



Original Contribution

Micropapillary adenocarcinoma of lung: Morphological criteria and diagnostic reproducibility among pulmonary pathologists

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ABSTRACT

Context: Invasive micropapillary adenocarcinoma (MPC) is an aggressive variant of lung adenocarcinoma, frequently manifesting with advanced stage lymph node metastasis and decreased survival.

Objective: Identification of this morphology is important, as it is strongly correlated with poor prognosis regardless of the amount of MPC component. To date, no study has investigated the morphological criteria used to objectively diagnose it.

Design: Herein, we selected 30 cases of potential MPC of lung, and distributed 2 digital images per case among 15 pulmonary pathology experts. Reviewers were requested to diagnostically interpret, assign the percentage of MPC component, and record the morphological features they identified. The noted features included: columnar cells, elongated slender cell nests, extensive stromal retraction, lumen formation with internal epithelial tufting, epithelial signet ring-like forms, intracytoplasmic vacuolization, multiple nests in the same alveolar space, back-to-back lacunar spaces, epithelial nest anastomosis, marked pleomorphism, peripherally oriented nuclei, randomly distributed nuclei, small/medium/large tumor nest size, fibrovascular cores, and spread through air-spaces (STAS).

Results: Cluster analysis revealed three subgroups with the following diagnoses: “MPC”, “combined papillary and MPC”, and “others”. The subgroups correlated with the reported median percentage of MPC. Intracytoplasmic vacuolization, epithelial nest anastomosis/confluence, multiple nests in the same alveolar space, and small/medium tumor nest size were the most common criteria identified in the cases diagnosed as MPC. Peripherally oriented nuclei and epithelial signet ring-like forms were frequently identified in both the “MPC” and “combined papillary and MPC” groups.

Conclusions: Our study provides objective diagnostic criteria to diagnose MPC of lung.

1. Introduction

Among carcinomas, micropapillary patterns have been recognized

as aggressive variants in organs such as breast, bladder, salivary gland, and colon [1–10]. Invasive adenocarcinoma with a micropapillary pattern has also been identified in the lung, and the majority of lesions

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have been determined to be primary tumors rather than metastatic lesions, with both cytokeratin 7 (CK7) and thyroid transcription factor (TTF1) reactivity [11]. Invasive micropapillary adenocarcinoma (MPC) has also been reported to have adverse prognosis in the lung [12–17]. Studies have shown that both intra- and extra-pulmonary metastasis as well as lymphovascular invasion (LVI) are common; and the 5 year survival rate after diagnosis is significantly lower compared to patients with other morphological variants and the same tumor stage [11,18–23]. Recurrence is more frequent in patients with MPC of lung compared to other patterns such as acinar or papillary [20,24–28]. Thus, overall, MPC of lung has been recognized as a distinct morphological marker with poor prognosis, even when it is present in small amounts [8,11]. Interestingly, the prognostic findings are similar regardless of the percentage of MPC found within the primary tumor [8,11].

To date, the precise morphological criteria to diagnose MPC of lung have not been objectively determined. Furthermore, diagnostic accuracy in equivocal cases has not been thoroughly assessed. Several research groups have proposed adherence to arbitrary definitions in breast, urinary bladder, lung, and other organs; nonetheless, these studies lack objectiveness in criteria selection [6,10,29–32]. Morphological features that have been described in MPCs of lung include: small papillary tufts freely lying within alveolar spaces/encased within thin walls of connective tissue spaces, few or no fibrovascular cores, spaces with flat endothelial or no lining, micropapillary tufts in lymphovascular spaces, and variants consisting of micropapillary tufts floating within cystic spaces lined with tumor cells [8,11,17,19,33]. Stromal retraction was also noted in many cases [18]. Size variation, including very small nests, has been a controversial topic, and researchers have studied consecutive sections of paraffin-embedded blocks to determine when cellular tufts extend or disappear in consecutive cuts (to address the size concern: small vs. medium or large nests) [19]. However, diagnostic reproducibility using specific criteria has not been thoroughly evaluated, and we continue to find inconsistencies in the studies reporting MPC of lung. For example, in 4 studies of lung adenocarcinoma (including > 200 patients), the average number of patients diagnosed with the micropapillary variant was markedly different; the average ranged between 3.4 and 41% among the studies [34–37]. The aforementioned findings indicate lack of diagnostic consistency, which should be addressed in our opinion.

Given the clinical needs, in our study, we aimed to evaluate the diagnostic interpretations among pulmonary pathologists in both unequivocal cases of MPC of lung and challenging ones where diagnosis was not straightforward. Furthermore, our intention was to determine the morphological features identified in cases that were diagnosed as MPC of lung with high percentage of agreement on diagnoses among reviewers.

2. Materials and methods

A total of 30 cases of adenocarcinoma of lung (only one to two slides per case), submitted by all the coauthors, were reviewed and selected by two of the authors (RM, JR). The selected cases included tumors with unequivocal areas of MPC morphology, others with areas of mixed histological patterns including MPC (but also papillary, acinar, etc.), and cases with areas that could be classified as “non-diagnostic for MPC” (Fig. 1A–F). Only cases with wedge resection, segmentectomy, lobectomy or pneumonectomy were included; biopsies were excluded from the study. A digital survey, consisting of three questions and two digital still images per case, was distributed to 15 pulmonary pathologists. Our goal was to determine if specific morphological criteria- reproducible among pathologists- were present to diagnose this entity, even when limited amount of tissue was available. The 15 participating pathologists (ACR, AC, RB, PC, YG, HDT, MLS, BTL, LMS, MBB, AB, KR, AA, RM, JR) were from 10 different institutions, including experts that have practiced outside the United States. Each pathologist was assigned a numerical code (1 to 15) to secure anonymity of the individual

responses.

The digital survey involved interpretation of two static images of hematoxylin- and eosin- stained slides. The images included areas of parenchyma and others near the periphery. The criteria chosen for the study were selected by two authors (RM, JR), according to those established by the 2015 WHO Lung Tumor Classification, and relevant criteria identified in other cancer types where micropapillary patterns have been extensively described [5–10,11]. The images were chosen for illustrative purposes to solicit potential diversity in opinions; they were representative of parts of the tumors selected, an included a high power (40–60X) and a low power (10–20X) in each case. The remainder of the slide was not included for complete review. The survey comprised three questions addressing the following: diagnostic interpretation, percentage of MPC component observed in the images (quantitative assessment), and determination of the morphological criteria used to make a diagnosis, initially recorded in diagnostic interpretation (Table 1). Pathologists were given several weeks to answer the survey, and the survey could be interrupted and continued at any moment during the period of time provided (to prevent bias based on the survey including 30 cases with multiple choices available for each question). Given our study design, we predicted that we would be able to identify morphological criteria used by the pathologist to diagnose MPC and other types of adenocarcinoma such as papillary, acinar, and others.

Statistical analyses were performed using the R software for statistical computing (<http://www.r-project.org/>). The relationships of the pathologists' diagnostic interpretations and mean percentage of morphological criteria reported by case were assessed using unsupervised hierarchical clustering with Euclidean distance and complete linkage. The clustering analysis of the tumor cases, with the morphological criteria and diagnostic impressions, was visualized on a heatmap dendrogram using the “heatmap.2” function in the R package “gplots”. Weighted kappa was used to compare contingency tables with the subgrouping cluster, predicted by morphological criteria and diagnostic classification.

Additionally, the participating pathologists were separated in 2 groups, based on their years of experience in the field. The groups were designated as “less than 15 years” vs. “15 years or more”. Using heatmap dendrograms, we compared the diagnostic impressions and inter-observer reproducibility among each group. Overall percent of agreement (OPA) values were used to assess inter-observer reproducibility; OPA values were calculated by comparing each possible permutation between reviewers (pathologists) data and sub-groups. An analysis-of-variance (ANOVA) model was created to determine differences in the median MPC percentage and morphological criteria in each subgroup, as reported by the pathologists in the hierarchical subgroups. Differences between groups were assessed with a Tukey-Kramer pairwise comparison test. All statistics were two-sided, and a p -value < 0.05 was considered statistically significant.

3. Results

Hierarchical clustering analysis of the diagnostic interpretations among pulmonary pathologists (addressed in the first question of the survey) resulted in three subgroups with different predominant diagnoses: “others”, “mixed papillary with MPC”, and “MPC” (Fig. 2A). The aforementioned subgroups were designated as MPC0, MPC1, and MPC2, respectively, depending on the total amount of MPC component. The overall diagnostic percentage of agreement among all reviewers and all cases was 45.0% (CI, 43.2–46.7). The overall percent of agreement that the reviewers reached within the subgroups was 11.3% (CI, 10.6–11.9) for the 9 cases of MP0; 47.9% (CI, 47.3–48.6) for the 8 cases of MP1; and 57.5% (CI, 56.8–58.2) for 13 cases of MP2 (Fig. 2B). The median percentage of the MPC component identified among cases in each subgroup correlated with the diagnostic impressions initially recorded (addressed in the second question of the survey, Fig. 2C).

We calculated the average percentage reported for each criterion in

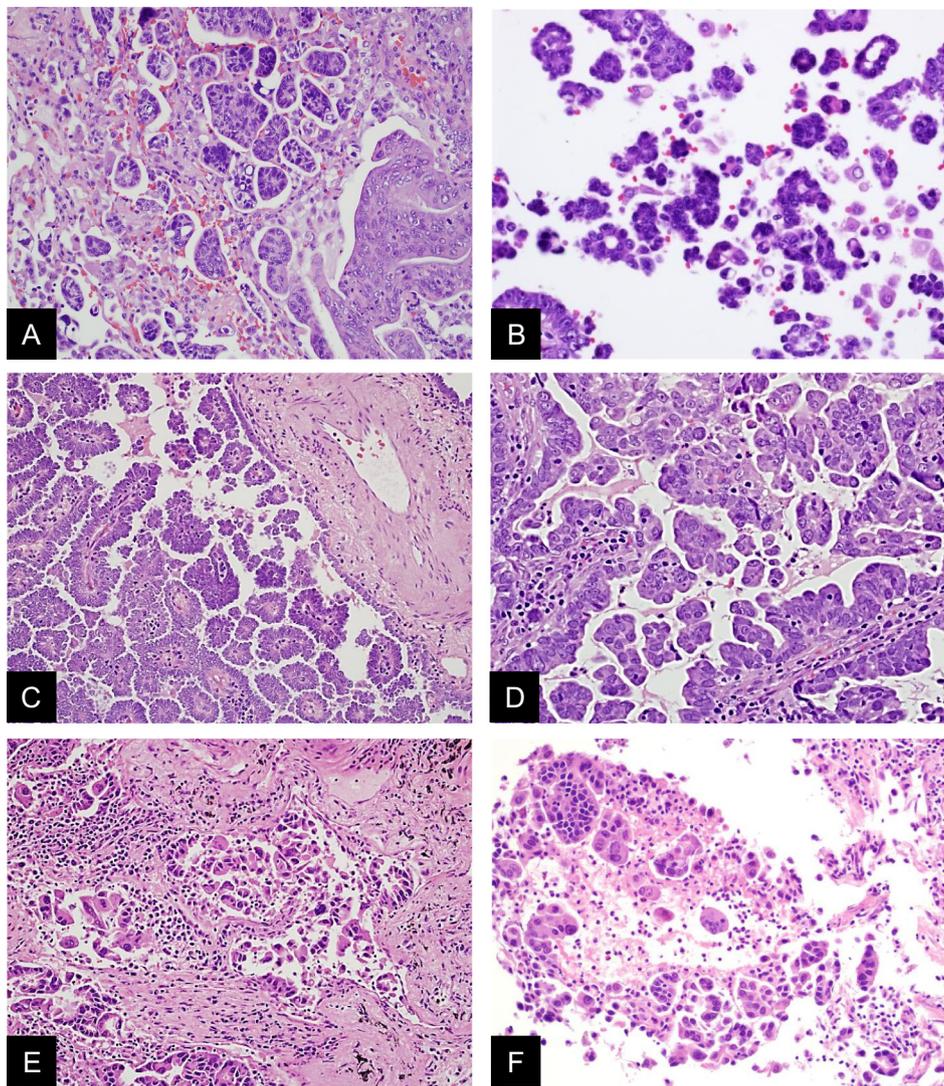


Fig. 1. Digital images of representative tumors chosen for the study are shown in A-F. Cases with unequivocal areas of micropapillary morphology were selected (A-B). Cases with admixed morphologies, such as papillary and micropapillary, were also selected (C-D). Additionally, tumors with challenging areas “suspicious for micropapillary” were selected for the study (E-F).

the subgroups (MPC0, MPC1, MPC2) for the 17 morphological features (addressed in the third question of the survey; Table 2). We further selected the morphological criteria with reported averages that were significantly different between at least two hierarchical subgroups ($n = 8$), and we eliminated those that overlapped in all three ($n = 9$) (Table 3). Our results showed that the percentage of MPC gradually increased in the hierarchical subgroups (MPC2 > MPC1 > MPC0), and 4 morphological criteria gradually increased within the same subgroups: “intracytoplasmic vacuolization”, “multiple nests in same lacunar/alveolar space”, “small tumor nest size (< 4 cells)”, and “medium tumor nest size (< 12 cells)” (the former 3 were seen in MPC1 and MPC2 in $\geq 50\%$ of the cases; $p = 0.048, 0.005, 0.001, 0.004$, respectively; Fig. 3A-D). In addition, the only criterion that was present almost exclusively within the MPC2 subgroup (predominantly MPC), in > 50% of the cases, was “epithelial nest anastomosis/confluence” (Fig. 3E).

We further evaluated the frequency of morphological criteria in groups with MPC histology (MPC1 and 2) as compared to MPC0 (small to no MPC component). Our results showed that “peripherally oriented nuclei” and “epithelial signet ring-like forms” were more commonly seen in MPC1 and MPC2, and were low to absent in the MPC0 subgroup ($p = 0.002, 0.047$, respectively). The detected percentage for both

criteria did not differ significantly within the MPC1 and MPC2 groups (Tables 2 and 3). On a separate note, we identified that “central fibrovascular core” was present almost exclusively in the MCP subgroup (30% in MPC1, vs. 0–10% in MPC0 and MPC2, $p \leq 0.001$) (Table 2). Furthermore, cluster analysis, using the 8 morphological criteria, with significant mean differences between sub-groups in Table 2 yielded a dendrogram that also separated our 30 cases into 3 sub-groups (Fig. 4). The subgroups reported by the clustering analysis of diagnostic impression (Fig. 2A) and morphological criteria (Table 3) were in substantial agreement with each other (Kappa = 0.631, [CI, 0.420–0.843]).

Finally, we arbitrarily divided our reviewers into 2 groups based on their years of experience in the field. The groups were designated as “less than 15 years” vs. “15 years or more”. We then assessed their diagnostic impressions among the tumor cases (Fig. 5). Statistical analysis showed that the overall percent of agreement reached within the pathologists of each group was significantly high and similar among the pathologists with less vs. > 15 years of experience: 50.5% vs. 41.3% (CI, 48.8–52.4; CI, 40.0–42.8, respectively).

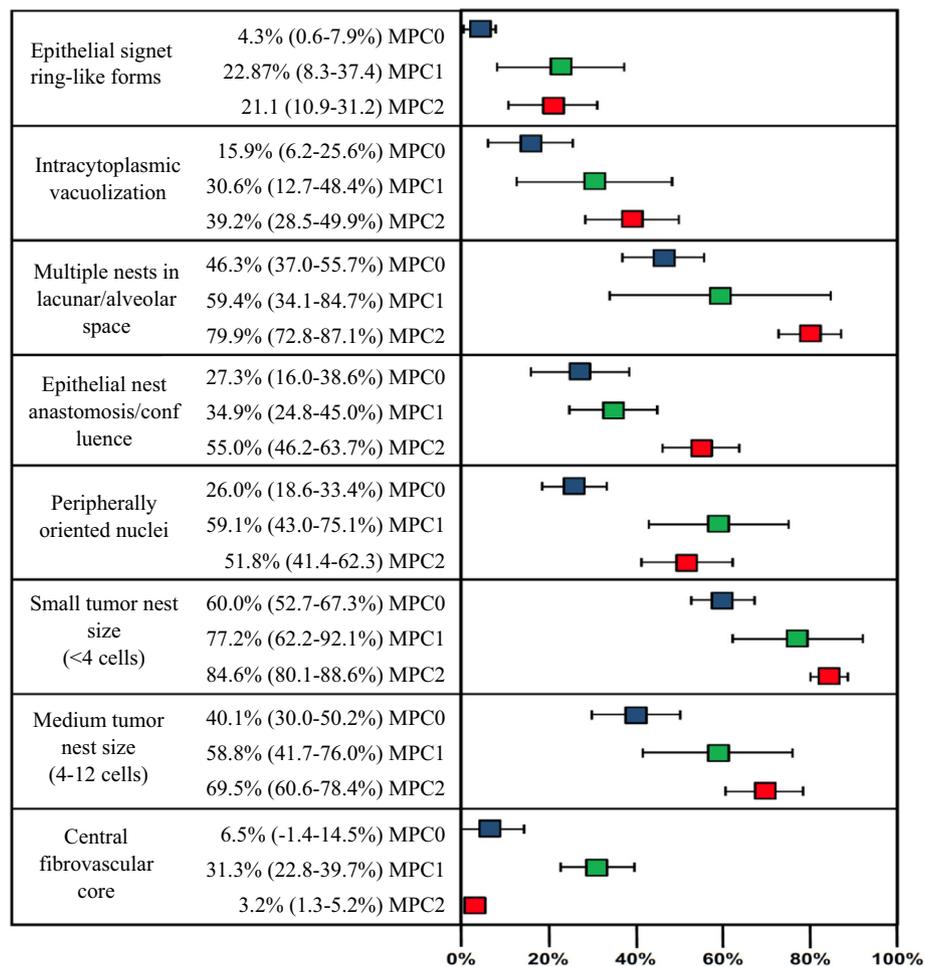
Table 2

Mean percentage of morphological features reported by pathologists as present per hierarchical subgroups. Values in parenthesis highlight the confidence intervals of the averages.

Morphologic criteria	MPC0 (n = 9)	MPC1 (n = 8)	MPC2 (n = 13)
Columnar cells	36.5% (26.3–46.9%)	38.0% (32.1–43.9%)	26.5% (21.0–32.0%)
Elongated slender cell nests processes	32.3% (23.2–41.3%)	42.2% (30.8–53.5%)	38.0% (32.0–44.0%)
Extensive stromal retraction	3.4% (0.8–5.9%)	7.1% (1.4–12.8%)	16.8% (5.8%–27.8%)
Lumen formation with internal epithelial tufting	26.2% (18.0–34.4%)	43.5% (22.9–64.2%)	36.8% (28.0–45.7%)
Epithelial signet ring-like forms	4.3% (0.6–7.9%)	22.8% (8.3–37.4%)	21.1% (10.9–31.2%)
Intracytoplasmic vacuolization	15.9% (6.2–25.6%)	30.6% (12.7–48.4%)	39.2% (28.5–49.9%)
Multiple nests in same space	46.3% (37.0–55.7%)	59.4% (34.1–84.7%)	79.9% (72.8–87.1%)
Back-to-back lacunar spaces	9.7% (6.3–13.0%)	24.0% (8.7–39.3%)	29.4% (13.1–45.8%)
Epithelial nest anastomosis/confluence	27.3% (16.0–38.6%)	34.9% (24.8–45.0%)	55.0% (46.2–63.78%)
Marked nuclear pleomorphism	31.4% (19.4–43.4%)	33.0% (23.5–42.5%)	35.0% (27.4–42.5%)
Peripherally oriented nuclei	26.0% (18.6–33.4%)	59.1% (43.0–75.1%)	51.8% (41.4–62.3%)
Randomly distributed nuclei	57.8% (48.9–66.7%)	43.2% (27.0–59.4%)	68.7% (62.0–75.3%)
Small tumor nest size	60.0% (52.7–67.3%)	77.2% (62.2–92.1%)	84.6% (80.1–88.6%)
Medium tumor nest size (4–12 cells)	40.1% (30.0–50.2%)	58.8% (41.7–76.0%)	69.5% (60.6–78.4%)
Large tumor nest size (> 12 cells)	9.6% (0–19.3%)	21.7% (11.9–31.6%)	29.6% (18.9–40.3%)
Nest cells central fibrovascular core	6.5% (-1.4–14.5%)	31.3% (22.8–39.7%)	3.2% (1.3%–5.2%)
STAS (spread through airspaces)	49.9% (37.6–62.1%)	51.9% (44.4–59.3%)	43.4% (34.1–52.8%)

Table 3

Mean percentage of the 8 morphologic criteria with significant differences among the hierarchical subgroups.



multiple nests in same lacunar/alveolar space, small tumor nest size, and medium tumor nest size. In particular, the last 4 criteria were seen in $\geq 30\%$ of the cases within the MPC1 subgroup, and $\geq 55\%$ of the cases within the MPC2 subgroup, suggesting that their presence within an adenocarcinoma of the lung is associated or linked to MPC histology. The cases with MPC histology were also associated with

intracytoplasmic vacuolization; however, this occurred less frequently (approximately 30% in MPC1 and 40% in MPC2). Epithelial nest anastomosis/confluence seemed to be the least reliable criterion to distinguish between tumors with admixed histological patterns, including MPC and papillary (MPC1 subgroup), and tumors without an MPC component (MPC0, referred to as “others” in our study).

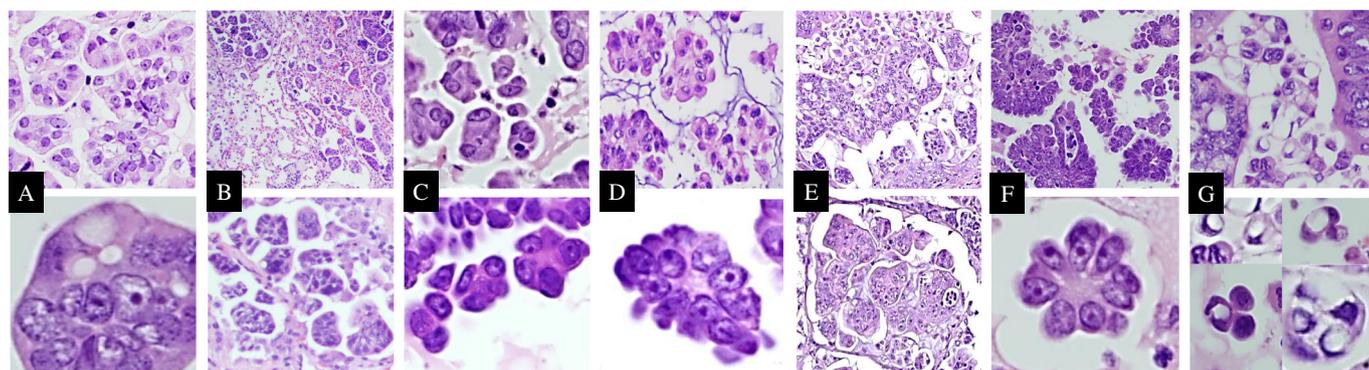


Fig. 3. The most commonly identified morphological criteria associated with diagnosis of MPC (both MPC1 and MPC2) were: intracytoplasmic vacuolization (A), multiple nests in same lacunar/alveolar space (B), small tumor nest size (< 4 cells) (C), medium tumor nest size (< 12 cells) (D), epithelial nest anastomosis/confluence (E), peripherally oriented nuclei (F), and epithelial signet ring-like forms (G).

On another note, the criteria “peripherally oriented nuclei” and “epithelial signet ring-like forms” were encountered almost equally in the MPC1-2 subgroups (percentages were not significantly different), and were nearly absent in MPC0. However, the former criterion was identified more frequently in the MPC1–2 groups ($\geq 50\%$) compared to MPC0 (20–25%). This suggests that the presence of “peripherally oriented nuclei” is a slightly more reliable criterion to diagnose a tumor with a component of MPC of lung and/or with MPC as a unique histological pattern. Additionally, our results showed that the presence of a central fibrovascular core favors diagnosis of a tumor with papillary morphology (a feature considered to be characteristic of this histological variant) but is not helpful to distinguish micropapillary from other adenocarcinoma types.

The internal validity of our results is supported by a strong agreement of diagnostic sub-groups, predicted by two different cluster analyses based on two independent criteria: morphological and diagnostic interpretation. These results were independent of the years of experience that each individual pathologist had in the field.

Our study had certain limiting factors that could have caused significant bias, for example, the fact that the reviewers knew that the

study aimed to improve diagnosis of MPC of lung. Also, the reproducibility of our results could have been impacted because an exact definition or illustrative image for each criterion was not provided. Additionally, we were unable to determine the sensitivity/specificity of our criteria (which have been assessed in similar studies in the past) [5]. This limitation is attributed to the fact that currently, neither a gold standard nor an objective assessment to determine whether a case has or does not have MPC histology can be found in the literature (hence the importance of this study).

Other constraints of our study include the limited heterogeneity observed between the dataset among raters. This did not allow us to reliably trust the Fleiss' kappa function in most of our statistical analyses (executed with the “kappa.fleiss” in R). Low heterogeneity can influence the formula to calculate Fleiss' kappa, creating a paradoxical result that does not correlate with the real agreement [15]. Homogeneity of the data increases the probability of responses arising by chance when applying this statistical tool (this topic has been extensively studied in the past, and further details are beyond the scope of this manuscript) [38-39]. Thus, in our cases, we decided to report OPA to show the high concordance that was truly seen between raters.

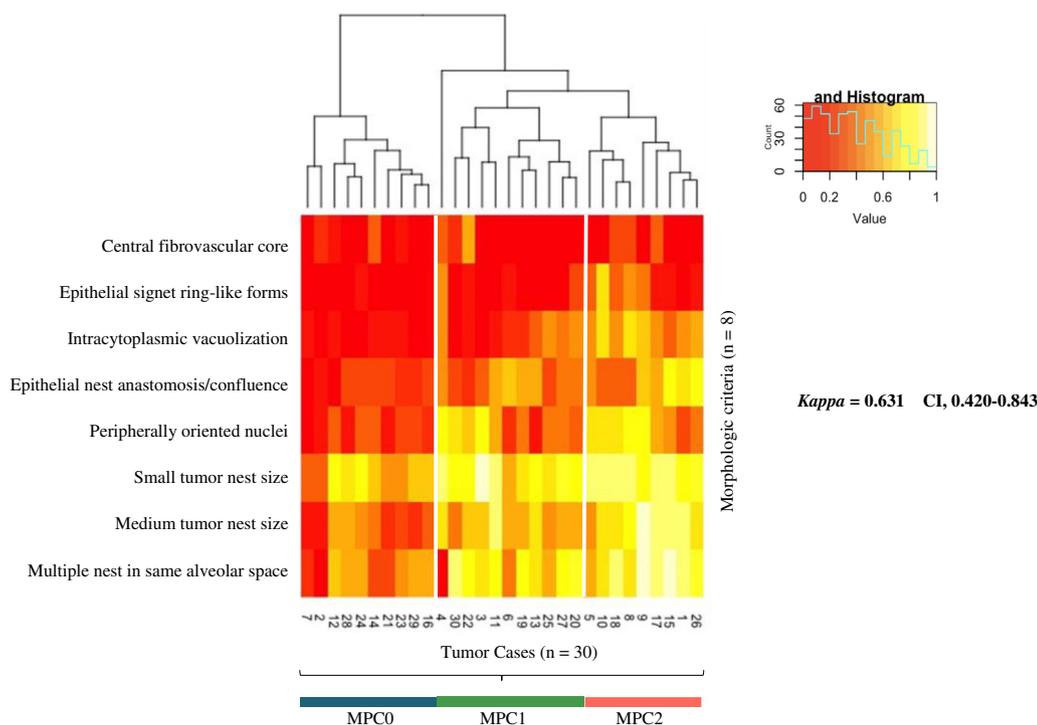


Fig. 4. A total of 8/17 morphological criteria demonstrated significant differences among hierarchical sub-groups. These criteria were compared with the diagnostic interpretations of the pathologists through hierarchical clustering analysis. Results from the dendrogram demonstrated strong agreement among the subgroups from the hierarchical cluster based on both diagnostic impression and the reported morphological criteria (Kappa = 0.631 CI, 0.420–0.843).

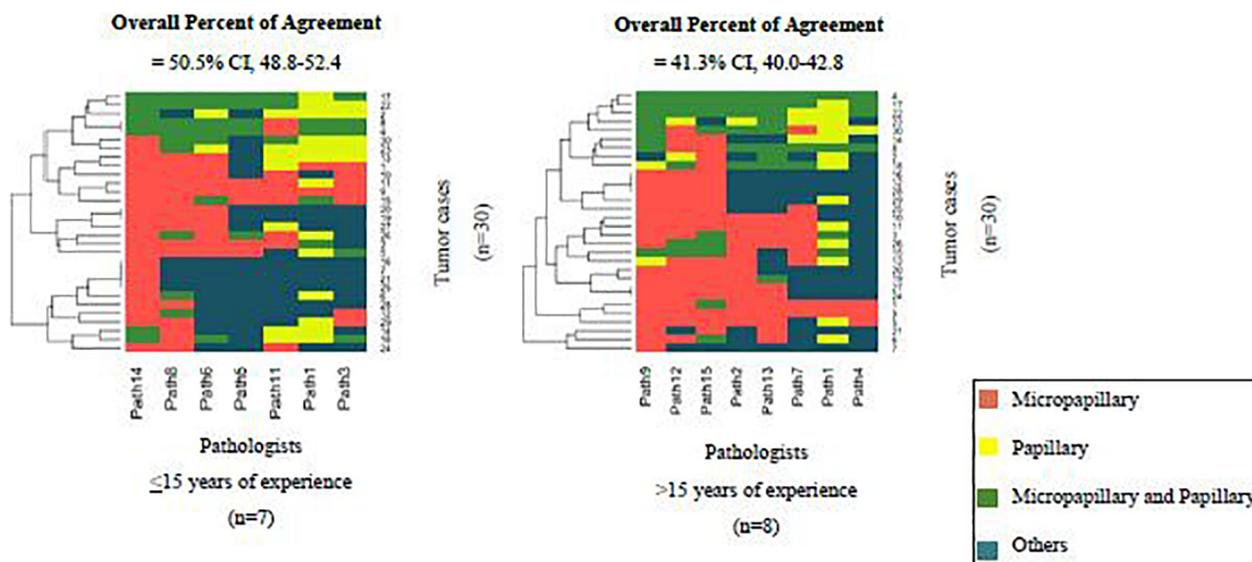


Fig. 5. Hierarchical clustering analysis of the diagnostic interpretations of pulmonary pathologists with 15 years of experience or less (50.5%) vs. pathologists with > 15 years of experience in the field (41.3%) showed that no significant differences existed between the groups.

Another example showing the absence of higher heterogeneity in our study was the use of narrow/limited options for diagnostic interpretations. Our design preset the answer to our first question to four specific variants: MPC, papillary, mixed papillary and MPC, or others. More so, the pathologists were provided with two pictures of representative areas selected of each slide. This could have biased the pathologists towards a particular diagnosis, and could have influenced the number of cases diagnosed with MPC and papillary components. By not providing additional options for mixed histological patterns (other than papillary mixed with MPC), we could have not taken into account MPC cases exhibiting mixed patterns with non-papillary variants such as acinar, lepidic or solid. This is evidenced by the rare cases within the MPC0 group, for which a percentage of MPC was provided in the second question of the survey, even though diagnosis of “others” had been selected.

Importantly, we had to take into consideration in the interpretation of our results the fact that the static digital images used in the study showed a limited area of tumor; and that we decided to also include pictures of particular areas that may have not been classic of the main tumor diagnosis. Our goal was not to assess consistent reproducibility of the correct diagnosis for the cases included in the study (whole slide imaging would have been a more sensitive technique to achieve this). Moreover, we aimed to address the opinion of experts regarding particular tumor areas including some where the diagnosis could have been. These areas spanned a morphological spectrum that included MPC of lung mimics, MPC of lung mixed with papillary, and areas with more “classic” MPC.

Despite these potential limitations, our study provides an unprecedented, strong, multi-institutional, detailed diagnostic approach among experts in the field; we believe that this study will assist the identification of MPC of the lung. Given that currently there is no gold standard to diagnose this entity in the lung, our study aimed to address the morphological criteria encountered in MPC of lung cases in which pulmonary pathologists agreed on the diagnoses.

Several interesting issues remain to be addressed in future studies, notwithstanding the fact that they are beyond the scope of this study. For example, intra-observer variability amongst pathologists who diagnosed the same cases over time. This parameter would objectively test consistency, precision, and internal reproducibility more thoroughly. More so, it would have been interesting to determine the association of clinicopathological associations and overall prognosis to individual morphological features observed in MPC of lung.

Additionally, it would be interesting to assess the interpretation of cases and compare the reproducibility and overall percent of agreement among pathologists, after proper training sessions regarding diagnostic morphological features have been evaluated in the study. Also, it is notable that papillary lung adenocarcinoma showed a tendency to cluster with MPC; this illustrates the challenge to separate the two entities in cases with limited tissue sampling. Finally, considering the genotypic correlation and examining driver mutations and lineage markers in tumors with micropapillary morphology among lung adenocarcinomas has relevant research potential. Further studies are necessary to address this.

In summary, our study showed that the significant features for MPC diagnosis are: “multiple nests in same lacuna or alveolar space”, “small to medium tumor nest size”, and “epithelial nests anastomosis/confluence”. In addition, “intracytoplasmic vacuolization” could also be useful; however, it was not identified in a significant number of the cases in this study, and thus we did not consider it as important as the other criteria. Also, “peripherally oriented nuclei” and “epithelial signet ring-like forms” can be useful in cases with admixed tumor types. Finally, as it has been previously described, our study showed that the presence of fibrovascular cores corresponds to diagnosis of papillary adenocarcinoma and not MPC. However, it is possible that this feature can be a confounding factor favoring sole papillary adenocarcinoma diagnosis in cases with mixed papillary and MPC histology. In these cases, we suggest searching for additional features such as “peripherally oriented nuclei” and “epithelial signet ring-like forms”. These two additional features were commonly seen in the MPC1 (mixed papillary and MPC) subgroup of our study, and could prove to be useful.

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Disclosure

The authors of this manuscript have no disclosures.

References

[1] Bertz S, Wach S, Taubert H, et al. Micropapillary morphology is an indicator of poor

- prognosis in patients with urothelial carcinoma treated with transurethral resection and radiochemotherapy. *Virchows Arch* 2016;469(3):339–44.
- [2] Barresi V, Branca G, Vitarelli E, Tuccari G. Micropapillary pattern and poorly differentiated clusters represent the same biological phenomenon in colorectal cancer: a proposal for a change in terminology. *Am J Clin Pathol* 2014;142(3):375–83.
 - [3] Eom DW, Kang GH, Han SH, et al. Gastric micropapillary carcinoma: a distinct subtype with a significantly worse prognosis in TNM stages I and II. *Am J Surg Pathol* 2011;35(1):84–91.
 - [4] Wu Y, Zhang N, Yang Q. The prognosis of invasive micropapillary carcinoma compared with invasive ductal carcinoma in the breast: a meta-analysis. *BMC Cancer* 2017;17(1):839–48.
 - [5] Sangoi AR, Beck AH, Amin MB, et al. Interobserver reproducibility in the diagnosis of invasive micropapillary carcinoma of the urinary tract among urologic pathologists. *Am J Surg Pathol* 2010;34(9):1367–76.
 - [6] Amin MB, Tamboli P, Merchant SH, et al. Micropapillary variant of transitional cell carcinoma of the urinary bladder. Histologic pattern resembling ovarian papillary serous carcinoma. *Am J Surg Pathol* 1994;18:1224–32.
 - [7] Nagao T, Gaffey TA, Visscher DW, et al. Invasive micropapillary salivary duct carcinoma: a distinct histologic variant with biologic significance. *Am J Surg Pathol* 2004;28(3):319–26.
 - [8] Nassar H. Carcinomas with micropapillary morphology: clinical significance and current concepts. *Adv Anat Pathol* 2004;11(6):297–303.
 - [9] Sakamoto K, Watanabe M, De La Cruz C, et al. Primary invasive micropapillary carcinoma of the colon. *Histopathology* 2005;47(5):479–84.
 - [10] Sriaunkul S, Tavassoli FA. Invasive micropapillary carcinoma of the breast. *Mod Pathol* 1993;6(6):660–2.
 - [11] Amin MB, Tamboli P, Merchant SH, et al. Micropapillary component in lung adenocarcinoma: a distinctive histologic feature with possible prognostic significance. *Am J Surg Pathol* 2002;26(3):358–64.
 - [12] Yanagawa N, Shiono S, Abiko M, Katahira M, Osakabe M, Ogata SY. The clinical impact of solid and micropapillary patterns in resected lung adenocarcinoma. *J Thorac Oncol* 2016;11(11):1976–83.
 - [13] Zhao Y, Wang R, Shen X, et al. Minor components of micropapillary and solid subtypes in lung adenocarcinoma are predictors of lymph node metastasis and poor prognosis. *Ann Surg Oncol* 2016;23(6):2099–105.
 - [14] Lee MC, Buitrago DH, Kadota K, Jones DR, Adusumilli PS. Recent advances and clinical implications of the micropapillary histological subtype in lung adenocarcinomas. *Lung Cancer Manag* 2014;1(3(3)):245–53.
 - [15] Moon Y, Kim KS, Sung SW, et al. Correlation of histological components with tumor invasion in pulmonary adenocarcinoma. *World J Surg Oncol* 2014;17(12):388.
 - [16] Tsutsumida H, Nomoto M, Goto M, et al. A micropapillary pattern is predictive of a poor prognosis in lung adenocarcinoma, and reduced surfactant apoprotein A expression in the micropapillary pattern is an excellent indicator of a poor prognosis. *Mod Pathol* 2007;20(6):638.
 - [17] Pyo JS, Kim JH. Clinicopathological significance of micropapillary pattern in lung adenocarcinoma. *Pathol Oncol Res* 2017;6.
 - [18] Makimoto Y, Nabeshima K, Iwasaki H, et al. Micropapillary pattern: a distinct pathological marker to subclassify tumours with a significantly poor prognosis within small peripheral lung adenocarcinoma (< / = 20 mm) with mixed bronchioalveolar and invasive subtypes (Noguchi's type C tumours). *Histopathology* 2005;46(6):677–84.
 - [19] Kamiya K, Hayashi Y, Douguchi J, et al. Histopathological features and prognostic significance of the micropapillary pattern in lung adenocarcinoma. *Mod Pathol* 2008;21(8):992–1001.
 - [20] Hung JJ, Yeb YC, Jeng WJ, et al. Prognostic factors of survival after recurrence in patients with resected lung adenocarcinoma. *J Thorac Oncol* 2015;10(9):1328–36.
 - [21] Xu CH, Wang W, Wei Y, et al. Prognostic value of the new International Association for the Study of Lung Cancer/American Thoracic Society/European Respiratory Society classification in stage IB lung adenocarcinoma. *Eur J Surg Oncol* 2015;41(10):1430–6.
 - [22] Lee G, Lee HY, Jeong JY, et al. Clinical impact of minimal micropapillary pattern in invasive lung adenocarcinoma: prognostic significance and survival outcomes. *Am J Surg Pathol* 2015;39(5):660–6.
 - [23] Cha MJ, Lee HY, Lee KS, et al. Micropapillary and solid subtypes of invasive lung adenocarcinoma: clinical predictors of histopathology and outcome. *J Thorac Cardiovasc Surg* 2014;147(3):921–928.e2.
 - [24] Zhao ZR, To KF, Mok TS, Ng CS. Is there significance in identification of non-predominant micropapillary or solid components in early-stage lung adenocarcinoma? *Interact Cardiovasc Thorac Surg* 2017;24(1):121–5.
 - [25] Sumiyoshi S, Yoshizawa A, Sonobe M, et al. Pulmonary adenocarcinomas with micropapillary component significantly correlate with recurrence, but can be well controlled with EGFR tyrosine kinase inhibitors in the early stages. *Lung Cancer* 2013;81(1):53–9.
 - [26] Zhao W, Wang H, Xie J, Tian B. A clinicopathological study of small lung adenocarcinoma 1 cm or less in size: emphasis on histological subtypes associated with lymph node metastasis and recurrence. *Int J Surg Pathol* 2018;26(1):4–11. Feb.
 - [27] Yi E, Bae MK, Cho S, Chung JH, Jheon S, Kim K. Pathological prognostic factors of recurrence in early stage lung adenocarcinoma. *ANZ J Surg* 2018;88(4):327–31. <https://doi.org/10.1111/ans.14033>. April.
 - [28] Hung JJ, Jeng WJ, Chou TY, et al. Prognostic value of the new International Association for the Study of Lung Cancer/American Thoracic Society/European Respiratory Society lung adenocarcinoma classification on death and recurrence in completely resected stage I lung adenocarcinoma. *Ann Surg* 2013;258(6):1079–86.
 - [29] Luna-More S, Gonzalez B, Acedo C, Rodrigo I, Luna C. Invasive micropapillary carcinoma of breast. A new special type of invasive mammary carcinoma. *Pathol Res Pract* 1995;190:668–74.
 - [30] Johansson SL, Borghede G, Holmang S. Micropapillary bladder carcinoma: a clinicopathological study of 20 cases. *J Urol* 1999;161:1798–802.
 - [31] De La Cruz C, Moriya T, Endoh M, et al. Invasive micropapillary carcinoma of the breast: clinicopathological and immunohistochemical study. *Pathol Int* 2004;44:18–23.
 - [32] Pettinato G, Manivel MJ, Panico L, Sparano L, Petrella G. Invasive micropapillary carcinoma of the breast: clinicopathologic study of 62 cases of a poorly recognized variant with highly aggressive behavior. *Am J Clin Pathol* 2004;2004(121):851–66.
 - [33] Hayakawa T, Tajima S, Tsukui M, Takanashi Y, Neyatani H, Funai K. Stromal micropapillary predominant lung adenocarcinoma: a rare histological phenotype with poor prognosis. *Respir Case Rep* 2016;4(6):e00203.
 - [34] Dai C, Xie H, Kadeer X, et al. Relationship of lymph node micrometastasis and micropapillary component and their joint influence on prognosis of patients with Stage I lung adenocarcinoma. *Am J Surg Pathol* 2017;41(9):1212–20.
 - [35] Haryju T, Wakahara M, Matsuoka Y, et al. Clinicopathological characteristics of lung adenocarcinoma with unexpected lymph node metastasis. *Ann Thorac Cardiovasc Surg* 2017;23(4):181–7.
 - [36] Yoshida Y, Nitadori JI, Shinozaki-Ushiku A, et al. Micropapillary histological subtype in lung adenocarcinoma of 2 cm or less: impact on recurrence and clinical predictors. *Gen Thorac Cardiovasc Surg* 2017;65(5):273–9.
 - [37] Ohe M, Yokose T, Sakuma Y, et al. Stromal micropapillary component as a novel unfavorable prognostic factor of lung adenocarcinoma. *Diagn Pathol* 2012;7:3.
 - [38] Marasini D, Quatto P, Ripamonti E. Assessing the inter-rater agreement for ordinal data through weighted indexes. *Stat Methods Med Res* 2016;25(6):2611–33.
 - [39] Feinstein AR, Cicchetti DV. High agreement but low kappa: I. the problems of two paradoxes. *J Clin Epidemiol* 1990;43(6):543–9.