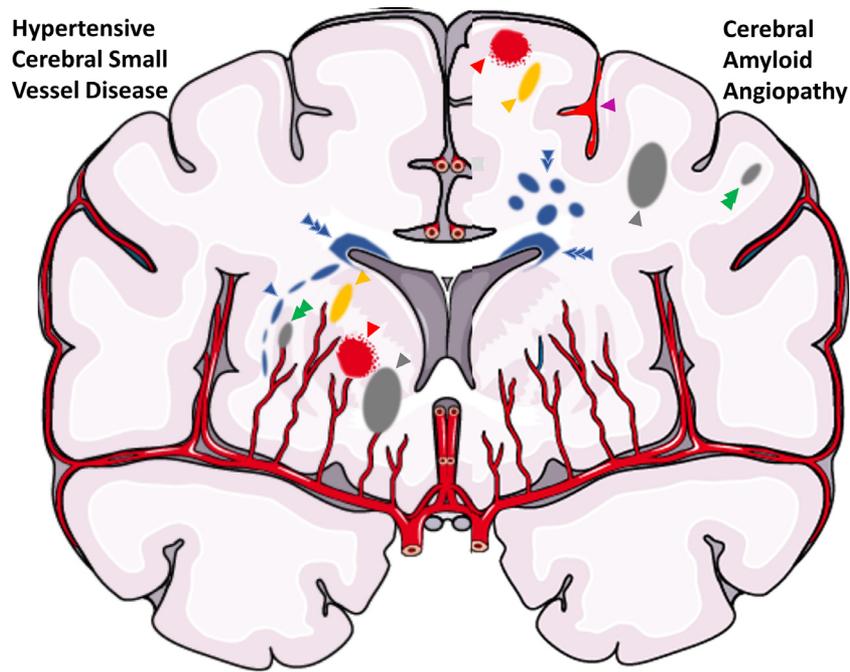


Figure 2. Illustration of a coronal brain section with parenchymal findings of cerebral small vessel disease. The right hemisphere includes findings suggestive of hypertensive cerebral small vessel disease, including a peribasal ganglia white matter hyperintensity (blue arrow), dilated perivascular space (yellow arrow), deep lacune (gray arrow), and microbleed (red arrow). The left hemisphere includes findings suggestive of cerebral amyloid angiopathy, including a microbleed (red arrow), sulcal siderosis (purple arrow), dilated perivascular space (yellow arrow), subcortical white matter hyperintensity spots (double blue arrow), and lobar lacune (gray arrow). There are also perivascular white matter hyperintensities (triple blue arrow) and microinfarcts (double green arrow), which are associated with both small vessel diseases.

from the carotid⁴⁹ or aortic arch,⁵⁰ and cardioembolism.⁴⁷ While the aforementioned studies show LS may be associated with non-cSVD etiologies, it is important to recognize that cSVD shares risk factors with these other etiologies, and it can be difficult to rule out concomitant processes.⁵¹ Indeed, another study of a cohort of ICH patients showed that there was no association between intracranial atherosclerosis (vessel calcification and stenoses) and markers of cSVD (WMH and CMB), supporting atherosclerosis and cSVD are distinct unrelated processes.⁵² Most LS result from intrinsic disease of the small perforating arteries. Furthermore, to conceptualize LS and devise novel therapeutic targets, it is important to distinguish the vessel pathology from the parenchymal pathology, although they are intimately related (Fig 3).

Vessel Histopathology

In Fisher’s early studies of serial sections through over 68 lacunes in 18 human cadaveric brains, he noted that most were found distal to occlusive lesions of small

perforating arteries.^{9,10,53-57} These occlusive lesions are created by distinct vessel pathologies that are based on the size of the vessel and include arteriosclerosis/atherosclerosis, arteriolosclerosis, and lipohyalinosis.^{58,59} Arteriosclerosis/atherosclerosis affects vessels of 200-800 μm and involves similar mechanisms as atherosclerosis of larger vessels (although calcifications are less prominent). In Fisher’s descriptions, atherosclerotic plaques with foam cells were described as microatheromas in the proximal perforating arteries, junctional atheromas in the origin of a perforator, or mural atheromas.¹¹ Arteriolosclerosis affects vessels of 40-150 μm and causes concentric hyaline thickening of the vessel wall, as seen in other organs such as the kidney.⁵⁹ The third mechanism of LS is a process that Fisher termed lipohyalinosis. This process affects vessels with a size ranging from 40 to 300 μm and is characterized by the aggregation of lipid and proteins.⁶⁰ This histopathological term is thought to be the major driver behind cSVD, and underlies most LS.

Because of his pathological observations, Fisher defined lipohyalinosis as a “hypertensive cerebral vasculopathy

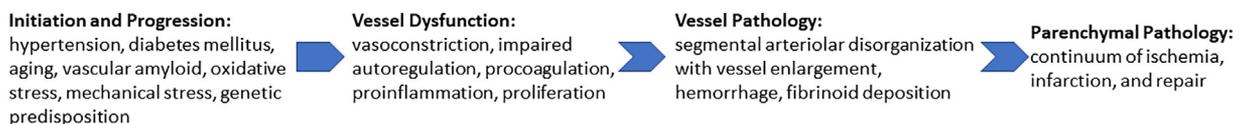


Figure 3. Lacunar stroke timeline, portraying both the vessel and parenchymal disease processes. These processes can be thought of separate therapeutic targets for intervention, but they are intimately related.

in which the lumen of an artery, usually less than 200 μm in diameter, is occluded, the wall of the artery is . . . reduced to connective tissue shreds, hemosiderin-filled macrophages lie scattered in the vicinity, and the wall stains bright red with oil-red-O."¹¹ The central feature of this pathology is noninflammatory fibrinoid vessel wall necrosis. Connective tissue largely replaces the affected vessel walls to variable degrees, extending to the central region of most vessels and occluding the lumen. Muscular elements and elastic elements were often unrecognizable. Few fatty macrophages were observed between lamellae of the disintegrating vessel wall. The vessels did not contain polymorphonuclear leukocytes or pleomorphic histiocytes. Fibroblast proliferation was noted in the adventitia. Interestingly, this disorganization was a focal process, usually extending 2-3 times the vessel diameter. This "segmental arterial disorganization" was often found at sites of bifurcation and branching.¹⁰

Segmental arteriolar disorganization is associated with 3 prominent, but often overlapping findings: vessel enlargement, hemorrhage, and fibrinoid deposition.¹⁰ In most of the lesions, enlargement was thought to be primarily from connective tissue external to the vessel lumen, increasing the diameter 2- to 3-fold. Fisher argued that there was no clear evidence of lumen dilation; rather the fibrosis and disorganization obscured the boundary between wall and lumen, increasing the diameter due to wall thickening. The forms of the enlargements varied, but most were asymmetric. In some cases, single arteries were identified with multiple focal areas of disorganization/enlargement of different ages. Hemorrhage was also present in some lesions via a destruction of vessel walls.¹⁰ Some acute lesions had red blood cells within the damaged wall itself, suggesting a possible local microdissection. Chronic lesions were associated with hemosiderin laden macrophages, suggesting prior hemorrhage. Astroglial proliferation was observed around the hemorrhage. It was also noted that hemorrhage could be found in lesions that did not appear to have enlargement, arguing against a microaneurysm rupture hypothesis. Many lesions also demonstrated evidence of fibrinoid deposition.¹⁰ This deposit could be found in acute or chronic lesions. Furthermore, the fibrinoid deposition within the vessel appeared to cause both lumen reduction and outward wall bulging.

In similar fashion to larger vessel occlusions, connective tissue was often found proximal and distal the lesion, indicating thrombosis and subsequent organization. Retrograde occlusion was noted within pontine penetrators in particular, where normal restoration of flow occurred at the junction of the parent artery. The organized connective tissue varied from fine threads to dense collagenous strands. There were infrequently noted small "channels of recanalization." In areas of arteries affected by secondary thrombosis, walls were thinned, poorly stained, and atrophic, resembling veins.¹⁰ The progression

of this pathology has been described to affect the brain in 3 stages. In the first stage, lipohyalinosis preferentially affects the basal ganglia. In the second stage, lipohyalinosis involves the deep white matter, and also the cerebellum, thalamus, and leptomeningeal arteries. In the last stage, lipohyalinosis affects the brainstem.¹¹

Vessel Pathophysiology—Initiation and Progression

Much of Fisher's work aimed to elucidate the pathophysiology of lipohyalinosis and segmental arterial disorganization, which we refer to as cSVD. At the time of his detailed description, the process of small-vessel destruction had been given several other names including fibrinoid necrosis,⁶¹ plasmatic destruction,⁶² hyalinosis,⁶³ angionecrosis,⁶⁴ and fibrinoid arteritis.⁶⁵ Regardless of name, the mechanism of cSVD is incompletely understood mostly because of the inherent limitations of human autopsy studies. Most autopsy studies of diseased vessels are of old, healed lesions given the low mortality rate of acute LS.⁶⁶ With only histologic descriptions, one must be cautious when making inferences about mechanism. Novel imaging techniques are enhancing our understanding of the disease, but do not allow for direct visualization of the human small vessels *in vivo*. It is now believed that the vessel pathophysiology underlying cSVD likely involves 2 main pathogenic mechanisms: endothelial dysfunction and blood-brain-barrier (BBB) disruption.⁴⁵ In cSVD, there is a shift in the balance of basic functions of both the endothelium and the BBB.

The endothelium is important for the regulation of vessel tone, fibrinolysis/coagulation, inflammation, and angiogenesis. Endothelial dysfunction can be best understood in terms of a shift in each of these functions toward vasoconstriction with impaired autoregulation, procoagulation, proinflammation, and proliferation.⁶⁷ There are different theories regarding how this shift occurs. It was initially hypothesized that HTN was the most important factor, where abnormal mechanical forces on the vessel were secondary to elevated pressure. However, the prevalence of cSVD remains high despite today's effective and widely used antihypertensive agents. As the disease progresses, there is a failure of autoregulation, and the vessel is unable to dilate to maintain perfusion distally.⁶⁸ "Forced dilatation" occurs, as connective tissue accumulates in the vessel wall that leaves downstream vessels subject to even higher pressures and mechanical damage.⁶⁹⁻⁷¹ The thickening worsens and eventual narrowing of the lumen occurs with thrombosis and occlusion.⁴⁵ Indeed, experimental animal models of HTN can induce fibrinoid necrosis.⁷² As the necrosis matures, there is subsequent worsening disorganization and occlusion of the arterial lumen.

Other theories of endothelial dysfunction are proposed, some less widely accepted.⁶⁰ Rather than direct effects of

HTN on mechanical vessel wall destruction, there may be excessive early autoregulatory vasoconstriction that causes focal ischemia. Interestingly, 1 model of experimental HTN showed that fibrinoid necrosis occurred in only localized segments that demonstrated areas of alternating vasoconstriction and dilation.^{71,73} Repeated administration of the vasoconstrictor vasopressin in a rat model demonstrated that it is possible to induce fibrinoid necrosis.⁷⁴ More recent studies have focused on other targets. Thrombomodulin, a marker of endothelial dysfunction, may also be involved. A study of cSVD brains compared to aged controls showed that it was elevated in cSVD brains.⁷⁵ Vascular endothelial growth factor has also been considered for a potential role, though it failed to correlate with disease severity in 1 study.⁷⁶ Yet another autopsy study of cSVD brains showed that white matter penetrating arterioles had impaired vasodilation compared to those in the pia.⁷⁷

The BBB is also important in the vessel pathology of LS. The BBB is comprised of several inter-related structures: tight junctions joining endothelial cells, basement membranes, associated perivascular spaces, pericytes, glia limitans, and astrocyte end feet.⁶⁸ High arterial pressure has been proposed to degrade the BBB. This results in local edema and plasma protein deposition in the vessel wall, which deteriorates the wall structure and damages smooth muscle cells.⁷⁸ However, as with endothelial dysfunction, it is becoming clear that other factors besides HTN are important. BBB degradation likely also underlies the development of more slowly progressing WMH, which are intimately related to LS. Edema develops surrounding leaky vessels, which results in gliosis and appears as WMH radiographically.⁴⁵ In patients with LS, poor functional outcome was associated with increased basal ganglia BBB permeability as measured by contrast-enhanced MRI 3 years later.⁷⁹ Additional studies showed that BBB leakage may mediate cSVD, and HTN was associated with the severity of leakage.⁸⁰ In patients with recent LS, global cSVD burden was associated with increased BBB degradation, interestingly at both the acute infarct site and nonischemic areas.⁸¹

To understand what shifts the balance of these basic functions of the endothelium and BBB, leading to endothelial dysfunction and BBB degradation, respectively, it is useful to think in terms of initiation and progression of the disease. What causes the disease process in the first place and why does it get worse over time? A complex interplay of several factors, including HTN, diabetes mellitus (DM), aging, oxidative stress, mechanical stress, and genetic predisposition are likely involved. Epidemiological studies and genetics studies offer insight (discussed below). There have been several recent advancements in our understanding of the risk factors for cSVD and LS.⁴³ HTN has been a long known risk factor⁴⁵ for fibrinoid necrosis in the small vessels of the brain and other organs.⁸² Interestingly, the vessels of the brain appear to be particularly susceptible as they undergo

fibrinoid necrosis secondary to milder elevations in blood pressure.^{65,83,84} Different organs have been shown to have varying degrees of susceptibility to HTN induced damage in an animal model of inducible HTN.⁸⁵ However, it has been proposed that HTN may be an epiphenomenon in the development of cSVD. Using a transgenic rodent model expressing prorenin solely in the liver, it was shown that "hypertensive" end-organ damage, including fibrinoid necrosis, could be seen in normotensive rats.⁸⁶ This finding suggested that HTN may not be required to induce vessel pathology, but it may be secondary to local generation of an excess of vasoconstrictors, such as angiotensin II. This peptide is not only a potent vasoconstrictor but also plays a role in vascular remodeling.⁸⁷

DM is another well-known risk factor for LS.⁴⁵ Using data from the Atherosclerosis Risk in Communities study, it was shown that smaller lacunes less than 7 mm were associated with DM and HbA1C levels, while larger ones were associated with low-density lipoprotein.⁸⁸ This supports DM plays more of a role in cSVD compared to atherosclerosis and other diseases of the smaller cerebral vessels. Another study using Framingham Heart data showed that incident LS were associated with HTN and DM, but incident intracerebral hemorrhages were associated only with HTN.⁸⁹ Perhaps DM is more important for the small-vessel pathology that causes LS. A genetic predisposition to longevity may also be protective for cSVD and LS. Patients from the Leiden Longevity Study (offspring of nonagenarians) had lower susceptibility to WMH and lacunes, but not CMB.⁹⁰ Similarly, stroke in the young appears to be associated with the development of a higher burden of cSVD in mid-life, where these patients had accelerated cerebral aging by 10-20 years.⁹¹

Parenchymal Histopathology

In addition to the vessel pathology, there is also a unique parenchymal pathology. It is important to make this distinction to understand the pathophysiology of LS and identify novel therapeutic targets, but the vessel pathology and parenchymal pathology are intimately related. In regard to the brain parenchyma, Fisher examined lacunes of varying ages and observed that more chronic lacunes had trabeculated cavities that were surrounded by gliosis and few macrophages, while recent lacunes contained numerous macrophages and had minimal gliosis. Due to the wide-ranging appearance of these lesions, Poirier and Derouesné proposed a classification schema for these lesions in 1984.⁹² This classification system included 3 types of lacunes: Type I are ischemic lesions, Type II are microbleeds where the lacunar cavity is filled with hemosiderin-laden macrophages, and Type III are dilated perivascular spaces. Recent advancements in neuroimaging (described above) have since allowed the radiographic correlates to be easily distinguished. On T2-weighted MRI, lacunes have a surrounding

hyperintense rim compared to dilated perivascular spaces, which lack this finding and are typically smaller than lacunes (<3 mm).^{38,93,94}

The pathologic description of the subtypes of Type I ischemic lesions, Ia and Ib, provide additional insight into cSVD pathogenesis. Type I lesions are characterized by an irregular cavity ranging from 1 to 20 mm in length, and are most commonly found in the putamen, caudate, thalamus, pons, internal capsule, and hemispheric white matter.^{9,95} Within this group, Type Ia lesions are old infarcts with pan-necrotic cavitation and scattered macrophages. Type Ib lesions represent incomplete infarcts with selective loss of only some vulnerable elements.⁹⁶ Interestingly, in a study by Lammie et al examining 172 autopsy brains, Type Ib incomplete infarcts were seen in the majority of cadaveric brains that contained the classic Type Ia lesions.⁹⁶ These lesions exist on a spectrum, and can be graded from 1 to 3,⁹⁶ based on their size, parenchymal integrity, surviving neurons, surviving glia, presence of reactive glia, and presence of inflammatory cells. Grade 1 lesions are about .5 × .5 mm, have parenchymal rarefaction/pallor, a mild reduction in neuron number with occasional “ghost” neurons, preserved oligodendroglia, absent reactive astrocytes, and absent inflammatory cells (lymphocytes, macrophages, siderophages). Grade 3 lesions, on the other hand, are about 10 × 1 mm, and exhibit spongiosis, rarefaction with small central cavitation, no surviving neurons with only occasional surviving oligodendroglia, gliovascular septa in cavities with surrounding gliosis, and inflammatory cells scattered in and around the cavity. Grade 2 lesions are described as an intermediary of the 2.

The etiology of Type Ib parenchymal lesions has been debated; understanding the cause of these lesions has implications for therapeutic targets. It has been contended by some that Type Ib lesions are secondary to local edema instead of ischemia.⁹⁷ Ma and Olsson first described “edema-associated gliosis”⁹⁷ that appears similar in morphology to these lesions.⁹⁸ Furthermore, similar findings have been recapitulated in animal models.⁹⁹⁻¹⁰¹ As mentioned above, this pathology was originally described as resulting from destructive vaginitis in which the blood vessel remained patent, but there was perivascular space dilatation, which destroyed surrounding parenchyma (similar to Type III lesions). Still, most speculate that Type Ib lesions result from ischemia, similar to Type Ia lesions.¹⁰² Most likely, these incomplete Type Ib lesions differ from Type Ia lesions secondary to a less-severe degree of ischemia, a shorter duration of ischemia, and less likely a different etiology of ischemia.

Parenchymal Pathophysiology—Ischemia, Infarction, and Repair

Three principles underlie the dynamic processes of injury and recovery in LS: ischemia, infarction, and repair.

Ischemia occurs when parenchyma is deprived of blood affecting its function but not causing apoptosis and necrosis. Infarct refers to parenchyma that has undergone apoptosis and necrosis. These likely exist on a continuum where blood flow changes can tip the balance. While different from large-vessel occlusions,¹⁰³ there is also mounting evidence supporting a role of decreased blood flow in lacunes, small subcortical infarcts, and WMH. A recent study showed WMH had decreased cerebral blood flow (CBF) and decreased vascular reactivity compared to contralateral normal white matter.¹⁰⁴ Another study demonstrated that at 4-year follow-up, baseline WMH volume was associated with decreased CBF over time, but baseline CBF was not associated with progression of lacunes or WMH.¹⁰⁵ In contrast, another study of minor strokes showed that at 18-month follow-up, regions of low baseline CBF were associated with the development of new WMH.¹⁰⁶ Many hypothesize that lower CBF precedes the development of LS and WMH, but the data are conflicting.¹⁰⁷ Interestingly, the peri-infarct region was recently pathologically examined in lacunes,¹⁰⁸ and axons up to 150% of the infarct diameter away showed changes reflecting a loss of normal cell-cell adhesion and signaling between axons and oligodendrocytes. This may reflect injury to myelin out of proportion to axons. These findings have been replicated in another study demonstrating enlargement/progression of lacunes by serial imaging.¹⁰⁹ This slow stroke progression may prove a viable target for therapeutic intervention in human patients.

Repair refers to processes involving proliferating surviving elements and neuroplasticity, including neurogenesis, gliogenesis, axonal sprouting, remyelination, and altered neuronal excitability.¹¹⁰ Importantly, the repair phase occurs after the initial ischemia and infarct phases. There is a biphasic nature to this progression, emphasizing the importance of when, where, and how damaged brain makes the transition from injury to repair.¹¹¹ This is exemplified at the structural level by matrix metalloproteinases, which have a deleterious role acutely but are essential for neurovascular remodeling during subsequent repair,¹¹² and at the functional level by changes in excitability, in which excitotoxicity leads to further injury acutely but increasing excitability promotes recovery in the repair phase.^{113,114} This concept may also be pertinent for LS and injury isolated to white matter, although the structural and functional elements are different and only beginning to be understood. There are likely spontaneous processes that stabilize the BBB, modulate glial scarring, regulate remyelination, slow axonal degeneration, and promote the health of the oligovascular unit.^{108,115,116} These endogenous mechanisms involve oligodendrocyte precursor cells,¹¹⁷ which proliferate, migrate, and differentiate into myelinating oligodendrocytes.^{118,119} There may also be endogenous processes that can hinder repair. Glial scar formation involves several cell surface and extracellular matrix proteins that can inhibit axonal growth and impair the ability of oligodendrocytes to

myelinate new axons. These detrimental proteins include neurite outgrowth inhibitor (Nogo), ephrin ligands, and chondroitin proteoglycans.¹²⁰ A recent mouse study showed that Nogo receptor 1 blockade could overcome remyelination failure and stimulate functional recovery after LS.¹²¹ Further demonstrating the complexity and biphasic nature of repair, reactive astrogliosis can be beneficial both acutely and under certain circumstances during repair.^{122,123}

As expected, these parenchymal processes are difficult to study *in vivo*, especially in human patients. However, serial imaging studies of small subcortical strokes, lacunes, WMH, and other imaging biomarkers of cSVD may help us understand the progression of disease. These imaging sequelae are inter-related and show changes consistent with both progression and regression of severity.¹²⁴ Indeed, some small subcortical infarcts can become undetectable⁴¹ and some WMH can regress on repeat imaging.¹²⁵ This may occur through alteration in blood flow, focal small-vessel changes, and parenchymal repair processes. Interactions between small subcortical infarcts, lacunes, and WMH have been studied, and a WMH “penumbra” hypothesis has been proposed as incident lacunes have a predilection for the edge of WMH.¹²⁶ However, another study confirmed their main axes aligned with the orientation of perforating arteries and not white fiber tracts,¹²⁷ supporting the widely accepted vascular etiology. Perhaps flow-limiting areas of vessel disease affecting lower-order branched vessels contribute to downstream hypoperfusion and WMH. When subsequent branches develop worsening pathology and/or when perfusion decreases further in these already vulnerable areas, small subcortical infarcts occur and WMHs progress.²⁹

In addition to relationships to each other, lacunes, small subcortical infarcts, and WMHs are also related to CMI. While smaller than small subcortical infarcts and lacunes, CMI may be linked to LS in their underlying etiology. CMI have been classically observed in neuropathological studies¹²⁸ and have been more recently observed by 3T and 7T MRI scanners.^{42,129-132} In a cadaveric study, *in vivo* 1.5T MRI was compared to post mortem *ex vivo* 7T MRI and histology. The authors noted that the vast majority of CMI are under the detection limits of clinical *in vivo* MRI.¹³³ In another study, silent DWI positive CMI were detected in 3.8% of cognitively impaired, 5% of normal, and 12.9% of past ICH patients.¹³⁴ Histologic sampling showed that even a single DWI lesion suggested an annual incidence of hundreds of new CMI.¹³⁴ CMI are becoming more clinically relevant as they become better understood. Asymptomatic DWI positive CMI have been shown to produce chronic local microstructural injury¹³⁵; they are a common finding in memory disorder populations and are associated with vascular dementia.^{130,131,136}

CMI likely have a heterogeneous etiology but many result from cSVD of high order branched vessels. Indeed, the location of CMI has shown some predictive value. While

deep CMI are likely secondary to cSVD,^{42,137} cortical CMI are more heterogeneous in etiology as they are related to CAA,^{42,137} cSVD and LS,¹³² and intracranial stenosis and large cortical infarcts.¹³² A study of patients with recent ICH showed that the lobar location was associated with CMI in the frontal lobe, while the deep location was associated with CMI in the parietal lobe.¹³⁸ In another study of patients with ischemic stroke, CMI were found in 28%, 71% of which were in the vascular territory of the primary infarct. WMH was associated with CMI outside the territory and large-vessel atherosclerosis was associated with CMI within it.¹³⁹ Understanding the role of CMI may provide further insight into LS pathogenesis.

These imaging findings in cSVD have also been found to be associated with other imaging biomarkers, including ICH. One study showed that lobar lacunes, defined as lacunes by the STRIVE definition that are located in subcortical lobar white matter, were associated with CAA-related ICH, while deep lacunes were associated with HTN-related ICH. Furthermore, lobar lacunes had an association with WMH.⁴⁶ This finding was confirmed in another study, which also demonstrated an association of lobar lacunes with brain amyloid deposition on positron emission tomography imaging.¹⁴⁰ These associations prompt investigation into the role of CAA in lacune formation. While lacunes are more typically found in deep brain structures, they have been reported in the subcortical white matter such as the centrum semiovale, even by Fisher.^{9,10} However, we acknowledge that lacunes from infarction are sometimes difficult to distinguish from dilated perivascular spaces, which are also associated with CAA.¹⁴¹ Another study demonstrated that subcortical WMH were more common in CAA, while peribasal ganglia WMH were more common in cSVD. Predictors of peribasal ganglia WMH included deep CMB, total WMH volume, high BG perivascular spaces.¹⁴² Indeed, WMH are related to both cSVD and CAA. In patients with CAA, WMH increase over time, and are associated with incident lobar ICH. This supports that WMH in CAA is related to progressive vessel disease.¹⁴³ Yet another study showed that WMH have a predilection for areas of lower perfusion, regardless of underlying diagnosis.¹⁴⁴ Perivascular spaces may also provide insight into the underlying microangiopathy. In a study of patients with ICH, centrum semiovale perivascular spaces were associated with lobar CMB and cortical superficial siderosis, while basal ganglia perivascular spaces were associated with deep ICH and WMH volumes.¹⁴⁵ A similar finding was shown in patients with cognitive impairment, where white matter perivascular spaces were associated with lobar CMB, and basal ganglia perivascular spaces were associated with HTN.¹⁴⁶

Biomarkers in LS and Systemic Disease

The development of biomarkers for LS and cSVD is important for predicting disease severity and prognosis and for preventing disease progression (Table 2).

Table 2. Biomarkers in lacunar stroke and cerebral small-vessel disease

Biomarker	Association	Comparison	Reference
<i>Coagulation/fibrinolysis</i>			
Tissue plasminogen activator	↑	LS versus patients without stroke	147
Tissue plasminogen activator	↓	LS versus cortical stroke	148
Fibrinogen alpha chain	↑	LS versus patients without stroke	158
Fibrinogen beta chain	↑	LS versus patients without stroke	158
Fibrinogen	↓	LS versus cortical stroke	147
Fibrinogen	↑	LS with adverse outcome versus LS without	158
D-dimer	↑	LS versus patients without stroke	147
D-dimer	↓	LS versus cortical stroke	147
Thrombin-antithrombin complex	↑	Blacks with LS versus those without	153
<i>Endothelial dysfunction</i>			
von Willebrand factor	↑	LS versus patients without stroke	149
von Willebrand factor	↓	LS versus cortical stroke	149
Homocysteine	↑	LS versus patients without stroke	149
Homocysteine	None	LS versus cortical stroke	149
Endothelial progenitor cells	↓	LS and WMH versus hypertensive controls	150
Angiogenic T-cells	↓	LS and WMH versus hypertensive controls	150
<i>Platelet activation</i>			
Activated glycoprotein IIb/IIIa	None	LS versus patients without stroke	149
P-selectin	None	LS versus patients without stroke	149
Platelet microparticles	None	LS versus patients without stroke	149
<i>Inflammation</i>			
Alkaline phosphatase	↑	Silent LS versus large-vessel atherosclerosis	151
C-reactive protein	↑	LS versus patients without stroke	152
C-reactive protein	None	LS versus cortical stroke	152
Interleukin-6	↑	LS versus patients without stroke	147
Interleukin-6	↓	LS versus cortical stroke	147
Interleukin-6	↑	LS with major vascular events versus without	155
Tumor necrosis factor	↑	LS versus patients without stroke	147
Tumor necrosis factor	↑	LS with major vascular events versus without	155
Neopterin	↑	LS versus hypertensive controls	154
<i>Adhesion</i>			
Intercellular adhesion molecule-1	↑	LS versus hypertensive controls	154
Vascular cadherin adhesion molecule-1	↑	LS versus hypertensive controls	154
Vascular cadherin adhesion molecule-1	↑	Blacks with LS versus those without	153
Integrin alpha-IIb	↑	LS with adverse outcome versus LS without	158
Talin-1	↑	LS with adverse outcome versus LS without	158
Filamin-A	↑	LS with adverse outcome versus LS without	158
<i>Lipids</i>			
Glucosylceramide	↑	LS versus patients without stroke	156
Phosphatidylethanolamine	↑	LS versus patients without stroke	156
Free fatty acids	↑	LS versus patients without stroke	156
Triacylglycerol	↑	LS versus patients without stroke	156
Omega 3-polyunsaturated fatty acids	↓	LS with cSVD versus without	157
<i>Other</i>			
Myelin basic protein	↑	LS with adverse outcome versus LS without	158
Albumin	↓	LS with adverse outcome versus LS without	158

Abbreviations: cSVD, cerebral small vessel disease; LS, lacunar stroke; WMH, white matter hyperintensity.

Biomarkers can also shed light on mechanisms and serve as outcomes for future studies. A meta-analysis showed that markers of coagulation/fibrinolysis, endothelial dysfunction, and inflammation were altered in LS compared to other stroke subtypes and/or healthy controls. This suggests that these 3 pathways are clinically relevant mechanisms to LS pathogenesis.¹⁴⁷

One comprehensive study compared several biomarkers in nondisabling LS versus cortical stroke after 1 month; the LS group had decreased tissue plasminogen activator, which was the only biomarker found to be significantly different.¹⁴⁸ Another study enrolled recent LS patients and showed that markers of platelet activation (activated glycoprotein IIb/IIIa, P-selectin, and platelet microparticles)

were not different from healthy controls; however, markers of endothelial dysfunction (von Willebrand factor and homocysteine) were increased.¹⁴⁹ Patients with essential HTN and WMH or lacunes, versus HTN alone, were enrolled in another study that showed that endothelial progenitor cells (which may ameliorate endothelial dysfunction) and angiogenic T-cells (which stimulate endothelial progenitor cells) were both decreased in number.¹⁵⁰

Serum alkaline phosphatase, posited to be a marker of vascular calcification, was shown to be associated with silent lacunes but not large-vessel atherosclerosis in neurologically healthy patients.¹⁵¹ Alkaline phosphatase is an acute phase reactant so this association could be due to inflammation, which underlies the pathophysiology of LS, at least in part.⁴⁵ Another study of healthy patients showed a correlation of higher C-reactive protein levels and number of silent lacunes.¹⁵² One small study of African Americans with LS revealed that they had elevated proinflammatory biomarkers, soluble vascular cadherin adhesion molecule-1 and thrombin antithrombin, and an abnormal response to acute immune challenge 6 weeks after stroke.¹⁵³ Vascular inflammation was also assessed in a study of first-time LS and essential HTN controls. Neopterin and circulating adhesion molecules were higher in patients with MRI-confirmed cSVD.¹⁵⁴ The higher Levels of Inflammatory Markers in the Treatment of Stroke study (ancillary biomarker study within SPS3) examined patients with recent LS, and showed interleukin-6 and tumor necrosis factor receptor concentrations predicted risk of recurrent vascular events.¹⁵⁵

In addition to these 3 categories of biomarkers, other novel potential biomarkers are under evaluation. One group developed a plasma panel for lipid species, including glucosylceramide, phosphatidylethanolamine, free fatty acid, and triacylglycerol, that was 93% sensitive and 97% specific for LS compared to healthy controls.¹⁵⁶ Another study found that low levels of plasma omega 3-polyunsaturated fatty acids were associated with markers of cSVD, though perhaps less so in LS, when patients were enrolled 7 days after stroke.¹⁵⁷ Using quantitative proteomics of microvesicle enriched plasma, 1 study showed that the patients with adverse outcomes had decreased albumin and increased brain-specific proteins (myelin basic protein), coagulation cascade proteins (fibrinogen alpha chain, fibrinogen beta chain), and focal adhesion proteins (integrin alpha-IIb, talin-1, and filamin-A).¹⁵⁸

The literature investigating biomarkers can be difficult to interpret given the heterogeneity of stroke and the many variables involved. Future biomarker studies should be hypothesis-driven, specifically evaluating the unique properties of cSVD-induced LS compared to other stroke subtypes. This involves including controls, such as patients with HTN without cSVD and patients with other subtypes of stroke. Duration after stroke onset, size of stroke, and location of stroke are all also important considerations. Lastly, studies should evaluate features that are unique to the cerebral vasculature

that may differentiate cSVD from systemic thrombosis and systemic vessel diseases. Indeed, it has been suggested that cSVD may be a component of a systemic disease, affecting multiple organs by similar pathophysiological mechanisms causing systemic arteriolar dysfunction.¹⁵⁹ An example is the association between WMH, dementia, and retinal vascular abnormalities.¹⁵⁹ Moreover, chronic kidney disease may be linked to WMH, demonstrated in a study that showed patients with kidney disease had increased WMH.¹⁶⁰ Further studies showed that patients with reduced creatinine clearance have increased WMH volume.¹⁶¹

Genetics of LS

Although LS is a multifactorial disease that is associated with numerous risk factors, there are considerable differences in susceptibility to these risk factors when comparing individuals. Moreover, there are examples of patients who develop cSVD in the absence of risk factors, suggesting a role for genetic modulation.¹⁶² Family history of vascular disease is an independent risk factor for LS.¹⁶³ There is also evidence from twin and family studies that WMH, related to LS, have a genetic basis.^{21,164,165} One study examined 74 monozygotic and 71 dizygotic twins, showing a 73% heritability for WMH volume.¹⁶⁶ Moreover, concordance rates for WMH were 61% among monozygotic twins and 38% in dizygotic twins. Another heritability study utilized a family-based study design from the Framingham Heart cohort.¹⁶⁷ WMH volume was assessed in over 2000 patients who were free of stroke. The results of this study demonstrated a WMH volume heritability of 78% among women and 52% among men.

Apart from these heritability studies, there are several single-gene disorders that are linked to mutations that precipitate cSVD. These account for about 1%-5% of all strokes.¹⁶⁸ Among the major diseases are Fabry's disease, CADASIL, CARASIL,¹⁶⁹ COL4A1-related cSVD,¹⁷⁰ and autosomal dominant retinal vasculopathy with cerebral leukodystrophy.¹⁷¹ The contribution of these individual genes in developing spontaneous cSVD is unknown; however, a more complete understanding of these disorders may provide insight into its pathogenesis.¹⁷²

Candidate gene studies have been a popular method to study complex polygenic disorders, and employ the use of single nucleotide polymorphism (SNP) analysis in genes that are suspected to play a role in the disease.¹⁷³ Genes that encode for angiotensin-converting enzyme (ACE) and endothelial nitric oxide synthase (eNOS) are among some candidate genes that have been examined. The deletion (DD) genotype for ACE and the GG genotype of Glu298Asp eNOS polymorphism¹⁷⁴ have been associated with all ischemic stroke¹⁷⁵ and specifically LS.¹⁷⁶ Another candidate gene that has been extensively studied is the *apolipoprotein E (ApoE)* gene. ApoE ϵ 4 allele carriers had a significantly higher subcortical (and not periventricular) WMH burden when controlling for

HTN.¹⁷⁷ Since *ApoE* is involved in lipid metabolism¹⁷⁸ and neuronal repair,¹⁷⁹ it raises the question whether it may be involved in the pathophysiology of subcortical LS. Some speculate that the D allele of *ACE* may share a similar pathway as *ApoE* ϵ 4 and may be implicated in the development of WMH.¹⁸⁰ As with many candidate gene studies, sampling bias and poor phenotyping are limitations.¹⁸¹ Larger studies failed to replicate this finding, and only showed an association between the *ACE* I/D polymorphism and WMH.¹⁸²

One novel method that circumvents the arduous task of examining individual genes has been genome wide association studies (GWAS). Applying this methodology to 2 independent Japanese samples with LS revealed a nonsynonymous SNP in a member of the protein kinase C family, *PRKCH*.¹⁸³ This protein is expressed in endothelial cells, macrophages, and smooth muscle cells of atherosclerotic plaques. Similar findings were recapitulated in subjects with silent lacunes.¹⁸⁴ In GWAS examining WMH, 6 SNPs were associated with larger WMH volumes (although the distribution of these lesions was not examined). These SNPs were located near *WBP2*, *TRIM65*, *TRIM47*, *MRPL38*, *FBF1*, and *ACOX*.¹⁸⁵ In another such GWAS study conducted in the Netherlands, chromosome 5q12 was shown to increase susceptibility to stroke.¹⁸⁶ Further mapping of this locus revealed that SNPs near the *PDE4E* gene are intimately tied to stroke risk.¹⁸⁷ The authors showed that the C allele of SNP45 and the T allele of SNP39 increased the risk of LS by 4.8 and 6.3 times, respectively.¹⁸⁸ A recent GWAS was performed on over 1000 early-onset LS patients.¹⁸⁹ On the basis of histopathological features and radiographic findings, the authors defined 2 subsets of LS: isolated LS and multiple LS with leukoaraiosis. There was a significant heritable component to LS of around 20%-25%, with isolated LS at 15%-18% and multiple LS/leukoaraiosis at 23%-28%. This suggests the presence of 2 LS subtypes.

GWAS have been criticized as they are underpowered to detect rare variants that could contribute higher risk than common variants, which generally confer a small relative risk.¹⁸¹ To assess for this, the use of non-hypothesis-driven methodology using large sample sizes is required. In this regard, whole exome sequencing using next-generation technology allows for the analysis of the entire coding sequences in the human genome. A small study using exome sequencing was performed in the Chinese Han population.¹⁹⁰ A SNP in the *c1orf156* gene located on 1q24 and another in the *XYLB* gene located on 3p21 were identified as being related to ischemic stroke. These genes have been implicated broadly in cardiovascular disease. Another whole exome sequencing study examined 10 patients, 5 of which had LS.¹⁹¹ Two genes, *CSN3* and *HLA-DPB1*, were implicated among the LS group. Interestingly, the *CSN3* gene has been associated with DM and coronary artery atherosclerosis.^{192,193} A study utilizing next generation sequencing on LS specifically is currently underway¹⁹⁴; it may provide further insight into LS pathogenesis. Although the exact

genetic determinants of cSVD remain to be clarified, it is likely that it is a polygenic phenomenon involving an interplay between many genetic variants across the genome and environmental factors.¹⁷²

Modeling Small-Vessel Disease and LS

A major barrier to the study of cSVD and LS is that animal and cellular models that accurately replicate the vessel and parenchymal pathology in humans are only recently being developed. For many of the basic questions regarding mechanism, experimental models may enhance our understanding. Models must be chosen carefully, however, as no single model perfectly replicates the human phenotype. Furthermore, several models may be required in complementary fashion to capture both vessel and parenchymal pathology and replicate findings. There are 3 main categories of animal models: chronically hypertensive animals, chronically hypoperfused animals, and animals that undergo targeted small focal infarcts.¹⁹⁵

The stroke-prone spontaneously hypertensive rat (SHR-SP) is an example of a chronically hypertensive model.¹⁹⁶ Compared to control rats, SHR-SP show reduced endothelial tight junctions, reduced nitric oxide bioavailability, impaired myelination, increased glial and microglial activity, impaired matrix proteins, impaired vascular reactivity, and reduced albumin.¹⁹⁷ Established in 1974,^{198,199} this inbred rat strain spontaneously develops an arteriopathy that may include cerebral edema, ischemia/infarction, and ICH.^{78,196,200,201} The pathology was similar to humans, in that strokes occurred at arteries that branch from larger ones.²⁰¹ A decade later, a more detailed histologic description was made through serial sectioning that showed BBB degradation and vasogenic edema.²⁰⁰ At the leakage sites, they identified fibrinoid degeneration of penetrating arterioles. There was a loss of detectable vessel wall components and a thickening of the vessels with expanded perivascular tissue space. They also described occlusive fibrin thrombi and noted the formation of necrotic cysts, though some parenchymal lesions were a milder variation of edema. Fibrin thrombi were found close to the cysts, but it was unclear if the thrombi caused infarcts or if they formed secondarily to a primary leakage event. The same group later showed that damaged endothelial and smooth muscle cells were replaced by fibrin-like material, basement membrane thickening, and bundles of collagen fibrils.⁷⁸ They found these vascular lesions occurred in short segments similar to the description of human lesions by Fisher. Interestingly, more modern studies of SHR-SP have shown the endothelial structural changes predate HTN. The changes included reduced endothelial tight junctions, increased smooth muscle actin and GFAP, as well as altered matrix metalloproteinase 9 compared to controls.²⁰² A recent detailed systematic review deemed SHRSP the most appropriate

model for human cSVD available,²⁰³ and it lends itself to easy study of neuroprotective agents.¹⁹⁶ However, another recent study suggests it also models posterior reversible encephalopathy syndrome.²⁰⁴

In addition to modeling the vascular pathology, novel models are being developed to study the parenchymal pathology including cell, tissue, and animal-based platforms.²⁰⁵ While the traditional middle cerebral artery occlusion model does damage oligodendrocytes and white matter in response to ischemia,²⁰⁶ it does not target white matter specifically.^{207,208} The bilateral carotid occlusion model of chronic hypoperfusion has been shown to induce white matter ischemia.^{209,210} This model causes changes in white matter that resemble human WMH²¹¹⁻²¹³ and cognitive impairment.²¹⁴

Key Points

- Since the 1800s, lacunes were recognized as infarcts involving the distal small penetrating branches of larger cerebral arteries.
- C. Miller Fisher performed numerous pathological studies of lacunes and described lipohyalinosis, the unique underlying mechanism of cerebral small vessel disease.
- Lacunar strokes comprise up to 20-30% of all acute ischemic strokes.
- To conceptualize lacunar stroke and devise novel therapeutic targets, it is important to distinguish the vessel pathology from the parenchymal pathology.
- Vessel histopathology is characterized by segmental arteriolar disorganization, which is associated with three prominent, often overlapping findings: vessel enlargement, hemorrhage, and fibrinoid deposition.
- The initiation and progression of cerebral small vessel disease is related to hypertension, diabetes mellitus, aging, oxidative stress, mechanical stress, and genetic predisposition.
- A physiological balance is tipped toward vasoconstriction with impaired autoregulation, procoagulation, proinflammation, and proliferation.
- The parenchymal pathophysiology involves is a continuum of reduced blood flow, ischemia, infarction, and repair.
- Recent studies suggest that lobar lacunes may be associated with cerebral amyloid angiopathy.
- Other areas of future research include understanding the genetic underpinnings of lacunar stroke, uncovering novel biomarkers, and studying cerebral small vessel disease as part of a systemic process involving other vascular beds.
- Preclinical models of cerebral small vessel disease and lacunar stroke include three types: chronically hypertensive animals, chronically hypoperfused animals, and animals that undergo targeted small focal infarcts.
- Further research using a variety of strategies including preclinical models and advanced neuroimaging is needed to further unravel the processes that underlie this common disease of older adults.

For focal infarcts targeted to any desired brain region, laser coagulation models²¹⁵ and stereotactic parenchymal injection of vasoconstrictors can be utilized. Common vasoconstrictors include endothelin-1²⁰⁸ and N5-(1-iminoethyl)-L-ornithine,²¹⁶ the latter being preferred as it avoids receptor mediated effects. If injected into the subcortical white matter, N5-(1-iminoethyl)-L-ornithine creates a lesion presumably from small-vessel occlusion and induces a phenotype similar to human LS.²¹⁶ Cognitive impairment can also be assessed in these models, further demonstrating translatability.²¹⁷ In this model, several histologic changes have been reported after white matter stroke, including focal edema, demyelination, axonal damage, loss of oligodendrocytes, and local activation of astrocytes and microglia.¹¹⁷ One mouse model showed that the size of these strokes grew over the first month,¹¹⁷ possibly representing vulnerable hypomyelinated peri-infarct axons. In this peri-infarct region, myelin loss was found to exceed axon damage. Furthermore, microglia/macrophages were found near injured axons at day 1, and their presence continued to increase until day 7.

In addition, transgenic mouse models can be utilized. *COL4A1/2* models replicate cSVD with ICH,²¹⁸ while *Notch3* models exhibit WMH largely without vessel pathology.²¹⁹ Lastly, there are in vitro models of the BBB and also of neurons, oligodendrocytes, and other glia culture systems that can undergo a multitude of experiments, including oxygen/glucose deprivation.²²⁰

Conclusion

LS pose a major burden to society in terms of morbidity and mortality. They comprise a large portion of strokes and can be particularly devastating given their stuttering progressive nature, their risk of recurrence, and their cumulative effects on cognition. Since their early discovery in the late 1800s and their detailed description by Fisher in the 1960s, there has been steady progress in the understanding of their pathophysiology. Although LS is multifactorial, cSVD is the major mechanism that contributes to LS. CAA is another common cerebrovascular pathology emerging as a cause of both hemorrhagic and ischemic brain lesions, including LS. It is important to conceptualize LS as a process involving both small vessel and parenchyma pathologies, though they are intimately related. With developments in imaging, CMI may provide additional clues into LS pathogenesis and the relationship of LS to other imaging markers of cSVD can be further examined. Until recently, cSVD has been poorly understood as it is a complex phenomenon that is influenced by genetics and environmental risk factors. Although there has been improvement in understanding this disease, it has not translated into effective targeted therapies for LS. Further research using a variety of strategies including preclinical models and advanced neuroimaging is needed

to further unravel the processes that underlie this common disease of older adults.

Conflicts of Interest

The authors report no conflicts of interest or disclosures.

References

- Pearce J. Dechambre's description of lacunes, 1838. *J Neurol Neurosurg Psychiatry* 1990;53:134.
- Dechambre A. Memoire sur la curabilite du ramollissement cerebral. *Gaz Medicale, Paris* 1838;6:305-314.
- Roman GC. The original description of lacunes. *Neurology* 1986;36:85.
- Roman GC. On the history of lacunes, etat criblé, and the white matter lesions of vascular dementia. *Cerebrovasc Dis* 2002;13(Suppl 2):1-6.
- Durand-Fardel M. Traite du ramollissement du cerveau. JB Baillière, Paris.
- Poirier J, Derouesne C. The concept of cerebral lacunae from 1838 to the present. *Rev Neurol (Paris)* 1985;141:3-17.
- Marie P. Des foyers lacunaires de désintégration et des differents autres états cavitaires du cerveau. *Rev Méd* 1901;21:281-298.
- Román GC. On the history of lacunes, etat criblé, and the white matter lesions of vascular dementia. *Cerebrovasc Dis* 2002;13(Suppl 2):1-6.
- Fisher CM. Lacunes: small, deep cerebral infarcts. *Neurology* 1965;15:774-784.
- Fisher CM. The arterial lesions underlying lacunes. *Acta Neuropathol* 1968;12:1-15.
- Fisher CM. Lacunar strokes and infarcts: a review. *Neurology* 1982;32:871-876.
- Moran C, Phan TG, Srikanth VK. Cerebral small vessel disease: a review of clinical, radiological, and histopathological phenotypes. *Int J Stroke* 2012;7:36-46.
- Marti-Vilalta JL, Arboix A. The Barcelona Stroke Registry. *Eur Neurol* 1999;41:135-142.
- Caplan LR. Worsening in ischemic stroke patients: is it time for a new strategy? *Stroke* 2002;33:1443-1445.
- Mohr JP, Caplan LR, Melski JW, et al. The Harvard Cooperative Stroke Registry: a prospective registry. *Neurology* 1978;28:754-762.
- Steinke W, Ley SC. Lacunar stroke is the major cause of progressive motor deficits. *Stroke* 2002;33:1510-1516.
- Samuelsson M, Soderfeldt B, Olsson GB. Functional outcome in patients with lacunar infarction. *Stroke* 1996;27:842-846.
- Norrving B. Long-term prognosis after lacunar infarction. *Lancet Neurol* 2003;2:238-245.
- Bejot Y, Catteau A, Caillier M, et al. Trends in incidence, risk factors, and survival in symptomatic lacunar stroke in Dijon, France, from 1989 to 2006: a population-based study. *Stroke* 2008;39:1945-1951.
- Leary MC, Saver JL. Annual incidence of first silent stroke in the United States: a preliminary estimate. *Cerebrovasc Dis* 2003;16:280-285.
- Vermeer SE, Longstreth WT, Koudstaal PJ. Silent brain infarcts: a systematic review. *Lancet Neurol* 2007;6:611-619.
- Regenhardt RW, Biseko MR, Shayo AF, et al. Opportunities for intervention: stroke treatments, disability and mortality in urban Tanzania. *Int J Qual Heal Care* 2019;31:385-392.
- Berkhemer OA, Fransen PSS, Beumer D, et al. A randomized trial of intraarterial treatment for acute ischemic stroke. *New Engl J Med* 2014;372:11-20.
- Jovin TG, Chamorro A, Cobo E, et al. Thrombectomy within 8 hours after symptom onset in ischemic stroke. *N Engl J Med* 2015;372:1-11.
- Goyal M, Menon BK, Zwam WH, et al. Endovascular thrombectomy after large-vessel ischaemic stroke: a meta-analysis of individual patient data from five randomised trials. *Lancet* 2016;387:1723-1731.
- Saver JL, Goyal M, Bonafe A, et al. Stent-retriever thrombectomy after intravenous t-PA vs. t-PA alone in stroke. *N Engl J Med* 2015;372:2285-2295.
- Campbell BCV, Mitchell PJ, Kleinig TJ, et al. Endovascular therapy for ischemic stroke with perfusion-imaging selection. *N Engl J Med* 2015;372:1009-1018.
- Regenhardt RW, Mecca AP, Flavin SA, et al. Delays in the air or ground transfer of patients for endovascular thrombectomy. *Stroke* 2018;49:1419-1425.
- Regenhardt RW, Das AS, Lo EH, et al. Advances in understanding the pathophysiology of lacunar stroke. *JAMA Neurol* 2018;75:1273-1281.
- Fisher CM, Curry HB. Pure motor hemiplegia of vascular origin. *Arch Neurol* 1965;13:30-44.
- Silverman I, Rymer M. Ischemic stroke: an atlas of investigation and treatment. Oxford, UK: Clinical Publishing; 2009.
- Alexandrov A. Neurovascular examination: the rapid evaluation of stroke patients using ultrasound waveform interpretation. Oxford, UK: Wiley-Blackwell; 2013.
- Mohr JP. Lacunes. *Neurol Clin* 1983;1:201-221.
- Adams Jr HP, Bendixen BH, Kappelle LJ, et al. Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. TOAST. Trial of Org 10172 in Acute Stroke Treatment. *Stroke* 1993;24:35-41.
- Kang D-W, Chalela JA, Ezzeddine MA, et al. Association of ischemic lesion patterns on early diffusion-weighted imaging with TOAST stroke subtypes. *Arch Neurol* 2003;60:1730-1734.
- Arsava EM, Ballabio E, Benner T, et al. The Causative Classification of Stroke system: an international reliability and optimization study. *Neurology* 2010;75:1277-1284.
- Amarenco P, Bogousslavsky J, Caplan LR, et al. The ASCOD phenotyping of ischemic stroke (updated ASCO phenotyping). *Cerebrovasc Dis* 2013;36:1-5.
- Wardlaw JM, Smith EE, Biessels GJ, et al. Neuroimaging standards for research into small vessel disease and its contribution to ageing and neurodegeneration. *Lancet Neurol* 2013;12:822-838.
- Wardlaw JM. What is a lacune? *Stroke* 2008;39:2921-2922.
- Moreau F, Patel S, Lauzon ML, et al. Cavitation after acute symptomatic lacunar stroke depends on time, location, and MRI sequence. *Stroke* 2012;43:1837-1842.
- Shi Y, Wardlaw JM. Update on cerebral small vessel disease: a dynamic whole-brain disease. *Stroke Vasc Neurol* 2016;1:83-92.
- Smith EE, Schneider JA, Wardlaw JM, et al. Cerebral microinfarcts: the invisible lesions. *Lancet Neurol* 2012;11:272-282.
- Pantoni L. Cerebral small vessel disease: from pathogenesis and clinical characteristics to therapeutic challenges. *Lancet Neurol* 2010;9:689-701.
- Chowdhury D, Wardlaw JM, Dennis MS. Are multiple acute small subcortical infarctions caused by embolic mechanisms? *J Neurol Neurosurg Psychiatry* 2004;75:1416-1420.
- Caplan LR. Lacunar infarction and small vessel disease: pathology and pathophysiology. *J stroke* 2015;17:2-6.
- Pasi M, Boulouis G, Fotiadis P, et al. Distribution of lacunes in cerebral amyloid angiopathy and hypertensive small vessel disease. *Neurology* 2017;88:2162-2168.

47. Del Bene A, Makin SDJ, Doubal FN. Variation in risk factors for recent small subcortical infarcts with infarct size, shape, and location. *Stroke* 2013;44:3000-3006.
48. Tan MYQ, Singhal S, Ma H, et al. Examining subcortical infarcts in the era of acute multimodality CT imaging. *Front Neurol* 2016;7:220.
49. Del Brutto OH, Mera RM, Gillman J, et al. Calcifications in the carotid siphon correlate with silent cerebral small vessel disease in community-dwelling older adults: a population-based study in rural Ecuador. *Geriatr Gerontol Int* 2016;16:1063-1067.
50. Song T-J, Kim YD, Yoo J, et al. Association between aortic atheroma and cerebral small vessel disease in patients with ischemic stroke. *J Stroke* 2016;18:312-320.
51. Park YS, Chung PW, Kim YB, et al. Small deep infarction in patients with atrial fibrillation: evidence of lacunar pathogenesis. *Cerebrovasc Dis* 2013;36:205-210.
52. Boulouis G, Charidimou A, Auriel E, et al. Intracranial atherosclerosis and cerebral small vessel disease in intracerebral hemorrhage patients. *J Neurol Sci* 2016;369:324-329.
53. Fisher CM, Caplan LR. Basilar artery branch occlusion: a cause of pontine infarction. *Neurology* 1971;21:900-905.
54. Fisher CM. Bilateral occlusion of basilar artery branches. *J Neurol Neurosurg Psychiatry* 1977;40:1182-1189.
55. Fisher CM. Thalamic pure sensory stroke: a pathologic study. *Neurology* 1978;28:1141-1144.
56. Fisher CM. Capsular infarcts: the underlying vascular lesions. *Arch Neurol* 1979;36:65-73.
57. Fisher CM, Tapia J. Lateral medullary infarction extending to the lower pons. *J Neurol Neurosurg Psychiatry* 1987;50:620-624.
58. Grinberg LT, Thal DR. Vascular pathology in the aged human brain. *Acta Neuropathol* 2010;119:277-290.
59. Thal DR, Grinberg LT, Attems J. Vascular dementia: different forms of vessel disorders contribute to the development of dementia in the elderly brain. *Exp Gerontol* 2012;47:816-824.
60. Lammie GA. Hypertensive cerebral small vessel disease and stroke. *Brain Pathol* 2002;12:358-370.
61. Staemmler M. Zur Lehre von der Entstehung des Schlaganfalles. *Klin Wschr* 1936;15:1300.
62. Wolff K. Untersuchungen und Bemerkungen zur Lehre von der hypertensiven apoplektischen Hirnblutung. *Virchows Arch Path Anat* 1937;299:573.
63. Scholz W, Nieto D. Studien zur Pathologie der Hirngefäße. I. Fibrose und Hyalinose. *Z ges Neurol Psychiat* 1938;162:675.
64. Matsuoka S. Studien über Hirnblutung und Erweichung (III. Mitteilung). Über kleine Aneurysmen in den Gehirnen ohne Blutung bzw. Blutungsfreien Hirnpartien. *Trans Soc Path Jap* 1939;29:449.
65. Feigin I, Prose P. Hypertensive fibrinoid arteritis of the brain and gross cerebral hemorrhage: a form of "hyalinosis". *Arch Neurol* 1959;1:98-110.
66. Bamford J, Sandercock P, Jones L, et al. The natural history of lacunar infarction: the Oxfordshire Community Stroke Project. *Stroke* 1987;18:545-551.
67. Poggesi A, Pasi M, Pescini F, et al. Circulating biologic markers of endothelial dysfunction in cerebral small vessel disease: a review. *J Cereb Blood Flow Metab* 2016;36:72-94.
68. Wardlaw JM, Smith C, Dichgans M. Mechanisms of sporadic cerebral small vessel disease: insights from neuroimaging. *Lancet Neurol* 2013;12:483-497.
69. Lassen NA, Agnoli A. The upper limit of autoregulation of cerebral blood flow—on the pathogenesis of hypertensive encephalopathy. *Scand J Clin Lab Invest* 1972;30:113-116.
70. Harper AM, Lassen NA, MacKenzie ET, et al. Proceedings: the upper limit of "autoregulation" of cerebral blood flow in the baboon. *J Physiol* 1973;234:61P-62P.
71. MacKenzie ET, Strandgaard S, Graham DJ, et al. Effects of acutely induced hypertension in cats on pial arteriolar caliber, local cerebral blood flow, and the blood-brain barrier. *Circ Res* 1976;39:33-41.
72. Hill GS. Studies on the pathogenesis of hypertensive vascular disease. Effect of high-pressure intra-arterial injections in rats. *Circ Res* 1970;27:657-668.
73. Byrom FB. The pathogenesis of hypertensive encephalopathy and its relation to the malignant phase of hypertension; experimental evidence from the hypertensive rat. *Lancet* 1954;267:201-211.
74. Byrom FB. Morbid effects of vasopressin on the organs and vessels of rats. *J Pathol Bacteriol* 1937;45:1-16.
75. Giwa MO, Williams J, Elderfield K, et al. Neuropathologic evidence of endothelial changes in cerebral small vessel disease. *Neurology* 2012;78:167-174.
76. Ahmed-Jushuf F, Jiwa NS, Arwani AS, et al. Age-dependent expression of VEGFR2 in deep brain arteries in small vessel disease, CADASIL, and healthy brains. *Neurobiol Aging* 2016;42:110-115.
77. Bagi Z, Brandner DD, Le P, et al. Vasodilator dysfunction and oligodendrocyte dysmaturation in aging white matter. *Ann Neurol* 2017;83:142-152.
78. Fredriksson K, Nordborg C, Kalimo H, et al. Cerebral microangiopathy in stroke-prone spontaneously hypertensive rats. An immunohistochemical and ultrastructural study. *Acta Neuropathol* 1988;75:241-252.
79. Wardlaw JM, Doubal FN, Valdes-Hernandez M, et al. Blood-brain barrier permeability and long-term clinical and imaging outcomes in cerebral small vessel disease. *Stroke* 2013;44:525-527.
80. Muñoz Maniega S, Chappell FM, Valdés Hernández MC, et al. Integrity of normal-appearing white matter: influence of age, visible lesion burden and hypertension in patients with small-vessel disease. *J Cereb Blood Flow Metab* 2017;37:644-656.
81. Arba F, Leigh R, Inzitari D, et al. Blood-brain barrier leakage increases with small vessel disease in acute ischemic stroke. *Neurology* 2017;89:2143-2150.
82. Gustafsson F. Hypertensive arteriolar necrosis revisited. *Blood Press* 1997;6:71-77.
83. Rosenblum WI. Miliary aneurysms and "fibrinoid" degeneration of cerebral blood vessels. *Hum Pathol* 1977;8:133-139.
84. Rosenblum WI. The importance of fibrinoid necrosis as the cause of cerebral hemorrhage in hypertension. *Commentary. J Neuropathol Exp Neurol*. 1993;52:11-13.
85. Kantachuesiri S, Fleming S, Peters J, et al. Controlled hypertension, a transgenic toggle switch reveals differential mechanisms underlying vascular disease. *J Biol Chem* 2001;276:36727-36733.
86. Veniant M, Menard J, Bruneval P, et al. Vascular damage without hypertension in transgenic rats expressing prorenin exclusively in the liver. *J Clin Invest* 1996;98:1966-1970.
87. Gibbons GH, Dzau VJ. The emerging concept of vascular remodeling. *N Engl J Med* 1994;330:1431-1438.
88. Bezerra DC, Sharrett AR, Matsushita K, et al. Risk factors for lacune subtypes in the Atherosclerosis Risk in Communities (ARIC) Study. *Neurology* 2012;78:102-108.

89. Lioutas V-A, Beiser A, Himali J, et al. Lacunar infarcts and intracerebral hemorrhage differences. *Stroke* 2017;48:486-489.
90. Altmann-Schneider I, van der Grond J, Slagboom PE, et al. Lower susceptibility to cerebral small vessel disease in human familial longevity: the Leiden Longevity Study. *Stroke* 2013;44:9-14.
91. Arntz RM, van den Broek SMA, van Uden IWM, et al. Accelerated development of cerebral small vessel disease in young stroke patients. *Neurology* 2016;87:1212-1219.
92. Poirier J, Derouesne C. Cerebral lacunae. A proposed new classification. *Clin Neuropathol* 1984;3:266.
93. Bokura H, Kobayashi S, Yamaguchi S. Distinguishing silent lacunar infarction from enlarged Virchow-Robin spaces: a magnetic resonance imaging and pathological study. *J Neurol* 1998;245:116-122.
94. Ding J, Sigurdsson S, Jónsson PV, et al. Large perivascular spaces visible on magnetic resonance imaging, cerebral small vessel disease progression, and risk of dementia. *JAMA Neurol* 2017;74:1105.
95. Tuszynski MH, Petito CK, Levy DE. Risk factors and clinical manifestations of pathologically verified lacunar infarctions. *Stroke* 1989;20:990-999.
96. Lammie GA, Brannan F, Wardlaw JM. Incomplete lacunar infarction (type Ib lacunes). *Acta Neuropathol* 1998;96:163-171.
97. Ma KC, Olsson Y. Structural and vascular permeability abnormalities associated with lacunes of the human brain. *Acta Neurol Scand* 1993;88:100-107.
98. Lassen NA. Incomplete cerebral infarction—focal incomplete ischemic tissue necrosis not leading to emolliation. *Stroke* 1982;13:522-523.
99. Fredriksson K, Kalimo H, Nordborg C, et al. Cyst formation and glial response in the brain lesions of stroke-prone spontaneously hypertensive rats. *Acta Neuropathol* 1988;76:441-450.
100. Nag S. Cerebral changes in chronic hypertension: combined permeability and immunohistochemical studies. *Acta Neuropathol* 1984;62:178-184.
101. Ogata J, Fujishima M, Tamaki K, et al. Vascular changes underlying cerebral lesions in stroke-prone spontaneously hypertensive rats. A serial section study. *Acta Neuropathol*. 1981;54:183-188.
102. Lammie A. The role of oedema in lacune formation. *Cerebrovasc Dis* 1998;8:246.
103. Regenhardt RW, Das AS, Stapleton CJ, et al. Blood pressure and penumbral sustenance in stroke from large vessel occlusion. *Front Neurol* 2017;8:317. <https://doi.org/10.3389/fneur.2017.00317>. eCollection 2017.
104. Sam K, Crawley AP, Poublic J, et al. Vascular dysfunction in leukoaraiosis. *Am J Neuroradiol* 2016;37:2258-2264.
105. van der Veen PH, Muller M, Vincken KL, et al. Longitudinal relationship between cerebral small-vessel disease and cerebral blood flow: the second manifestations of arterial disease-magnetic resonance study. *Stroke* 2015;46:1233-1238.
106. Bernbaum M, Menon BK, Fick G, et al. Reduced blood flow in normal white matter predicts development of leukoaraiosis. *J Cereb Blood Flow Metab* 2015;35:1610-1615.
107. Shi Y, Thrippleton MJ, Makin SD, et al. Cerebral blood flow in small vessel disease: a systematic review and meta-analysis. *J Cereb Blood Flow Metab* 2016;36:1653-1667.
108. Hinman JD, Lee MD, Tung S, et al. Molecular disorganization of axons adjacent to human lacunar infarcts. *Brain* 2015;138:736-745.
109. Gouw AA, van der Flier WM, Pantoni L, et al. On the etiology of incident brain lacunes: longitudinal observations from the LADIS study. *Stroke* 2008;39:3083-3085.
110. Carmichael ST. Emergent properties of neural repair: elemental biology to therapeutic concepts. *Ann Neurol* 2016;79:895-906.
111. Lo EH. A new penumbra: transitioning from injury into repair after stroke. *Nat Med* 2008;14:497-500.
112. Zhao B-Q, Wang S, Kim H-Y, et al. Role of matrix metalloproteinases in delayed cortical responses after stroke. *Nat Med* 2006;12:441-445.
113. Carmichael ST. Brain excitability in stroke: the yin and yang of stroke progression. *Arch Neurol* 2012;69:161-167.
114. Clarkson AN, Huang BS, Macisaac SE, et al. Reducing excessive GABA-mediated tonic inhibition promotes functional recovery after stroke. *Nature* 2010;468:305-309.
115. Xiao G, Hinman J. Concepts and opportunities for repair in cerebral microvascular disease and white matter stroke. *Neural Regen Res* 2016;11:1398-1400.
116. Hinman JD. The back and forth of axonal injury and repair after stroke. *Curr Opin Neurol* 2014;27:1.
117. Sozmen EG, Kolekar A, Havton LA, et al. A white matter stroke model in the mouse: axonal damage, progenitor responses and MRI correlates. *J Neurosci Methods* 2009;180:261-272.
118. Franklin RJM. Remyelination of the demyelinated CNS: the case for and against transplantation of central, peripheral and olfactory glia. *Brain Res Bull* 2002;57:827-832.
119. Shindo A, Liang AC, Maki T, et al. Subcortical ischemic vascular disease: roles of oligodendrocyte function in experimental models of subcortical white-matter injury. *J Cereb Blood Flow Metab* 2016;36:187-198.
120. Giger RJ, Venkatesh K, Chivatakarn O, et al. Mechanisms of CNS myelin inhibition: evidence for distinct and neuronal cell type specific receptor systems. *Restor Neurol Neurosci* 2008;26:97-115.
121. Sozmen EG, Rosenzweig S, Llorente IL, et al. Nogo receptor blockade overcomes remyelination failure after white matter stroke and stimulates functional recovery in aged mice. *Proc Natl Acad Sci* 2016;113:E8453-E8462.
122. Hayakawa K, Pham L-DD, Katusic ZS, et al. Astrocytic high-mobility group box 1 promotes endothelial progenitor cell-mediated neurovascular remodeling during stroke recovery. *Proc Natl Acad Sci USA* 2012;109:7505-7510.
123. Sofroniew MV. Molecular dissection of reactive astrogliosis and glial scar formation. *Trends Neurosci* 2009;32:638-647.
124. Schmidt R, Seiler S, Loitfelder M. Longitudinal change of small-vessel disease-related brain abnormalities. *J Cereb Blood Flow Metab* 2016;36:26-39.
125. Wardlaw JM, Chappell FM, Valdés Hernández M, et al. White matter hyperintensity reduction and outcomes after minor stroke. *Neurology* 2017;89:1003-1010.
126. Duering M, Csanadi E, Gesierich B, et al. Incident lacunes preferentially localize to the edge of white matter hyperintensities: insights into the pathophysiology of cerebral small vessel disease. *Brain* 2013;136:2717-2726.
127. Gesierich B, Duchesnay E, Jouvent E, et al. Features and determinants of lacune shape. *Stroke* 2016;47:1258-1264.
128. Brundel M, de Bresser J, van Dillen JJ, et al. Cerebral microinfarcts: a systematic review of neuropathological studies. *J Cereb Blood Flow Metab* 2012;32:425-436.
129. Wityk RJ. Cerebral cortical microinfarcts on 3-T magnetic resonance imaging. *JAMA Neurol* 2017;74:385.

130. van Veluw SJ, Hilal S, Kuijf HJ, et al. Cortical microinfarcts on 3T MRI: clinical correlates in memory-clinic patients. *Alzheimers Dement* 2015;11:1500-1509.
131. Hilal S, Sikking E, Shaik MA, et al. Cortical cerebral microinfarcts on 3T MRI: a novel marker of cerebrovascular disease. *Neurology* 2016;87:1583-1590.
132. Hilal S, Sikking E, Shaik MA, et al. Cortical cerebral microinfarcts on 3T MRI. *Neurology* 2016;87:1583-1590.
133. van Veluw SJ, Charidimou A, van der Kouwe AJ, et al. Microbleed and microinfarct detection in amyloid angiopathy: a high-resolution MRI-histopathology study. *Brain* 2016;139:3151-3162.
134. Auriel E, Westover MB, Bianchi MT, et al. Estimating total cerebral microinfarct burden from diffusion-weighted imaging. *Stroke* 2015;46:2129-2135.
135. Auriel E, Edlow BL, Reijmer YD, et al. Microinfarct disruption of white matter structure: a longitudinal diffusion tensor analysis. *Neurology* 2014;83:182-188.
136. Longstreth WT, Sonnen JA, Koepsell TD, et al. Associations between microinfarcts and other macroscopic vascular findings on neuropathologic examination in 2 databases. *Alzheimer Dis Assoc Disord* 2009;23:291-294.
137. Arvanitakis Z, Capuano AW, Leurgans SE, et al. The relationship of cerebral vessel pathology to brain microinfarcts. *Brain Pathol* 2017;27:77-85.
138. Auriel E, Gurol ME, Ayres A, et al. Characteristic distributions of intracerebral hemorrhage-associated diffusion-weighted lesions. *Neurology* 2012;79:2335-2341.
139. Oliveira-Filho J, Ay H, Shoamanesh A, et al. Incidence and etiology of microinfarcts in patients with ischemic stroke. *J Neuroimaging* 2018;28:406-411.
140. Tsai H-H, Pasi M, Tsai L-K, et al. Distribution of lacunar infarcts in asians with intracerebral hemorrhage: a magnetic resonance imaging and amyloid positron emission tomography study. *Stroke* 2018;49:1515-1517.
141. Smith EE, Lee J-M. Lacunes. *Neurology* 2017;88:2158-2159.
142. Charidimou A, Boulouis G, Haley K, et al. White matter hyperintensity patterns in cerebral amyloid angiopathy and hypertensive arteriopathy. *Neurology* 2016;86:505-511.
143. Chen YW, Gurol ME, Rosand J, et al. Progression of white matter lesions and hemorrhages in cerebral amyloid angiopathy 204. *Neurology* 2006;67:83-87.
144. Holland CM, Smith EE, Csapo I, et al. Spatial distribution of white-matter hyperintensities in Alzheimer disease, cerebral amyloid angiopathy, and healthy aging. *Stroke* 2008;39:1127-1133.
145. Charidimou A, Boulouis G, Pasi M, et al. MRI-visible perivascular spaces in cerebral amyloid angiopathy and hypertensive arteriopathy. *Neurology* 2017;88:1157-1164.
146. Martinez-Ramirez S, Pontes-Neto OM, Dumas AP, et al. Topography of dilated perivascular spaces in subjects from a memory clinic cohort. *Neurology* 2013;80:1551-1556.
147. Wiseman S, Marlborough F, Doubal F, et al. Blood markers of coagulation, fibrinolysis, endothelial dysfunction and inflammation in lacunar stroke versus non-lacunar stroke and non-stroke: systematic review and meta-analysis. *Cerebrovasc Dis* 2014;37:64-75.
148. Wiseman SJ, Doubal FN, Chappell FM, et al. Plasma biomarkers of inflammation, endothelial function and hemostasis in cerebral small vessel disease. *Cerebrovasc Dis* 2015;40:157-164.
149. Lavallée PC, Labreuche J, Faille D, et al. Circulating markers of endothelial dysfunction and platelet activation in patients with severe symptomatic cerebral small vessel disease. *Cerebrovasc Dis* 2013;36:131-138.
150. Rouhl RP, Mertens AE, van Oostenbrugge RJ, et al. Angiogenic T-cells and putative endothelial progenitor cells in hypertension-related cerebral small vessel disease. *Stroke* 2012;43:256-258.
151. Lee H-B, Kim J, Kim S-H, et al. Association between serum alkaline phosphatase level and cerebral small vessel disease Shimomura T, editor *PLoS One* 2015;10:e0143355.
152. Mitaki S, Nagai A, Oguro H, Yamaguchi S. C-reactive protein levels are associated with cerebral small vessel-related lesions. *Acta Neurol Scand* 2016;133:68-74.
153. Brown CM, Bushnell CD, Samsa GP, et al. Chronic systemic immune dysfunction in African-Americans with small vessel-type ischemic stroke. *Transl Stroke Res* 2015;6:430-436.
154. Rouhl RP, Damoiseaux JG, Lodder J, et al. Vascular inflammation in cerebral small vessel disease. *Neurobiol Aging* 2012;33:1800-1806.
155. Boehme AK, McClure LA, Zhang Y, et al. Inflammatory markers and outcomes after lacunar stroke. *Stroke* 2016;47:659-667. [STROKEAHA.115.012166](https://doi.org/10.1161/STROKEAHA.115.012166).
156. Yang L, Lv P, Ai W, et al. Lipidomic analysis of plasma in patients with lacunar infarction using normal-phase/reversed-phase two-dimensional liquid chromatography-quadrupole time-of-flight mass spectrometry. *Anal Bioanal Chem* 2017;409:3211-3222.
157. Song T-J, Chang Y, Shin M-J, et al. Low levels of plasma omega 3-polyunsaturated fatty acids are associated with cerebral small vessel diseases in acute ischemic stroke patients. *Nutr Res* 2015;35:368-374.
158. Datta A, Chen CP, Sze SK. Discovery of prognostic biomarker candidates of lacunar infarction by quantitative proteomics of microvesicles enriched plasma Koomen JM, editor *PLoS One* 2014;9:e94663.
159. Thompson CS, Hakim AM. Living beyond our physiological means: small vessel disease of the brain is an expression of a systemic failure in arteriolar function: A unifying hypothesis. *Stroke* 2009;40:322-331.
160. Martinez-Vea A, Salvado E, Bardaji A, et al. Silent cerebral white matter lesions and their relationship with vascular risk factors in middle-aged predialysis patients with CKD. *Am J Kidney Dis* 2006;47:241-250.
161. Khatri M, Wright CB, Nickolas TL, et al. Chronic kidney disease is associated with white matter hyperintensity volume: the Northern Manhattan Study (NOMAS). *Stroke* 2007;38:3121-3126.
162. Lammie GA, Brannan F, Slattery J, et al. Nonhypertensive cerebral small-vessel disease. An autopsy study. *Stroke* 1997;28:2222-2229.
163. Jerrard-Dunne P, Cloud G, Hassan A, et al. Evaluating the genetic component of ischemic stroke subtypes: a family history study. *Stroke* 2003;34:1364-1369.
164. Vermeer SE, Prins ND, den Heijer T, et al. Silent brain infarcts and the risk of dementia and cognitive decline. *N Engl J Med* 2003;348:1215-1222.
165. Debette S, Markus HS. The clinical importance of white matter hyperintensities on brain magnetic resonance imaging: systematic review and meta-analysis. *BMJ* 2010;341:c3666.
166. Carmelli D, DeCarli C, Swan GE, et al. Evidence for genetic variance in white matter hyperintensity volume in normal elderly male twins. *Stroke* 1998;29:1177-1181.
167. Atwood LD, Wolf PA, Heard-Costa NL, et al. Genetic variation in white matter hyperintensity volume in the Framingham study. *Stroke* 2004;35:1609-1613.

168. Bersano A, Baron P, Lanfranconi S, et al. Lombardia GENS: a collaborative registry for monogenic diseases associated with stroke. *Funct Neurol* 2012;27:107-117.
169. Choi JC. Genetics of cerebral small vessel disease. *J Stroke* 2015;17:7-16.
170. Lanfranconi S, Markus HS. COL4A1 mutations as a monogenic cause of cerebral small vessel disease: a systematic review. *Stroke* 2010;41:e513-e518.
171. Jen J, Cohen AH, Yue Q, et al. Hereditary endotheliopathy with retinopathy, nephropathy, and stroke (HERNS). *Neurology* 1997;49:1322-1330.
172. Meschia JF. Ischemic stroke as a complex genetic disorder. *Semin Neurol* 2006;26:49-56.
173. Tabor HK, Risch NJ, Myers RM. Candidate-gene approaches for studying complex genetic traits: practical considerations. *Nat Rev* 2002;3:391-397.
174. Elbaz A, Poirier O, Moulin T, et al. Association between the Glu298Asp polymorphism in the endothelial constitutive nitric oxide synthase gene and brain infarction. *The GENIC Investigators. Stroke* 2000;31:1634-1639.
175. Sharma P. Meta-analysis of the ACE gene in ischaemic stroke. *J Neurol Neurosurg Psychiatry* 1998;64:227-230.
176. Markus HS, Barley J, Lunt R, et al. Angiotensin-converting enzyme gene deletion polymorphism. A new risk factor for lacunar stroke but not carotid atheroma. *Stroke* 1995;26:1329-1333.
177. de Leeuw FE, Richard F, De Groot JC, et al. Interaction between hypertension, apoE, and cerebral white matter lesions. *Stroke* 2004;35:1057-1060.
178. Davignon J, Gregg RE, Sing CF. Apolipoprotein E polymorphism and atherosclerosis. *Arteriosclerosis* 1988;8:1-21.
179. Huang Y. A β -independent roles of apolipoprotein E4 in the pathogenesis of Alzheimer's disease. *Trends Mol Med* 2010;16:287-294.
180. Leys D, Pasquier F. Editorial comment—not all hypertensive subjects have similar risks for white matter lesions: influence of genetic factors. *Stroke* 2004;35:1061-1062.
181. Bevan S, Markus HS. Genetics of common polygenic ischaemic stroke: current understanding and future challenges. *Stroke Res Treat* 2011;2011:179061.
182. Paternoster L, Chen W, Sudlow CL. Genetic determinants of white matter hyperintensities on brain scans: a systematic assessment of 19 candidate gene polymorphisms in 46 studies in 19,000 subjects. *Stroke* 2009;40:2020-2026.
183. Kubo M, Hata J, Ninomiya T, et al. A nonsynonymous SNP in PRKCH (protein kinase C ϵ) increases the risk of cerebral infarction. *Nat Genet* 2007;39:212-217.
184. Serizawa M, Nabika T, Ochiai Y, et al. Association between PRKCH gene polymorphisms and subcortical silent brain infarction. *Atherosclerosis* 2008;199:340-345.
185. Fornage M, Dobbins M, Bis JC, et al. Genome-wide association studies of cerebral white matter lesion burden: the CHARGE consortium. *Ann Neurol* 2011;69:928-939.
186. Gretarsdottir S, Sveinbjornsdottir S, Jonsson HH, et al. Localization of a susceptibility gene for common forms of stroke to 5q12. *Am J Hum Genet* 2002;70:593-603.
187. Gretarsdottir S, Thorleifsson G, Reynisdottir ST, et al. The gene encoding phosphodiesterase 4D confers risk of ischemic stroke. *Nat Genet* 2003;35:131-138.
188. van Rijn MJ, Slooter AJ, Schut AF, et al. Familial aggregation, the PDE4D gene, and ischemic stroke in a genetically isolated population. *Neurology* 2005;65:1203-1209.
189. Traylor M, Bevan S, Baron JC, et al. Genetic architecture of lacunar stroke. *Stroke* 2015;46:2407-2412.
190. Zhang Y, Tong Y, Zhang Y, et al. Two novel susceptibility SNPs for ischemic stroke using exome sequencing in Chinese Han population. *Mol Neurobiol* 2014;49:852-862.
191. Cole JW, Stine OC, Liu X, et al. Rare variants in ischemic stroke: an exome pilot study. *PLoS One* 2012;7:e35591.
192. King JY, Ferrara R, Tabibiazar R, et al. Pathway analysis of coronary atherosclerosis. *Physiol Genomics* 2005;23:103-118.
193. Rampersaud E, Damcott CM, Fu M, et al. Identification of novel candidate genes for type 2 diabetes from a genome-wide association scan in the old order Amish: evidence for replication from diabetes-related quantitative traits and from independent populations. *Diabetes* 2007;56:3053-3062.
194. Bersano A, Zuffardi O, Pantoni L, et al. Next generation sequencing for systematic assessment of genetics of small-vessel disease and lacunar stroke. *J Stroke Cerebrovasc Dis* 2015;24:759-765.
195. Hainsworth AH, Brittain JF, Khatun H. Pre-clinical models of human cerebral small vessel disease: utility for clinical application. *J Neurol Sci* 2012;322:237-240.
196. Regenhardt RW, Mecca AP, Desland F, et al. Centrally administered angiotensin-(1-7) increases the survival of stroke-prone spontaneously hypertensive rats. *Exp Physiol* 2014;99:442-453.
197. Bailey EL, McBride MW, Beattie W, et al. Differential gene expression in multiple neurological, inflammatory and connective tissue pathways in a spontaneous model of human small vessel stroke. *Neuropathol Appl Neurobiol* 2014;40:855-872.
198. Okamoto K, Yamori Y, Nagaoka A. Establishment of the stroke-prone spontaneously hypertensive rat (SHR). *Circ Res* 1974;34:143.
199. Yamori Y, Horie R, Sato M, et al. Experimental studies on the pathogenesis and prophylaxis of stroke in stroke-prone spontaneously hypertensive rats (SHR). (1) Quantitative estimation of cerebrovascular permeability. *Jpn Circ J* 1975;39:611-615.
200. Fredriksson K, Auer RN, Kalimo H, et al. Cerebrovascular lesions in stroke-prone spontaneously hypertensive rats. *Acta Neuropathol* 1985;68:284-294.
201. Yamori Y, Horie R, Handa H, et al. Pathogenetic similarity of strokes in stroke-prone spontaneously hypertensive rats and humans. *Stroke* 1976;7:46-53.
202. Bailey EL, Wardlaw JM, Graham D, et al. Cerebral small vessel endothelial structural changes predate hypertension in stroke-prone spontaneously hypertensive rats: a blinded, controlled immunohistochemical study of 5- to 21-week-old rats. *Neuropathol Appl Neurobiol* 2011;37:711-726.
203. Bailey EL, Smith C, Sudlow CL, et al. Is the spontaneously hypertensive stroke prone rat a pertinent model of sub cortical ischemic stroke? A systematic review. *Int J Stroke* 2011;6:434-444.
204. Herisson F, Zhou I, Mawet J, et al. Posterior reversible encephalopathy syndrome in stroke-prone spontaneously hypertensive rats on high-salt diet. *J Cereb Blood Flow Metab* 2018;19. <https://doi.org/10.1177/0271678X17752795>. [Epub ahead of print].
205. Sozmen EG, Hinman JD, Carmichael ST. Models that matter: white matter stroke models. *Neurotherapeutics* 2012;9:349-358.
206. Irving EA, Yatsushiro K, McCulloch J, et al. Rapid alteration of tau in oligodendrocytes after focal ischemic injury in the rat: involvement of free radicals. *J Cereb Blood Flow Metab* 1997;17:612-622.

207. Regenhardt RW, Bennion DM, Sumners C. Cerebroprotective action of angiotensin peptides in stroke. *Clin Sci* 2014;126:195-205.
208. Regenhardt RW, Desland F, Mecca AP, et al. Anti-inflammatory effects of angiotensin-(1-7) in ischemic stroke. *Neuropharmacology* 2013;71:154-163.
209. Sarti C, Pantoni L, Bartolini L, et al. Cognitive impairment and chronic cerebral hypoperfusion: what can be learned from experimental models. *J Neurol Sci* 2002;203-204:263-266.
210. Farkas E, Luiten PG, Bari F. Permanent, bilateral common carotid artery occlusion in the rat: a model for chronic cerebral hypoperfusion-related neurodegenerative diseases. *Brain Res Rev* 2007;54:162-180.
211. Farkas E, Donka G, de Vos RA, et al. Experimental cerebral hypoperfusion induces white matter injury and microglial activation in the rat brain. *Acta Neuropathol* 2004;108:57-64.
212. Wakita H, Tomimoto H, Akiguchi I, et al. Axonal damage and demyelination in the white matter after chronic cerebral hypoperfusion in the rat. *Brain Res* 2002;924:63-70.
213. Hattori Y, Enmi J-I, Kitamura A, et al. A Novel mouse model of subcortical infarcts with dementia. *J Neurosci* 2015;35:3915-3928.
214. Farkas E, Luiten PG. Cerebral microvascular pathology in aging and Alzheimer's disease. *Prog Neurobiol* 2001;64:575-611.
215. Shih AY, Blinder P, Tsai PS, et al. The smallest stroke: occlusion of one penetrating vessel leads to infarction and a cognitive deficit. *Nat Neurosci* 2013;16:55-63.
216. Rosenzweig S, Carmichael ST. Age-dependent exacerbation of white matter stroke outcomes: a role for oxidative damage and inflammatory mediators. *Stroke* 2013;44:2579-2586.
217. Yang Y, Kimura-Ohba S, Thompson J, et al. Rodent models of vascular cognitive impairment. *Transl Stroke Res* 2016;7:407-414.
218. Weng Y-C, Sonni A, Labelle-Dumais C, et al. COL4A1 mutations in patients with sporadic late-onset intracerebral hemorrhage. *Ann Neurol* 2012;71:470-477.
219. Joutel A, Faraci FM. Cerebral small vessel disease: insights and opportunities from mouse models of collagen IV-related small vessel disease and cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy. *Stroke* 2014;45:1215-1221.
220. Naik P, Cucullo L. In vitro blood-brain barrier models: current and perspective technologies. *J Pharm Sci* 2012;101:1337-1354.
221. Das AS, Regenhardt RW, Vernooij MW, Blacker D, Charidimou A, Viswanathan A. Asymptomatic Cerebral Small Vessel Disease: Insights from Population-Based Studies. *J Stroke* 2019. <https://doi.org/10.5853/jos.2018.03608>. [Epub ahead of print].