



Selective Targeting of Perirhinal Cortex Projection to Hippocampal CA1 Interneurons

Xiang Li¹ · Yiding Li¹ · Junhui Zhang² · Xiaohui Zhang¹

Received: 7 September 2018 / Accepted: 13 October 2018 / Published online: 16 March 2019
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Dear Editor,

The perirhinal cortex (PER) is conceptually important in the recognition memory, especially in familiarity discrimination, whereas the hippocampus is important for association and recollection [1]. This notion is known as the dual-process model of recognition memory. An alternative idea suggests that, like the hippocampus, the PER contributes to both familiarity and recollection [2], known for the unitary process model. Both models have made strong emphasis on a vital role of the PER and hippocampus in recognition memory, suggesting that they are highly interconnected. Indeed, previous animal studies using anterograde and retrograde tracers revealed direct anatomic projections from the PER to hippocampus in rodents and monkeys [3–5]. However, the latter projection is less prominent compared to that of PER to entorhinal cortex (EC) or that of EC to hippocampus. This may implicate that information flow from PER to hippocampus is primarily through an indirect path, where the EC is a major mediator [3–6]. Consistently, the *in vivo* electrical stimulation results suggested a weak functional projection from PER directly

to hippocampal dentate gyrus (DG) and CA1 areas, respectively [7, 8].

It remains unknown how the PER projections innervate different types of hippocampal CA1 neurons. To address this question, we used the channelrhodopsin-2 (ChR2) assisted circuit mapping (CRACM) method [9, 10] to map direct functional connections from the PER to dorsal hippocampal CA1 (d-HP CA1). Similar to our recent study that mapped the EC direct projection to dCA1 [10], we firstly injected the *AAV-hsynapsin-ChR2-mcherry* into the PER in the wild-type C57BL/6 mice at postnatal days 14–16, in order to achieve selective expression of light-activated optogenetic channels channelrhodopsin-2 (ChR2) in the PER neurons. After 4 weeks, ChR2(-mcherry) expression could be detected in the PER neurons and their projection axons within the DG molecular (MO) layer and the CA1 stratum lacunosum-moleculare (SLM) layer in the dorsal hippocampus (Fig. 1A), which is consistent with the previous observation [6].

In the CRACM experiment as previously described [10], activating those ChR2-expressing PER axons in the dCA1 with a 10×10 grid of laser beam stimuli on acute hippocampal slices allowed us to specifically map out their synaptic connections to targeted hippocampal neurons. The grid of 473-nm laser spot stimuli spaced by 40- μ m intervals, covering the CA1 SLM and DG MO layers, was used to activate ChR2 channels on the PER axons (Fig. 1B). The resultant excitation by 0.5 mW–2 mW laser pulse (1-ms duration) could elicit synaptic transmission of direct PER-CA1 connections on distal apical dendrites of a dCA1 neuron so that excitatory postsynaptic currents (EPSC_{CRACM}) were detected by whole-cell recording on the soma. The mean latency of evoked EPSC_{CRACM} was 5.84 ± 0.40 ms ($n = 5$), and averaged EPSC_{CRACM} amplitudes of corresponding stimulation locations were

Electronic supplementary material The online version of this article (<https://doi.org/10.1007/s12264-019-00363-y>) contains supplementary material, which is available to authorized users.

✉ Junhui Zhang
jhzhang933@sohu.com

✉ Xiaohui Zhang
xhzhang@bnu.edu.cn

¹ State Key Laboratory of Cognitive Neuroscience & Learning, IDG/McGovern Institute for Brain Research, Beijing Normal University, Beijing 100875, China

² Department of Joint Surgery, The 6th Hospital of Ningbo, Ningbo 31500, China

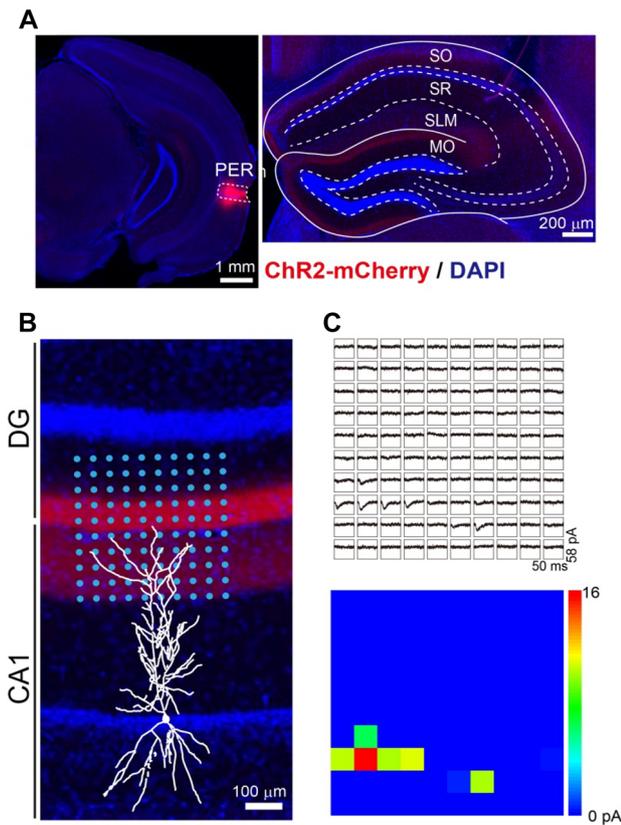


Fig. 1 Viral expression of ChR2 in the PER and CRACM of the PER-dCA1 connections. **A** Fluorescence images showing ChR2(-mcherry) expression in the PER 4 weeks after a stereotaxic injection of *AAV-hsynapsin-ChR2-mcherry* into the PER (left) and the distribution of PER axons in the dorsal hippocampus (right). **B** Diagram of CRACM experiment with a 10×10 grid of laser spot stimulation. **C** Averaged EPSC_{CRACM} traces of 5 sweeps of laser stimulation to individual locations (upper) and the heat-map of EPSC_{CRACM} amplitude (bottom) for a recorded CA1 PC.

transformed into a spatial heat-map showing dendritic field of the synapse (Fig. 1C). Cells with no apparent EPSC_{CRACM} under laser stimuli at maximal 5 mW laser power were defined as the unresponsive cells, indicating no direct functional connections.

Based on post-recording construction of neuronal morphology, the recorded excitatory pyramidal cells (PCs) were located mainly within the CA1 stratum pyramidale (SP) layer and were further differentiated into two major subtypes, the simple PC (sPC) and complex PC (cPC) according to our recent study [10]. Typically, sPCs had a single apical dendritic trunk and their cell body preferentially resided in the deep SP layer, while cPCs distinctively exhibited twin apical trunks that branched out a position near the soma (Fig. 2A). Meanwhile, the recorded inhibitory interneurons (INs) were defined mainly based on reconstructed neuronal morphology and their location within the SR-layer [11]. In total, we recorded 25 sPCs, 24

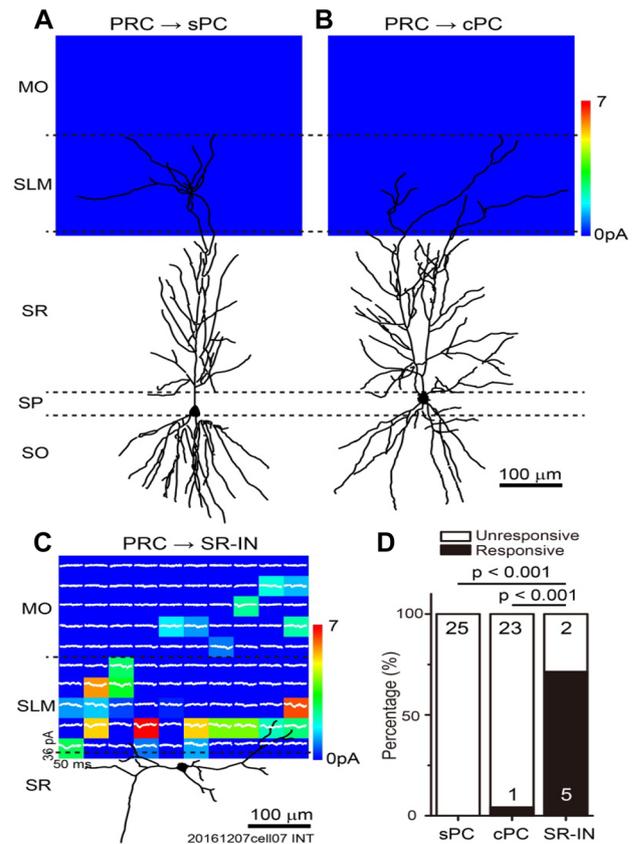


Fig. 2 PER axons selectively innervate dCA1 inhibitory interneurons. **A–C** The responsive dendritic heat-maps for the PER inputs (EPSC_{CRACM} activated by laser spots at 2-mW) overlaid on the reconstructed morphologies of excitatory sPC (**A**) or cPC (**B**) within the SP layer or an inhibitory IN in the SR layer (**C**). **D** Percentages of sPCs, cPCs and SR-INs that were responsive (with EPSC_{CRACM}, black) or unresponsive (white) to laser activation of ChR2-expressing PER axons at maximum 5 mW power. The *P* value: Fisher exact test; *n*: number of examined cells.

cPCs and 7 inhibitory INs located within the SR and SLM layers (SR-IN) in the dCA1, respectively, in the hippocampal slices prepared from 4 mice four weeks after stereotaxic injection of *AAV-hSyn-ChR2* in the PER. We found that no EPSC_{CRACM} was detected in all examined sPCs (100% non-responsive, $n = 25$) and in nearly all cPCs (95.8% non-responsive, $n = 24$) under stimulation of laser beam at maximal power (Fig. 2A, B, D). In contrast, evoked EPSC_{CRACM} events could be detected in 5 out of 7 recorded inhibitory SR-INs. It was noted that the soma of responsive INs resided mainly along the border of SLM and SR layers (Fig. 2C, D; Fig. S1). These results suggest that in the dCA1, PER projection axons preferentially form their direct excitatory synapses on the inhibitory SR-INs, but innervate very few excitatory PCs. Such distinct PER-CA1 connections can result in a feed-forward inhibition, rather than excitation, to the CA1 principal PC population when the PER is activated.

This finding of selective targeting of the PER projection to hippocampal CA1 inhibitory INs can provide an circuit basis for a previous finding that electric stimulation to the PER elicited a rapid field potential sink (or excitation) in the DG MO layer only, but not in the CA1 SLM layer [7, 8]. These results together support a notion that the PER provide very few excitatory inputs to most CA1 PCs. Anatomically, PER projection axons were found to be sparsely dispersed within the CA1 SLM layer and substantially less dense in other hippocampal areas compared to the EC projections [3–6].

In addition to its direct projection to hippocampal CA1 INs (Fig. 2), the PER also indirectly transfer synaptic information into the CA1 *via* the EC-hippocampus path. It is supported by the observation of dense PER projection axons in the superficial layers of both lateral and medial parts of EC [5, 6, 12, 13]. EC granule cells and PCs in the superficial 2/3 layers are known to send their excitatory projections into hippocampal DG and CA1 areas, respectively [10, 14]. Moreover, the direct PER projection could also be observed to the subiculum [8, 15], one of major downstream recipients of hippocampal CA1 outputs. Thus, it is of interest to further elucidate how the PER is interwired with other parts of hippocampal network and how these PER-associated circuits differentially contribute to hippocampus-dependent recognition memory.

Acknowledgements This work was supported by grants from the Beijing Municipal Science & Technology Commission (Z181100001518001) and the Interdisciplinary Research Funds of Beijing Normal University.

References

1. Brown MW, Aggleton JP. Recognition memory: what are the roles of the perirhinal cortex and hippocampus? *Nat Rev Neurosci* 2001, 2: 51–61.

2. Suzuki WA, Naya Y. The perirhinal cortex. *Annu Rev Neurosci* 2014, 37: 39–53.
3. Witter MP, Groenewegen HJ, Lopes da Silva FH, Lohman AH. Functional organization of the extrinsic and intrinsic circuitry of the parahippocampal region. *Prog Neurobiol* 1989, 33: 161–253.
4. Burwell RD, Witter MP, Amaral DG. Perirhinal and postrhinal cortices of the rat: a review of the neuroanatomical literature and comparison with findings from the monkey brain. *Hippocampus* 1995, 5: 390–408.
5. Lavenex P, Amaral DG. Hippocampal-neocortical interaction: a hierarchy of associativity. *Hippocampus* 2000, 10: 420–430.
6. Burwell RD, Amaral DG. Perirhinal and postrhinal cortices of the rat: Interconnectivity and connections with the entorhinal cortex. *J Compar Neurol* 1998, 391: 293–321.
7. Canning KJ, Leung LS. Lateral entorhinal, perirhinal, and amygdala-entorhinal transition projections to hippocampal CA1 and dentate gyrus in the rat: a current source density study. *Hippocampus* 1997, 7: 643–655.
8. Naber PA, Witter MP, Lopez da Silva, FH. Perirhinal cortex input to the hippocampus in the rat: evidence for parallel pathways, both direct and indirect. A combined physiological and anatomical study. *Eur J Neurosci* 1999, 11: 4119–4133.
9. Petreanu L, Mao T, Sternson SM, Svoboda K. The subcellular organization of neocortical excitatory connections. *Nature* 2009, 457: 1142–1145.
10. Li YD, Xu JM, Liu YF, Zhu J, Liu N, Zeng WB, *et al.* A distinct entorhinal cortex to hippocampal CA1 direct circuit for olfactory associative learning. *Nat Neurosci* 2017, 20: 559–570.
11. Freund TF, Buzsaki G. Interneurons of the hippocampus. *Hippocampus* 1996, 6: 347–470.
12. Furtak SC, Wei SM, Agster KL, Burwell RD. Functional neuroanatomy of the parahippocampal region in the rat: The perirhinal and postrhinal cortices. *Hippocampus* 2007, 17: 709–722.
13. Biella G, Uva L, de Curtis M. Propagation of neuronal activity along the neocortical–perirhinal–entorhinal pathway in the guinea pig. *J Neurosci* 2002, 22: 9972–9979.
14. van Strien NM, Cappaert NLM, Witter MP. The anatomy of memory: an interactive overview of the parahippocampal-hippocampal network. *Nat Rev Neurosci* 2009, 10: 272–282.
15. Kosel KC, Van Hoesen GW, Rosene DL. A direct projection from the perirhinal cortex (area 35) to the subiculum in the rat. *Brain Res* 1983, 269: 347–351.