

REVIEW ARTICLE

A Review of Prostate-Specific Membrane Antigen (PSMA) Positron Emission Tomography (PET) in Renal Cell Carcinoma (RCC)

Thomas Ahn^{1,2}, Matthew J. Roberts^{1,2,3}, Aous Abduljabar², Andre Joshi^{4,5,6}, Marlon Perera^{3,7}, Handoo Rhee^{2,3,5}, Simon Wood^{3,6}, Ian Vela^{2,3,8}

¹Department of Urology, Greenslopes Private Hospital, Brisbane, Queensland, Australia

²Faculty of Medicine, The University of Queensland, Brisbane, Queensland, Australia

³Department of Urology, Princess Alexandra Hospital, Brisbane, Queensland, Australia

⁴Department of Urology, Townsville Hospital, Brisbane, Queensland, Australia

⁵Centre for Clinical Research, The University of Queensland, Brisbane, Queensland, Australia

⁶Centre for Kidney Disease Research, Translational Research Institute, Brisbane, Australia

⁷Department of Surgery, Austin Health, The University of Melbourne, Parkville, Victoria, Australia

⁸Australian Prostate Cancer Research Center Queensland, Institute of Health and Biomedical Innovation, Queensland University of Technology, Brisbane, Queensland, Australia

Abstract

Metastatic renal cell carcinoma (mRCC) is a disease that portends poor prognosis despite an increasing number of novel systemic treatment options including new targeted therapies and immunotherapy. Ablative intervention directed at oligometastatic RCC has demonstrated survival benefit. Consequently, developing techniques for improved staging of mRCC on contemporary imaging modalities including X-ray computed tomography (CT), magnetic resonance imaging (MRI) and/or bone scan (BS) is a clinical priority. This is relevant for metastatic deposits too small to characterize or lymph nodes within physiological normality. Prostate-specific membrane antigen (PSMA) is a type II transmembrane glycoprotein highly expressed on prostate cancer epithelial cells. Recently, small molecules targeting the PSMA receptor, linked to radioactive isotopes have been developed for use with positron emission tomography (PET). Despite its nomenclature, PSMA has also been found to be expressed in the neovasculature of non-prostate cancers such as renal cell carcinoma (RCC) and hence PSMA PET/CT imaging has been proposed as an alternative staging modality. Preliminary small studies involving the use of PSMA PET/CT imaging in mRCC have been encouraging with evidence of improved staging sensitivity which has directly led to change in management in some cases. Given these early encouraging reports, we performed a comprehensive narrative review on the available evidence, including the scientific basis for PSMA expression in RCC, the role of PSMA PET/CT imaging with potential clinical implications in mRCC, its limitations and future opportunities.

Key words: Positron emission tomography, Prostate-specific membrane antigen, Gallium, Renal cell carcinoma, Metastasis

Epidemiology and Issues with Renal Cell Carcinoma

Kidney cancer is the ninth and 14th most common cancer worldwide in men and women respectively, of which renal cell carcinoma (RCC) makes up greater than 90 % of these [1]. Approximately 20–30 % of patients with RCC present with metastatic disease [2, 3] whilst the incidence of post-operative RCC metastasis is much lower at less than 7 % at median follow-up between 1.5 to 5 years [4–6]. Metastatic RCC exhibits poor prognosis with a 5-year survival rate between 10 and 20 % and overall median survival of less than 1 year after diagnosis [7–9].

Staging and precise localization of tumour dissemination determines patient management and prognosis and hence prompt and accurate diagnosis is desirable. Traditionally, computed tomography (CT) with intravenous contrast agent has been the mainstay imaging modality in RCC staging. This is due to its widespread availability; however, definitive detection of small metastatic foci remains difficult [10]. Improved identification of metastases has significant clinical diagnostic and treatment relevance, especially in oligometastatic disease with the potential for definitive local therapy and subsequent better survival and prognosis [11].

Molecular Data Relating PSMA to RCC

Prostate-specific membrane antigen (PSMA) is a type II transmembrane glycoprotein. It has a unique three-part structure: an 18-amino-acid internal portion, a 25-amino-acid transmembrane portion and a 707-amino-acid external portion [12]. PSMA is encoded by a gene (Folate hydrolase 1; FOLH1) which is located on the short arm of chromosome 11 [13]. PSMA is significantly overexpressed in most prostate cancer cells associated with increasing tumour grade and stage [14, 15]. Subsequent initial clinical experience using PSMA PET/CT imaging has demonstrated superior sensitivity and specificity compared to standard of care (SOC) imaging in primary, metastatic and biochemically recurrent prostate cancer [16, 17].

Despite the specificity implied by its name, PSMA is physiologically widely expressed in numerous other tissues including kidney, liver, spleen, small and large bowel, rectum, bladder, thyroid gland and salivary gland [14, 18, 19]. PSMA has also been shown to be expressed in the neovasculature of many solid tumours including RCC [20, 21]. Consequently, the extracellular portion of the PSMA protein is a potential site for targeted imaging strategies.

The von Hippel-Lindau (VHL) gene, a tumour suppressor gene, was identified in the past few decades as an important step in renal carcinogenesis. Gene mutation or deletion of the VHL gene is most commonly seen in clear cell RCC (ccRCC), resulting in the overexpression of hypoxia-inducible factor (HIF). This overexpression subsequently leads to increased pro-angiogenic growth factors including

vascular endothelial growth factor (VEGF), epidermal growth factor (EGF) and platelet-derived growth factor [22]. These growth factors induce cell proliferation and angiogenesis. PSMA expression in the neovasculature of benign and malignant tumours including RCC has been previously identified, with a number of published histopathologic literature consistently reporting high PSMA expression rates in ccRCC [20, 21, 23–28]. Chang et al. found PSMA expression in the neovascular endothelial cells of 20 metastatic ccRCC lesions involving a number of different metastatic sites (Table 1) [21]. Baccala et al. also reported similar findings with positive PSMA staining in tumour-associated neovascular cells in 76.2 % of ccRCC, 31.2 % of chromophobe RCC, 52.6 % of renal oncocytoma and 21.4 % of transitional RCC. Interestingly, papillary RCC and angiomyolipoma (AML) were PSMA negative and PSMA expression did not correlate with the pathologic stage of ccRCC [20]. This is in contrast to findings of Spatz et al. where higher grade and stage ccRCC were associated with increased PSMA expression in tumour vessels, and increased PSMA staining intensity was associated with poorer overall survival [23]. Furthermore, there is conflicting data regarding PSMA expression in primary RCC [18, 29] with the majority of the literature findings supporting PSMA expression only in tumour-associated neovasculature [14, 21, 30]. Due to avid physiological renal uptake of PSMA tracers, primary RCCs are poorly visualized [20, 31, 32]. Overall, the translation of these immunohistochemical findings to clinical imaging diagnostics is unclear; however, early small studies (Table 2) have been encouraging with improved metastatic lesion detection *via* PSMA PET/CT for ccRCC.

Role of PSMA in RCC Imaging and Clinical Implications

The role of PSMA PET/CT imaging has been well-established in the staging of primary, metastatic and biochemically recurrent prostate cancer [16, 17, 42–44]. There has been an increasing interest in the role of PSMA PET/CT imaging in non-prostate malignancies [45, 46]. This is somewhat limited with regard to RCC specifically, with few reviews in the literature to date [47]. Clinically, there has been growing interest in the utilization of PSMA PET/CT for improved detection of metastatic lesions from RCC, especially that of ccRCC. ccRCC is the most common RCC (80–90 %) and is a highly vascularized [44] tumour with PSMA expression detected in tumour-associated vasculature [20, 21, 23–28, 34, 38]. A number of PSMA-directed ligands have been used thus far for PET imaging of mRCC (Table 2).

In 2014, Demirci et al. published the first case report utilizing Ga-68 labelled Glu-NH-CO-NH-Lys-(Ahx) (^{68}Ga]Ga-PSMA-HBED-CC) PET/CT in the diagnosis of metastatic ccRCC [33]. They found that ^{68}Ga]Ga-PSMA-HBED-CC PET/CT had better radiotracer uptake for

Table 1. Summary of literature renal cell carcinoma-associated neovascular immunohistochemical staining

Study	Type	Year (study period)	Location	Number of patients/nephrectomies	Objectives	Number and biopsy sites	Immunohistochemical agent	Immunohistochemical staining agents and results
Chang et al. [21]	R	2001	USA	20 patients	Assess expression of PSMA on tumour-associated neovasculature of met CaP and met ccRCC	22 CaP met lesions (7 bone, 4 LN, 2 liver, 1 lung, 8 soft tissue) 20 ccRCC met lesions (5 bone, 5 LN, 1 liver, 3 lung, 2 soft tissue, 3 brain, 1 adrenal gland)	Anti-PSMA mAb PM2 J004.5 Anti-PSMA mAb 7E11 Anti-endothelial cell mAb CD34	For ccRCC met lesion: – Anti-PSMA mAb PM2 J004.5 positive 20/20 specimens; – Anti-PSMA mAb 7E11 positive 15/20; – Anti-endothelial cell mAb D34 20/20 Metastatic CaP only 2 of 22 had neovasculature PSMA expression PSMA was expressed in the proximal tubules of the normal kidney and in the tumour-associated vasculature in the renal tumours. Positive PSMA staining was detected in: 76.2 % (16/21) of ccRCC 31.2 % (5/16) of chrRCC 52.6 % (10/19) of oncocytoma 21.4 % (3/14) of TCC 0 % of pRCC and AML Of 30 patients with metastasis the primary tumour vessels were positive for PSMA in 29 (96.7 %) with ccRCC, of whom 60 % showed immunohistochemically moderate to highly reactive results. All 14 patients with ccRCC and LN metastases (pNI) showed positive PSMA results In all cases, immunoreactivity was detected only in the tumour-associated neovasculature and not in tumour cells. ccRCC showed the most diffuse staining pattern, where 24/30 cases (80 %) had >50 % reactive vessels ChrRCC (9/15; 60 %) Oncocytoma (5/15, 33 %). No diffuse staining was detected in any of the
Baccala et al. [20]	R	2007 (2003)	USA	109 RN or PN	Expression of PSMA in different primary renal tumours	169 lesions: 60 normal kidneys 21 ccRCC 20 pRCC 16 chrRCC 19 oncocytoma 14 TCC 19 AML	Anti-PSMA mAb PM2 J004.5 Anti-endothelial cell mAb CD34	
Spatz et al. [23]	R	2017 (1992–2004)	Germany	257 patients	Find rationale for PSMA based imaging and investigate the prognostic role of vascular PSMA expression in patients with RCC	257 nephrectomies. 2 cores of non-necrotic tumour and 2 cores of adjacent normal tissues were extracted	Anti-endothelial cell mAb CD34	
Al-Ahmadie et al. [28]	R	2008	USA	75 nephrectomies	To determine whether PSMA is differentially expressed in the vessels associated with distinct types of renal epithelial neoplasms and whether it could be used as a marker of subtype.	75 nephrectomies. The staining pattern was assessed in tumour cells, tumour-associated vessels and adjacent normal kidney.	Anti-PSMA mAb 13D6 Anti-endothelial mAb CD31	

Table 1. (continued)

Study	Type	Year (study period)	Location	Number of patients/nephrectomies	Objectives	Number and biopsy sites	Immunohistochemical agent	Immunohistochemical staining agents and results
								pRCC and only focal staining was detected in 11 cases (11/15; 73 %)

AML, angiolymphoma; CaP, prostate cancer; ccRCC, clear cell renal cell carcinoma; chRCC, chromophobe renal cell carcinoma; LN, lymph node; met, metastatic; mAb, monoclonal antibody; pRCC, papillary renal cell carcinoma; P, prospective study; PN, partial nephrectomy; PSMA, prostate-specific membrane antigen; pRCC, papillary renal cell carcinoma; R, retrospective study; RCC, radical nephrectomy; TCC, transitional cell carcinoma

multiple bone metastasis compared to 2-deoxy-2-[¹⁸F]fluoro-*D*-glucose ([¹⁸F]FDG) PET/CT resulting in improved visual detectability [33]. In 2015, Rowe et al. compared PSMA-targeted F-18-labelled-low-molecular-weight PSMA ligand 2-(3{1-carboxy-5-[(6-[¹⁸F]fluoro-pyridine-3-carbonyl)-amino]-pentyl}-ureido)-pentanedioic acid) also known as [¹⁸F] DCFPyL PET/CT to conventional imaging (CT or MRI) in five patients and demonstrated superior sensitivity (95 versus 79 %) [34]. Unfortunately, histopathology confirmation was not performed in this study; however, this was offset by a later study which revealed PSMA expression in the majority of PSMA PET/CT positive and conventional CT-negative lesions in a rapid autopsy series [24]. In 2016, Rowe et al. reported a comparison of [¹⁸F] FDG PET/CT and [¹⁸F] DCFPyL PET/CT in a single patient with mRCC. [¹⁸F] DCFPyL PET/CT demonstrated improved sensitivity and better radiotracer uptake in detecting occult lesions [35]. Rhee et al. investigated ten patients (eight patients with proven ccRCC) and found [⁶⁸Ga]Ga-PSMA-HBED-CC PET/CT had greater sensitivity (92 versus 69 %) and higher positive predictive value (97 versus 80 %) compared to conventional CT for detection of metastatic disease [26]. In this study, histopathological correlation was based on 36 tumour sites identified on either PSMA PET/CT or conventional CT. This improved staging ultimately led to a change in management of two patients, with the detection of small hepatic metastasis in one patient and superior delineation of IVC tumour in another. Both were subsequently excised. Siva et al. explored [⁶⁸Ga]Ga-PSMA-HBED-CC PET/CT and [¹⁸F] FDG PET/CT in six cases of ccRCC with concordance in five patients whilst the former was able to detect two more lesions that led to a change in patient management [38].

PSMA is therefore a potential marker for the identification of tumour neoangiogenesis in ccRCC, even in small metastatic lesions. PSMA PET/CT may aid in improved early diagnosis and staging leading to subsequent interventions. It also has the extra added benefit of being able to be performed in patients with renal impairment or contrast allergy without risk of adverse events. These potential advantages may be limited to ccRCC, however, as non-clear cell metastasis is inconsistently detected. In a comparison between [¹⁸F] DCFPyL PET/CT and conventional imaging (CT and/or MRI) in non-ccRCC, Yin et al. reported that only 10 of 73 (13.7 %) suspected metastatic lesions had definitive radiotracer uptake with no additional lesions identified through the use of molecular imaging [41].

Limitations and Future Opportunities

Currently, there are limitations for the clinical use of PSMA PET/CT imaging in RCC highlighted by the lack of large, well-designed clinical trials in the literature. PSMA PET/CT imaging has minimal value in evaluation of primary renal tumours. PSMA is expressed physiologically in the proximal tubules of normal kidneys and exhibits no increased

Table 2. Summary of literature utilizing PSMA-PET CT imaging for detection of metastatic RCC (mRCC)

Study	Type	Year	Location	Objectives	Number of patients	Median age (Q1,Q3)	Histology	Radiotracer/ligand	Comparative conventional imaging	Metastatic RCC lesion detection
Demirci et al. [33]	CR	2014	Turkey	Compare ⁶⁸ Ga[PSMA-HBED-CC PET/CT and ¹⁸ F] FDG PET/CT for detection of mRCC	1	65	ccRCC	⁶⁸ Ga[PSMA-HBED-CC	[¹⁸ F] FDG PET/CT	[⁶⁸ Ga]Ga-PSMA-HBED-CC provided improved radiotracer uptake in metastatic bone lesions (seventh cervical vertebrae and acromion left scapula SUVmax = 35 for PSMA, FDG = 7.2; sternum SUVmax = 28.3 for PSMA, FDG = 5.15; Right ischial tuberosity SUVmax = 34.1 for PSMA, FDG = 5.3)
Rowe et al. [34]	P	2015	USA	Evaluate the utility of [¹⁸ F] DCFPyL PET/CT in imaging 5 patients with metastatic ccRCC	5	58 (54-61)	10 ccRCC	[¹⁸ F]DCFpyL	Contrast-enhanced CT or MRI	[¹⁸ F] DCFPyL PET/CT → 28 lesions were identified, 17 of which corresponded to sites of disease on conventional imaging; identified lesions not detected on CT/MRI (small LNs in mediastinum and RP, paraspinous muscle, perineal SC soft tissue, bone lesions) → sensitivity 94.7 % CT/MRI → 18 lesions were identified, able to detect one lesion that [¹⁸ F] DCFPyL was unable (6 mm metastatic liver lesion) → sensitivity 78.9 % [¹⁸ F] DCFPyL PET/CT has improved sensitivity and better radiotracer uptake compared to [¹⁸ F] FDG PET/CT
Rowe et al. [35]	CR	2016	USA	Compare [¹⁸ F] DCFPyL PET/CT and [¹⁸ F] FDG PET/CT for detection of metastatic lesions in ccRCC	1	58	ccRCC	[¹⁸ F]DCFpyl	[¹⁸ F] FDG PET/CT	<ul style="list-style-type: none"> • Additional subtle uptake in left posterior ninth rib and left iliac bone found in [¹⁸F] DCFPyL PET/CT • Right iliac lesion (SUVmax = 16.6 for [¹⁸F] DCFPyL, 3.3 for [¹⁸F]FDG); left iliac lesion (SUVmax = 13.9 for [¹⁸F] DCFPyL, 4.0 for [¹⁸F]FDG)
Rhee et al. [25]	P	2016	Australia	Evaluate the diagnostic potential of ⁶⁸ Ga] Ga-PSMA-HBED-CC PET/CT in detecting met renal tumour lesions	10	57 (45-69)	8 ccRCC 1 pRCC 1 unclassified	[⁶⁸ Ga]Ga-PSMA-HBED-CC	CT ± MRI/US/BS	<ul style="list-style-type: none"> • Detection of mRCC lesions: Conventional sensitivity 68.6 %; PPV 80 %; • PSMA PET/CT sensitivity 92.11 %; PPV 97.22 %
Einspieler et al. [36]	CR	2016	Germany	-	1	78	-	[⁶⁸ Ga]Ga-PSMA-HBED-CC	-	<ul style="list-style-type: none"> • PSMA expression in mediastinal, RP and iliac LN with subsequent Bx LN → RCC metastatic lesion

Table 2. (continued)

Study	Type	Year	Location	Objectives	Number of patients	Median age (Q1,Q3)	Histology	Radiotracer/ ligand	Comparative conventional imaging	Metastatic RCC lesion detection
Sawicki et al. [27]	R	2016	Germany	Evaluate the diagnostic potential of whole-body PET/CT using ⁶⁸ Ga-labelled PSMA ligand in histology proven RCC	6	72.3 (65–80)	Histology proven primary or metastatic/recurrent RCC: 4 ccRCC 1 pRCC1 chrCC	[⁶⁸ Ga]Ga-PSMA-HBED-CC	–	22 lesions detected: • 21 malignant lesions – 5 primary RCCs in 5 patients however no significant differentiating uptake between normal parenchyma and tumour – 8/16 metastatic lesions in 2 ccRCC patients showed focal ⁶⁸ Ga-PSMA uptake • 1 benign lesion → ectopic salivary tissue in left masseter Intensely PSMA tracer concentrating lesions in left suprarenal region, mediastinal LN, lytic bone lesions, thyroid nodules, and mild abnormal tracer-avid lung nodules; FDG showed mild to no significant abnormal uptake in these lesions. • 4 patients with ccRCC met lesions (lung, adrenal) → intense PSMA avidity versus low-mod FDG avidity • 1 patient post ccRCC resection with adrenal met → no uptake FDG or PSMA although histology positive • 1 patient with ccRCC GT met lesion → avid FDG and PSMA uptake however greater FDG intensity • 1 patient bed recurrence pRCC → FDG-avid, not avid on PSMA 3 PET avid lesions (RP LN and thyroid gland histology RCC; pelvic LN histology CaP) Detected 4 separate lesions in prostate, left thyroid, right kidney, left posterior iliac crest. Histology confirmed CaP (4 + 4), papillary carcinoma of thyroid, ccRCC and benign bone Bx 73 metastatic lesions and 3 primary renal lesions were identified on conventional imaging. No lesions were identified on [¹⁸ F] DCFPyI PET/CT without
Sasikumar et al. [37]	CR	2016	India	–	1	80	ccRCC	[⁶⁸ Ga]Ga-PSMA-HBED-CC	[¹⁸ F] FDG PET/CT	
Siva et al. [38]	R	2017	Australia	To assess the utility of PSMA-PET/CT for diagnostic evaluation of isolated or limited metastases planned for local surgery or radiation + therapeutic response assessment post SABR	8	65 (50–78)	7 ccRCC 1 pRCC	[⁶⁸ Ga]Ga-PSMA-HBED-CC	[¹⁸ F] FDG PET/CT	
Zacho et al. [39]	CR	2017	Denmark	–	1	75	–	[⁶⁸ Ga]Ga-PSMA-HBED-CC	–	
Joshi et al. [40]	CR	2017	Australia	–	1	67	–	[⁶⁸ Ga]Ga-PSMA-HBED-CC	–	
Yin et al. [41]	P	2018	USA	To investigate the utility of PSMA-targeted [¹⁸ F] DCFPyI PET/CT for detection of metastatic lesion of non-	8	61 (54–66)	Histology proven non-clear cell RCC: 3 pRCC 2chrCC 2 unclassified 1 Xp11 translocation	[¹⁸ F] DCFPyI	CT or MRI	

Table 2. (continued)

Study	Type	Year	Location	Objectives	Number of patients	Median age (Q1,Q3)	Histology	Radiotracer/ligand	Comparative conventional imaging	Metastatic RCC lesion detection
				clear cell RCC						corresponding finding on conventional imaging. <ul style="list-style-type: none"> • Only 10 of 73 (13.7 %) suspected metastatic lesions were classified as having definitive radiotracer uptake • 14 (19.2 %) equivocal • 49 (67.1 %) no significant uptake
AML, angiomyolipoma; BS, bone scan; Bx, biopsy; CaP, prostate cancer; ccRCC, clear cell renal cell carcinoma; chRCC, chromophobe renal cell carcinoma; CR, case report; CT, computed tomography; 18F-DCFPyl, 18F-labelled low-molecular weight PSMA ligand 2-(3-{1-carboxy-5-[(6-[18F]fluoro-pyridine-3-carbonyl)-amino]-pentyl}-ureido)-pentanedioic acid); 68Ga, gallium-68; GT, greater trochanter; LN, lymph node; mAb, monoclonal antibody; met, metastatic; mRCC, metastatic renal cell carcinoma; MRI, magnetic resonance imaging; pRCC, papillary renal cell carcinoma; P, prospective study; PET, positron emission tomography; PN, partial nephrectomy; PPV, positive predictive value; PSMA, Prostate-specific membrane antigen; pRCC, papillary renal cell carcinoma; RP, retro-peritoneal; R, retrospective study; RN, radical nephrectomy; SABR, stereotactic ablative body radiotherapy; SC, subcutaneous; SUV max, maximum standardized uptake value; TCC, transitional cell carcinoma; US, ultrasound, USA, United States of America										

expression in the primary tumour, compared to the normal surrounding renal parenchyma [20, 31, 32]. Other factors affecting specificity include wide PSMA expression across a number of normal organs (liver, spleen, salivary glands) [14, 18, 19], and expression in other solid organ tumours (e.g. bladder, colorectal, lung) [14, 48–50]. Other benign specific (sarcoidosis, tuberculosis) [51, 52] and non-specific conditions (regeneration, repair) [19, 53] can also lead to false positive imaging findings. Additionally, there is an inability to distinguish between benign and malignant renal tumour-associated neovasculature or RCC subtypes; however, PSMA ligand binding is more likely in ccRCC compared to pRCC or AML [20]. There is also conflicting evidence on the prognostic role of PSMA expression in the neovasculature of ccRCC [20, 23].

The small studies thus far, however, suggest that PSMA PET/CT imaging appears to facilitate improved detection of mRCC lesions especially in ccRCC. Studies have reported multiple biopsy proven metastatic ccRCC lesions, missed by conventional CT imaging indicating a potential role for more accurate staging of advanced and high-risk ccRCC [26]. This improved sensitivity in detection of metastatic ccRCC may significantly impact patient outcomes, especially with early detection of potentially curable oligometastatic disease [26].

PSMA PET/CT imaging could also be potentially utilized as a way of monitoring, assessing and/or predicting response to systemic neovasculature agents targeting angiogenesis in ccRCC. It may provide functional imaging feedback as a measure of the *in vivo* density of the neovascularity of metastatic lesions. Currently, no PSMA-targeted therapy exists although anti-angiogenesis agents VEGF and platelet-derived growth factor receptor (sunitinib, pazopanib) and isolated VEGF inhibitors (bevacizumab) have had varying degrees of success when used in clinical trials in advanced stage ccRCC [8, 54–56]. The potential therapeutic success and good tolerability of PSMA-targeted therapies in prostate cancer (with radionucleotides such as 177Lu-labelled PSMA) suggests promising potential further development. Further larger prospective trials are required in both diagnostic and treatment-related PSMA utilization [57, 58].

Currently, there are two clinical trials currently underway in the US investigating the role of PSMA imaging in RCC. The aim of one study is to evaluate and compare the diagnostic accuracy of PSMA-targeted [18F] DCFPyL PET/CT with conventional imaging in 30 patients with various stages of mRCC [59]. The second trial is evaluating the feasibility and uptake of 68Ga P16-093 PSMA PET/CT in 30 patients with known or suspected metastatic RCC or prostate cancer [60].

Conclusion

PSMA expression in the tumour neovasculature of RCC has been established especially for ccRCC. Although there is a sound scientific basis for the use of PSMA-PET/CT in

detecting ccRCC metastatic foci, its translation into clinical practice is currently not routinely recommended. Early small studies involving PSMA PET/CT imaging have been encouraging with evidence of improved sensitivity compared to conventional imaging modalities. Further investigation is needed to confirm utility through larger studies in this clinical setting.

Funding Information. This research received no specific grant from any funding agency in public, commercial, or not-for-profit sectors. IV is supported by a Movember Clinician Scientist Award awarded through Prostate Cancer Foundation of Australia's Research Program.

Compliance with Ethical Standards

Conflict of Interest

All authors declare that there is no conflict of interest.

Human and Animal Rights

This article does not contain any studies with human participants or animals performed by any of the authors.

Publisher's Note. Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

References

- Znaor A, Lortet-Tieulent J, Laversanne MJ et al (2015) International variations and trends in renal cell carcinoma incidence and mortality. *Eur Urol* 67:519–530
- Gupta K, Miller JD, Li JZ, Russell MW, Charbonneau C (2008) Epidemiologic and socioeconomic burden of metastatic renal cell carcinoma (mRCC): a literature review. *Cancer Treat Rev* 34:193–205
- Dabestani S, Thorstenson A, Lindblad P, Harmenberg U, Ljungberg B, Lundstam S (2016) Renal cell carcinoma recurrences and metastases in primary non-metastatic patients: a population-based study. *World J Urol* 34:1081–1086
- Thompson RH, Hill JR, Babayev Y, Cronin A, Kaag M, Kundu S, Bernstein M, Coleman J, Dalbagni G, Touijer K, Russo P (2009) Metastatic renal cell carcinoma risk according to tumor size. *J Urol* 182:41–45
- Capogrosso P, Capitanio U, La Croce G et al (2016) Follow-up after treatment for renal cell carcinoma: the evidence beyond the guidelines. *Eur Urol Focus* 1:272–281
- Pierorazio PM, Johnson MH, Patel HD, Sozio SM, Sharma R, Iyoha E, Bass EB, Allaf ME (2016) Management of Renal Masses and Localized Renal Cancer: systematic review and meta-analysis. *J Urol* 196:989–999
- Pantuck AJ, Zisman A, Beldegrun AS (2001) The changing natural history of renal cell carcinoma. *J Urol* 166:1611–1623
- Escudier B, Pluzanska A, Koralewski P, Ravaud A, Bracarda S, Szczylik C, Chevreau C, Flipek M, Melichar B, Bajetta E, Gorbunova V, Bay JO, Bodrogi I, Jagiello-Gruszfeld A, Moore N (2007) Bevacizumab plus interferon alfa-2a for treatment of metastatic renal cell carcinoma: a randomised, double-blind phase III trial. *Lancet* 370:2103–2111
- Motzer RJ, Russo P (2000) Systemic therapy for renal cell carcinoma. *J Urol* 163:408–417
- Brufau BP, Cerqueda CS, Villalba LB, Izquierdo RS, González BM, Molina CN (2013) Metastatic renal cell carcinoma: radiologic findings and assessment of response to targeted antiangiogenic therapy by using multidetector CT. *Radiographics* 33:1691–1716
- Loh J, Davis ID, Martin JM, Siva S (2014) Extracranial oligometastatic renal cell carcinoma: current management and future directions. *Future Oncol* 10:761–774
- Ghosh A, Heston WD (2004) Tumor target prostate specific membrane antigen (PSMA) and its regulation in prostate cancer. *J Cell Biochem* 91:528–539
- Leek J, Lench N, Maraj B, Bailey A, Carr IM, Andersen S, Cross J, Whelan P, MacLennan K, Meredith DM (1995) Prostate-specific membrane antigen: evidence for the existence of a second related human gene. *Br J Cancer* 72:583–588
- Silver DA, Pellicer I, Fair WR, Heston WD, Cordon-Cardo C (1997) Prostate-specific membrane antigen expression in normal and malignant human tissues. *Clin Cancer Res* 3:81–85
- Bostwick DG, Pacelli A, Blute M, Roche P, Murphy GP (1998) Prostate specific membrane antigen expression in prostatic intraepithelial neoplasia and adenocarcinoma: a study of 184 cases. *Cancer* 82:2256–2261
- Perera M, Papa N, Christidis D, Wetherell D, Hofman MS, Murphy DG, Bolton D, Lawrentschuk N (2016) Sensitivity, specificity, and predictors of positive ⁶⁸Ga-prostate-specific membrane antigen positron emission tomography in advanced prostate cancer: a systematic review and meta-analysis. *Eur Urol* 70:926–937
- Perera M, Murphy D, Lawrentschuk N (2018) Prostate-specific membrane antigen positron emission tomography/computed tomography in locally advanced, recurrent, and metastatic prostate cancer. *JAMA Oncol* 4:748–749
- Kinoshita Y, Kuratsukuri K, Landas S, Imaida K, Rovito PM Jr, Wang CY, Haas GP (2006) Expression of prostate-specific membrane antigen in normal and malignant human tissues. *World J Surg* 30:628–636
- Kirchner J, Schaarschmidt BM, Sawicki LM, Heusch P, Hautzel H, Ermert J, Rabenalt R, Antoch G, Buchbender C (2017) Evaluation of practical interpretation hurdles in ⁶⁸Ga-PSMA PET/CT in 55 patients: physiological tracer distribution and incidental tracer uptake. *Clin Nucl Med* 42:e322–e327
- Baccala A, Sercia L, Li J, Heston W, Zhou M (2007) Expression of prostate-specific membrane antigen in tumor-associated neovasculature of renal neoplasms. *Urology* 70:385–390
- Chang SS, Reuter VE, Heston WD, Gaudin PB (2001) Metastatic renal cell carcinoma neovasculature expresses prostate-specific membrane antigen. *Urology* 57:801–805
- Cohen D, Zhou M (2005) Molecular genetics of familial renal cell carcinoma syndromes. *Clin Lab Med* 25:259–277
- Spatz S, Tolkach Y, Jung K, Stephan C, Busch J, Ralla B, Rabien A, Feldmann G, Brossart P, Bundschuh RA, Ahmadzadehfar H, Essler M, Toma M, Müller SC, Ellinger J, Hauser S, Kristiansen G (2018) Comprehensive evaluation of prostate specific membrane antigen expression in the vasculature of renal tumors: implications for imaging studies and prognostic role. *J Urol* 199:370–377
- Gorin MA, Rowe SP, Hooper JE, Kates M, Hammers HJ, Szabo Z, Pomper MG, Allaf ME (2017) PSMA-targeted ¹⁸F-DCFPyL PET/CT imaging of clear cell renal cell carcinoma: results from a rapid autopsy. *Eur Urol* 71:145–146
- Rhee H, Ng KL, Tse BW et al (2016) Using prostate specific membrane antigen (PSMA) expression in clear cell renal cell carcinoma for imaging advanced disease. *Pathology* 48:613–616
- Rhee H, Blazak J, Tham CM, Ng KL, Shepherd B, Lawson M, Preston J, Vela I, Thomas P, Wood S (2016) Pilot study: use of gallium-68 PSMA PET for detection of metastatic lesions in patients with renal tumour. *EJNMMI Res* 6:76
- Sawicki LM, Buchbender C, Boos J, Giessing M, Ermert J, Antke C, Antoch G, Hautzel H (2017) Diagnostic potential of PET/CT using a ⁶⁸Ga-labelled prostate-specific membrane antigen ligand in whole-body staging of renal cell carcinoma: initial experience. *Eur J Nucl Med Mol Imaging* 44:102–107
- Al-Ahmadie HA, Olgac S, Gregor PD et al (2008) Expression of prostate-specific membrane antigen in renal cortical tumors. *Mod Pathol* 21:727–732
- Dumas F, Gala JL, Berteau P et al (1999) Molecular expression of PSMA mRNA and protein in primary renal tumors. *Int J Cancer* 80:799–803
- Chang SS, Reuter VE, Heston WD, Bander NH, Grauer LS, Gaudin PB (1999) Five different anti-prostate-specific membrane antigen (PSMA) antibodies confirm PSMA expression in tumor-associated neovasculature. *Cancer Res* 59:3192–3198
- Martinez de Llano SR, Delgado-Bolton RC, Jimenez-Vicioso A et al (2007) Meta-analysis of the diagnostic performance of ¹⁸F-FDG PET in renal cell carcinoma. *Rev Esp Med Nucl* 26:19–29
- Wang HY, Ding HJ, Chen JH, Chao CH, Lu YY, Lin WY, Kao CH (2012) Meta-analysis of the diagnostic performance of [¹⁸F]FDG-PET and PET/CT in renal cell carcinoma. *Cancer Imaging* 12:464–474

33. Demirci E, Ocak M, Kabasakal L, Decristoforo C, Talat Z, Halaç M, Kanmaz B (2014) ^{68}Ga -PSMA PET/CT imaging of metastatic clear cell renal cell carcinoma. *Eur J Nucl Med Mol Imaging* 41:1461–1462
34. Rowe SP, Gorin MA, Hammers HJ, Som Javadi M, Hawasli H, Szabo Z, Cho SY, Pomper MG, Allaf ME (2015) Imaging of metastatic clear cell renal cell carcinoma with PSMA-targeted ^{18}F -DCFPyL PET/CT. *Ann Nucl Med* 29:877–882
35. Rowe SP, Gorin MA, Hammers HJ, Pomper MG, Allaf ME, Javadi MS (2016) Detection of ^{18}F -FDG PET/CT occult lesions with ^{18}F -DCFPyL PET/CT in a patient with metastatic renal cell carcinoma. *Clin Nucl Med* 41:83–85
36. Einspieler I, Tauber R, Maurer T, Schwaiger M, Eiber M (2016) ^{68}Ga prostate-specific membrane antigen uptake in renal cell cancer lymph node metastases. *Clin Nucl Med* 41:e261–e262
37. Sasikumar A, Joy A, Nanabala R, Unni M, TK P (2016) Complimentary pattern of uptake in ^{18}F -FDG PET/CT and ^{68}Ga -prostate-specific membrane antigen PET/CT in a case of metastatic clear cell renal carcinoma. *Clin Nucl Med* 41:e517–e519
38. Siva S, Callahan J, Pryor D, Martin J, Lawrentschuk N, Hofman MS (2017) Utility of (68) Ga prostate specific membrane antigen - positron emission tomography in diagnosis and response assessment of recurrent renal cell carcinoma. *J Med Imaging Radiat Oncol* 61:372–378
39. Zacho HD, Nielsen JB, Dettmann K, Haberkorn U, Petersen LJ (2017) Incidental detection of thyroid metastases from renal cell carcinoma using ^{68}Ga -PSMA PET/CT to assess prostate cancer recurrence. *Clin Nucl Med* 42:221–222
40. Joshi A, Nicholson C, Rhee H, Gustafson S, Miles K, Vela I (2017) Incidental malignancies identified during staging for prostate cancer with ^{68}Ga prostate-specific membrane antigen HBED-CC positron emission tomography imaging. *Urology* 104:e3–e4
41. Yin Y, Campbell SP, Markowski MC, Pierorazio PM, Pomper MG, Allaf ME, Rowe SP, Gorin MA (2018) Inconsistent detection of sites of metastatic non-clear cell renal cell carcinoma with PSMA-targeted [^{18}F] DCFPyL PET/CT. *Mol Imaging Biol*
42. Corfield J, Perera M, Bolton D, Lawrentschuk N (2018) ^{68}Ga -prostate specific membrane antigen (PSMA) positron emission tomography (PET) for primary staging of high-risk prostate cancer: a systematic review. *World J Urol* 36:519–527
43. Udovicich C, Perera M, Hofman MS, Siva S, del Rio A, Murphy DG, Lawrentschuk N (2017) ^{68}Ga -prostate-specific membrane antigen-positron emission tomography/computed tomography in advanced prostate cancer: current state and future trends. *Prostate Int* 5:125–129
44. Rhee H, Thomas P, Shepherd B, Gustafson S, Vela I, Russell PJ, Nelson C, Chung E, Wood G, Malone G, Wood S, Heathcote P (2016) Prostate specific membrane antigen positron emission tomography may improve the diagnostic accuracy of multiparametric magnetic resonance imaging in localized prostate cancer. *J Urol* 196:1261–1267
45. Backhaus P, Noto B, Avramovic N, Grubert LS, Huss S, Bögemann M, Stegger L, Weckesser M, Schäfers M, Rahbar K (2018) Targeting PSMA by radioligands in non-prostate disease-current status and future perspectives. *Eur J Nucl Med Mol Imaging* 45:860–877
46. Salas Fragomeni RA, Amir T, Sheikhabahaei S, Harvey SC, Javadi MS, Solnes LB, Kiess AP, Allaf ME, Pomper MG, Gorin MA, Rowe SP (2018) Imaging of nonprostate cancers using PSMA-targeted radiotracers: rationale, current state of the field, and a call to arms. *J Nucl Med* 59:871–877
47. Evangelista L, Basso U, Maruzzo M, Novara G (2018) The role of radiolabeled prostate-specific membrane antigen positron emission tomography/computed tomography for the evaluation of renal cancer. *Eur Urol Focus* S2405–4569(18)30223–30232. <https://doi.org/10.1016/j.euf.2018.08.004>
48. Hangaard L, Jochumsen MR, Vendelbo MH, Bouchelouche K (2017) Metastases from colorectal cancer avid on ^{68}Ga -PSMA PET/CT. *Clin Nucl Med* 42:532–533
49. Roy SG, Parida GK, Tripathy S, Singhal A, Tripathi M, Bal C (2017) *In vivo* demonstration of PSMA expression in adenocarcinoma urinary bladder using ^{68}Ga -PSMA 11 PET/CT. *Clin Nucl Med* 42:542–543
50. Wang HL, Wang SS, Song WH, Pan Y, Yu HP, Si TG, Liu Y, Cui XN, Guo Z (2015) Expression of prostate-specific membrane antigen in lung cancer cells and tumor neovasculature endothelial cells and its clinical significance. *PLoS One* 10:e0125924
51. Ardies PJ, Gykiere P, Goethals L, de Mey J, de Geeter F, Everaert H (2017) PSMA uptake in mediastinal sarcoidosis. *Clin Nucl Med* 42:303–305
52. Pyka T, Weirich G, Einspieler I, Maurer T, Theisen J, Hatzichristodoulou G, Schwamborn K, Schwaiger M, Eiber M (2016) ^{68}Ga -PSMA-HBED-CC PET for differential diagnosis of suggestive lung lesions in patients with prostate cancer. *J Nucl Med* 57:367–371
53. Gordon IO, Tretiakova MS, Noffsinger AE, Hart J, Reuter VE, al-Ahmadie HA (2008) Prostate-specific membrane antigen expression in regeneration and repair. *Mod Pathol* 21:1421–1427
54. Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Rixe O, Oudard S, Negrier S, Szczylik C, Kim ST, Chen I, Bycott PW, Baum CM, Figlin RA (2007) Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. *N Engl J Med* 356:115–124
55. Sternberg CN, Hawkins RE, Wagstaff J, Salman P, Mardiak J, Barrios CH, Zarba JJ, Gladkov OA, Lee E, Szczylik C, McCann L, Rubin SD, Chen M, Davis ID (2013) A randomised, double-blind phase III study of pazopanib in patients with advanced and/or metastatic renal cell carcinoma: final overall survival results and safety update. *Eur J Cancer* 49:1287–1296
56. Choueiri TK, Motzer RJ (2017) Systemic therapy for metastatic renal-cell carcinoma. *N Engl J Med* 376:354–366
57. Baum RP, Kulkarni HR, Schuchardt C, Singh A, Wirtz M, Wiessalla S, Schottelius M, Mueller D, Klette I, Wester HJ (2016) ^{177}Lu -labeled prostate-specific membrane antigen radioligand therapy of metastatic castration-resistant prostate cancer: safety and efficacy. *J Nucl Med* 57:1006–1013
58. Ahmadzadehfar H, Rahbar K, Kurpig S et al (2015) Early side effects and first results of radioligand therapy with ^{177}Lu -DKFZ-617 PSMA of castrate-resistant metastatic prostate cancer: a two-centre study. *EJNMMI Res* 5:114
59. National Institute of Health [ClinicalTrials.gov](https://clinicaltrials.gov). STUDY of PSMA-targeted ^{18}F -DCFPyL PET/CT in the evaluation of patients with renal cell carcinoma. <https://clinicaltrials.gov/ct2/show/NCT02687139>. (accessed 4th May 2018)
60. National Institute of Health [ClinicalTrials.gov](https://clinicaltrials.gov) preliminary evaluation of uptake of [^{68}Ga]P16-093 in Metastatic Prostate and Renal Cancer. <https://clinicaltrials.gov/ct2/show/NCT03073395> (accessed May 4th)