

Microglia, Monocytes, and the Recurrence of Anxiety in Stress-Sensitized Mice

To the Editor:

We read with great interest the article by Weber *et al.* (1) in *Biological Psychiatry* describing the effects of microglia elimination and repopulation on stress sensitization induced by repeated social defeat (RSD). The article highlights brain-immune interactions and, in particular, the importance of stress-primed microglia for monocyte accumulation in the brain of RSD-sensitized mice following acute stress. The transcriptomic analysis of microglia 24 days after RSD could be very useful to other researchers, so the authors may wish to make this information accessible to the community by depositing it to an appropriate data repository.

Several earlier studies by the same group have established robust trafficking of myeloid cells to the brain as a consequence of RSD (2–5). Importantly, the authors have previously reported that transcatheter perfusion did not affect the number of Ly6C^{hi} macrophages in the central nervous system after RSD (2). If replicated, this exciting finding would strongly suggest that prolonged stress over a span of several days increases the presence of blood-derived cells either in the perivascular space or in the brain parenchyma (2). At this time, the fate of these engrafted cells in the brain remains to be further elucidated.

In the current study, the authors focus on the effects of a new cycle of acute defeat. They show that the number of CD11b⁺/CD45^{hi} cells in brain was more than doubled 14 hours after acute defeat in stress-sensitized mice, i.e., mice that had undergone the RSD procedure 24 days previously. Stress-naïve mice subjected to one episode of acute defeat served as controls (1). It should be noted in this context that 14 hours seems like a relatively short period of time for blood-derived cells to engraft the central nervous system, especially as compared with more severe insults with breakdown of the blood-brain barrier such as focal brain ischemia (6). As data on stress-sensitized mice without a new cycle of acute defeat are missing, it is difficult to judge the contribution of previously engrafted CD11b⁺/CD45^{hi} cells to the increased number of these cells observed here after acute defeat. Again, it would be good if the authors could clarify if brains were perfused.

In line with two earlier reports (7,8), the authors find that the repopulated microglia after treatment with PLX5622 originate from the small number of microglia surviving in the brain (1). However, other studies have yielded strong evidence in support of a key role for circulating monocytes in repopulating the microglial niche (9,10), so the special effects of PLX5622 may not be generalizable to microglia depletion per se. The underlying data obtained by Weber *et al.* (1) from CX₃CR1^{CreER-YFP}/R26^{tdTOM} mice also raise a number of issues. First, the authors report that 46% of CD11b⁺/CD45^{hi} cells isolated from tamoxifen-injected mice after microglial elimination and repopulation lacked yellow fluorescent protein. This lack of yellow fluorescent protein indicates that, at least after treatment with PLX5622, CD11b⁺/CD45^{hi} cells form a quite heterogeneous

population and suggests that the presence of other leukocyte subsets besides macrophages should be investigated (e.g., expression of markers for T cells, natural killer cells, and granulocytes). Second, because of the leakiness of the Cre recombinase acknowledged by the authors, we would rather caution against drawing any firm inferences. In this respect, we also note that additional information from the microglia elimination and repopulation experiment concerning yellow fluorescent protein and tdTOM expression in brain macrophages harvested from CX₃CR1^{CreER-YFP}/R26^{tdTOM} mice not treated with tamoxifen would have been very valuable. Finally, and more generally, parameters such as the common CD45^{hi/lo} expression by flow cytometry may conceivably change with injury, treatment, and disease (11,12).

Although blood-derived brain macrophages and resident microglia share many similarities, recent research has highlighted important differences between these two cell types that may be directly relevant to disease outcomes (13–15). By showing that primed microglia and blood-derived cells cooperate to rekindle and augment anxiety, the authors add a new twist to this evolving story. It will be interesting to further delimit the precise roles played by myeloid cells in stress-induced anxiety.

Golo Kronenberg
Ria Uhlemann
Matthias Endres
Karen Gertz

Acknowledgments and Disclosures

This work was supported by the Deutsche Forschungsgemeinschaft (Grant Nos. SFB TRR43 and Exc257 [to ME], Grant No. KR 2956/4-1 [to GK], and Grant No. 2576/2-1 [to KG]), Bundesministerium für Bildung und Forschung (Grant No. CSB 01 EO 1301 [to ME, KG, and GK]), German Center for Neurodegenerative Diseases (to ME), German Center for Cardiovascular Research (to ME), and Corona Foundation (to ME).

The authors report no biomedical financial interests or potential conflicts of interest.

Article Information

From the University of Leicester and Leicestershire Partnership NHS Trust (GK), Leicester, Leicestershire, United Kingdom; and Charité – Universitätsmedizin Berlin (RU, ME, KG), corporate member of Freie Universität Berlin, Humboldt-Universität zu Berlin, and Berlin Institute of Health, Klinik und Hochschulambulanz für Neurologie und Centrum für Schlaganfallforschung Berlin, Berlin, Germany.

Address correspondence to Karen Gertz, M.D., Klinik und Hochschulambulanz für Neurologie, Abteilung für Experimentelle Neurologie und Centrum für Schlaganfallforschung Berlin, Charité – Universitätsmedizin Berlin, Charitéplatz 1, Berlin 10117, Germany; E-mail: karen.gertz@charite.de.

See also associated correspondence: <https://doi.org/10.1016/j.biopsych.2019.01.026>.

Received Nov 30, 2018; accepted Jan 30, 2019.

References

1. Weber MD, McKim DB, Niraula A, Witcher KG, Yin W, Sobol CG, *et al.* (2019): The influence of microglial elimination and repopulation on stress sensitization induced by repeated social defeat. *Biol Psychiatry* 85:667–678.

2. Wohleb ES, Hanke ML, Corona AW, Powell ND, Stiner LM, Bailey MT, *et al.* (2011): Beta-adrenergic receptor antagonism prevents anxiety-like behavior and microglial reactivity induced by repeated social defeat. *J Neurosci* 31:6277–6288.
3. Wohleb ES, Powell ND, Godbout JP, Sheridan JF (2013): Stress-induced recruitment of bone marrow-derived monocytes to the brain promotes anxiety-like behavior. *J Neurosci* 33:13820–13833.
4. Wohleb ES, Patterson JM, Sharma V, Quan N, Godbout JP, Sheridan JF (2014): Knockdown of interleukin-1 receptor type-1 on endothelial cells attenuated stress-induced neuroinflammation and prevented anxiety-like behavior. *J Neurosci* 34:2583–2591.
5. McKim DB, Weber MD, Niraula A, Sawicki CM, Liu X, Jarrett BL, *et al.* (2018): Microglial recruitment of IL-1beta-producing monocytes to brain endothelium causes stress-induced anxiety. *Mol Psychiatry* 23:1421–1431.
6. Gertz K, Kronenberg G, Kalin RE, Baldinger T, Werner C, Balkaya M, *et al.* (2012): Essential role of interleukin-6 in post-stroke angiogenesis. *Brain* 135:1964–1980.
7. Elmore MR, Najafi AR, Koike MA, Dagher NN, Spangenberg EE, Rice RA, *et al.* (2014): Colony-stimulating factor 1 receptor signaling is necessary for microglia viability, unmasking a microglia progenitor cell in the adult brain. *Neuron* 82:380–397.
8. Cronk JC, Filiano AJ, Louveau A, Marin I, Marsh R, Ji E, *et al.* (2018): Peripherally derived macrophages can engraft the brain independent of irradiation and maintain an identity distinct from microglia. *J Exp Med* 215:1627–1647.
9. Varvel NH, Grathwohl SA, Baumann F, Liebig C, Bosch A, Brawek B, *et al.* (2012): Microglial repopulation model reveals a robust homeostatic process for replacing CNS myeloid cells. *Proc Natl Acad Sci U S A* 109:18150–18155.
10. Lund H, Pieber M, Parsa R, Han J, Grommisch D, Ewing E, *et al.* (2018): Competitive repopulation of an empty microglial niche yields functionally distinct subsets of microglia-like cells. *Nat Commun* 9:4845.
11. Bennett ML, Bennett FC, Liddelov SA, Ajami B, Zamanian JL, Fernhoff NB, *et al.* (2016): New tools for studying microglia in the mouse and human CNS. *Proc Natl Acad Sci U S A* 113:E1738–1746.
12. Trahanas DM, Cuda CM, Perlman H, Schwulst SJ (2015): Differential activation of infiltrating monocyte-derived cells after mild and severe traumatic brain injury. *Shock* 43:255–260.
13. Kronenberg G, Uhlemann R, Richter N, Klempin F, Wegner S, Staerck L, *et al.* (2018): Distinguishing features of microglia- and monocyte-derived macrophages after stroke. *Acta Neuropathol* 135:551–568.
14. Varvel NH, Neher JJ, Bosch A, Wang W, Ransohoff RM, Miller RJ, *et al.* (2016): Infiltrating monocytes promote brain inflammation and exacerbate neuronal damage after status epilepticus. *Proc Natl Acad Sci U S A* 113:E5665–5674.
15. Yamasaki R, Lu H, Butovsky O, Ohno N, Rietsch AM, Cialic R, *et al.* (2014): Differential roles of microglia and monocytes in the inflamed central nervous system. *J Exp Med* 211:1533–1549.