



An Integrative Overview of Non-Amyloid and Non-Tau Pathologies in Alzheimer's Disease

Blaise W. Menta^{1,2,3} · Russell H. Swerdlow^{1,2,3,4,5,6}

Received: 27 June 2018 / Revised: 27 July 2018 / Accepted: 30 July 2018 / Published online: 6 August 2018
© Springer Science+Business Media, LLC, part of Springer Nature 2018

Abstract

Alzheimer's disease (AD) is a neurodegenerative disease that devastates the lives of its victims, and challenges the family members and health care infrastructures that care for them. Clinically, attempts to understand AD have focused on trying to predict the presence of, and more recently demonstrate the presence of, its characteristic amyloid plaque and neurofibrillary tangle pathologies. Fundamental research has also traditionally focused on understanding the generation, content, and pathogenicity of plaques and tangles, but in addition to this there is now an emerging independent interest in other molecular phenomena including apolipoprotein E, lipid metabolism, neuroinflammation, and mitochondrial function. While studies emphasizing the role of these phenomena have provided valuable AD insights, it is interesting that at the molecular level these entities extensively intertwine and interact. In this review, we provide a brief overview of why apolipoprotein E, lipid metabolism, neuroinflammation, and mitochondrial research have become increasingly ascendant in the AD research field, and present the case for studying these phenomena from an integrated perspective.

Keywords Alzheimer's · Apolipoprotein E · Lipids · Mitochondria · Neuroinflammation

Introduction

Alzheimer's disease (AD) is a neurodegenerative disorder that, over time, degrades an individual's memory and other cognitive abilities [1]. AD directly afflicts millions around the world, additionally affects family members who become de facto care-providers, and has devastating personal and national financial consequences [2, 3]. AD manifests in two

forms, a rare autosomal dominant familial form and the more common sporadic form; in this review we focus on the later.

Symptoms typically begin slowly, most frequently beginning after the age of 65, although in a considerable minority it arises some or potentially many years earlier [4]. Further, when considering the actual start-point, it is well-recognized that AD pathologies predate the onset of clinical symptoms for periods that can exceed decades [5].

The majority of AD pathology studies emphasize the plaques and tangles of the disease, which were described in Alois Alzheimer's initial reports [6, 7]. More recently, an enhanced interest in non-plaque, non-tangle pathologies has emerged. This review considers four well-recognized non-plaque, non-tangle AD-relevant phenomena: apolipoprotein E, neuroinflammation, mitochondrial dysfunction, and lipid homeostasis. Whether these phenomena primarily function as mediators of plaque and tangle-induced damage, or exist as independent or perhaps even driving events is not entirely clear at this time. An improved appreciation of these four entities could provide insight into how AD arises and, consequently, how to treat it.

✉ Russell H. Swerdlow
rswerdlow@kumc.edu

¹ University of Kansas Alzheimer's Disease Center, Fairway, KS, USA

² Neuroscience Graduate Program, University of Kansas Medical Center, Lawrence, KS, USA

³ Department of Biochemistry and Molecular Biology, University of Kansas Medical Center, Lawrence, KS, USA

⁴ Department of Molecular and Integrative Physiology, University of Kansas Medical Center, Lawrence, KS, USA

⁵ Department of Neurology, University of Kansas Medical Center, Lawrence, KS, USA

⁶ Landon Center on Aging, MS 2012, 3901 Rainbow Blvd, Kansas City, KS 66160, USA

Apolipoprotein E

Apolipoproteins are proteins that possess the ability to bind lipids. The protein-lipid complexes that subsequently arise from these associations are called lipoproteins [8]. The classically recognized function of apolipoproteins is to facilitate the movement of hydrophobic lipids within hydrophilic plasma and cerebrospinal fluid (CSF) compartments. Lipoproteins passing within aqueous environments enable the delivery of the lipid components to cells, which are internalized in conjunction with their apolipoprotein chaperones via receptor-mediated endocytosis.

Apolipoprotein E (ApoE) serves as the brain's major apolipoprotein [9–11]. As the brain contains approximately 25% of an individual's total cholesterol, ApoE is presumably crucial to maintaining brain function and structure. ApoE-cholesterol complexes trigger endocytosis upon binding the low-density lipoprotein receptor (LDLR), low density lipoprotein receptor-related protein 1 (LRP1), apolipoprotein E receptor 2 (Apoer2), or very low density lipoprotein receptor (VLDLR), which collectively comprise the low density lipoprotein (LDL) receptor family [12]. The apolipoprotein E-mediated delivery of lipids to cells comprises a crucial component of neuron repair following brain injury [13].

The gene for ApoE is located at chromosome 19q13.32, and contains four exons and three introns [14]. Two polymorphisms, one at rs429358 and the other at rs7412, define three different *APOE* alleles: ϵ 2, ϵ 3, and ϵ 4. The ϵ 4 allele is considered the ancestral form, and most closely resembles the chimpanzee gene. The ϵ 2 and ϵ 3 versions, on the other hand, have increased in frequency over the last 200,000 years [15]. The general frequencies of the different *APOE* alleles are approximately 6–7% for ϵ 2, 78–79% for ϵ 3, & 14–15% for ϵ 4, although the ϵ 4 allele is seen with higher frequency in populations closer to the poles and equator [16].

The alleles of *APOE* differ from each other by single amino acid substitutions. These occur at amino acids 112 & 158, with ϵ 2 having two cysteines, ϵ 3 having a cysteine and an arginine respectively, and ϵ 4 having two arginines [8]. ApoE has two domains, the N-terminal and C-terminal, which are connected by an intermediate hinge region [17].

This amino acid variation leads to structural differences between the variants. Some researchers have found the ϵ 4 version shows decreased terminal domain stability in both its C-terminal & N-terminal domains, [18] while others report the ϵ 4 version shows decreased stability in only the N-terminal domain [19]. This ϵ 4-associated loss of stability leads to a higher rate of protein turnover, and reduced levels of the protein in plasma [20]. The ϵ 4 variant also

shows a higher binding affinity for larger very low density lipoprotein (VLDL) and LDL particles, while ϵ 2 and ϵ 3 variants preferentially bind the smaller high density lipoprotein (HDL) [21].

ApoE is mainly produced in the liver and separately in the CNS [9]. CNS production is necessary given the high concentration of cholesterol in the brain, as well as cholesterol's inability to cross the blood brain barrier (BBB). Specifically, astrocytes and microglia produce ApoE under normal physiological conditions, while stress or injury can induce neuron production [22]. Neuron synthesis may reflect an attempt to repair intracellular damage, perhaps by increasing lipid delivery to the damaged cell.

The different variants of *APOE* can undergo dissimilar processing within neurons. Notably, a chymotrypsin-like serine protease cleaves the ϵ 4 variant more efficiently than the ϵ 3 form [23]. This cleavage generates fragments that may perturb other cell functions, and in particular mitochondrial function. A proportion of the cleaved ϵ 4 fragments bind to subunits of complexes III and IV of the mitochondrial respiratory chain on the inner mitochondrial membrane, with a subsequent inhibition of enzymatic activity [24]. In neurons, the protease-generated fragments also appear to associate with neurofibrillary tangles (NFT) [25, 26]. Thus, preferential cleavage of the ϵ 4 protein may confer neurotoxic qualities, which exist independent of its reduced lipid binding capacity.

APOE ϵ 4 alleles strongly associate with an increased lifetime risk of AD, and *APOE* ϵ 4 alleles currently represent the best-recognized genetic risk factor for late onset, sporadic AD [27]. Having one copy of the allele increases an individual's probability of developing AD by 2–3 fold, while having two copies confers an increase of approximately eightfold [28]. The *APOE* ϵ 2 variant, on the other hand, associates with a reduced lifetime risk of AD [29].

Although ϵ 2, ϵ 3, and ϵ 4 have different lipid binding affinities, whether or not ApoE's role in maintaining lipid homeostasis drives its AD associations is not fully understood. It is possible that the different ApoE isoforms could influence AD risk through mechanisms that are independent of its role in lipid homeostasis [30]. ApoE ϵ 4 has been shown to localize with amyloid- β (A β) in plaques, [31] and ApoE forms complexes with A β in the brain and facilitates its clearance from the brain. Notably, the ϵ 4 variant redirects these complexes away from LRP1 receptors and will only interact with the VLDLR. This disrupts clearance, and results in brain A β accumulation [32].

Lipid Homeostasis

Lipids are an integral part of the CNS and their homeostasis is vital to the proper functioning of the nervous system. In his original paper describing AD, Alois Alzheimer

noted many glial cells contained adipose saccules [33]. The processes primarily responsible for maintaining lipid homeostasis include uptake, catabolism, storage, and synthesis [34]. The brain itself holds large amounts of cholesterol, especially within neuron plasma cell membranes, glia cell plasma membranes, and oligodendrocyte-generated myelin sheaths [35].

Genes involved in lipid homeostasis influence AD risk. Two early genome wide association studies (GWAS) identified a contribution for two such genes, *CLU* and *PICALM* [36, 37]. The *CLU* gene produces the protein clusterin, also known as apolipoprotein J, which participates in membrane recycling [38, 39]. The phosphatidylinositol binding clathrin assembly protein expressed by *PICALM* plays a role in clathrin-mediated endocytosis.

In addition to *CLU* and *PICALM*, the bridging integrator-1 (*BINI*) gene also associates with AD. Similar to *PICALM*, *BINI* is relevant to endocytosis [40]. Although it reportedly influences A β levels, neither *BINI* knock-down nor over-expression alter APP processing [40]. This suggests *BINI* may influence AD through its effects on lipid biology. Finally, a subsequent GWAS revealed an association between AD and the gene that encodes the ATP-binding cassette sub-family A member 7 (*ABCA7*) [41]. The ATP-binding cassette family is involved in lipid trafficking [42].

A component of the plasma membrane is lipid rafts, which are composed of proteins, glycosphingolipids, and cholesterol that transit as a group independently along the cytoplasmic leaflet of the membrane [43]. In individuals with AD these rafts contain altered lipid profiles, which are characterized by low levels of long chain polyunsaturated fatty acids (LC-PUFA) and perturbed interactions between phospholipids and fatty acids [43]. Differences in lipid composition between individuals with AD and healthy individuals are seen both at the general cellular level, as well as by brain region. For example, the prefrontal cortex of AD patients exhibits altered levels of diacylglycerol and sphingolipids when compared to brains from healthy, age-matched individuals [43].

Higher serum cholesterol levels during midlife reportedly associate with an increased risk of late-onset AD [44]. Observational studies of individuals taking cholesterol-lowering statin drugs suggested this intervention might reduce the risk of developing AD or other dementias, [45] although clinical trials of statin drugs performed in AD participants showed no benefit [46].

Sterols used to generate myelin are produced via local synthesis, and not through the input of cholesterol into the brain [47]. The CNS is highly enriched in LC-PUFAs, the most important of which is docosahexaenoic acid (DHA), an omega-3 fatty acid [48]. Epidemiologic studies have reported that individuals with a reduced intake of omega-3

fatty acids have an increased risk of developing cognitive deficits [49].

Under baseline conditions, neurons are believed to utilize glucose to produce energy. They appear unable to perform fatty acid beta-oxidation, although astrocytes can successfully execute this biochemical pathway [50, 51]. Potential advantages of avoiding neuron beta-oxidation include preserving fatty acids for use in membrane expansion or maintenance, and also limiting beta-oxidation associated reactive oxygen species (ROS) production [52]. Advantages of pursuing astrocyte fatty acid beta-oxidation include the production of ketone bodies, which are potentially shuttled to neurons and used there to support energy production [53].

Cells use lipids for a number of functions during normal conditions, and when there is a change in the environment they can adapt to meet the new needs. The lipidome is a dynamic part of cells that will actively respond to changes in physiologic conditions [54, 55]. The downstream effects of an altered lipidome, though, are complex and could potentially result in dysfunction or disease, or simply reflect a downstream effect of dysfunction or disease.

Neuroinflammation

Inflammation represents the response of the immune system to endogenous or exogenous-induced tissue damage. When this response occurs in the central nervous system (CNS) it is referred to as neuroinflammation [56, 57]. In the short term, such reactions can minimize tissue damage or initiate repair, while chronic activation can conversely cause harm. Neuroinflammation is frequently present in neurodegenerative conditions, where it may reflect a byproduct of an upstream neurodegenerative pathology, or alternatively potentially instigate neurodegenerative pathology [58]. Neuroinflammation is observed in the early stages of AD, and may actually precede A β aggregation and tau neurofibrillary tangles [59].

Recent research identifies links between AD and variants within genes that influence inflammation, including *TREM2*, *CRI*, *HLA-DRB5/DRB1*, *INPPD5*, *MEFC2*, and *PTK2B* [37, 60, 61]. The triggering receptor expressed on myeloid cells 2 (*TREM2*) gene expresses a protein that is highly expressed on microglia, and which regulates cytokine release [62]. Complement receptor type 1 (*CRI*) is involved in the innate immune system's regulation of complement [63]. Human Leukocyte Antigen-antigen D Related beta chain (*HLA-DRB*) is implicated in Multiple Sclerosis, a degenerative-autoimmune disease that is characterized by inflammation [64].

The BBB ideally restricts the access of systemic immune cells to the CNS, thereby conferring an “immune privileged” status to the brain [65]. Accordingly, the brain

has its own resident cells that fill the role of an immune system, the microglia. Under baseline conditions they typically show phenotypic conformity, but in the context of homeostatic changes or frank damage morphologic and gene expression changes occur [66]. Neuron damage activates local microglia, and activation of local microglia in turn stimulates more remote microglia and amplifies the overall neuroinflammation state [67].

Activated microglia can produce a number of potentially neurotoxic pro-inflammatory cytokines, including interleukins (IL-1, IL-6), and tumor necrosis factor (TNF- α) [68]. Microglia further act as the macrophages of the nervous system, and remove debris, pathogens, and unwanted cells. It is well recognized that microglia actively disassemble synapses formed by damaged neurons [69]. Interestingly, in AD, microglia can be seen in the vicinity of otherwise intact neurons whose synapses are separating [70]. This raises the possibility that microglia-mediated synaptic stripping might represent an early pathologic event that, in its most extreme form, independently contributes to neurodegeneration. In AD, synapse degradation correlates reasonably well with cognitive function [71].

Microglia constitute only one neuroinflammation component. Astrocytes, oligodendrocytes, and the BBB also play a role in neuroinflammatory responses. The different components mediate specific effects that arise at different time-points in the course of a neuroinflammatory response. For this reason, inflammation that occurs within acute settings, such as a traumatic brain injury or a stroke, can dramatically differ from more chronic neuroinflammatory responses that arise during the course of multiple sclerosis or AD [57].

BBB disruption is a frequent component of neuroinflammation, to the point that myeloid cells transit from the circulation to the CNS parenchyma. In fact, myeloid cells only cross the BBB when there is a breakdown in its integrity [72]. During normal ageing the BBB becomes less restrictive, which allows for a more extensive interaction of blood immune cells with the brain parenchyma [73]. This upsets CNS homeostasis and further amplifies the inflammatory response, to the point that cells with immune-response potential that would normally remain quiescent become active. For example, astrocytes can be induced to release relatively large volumes of chemokines, which further perpetuates the pro-inflammatory cytokine response [74].

Due to the multitude of response amplifications that can occur, acute activation can potentially evolve into a chronic response that fuels its own perpetuation. In diseases such as AD, it is still not clear why neuroinflammation arises, or if it is initiated through an acute insult.

Mitochondrial Dysfunction

Mitochondria are considered the “powerhouse” of the cell because they produce most of the ATP used by cells. Mitochondria are dynamic, mobile organelles that can divide or fuse to form branching complex structures [75]. Such adaptations help cells meet their overall energy demands, address local energy stresses at specific cell regions, and repair or remove damaged mitochondria. To increase mitochondrial mass, cells can undergo a process called mitochondrial biogenesis. Mitochondrial biogenesis primarily occurs within neuron cell bodies, although it can also occur in axons as well [76]. Similarly, while most lysosome-mediated elimination of cell waste takes place in the soma, axons can also perform localized mitophagy via the PINK1 & Parkin pathway [77]. The flexible nature of the mitochondrial pool helps cells more efficiently meet their energy needs, and the loss of this flexibility can result in dysfunction or disease.

Mitochondrial dysfunction is seen in a number of neurodegenerative conditions including AD, Leber’s hereditary optic neuropathy (LHON), Parkinson’s disease, and amyotrophic lateral sclerosis (ALS) [78]. Mitochondria may account for observed relationships between advancing age and an age-related increase in the incidence of various neurodegenerative diseases, as mitochondrial function declines with advancing age [79]. In various tissues, including the nervous system, somatic mutations accumulate within mitochondrial DNA (mtDNA), which may contribute to or compound age-related changes in mitochondrial function [80, 81]. Some have proposed declines in mitochondrial function that exceed a threshold can contribute to the onset or progression of neurodegenerative diseases [82, 83]. According to one scheme, an individual’s genetic inheritance helps to define a baseline level of mitochondrial function, and the rate at which that individual’s mitochondria decline over decades further determines how rapidly the individual approaches a functional threshold that allows for the manifestation of an age related disease such as AD [84].

Dysfunctional mitochondria may produce reactive oxygen species (ROS), which can damage lipids, proteins, and DNA [85]. Multiple markers of oxidative stress are consistently elevated in AD subject autopsy brains, and perhaps other tissues as well [86]. Whether oxidative stress, either as a byproduct of mitochondrial dysfunction or some other generator, mediates pathology or primarily serves as a marker of mitochondrial dysfunction, is unclear.

Interactions and Integration

ApoE, lipid biology, neuroinflammation, and mitochondria are functionally inter-related. In one study that utilized *APOE* knock-in mice, the expression of two human $\epsilon 4$ transgenes resulted in higher serum cholesterol, lower brain cholesterol, and lower brain phospholipid levels [87]. On the other hand, in another study that featured $\epsilon 4$ knock-in mice, the synapse plasma membrane exofacial leaflets contained increased cholesterol [88, 89]. Therefore, although cholesterol levels in the brains of mice expressing the *APOE* $\epsilon 4$ allele may show an overall decrease, that decrease is not evenly distributed and localized increased levels of cholesterol are possible. ApoE $\epsilon 4$ expression in neurons, but not astrocytes, also reduces mitochondrial respiration [90]. It therefore appears that ApoE affects both lipid composition and mitochondrial respiration; whether these effects occur independently, or are mechanistically linked, is difficult to resolve.

Some note inflammation can influence lipid mobilization, at least in adipose tissue [91, 92]. Others have proposed that in the CNS, lipid mobilization may serve to alleviate energy stress [93]. Potentially relevant data suggest neuroinflammation may also regulate proteins that manage CNS lipid handling. For instance, following treatment with IL-1 β , rat glial cell cultures showed a significant increase in extracellular ApoE, while TNF- α treatment reduced extracellular ApoE [94]. This suggests the ability of neuroinflammatory responses to regulate ApoE action can be both precise yet flexible (Fig. 1).

Further research that highlights interactions among ApoE, lipid biology, and neuroinflammation include a study that compared mice expressing *APOE* $\epsilon 4$ versus $\epsilon 3$ transgenes. The $\epsilon 4$ -expressing mice showed an increased lipopolysaccharide (LPS) challenge-induced response of their NF- κ B signaling pathway, as well as increased microglial activation [95]. These results suggest that the *APOE* $\epsilon 4$ isoform potentiates neuroinflammation responses. To this point, the $\epsilon 2$ and $\epsilon 3$ isoforms can better interact with the low-density lipoprotein receptor family, with a resulting suppression of c-Jun N-terminal kinase (JNK) activation and a reduction in pro-inflammation signaling [96].

ApoE also participates in cerebrovascular regulation. By regulating a CypA-NF κ B-matrix metalloproteinase 9 pathway in pericytes, *APOE* $\epsilon 2$ and $\epsilon 3$ isoforms promote BBB integrity, while expression of $\epsilon 4$ promotes BBB permeability [97]. Prior studies, therefore, identify multiple pathways through which ApoE may facilitate, modulate, or otherwise influence neuroinflammation.

Interactions between mitochondria and neuroinflammation are also well recognized. BV2 and SH-SY5Y cells exposed to extracellular mtDNA increased their production

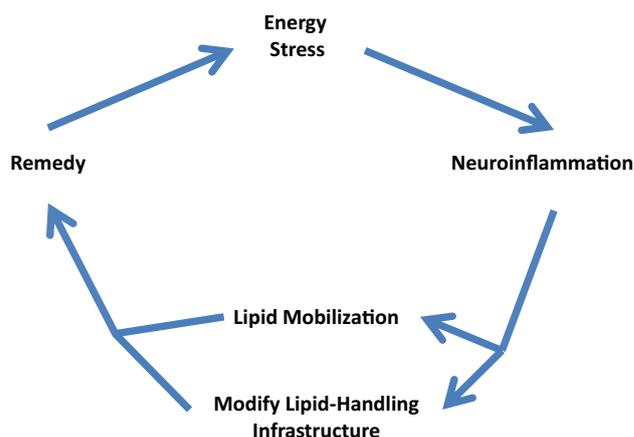


Fig. 1 Neuroinflammation may help cells respond to energy stress. As depicted, neuroinflammation activates both lipid mobilization and proteins (such as ApoE) that assist in the utilization of those lipids. These actions can serve to alleviate an energy stress, although since energy stress can also initiate neuroinflammation, this process could also potentially compensate for a primary energy deficit

of pro-inflammatory cytokines [98]. Astrocytes exposed to IL-1 β , one of the major pro-inflammatory cytokines, exhibited altered mitochondrial cycle dynamics, with an increase in mitochondrial fission [99].

In terms of relationships between mitochondria and lipid homeostasis, it is important to note mitochondria play essential roles in both lipid catabolism and synthesis [100]. Mitochondrial function, therefore, should presumably affect the lipidome.

BV2 microglia cells exposed to LPS showed an increased saturated fatty acid content, as well as a concomitant decrease in monounsaturated fatty acid levels [101]. In the setting of inflammation, macrophage ApoE expression declines due to changes in AP-1 and NF- κ B regulation [102]. Consequently, inflammation plays a role in the regulation of both ApoE and lipid levels. Figure 2 schematically illustrates potential connections between ApoE, lipid metabolism, neuroinflammation, and mitochondria.

Future Directions

ApoE, mitochondrial dysfunction, lipid metabolism, and neuroinflammation are implicated in AD. Strong biological connections functionally link ApoE, mitochondrial dysfunction, lipid metabolism, and neuroinflammation. The extent to which these components act independently to influence AD, or work through interactions to mediate the disease, remains to be seen. Addressing this question could facilitate a deeper understanding of what causes AD, and ideally how to treat it.

After over a century of studying AD, the field is finally reaching the point that it can visualize its classic

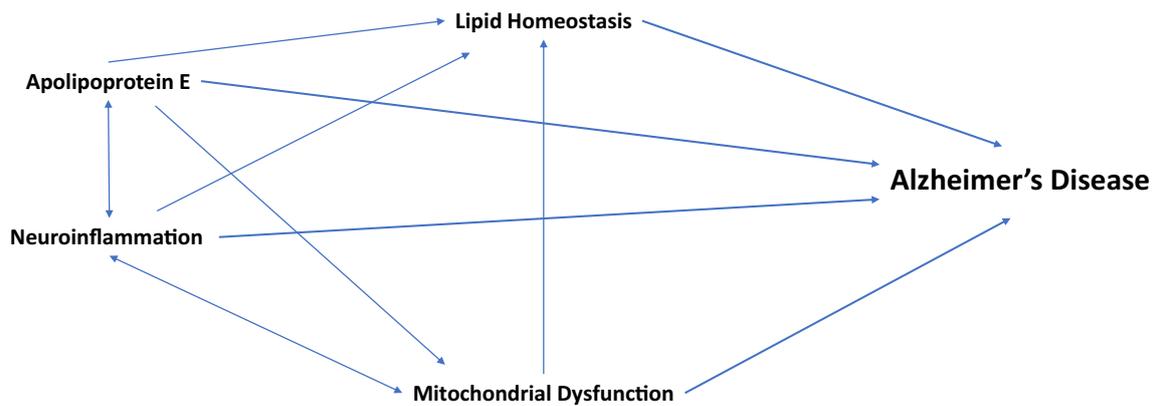


Fig. 2 Interactions between ApoE, lipid metabolism, neuroinflammation, and mitochondria may cooperatively influence the development of AD

histopathologies, the plaques and tangles, in living patients. Having achieved these milestones, the AD field is hopefully now poised to address the critical question of why plaques and tangles appear. A fuller understanding of ApoE, mitochondrial, lipid metabolism, and neuroinflammation biology could potentially inform this critical challenge.

Acknowledgements The authors are supported by the University of Kansas Alzheimer's Disease Center (NIA P30AG035982).

References

1. Querfurth HW, LaFerla FM (2010) Alzheimer's disease. *N Engl J Med* 362(4):329–344. <https://doi.org/10.1056/NEJMra0909142>
2. Thies W, Bleiler L (2011) 2011 Alzheimer's disease facts and figures. *Alzheimers Dement* 7(2):208–244. <https://doi.org/10.1016/j.jalz.2011.02.004>
3. Hebert LE, Scherr PA, Bienias JL, Bennett DA, Evans DA (2003) Alzheimer disease in the US population: prevalence estimates using the 2000 census. *Arch Neurol* 60(8):1119–1122. <https://doi.org/10.1001/archneur.60.8.1119>
4. Swerdlow RH (2007) Is aging part of Alzheimer's disease, or is Alzheimer's disease part of aging? *Neurobiol Aging* 28(10):1465–1480. <https://doi.org/10.1016/j.neurobiolaging.2006.06.021>
5. Jack CR Jr, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, Petersen RC, Trojanowski JQ (2010) Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. *Lancet Neurol* 9(1):119–128. [https://doi.org/10.1016/S1474-4422\(09\)70299-6](https://doi.org/10.1016/S1474-4422(09)70299-6)
6. Alzheimer A (1907) Über eine eigenartige Erkrankung der Hirnrinde. *Allg Z Psychiat Psych-Gerichtl Med* 64:146–148
7. Alzheimer A (1911) Über eigenartige Krankheitsfälle des späteren Alters. *Z die Gesamte Neurologie Psychiatrie* 4:456–485
8. Mahley RW, Innerarity TL, Rall SC Jr, Weisgraber KH (1984) Plasma lipoproteins: apolipoprotein structure and function. *J Lipid Res* 25(12):1277–1294
9. Elshourbagy NA, Liao WS, Mahley RW, Taylor JM (1985) Apolipoprotein E mRNA is abundant in the brain and adrenals, as well as in the liver, and is present in other peripheral tissues of rats and marmosets. *Proc Natl Acad Sci USA* 82(1):203–207
10. Mahley RW, Huang Y (2012) Apolipoprotein e sets the stage: response to injury triggers neuropathology. *Neuron* 76(5):871–885. <https://doi.org/10.1016/j.neuron.2012.11.020>
11. Pitas RE, Boyles JK, Lee SH, Foss D, Mahley RW (1987) Astrocytes synthesize apolipoprotein E and metabolize apolipoprotein E-containing lipoproteins. *Biochim Biophys Acta* 917(1):148–161
12. Holtzman DM, Herz J, Bu G (2012) Apolipoprotein E and apolipoprotein E receptors: normal biology and roles in Alzheimer disease. *Cold Spring Harb Perspect Med* 2(3):a006312. <https://doi.org/10.1101/cshperspect.a006312>
13. Liu CC, Liu CC, Kanekiyo T, Xu H, Bu G (2013) Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. *Nat Rev Neurol* 9(2):106–118. <https://doi.org/10.1038/nrneurol.2012.263>
14. Zerbino DR, Achuthan P, Akanni W, Amode MR, Barrell D, Bhai J, Billis K, Cummins C, Gall A, Giron CG, Gil L, Gordon L, Haggerty L, Haskell E, Hourlier T, Izuogu OG, Janacek SH, Juettemann T, To JK, Laird MR, Lavidas I, Liu Z, Loveland JE, Maurel T, McLaren W, Moore B, Mudge J, Murphy DN, Newman V, Nuhn M, Ogeh D, Ong CK, Parker A, Patricio M, Riat HS, Schuilenburg H, Sheppard D, Sparrow H, Taylor K, Thormann A, Vullo A, Walts B, Zadissa A, Frankish A, Hunt SE, Kostadima M, Langridge N, Martin FJ, Muffato M, Perry E, Ruffier M, Staines DM, Trevanion SJ, Aken BL, Cunningham F, Yates A, Flicek P (2018) Ensembl 2018. *Nucleic Acids Res* 46(D1):D754–D761. <https://doi.org/10.1093/nar/gkx1098>
15. Fullerton SM, Clark AG, Weiss KM, Nickerson DA, Taylor SL, Stengard JH, Salomaa V, Vartiainen E, Perola M, Boerwinkle E, Sing CF (2000) Apolipoprotein E variation at the sequence haplotype level: implications for the origin and maintenance of a major human polymorphism. *Am J Hum Genet* 67(4):881–900. <https://doi.org/10.1086/303070>
16. Eisenberg DT, Kuzawa CW, Hayes MG (2010) Worldwide allele frequencies of the human apolipoprotein E gene: climate, local adaptations, and evolutionary history. *Am J Phys Anthropol* 143(1):100–111. <https://doi.org/10.1002/ajpa.21298>
17. Wetterau JR, Aggerbeck LP, Rall SC Jr, Weisgraber KH (1988) Human apolipoprotein E3 in aqueous solution. I. Evidence for two structural domains. *J Biol Chem* 263(13):6240–6248
18. Mizuguchi C, Hata M, Dhanasekaran P, Nickel M, Okuhira K, Phillips MC, Lund-Katz S, Saito H (2014) Fluorescence study of domain structure and lipid interaction of human apolipoproteins E3 and E4. *Biochim Biophys Acta* 1841(12):1716–1724. <https://doi.org/10.1016/j.bbaliip.2014.09.019>
19. Morrow JA, Segall ML, Lund-Katz S, Phillips MC, Knapp M, Rupp B, Weisgraber KH (2000) Differences in stability among

- the human apolipoprotein E isoforms determined by the amino-terminal domain. *Biochemistry* 39(38):11657–11666
20. Ramaswamy G, Xu Q, Huang Y, Weisgraber KH (2005) Effect of domain interaction on apolipoprotein E levels in mouse brain. *J Neurosci* 25(46):10658–10663. <https://doi.org/10.1523/JNEUROSCI.1922-05.2005>
 21. Hatters DM, Peters-Libeu CA, Weisgraber KH (2006) Apolipoprotein E structure: insights into function. *Trends Biochem Sci* 31(8):445–454. <https://doi.org/10.1016/j.tibs.2006.06.008>
 22. Aoki K, Uchihara T, Sanjo N, Nakamura A, Ikeda K, Tsuchiya K, Wakayama Y (2003) Increased expression of neuronal apolipoprotein E in human brain with cerebral infarction. *Stroke* 34(4):875–880. <https://doi.org/10.1161/01.STR.0000064320.73388.C6>
 23. Harris FM, Brecht WJ, Xu Q, Tesseur I, Kekoni L, Wyss-Coray T, Fish JD, Masliah E, Hopkins PC, Scearce-Levie K, Weisgraber KH, Mucke L, Mahley RW, Huang Y (2003) Carboxyl-terminal-truncated apolipoprotein E4 causes Alzheimer's disease-like neurodegeneration and behavioral deficits in transgenic mice. *Proc Natl Acad Sci USA* 100(19):10966–10971. <https://doi.org/10.1073/pnas.1434398100>
 24. Nakamura T, Watanabe A, Fujino T, Hosono T, Michikawa M (2009) Apolipoprotein E4 (1-272) fragment is associated with mitochondrial proteins and affects mitochondrial function in neuronal cells. *Mol Neurodegener* 4:35. <https://doi.org/10.1186/1750-1326-4-35>
 25. Huang Y, Liu XQ, Wyss-Coray T, Brecht WJ, Sanan DA, Mahley RW (2001) Apolipoprotein E fragments present in Alzheimer's disease brains induce neurofibrillary tangle-like intracellular inclusions in neurons. *Proc Natl Acad Sci USA* 98(15):8838–8843. <https://doi.org/10.1073/pnas.151254698>
 26. Rohn TT, Catlin LW, Coonse KG, Habig JW (2012) Identification of an amino-terminal fragment of apolipoprotein E4 that localizes to neurofibrillary tangles of the Alzheimer's disease brain. *Brain Res* 1475:106–115. <https://doi.org/10.1016/j.brainres.2012.08.003>
 27. Strittmatter WJ, Saunders AM, Schmechel D, Pericak-Vance M, Enghild J, Salvesen GS, Roses AD (1993) Apolipoprotein E: high-avidity binding to beta-amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc Natl Acad Sci USA* 90(5):1977–1981
 28. Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, Roses AD, Haines JL, Pericak-Vance MA (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261(5123):921–923
 29. Conejero-Goldberg C, Gomar JJ, Bobes-Bascaran T, Hyde TM, Kleinman JE, Herman MM, Chen S, Davies P, Goldberg TE (2014) APOE2 enhances neuroprotection against Alzheimer's disease through multiple molecular mechanisms. *Mol Psychiatry* 19(11):1243–1250. <https://doi.org/10.1038/mp.2013.194>
 30. Mahley RW, Rall SC Jr (2000) Apolipoprotein E: far more than a lipid transport protein. *Annu Rev Genom Hum Genet* 1:507–537. <https://doi.org/10.1146/annurev.genom.1.1.507>
 31. Koffie RM, Hashimoto T, Tai HC, Kay KR, Serrano-Pozo A, Joyner D, Hou S, Kopeikina KJ, Frosch MP, Lee VM, Holtzman DM, Hyman BT, Spire-Jones TL (2012) Apolipoprotein E4 effects in Alzheimer's disease are mediated by synaptotoxic oligomeric amyloid-beta. *Brain* 135(Pt 7):2155–2168. <https://doi.org/10.1093/brain/aw127>
 32. Deane R, Sagare A, Hamm K, Parisi M, Lane S, Finn MB, Holtzman DM, Zlokovic BV (2008) apoE isoform-specific disruption of amyloid beta peptide clearance from mouse brain. *J Clin Invest* 118(12):4002–4013. <https://doi.org/10.1172/JCI36663>
 33. Alzheimer A, Stelzmann RA, Schnitzlein HN, Murtagh FR (1995) An English translation of Alzheimer's 1907 paper, "Über eine eigenartige Erkrankung der Hirnrinde". *Clin Anat* 8(6):429–431. <https://doi.org/10.1002/ca.980080612>
 34. Grosvenor S, Grimm MO, Friess P, Hartmann T (2010) Role of amyloid beta in lipid homeostasis. *Biochim Biophys Acta* 1801(8):966–974. <https://doi.org/10.1016/j.bbali.2010.05.002>
 35. Dietschy JM, Turley SD (2001) Cholesterol metabolism in the brain. *Curr Opin Lipidol* 12(2):105–112
 36. Harold D, Abraham R, Hollingworth P, Sims R, Gerrish A, Hamshere ML, Pahwa JS, Moskva V, Dowzell K, Williams A, Jones N, Thomas C, Stretton A, Morgan AR, Lovestone S, Powell J, Proitsi P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Morgan K, Brown KS, Passmore PA, Craig D, McGuinness B, Todd S, Holmes C, Mann D, Smith AD, Love S, Kehoe PG, Hardy J, Mead S, Fox N, Rossor M, Collinge J, Maier W, Jessen F, Schurmann B, Heun R, van den Bussche H, Heuser I, Kornhuber J, Wiltfang J, Dichgans M, Frolich L, Hampel H, Hull M, Rujescu D, Goate AM, Kauwe JS, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K, Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin A, Gilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tzolaki M, Singleton AB, Guerreiro R, Muhleisen TW, Nothen MM, Moebus S, Jockel KH, Klopp N, Wichmann HE, Carrasquillo MM, Pankratz VS, Younkin SG, Holmans PA, O'Donovan M, Owen MJ, Williams J (2009) Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet* 41(10):1088–1093. <https://doi.org/10.1038/ng.440>
 37. Lambert JC, Heath S, Even G, Campion D, Sleegers K, Hiltnen M, Combarros O, Zelenika D, Bullido MJ, Tavernier B, Letenneur L, Bettens K, Berr C, Pasquier F, Fievet N, Barberger-Gateau P, Engelborghs S, De Deyn P, Mateo I, Franck A, Helisalmi S, Porcellini E, Hanon O, European Alzheimer's Disease Initiative I, de Pancorbo MM, Lendon C, Dufouil C, Jaillard C, Leveillard T, Alvarez V, Bosco P, Mancuso M, Panza F, Nacmias B, Bossu P, Piccardi P, Annoni G, Seripa D, Galimberti D, Hannequin D, Licastro F, Soininen H, Ritchie K, Blanche H, Dartigues JF, Tzourio C, Gut I, Van Broeckhoven C, Alperovitch A, Lathrop M, Amouyel P (2009) Genome-wide association study identifies variants at CLU and CR1 associated with Alzheimer's disease. *Nat Genet* 41(10):1094–1099. <https://doi.org/10.1038/ng.439>
 38. Jones SE, Jomary C (2002) Clusterin. *Int J Biochem Cell Biol* 34(5):427–431
 39. Calero M, Rostagno A, Matsubara E, Zlokovic B, Frangione B, Ghiso J (2000) Apolipoprotein J (clusterin) and Alzheimer's disease. *Microsc Res Tech* 50 (4):305–315. [https://doi.org/10.1002/1097-0029\(20000815\)50:4<3C305::AID-JEMT10%3E3.0.CO;2-L](https://doi.org/10.1002/1097-0029(20000815)50:4<3C305::AID-JEMT10%3E3.0.CO;2-L)
 40. Glennon EB, Whitehouse II, Miners JS, Kehoe PG, Love S, Kellert KA, Hooper NM (2013) BIN1 is decreased in sporadic but not familial Alzheimer's disease or in aging. *PLoS ONE* 8(10):e78806. <https://doi.org/10.1371/journal.pone.0078806>
 41. Hollingworth P, Harold D, Sims R, Gerrish A, Lambert JC, Carrasquillo MM, Abraham R, Hamshere ML, Pahwa JS, Moskva V, Dowzell K, Jones N, Stretton A, Thomas C, Richards A, Ivanov D, Widdowson C, Chapman J, Lovestone S, Powell J, Proitsi P, Lupton MK, Brayne C, Rubinsztein DC, Gill M, Lawlor B, Lynch A, Brown KS, Passmore PA, Craig D, McGuinness B, Todd S, Holmes C, Mann D, Smith AD, Beaumont H, Warden D, Wilcock G, Love S, Kehoe PG, Hooper NM, Vardy ER, Hardy J, Mead S, Fox NC, Rossor M, Collinge J, Maier W, Jessen F, Ruther E, Schurmann B, Heun R, Kolsch H, van den Bussche H, Heuser I, Kornhuber J, Wiltfang J, Dichgans M, Frolich L, Hampel H, Gallacher J, Hull M, Rujescu D, Giegling I, Goate AM, Kauwe JS, Cruchaga C, Nowotny P, Morris JC, Mayo K, Sleegers K, Bettens K,

- Engelborghs S, De Deyn PP, Van Broeckhoven C, Livingston G, Bass NJ, Gurling H, McQuillin A, Gwilliam R, Deloukas P, Al-Chalabi A, Shaw CE, Tsolaki M, Singleton AB, Guerreiro R, Muhleisen TW, Nothen MM, Moebus S, Jockel KH, Klopp N, Wichmann HE, Pankratz VS, Sando SB, Aasly JO, Barcikowska M, Wszolek ZK, Dickson DW, Graff-Radford NR, Petersen RC, Alzheimer's Disease Neuroimaging I, van Duijn CM, Breteler MM, Ikram MA, DeStefano AL, Fitzpatrick AL, Lopez O, Launer LJ, Seshadri S, consortium C, Berr C, Campion D, Epelbaum J, Dartigues JF, Tzourio C, Alperovitch A, Lathrop M, consortium E, Feulner TM, Friedrich P, Riehle C, Krawczak M, Schreiber S, Mayhaus M, Nicolhaus S, Wagenpfeil S, Steinberg S, Stefansson H, Stefansson K, Snaedal J, Bjornsson S, Jonsson PV, Chouraki V, Genier-Boley B, Hiltunen M, Soininen H, Combarros O, Zelenika D, Delepine M, Bullido MJ, Pasquier F, Mateo I, Frank-Garcia A, Porcellini E, Hanon O, Coto E, Alvarez V, Bosco P, Siciliano G, Mancuso M, Panza F, Solfrizzi V, Nacmias B, Sorbi S, Bossu P, Piccardi P, Arosio B, Annoni G, Seripa D, Pilotto A, Scarpini E, Galimberti D, Brice A, Hannequin D, Licastrò F, Jones L, Holmans PA, Jonsson T, Riemenschneider M, Morgan K, Younkin SG, Owen MJ, O'Donovan M, Amouyel P, Williams J (2011) Common variants at ABCA7, MS4A6A/MS4A4E, EPHA1, CD33 and CD2AP are associated with Alzheimer's disease. *Nat Genet* 43(5):429–435. <https://doi.org/10.1038/ng.803>
42. Villegas-Llerena C, Phillips A, Garcia-Reitboeck P, Hardy J, Pocock JM (2016) Microglial genes regulating neuroinflammation in the progression of Alzheimer's disease. *Curr Opin Neurobiol* 36:74–81. <https://doi.org/10.1016/j.conb.2015.10.004>
43. Martin V, Fabelo N, Santpere G, Puig B, Marin R, Ferrer I, Diaz M (2010) Lipid alterations in lipid rafts from Alzheimer's disease human brain cortex. *J Alzheimers Dis* 19(2):489–502. <https://doi.org/10.3233/JAD-2010-1242>
44. Solomon A, Kivipelto M, Wolozin B, Zhou J, Whitmer RA (2009) Midlife serum cholesterol and increased risk of Alzheimer's and vascular dementia three decades later. *Dement Geriatr Cogn Disord* 28(1):75–80. <https://doi.org/10.1159/000231980>
45. Macedo AF, Taylor FC, Casas JP, Adler A, Prieto-Merino D, Ebrahim S (2014) Unintended effects of statins from observational studies in the general population: systematic review and meta-analysis. *BMC Med* 12:51. <https://doi.org/10.1186/1741-7015-12-51>
46. Sano M, Bell KL, Galasko D, Galvin JE, Thomas RG, van Dyck CH, Aisen PS (2011) A randomized, double-blind, placebo-controlled trial of simvastatin to treat Alzheimer disease. *Neurology* 77(6):556–563. <https://doi.org/10.1212/WNL.0b013e318228bfl1>
47. Jurevics H, Morell P (1995) Cholesterol for synthesis of myelin is made locally, not imported into brain. *J Neurochem* 64(2):895–901
48. Zarate R, El Jaber-Vazdekis N, Tejera N, Perez JA, Rodriguez C (2017) Significance of long chain polyunsaturated fatty acids in human health. *Clin Transl Med* 6(1):25. <https://doi.org/10.1186/s40169-017-0153-6>
49. Cole GM, Ma QL, Frautschy SA (2009) Omega-3 fatty acids and dementia. *Prostaglandins Leukot Essent Fatty Acids* 81(2–3):213–221. <https://doi.org/10.1016/j.plefa.2009.05.015>
50. Mergenthaler P, Lindauer U, Dienel GA, Meisel A (2013) Sugar for the brain: the role of glucose in physiological and pathological brain function. *Trends Neurosci* 36(10):587–597. <https://doi.org/10.1016/j.tins.2013.07.001>
51. Edmond J, Robbins RA, Bergstrom JD, Cole RA, de Vellis J (1987) Capacity for substrate utilization in oxidative metabolism by neurons, astrocytes, and oligodendrocytes from developing brain in primary culture. *J Neurosci Res* 18(4):551–561. <https://doi.org/10.1002/jnr.490180407>
52. Tracey TJ, Steyn FJ, Wolvetang EJ, Ngo ST (2018) Neuronal lipid metabolism: multiple pathways driving functional outcomes in health and disease. *Front Mol Neurosci* 11:10. <https://doi.org/10.3389/fnmol.2018.00010>
53. Guzman M, Blazquez C (2001) Is there an astrocyte-neuron ketone body shuttle? *Trends in endocrinology and metabolism*. *TEM* 12(4):169–173
54. Garcia-Canaveras JC, Peris-Diaz MD, Alcoriza-Balaguer MI, Cerdan-Calero M, Donato MT, Lahoz A (2017) A lipidomic cell-based assay for studying drug-induced phospholipidosis and steatosis. *Electrophoresis* 38(18):2331–2340. <https://doi.org/10.1002/elps.201700079>
55. Lydic TA, Goo YH (2018) Lipidomics unveils the complexity of the lipidome in metabolic diseases. *Clin Transl Med* 7(1):4. <https://doi.org/10.1186/s40169-018-0182-9>
56. Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, Jacobs AH, Wyss-Coray T, Vitorica J, Ransohoff RM, Herrup K, Frautschy SA, Finsen B, Brown GC, Verkhratsky A, Yamanaka K, Koistinaho J, Latz E, Halle A, Petzold GC, Town T, Morgan D, Shinohara ML, Perry VH, Holmes C, Bazan NG, Brooks DJ, Hunot S, Joseph B, Deigendesch N, Garaschuk O, Boddeke E, Dinarello CA, Breitner JC, Cole GM, Golenbock DT, Kummer MP (2015) Neuroinflammation in Alzheimer's disease. *Lancet Neurol* 14(4):388–405. [https://doi.org/10.1016/S1474-4422\(15\)70016-5](https://doi.org/10.1016/S1474-4422(15)70016-5)
57. Shi FD (2015) Neuroinflammation. *Neurosci Bull* 31(6):714–716. <https://doi.org/10.1007/s12264-015-1568-y>
58. Obermeier B, Daneman R, Ransohoff RM (2013) Development, maintenance and disruption of the blood-brain barrier. *Nat Med* 19(12):1584–1596. <https://doi.org/10.1038/nm.3407>
59. Eikelenboom P, van Exel E, Hoozemans JJ, Veerhuis R, Roze-muller AJ, van Gool WA (2010) Neuroinflammation—an early event in both the history and pathogenesis of Alzheimer's disease. *Neurodegener Dis* 7(1–3):38–41. <https://doi.org/10.1159/000283480>
60. Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, DeStafano AL, Bis JC, Beecham GW, Grenier-Boley B, Russo G, Thornton-Wells TA, Jones N, Smith AV, Chouraki V, Thomas C, Ikram MA, Zelenika D, Vardarajan BN, Kamatani Y, Lin CF, Gerrish A, Schmidt H, Kunkle B, Dunstan ML, Ruiz A, Bihoreau MT, Choi SH, Reitz C, Pasquier F, Cruchaga C, Craig D, Amin N, Berr C, Lopez OL, De Jager PL, Deramecourt V, Johnston JA, Evans D, Lovestone S, Letenneur L, Moron FJ, Rubinsztein DC, Eiriksdottir G, Sleegers K, Goate AM, Fievet N, Huettelmann MW, Gill M, Brown K, Kamboh MI, Keller L, Barberger-Gateau P, McGuinness B, Larson EB, Green R, Myers AJ, Dufouil C, Todd S, Wallon D, Love S, Rogava E, Gallacher J, St George-Hyslop P, Clarimon J, Lleo A, Bayer A, Tsuang DW, Yu L, Tsolaki M, Bossu P, Spalletta G, Proitsi P, Collinge J, Sorbi S, Sanchez-Garcia F, Fox NC, Hardy J, Deniz Naranjo MC, Bosco P, Clarke R, Brayne C, Galimberti D, Mancuso M, Matthews F, European Alzheimer's Disease I, Genetic, Environmental Risk in Alzheimer's D, Alzheimer's Disease Genetic C, Cohorts for H, Aging Research in Genomic E, Moebus S, Mecocci P, Del Zompo M, Maier W, Hampel H, Pilotto A, Bullido M, Panza F, Caffarra P, Nacmias B, Gilbert JR, Mayhaus M, Lannefelt L, Hakonarson H, Pichler S, Carrasquillo MM, Ingelsson M, Beekly D, Alvarez V, Zou F, Valladares O, Younkin SG, Coto E, Hamilton-Nelson KL, Gu W, Razquin C, Pastor P, Mateo I, Owen MJ, Faber KM, Jonsson PV, Combarros O, O'Donovan MC, Cantwell LB, Soininen H, Blacker D, Mead S, Mosley Jr. TH, Bennett DA, Harris TB, Fratiglioni L, Holmes C, de Bruijn RF, Passmore P, Montine TJ, Bettens K, Rotter JJ, Brice A, Morgan K, Foroud TM, Kukull WA, Hannequin D, Powell JF, Nalls MA, Ritchie K, Lunetta KL, Kauwe JS, Boerwinkle E, Riemenschneider M, Boada M, Hiltunen M, Martin

- ER, Schmidt R, Rujescu D, Wang LS, Dartigues JF, Mayeux R, Tzourio C, Hofman A, Nothen MM, Graff C, Psaty BM, Jones L, Haines JL, Holmans PA, Lathrop M, Pericak-Vance MA, Launer LJ, Farrer LA, van Duijn CM, Van Broeckhoven C, Moskvina V, Seshadri S, Williams J, Schellenberg GD, Amouyel P (2013) Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet* 45(12):1452–1458. <https://doi.org/10.1038/ng.2802>
61. Wilkins HM, Carl SM, Greenleaf AC, Festoff BW, Swerdlow RH (2014) Bioenergetic dysfunction and inflammation in Alzheimer's disease: a possible connection. *Front Aging Neurosci* 6:311. <https://doi.org/10.3389/fnagi.2014.00311>
 62. Atagi Y, Liu CC, Painter MM, Chen XF, Verbeeck C, Zheng H, Li X, Rademakers R, Kang SS, Xu H, Younkin S, Das P, Fryer JD, Bu G (2015) Apolipoprotein E is a ligand for triggering receptor expressed on myeloid cells 2 (TREM2). *J Biol Chem* 290(43):26043–26050. <https://doi.org/10.1074/jbc.M115.679043>
 63. Karch CM, Goate AM (2015) Alzheimer's disease risk genes and mechanisms of disease pathogenesis. *Biol Psychiatry* 77(1):43–51. <https://doi.org/10.1016/j.biopsych.2014.05.006>
 64. Hollenbach JA, Oksenberg JR (2015) The immunogenetics of multiple sclerosis: a comprehensive review. *J Autoimmun* 64:13–25. <https://doi.org/10.1016/j.jaut.2015.06.010>
 65. Galea I, Bechmann I, Perry VH (2007) What is immune privilege (not)? *Trends Immunol* 28(1):12–18. <https://doi.org/10.1016/j.it.2006.11.004>
 66. Perry VH, Nicoll JA, Holmes C (2010) Microglia in neurodegenerative disease. *Nat Rev Neurol* 6(4):193–201. <https://doi.org/10.1038/nrneurol.2010.17>
 67. Block ML (2008) NADPH oxidase as a therapeutic target in Alzheimer's disease. *BMC Neurosci* 9(Suppl 2):S8. <https://doi.org/10.1186/1471-2202-9-S2-S8>
 68. Smith JA, Das A, Ray SK, Banik NL (2012) Role of pro-inflammatory cytokines released from microglia in neurodegenerative diseases. *Brain Res Bull* 87(1):10–20. <https://doi.org/10.1016/j.brainresbull.2011.10.004>
 69. Kettenmann H, Kirchhoff F, Verkhratsky A (2013) Microglia: new roles for the synaptic stripper. *Neuron* 77(1):10–18. <https://doi.org/10.1016/j.neuron.2012.12.023>
 70. Perry VH, O'Connor V (2010) The role of microglia in synaptic stripping and synaptic degeneration: a revised perspective. *ASN Neuro* 2(5):e00047. <https://doi.org/10.1042/AN20100024>
 71. DeKosky ST, Scheff SW (1990) Synapse loss in frontal cortex biopsies in Alzheimer's disease: correlation with cognitive severity. *Ann Neurol* 27(5):457–464. <https://doi.org/10.1002/ana.410270502>
 72. Ransohoff RM, Engelhardt B (2012) The anatomical and cellular basis of immune surveillance in the central nervous system. *Nat Rev Immunol* 12(9):623–635. <https://doi.org/10.1038/nri3265>
 73. Marques F, Sousa JC, Sousa N, Palha JA (2013) Blood-brain-barriers in aging and in Alzheimer's disease. *Mol Neurodegener* 8:38. <https://doi.org/10.1186/1750-1326-8-38>
 74. Hennessy E, Griffin EW, Cunningham C (2015) Astrocytes are primed by chronic neurodegeneration to produce exaggerated chemokine and cell infiltration responses to acute stimulation with the cytokines IL-1beta and TNF-alpha. *J Neurosci* 35(22):8411–8422. <https://doi.org/10.1523/JNEUROSCI.2745-14.2015>
 75. Hales KG (2004) The machinery of mitochondrial fusion, division, and distribution, and emerging connections to apoptosis. *Mitochondrion* 4(4):285–308. <https://doi.org/10.1016/j.mito.2004.05.007>
 76. Amiri M, Hollenbeck PJ (2008) Mitochondrial biogenesis in the axons of vertebrate peripheral neurons. *Dev Neurobiol* 68(11):1348–1361. <https://doi.org/10.1002/dneu.20668>
 77. Ashrafi G, Schlehe JS, LaVoie MJ, Schwarz TL (2014) Mitophagy of damaged mitochondria occurs locally in distal neuronal axons and requires PINK1 and Parkin. *J Cell Biol* 206(5):655–670. <https://doi.org/10.1083/jcb.201401070>
 78. Swerdlow RH (2009) The neurodegenerative mitochondrialopathies. *J Alzheimers Dis* 17(4):737–751. <https://doi.org/10.3233/JAD-2009-1095>
 79. Swerdlow RH (2011) Brain aging, Alzheimer's disease, and mitochondria. *Biochim Biophys Acta* 1812(12):1630–1639. <https://doi.org/10.1016/j.bbadis.2011.08.012>
 80. Swerdlow RH, Koppel S, Weidling I, Hayley C, Ji Y, Wilkins HM (2017) Mitochondria, cybrids, aging, and Alzheimer's disease. *Progress in molecular biology translational science* 146:259–302. <https://doi.org/10.1016/bs.pmbts.2016.12.017>
 81. Wallace DC (1992) Mitochondrial genetics: a paradigm for aging and degenerative diseases? *Science* 256(5057):628–632
 82. Breuer ME, Koopman WJ, Koene S, Nooteboom M, Rodenburg RJ, Willems PH, Smeitink JA (2013) The role of mitochondrial OXPHOS dysfunction in the development of neurologic diseases. *Neurobiol Dis* 51:27–34. <https://doi.org/10.1016/j.nbd.2012.03.007>
 83. Lin MT, Beal MF (2006) Mitochondrial dysfunction and oxidative stress in neurodegenerative diseases. *Nature* 443(7113):787–795. <https://doi.org/10.1038/nature05292>
 84. Swerdlow RH, Khan SM (2004) A “mitochondrial cascade hypothesis” for sporadic Alzheimer's disease. *Med Hypotheses* 63(1):8–20. <https://doi.org/10.1016/j.mehy.2003.12.045>
 85. Schieber M, Chandel NS (2014) ROS function in redox signaling and oxidative stress. *Curr Biol* 24(10):R453–R462. <https://doi.org/10.1016/j.cub.2014.03.034>
 86. Good PF, Werner P, Hsu A, Olanow CW, Perl DP (1996) Evidence of neuronal oxidative damage in Alzheimer's disease. *Am J Pathol* 149(1):21–28
 87. Hamanaka H, Katoh-Fukui Y, Suzuki K, Kobayashi M, Suzuki R, Motegi Y, Nakahara Y, Takeshita A, Kawai M, Ishiguro K, Yokoyama M, Fujita SC (2000) Altered cholesterol metabolism in human apolipoprotein E4 knock-in mice. *Hum Mol Genet* 9(3):353–361
 88. Hayashi H, Igbavboa U, Hamanaka H, Kobayashi M, Fujita SC, Wood WG, Yanagisawa K (2002) Cholesterol is increased in the exofacial leaflet of synaptic plasma membranes of human apolipoprotein E4 knock-in mice. *Neuroreport* 13(4):383–386
 89. Cossec JC, Marquer C, Panchal M, Lazar AN, Duyckaerts C, Potier MC (2010) Cholesterol changes in Alzheimer's disease: methods of analysis and impact on the formation of enlarged endosomes. *Biochim Biophys Acta* 1801(8):839–845. <https://doi.org/10.1016/j.bbalip.2010.03.010>
 90. Chen HK, Ji ZS, Dodson SE, Miranda RD, Rosenblum CI, Reynolds IJ, Freedman SB, Weisgraber KH, Huang Y, Mahley RW (2011) Apolipoprotein E4 domain interaction mediates detrimental effects on mitochondria and is a potential therapeutic target for Alzheimer disease. *J Biol Chem* 286(7):5215–5221. <https://doi.org/10.1074/jbc.M110.151084>
 91. Coppack SW (2001) Pro-inflammatory cytokines and adipose tissue. *Proc Nutr Soc* 60(3):349–356
 92. Ogawa H, Nielsen S, Kawakami M (1989) Cachectin/tumor necrosis factor and interleukin-1 show different modes of combined effect on lipoprotein lipase activity and intracellular lipolysis in 3T3-L1 cells. *Biochim Biophys Acta* 1003(2):131–135
 93. Klosinski LP, Yao J, Yin F, Fonteh AN, Harrington MG, Christensen TA, Trushina E, Brinton RD (2015) White matter lipids as a ketogenic fuel supply in aging female brain: implications for Alzheimer's disease. *EBioMedicine* 2(12):1888–1904. <https://doi.org/10.1016/j.ebiom.2015.11.002>

94. Aleong R, Blain JF, Poirier J (2008) Pro-inflammatory cytokines modulate glial apolipoprotein E secretion. *Curr Alzheimer Res* 5(1):33–37
95. Ophir G, Amariglio N, Jacob-Hirsch J, Elkon R, Rechavi G, Michaelson DM (2005) Apolipoprotein E4 enhances brain inflammation by modulation of the NF-kappaB signaling cascade. *Neurobiol Dis* 20(3):709–718. <https://doi.org/10.1016/j.nbd.2005.05.002>
96. Pocivavsek A, Mikhailenko I, Strickland DK, Rebeck GW (2009) Microglial low-density lipoprotein receptor-related protein 1 modulates c-Jun N-terminal kinase activation. *J Neuroimmunol* 214(1–2):25–32. <https://doi.org/10.1016/j.jneuroim.2009.06.010>
97. Bell RD, Winkler EA, Singh I, Sagare AP, Deane R, Wu Z, Holtzman DM, Betsholtz C, Armulik A, Sallstrom J, Berk BC, Zlokovic BV (2012) Apolipoprotein E controls cerebrovascular integrity via cyclophilin A. *Nature* 485(7399):512–516. <https://doi.org/10.1038/nature11087>
98. Wilkins HM, Carl SM, Weber SG, Ramanujan SA, Festoff BW, Linseman DA, Swerdlow RH (2015) Mitochondrial lysates induce inflammation and Alzheimer’s disease-relevant changes in microglial and neuronal cells. *J Alzheimers Dis* 45(1):305–318. <https://doi.org/10.3233/JAD-142334>
99. Motori E, Puyal J, Toni N, Ghanem A, Angeloni C, Malaguti M, Cantelli-Forti G, Berninger B, Conzelmann KK, Gotz M, Winklhofer KF, Hrelia S, Bergami M (2013) Inflammation-induced alteration of astrocyte mitochondrial dynamics requires autophagy for mitochondrial network maintenance. *Cell Metab* 18(6):844–859. <https://doi.org/10.1016/j.cmet.2013.11.005>
100. Scheffler IE (2008) *Mitochondria*, 2nd edn. Wiley-Liss, Hoboken
101. Button EB, Mitchell AS, Domingos MM, Chung JH, Bradley RM, Hashemi A, Marvyn PM, Patterson AC, Stark KD, Quadri-latero J, Duncan RE (2014) Microglial cell activation increases saturated and decreases monounsaturated fatty acid content, but both lipid species are proinflammatory. *Lipids* 49(4):305–316. <https://doi.org/10.1007/s11745-014-3882-y>
102. Gafencu AV, Robciuc MR, Fuior E, Zannis VI, Kardassis D, Simionescu M (2007) Inflammatory signaling pathways regulating ApoE gene expression in macrophages. *J Biol Chem* 282(30):21776–21785. <https://doi.org/10.1074/jbc.M611422200>