



Alteration in NMDA Receptor Mediated Glutamatergic Neurotransmission in the Hippocampus During Senescence

Ashok Kumar¹ · Thomas C. Foster^{1,2}

Received: 13 June 2018 / Revised: 7 September 2018 / Accepted: 8 September 2018 / Published online: 12 September 2018
© Springer Science+Business Media, LLC, part of Springer Nature 2018

Abstract

Glutamate is the primary excitatory neurotransmitter in neurons and glia. *N*-methyl-D-aspartate (NMDA), α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA), and kainate receptors are major ionotropic glutamate receptors. Glutamatergic neurotransmission is strongly linked with Ca^{2+} homeostasis. Research has provided ample evidence that brain aging is associated with altered glutamatergic neurotransmission and Ca^{2+} dysregulation. Much of the work has focused on the hippocampus, a brain region critically involved in learning and memory, which is particularly susceptible to dysfunction during senescence. The current review examines Ca^{2+} regulation with a focus on the NMDA receptors in the hippocampus. Integrating the knowledge of the complexity of age-related alterations in Ca^{2+} homeostasis and NMDA receptor-mediated glutamatergic neurotransmission will positively shape the development of highly effective therapeutics to treat brain disorders including cognitive impairment.

Keywords Aging · Calcium homeostasis · Hippocampus · Glutamatergic neurotransmission · *N*-Methyl-D-aspartate receptor · Synaptic function · LTP · LTD

Introduction

The hypothesized role of senescent glutamatergic synapses in age-related memory decline is founded on studies examining the mechanisms for age-related changes in calcium (Ca^{2+})-dependent synaptic plasticity. The current review focuses on assessing age-associated changes in Ca^{2+} regulation and *N*-methyl-D-aspartate (NMDA) receptor-mediated synaptic transmission, in the hippocampus. In particular, we focus on possible mechanisms for an age-related decline in the function of NMDA glutamate receptors. The activity of NMDA receptors is critical for synaptic plasticity, long-term

potentiation (LTP), and long-term depression (LTD), in the hippocampus.

An Age-Related Shift in Synaptic Plasticity

LTP is a rapid and long lasting increase in synaptic transmission in response to intense synaptic activity. The induction of LTP requires activation of postsynaptic NMDA receptors resulting in a large, yet brief, influx of Ca^{2+} through the NMDA receptor channel. In turn, this large rise in intracellular Ca^{2+} activates Ca^{2+} sensitive kinases. Kinase activity increases the strength of the synaptic response through phosphorylation of α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) glutamate receptors, which leads to insertion of additional AMPA–glutamate channels into the post-synaptic membrane [1]. Early studies demonstrated that decay of LTP increased in aged animals and correlated with forgetting, suggesting that impaired acquisition and retention of information were related to impairment in the induction and maintenance of LTP [2].

In contrast to LTP, which requires a brief but large increase in intracellular Ca^{2+} , LTD is induced by a modest and prolonged rise in intracellular Ca^{2+} . The small and sustained rise in Ca^{2+} activates Ca^{2+} -sensitive phosphatases that

Special Issue: In honor of Eli Michaelis.

✉ Ashok Kumar
kash@ufl.edu

✉ Thomas C. Foster
foster1@ufl.edu

¹ Department of Neuroscience, McKnight Brain Institute, University of Florida, PO Box 100244, Gainesville, FL 32610-0244, USA

² Genetics and Genomics Program, University of Florida, Gainesville, FL 32611, USA

decrease synaptic transmission through dephosphorylation of AMPA–glutamate receptors, resulting in their removal from the post-synaptic membrane [3]. Initial studies of LTD indicated developmental regulation, such that the ability to induce LTD in the hippocampus declined from neonatal to adult periods. Thus, the discovery that aged animals exhibit increase in susceptibility to induction of LTD was unexpected [4–7]. The increase in susceptibility to LTD was shown to contribute to the decay of LTP and the induction or magnitude of LTD correlated with increased forgetting in older animals [7–9]. The results implicate LTD as a mechanism for enhancing the decay of LTP and an increased level of forgetting observed with advanced age.

The shift in synaptic plasticity during aging, favoring LTD over LTP, likely contributes to other correlates of cognitive aging including a decrease in synaptic strength and reduced transmission through the hippocampus of older animals [8, 9]. In turn, the weakening of synaptic transmission could contribute to the decrease in activation of the hippocampus of aging-memory impaired humans, recorded as a decrease in the functional magnetic resonance imaging (fMRI) blood oxygen-level dependent (BOLD) signal during learning or recall [10]. Moreover, LTD is involved in synapse removal, such that LTD may decrease synaptic connectivity and contribute to the reduction in hippocampal volume [11–13]. Together, these results point to Ca^{2+} dysregulation as a mechanism for senescent physiology characterized by decreased LTP, increased LTD, and decreased synaptic transmission.

Ca^{2+} -Dysregulation with Advance Age

Thirty years ago, observations of age-related changes in how neurons handle Ca^{2+} led to the formulation of the Ca^{2+} hypothesis of brain aging [14]. The initial Ca^{2+} hypothesis proposed that a small and prolonged increase in Ca^{2+} would, over time, have toxic effects, resulting in neuronal death, similar to that observed following a large increase over a short period. Initial studies provided an evidence for a small and sustained upsurge in intracellular free Ca^{2+} with advance age [15]. Due to the discoveries that normal aging is not associated with a loss of neurons [16, 17], the hypothesis has changed to reflect altered Ca^{2+} -dependent physiology, including senescent synaptic function [8, 18, 19].

The Ca^{2+} ion is a central signaling molecule in numerous cellular functions including apoptosis, energy production, gene regulation, cell proliferation, membrane excitability, synaptic transmission, and plasticity. Due to the ubiquitous nature of Ca^{2+} signaling, Ca^{2+} is one of the most highly regulated ions with the concentration inside the cell maintained at a level 10,000 times lower than the concentration in the extracellular space [20–22]. Accordingly, any change in Ca^{2+} regulating mechanisms can result in an alteration

in cell function. Age related changes have been reported for Ca^{2+} -buffering and extrusion mechanisms [23–25]. In addition, aging hippocampal neurons exhibit a shift in the sources of Ca^{2+} . Moreover, the shift in level of different Ca^{2+} sources likely results in changes in the subcellular localization of Ca^{2+} at the synapse, dendrite, and soma. During neural activity, intracellular Ca^{2+} rises at the synapse mainly due to influx of Ca^{2+} into the cell through NMDA receptors. In the soma and dendrites, Ca^{2+} signals arise due to receptor activity; however, much of the Ca^{2+} arises from voltage-dependent Ca^{2+} channels (VDCCs) and release of Ca^{2+} from intercellular Ca^{2+} stores (ICS). In the case of aging, Ca^{2+} from NMDA receptors appears to decrease and that from VDCCs and ICS increases (Fig. 1).

The NMDA Receptor

NMDA receptors represent one of the ligand-gated non-selective cation ionotropic glutamate receptors, which are present in high density within the hippocampus and play pivotal physiological and pathophysiological roles in the central nervous system [26–28]. NMDA receptors are hetero-tetrameric protein complexes composed of two classes of related subunits from seven homologous genes, GluN1, GluN2A–GluN2D, and GluN3A–GluN3B [29–36]. The majority of NMDA receptors are assemblies of two GluN1 subunits, the ubiquitously expressed and obligatory subunit, and two GluN2A–D subunits, a modulatory subunit. In addition, GluN3 subunits (GluN3A and GluN3B), without involving GluN2 subunits, can assemble with GluN1 subunits to form functional receptors [35, 37–40]. Developmentally, the expression of GluN1, GluN2B, and GluN3A decreases with age compared to adulthood, while an increase in the expression of GluN2A and GluN3B is reported during development [40].

NMDA receptors, along with AMPA and kainite receptors, are critical for the rapid regulation of synaptic plasticity including LTP and LTD, which are important cellular correlates for learning and memory function [41–47]. The induction of LTP and LTD involves activation of NMDA receptors, Ca^{2+} entry, and differential activation of kinases/phosphatases [18]. Interestingly, recent work indicates that amyloid beta could act on GluN2B subunits, through metabotropic mechanisms, to influence phosphatase/kinase activity, influencing synaptic function and spine loss [48, 49].

Physiological studies consistently indicate that NMDA receptor mediated excitatory postsynaptic potentials in the Schaffer collateral pathway of the hippocampus are reduced by approximately 50–60% in aged animals [50–57]. In turn, a decrease in NMDA receptor function is likely to influence induction of AMPA receptor mediated synaptic plasticity [48, 58] (i.e. metaplasticity [59]).

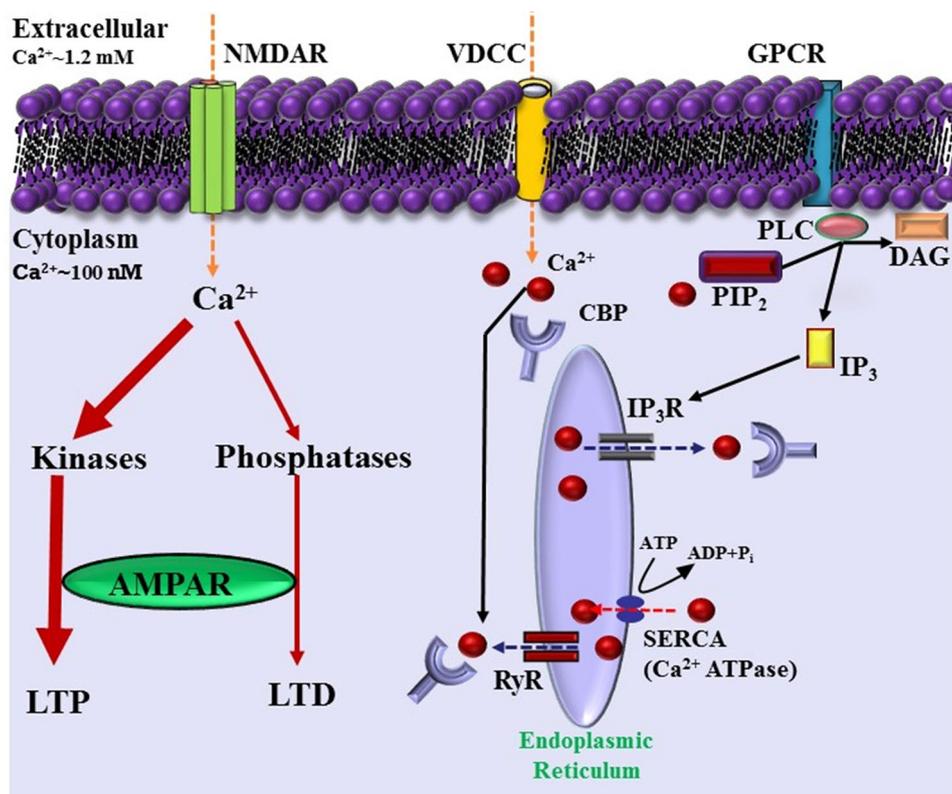


Fig. 1 Model illustrating various Ca^{2+} sources including NMDA receptor (NMDAR), voltage-dependent Ca^{2+} channels (VDCC), and G protein-coupled receptor (GPCR). Ca^{2+} (red balls) influxes into the cytosol (yellow dashed arrows) through these sources in a healthy neuron. The release of Ca^{2+} into the cytoplasm also occurs from the intracellular Ca^{2+} stores through inositol (1,4,5)-trisphosphate receptor (IP_3R) and ryanodine receptors (RyR) involving phospholipase C

(PLC), diacylglycerol (DAG) and inositol (1,4,5)-trisphosphate (IP_3). Organelles, including the endoplasmic reticulum, act as a Ca^{2+} buffering system, releasing and sequestering Ca^{2+} . Further, the model depicts Ca^{2+} buffering and extrusion pathways (red dashed arrows), involving plasma membrane Ca^{2+} ATPase, sarcoplasmic reticulum Ca^{2+} ATPases (SERCA), and various Ca^{2+} binding proteins (CBP). (Color figure online)

Expression of NMDA Receptor During Aging

Alteration in expression of specific NMDA receptor subunits might be a potential mechanism for the observed decrease in the NMDA receptor function [60]. GluN1 subunit of NMDA receptor is highly expressed in the hippocampus; results demonstrate that GluN1 subunit is susceptible to aging process. A significant decrease in the expression of GluN1 protein levels with advancing age is observed in the hippocampus [51, 61–65]. GluN1 mRNA expression of GluN1 subunit also declines with increasing age in the hippocampus [60, 66]. In contrast, other studies report modest or no age-related decrease in GluN1 protein expression in the whole hippocampus [67, 68]. These studies suggest that the GluN1 subunit of the NMDA receptor is variably susceptible to the influence of aging process.

The GluN2B subunit is highly expressed throughout the brain during early stages of development and declines at the onset of sexual maturity; GluN2A subunit-containing NMDA receptors increase across the same life span [69–74].

A shift in GluN2A and GluN2B expression in the hippocampus is thought to contribute to developmental changes in cognition and synaptic function [75]. GluN2A subunit of NMDA receptor is highly expressed in the hippocampus and other brain regions. Aging is associated with no change or a modest decrease in the expression of GluN2A mRNA expression in the hippocampus [60, 76, 77]. Similarly, there is some indication that expression of GluN2A protein is decreased in the hippocampus of aged animals when compared with middle aged animals [63], while other studies indicate no age-related change in the GluN2A subunit in the hippocampus [62, 78, 79].

Interestingly, GluN2B subunits of NMDA receptor display slower channel kinetics and greater Ca^{2+} conductance. These channels, by taking longer duration to close, allow more Ca^{2+} influx into the cell over a longer period, and are therefore thought to be more conducive to the induction of activity dependent synaptic plasticity. Additionally, upregulation of GluN2B significantly augments LTP and memory function in rodents [80, 81], including aged mice [82].

Finally, studies that involve viral vector-mediated upregulation of GluN2B expression in adult hippocampus suggest that increasing the level of GluN2B expression can improve cognitive function during aging [54, 79].

In contrast to GluN2A, expression of GluN2B protein [62–64, 67, 68, 78, 83, 84] and GluN2B mRNA [60, 76–78, 83, 85] is generally reported to decline in the hippocampus with advanced age. One problem is that few studies have examined the expression of both subunits in the same animal. For studies that examine the protein expression of both subunits, some studies suggest that decreased expression, mainly of GluN2B, increases the ratio of GluN2A/GluN2B protein in several brain regions [62, 78, 86]. However, other reports indicate that both subunits decline equally with advanced age [63, 67].

Modification of Existing Receptors

In addition, to the required binding of glutamate and postsynaptic depolarization, NMDA receptors are regulated by posttranslational modification of the receptor. In adults, the NMDA receptor synaptic responses can undergo LTP (NMDAR–LTP) and LTD (NMDAR–LTD) [87]. NMDAR–LTP depends on the activity of NMDARs, a subsequent increase in intracellular Ca^{2+} , and kinase activity [88–90]. Activation of kinases, such as tyrosine kinase [91, 92], protein kinase C (PKC) [93, 94], protein kinase A [95], and CAMKII increases NMDA receptor mediated currents. Interestingly, aging is associated with a shift in the balance of kinase/phosphatase activity, favoring an increase in the phosphatase activity [96–98].

With advancing age, the cellular localization, basal or stimulation induced kinase activity may be altered [53, 99]. In contrast to kinases, the activity of protein phosphatase appears to be augmented with normal aging, influencing synaptic function [96, 97, 100]. Protein phosphatases, including calcineurin and protein phosphatase 1, decrease NMDA receptor currents [92, 95, 101]. Phosphorylation state of GluN1, GluN2A or GluN2B subunits can rapidly regulate surface expression and localization of the NMDA receptors [102–105]. Inhibition of phosphatase activity increases the AMPA receptor component of synaptic transmission, specifically in aged animals [97]. The increase in synaptic transmission due to phosphatase inhibition may be linked to increased susceptibility to induction of LTD and thus, represent reversal of LTP. Inhibition of phosphatase activity also increases NMDA receptor-mediated synaptic transmission in aged animals; however, the increase is relatively small relative to the decrease in the NMDA synaptic response associated with aging [53].

Recent work suggests that redox regulation of NMDA receptor function contributes to schizophrenia [106], stressor-induced depressive-like behavior [107], and

synaptic plasticity during development [108]. In young animals, NMDA receptor function is modulated by redox state, such that under oxidizing conditions, disulfide bonds can form between cysteine residues in the NMDA receptor subunits [109–111]. Three pairs of cysteine residues are located within the N-terminal regulatory domain of the receptor (two pairs reside in GluN1 and one pair resides in GluN2A subunit) [110, 112, 113]. The formation of disulfide bonds between cysteine residues is thought to decrease the current through the NMDA receptor. In contrast to younger animals, little or no effect of oxidizing agents was observed for older animals, suggesting that cells were already in an oxidized state. In contrast, reducing conditions enhanced NMDA receptor mediated synaptic responses in hippocampus of aged animals [53, 55, 56, 114–116]. Recent results provide evidence for a link between the redox-mediated decline in NMDA receptor function and the emergence of an age-related cognitive phenotype with impairment in the rapid acquisition and retention of novel spatial information [55, 56]. These results demonstrate that the age-related decrease in NMDA receptor-mediated synaptic responses at CA3–CA1 hippocampal synapses with advanced age is related to redox state such that the reducing agent, dithiothreitol (DTT) significantly enhanced the NMDA receptor component of the synaptic response to a greater extent in cognitively impaired animals relative to unimpaired animals [55] (Fig. 2).

The mechanism for redox regulation of NMDA receptor function is unclear. An age-related shift in the composition of NMDA receptor subunits could render the NMDA receptor more susceptible to redox regulation. As noted above, the GluN2A subunit has unique extracellular cysteine residues, and reducing the extracellular disulfide bonds, by the addition of extracellular glutathione, increases the NMDA receptor response of diheteromeric GluN1–GluN2A receptors [117]. However, in the case of aging, addition of extracellular glutathione and other antioxidants that do not readily pass the cell membrane do not increase the NMDA receptor response in aged animals [53, 118]. Rather, intracellular application of glutathione increased the NMDA receptor response demonstrating an important role for intracellular redox state [53]. Furthermore, the DTT-mediated growth of the NMDA receptor response depends on the activity of Ca^{2+} /calmodulin-dependent protein kinase II (CaMKII) [53]. Interestingly, NMDAR–LTP is sensitive to redox conditions, [119, 120] and is impaired in aged animals [79], suggesting that redox mechanisms may underlie an age-related decrease in NMDAR–LTP.

Modification of NMDA Receptor Function by Agonist

In addition to binding glutamate, glycine acts as a co-agonist, binding to the GluN1 subunits. D-Serine might represent

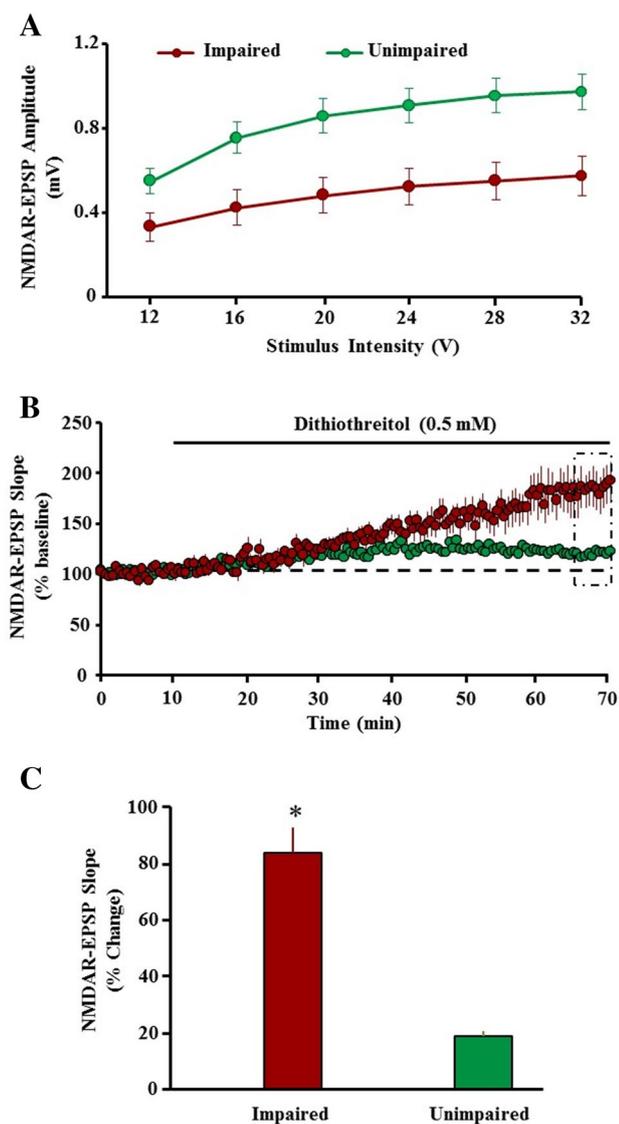


Fig. 2 Redox environment contributes to the decline in NMDA receptor function associated with cognitive impairment. **a** Input–output curves for the mean NMDA receptor EPSP (NMDAR–EPSP) amplitude evoked by increasing stimulating voltage. **b** Time course of changes in the slope of NMDA receptor-mediated EPSP obtained from hippocampal slices 10 min before and 60 min after bath application of the reducing agent DTT (0.5 mM, solid line) in unimpaired (red circles) and impaired (green circles) animals. **c** Bars represent the mean \pm SEM change in NMDAR–EPSP slope after the application of DTT in unimpaired (red circles) and impaired (green circles) animals. The asterisk indicates a significant increase in NMDAR–EPSP slope in impaired animals when compared with unimpaired. Reproduced with permission from Kumar and Foster [55]. (Color figure online)

another physiological co-agonist of the NMDA receptor as it can bind at the glycine-binding site [121–125]. D-Serine acts as a neuronal signaling molecule leading to upregulation of NMDA receptors. In addition, D-serine is highly expressed in the brain and released from astrocytes [122, 126–129].

D-Serine is required for NMDA receptor activation, and may have preferential affinity/effectiveness for NMDA receptors that contain GluN2B subunits [123]. The levels of D-serine are dramatically reduced with advanced age [130–132]. One possibility is a loss of the serine racemase enzyme, which generates D-serine from L-serine [133, 134]. A decline in serine racemase mRNA is observed in the hippocampus of aged rats [132]. The enzyme serine racemase generates D-serine from L-serine; pharmacological or viral gene delivery tools could be employed to increase endogenous levels of D-serine or serine racemase expression. Future studies to upregulate the expression of serine racemase, in order to enhance the endogenous level of D-serine, could provide another avenue to restore impaired NMDA receptor function during aging and under pathological conditions.

Influence of VDCCs and ICS on NMDA Receptor Function

In addition to binding of the transmitter glutamate, NMDA receptor activation requires postsynaptic depolarization to relieve the Mg^{2+} block of the channel. For CA1 neurons from aged animals, depolarization induced by a burst of afferent activity is reduced, due to activation of Ca^{2+} -dependent K^+ channels. The augmentation in the after hyperpolarization (AHP) amplitude diminishes the activation of NMDA channels, further contributing to impaired synaptic plasticity [18, 19]. The enhanced AHP with age is linked to increased involvement of VDCCs and internal Ca^{2+} stores [114, 135–149]. Thus, changes in Ca^{2+} from VDCCs or ICS can act through the AHP to impair NMDA receptor function.

VDCCs are ion channels in the plasma membrane open in response to membrane depolarization and allow Ca^{2+} influx into the cell from the extracellular space. In hippocampal CA1 pyramidal neurons of the rat, the L-type Ca^{2+} currents are increased [150, 151] and an increase in the density of functional L-type VDCCs have been reported for aged animals [152]. The idea that L-channels are increased in the hippocampus during senescence is also supported by mRNA and protein expression studies indicating an increase in $Ca_v1.3$ [153–155]. Treatments to reduce the AHP permits increase activation of NMDA receptor, to shift the threshold for induction of synaptic plasticity [156, 157]. In aged rats, under L-channel blockade, the induction of LTP is facilitated for low level synaptic activation, which would not induce synaptic modification in young animals [156]. It should be noted that L-channel blockade does not completely ameliorate age-related differences. The AHP amplitude is reduced but not to the levels observed in young animals [114, 148].

In addition to Ca^{2+} influx from outside the cell, ICS play a major role in regulating larger Ca^{2+} signals [158, 159]. Organelles, including the endoplasmic reticulum, mitochondria, and lysosomes act as Ca^{2+} buffering systems—releasing and

sequestering Ca^{2+} [160–166]. Thus, there are at least two possible mechanisms by which ICS can regulate Ca^{2+} homeostasis: (1) release of stored Ca^{2+} to enhance Ca^{2+} signals and (2) removing cytosolic Ca^{2+} following a large influx.

Two pathways control the release of Ca^{2+} from the endoplasmic reticulum, Ca^{2+} -induced Ca^{2+} release (CICR) and the inositol (1,4,5)-trisphosphate (IP_3) pathway activated by G protein-coupled receptors. G protein-coupled receptors activate phospholipase C to form diacylglycerol and IP_3 , which act on IP_3 receptors (IP_3Rs) to release Ca^{2+} from ICS. Previous studies have observed an age associated decrease in IP_3Rs in several brain regions [167–170]. Despite a general decrease in the receptor, the literature suggests that a decrease in IP_3 induced Ca^{2+} release is either limited to cortical cells [167] or no age-related change is observed [171]. The disconnect between a reduction in IP_3R expression and the apparent absence of an effect of age on IP_3 -induced Ca^{2+} release may be due to increased oxidation of the IP_3Rs which has been demonstrated to increase IP_3R function in brain cells [172, 173]. As such, reduced expression may act as compensation for an altered redox state, in order to maintain proper IP_3 signaling.

CICR is a Ca^{2+} amplification process that is initiated by influx of Ca^{2+} through membrane channels (i.e. VDCCs) (Fig. 1). The intracellular Ca^{2+} binds ryanodine receptors (RyRs) to release additional Ca^{2+} into the cytosol from the endoplasmic reticulum. Accumulating evidence supports a role of altered CICR in contributing to altered physiology of normal aging. The increased involvement of RyRs does not appear to be due to increased RyR expression [169]. Rather, an age-related increase in oxidative stress and a shift in the intracellular redox state may enhance the responsiveness of RyRs to intracellular Ca^{2+} [114, 174–176]. The redox state influences the formation of cysteine disulfide bonds. The disulfide bonds of RyR for ICS determine Ca^{2+} release and the amplitude of the AHP. In the case of NMDA receptors, redox sensitive disulfide bonds are localized to NMDA receptor subunits and to molecules such as Ca^{2+} /calmodulin-dependent protein kinase II that modify NMDA receptor function. The age-dependent specificity of oxidizing agents and DTT provide strong support for the tenet that redox stress mediates senescent physiology. Increased CICR appears to contribute to the larger AHP during aging [5, 140, 149]. As noted above, hippocampal cells exhibit increase Ca^{2+} from L-type Ca^{2+} channels, which could provide a source of Ca^{2+} to fill ICS and activate CICR from ICS.

The increase in CICR activates Ca^{2+} -dependent potassium channels in the membrane, inducing larger AHP. Attenuating CICR, by blocking RyR or depletion of Ca^{2+} from ICS, has a greater influence in reducing the amplitude of the AHP in aged animals [114, 140], indicating an aging-specific mechanism. Moreover, attenuation of CICR promotes induction of LTP and inhibits LTD during senescence [5, 140].

Thus, the relative shift in Ca^{2+} sources, with reduced extracellular influx of Ca^{2+} from NMDA receptors and increased release of Ca^{2+} from ICS, underlies senescent physiology, which is characterized by enhanced amplitude of AHP, a decrease in NMDA voltage-gated channel activity, decreased synaptic transmission, and altered synaptic plasticity.

Conclusion

Ideas about the role of senescent glutamatergic synapses in contributing to the age-related cognitive impairment are based on studies delineating the mechanisms for age-associated changes in Ca^{2+} -dependent synaptic plasticity. Specifically, aging associated alterations in Ca^{2+} regulation modify NMDA glutamate receptor mediated synaptic transmission including impaired LTP and enhanced LTD. In addition to altered synaptic plasticity, alterations in NMDA receptor subunit expression profile, molecular and biochemical modulatory mechanisms, and alterations in Ca^{2+} sources provide impetus for altering the NMDA receptor-mediated synaptic transmission. These senescent glutamatergic synaptic plasticity mechanisms contribute to cognitive aging. Due to the critical importance of NMDA glutamate receptors in synaptic transmission and cognitive function, a selective upregulation of NMDA receptor function may provide an avenue for treating age-associated cognitive deficits. Clearly, future research will need to delineate the contributions of several mechanisms in optimizing specific subunit contribution and influence of upregulation in mediating cognition. Thus, it will be imperative for future research to determine whether enhancing or inhibiting NMDA receptor function by upregulating or downregulating different subunits expression configurations will be beneficial in preserving cognitive domain and promoting successful cognitive aging.

Acknowledgements Supported by National Institute of Aging Grants R37AG036800, RO1049711, RO1037984, and RO1052258 and the Evelyn F. McKnight Brain Research Foundation. This work was partially supported by the University of Florida Claude D. Pepper Older Americans Independence Center (P30-AG028740).

References

1. Nicoll RA, Malenka RC (1999) Expression mechanisms underlying NMDA receptor-dependent long-term potentiation. *Ann N Y Acad Sci* 868:515–525
2. Barnes CA, McNaughton BL (1985) An age comparison of the rates of acquisition and forgetting of spatial information in relation to long-term enhancement of hippocampal synapses. *Behav Neurosci* 99:1040–1048
3. Lee HK, Kameyama K, Haganir RL, Bear MF (1998) NMDA induces long-term synaptic depression and dephosphorylation of the GluR1 subunit of AMPA receptors in hippocampus. *Neuron* 21:1151–1162

4. Kumar A (2010) Carbachol-induced long-term synaptic depression is enhanced during senescence at hippocampal CA3-CA1 synapses. *J Neurophysiol* 104:607–616
5. Kumar A, Foster TC (2005) Intracellular calcium stores contribute to increased susceptibility to LTD induction during aging. *Brain Res* 1031:125–128
6. Kumar A, Foster TC (2007) Shift in induction mechanisms underlies an age-dependent increase in DHPG-induced synaptic depression at CA3 CA1 synapses. *J Neurophysiol* 98:2729–2736
7. Norris CM, Korol DL, Foster TC (1996) Increased susceptibility to induction of long-term depression and long-term potentiation reversal during aging. *J Neurosci* 16:5382–5392
8. Foster TC (1999) Involvement of hippocampal synaptic plasticity in age-related memory decline. *Brain Res Rev* 30:236–249
9. Foster TC, Kumar A (2007) Susceptibility to induction of long-term depression is associated with impaired memory in aged Fischer 344 rats. *Neurobiol Learn Mem* 87:522–535
10. Febo M, Foster TC (2016) Preclinical magnetic resonance imaging and spectroscopy studies of memory, aging, and cognitive decline. *Front Aging Neurosci* 8:158
11. Zhou Q, Homma KJ, Poo MM (2004) Shrinkage of dendritic spines associated with long-term depression of hippocampal synapses. *Neuron* 44:749–757
12. Foster TC (2006) Biological markers of age-related memory deficits: treatment of senescent physiology. *CNS Drugs* 20:153–166
13. Shinoda Y, Kamikubo Y, Egashira Y, Tominaga-Yoshino K, Ogura A (2005) Repetition of mGluR-dependent LTD causes slowly developing persistent reduction in synaptic strength accompanied by synapse elimination. *Brain Res* 1042:99–107
14. Khachaturian ZS (1989) Calcium, membranes, aging, and Alzheimer's disease. Introduction and overview. *Ann N Y Acad Sci* 568:1–4
15. Michaelis ML, Johe K, Kito TE (1984) Age-dependent alterations in synaptic membrane systems for Ca^{2+} regulation. *Mech Ageing Dev* 25:215–225
16. Rapp PR, Gallagher M (1996) Preserved neuron number in the hippocampus of aged rats with spatial learning deficits. *Proc Natl Acad Sci USA* 93:9926–9930
17. West MJ (1993) Regionally specific loss of neurons in the aging human hippocampus. *Neurobiol Aging* 14:287–293
18. Foster TC (2012) Dissecting the age-related decline on spatial learning and memory tasks in rodent models: N-methyl-D-aspartate receptors and voltage-dependent Ca^{2+} channels in senescent synaptic plasticity. *Prog Neurobiol* 96:283–303
19. Foster TC, Norris CM (1997) Age-associated changes in Ca^{2+} -dependent processes: relation to hippocampal synaptic plasticity. *Hippocampus* 7:602–612
20. Berridge MJ, Lipp P, Bootman MD (2000) The versatility and universality of calcium signalling. *Nat Rev Mol Cell Biol* 1:11–21
21. Orrenius S, Zhivotovsky B, Nicotera P (2003) Regulation of cell death: the calcium-apoptosis link. *Nat Rev Mol Cell Biol* 4:552–565
22. Rizzuto R (2001) Intracellular Ca^{2+} pools in neuronal signalling. *Curr Opin Neurobiol* 11:306–311
23. Verkhratsky A, Toescu EC (1998) Calcium and neuronal ageing. *Trends Neurosci* 21:2–7
24. Brewer LD, Porter NM, Kerr DS, Landfield PW, Thibault O (2006) Chronic $1\alpha,25\text{-(OH)}_2$ vitamin D3 treatment reduces Ca^{2+} -mediated hippocampal biomarkers of aging. *Cell Calcium* 40:277–286
25. Kumar A, Bodhinathan K, Foster TC (2009) Susceptibility to calcium dysregulation during brain aging. *Front Aging Neurosci* 1:2
26. Cotman C, Monaghan D (1989) Multiple excitatory amino acid receptor regulation of intracellular Ca^{2+} . Implications for aging and Alzheimer's disease. *Ann N Y Acad Sci* 568:138–148
27. Cotman CW, Geddes JW, Bridges RJ, Monaghan DT (1989) N-methyl-D-aspartate receptors and Alzheimer's disease. *Neurobiol Aging* 10:603–605 (**discussion 618–620**)
28. Morris RG (2013) NMDA receptors and memory encoding. *Neuropharmacology* 74:32–40
29. Dingledine R, Borges K, Bowe D, Traynelis SF (1999) The glutamate receptor ion channels. *Pharmacol Rev* 51:7–61
30. Cull-Candy S, Brickley S, Farrant M (2001) NMDA receptor subunits: diversity, development and disease. *Curr Opin Neurobiol* 11:327–335
31. Kutsuwada T, Kashiwabuchi N, Mori H, Sakimura K, Kushiya E, Araki K, Meguro H, Masaki H, Kumanishi T, Arakawa M et al (1992) Molecular diversity of the NMDA receptor channel. *Nature* 358:36–41
32. Meguro H, Mori H, Araki K, Kushiya E, Kutsuwada T, Yamazaki M, Kumanishi T, Arakawa M, Sakimura K, Mishina M (1992) Functional characterization of a heteromeric NMDA receptor channel expressed from cloned cDNAs. *Nature* 357:70–74
33. Monyer H, Sprengel R, Schoepfer R, Herb A, Higuchi M, Lomeli H, Burnashev N, Sakmann B, Seeburg PH (1992) Heteromeric NMDA receptors: molecular and functional distinction of subtypes. *Science* 256:1217–1221
34. Moriyoshi K, Masu M, Ishii T, Shigemoto R, Mizuno N, Nakanishi S (1991) Molecular cloning and characterization of the rat NMDA receptor. *Nature* 354:31–37
35. Laube B, Kuhse J, Betz H (1998) Evidence for a tetrameric structure of recombinant NMDA receptors. *J Neurosci* 18:2954–2961
36. Kumar A (2015) NMDA receptor function during senescence: implication on cognitive performance. *Front Neurosci* 9:473
37. Al-Hallaq RA, Jarabek BR, Fu Z, Vicini S, Wolfe BB, Yasuda RP (2002) Association of NR3A with the N-methyl-D-aspartate receptor NR1 and NR2 subunits. *Mol Pharmacol* 62:1119–1127
38. Schuler T, Mesic I, Madry C, Bartholomaeus I, Laube B (2008) Formation of NR1/NR2 and NR1/NR3 heterodimers constitutes the initial step in N-methyl-D-aspartate receptor assembly. *J Biol Chem* 283:37–46
39. Sucher NJ, Akbarian S, Chi CL, Leclerc CL, Awobuluyi M, Deitcher DL, Wu MK, Yuan JP, Jones EG, Lipton SA (1995) Developmental and regional expression pattern of a novel NMDA receptor-like subunit (NMDAR-L) in the rodent brain. *J Neurosci* 15:6509–6520
40. Low CM, Wee KS (2010) New insights into the not-so-new NR3 subunits of N-methyl-D-aspartate receptor: localization, structure, and function. *Mol Pharmacol* 78:1–11
41. Collingridge G (1987) Synaptic plasticity. The role of NMDA receptors in learning and memory. *Nature* 330:604–605
42. Morris RG, Anderson E, Lynch GS, Baudry M (1986) Selective impairment of learning and blockade of long-term potentiation by an N-methyl-D-aspartate receptor antagonist, AP5. *Nature* 319:774–776
43. Morris RG (1989) Synaptic plasticity and learning: selective impairment of learning rats and blockade of long-term potentiation in vivo by the N-methyl-D-aspartate receptor antagonist AP5. *J Neurosci* 9:3040–3057
44. Lisman JE, Fellous JM, Wang XJ (1998) A role for NMDA-receptor channels in working memory. *Nat Neurosci* 1:273–275
45. Mondadori C, Weiskrantz L (1993) NMDA receptor blockers facilitate and impair learning via different mechanisms. *Behav Neural Biol* 60:205–210
46. Mondadori C, Weiskrantz L, Buerki H, Petschke F, Fagg GE (1989) NMDA receptor antagonists can enhance or impair learning performance in animals. *Exp Brain Res* 75:449–456

47. Martin SJ, Grimwood PD, Morris RG (2000) Synaptic plasticity and memory: an evaluation of the hypothesis. *Annu Rev Neurosci* 23:649–711
48. Dore K, Stein IS, Brock JA, Castillo PE, Zito K, Sjöström PJ (2017) Unconventional NMDA receptor signaling. *J Neurosci* 37:10800–10807
49. Foster TC, Kyritsopoulos C, Kumar A (2017) Central role for NMDA receptors in redox mediated impairment of synaptic function during aging and Alzheimer's disease. *Behav Brain Res* 322:223–232
50. Barnes CA, Rao G, Shen J (1997) Age-related decrease in the N-methyl-D-aspartate-mediated excitatory postsynaptic potential in hippocampal region CA1. *Neurobiol Aging* 18:445–452
51. Eckles-Smith K, Clayton D, Bickford P, Browning MD (2000) Caloric restriction prevents age-related deficits in LTP and in NMDA receptor expression. *Brain Res Mol Brain Res* 78:154–162
52. Billard JM, Rouaud E (2007) Deficit of NMDA receptor activation in CA1 hippocampal area of aged rats is rescued by D-cycloserine. *Eur J Neurosci* 25:2260–2268
53. Bodhinathan K, Kumar A, Foster TC (2010) Intracellular redox state alters NMDA receptor response during aging through Ca²⁺/calmodulin-dependent protein kinase II. *J Neurosci* 30:1914–1924
54. Brim BL, Haskell R, Awedikian R, Ellinwood NM, Jin L, Kumar A, Foster TC, Magnusson K (2013) Memory in aged mice is rescued by enhanced expression of the GluN2B subunit of the NMDA receptor. *Behav Brain Res* 238:211–226
55. Kumar A, Foster TC (2013) Linking redox regulation of NMDAR synaptic function to cognitive decline during aging. *J Neurosci* 33:15710–15715
56. Lee WH, Kumar A, Rani A, Foster TC (2014) Role of antioxidant enzymes in redox regulation of N-methyl-D-aspartate receptor function and memory in middle-aged rats. *Neurobiol Aging* 35:1459–1468
57. Kumar A, Rani A, Scheinert RB, Ormerod BK, Foster TC (2018) Nonsteroidal anti-inflammatory drug, indomethacin improves spatial memory and NMDA receptor function in aged animals. *Neurobiol Aging* 70:184–193
58. Zorumski CF, Izumi Y (2012) NMDA receptors and metaplasticity: mechanisms and possible roles in neuropsychiatric disorders. *Neurosci Biobehav Rev* 36:989–1000
59. Abraham WC (2008) Metaplasticity: tuning synapses and networks for plasticity. *Nat Rev Neurosci* 9:387
60. Magnusson KR (2000) Declines in mRNA expression of different subunits may account for differential effects of aging on agonist and antagonist binding to the NMDA receptor. *J Neurosci* 20:1666–1674
61. Liu F, Day M, Muniz LC, Bitran D, Arias R, Revilla-Sanchez R, Grauer S, Zhang G, Kelley C, Pulito V, Sung A, Mervis RF, Navarra R, Hirst WD, Reinhart PH, Marquis KL, Moss SJ, Pangalos MN, Brandon NJ (2008) Activation of estrogen receptor-beta regulates hippocampal synaptic plasticity and improves memory. *Nat Neurosci* 11:334–343
62. Mesches MH, Gemma C, Veng LM, Allgeier C, Young DA, Browning MD, Bickford PC (2004) Sulindac improves memory and increases NMDA receptor subunits in aged Fischer 344 rats. *Neurobiol Aging* 25:315–324
63. Magnusson KR, Nelson SE, Young AB (2002) Age-related changes in the protein expression of subunits of the NMDA receptor. *Brain Res Mol Brain Res* 99:40–45
64. Coultrap SJ, Bickford PC, Browning MD (2008) Blueberry-enriched diet ameliorates age-related declines in NMDA receptor-dependent LTP. *Age* 30:263–272
65. Newton IG, Forbes ME, Linville MC, Pang H, Tucker EW, Riddle DR, Brunso-Bechtold JK (2008) Effects of aging and caloric restriction on dentate gyrus synapses and glutamate receptor subunits. *Neurobiol Aging* 29:1308–1318
66. Adams MM, Morrison JH, Gore AC (2001) N-methyl-D-aspartate receptor mRNA levels change during reproductive senescence in the hippocampus of female rats. *Exp Neurol* 170:171–179
67. Sonntag WE, Bennett SA, Khan AS, Thornton PL, Xu X, Ingram RL, Brunso-Bechtold JK (2000) Age and insulin-like growth factor-1 modulate N-methyl-D-aspartate receptor subtype expression in rats. *Brain Res Bull* 51:331–338
68. Zhao X, Rosenke R, Kronemann D, Brim B, Das SR, Dunah AW, Magnusson KR (2009) The effects of aging on N-methyl-D-aspartate receptor subunits in the synaptic membrane and relationships to long-term spatial memory. *Neuroscience* 162:933–945
69. Liu XB, Murray KD, Jones EG (2004) Switching of NMDA receptor 2A and 2B subunits at thalamic and cortical synapses during early postnatal development. *J Neurosci* 24:8885–8895
70. Monyer H, Burnashev N, Laurie DJ, Sakmann B, Seeburg PH (1994) Developmental and regional expression in the rat brain and functional properties of four NMDA receptors. *Neuron* 12:529–540
71. Laurie DJ, Bartke I, Schoepfer R, Naujoks K, Seeburg PH (1997) Regional, developmental and interspecies expression of the four NMDAR2 subunits, examined using monoclonal antibodies. *Brain Res Mol Brain Res* 51:23–32
72. Laurie DJ, Seeburg PH (1994) Regional and developmental heterogeneity in splicing of the rat brain NMDAR1 mRNA. *J Neurosci* 14:3180–3194
73. Law AJ, Weickert CS, Webster MJ, Herman MM, Kleinman JE, Harrison PJ (2003) Changes in NMDA receptor subunit mRNAs and cyclophilin mRNA during development of the human hippocampus. *Ann N Y Acad Sci* 1003:426–430
74. Law AJ, Weickert CS, Webster MJ, Herman MM, Kleinman JE, Harrison PJ (2003) Expression of NMDA receptor NR1, NR2A and NR2B subunit mRNAs during development of the human hippocampal formation. *Eur J Neurosci* 18:1197–1205
75. Dumas TC (2005) Developmental regulation of cognitive abilities: modified composition of a molecular switch turns on associative learning. *Prog Neurobiol* 76:189–211
76. Magnusson KR (2001) Influence of diet restriction on NMDA receptor subunits and learning during aging. *Neurobiol Aging* 22:613–627
77. Magnusson KR, Kresge D, Supon J (2006) Differential effects of aging on NMDA receptors in the intermediate versus the dorsal hippocampus. *Neurobiol Aging* 27:324–333
78. Clayton DA, Browning MD (2001) Deficits in the expression of the NR2B subunit in the hippocampus of aged Fisher 344 rats. *Neurobiol Aging* 22:165–168
79. Clayton DA, Mesches MH, Alvarez E, Bickford PC, Browning MD (2002) A hippocampal NR2B deficit can mimic age-related changes in long-term potentiation and spatial learning in the Fischer 344 rat. *J Neurosci* 22:3628–3637
80. Cui Y, Jin J, Zhang X, Xu H, Yang L, Du D, Zeng Q, Tsien JZ, Yu H, Cao X (2011) Forebrain NR2B overexpression facilitating the prefrontal cortex long-term potentiation and enhancing working memory function in mice. *PLoS ONE* 6:e20312
81. Tang YP, Shimizu E, Dube GR, Rampon C, Kerchner GA, Zhuo M, Liu G, Tsien JZ (1999) Genetic enhancement of learning and memory in mice. *Nature* 401:63–69
82. Cao X, Cui Z, Feng R, Tang YP, Qin Z, Mei B, Tsien JZ (2007) Maintenance of superior learning and memory function in NR2B transgenic mice during ageing. *Eur J Neurosci* 25:1815–1822
83. Dyllal SC, Michael GJ, Whelpton R, Scott AG, Michael-Titus AT (2007) Dietary enrichment with omega-3 polyunsaturated fatty acids reverses age-related decreases in the GluR2 and NR2B

- glutamate receptor subunits in rat forebrain. *Neurobiol Aging* 28:424–439
84. Liu P, Smith PF, Darlington CL (2008) Glutamate receptor subunits expression in memory-associated brain structures: regional variations and effects of aging. *Synapse* 62:834–841
 85. Marquez Loza A, Elias V, Wong CP, Ho E, Bermudez M, Magnusson KR (2017) Effects of ibuprofen on cognition and NMDA receptor subunit expression across aging. *Neuroscience* 344:276–292
 86. Magnusson KR, Scruggs B, Zhao X, Hammersmark R (2007) Age-related declines in a two-day reference memory task are associated with changes in NMDA receptor subunits in mice. *BMC Neurosci* 8:43
 87. Hunt DL, Castillo PE (2012) Synaptic plasticity of NMDA receptors: mechanisms and functional implications. *Curr Opin Neurobiol* 22:496–508
 88. Aniksztejn L, Ben-Ari Y (1995) Expression of LTP by AMPA and/or NMDA receptors is determined by the extent of NMDA receptors activation during the tetanus. *J Neurophysiol* 74:2349–2357
 89. Li HB, Jackson MF, Yang K, Trepanier C, Salter MW, Orser BA, Macdonald JF (2011) Plasticity of synaptic GluN receptors is required for the Src-dependent induction of long-term potentiation at CA3-CA1 synapses. *Hippocampus* 21:1053–1061
 90. Peng Y, Zhao J, Gu QH, Chen RQ, Xu Z, Yan JZ, Wang SH, Liu SY, Chen Z, Lu W (2010) Distinct trafficking and expression mechanisms underlie LTP and LTD of NMDA receptor-mediated synaptic responses. *Hippocampus* 20:646–658
 91. Heidinger V, Manzerra P, Wang XQ, Strasser U, Yu SP, Choi DW, Behrens MM (2002) Metabotropic glutamate receptor 1-induced upregulation of NMDA receptor current: mediation through the Pyk2/Src-family kinase pathway in cortical neurons. *J Neurosci* 22:5452–5461
 92. Wang LY, Orser BA, Brautigam DL, MacDonald JF (1994) Regulation of NMDA receptors in cultured hippocampal neurons by protein phosphatases 1 and 2A. *Nature* 369:230–232
 93. Ben-Ari Y, Aniksztejn L, Bregestovski P (1992) Protein kinase C modulation of NMDA currents: an important link for LTP induction. *Trends Neurosci* 15:333–339
 94. Chen L, Huang LY (1992) Protein kinase C reduces Mg^{2+} block of NMDA-receptor channels as a mechanism of modulation. *Nature* 356:521–523
 95. Raman IM, Tong G, Jahr CE (1996) Beta-adrenergic regulation of synaptic NMDA receptors by cAMP-dependent protein kinase. *Neuron* 16:415–421
 96. Foster TC, Sharrow KM, Masse JR, Norris CM, Kumar A (2001) Calcineurin links Ca^{2+} dysregulation with brain aging. *J Neurosci* 21:4066–4073
 97. Norris CM, Halpain S, Foster TC (1998) Alterations in the balance of protein kinase/phosphatase activities parallel reduced synaptic strength during aging. *J Neurophysiol* 80:1567–1570
 98. Foster TC (2007) Calcium homeostasis and modulation of synaptic plasticity in the aged brain. *Aging Cell* 6:319–325
 99. Foster TC (2004) Age-related changes in synaptic phosphorylation and dephosphorylation. In: Mattson M (ed) *Protein Phosphorylation in Aging and Age-Related Disease*. Elsevier, Amsterdam, pp 133–152
 100. Mulkey RM, Endo S, Shenolikar S, Malenka RC (1994) Involvement of a calcineurin/inhibitor-1 phosphatase cascade in hippocampal long-term depression. *Nature* 369:486–488
 101. Lieberman DN, Mody I (1994) Regulation of NMDA channel function by endogenous $Ca(2+)$ -dependent phosphatase. *Nature* 369:235–239
 102. Chung HJ, Huang YH, Lau LF, Huganir RL (2004) Regulation of the NMDA receptor complex and trafficking by activity-dependent phosphorylation of the NR2B subunit PDZ ligand. *J Neurosci* 24:10248–10259
 103. Gardoni F, Schrama LH, Kamal A, Gispén WH, Cattabeni F, Di Luca M (2001) Hippocampal synaptic plasticity involves competition between Ca^{2+} /calmodulin-dependent protein kinase II and postsynaptic density 95 for binding to the NR2A subunit of the NMDA receptor. *J Neurosci* 21:1501–1509
 104. Hallett PJ, Spoelgen R, Hyman BT, Standaert DG, Dunah AW (2006) Dopamine D1 activation potentiates striatal NMDA receptors by tyrosine phosphorylation-dependent subunit trafficking. *J Neurosci* 26:4690–4700
 105. Lin Y, Jover-Mengual T, Wong J, Bennett MV, Zukin RS (2006) PSD-95 and PKC converge in regulating NMDA receptor trafficking and gating. *Proc Natl Acad Sci USA* 103:19902–19907
 106. Steullet P, Cabungcal JH, Monin A, Dwir D, O'Donnell P, Cuenod M, Do KQ (2014) Redox dysregulation, neuroinflammation, and NMDA receptor hypofunction: A “central hub” in schizophrenia pathophysiology? *Schizophr Res*. <https://doi.org/10.1016/j.schres.2014.06.021>
 107. Ibi M, Liu J, Arakawa N, Kitaoka S, Kawaji, Matsuda KI, Iwata K, Matsumoto M, Katsuyama, M, Zhu K, Teramukai S, Furuyashiki T, Yabe-Nishimura C (2017) Depressive-like behaviors are regulated by NOX1/NADPH oxidase by redox modification of NMDA receptor 1. *J Neurosci* 37:4200–4212. <https://doi.org/10.1523/JNEUROSCI.2988-16.2017>
 108. Francis-Oliveira J, Vilar Higa GS, Mendonca Munhoz Dati L, Carvalho Shieh I, De Pasquale R (2018) Metaplasticity in the visual cortex: crosstalk between visual experience and reactive oxygen species. *J Neurosci* 38:5649–5665
 109. Aizenman E, Hartnett KA, Reynolds IJ (1990) Oxygen free radicals regulate NMDA receptor function via a redox modulatory site. *Neuron* 5:841–846
 110. Choi Y, Chen HV, Lipton SA (2001) Three pairs of cysteine residues mediate both redox and Zn^{2+} modulation of the nmda receptor. *J Neurosci* 21:392–400
 111. Sullivan JM, Traynelis SF, Chen HS, Escobar W, Heinemann SF, Lipton SA (1994) Identification of two cysteine residues that are required for redox modulation of the NMDA subtype of glutamate receptor. *Neuron* 13:929–936
 112. Choi YB, Lipton SA (2000) Redox modulation of the NMDA receptor. *Cell Mol Life Sci* 57:1535–1541
 113. Lipton SA, Choi YB, Takahashi H, Zhang D, Li W, Godzik A, Bankston LA (2002) Cysteine regulation of protein function—as exemplified by NMDA-receptor modulation. *Trends Neurosci* 25:474–480
 114. Bodhinathan K, Kumar A, Foster TC (2010) Redox sensitive calcium stores underlie enhanced after hyperpolarization of aged neurons: role for ryanodine receptor mediated calcium signaling. *J Neurophysiol* 104:2586–2593
 115. Haxaire C, Turpin FR, Potier B, Kervern M, Sinet PM, Barbanel G, Mothet JP, Dutar P, Billard JM (2012) Reversal of age-related oxidative stress prevents hippocampal synaptic plasticity deficits by protecting d-serine-dependent NMDA receptor activation. *Aging Cell* 11:336–344
 116. Robillard JM, Gordon GR, Choi HB, Christie BR, MacVicar BA (2011) Glutathione restores the mechanism of synaptic plasticity in aged mice to that of the adult. *PLoS ONE* 6:e20676
 117. Kohr G, Eckardt S, Luddens H, Monyer H, Seeburg PH (1994) NMDA receptor channels: subunit-specific potentiation by reducing agents. *Neuron* 12:1031–1040
 118. Yang YJ, Wu PF, Long LH, Yu DF, Wu WN, Hu ZL, Fu H, Xie N, Jin Y, Ni L, Wang JZ, Wang F, Chen JG (2010) Reversal of aging-associated hippocampal synaptic plasticity deficits by reductants via regulation of thiol redox and NMDA receptor function. *Aging Cell* 9:709–721

119. Gozlan H, Chinesra P, Diabira D, Ben-Ari Y (1994) NMDA redox site modulates long-term potentiation of NMDA but not of AMPA receptors. *Eur J Pharmacol* 262:R3–R4
120. Bernard CL, Hirsch JC, Khazipov R, Ben-Ari Y, Gozlan H (1997) Redox modulation of synaptic responses and plasticity in rat CA1 hippocampal neurons. *Exp Brain Res* 113:343–352
121. Hood WF, Compton RP, Monahan JB (1989) D-cycloserine: a ligand for the N-methyl-D-aspartate coupled glycine receptor has partial agonist characteristics. *Neurosci Lett* 98:91–95
122. Mothet JP, Parent AT, Wolosker H, Brady RO Jr, Linden DJ, Ferris CD, Rogawski MA, Snyder SH (2000) D-serine is an endogenous ligand for the glycine site of the N-methyl-D-aspartate receptor. *Proc Natl Acad Sci USA* 97:4926–4931
123. Priestley T, Laughton P, Myers J, Le Bourdelles B, Kerby J, Whiting PJ (1995) Pharmacological properties of recombinant human N-methyl-D-aspartate receptors comprising NR1a/NR2A and NR1a/NR2B subunit assemblies expressed in permanently transfected mouse fibroblast cells. *Mol Pharmacol* 48:841–848
124. Labrie V, Roder JC (2010) The involvement of the NMDA receptor D-serine/glycine site in the pathophysiology and treatment of schizophrenia. *Neurosci Biobehav Rev* 34:351–372
125. Panatier A, Theodosis DT, Mothet JP, Touquet B, Pollegioni L, Poulain DA, Oliet SH (2006) Glia-derived D-serine controls NMDA receptor activity and synaptic memory. *Cell* 125:775–784
126. Nishikawa T (2005) Metabolism and functional roles of endogenous D-serine in mammalian brains. *Biol Pharm Bull* 28:1561–1565
127. Schell MJ, Molliver ME, Snyder SH (1995) D-serine, an endogenous synaptic modulator: localization to astrocytes and glutamate-stimulated release. *Proc Natl Acad Sci USA* 92:3948–3952
128. De Miranda J, Panizzutti R, Foltyn VN, Wolosker H (2002) Cofactors of serine racemase that physiologically stimulate the synthesis of the N-methyl-D-aspartate (NMDA) receptor co-agonist D-serine. *Proc Natl Acad Sci USA* 99:14542–14547
129. Fossat P, Turpin FR, Sacchi S, Dulong J, Shi T, Rivet JM, Sweedler JV, Pollegioni L, Millan MJ, Oliet SH, Mothet JP (2012) Glial D-serine gates NMDA receptors at excitatory synapses in prefrontal cortex. *Cereb Cortex* 22:595–606
130. Junjaud G, Rouaud E, Turpin F, Mothet JP, Billard JM (2006) Age-related effects of the neuromodulator D-serine on neurotransmission and synaptic potentiation in the CA1 hippocampal area of the rat. *J Neurochem* 98:1159–1166
131. Mothet JP, Rouaud E, Sinet PM, Potier B, Jouvenceau A, Dutar P, Videau C, Epelbaum J, Billard JM (2006) A critical role for the glial-derived neuromodulator D-serine in the age-related deficits of cellular mechanisms of learning and memory. *Aging Cell* 5:267–274
132. Turpin FR, Potier B, Dulong JR, Sinet PM, Alliot J, Oliet SH, Dutar P, Epelbaum J, Mothet JP, Billard JM (2011) Reduced serine racemase expression contributes to age-related deficits in hippocampal cognitive function. *Neurobiol Aging* 32:1495–1504
133. Wolosker H, Blackshaw S, Snyder SH (1999) Serine racemase: a glial enzyme synthesizing D-serine to regulate glutamate-N-methyl-D-aspartate neurotransmission. *Proc Natl Acad Sci USA* 96:13409–13414
134. Wolosker H, Sheth KN, Takahashi M, Mothet JP, Brady RO Jr, Ferris CD, Snyder SH (1999) Purification of serine racemase: biosynthesis of the neuromodulator D-serine. *Proc Natl Acad Sci USA* 96:721–725
135. Landfield PW, Pitler TA (1984) Prolonged Ca^{2+} -dependent afterhyperpolarizations in hippocampal neurons of aged rats. *Science* 226:1089–1092
136. Kerr DS, Campbell LW, Hao SY, Landfield PW (1989) Corticosteroid modulation of hippocampal potentials: increased effect with aging. *Science* 245:1505–1509
137. Pitler TA, Landfield PW (1990) Aging-related prolongation of calcium spike duration in rat hippocampal slice neurons. *Brain Res* 508:1–6
138. Gong LW, Gao TM, Huang H, Zhou KX, Tong Z (2002) ATP modulation of large conductance $Ca(2+)$ -activated $K(+)$ channels via a functionally associated protein kinase A in CA1 pyramidal neurons from rat hippocampus. *Brain Res* 951:130–134
139. Hsu KS, Huang CC, Liang YC, Wu HM, Chen YL, Lo SW, Ho WC (2002) Alterations in the balance of protein kinase and phosphatase activities and age-related impairments of synaptic transmission and long-term potentiation. *Hippocampus* 12:787–802
140. Kumar A, Foster TC (2004) Enhanced long-term potentiation during aging is masked by processes involving intracellular calcium stores. *J Neurophysiol* 91:2437–2444
141. Tombaugh GC, Rowe WB, Rose GM (2005) The slow afterhyperpolarization in hippocampal CA1 neurons covaries with spatial learning ability in aged Fisher 344 rats. *J Neurosci* 25:2609–2616
142. Kumar A, Foster TC (2002) 17beta-Estradiol benzoate decreases the AHP amplitude in CA1 pyramidal neurons. *J Neurophysiol* 88:621–626
143. Disterhoft JF, Moyer JR Jr, Thompson LT, Kowalska M (1993) Functional aspects of calcium-channel modulation. *Clin Neuropharmacol* 16(Suppl 1):S12–S24
144. Disterhoft JF, Thompson LT, Moyer JR, Mogul DJ (1996) Calcium-dependent afterhyperpolarization and learning in young and aging hippocampus. *Life Sci* 59:413–420
145. Moyer JR Jr, Thompson LT, Black JP, Disterhoft JF (1992) Nimodipine increases excitability of rabbit CA1 pyramidal neurons in an age- and concentration-dependent manner. *J Neurophysiol* 68:2100–2109
146. Power JM, Oh MM, Disterhoft JF (2001) Metrifonate decreases sI(AHP) in CA1 pyramidal neurons in vitro. *J Neurophysiol* 85:319–322
147. Moyer JR Jr, Power JM, Thompson LT, Disterhoft JF (2000) Increased excitability of aged rabbit CA1 neurons after trace eyeblink conditioning. *J Neurosci* 20:5476–5482
148. Power JM, Wu WW, Sametsky E, Oh MM, Disterhoft JF (2002) Age-related enhancement of the slow outward calcium-activated potassium current in hippocampal CA1 pyramidal neurons in vitro. *J Neurosci* 22:7234–7243
149. Gant JC, Sama MM, Landfield PW, Thibault O (2006) Early and simultaneous emergence of multiple hippocampal biomarkers of aging is mediated by Ca^{2+} -induced Ca^{2+} release. *J Neurosci* 26:3482–3490
150. Campbell LW, Hao SY, Thibault O, Blalock EM, Landfield PW (1996) Aging changes in voltage-gated calcium currents in hippocampal CA1 neurons. *J Neurosci* 16:6286–6295
151. Brewer LD, Dowling AL, Curran-Rauhut MA, Landfield PW, Porter NM, Blalock EM (2009) Estradiol reverses a calcium-related biomarker of brain aging in female rats. *J Neurosci* 29:6058–6067
152. Thibault O, Landfield PW (1996) Increase in single L-type calcium channels in hippocampal neurons during aging. *Science* 272:1017–1020
153. Herman JP, Chen KC, Booze R, Landfield PW (1998) Up-regulation of alpha1D Ca^{2+} channel subunit mRNA expression in the hippocampus of aged F344 rats. *Neurobiol Aging* 19:581–587
154. Veng LM, Mesches MH, Browning MD (2003) Age-related working memory impairment is correlated with increases in the L-type calcium channel protein alpha1D (Cav1.3) in area CA1 of the hippocampus and both are ameliorated by chronic nimodipine treatment. *Brain Res Mol Brain Res* 110:193–202
155. Chen KC, Blalock EM, Thibault O, Kaminker P, Landfield PW (2000) Expression of alpha 1D subunit mRNA is correlated with

- L-type Ca^{2+} channel activity in single neurons of hippocampal “zipper” slices. *Proc Natl Acad Sci USA* 97:4357–4362
156. Norris CM, Halpain S, Foster TC (1998) Reversal of age-related alterations in synaptic plasticity by blockade of L-type Ca^{2+} channels. *J Neurosci* 18:3171–3179
157. Shankar S, Teyler TJ, Robbins N (1998) Aging differentially alters forms of long-term potentiation in rat hippocampal area CA1. *J Neurophysiol* 79:334–341
158. Ly CV, Verstreken P (2006) Mitochondria at the synapse. *Neuroscientist* 12:291–299
159. Mattson MP, LaFerla FM, Chan SL, Leissring MA, Shepel PN, Geiger JD (2000) Calcium signaling in the ER: its role in neuronal plasticity and neurodegenerative disorders. *Trends Neurosci* 23:222–229
160. Murchison D, Griffith WH (2007) Calcium buffering systems and calcium signaling in aged rat basal forebrain neurons. *Aging Cell* 6:297–305
161. Toescu EC, Verkhratsky A (2004) Ca^{2+} and mitochondria as substrates for deficits in synaptic plasticity in normal brain ageing. *J Cell Mol Med* 8:181–190
162. Duchen MR (2000) Mitochondria and calcium: from cell signaling to cell death. *J Physiol* 529(Pt 1):57–68
163. Nicholls DG, Budd SL (2000) Mitochondria and neuronal survival. *Physiol Rev* 80:315–360
164. Solovyova N, Veselovsky N, Toescu EC, Verkhratsky A (2002) Ca^{2+} dynamics in the lumen of the endoplasmic reticulum in sensory neurons: direct visualization of Ca^{2+} -induced Ca^{2+} release triggered by physiological Ca^{2+} entry. *EMBO J* 21:622–630
165. McGuinness L, Bardo SJ, Emptage NJ (2007) The lysosome or lysosome-related organelle may serve as a Ca^{2+} store in the boutons of hippocampal pyramidal cells. *Neuropharmacology* 52:126–135
166. Toescu EC, Myronova N, Verkhratsky A (2000) Age-related structural and functional changes of brain mitochondria. *Cell Calcium* 28:329–338
167. Burnett DM, Daniell LC, Zahniser NR (1990) Decreased efficacy of inositol 1,4,5-trisphosphate to elicit calcium mobilization from cerebrocortical microsomes of aged rats. *Mol Pharmacol* 37:566–571
168. Igwe OJ, Ning L (1993) Inositol 1,4,5-trisphosphate arm of the phosphatidylinositol signal transduction pathway in the rat cerebellum during aging. *Neurosci Lett* 164:167–170
169. Martini A, Battaini F, Govoni S, Volpe P (1994) Inositol 1,4,5-trisphosphate receptor and ryanodine receptor in the aging brain of Wistar rats. *Neurobiol Aging* 15:203–206
170. Simonyi A, Xia J, Igbavboa U, Wood WG, Sun GY (1998) Age differences in the expression of metabotropic glutamate receptor 1 and inositol 1,4,5-trisphosphate receptor in mouse cerebellum. *Neurosci Lett* 244:29–32
171. Stutzmann GE, Smith I, Caccamo A, Oddo S, Laferla FM, Parker I (2006) Enhanced ryanodine receptor recruitment contributes to Ca^{2+} disruptions in young, adult, and aged Alzheimer’s disease mice. *J Neurosci* 26:5180–5189
172. Long LH, Liu J, Liu RL, Wang F, Hu ZL, Xie N, Fu H, Chen JG (2009) Differential effects of methionine and cysteine oxidation on $[\text{Ca}^{2+}]_i$ in cultured hippocampal neurons. *Cell Mol Neurobiol* 29:7–15
173. Peuchen S, Duchen MR, Clark JB (1996) Energy metabolism of adult astrocytes in vitro. *Neuroscience* 71:855–870
174. Bull R, Finkelstein JP, Humeres A, Behrens MI, Hidalgo C (2007) Effects of ATP, Mg^{2+} , and redox agents on the Ca^{2+} dependence of RyR channels from rat brain cortex. *Am J Physiol Cell Physiol* 293:C162–C171
175. Gokulrangan G, Zaidi A, Michaelis ML, Schoneich C (2007) Proteomic analysis of protein nitration in rat cerebellum: effect of biological aging. *J Neurochem* 100:1494–1504
176. Hidalgo C, Bull R, Behrens MI, Donoso P (2004) Redox regulation of RyR-mediated Ca^{2+} release in muscle and neurons. *Biol Res* 37:539–552