



Review Article

Molecular mechanisms of immunoreceptors in platelets

Fahd A. Kuriri, Cindy J. O'Malley, Denise E. Jackson*

Thrombosis and Vascular Diseases Laboratory, School of Health and Biomedical sciences, RMIT University, Bundoora, Melbourne, Australia



ARTICLE INFO

Keywords:

Platelets
ITIM
ITAM
Haemostasis
Integrin α IIB β 3

ABSTRACT

Background: The main role of platelets is to control haemostasis when there is a blood vessel injury in order to minimise blood loss at the injury site. Under normal circumstances, platelets flow freely within blood vessels as the endothelial cells provide a non-adhesion surface. Naturally, bioactive mediators are released from endothelial cells to prevent and control platelet activation. However, when the vascular endothelium is ruptured, the local concentration of nitric oxide and prostaglandin is diminished and receptors containing a sequence of amino acids known as, immunoreceptor tyrosine-based inhibition motifs (ITIMs), serve as natural inhibitors within platelets. The main role of ITIMs is to decrease immunoreceptor tyrosine-based activation motif (ITAM) signalling in platelets; however, some studies have revealed their novel role in integrin α IIB β 3 activation. This review highlights the main structural and functional features of immunoreceptors in platelets.

1. Introduction

Platelets are the smallest fragments in blood which are derived from megakaryocytes (MKs). The average lifespan of circulating platelets is generally accepted to be less than 10 days. During their lifespan, platelets decrease in size such that old platelets are smaller than the young platelets [1]. At the end of their lifespan platelets are destroyed by neutrophils and macrophages in the spleen and liver.

Platelets play an important role in physiological and pathological thrombosis. In addition, they play an important role in angiogenesis during adulthood and childhood [2,3]. Recently, platelets' contribution to the immune system has been widely investigated, and they are considered to contribute to innate immune responses, the inflammation response as well as to wound healing [4,5].

In the flowing blood, platelets are margined near the vessel wall to monitor blood vessel integrity, while red blood cells (RBCs) are concentrated in the centre of the blood stream. Under normal conditions, circulating platelets do not interact with endothelial cells, which provides a non-adhesive surface to platelets [1]. When endothelium is removed, platelets slow down at the site of injury and come in contact with the subendothelial matrix that contains both collagen and von Willebrand factor (vWF). Platelets then bind to vWF via the platelet receptor GPIb/IX/V complex (platelet tethering) [6]. Thereafter, platelets exposed to collagen will bind directly or indirectly to collagen receptors. Integrin $\alpha_2\beta_1$ binds to collagen and allows for further binding to collagen via GPVI which initiates transmembrane and intracellular signalling leading to platelet activation [7].

The signalling pathway by GPVI/FcR γ chain initiates with cross-linking of the glycine-proline-hydroxyproline (GPO) repeat motif in collagen with the immunoglobulin (Ig) domains of GPVI dimers. The intracytoplasmic FcR γ chain of the immunoreceptor tyrosine-based activation motif (ITAM) on the cytoplasmic portion of the FcR γ chain is then phosphorylated by the sarcoma (Src) family kinases (SFK) Fyn/Lyn [8]. Downstream signalling cascades lead to activation of phospholipase C γ 2 (PLC γ 2) and phosphoinositide-3 kinase (PI3K) which, in turn activate the fibrinogen receptor (integrin α IIB β 3) (inside-out signalling), and ultimately cause Ca²⁺ mobilisation, granule secretion and platelet aggregation [9].

Activated platelets undergo protein cytoskeleton reorganisation, resulting in extensive formation of spiny spheres with long filopodia extending from the plasma membrane [10]. This increases the surface area of the platelet, allowing more interaction with adjacent platelets and also provides greater surface for binding of the coagulation proteins of the secondary haemostatic mechanism. Platelets then release their granule contents via platelet exocytosis. Some released contents such as ADP and serotonin attract more platelets to the site of injured vessel wall and active them, while thromboxane A2 initiates vasoconstriction to minimise blood loss and activates surrounding platelets. ADP binding to its receptors such as P2Y₁ and P2Y₁₂ (ADP receptors) triggers signalling pathways that eventually converts the fibrinogen receptor (α IIB β 3) from the resting to an activated state [11–13]. Activation of integrin α IIB β 3 in the plasma but not on surface is essential for fibrinogen to bind and bridge adjacent platelets (aggregation). The two loops of plasma fibrinogen then binds to the extracellular domain of the

* Corresponding author at: Thrombosis and Vascular Diseases Laboratory, School of Health and Biomedical sciences, RMIT University, Melbourne, Australia.
E-mail address: denise.jackson@rmit.edu.au (D.E. Jackson).

activated integrin α Ib β 3 to form a huge meshwork that forms a platelet plug and arrests blood loss from the injured vessel.

The platelet plug will arrest blood flow but requires stabilisation by the secondary haemostatic process which results in a fibrin network throughout the platelet plug.

In the subendothelial space, the tissue factor bearing cells from a complex with factor VIIa which then activates factor X and factor IX [14]. A small amount of thrombin is then generated which can activate platelets and release factor VIII from vWF (initiation phase) [15]. Amplification of secondary haemostasis occurs upon platelet granule secretion and exposure of phosphatidyl serine on the platelet surface. The propagation phase of haemostasis results in massive thrombin generation which cleaves fibrinogen to fibrin and stabilises the platelet plug [16].

Critical to the platelet thrombosis and haemostasis processes is signalling provided by platelet immunoreceptors. With the increase in the number of immunoreceptors identified in platelets and their diverse functional roles in platelet activation or inhibition, we will review the recent literature that highlights and summarises structural and functional features of the following immunoreceptors: GPVI, Fc γ RIIa, CLEC-2, PECAM-1, CEACAM-1, CEACAM-2, G6b and TLT-1.

2. ITAM-containing receptors in platelets

The immunoreceptor tyrosine-based activation motifs (ITAMs) are important signalling motifs in immune cells and platelets. ITAMs have a unique structure which possess two identical signatures Yxx(I/L) interspersed by 6–12 amino acids [17]. There are three ITAM containing receptors: GPVI/Fc γ chain, Fc γ RIIa and the C-type lectin CLEC-2 expressed in human platelets, while Fc γ RIIa is not expressed in murine platelets.

During platelet activation, the integrin α Ib β 3 clusters and triggers the integrin associated Src family kinases (SFKs), to phosphorylate the ITAMs in the cytoplasmic domain of the receptor. Following the ITAM phosphorylation, ITAMs serve as a docking site for SH2-domain containing proteins, thus further downstream signalling pathways are propagated.

Platelet activation is regulated via ITAM and ITIM immunoreceptor families which have positive and negative regulation contribution toward integrin α Ib β 3 outside-in signalling [18].

2.1. GPVI

Glycoprotein (GP) VI is only expressed on MKs and platelets and it is the main collagen receptor [9]. It is a 55–63 kDa type 1 transmembrane glycoprotein that belongs to the immunoreceptor superfamily (IgSF). The extracellular N-terminal domain of GPVI has two immunoglobulin (Ig) like domains, followed by a highly O-glycosylated mucin-like region (Fig. 1). The 19 amino acid transmembrane domain is important for forming a salt bridge between GPVI Arg272 and Asp11 of Fc γ R chain which acts as co-receptor [19,20]. In addition, the cytoplasmic tail which is important for activating intracellular signalling pathway contains 51 amino acids including ITAM motifs.

GPVI and Fc γ R chain receptor form a synergistic signalling complex that mediates platelet activation by tyrosine kinase phosphorylation of the two Fc γ R cytoplasmic domains (ITAM). Moreover, murine knockout studies revealed the important role of Fc γ R for GPVI expression and signalling pathways. Fc γ R^{-/-}-derived platelets lack GPVI expression on their surface and have attenuated collagen-stimulated platelet activation and thrombus formation under flow conditions [21]. The signalling pathway by GPVI initiates with crosslinking the glycine-proline-hydroxyproline (GPO) repeat motif in the collagen with Ig domains of GPVI dimers [22]. Thereafter, the proline-rich domain in the cytoplasmic tail of GPVI provides a docking site for SFK members and Lyn, which are important in tyrosine phosphorylation of the two Fc γ R ITAMs. This initiates the downstream signalling events via Fc γ R chain

[23,24], which to ultimately activate phospholipase C γ 2 (PLC γ 2) and phosphoinositide-3 kinase (PI3K) that in turn induce integrin α Ib β 3 activation, Ca²⁺ mobilisation, granule secretion, and aggregation (Fig. 2) [25].

Most recent work have revealed a new important role of GPVI in thrombus growth and stability. In vivo thrombus studies showed that GPVI-deficient mice had a marked delay in occlusion or with constant embolisation [26,27]. Moreover, in vitro thrombus formation studies have showed that GPVI can bind to D-dimer and the D-domain site on fibrin but not fibrinogen to induce platelet spreading via GPVI activation [22,28]. Therefore, these findings suggest the novel role of fibrin-GPVI crosslinking in thrombus propagation and stability.

2.2. Fc γ RIIa

Fc γ RIIa is an important member of the immunoglobulin gene superfamily that is expressed on human platelets [29]. It is a 40 kDa transmembrane glycoprotein with two Ig-like domains and a cytoplasmic tail that contains two intrinsic ITAMs with a typical sequence Yxx(I/L)x(6–12)Yxx(I/L) (Fig. 1) [30].

Fc γ RIIa is an activating form of Fc receptor (FcR) which has low affinity to the Fc of IgG-containing immune complexes. Fc γ RIIa has been demonstrated to be a SFK target. Once cross-linked, the ITAMs become phosphorylated and create a docking site for the SH2 domain of tyrosine kinase Syk [31]. As a result of Syk activation, which is necessary for integrin-mediated outside-in signalling pathway this leads to activation of phospholipase C γ 2. Furthermore, intracellular Ca²⁺ concentration increases followed by granule secretion, integrin activation, platelet aggregation and then thrombus formation (Fig. 2) [32].

2.3. The C-type lectin CLEC-2 in platelets

C-type lectin-like receptor 2 (CLEC-2) is a 32 kDa type 2 transmembrane glycoprotein with an extracellular carbohydrate-like recognition domain (CRD-like) [33]. Unlike other ITAM containing receptors, CLEC-2 has only a single YxxL motif in its cytoplasmic domain that is known as atypical or hemITAM motif (Fig. 1) [34]. Historically, CLEC-2 was first discovered as the target receptor for the snake venom rhodocytin. Another ligand for CLEC-2 is the endogenous cellular podoplanin which presents on cancer cells and lymphatic endothelial cells [34].

CLEC-2 and GPVI roles in vascular processes have been studied and been found that they contribute in maintenance of vascular integrity, angiogenesis and lymphogenesis. CLEC-2 role also in haemostasis was unclear and has been reported as having no contribution or a slight contribution [35–37]. More recently, combined deletion of GPVI and CLEC-2 in mice demonstrated severely impaired haemostasis and a marked thrombus growth defect compared to single knockout mice [38]. Which is by far more than that in a single GPVI or CLEC-2 deleted mice.

CLEC-2 activation signalling pathway is similar to the GPVI or Fc γ RIIa signalling pathways [39,40]. Cross-linking of CLEC-2 with its ligand podoplanin or the snake venom rhodocytin initiates the single ITAM phosphorylation which becomes a docking site for Syk. Afterwards, Syk signalling pathway leads to the activation of PLC γ 2 which ultimately activates the fibrinogen receptor (α Ib β 3), Ca²⁺ mobilisation, granule secretion, and platelet aggregation (Fig. 2) [41].

3. ITIM-containing receptors in platelets

3.1. PECAM-1

Platelet endothelial cell adhesion molecule-1 (PECAM-1;CD31) is a 130-kDa member of the immunoglobulin superfamily (IgSF) that is expressed on the most of nonerythroid haematopoietic cells, including platelets and leukocytes [42,43]. PECAM-1 is also abundantly

ITAM and hemITAM containing receptors

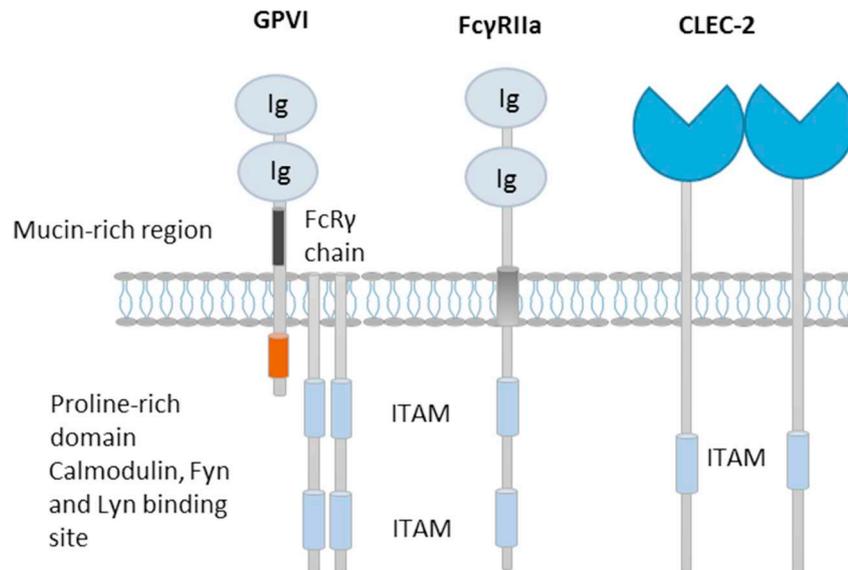


Fig. 1. Schematic diagram of ITAM containing receptors GPVI, FcγRIIIa and CLEC-2. The main structural features including the extracellular Ig like domains or a C terminal domain. The intracellular tail contains dual ITAMs in GPVI and FcγRIIIa but only a single ITAM in CLEC-2.

expressed at the intercellular junctions of endothelial cells [44]. PECAM-1 has a highly glycosylated extracellular domain that is composed of 6 immunoglobulin homology domains, a transmembrane domain, and a 118 amino acid cytoplasmic tail that mediates cell-cell interaction and cell signalling (Fig. 3) [45]. In humans, the localisation of the PECAM-1 gene has been determined as the long arm of chromosome 17q23, while it is on chromosome 6 in mice [46,47].

The function of PECAM-1 in intracellular junctions between endothelial cells is to mediate transmigration of leukocytes between the vasculature and subendothelial space [48]. In addition, it functions to maintain the cellular adhesive interactions to preserve the integrity of vascular endothelial cell junctions under inflammation and thrombotic stress conditions [49]. Moreover, PECAM-1 has been identified to mediate leukocyte migration through endothelial cells during diapedesis [50].

Previous studies have shown that, the cytoplasmic tail of PECAM-1 that conveys signalling functionality, contains an immunoreceptor tyrosine-based inhibitory motif (ITIM) and an immunoreceptor tyrosine-based motif (ITSM) [45,51–53]. Once platelets are activated, both ITIM and ITSM become tyrosine phosphorylated, thereby PECAM-1 becomes a docking site for the protein tyrosine phosphatases SHP-2 predominantly, but also SHP-1 [54]. These two protein tyrosine phosphatases initiate inhibitory effects by removing the phosphate group from the ITAM and thus dampening its signalling pathway (Fig. 2) [51,54].

The role of PECAM-1 in thrombus formation *in vitro* has been studied using type 1 collagen, and PECAM-1 deficient platelets display hyper-responsive platelet adhesion with larger and more stable thrombus [55]. This finding supports the concept that PECAM-1 acts as an inhibitory co-receptor in platelets to negatively regulate collagen-GPVI-mediated platelet responses. However, platelets isolated from PECAM-1 deficient mice show weak adhesion and restricted

cytoskeleton recognition on a fibrinogen-coated surface compared with wild type-derived platelets [56]. These findings raises the potential that PECAM-1 is also a positive regulator of the integrin α IIb β 3 mediated pathway [56]. However, the underlying mechanism of PECAM-1 into maintaining the normal integrin α IIb β 3-mediated platelet function is still not well understood.

The ITSM signalling pathway in leucocytes occurs by binding SLAM adaptor protein (SAP), SHP-2 and EWS-activated transcript-2 (EAT-2) to phosphorylated (but not required) ITSM that can then induce inhibitory signalling pathway that then binds and activates Fyn [57–59]. In platelets, CD150 and CD84 have been identified as ITSM containing receptors and have SAP and EAT-2 but they are not required to tyrosine phosphorylate CD150 and CD84 which is different compared to lymphocytes [60,61].

3.2. CEACAM-1

Carcinoembryonic antigen-related cell adhesion molecule 1 (CEACAM-1 or CD66a) is a transmembrane receptor which belongs to the CEA family of adhesion molecules [62]. Previously, CEACAM-1 was named biliary glycoprotein (BGP) as it was first isolated from human bile [63]. Moreover, CEACAM-1 is widely expressed on immune, endothelial, and blood cells. Interestingly, CEACAM-1 is the only isoform of CEACAM family that is expressed in both rodents and humans [64].

The cytoplasmic domain of CEACAM-1 has an ITIM and ITSM (Fig. 3) that mediates cell-cell interactions and cell signalling [45]. During platelet activation, ITIMs become tyrosine phosphorylated and become docking site for SH2 domain-containing protein tyrosine phosphatases (SHP-1 and SHP-2). SHP-1 and SHP-2 are recruited to inhibit cell signalling by reversing the tyrosine phosphorylation process that its brought about by the tyrosine kinases action (Fig. 2) [65]. On the other hand, the extracellular domain of CEACAM-1 contains one N-

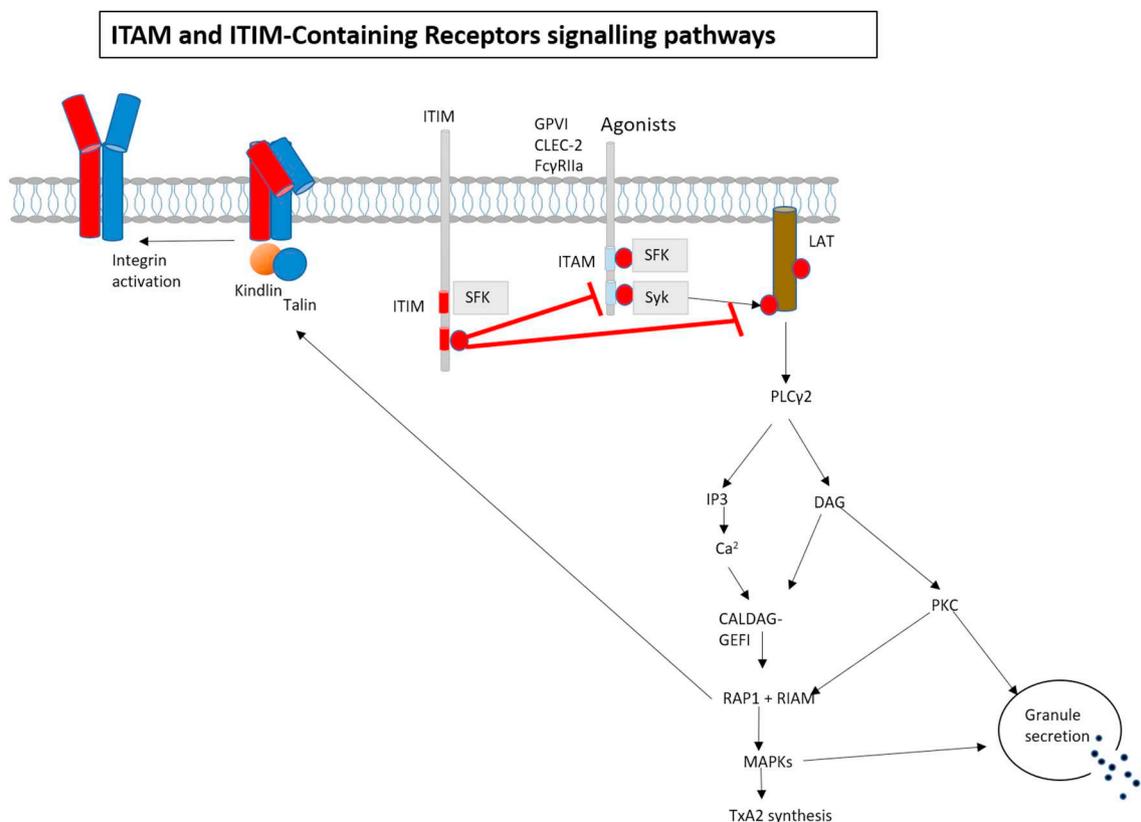


Fig. 2. Schematic diagram of ITAM containing receptors inside-out and outside-in signalling pathways. ITAM becomes phosphorylated and induces PLC γ 2 signalling pathway which ultimately results in (1) intracellular calcium elevation, (2) Granule secretion, (3) TxA2 synthesis and (4) integrin α IIb β 3 activation. ITIM-bound SHP-2 dampens ITAM signalling pathway by dephosphorylating and deactivating the key components of ITAM signalling pathway.

terminal immunoglobulin variable-region-like (IgV-like) domain that mediates homophilic adhesion, followed by three immunoglobulin constant-region-type-2-like (IgC2-like) domains [66,67].

The contribution of ITAM-mediated signalling that is initiated by ITAM-bearing GPVI/Fc γ chain, and its regulation by CEACAM-1 ITIMs has been well examined in response to type 1 collagen exposure, however, much less understood about the molecular effects that are produced by platelet exposure to other agonists [64]. This raises the question whether CEACAM-1 has a similar negative regulation effect on ITAM-bearing GPVI/Fc γ chain that might be initiated by GPIb/IX/V complex signalling or G-protein coupled signalling pathways.

CEACAM-1 roles in vascular cell types have been investigated. It plays a crucial role in maintaining the strength and vascular integrity [68]. Thus, CEACAM-1 knockout mice have been investigated and increased incidence of blood vessel leakage has been reported [69]. Moreover, CEACAM-1 role in carcinogenesis have been studied and been found that CEACAM-1 has a tumour-suppression function [70]. The tissue expression of CEACAM-1 in breast cancer was significantly reduced in the cancer area compared to normal tissue. Coupled with the inhibition of cancer proliferation by ITIM mediated signalling which shows the suppressive effects of CEACAM-1 on tumour development this suggests that a strategy to restore CEACAM1 expression may be helpful for the treatment of breast cancer [70]. Furthermore, healthy lung tissues lack CEACAM-1 expression, while it has been reported to be expressed on lung tumour cells [71,72]. These findings are questioning the relation of CEACAM-1 expression and function in lung cancer and tumour progression.

3.3. CEACAM-2

CEACAM-2 is a type I transmembrane receptor that belongs to the CEACAM superfamily. Similarly, CEACAM-1,2 share some overall

structure in terms of Ig domain in the extracellular part and with an ITIM and an ITSM in the cytoplasmic domain (Fig. 3). However, CEACAM-1 has four extracellular Ig-domains, while CEACAM-2 has only two Ig domains. CEACAM-2 is expressed in mouse platelets, but CEACAM-1 is predominant in human and mouse platelets [45]. Similar to CEACAM-1, the cytoplasmic tail of CEACAM-2 has a single ITIM and an ITSM that mediates cell-cell interactions and cell signalling [45].

3.4. G6b

G6b gene which codes for G6b-B is located in the major histocompatibility complex (MHC) region of chromosome 17 [73]. G6b-B is highly expressed on both mature megakaryocytes and platelets [74,75].

G6b-B consists of 242 amino acids consisting of an extracellular N-terminal domain of G6b-B that has a single extracellular variable Ig like (IgV) domain, and a single transmembrane domain. G6b-B cytoplasmic tail has an ITIM (Y194) and an ITSM (Y220) in its cytoplasmic domain (Fig. 3).

G6b-B works by inhibition of ITAM signalling pathway either from GPVI-Fc γ -chain complex, the hemITAM containing podoplanin activation receptor CLEC-2 or GPVI and ADP induced platelet aggregation after antibody-mediated crosslinking (Fig. 2) [76].

The physiological function of G6b-B was thoroughly investigated by using G6b-B knockout mouse models [77]. Interestingly, G6b-B knockout mice had a bleeding disorder due to impairment in platelet production and function [77]. Also, they exhibited significant macrothrombocytopenia with a functional defect, GPVI and GPIb α shedding. As a result, G6b-B was proposed to be an important megakaryocyte function and platelet formation regulator [77].

ITIM containing receptors PECAM-1, CEACAM-1, CEACAM-2, G6b-B and TLT-1.

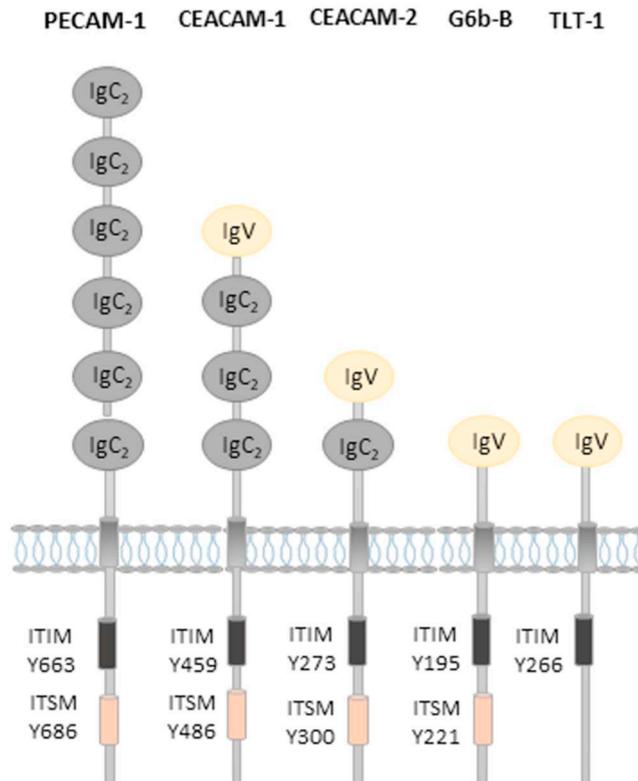


Fig. 3. Schematic diagram of ITIM containing receptors PECAM-1, CEACAM-1, CEACAM-2, G6b-B and TLT-1. The main structural features including the extracellular constant-region-type-2-like (IgC2-like) domains and/or variable Ig like (IgV) domains. The intracellular tail contains ITIM and ITSM or a single ITIM in TLT-1.

3.5. TLT-1

Triggering receptor expressed on myeloid cells (TREM) is an immunoreceptor that is involved in cell activation within the immune system. TREM contains a single V-type extracellular immunoglobulin domain, a short cytoplasmic tail and a trans-membrane domain (Fig. 3) [78]. TERM-like Transcript 1 (TLT-1) is a type 1 single IgV-containing protein that is localised exclusively in megakaryocytes and platelet α -granules [78]. TLT-1 is redistributed on the platelet surface during thrombin-induced platelet activation [79]. Moreover, two TLT-1 isoforms are expressed, the first isoform has a cytoplasmic tail with a potential ITIM, and the second isoform is characterised by a cytoplasmic tail lacking an ITIM [80].

TLT-1 is the only TREM family member that contains an ITIM in its cytoplasmic domain which acts as a docking site for the SHP family proteins. SHP-2, SHP-1 and SHIP1 are recruited to the ITIM (at Y281) to negate the ITAM-mediated signalling (Fig. 2) [81,82].

Pharmacologically inhibited TERM-1 using LR12 has been found to reduce platelet activation induced by ADP, thrombin and collagen in human platelets [83]. In addition, Trem-1 $-/-$ mice have reduced platelet aggregation to ADP, collagen and thrombin. Trem-1 inhibition reduces thrombus formation in a carotid artery model and protects mice from pulmonary embolism [83].

4. Conclusion

Megakaryocytes and platelets are very complex and express a number of immunoreceptors that participate in haemostasis and thrombosis control. Knockout mice models have been invaluable for

determining the role of ITAM and ITIM bearing receptors in the context of platelet haemostasis and thrombosis. Ig-ITIM bearing receptor knockout cleat a unique phenotype indicating they are redundant collectively, and the combined Ig-ITIM bearing receptors is only to play a potential role in regulating platelet-collagen interaction as a natural platelet inhibitor in vivo. Furthermore, questions have been raised about whether ITIM-containing receptors can regulate other signalling pathways other than GPVI signalling pathway. One unanticipated finding was that, ITIM-containing receptors have a positive regulatory effect on integrin α IIb β 3 activation, but the underlying mechanisms have not yet been clearly elucidated.

Acknowledgements

Shaqra University, Saudi Arabia.

Conflict of interests

Authors declare that there is no conflict of interest.

References

- [1] K. Jurk, B.E. Kehrel, Platelets: physiology and biochemistry, *Semin. Thromb. Hemost.* 31 (4) (2005) 381–392.
- [2] E.M. Battinelli, B.A. Markens, J.E. Italiano Jr., Release of angiogenesis regulatory proteins from platelet alpha granules: modulation of physiologic and pathologic angiogenesis, *Blood* 118 (5) (2011) 1359–1369.
- [3] G. Lakka Klement, E. Shai, D. Varon, Chapter 24 - the role of platelets in angiogenesis A2, in: Alan D. Michelson (Ed.), *Platelets*, Third edition, Academic Press, 2013, pp. 487–502.
- [4] R.A. Ali, L.M. Wuescher, R.G. Worth, *Platelets: essential components of the immune*

- system, *Curr. Trends Immunol.* 16 (2015) 65–78.
- [5] C.N. Morrell, A.A. Aggrey, L.M. Chapman, K.L. Modjeski, Emerging roles for platelets as immune and inflammatory cells, *Blood* 123 (18) (2014) 2759–2767.
 - [6] Y.F. Zhou, E.T. Eng, J. Zhu, C. Lu, T. Walz, T.A. Springer, Sequence and structure relationships within von Willebrand factor, *Blood* 120 (2) (2012) 449–458.
 - [7] B.P. Nuytens, T. Thijs, H. Deckmyn, K. Broos, Platelet adhesion to collagen, *Thromb. Res.* 127 (Suppl. 2) (2011) S26–S29.
 - [8] N.S. Poulter, A.Y. Pollitt, D.M. Owen, E.E. Gardiner, R.K. Andrews, H. Shimizu, et al., Clustering of glycoprotein VI (GPVI) dimers upon adhesion to collagen as a mechanism to regulate GPVI signaling in platelets, *J. Thromb. Haemost.* 15 (3) (2017) 549–564.
 - [9] B. Nieswandt, S.P. Watson, Platelet-collagen interaction: is GPVI the central receptor? *Blood* 102 (2) (2003) 449–461.
 - [10] D. Cerecedo, Platelet cytoskeleton and its hemostatic role, *Blood Coagul. Fibrinolysis* 24 (8) (2013) 798–808.
 - [11] Z. Li, M.K. Delaney, K.A. O'Brien, X. Du, Signaling during platelet adhesion and activation, *Arterioscler. Thromb. Vasc. Biol.* 30 (12) (2010) 2341–2349.
 - [12] C.V. Denis, D.D. Wagner, Platelet adhesion receptors and their ligands in mouse models of thrombosis, *Arterioscler. Thromb. Vasc. Biol.* 27 (4) (2007) 728–739.
 - [13] T. Gremmel, I.B. Yanachkov, M.I. Yanachkova, G.E. Wright, J. Wider, V.V. Undyala, et al., Synergistic inhibition of both P2Y1 and P2Y12 adenosine diphosphate receptors as novel approach to rapidly attenuate platelet-mediated thrombosis, *Arterioscler. Thromb. Vasc. Biol.* 36 (3) (2016) 501–509 (ATVBAHA. 115.306885).
 - [14] D.D. Monkovic, P.B. Tracy, Activation of human factor V by factor Xa and thrombin, *Biochemistry* 29 (5) (1990) 1118–1128.
 - [15] M. Diaz-Ricart, E. Estebanell, M. Lozano, J. Aznar-Salatti, J.G. White, A. Ordinas, et al., Thrombin facilitates primary platelet adhesion onto vascular surfaces in the absence of plasma adhesive proteins: studies under flow conditions, *Haematologica* 85 (3) (2000) 280–288.
 - [16] J.A. Oliver, D.M. Monroe, H.R. Roberts, M. Hoffman, Thrombin activates factor XI on activated platelets in the absence of factor XII, *Arterioscler. Thromb. Vasc. Biol.* 19 (1) (1999) 170–177.
 - [17] Y. Boulaftali, P.R. Hess, M.L. Kahn, W. Bergmeier, Platelet immunoreceptor tyrosine-based activation motif (ITAM) signaling and vascular integrity, *Circ. Res.* 114 (7) (2014) 1174–1184.
 - [18] H. Zhi, L. Rauova, V. Hayes, C. Gao, B. Boylan, D.K. Newman, et al., Cooperative integrin/ITAM signaling in platelets enhances thrombus formation in vitro and in vivo, *Blood* 121 (10) (2013) 1858–1867.
 - [19] J.M. Clemetson, J. Polgar, E. Magnenat, T.N. Wells, K.J. Clemetson, The platelet collagen receptor glycoprotein VI is a member of the immunoglobulin superfamily closely related to Fc α 1R and the natural killer receptors, *J. Biol. Chem.* 274 (41) (1999) 29019–29024.
 - [20] M. Jandrot-Perrus, S. Busfield, A.H. Lagrue, X. Xiong, N. Debili, T. Chickering, et al., Cloning, characterization, and functional studies of human and mouse glycoprotein VI: a platelet-specific collagen receptor from the immunoglobulin superfamily, *Blood* 96 (5) (2000) 1798–1807.
 - [21] B. Nieswandt, W. Bergmeier, V. Schulte, K. Rackebbrandt, J.E. Gessner, H. Zirngibl, Expression and function of the mouse collagen receptor glycoprotein VI is strictly dependent on its association with the Fc γ chain, *J. Biol. Chem.* 275 (31) (2000) 23998–24002.
 - [22] N.S. Poulter, A.Y. Pollitt, D.M. Owen, E.E. Gardiner, R.K. Andrews, H. Shimizu, et al., Clustering of glycoprotein VI (GPVI) dimers upon adhesion to collagen as a mechanism to regulate GPVI signaling in platelets, *J. Thromb. Haemost.* 15 (3) (2017) 549–564.
 - [23] K. Suzuki-Inoue, D. Tulasne, Y. Shen, T. Bori-Sanz, O. Inoue, S.M. Jung, et al., Association of Fyn and Lyn with the proline-rich domain of glycoprotein VI regulates intracellular signaling, *J. Biochem.* 277 (24) (2002) 21561–21566.
 - [24] L.S. Quek, J.M. Pasquet, I. Hers, R. Cornell, G. Knight, M. Barnes, et al., Fyn and Lyn phosphorylate the Fc receptor gamma chain downstream of glycoprotein VI in murine platelets, and Lyn regulates a novel feedback pathway, *Blood* 96 (13) (2000) 4246–4253.
 - [25] S.M. Jung, M. Moroi, K. Soejima, T. Nakagaki, Y. Miura, M.C. Berndt, et al., Constitutive dimerization of glycoprotein VI (GPVI) in resting platelets is essential for binding to collagen and activation in flowing blood, *J. Biol. Chem.* 287 (35) (2012) 30000–30013.
 - [26] O.M. Alshehri, C.E. Hughes, S. Montague, S.K. Watson, J. Frampton, M. Bender, et al., Fibrin activates GPVI in human and mouse platelets, *Blood* 126 (13) (2015) 1601–1608.
 - [27] E. Mammadova-Bach, V. Ollivier, S. Loyau, M. Schaff, B. Dumont, R. Favier, et al., Platelet glycoprotein VI binds to polymerized fibrin and promotes thrombin generation, *Blood* 126 (5) (2015) 683–691.
 - [28] M.-B. Onselae, A.T. Hardy, C. Wilson, X. Sanchez, A.K. Babar, J.L.C. Miller, et al., Fibrin and D-dimer bind to monomeric GPVI, *Blood Adv.* 1 (19) (2017) 1495–1504.
 - [29] R.H. Lee, W. Bergmeier, Platelet immunoreceptor tyrosine-based activation motif (ITAM) and hemITAM signaling and vascular integrity in inflammation and development, *J. Thromb. Haemost.* 14 (4) (2016) 645–654.
 - [30] H. Zhi, J. Dai, J. Liu, J. Zhu, D.K. Newman, C. Gao, et al., Platelet activation and thrombus formation over IgG immune complexes requires integrin α IIb β 3 and Lyn kinase, *PLoS One* 10 (8) (2015) e0135738.
 - [31] G.W. Chacko, A.M. Duchemin, K.M. Coggeshall, J.M. Osborne, J.T. Brandt, C.L. Anderson, Clustering of the platelet Fc gamma receptor induces noncovalent association with the tyrosine kinase p72syk, *J. Biol. Chem.* 269 (51) (1994) 32435–32440.
 - [32] M. Arman, K. Krauel, Human platelet IgG Fc receptor Fc γ RIIA in immunity and thrombosis, *J. Thromb. Haemost.* 13 (6) (2015) 893–908.
 - [33] M. Colonna, J. Samaridis, L. Angman, Molecular characterization of two novel C-type lectin-like receptors, one of which is selectively expressed in human dendritic cells, *Eur. J. Immunol.* 30 (2) (2000) 697–704.
 - [34] K. Suzuki-Inoue, G.L. Fuller, A. Garcia, J.A. Eble, S. Pohlmann, O. Inoue, et al., A novel Syk-dependent mechanism of platelet activation by the C-type lectin receptor CLEC-2, *Blood* 107 (2) (2006) 542–549.
 - [35] F. May, I. Hagedorn, I. Pleines, M. Bender, T. Vogtle, J. Eble, et al., CLEC-2 is an essential platelet-activating receptor in hemostasis and thrombosis, *Blood* 114 (16) (2009) 3464–3472.
 - [36] K. Suzuki-Inoue, Essential *in vivo* roles of the platelet activation receptor CLEC-2 in tumour metastasis, lymphangiogenesis and thrombus formation, *J. Biochem.* 150 (2) (2011) 127–132.
 - [37] C.E. Hughes, L. Navarro-Nunez, B.A. Finney, D. Mourao-Sa, A.Y. Pollitt, S.P. Watson, CLEC-2 is not required for platelet aggregation at arteriolar shear, *J. Thromb. Haemost.* 8 (10) (2010) 2328–2332.
 - [38] M. Bender, F. May, V. Lorenz, I. Thielmann, I. Hagedorn, B.A. Finney, et al., Combined *in vivo* depletion of glycoprotein VI and C-type lectin-like receptor 2 severely compromises hemostasis and abrogates arterial thrombosis in mice, *Arterioscler. Thromb. Vasc. Biol.* 33 (5) (2013) 926–934.
 - [39] C.E. Hughes, A.Y. Pollitt, J. Mori, J.A. Eble, M.G. Tomlinson, J.H. Hartwig, et al., CLEC-2 activates Syk through dimerization, *Blood* 115 (14) (2010) 2947–2955.
 - [40] S.P. Watson, J.M. Herbert, A.Y. Pollitt, GPVI and CLEC-2 in hemostasis and vascular integrity, *J. Thromb. Haemost.* 8 (7) (2010) 1456–1467.
 - [41] K. Suzuki-Inoue, Y. Kato, O. Inoue, M.K. Kaneko, K. Mishima, Y. Yatomi, et al., Involvement of the snake toxin receptor CLEC-2, in podoplanin-mediated platelet activation, by cancer cells, *J. Biochem.* 282 (36) (2007) 25993–26001.
 - [42] W.A. Muller, C.M. Ratti, S.L. McDonnell, Z.A. Cohn, A human endothelial cell-restricted, externally disposed plasmalemmal protein enriched in intercellular junctions, *J. Exp. Med.* 170 (2) (1989) 399–414.
 - [43] S.M. Albelda, P.D. Oliver, L.H. Romer, C.A. Buck, EndoCAM: a novel endothelial cell-cell adhesion molecule, *J. Cell Biol.* 110 (4) (1990) 1227–1237.
 - [44] C. Paddock, D. Zhou, P. Lertkiatmongkol, P.J. Newman, J. Zhu, Structural basis for PECAM-1 homophilic binding, *Blood* 127 (8) (2016) 1052–1061.
 - [45] C.H. Coxon, M.J. Geer, Y.A. Senis, ITIM receptors: more than just inhibitors of platelet activation, *Blood* 129 (26) (2017) 3407–3418.
 - [46] R.J. Gumina, N.E. Kirschbaum, P.N. Rao, P. van Tuinen, P.J. Newman, The human PECAM1 gene maps to 17q23, *Genomics* 34 (2) (1996) 229–232.
 - [47] Y. Xie, W.A. Muller, Fluorescence *in situ* hybridization mapping of the mouse platelet endothelial cell adhesion molecule-1 (PECAM1) to mouse chromosome 6, region F3-G1, *Genomics* 37 (2) (1996) 226–228.
 - [48] P.J. Newman, D.K. Newman, Signal transduction pathways mediated by PECAM-1: new roles for an old molecule in platelet and vascular cell biology, *Arterioscler. Thromb. Vasc. Biol.* 23 (6) (2003) 953–964.
 - [49] J.R. Privratsky, C.M. Paddock, O. Florey, D.K. Newman, W.A. Muller, P.J. Newman, Relative contribution of PECAM-1 adhesion and signaling to the maintenance of vascular integrity, *J. Cell Sci.* 124 (Pt 9) (2011) 1477–1485.
 - [50] W.A. Muller, The role of PECAM-1 (CD31) in leukocyte emigration: studies *in vitro* and *in vivo*, *J. Leukoc. Biol.* 57 (4) (1995) 523–528.
 - [51] V. Rathore, M.A. Stapleton, C.A. Hillery, R.R. Montgomery, T.C. Nichols, E.P. Merricks, et al., PECAM-1 negatively regulates GPIIb/IIIa signaling in murine platelets, *Blood* 102 (10) (2003) 3658–3664.
 - [52] B.E. Tourdot, M.K. Brenner, K.C. Keough, T. Holyst, P.J. Newman, D.K. Newman, Immunoreceptor Tyrosine-based Inhibitory Motif (ITIM)-mediated inhibitory signaling is regulated by sequential phosphorylation mediated by distinct nonreceptor tyrosine kinases: a case study involving PECAM-1, *Biochemistry* 52 (15) (2013) 2597–2608.
 - [53] C. Paddock, B.L. Lytle, F.C. Peterson, T. Holyst, P.J. Newman, B.F. Volkman, et al., Residues within a lipid-associated segment of the PECAM-1 cytoplasmic domain are susceptible to inducible, sequential phosphorylation, *Blood* 117 (22) (2011) 6012–6023.
 - [54] M. Cicmil, J.M. Thomas, M. Leduc, C. Bon, J.M. Gibbins, Platelet endothelial cell adhesion molecule-1 signaling inhibits the activation of human platelets, *Blood* 99 (1) (2002) 137–144.
 - [55] K.L. Jones, S.C. Hughan, S.M. Dopheide, R.W. Farndale, S.P. Jackson, D.E. Jackson, Platelet endothelial cell adhesion molecule-1 is a negative regulator of platelet-collagen interactions, *Blood* 98 (5) (2001) 1456–1463.
 - [56] J.L. Wee, D.E. Jackson, The Ig-ITIM superfamily member PECAM-1 regulates the "outside-in" signaling properties of integrin α IIb β 3 in platelets, *Blood* 106 (12) (2005) 3816–3823.
 - [57] M.L. Vilar, M.S. Frutuoso, S.M. Arruda, D.M. Lima, C.S. Bezerra, M.M. Pompeu, The role of the SLAM-SAP signaling pathway in the modulation of CD4+ T cell responses, *Braz. J. Med. Biol. Res.* 44 (4) (2011) 276–282.
 - [58] S. Latour, R. Roncagalli, R. Chen, M. Bakinowski, X. Shi, P.L. Schwartzberg, et al., Binding of SAP SH2 domain to FynT SH3 domain reveals a novel mechanism of receptor signalling in immune regulation, *Nat. Cell Biol.* 5 (2) (2003) 149–154.
 - [59] M. Simarro, A. Lanyi, D. Howie, F. Poy, J. Bruggeman, M. Choi, et al., SAP increases FynT kinase activity and is required for phosphorylation of SLAM and Ly9, *Int. Immunol.* 16 (5) (2004) 727–736.
 - [60] N. Nanda, P. Andre, M. Bao, K. Clauser, F. Deguzman, D. Howie, et al., Platelet aggregation induces platelet aggregate stability via SLAM family receptor signaling, *Blood* 106 (9) (2005) 3028–3034.
 - [61] S. Hofmann, A. Braun, R. Pozgaj, M. Morowski, T. Vogtle, B. Nieswandt, Mice lacking the SLAM family member CD84 display unaltered platelet function in hemostasis and thrombosis, *PLoS One* 9 (12) (2014) e115306.
 - [62] Y. Zhuo, J.Y. Yang, K.W. Moremen, J.H. Prestegard, Glycosylation alters dimerization properties of a cell-surface signaling protein, *Carbohydrate Antigen-related Cell Adhesion Molecule 1 (CEACAM1)*, *J. Biol. Chem.* 291 (38) (2016)

- 20085–20095.
- [63] T. Svenberg, Carcinoembryonic antigen-like substances of human bile. Isolation and partial characterization, *Int. J. Cancer* 17 (5) (1976) 588–596.
- [64] C. Wong, Y. Liu, J. Yip, R. Chand, J.L. Wee, L. Oates, et al., CEACAM1 negatively regulates platelet-collagen interactions and thrombus growth in vitro and in vivo, *Blood* 113 (8) (2009) 1818–1828.
- [65] M. Huber, L. Izzi, P. Grondin, C. Houde, T. Kunath, A. Veillette, et al., The carboxyl-terminal region of biliary glycoprotein controls its tyrosine phosphorylation and association with protein-tyrosine phosphatases SHP-1 and SHP-2 in epithelial cells, *J. Biol. Chem.* 274 (1) (1999) 335–344.
- [66] Y.H. Huang, C. Zhu, Y. Kondo, A.C. Anderson, A. Gandhi, A. Russell, et al., CEACAM1 regulates TIM-3-mediated tolerance and exhaustion, *Nature* 517 (7534) (2015) 386–390.
- [67] S.M. Watt, A.M. Teixeira, G.Q. Zhou, R. Doyonnas, Y. Zhang, F. Grunert, et al., Homophilic adhesion of human CEACAM1 involves N-terminal domain interactions: structural analysis of the binding site, *Blood* 98 (5) (2001) 1469–1479.
- [68] A.K. Horst, W.D. Ito, J. Dabelstein, U. Schumacher, H. Sander, C. Turbide, et al., Carcinoembryonic antigen-related cell adhesion molecule 1 modulates vascular remodeling in vitro and in vivo, *J. Clin. Investig.* 116 (6) (2006) 1596–1605.
- [69] A.L. Nouvion, M. Oubaha, S. Leblanc, E.C. Davis, H. Jastrow, R. Kammerer, et al., CEACAM1: a key regulator of vascular permeability, *J. Cell Sci.* 123 (Pt 24) (2010) 4221–4230.
- [70] B.B. Singer, E. Klaile, I. Scheffrahn, M.M. Muller, R. Kammerer, W. Reutter, et al., CEACAM1 (CD66a) mediates delay of spontaneous and Fas ligand-induced apoptosis in granulocytes, *Eur. J. Immunol.* 35 (6) (2005) 1949–1959.
- [71] W. Sienel, S. Dango, U. Woelfle, A. Morresi-Hauf, C. Wagener, J. Brummer, et al., Elevated expression of carcinoembryonic antigen-related cell adhesion molecule 1 promotes progression of non-small cell lung cancer, *Clin. Cancer Res.* 9 (6) (2003) 2260–2266.
- [72] I. Thom, O. Schult-Kronefeld, I. Burkholder, G. Schuch, B. Andritzky, H. Kastendieck, et al., Expression of CEACAM-1 in pulmonary adenocarcinomas and their metastases, *Anticancer Res.* 29 (1) (2009) 249–254.
- [73] E.C. de Vet, B. Aguado, R.D. Campbell, G6b, a novel immunoglobulin superfamily member encoded in the human major histocompatibility complex, interacts with SHP-1 and SHP-2, *J. Biol. Chem.* 276 (45) (2001) 42070–42076.
- [74] C.H. Coxon, A.J. Sadler, J. Huo, R.D. Campbell, An investigation of hierarchical protein recruitment to the inhibitory platelet receptor, G6B-b, *PLoS One* 7 (11) (2012) e49543.
- [75] E.C. de Vet, S.A. Newland, P.A. Lyons, B. Aguado, R.D. Campbell, The cell surface receptor G6b, a member of the immunoglobulin superfamily, binds heparin, *FEBS Lett.* 579 (11) (2005) 2355–2358.
- [76] S.A. Newland, I.C. Macaulay, A.R. Floto, E.C. de Vet, W.H. Ouwehand, N.A. Watkins, et al., The novel inhibitory receptor G6B is expressed on the surface of platelets and attenuates platelet function in vitro, *Blood* 109 (11) (2007) 4806–4809.
- [77] A. Mazharian, Y.J. Wang, J. Mori, D. Bem, B. Finney, S. Heising, et al., Mice lacking the ITIM-containing receptor G6b-B exhibit macrothrombocytopenia and aberrant platelet function, *Sci. Signal.* 5 (248) (2012) ra78.
- [78] A.V. Washington, L. Quigley, D.W. McVicar, Initial characterization of TREM-like transcript (TLT)-1: a putative inhibitory receptor within the TREM cluster, *Blood* 100 (10) (2002) 3822–3824.
- [79] A.V. Washington, R.L. Schubert, L. Quigley, T. Disipio, R. Feltz, E.H. Cho, et al., A TREM family member, TLT-1, is found exclusively in the alpha-granules of megakaryocytes and platelets, *Blood* 104 (4) (2004) 1042–1047.
- [80] A.D. Barrow, E. Astoul, A. Floto, G. Brooke, I.A. Relou, N.S. Jennings, et al., Cutting edge: TREM-like transcript-1, a platelet immunoreceptor tyrosine-based inhibition motif encoding costimulatory immunoreceptor that enhances, rather than inhibits, calcium signaling via SHP-2, *J. Immunol.* 172 (10) (2004) 5838–5842.
- [81] A.D. Barrow, J. Trowsdale, You say ITAM and I say ITIM, let's call the whole thing off: the ambiguity of immunoreceptor signalling, *Eur. J. Immunol.* 36 (7) (2006) 1646–1653.
- [82] S.H. Yoon, Y.D. Lee, J. Ha, Y. Lee, H.H. Kim, TLT-1s, alternative transcripts of triggering receptor expressed on myeloid cell-like transcript-1 (TLM-1), inhibits the triggering receptor expressed on myeloid cell-2 (TREM-2)-mediated signaling pathway during osteoclastogenesis, *J. Biol. Chem.* 287 (35) (2012) 29620–29626.
- [83] L. Jolly, J. Lemarie, K. Carrasco, B. Popovic, M. Derive, A. Boufenzler, et al., Triggering receptor expressed on myeloid cells-1: a new player in platelet aggregation, *Thromb. Haemost.* 117 (9) (2017) 1772–1781.