



# Association of long noncoding RNA H19 polymorphisms with the susceptibility and clinical features of ischemic stroke in southern Chinese Han population

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## Abstract

Stroke is the leading cause of death in China. Previous studies have demonstrated that long noncoding RNAs play important roles in ischemic stroke (IS). This study aimed to investigate long noncoding RNA H19 (lncRNA H19) expression in IS cases and the association between lncRNA H19 variants and IS risk and IS-related risk factors. A total of 550 IS cases and 550 controls were recruited for this study. LncRNA H19 expression was detected using quantitative real-time polymerase chain reaction. Genotyping was conducted by the Sequenom MassARRAY technology. LncRNA H19 level in peripheral blood of IS cases was significantly upregulated compared with healthy controls ( $P = 0.046$ ). No significant association was observed between lncRNA H19 rs217727 and rs4929984 polymorphisms with IS risk in all genetic models, and rs217727-rs4929984 haplotypes are not associated with IS susceptibility. Further meta-analysis also implied that the rs217727 and rs4929984 polymorphisms were not associated with IS in Chinese population. However, rs4929984 is significantly associated with the diastolic blood pressure level of IS patients (additive model:  $P_{\text{adj}} = 0.007$ ; dominant model:  $P_{\text{adj}} = 0.013$ ), whereas rs217727 is associated with international normalized ratio (additive model:  $P_{\text{adj}} = 0.019$ ; recessive model:  $P_{\text{adj}} = 0.004$ ), prothrombin time activity level (additive model:  $P_{\text{adj}} = 0.026$ ; recessive model:  $P_{\text{adj}} = 0.004$ ), and homocysteine level (recessive model:  $P_{\text{adj}} = 0.048$ ) in patients with IS. Our findings suggest that lncRNA H19 level may affect the occurrence of IS, and lncRNA H19 variants may influence blood pressure, coagulation function, and homocysteine metabolism of patients with IS in the southern Chinese Han population.

**Keywords** Long noncoding RNA H19 · Ischemic stroke · lncRNA H19 expression · Polymorphism · Risk

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Jiao Huang, Jialei Yang and Jinhong Li contributed equally to this work.

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## Introduction

Ischemic stroke (IS) accounts for approximately 87% of stroke occurrences worldwide (Mozaffarian et al. 2016), and is the third leading cause of death in developed countries (Feigin et al. 2015) and the most common cause of death in China (Chen et al. 2017). Approximately 1.12 million Chinese aged  $\geq 20$  years died of a stroke, and the age-standardized mortality rates per 100,000 person-years in aged  $\geq 20$  years were 56.5 for IS from September 1, 2012 to August 31, 2013 (Chen et al. 2017). The American Heart Association predicts that by 2030, the direct cost of caring for stroke patients will reach \$183.13 million (Ovbiagele et al. 2013), and because of high mortality, disability, and utilization of healthcare resources, stroke can bring a heavy economic cost to the world (Roy-O'Reilly and McCullough 2014). Nowadays, thrombolytic therapy is the most approved treatment for IS, but its effectiveness is limited on many strict conditions, such as in the first 3 h of thrombolysis after stroke

onset and other contraindications to treatment (Roy-O'Reilly and McCullough 2014). Thus, understanding the underlying mechanisms of IS and providing more evidence for diagnosis and therapy is urgent.

Long noncoding RNAs (lncRNAs) are a cluster of transcripts that lie in the non-protein coding regions of genes and have been the focus of considerable attention in recent years. Approximately 70%–90% of the human genome was transcribed to RNA, and lncRNAs account for more than 68% in human transcriptome (Iyer et al. 2015). Studies have provided evidence that the regulated mechanisms of lncRNAs include epigenetic, transcriptional, post-transcriptional, and translational regulations (Sun et al. 2017). In the transcriptional level, lncRNAs located in the nucleus could regulate and modify chromosomes and lead to the alteration of gene expression (Bao et al. 2018). Moreover, lncRNAs residing in the cytoplasm may act as a competing endogenous RNA for microRNAs and proteins to protect their target genes from degradation (Rashid et al. 2016). A growing number of studies have demonstrated that lncRNAs are involved in various multifactorial diseases in the human body, such as cancers, cardiovascular, autoimmune, and neurodegenerative diseases, and psychiatric disorders (Cipolla et al. 2018).

In the past years, lncRNAs have been involved in the development of IS. MALAT1, one of the first lncRNA identified to promote metastasis and proliferation of cancers, was increased significantly in oxygen-glucose deprivation (OGD) endothelial cells and middle cerebral artery occlusion mouse models of stroke and may act as a protective and healing properties of brain ischemia (Bao et al. 2018; Zhang et al. 2016). lncRNA FosDT promotes ischemic brain injury by binding chromatin-modifying proteins coREST and Sin3a to enable the formation of REST complex, providing a promising clue that lncRNA may be a potential therapy for post-stroke brain injury (Mehta et al. 2015). Moreover, lncRNA ANRIL was increased in rats with diabetes mellitus combining with cerebral infarction (Zhang et al. 2017). Increasing lncRNA ANRIL expression or mutation is also associated with stroke (Amouyel 2012). A large number of studies have shown that variants within chromosome 9p21.3, which overlaps with the lncRNA ANRIL (Kim et al. 2014) are associated with cardiovascular disease and IS risks (Chen et al. 2014; Zhang et al. 2012). For example, lncRNA ANRIL rs2383207 and rs1333049 are significantly associated with IS risk (Yang et al. 2018), and GG genotype of lncRNA ANRIL rs10757278 significantly increases stroke risk and recurrence in the Chinese population (Zhang et al. 2012), which suggests that lncRNAs may serve as a novel genetic marker for stroke.

Long noncoding RNA H19 (lncRNA H19) is a 2.3 kb RNA coded by *H19* gene, which plays a pivotal role in the embryonic development and growth control (Gabory et al. 2010). After birth, lncRNA H19 is rarely expressed in human

tissues except in the cardiac and skeletal muscles (Gabory et al. 2010). An abnormal expression of lncRNA H19 in the postnatal period was reported to be involved in tumors, such as the bladder, gastric, colorectal, and breast cancers (Luo et al. 2013; Soudyab et al. 2016; Sun et al. 2016). In addition, studies have found an aberrant re-expression of lncRNA H19 in atherosclerotic plaque (Han et al. 1996) and rat vascular smooth muscle cells after a carotid artery injury (Kim et al. 1994). lncRNA H19 is also expressed abnormally in a status of hypoxia, which works as a stimulus for cerebral ischemia and reperfusion (I/R) injury (Matouk et al. 2010). Recently, the abnormal lncRNA H19 expression is significantly correlated with the risk factors of IS, such as blood pressure (BP) (Tragante et al. 2014) and coronary artery disease (CAD) (Wang et al. 2017a). Significantly upregulated levels of lncRNA H19 in stroke patients, animals, and cell models of IS were also detected (Wang et al. 2017a, b). lncRNA H19 polymorphisms are reported to participate in the regulation of lncRNA H19 expression (Yang et al. 2015), hence, more attention has been given to those polymorphisms. For example, the single-nucleotide polymorphism (SNP) rs217727 is correlated with the susceptibility of oral squamous cell carcinoma (Guo et al. 2017), and with the risk of CAD (Gao et al. 2015).

More recently, lncRNA H19 variants (rs217727, rs2067051, rs2251375, rs492994, rs2839698 and rs10732516) were reported in Chinese case control studies (Wang et al. 2017a; Zhu et al. 2018). Wang et al. (2017a) found the rs217727 and rs4929984 variants are associated with the risk of ischemic stroke (152 IS patients and 150 controls) and the minor alleles of rs217727 (T) and rs4929984 (A) increased the risk of IS. Even though Zhu et al. (2018) obtained a significant association of rs217727 with ischemic stroke, they found the alleles of rs217727(T/C) in patients with IS were not differ from controls. These findings are compelling but still divergent. To further ascertain the roles of lncRNA H19 and its variants in the development of ischemic stroke, in the present study, we examine the lncRNA H19 expression in the patients with IS, investigate the genetic association of lncRNA H19 SNPs rs217727 and rs4929984 with IS susceptibility and the relationship between lncRNA H19 variants and clinical parameters of patients with IS in Southern Chinese Han Population.

## Material and methods

### Subjects

A total of 550 unrelated IS patients and 550 unrelated healthy control subjects were obtained from the First Affiliated Hospital of Guangxi University of Chinese Medicine from September 2010 to June 2017. All subjects were of Han Chinese descent. According to the Fourth National

Conference on diagnostic criteria for cerebrovascular diseases in China, IS cases were diagnosed using computed tomography (CT) or magnetic resonance imaging (MRI) and were confirmed by at least two independent clinical neurologists. Patients with IS caused by other conditions, such as transient ischemic attacks, brain trauma, tumors, and cerebrovascular malformation, were excluded from our study. Control subjects were enrolled from volunteers in examination centers and inpatients with other slight diseases, and healthy controls have no history of IS and other cerebrovascular diseases. All participants provided informed consent, and this study was approved by the Ethics Committee of the Guangxi University of Chinese Medicine.

### Quantitative real-time polymerase chain reaction (qRT-PCR)

Peripheral blood leukocytes were collected in the morning, and total RNA were extracted using TRIzol™ Reagent (Invitrogen™, USA) according to the manufacturer's protocol. cDNA reverse transcription was conducted using PrimeScript RT Reagent Kit with gDNA Eraser (Takara, Japan) and T100™ Thermal Cycler (BIO-RAD, USA). Primers were designed and synthesized by Shanghai Sangon Biological Engineering (Shanghai, China).

lncRNA H19: 5'-TTCAAGCCACGACTCT-3' (forward); 5'-GCTCACACTCACGCACACTC-3' (reverse).

Primer sequences of reference gene GAPDH: 5'-AGTCCTTCCACGATACCAAAGT-3' (forward); 5'-CATGAGAAGTATGACAACAGCCT-3' (reverse).

qRT-PCR experiment was conducted on the Applied Biosystems 7500 Real-Time PCR Systems (Applied Biosystems™, USA) with an SYBR Premix Ex Taq™ II kit (TaKaRa) according to the manufacturer's instruction. qRT-PCR was performed at the condition of 95 °C for 30 s, 95 °C for 5 s, and then 60 °C for 34 s, with a total of 40 cycles. The relative expression of lncRNA H19 was calculated using  $2^{-\Delta\Delta CT}$  method.

### Genotyping

Two milliliters of fasting peripheral blood sample were collected from the studied individuals, and DNA was extracted using the DNA kit from Aidlab Biotechnologies (Beijing, China). Genotyping was conducted using the Sequenom MassARRAY technology by Bio Miao Biological Technology (Beijing, China). Primer sequences are shown as follows:

rs217727: 5'-ACGTTGGATGAAAGACACCA TCGGAACAGC-3' (forward), 5'-ACGTTGGA TGAGCTCTGGGATGATGTGGTG-3' (reverse).

rs4929984: 5'-ACGTTGGATGTTTTGCAGAG GCGGTTTCAC-3' (forward), 5'-ACGTTGGATGATGC ATGGGCTCCTAGACAG-3' (reverse).

To confirm the quality of genotyping results, 5% of the DNA samples were selected to replicate detection and 100% consistency was obtained.

### Clinical biochemical markers measurement

Systolic blood pressure (SBP) and diastolic blood pressure (DBP) were measured with a calibrated mercury sphygmomanometer in the morning. The Mindray-6900 automatic blood corpuscle analyzer was used to measure platelet (PLT) level; activated partial thromboplastin time (APTT), D-dimer (D-D), fibrinogen (FIB), international normalized ratio (INR), prothrombin time (PT), prothrombin time activity (PTA), and thrombin time (TT) were detected using a StAgO Capact blood coagulation analyzer. The levels of C-reactive protein, homocysteine (Hcy), and uric acid (UA) were measured using the Hitachi 7600 automatic biochemistry analyzer. All measurements were completed by the First Affiliated Hospital of Guangxi University of Chinese Medicine according to strict laboratory procedures.

### Statistical analysis

Genetic association analysis was performed by PLINK software.  $\chi^2$  goodness-of-fit test was performed to determine Hardy–Weinberg Equilibrium (HWE). The distributions of genotype frequency between IS and control groups were compared using chi-square test. Non-conditional logistic regression analysis was used to evaluate the association of each loci with IS under different genetic models (additive, dominant, recessive, and allelic models). The generalized linear model

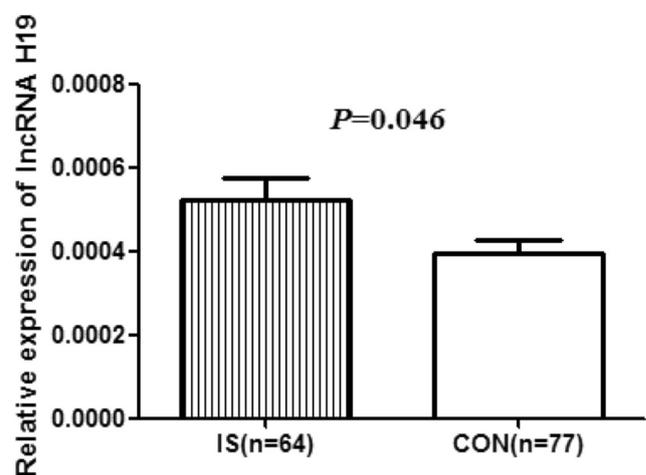


Fig. 1 Comparison of relative expression of lncRNA H19 in patients and healthy controls. IS: ischemic stroke, CON: healthy controls

**Table 1** Hardy-Weinberg equilibrium test for the healthy controls

| SNP_ID    | Allele (minor/major) | IS                            |                               |                               | control                       |                               |                               | $P_{\text{HWE}}$ | $\chi^2$ | $P$   |
|-----------|----------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|-------------------------------|------------------|----------|-------|
|           |                      | A <sub>1</sub> A <sub>1</sub> | A <sub>1</sub> A <sub>2</sub> | A <sub>2</sub> A <sub>2</sub> | A <sub>1</sub> A <sub>1</sub> | A <sub>1</sub> A <sub>2</sub> | A <sub>2</sub> A <sub>2</sub> |                  |          |       |
| rs217727  | A/G                  | 38                            | 221                           | 291                           | 47                            | 204                           | 298                           | 0.1609           | 1.715    | 0.424 |
| rs4929984 | A/C                  | 81                            | 243                           | 222                           | 80                            | 257                           | 208                           | 1.000            | 0.853    | 0.653 |

A1: minor allele-, A2: major allele-;  $P_{\text{HWE}}$ : HWE for control subjects

was used to determine the association between SNPs and clinical biochemical parameters. The relative expressions of lncRNA H19 in IS and control groups were compared through Student's *t* test, and the comparison of classified variables used chi-square test. These analyses were performed on the SPSS16.0 software. All significant results were set as two-tailed test and *P* value lower than 0.05.

### Meta-analysis

A meta-analysis was conducted to study the association of lncRNA H19 variants with IS susceptibility. Heterogeneity test was determined by the Q-test and  $I^2$  statistics, and if the  $I^2 > 50\%$  and  $P > 0.10$ , data were merged using the fixed-effect model (Mantel–Haenszel method). Otherwise, the random-effect model (The DeSimonian and Laird method) was used. The odds ratio (OR) with 95% confidence interval (CI) represents the effect strength. The above analysis was conducted using Stata 12.0. All tests were two tailed and a statistical significance was set as  $P < 0.05$ .

## Results

### Baseline characteristics of the study subjects

A total of 550 IS patients (306 males and 244 females) and 550 healthy controls (293 males and 257 females) were included in our study. The mean age was  $70.10 \pm 8.82$  in patients with IS and  $69.23 \pm 9.68$  in control subjects. Age ( $t = -1.560$ ,  $P = 0.119$ ) and gender ( $\chi^2 = 0.619$ ,  $P = 0.431$ ) in IS cases and controls did not differ significantly.

### lncRNA H19 expression in study subjects

We selected 64 IS patients and 77 healthy controls to determine lncRNA H19 expression. Figure 1 shows that compared with the control group, a significant upregulation of lncRNA H19 expression in peripheral blood in IS patients was obtained ( $t = 2.023$ ,  $P = 0.046$ ).

### Association analysis of lncRNA H19 variants with IS risk

A total of 550 IS patients and 550 healthy controls were genotyped, but 4 in IS group of rs4929984, 1 in control group of rs217727 and 5 in controls of rs4929984 were failed to genotype. Thus, 550 IS patients and 549 controls for rs217727 and 546 IS patients and 545 controls for rs4929984 were finally analyzed in the genetic association of lncRNA H19 polymorphisms with IS. The distributions of genotypes of rs217727 and rs4929984 in healthy controls were in Hardy–Weinberg Equilibrium ( $P > 0.05$ ). Table 1 shows that no significant difference was found in the distributions of genotype frequency of lncRNA H19 variants between IS and healthy controls (rs217727:  $\chi^2 = 1.715$ ,  $P = 0.424$ ; rs4929984:  $\chi^2 = 0.853$ ,  $P = 0.653$ ). What's more, rs217727 and rs4929984 showed no significant association with IS susceptibility under the additive, dominant, recessive, and allelic models (Table 2).

### Haplotype association analysis

The rs4929984 (A/C) and rs217727 (A/G) constituted C-A, A-G, and C-G haplotypes. No significant association between three haplotypes and IS susceptibility was observed ( $P > 0.05$ , Table 3).

### Meta-analysis

Four studies (including our study) were included in this meta-analysis. A total of 1269 IS cases and 1251 controls for rs217727, and 698 IS cases 695 controls for rs4929984 were analyzed (Supplemental Table 1). The genotypic distributions of rs217727 and rs4929984 in total controls were in accordance with HWE ( $P > 0.05$ ). After pooling all data, both the SNPs rs217727 and rs4929984 were not significantly associated with the risk of IS in the four genetic models (all  $P > 0.05$ , Table 4).

### Association analysis of lncRNA H19 variants and risk factors of patients with IS

As presented in Table 5, rs4929984 was significantly associated with the DBP level of IS patients in the additive ( $\beta$ :  $-2.02[-3.59\text{to}-0.45]$ ,  $P = 0.012$ ) and dominant models ( $\beta$ :  $-2.62[-4.86\text{to}-0.38]$ ,  $P = 0.022$ ). The significant association

**Table 2** Association of lncRNA H19 variants rs217727 and rs4929984 with IS risk

| SNP_ID          | Additive model   |       |                     | Dominant model   |       |                     | Recessive model  |       |                     | Allelic model    |       |  |
|-----------------|------------------|-------|---------------------|------------------|-------|---------------------|------------------|-------|---------------------|------------------|-------|--|
|                 | OR (95%CI)       | P     | Adjusted OR (95%CI) | OR (95%CI)       | P     | Adjusted OR (95%CI) | OR (95%CI)       | P     | Adjusted OR (95%CI) | OR (95%CI)       | P     |  |
| rs217727 (A/G)  | 0.99 (0.82–1.20) | 0.942 | 0.99 (0.82–1.20)    | 1.06 (0.83–1.34) | 0.649 | 1.06 (0.83–1.34)    | 0.79 (0.51–1.24) | 0.306 | 0.79 (0.50–1.23)    | 0.99 (0.82–1.20) | 0.941 |  |
| rs4929984 (A/C) | 0.95 (0.80–1.13) | 0.578 | 0.96 (0.81–1.14)    | 0.90 (0.71–1.15) | 0.399 | 0.91 (0.71–1.16)    | 1.01 (0.72–1.42) | 0.942 | 1.01 (0.73–1.42)    | 0.95 (0.80–1.13) | 0.573 |  |

was still observed after adjustment for age and gender under the additive ( $P_{\text{adj}} = 0.007$ ) and dominant models ( $P_{\text{adj}} = 0.013$ ). As summarized in Table 6, a significant association between rs217727 and INR level of IS patients was obtained (additive  $\beta$ : 1.39[0.25–2.53],  $P = 0.017$ ,  $P_{\text{adj}} = 0.019$ ; recessive  $\beta$ : 4.18[1.38–6.97],  $P = 0.004$ ,  $P_{\text{adj}} = 0.004$ ), and rs217727 was also significantly associated with PTA parameter (additive  $\beta$ :  $-3.09[-5.63\text{to}-0.54]$ ,  $P = 0.018$ ,  $P_{\text{adj}} = 0.026$ ; recessive  $\beta$ :  $-9.51[-15.68\text{to}-3.32]$ ,  $P = 0.003$ ,  $P_{\text{adj}} = 0.004$ ). Moreover, rs217727 was significantly associated with Hcy level in patients with IS under the recessive model ( $\beta$ : 6.18[0.07–12.18],  $P = 0.048$ ), and this association remained after the adjustment of age and gender ( $P_{\text{adj}} = 0.048$ , Table 7).

## Discussion

Our findings showed that lncRNA H19 level was upregulated in IS patients compared with healthy controls. The lncRNA H19 rs217727 and rs4929984 polymorphisms are not significantly associated with IS susceptibility in the southern Chinese Han population. Further, rs4929984 is significantly associated with the DBP level of IS cases, and rs217727 is significantly correlated with INR, PTA, and Hcy levels in IS patients (Table 7).

Upon the investigation on the effects of lncRNA H19 level on IS, we found an increased lncRNA H19 level in patients with IS. This result was in agreement with the findings of Wang (Wang et al. 2017a, b), whose results indicated an up-regulated lncRNA H19 level in stroke patients ( $N = 36$ ), brain tissues of mice with cerebral I/R injury, and human neuroblastoma cell (SH-SY5Y cell) after induction by OGD/R. Generally, lncRNA H19 inhibits autophagy by regulating DUSP5-ERK1/2 axis under the cellular OGD/R model (Wang et al. 2017a). Excessive autophagy in cerebral ischemia may induce apoptosis and mediate neuronal death (Puyal and Clarke 2009), and the inhibition of lncRNA H19 decreases cerebral infarct volume and brain edema and neurological deficits (Wang et al. 2017b). These data indicated that lncRNA H19 has a pivotal effect on the occurrence of IS.

As an imprinted gene, lncRNA H19 is expressed mainly in the period of the embryo and it executes the function of controlling embryo growth by targeting another imprinted gene, that is, the insulin-like growth factor 2 (Wilkin et al. 2000). lncRNA H19 is located in chromosome 11p15.5, and rs217727 and rs4929984 lying in *H19* are associated with the birth weight of newborns (Adkins et al. 2010; Hewage et al. 2015). Previous Chinese studies have reported that lncRNA H19 variants rs217727 and rs4929984 are significantly correlated with IS risk in the northern Chinese population (Wang et al. 2017a; Zhu et al. 2018). Gao et al. (2015) found that lncRNA H19 rs217727 is the risk loci of CAD. However, we failed to find any significant association

**Table 3** Haplotype association analysis of lncRNA H19 variants rs217727 and rs4929984 with IS

| Haplotype |   | Frequency-case | Frequency-control | OR   | <i>P</i> | Adjusted OR* | <i>P</i> <sub>adj</sub> |
|-----------|---|----------------|-------------------|------|----------|--------------|-------------------------|
| C         | A | 0.27           | 0.27              | 1.00 | 0.980    | 1.00         | 0.960                   |
| A         | G | 0.37           | 0.38              | 0.95 | 0.549    | 0.96         | 0.602                   |
| C         | G | 0.36           | 0.35              | 0.35 | 0.555    | 1.05         | 0.595                   |

*P*<sub>adj</sub>: adjusted by sex, age

between lncRNA H19 variants and IS susceptibility. On one hand, after combining previous studies, the results of meta-analysis show that the SNPs rs217727 and rs4929984 are not associated with IS risk, indicating that the small sample size is the major reason for our inconsistency. On the other hand, we speculate that a possible reason may be the genetic heterogeneity in different regions in China. Evidence proved that the Chinese population from north to south carries different allele frequencies of some human diseases (Yuan et al. 2013). In the study of previous studies (Wang et al. 2017a; Zhu et al. 2018), their participants were enrolled from the northern part of China, and may have reached to inconsistent findings as compared with ours. Thus, studies with larger sample size and multi-region are warranted to explain the correlation of lncRNA H19 variants with IS.

To investigate the association between lncRNA H19 variants and clinical biochemical parameters of IS patients, we collected clinical-related markers of patients with IS and performed an association analysis. As a result, we observed a significant correlation between rs4929984 and DBP level of patients with IS. SBP and DBP were demonstrated to be significantly associated with IS in the past years (Liu et al. 2013; Verdecchia et al. 2015). To the best of our knowledge, hypertension is defined as SBP  $\geq$  140 mmHg and DBP  $\geq$  90 mmHg (Kearney et al. 2005) and hypertension has been identified as a risk factor of IS. In recent years, genome-wide association studies (GWAS) have identified a number of genes and risk variants like *SH2B3* rs3184504 associated with BP through a combination study of a large sample in Europe ( $N = 133,661$ ), and these variants were confirmed to be strongly correlated

with SBP and DBP in individuals of East Asian ( $N = 29,719$ ), South Asian ( $N = 23,977$ ), and African ( $N = 19,775$ ) ancestry (International Consortium for Blood Pressure Genome-Wide Association et al. 2011). And genetic risk score assessing the association of the variants in aggregate BP was positively correlated with the incidence of stroke and CAD (International Consortium for Blood Pressure Genome-Wide Association et al. 2011). In addition, compared with the CC genotype, Feng et al. (2017) reported that *AMPD1* gene C34T polymorphism CT + TT genotype is significantly associated with decreased SBP of patients with cardiovascular diseases, which made one of the foundations for IS. rs4929984 is significantly associated with the blood pressure risk factor of IS, which suggests that lncRNA H19 variants involved in the development of IS may be related to the regulation of blood pressure.

Generally, abnormal blood coagulation plays a pivotal role in the pathogenesis of thrombus (Vazquez-Garza et al. 2017), which in turn leads to IS. Up to date, studies from GWAS and large sample subjects have identified plenty of risk loci and variants of thrombosis and IS (Chauhan and Debette 2016; Hinds et al. 2016). In investigating the correlation between gene variants and coagulation markers in stroke, Williams et al. (2013) found that *ABO* gene SNP rs505922 influencing blood coagulation is significantly associated with IS, with the T allele being a protective factor and the significance was confirmed in the subsequent subtype analysis of stroke. As a risk factor for atherosclerosis, ANRIL SNPs (rs10116277, rs7865618, rs564398, rs496892, and rs7044859) are significantly associated with IS and CAD (Cunnington et al. 2010)

**Table 4** Association between rs217727 and rs4929984 polymorphisms and blood pressure levels in patients with IS

| SNP_ID          | Number of cases/controls | Genetic model   | OR(95%CI)       | <i>P</i> | <i>I</i> <sup>2</sup> (%) | <i>P</i> <sub>Heterogeneity</sub> |
|-----------------|--------------------------|-----------------|-----------------|----------|---------------------------|-----------------------------------|
| rs217727 (A/G)  | 1269/1251                | Additive model  | 1.01(0.87–1.19) | 0.829    | 27.8                      | 0.251                             |
|                 |                          | Dominant model  | 1.40(0.87–2.26) | 0.162    | 85.4                      | 0.001                             |
|                 |                          | Recessive model | 1.41(0.80–2.48) | 0.239    | 81.5                      | 0.004                             |
|                 |                          | Allelic model   | 1.32(0.91–1.93) | 0.147    | 88.9                      | 0.000                             |
| rs4929984 (A/C) | 698/695                  | Additive model  | 1.04(0.89–1.21) | 0.631    | 0.0                       | 0.447                             |
|                 |                          | Dominant model  | 1.24(0.83–1.87) | 0.294    | 83.0                      | 0.003                             |
|                 |                          | Recessive model | 1.30(0.93–1.80) | 0.123    | 58.9                      | 0.088                             |
|                 |                          | Allelic model   | 1.23(0.90–1.69) | 0.194    | 86.1                      | 0.001                             |

**Table 5** Association between rs217727 and rs4929984 polymorphisms and blood coagulation markers in patients with IS

| SNP_ID          | Model     | SBP                |         |                    | DBP       |                      |         |                      |           |
|-----------------|-----------|--------------------|---------|--------------------|-----------|----------------------|---------|----------------------|-----------|
|                 |           | $\beta^c$ (95%CI)  | Crude P | $\beta^b$ (95%CI)  | $P_{adj}$ | $\beta^c$ (95%CI)    | Crude P | $\beta^b$ (95%CI)    | $P_{adj}$ |
| rs217727 (A/G)  | Additive  | 0.61 (-2.43–3.64)  | 0.695   | 0.63 (-2.41–3.66)  | 0.685     | 1.51 (-0.26–3.28)    | 0.094   | 1.52 (-0.19–3.24)    | 0.082     |
|                 | Dominant  | 1.04 (-2.74–4.81)  | 0.590   | 1.03 (-2.74–4.82)  | 0.591     | 1.85 (-0.35–4.05)    | 0.100   | 1.74 (-0.4–3.88)     | 0.111     |
|                 | Recessive | -0.39 (-7.82–7.05) | 0.919   | -0.23 (-7.68–7.21) | 0.951     | 1.92 (-2.42–6.26)    | 0.386   | 2.45 (-1.77–6.66)    | 0.255     |
| rs4929984 (A/C) | Additive  | -1.09 (-3.80–1.62) | 0.429   | -1.17 (-3.89–1.55) | 0.400     | -2.02 (-3.59to-0.45) | 0.012   | -2.02 (-3.66to-0.60) | 0.007     |
|                 | Dominant  | -1.19 (-5.04–2.67) | 0.547   | -1.31 (-5.19–2.57) | 0.507     | -2.62 (-4.86to-0.38) | 0.022   | -2.79 (-4.98to-0.60) | 0.013     |
|                 | Recessive | -1.97 (-7.29–3.36) | 0.470   | -2.01 (-7.34–3.33) | 0.461     | -2.80 (-5.90–0.31)   | 0.078   | -2.91 (-5.93–0.10)   | 0.059     |

$\beta^c$ : regression coefficient without adjusted;  $\beta^b$ : regression coefficient adjusted by age, sex;  $P_{adj}$ : adjusted by age, sex

although the underlying mechanism is unclear. LncRNA H19 has been reported aberrantly expressed in atherosclerosis plaque (Pan 2017), and we found a significant association between lncRNA H19 rs217727 and coagulation markers INR and PTA level, which suggests that coagulation dysfunction could be a reason for the involvement of lncRNA H19 polymorphism in the formation of thrombosis in IS occurrence. Functional studies need to verify our speculation.

In addition to BP and coagulation function, another risk factor, Hcy, also contributes to IS especially in hypertensive patients (Wang et al. 2014). Plasma Hcy level is commonly influenced by gene and variants. Zhao et al. (2017) investigated the association between the *MTHFR* C677T polymorphism and Hcy level in the incidence of IS and found that Hcy increases the risk of first stroke in subjects with the TT genotype, but this effect is significantly modified by *MTHFR* gene, indicating a significant gene–Hcy interaction in the occurrence of IS. Moreover, elevated Hcy-associated SNPs rs838133 and rs7422339 are correlated with IS (Cotlarciuc et al. 2014). A high Hcy level would tend the endothelial and vascular smooth muscle cells to injury, resulting in endothelial proliferation, activation of coagulation factors, and expression of plasminogen activator inhibitor, in turn, lead to PLT aggregation and subsequently IS (Hainsworth et al. 2016). Given that lncRNA H19 is involved in vascular endothelial cell function by regulating endothelial cell proliferation (Simion et al. 2018), lncRNA H19 rs217727 was significantly associated with the Hcy level of IS in this study, indicating that Hcy may be an intermediary between lncRNA H19 variations and the occurrence of IS. Taken together, lncRNA H19 might be involved in IS through the modification of blood pressure, coagulation function, and Hcy level. These findings need to be further verified.

**Limitations**

Several limitations need to be addressed. (i) This work is a single-center and retrospective study, and we only recruited Chinese Han population. Thus, selection bias may exist. Multi-center and prospective studies need to be conducted to demonstrate the association of lncRNA H19 with the occurrence of IS. (ii) Given that diverse IS subtypes may lead to different clinical endings, subtype analysis needs to be performed further. (iii) Although this study found an upregulation of lncRNA H19 level in 64 IS patients compared with 77 healthy controls and lncRNA H19 polymorphisms were associated with IS-related clinical parameters, our sample size was too small relative to the study of GWAS, and this size may not be sufficient to demonstrate the role of lncRNA H19 in IS. Hence, studies with large sample individuals are needed to verify our findings.

**Table 6** Association of rs217727 and rs4929984 polymorphisms with blood CRP, Hcy and UA levels in patients with IS

| Variables | SNP_ID    | Additive             |                |                      |                         | Dominant           |                |                     |                         | Recessive             |                |                       |                         |
|-----------|-----------|----------------------|----------------|----------------------|-------------------------|--------------------|----------------|---------------------|-------------------------|-----------------------|----------------|-----------------------|-------------------------|
|           |           | $\beta^a$ (95% CI)   | Crude <i>P</i> | $\beta^b$ (95% CI)   | <i>P</i> <sub>adj</sub> | $\beta^a$ (95% CI) | Crude <i>P</i> | $\beta^b$ (95% CI)  | <i>P</i> <sub>adj</sub> | $\beta^a$ (95% CI)    | Crude <i>P</i> | $\beta^b$ (95% CI)    | <i>P</i> <sub>adj</sub> |
| APTT      | rs217727  | 0.69 (-0.28–1.66)    | 0.164          | 0.67 (-0.30–1.64)    | 0.178                   | 0.61 (-0.60–1.83)  | 0.324          | 0.58 (-0.63–1.79)   | 0.349                   | 1.79 (-0.60–4.18)     | 0.142          | 1.79 (-0.60–4.18)     | 0.142                   |
|           | rs4929984 | -0.20 (-1.07–0.68)   | 0.659          | -0.14 (-1.01–0.73)   | 0.751                   | -0.62 (-1.86–0.62) | 0.328          | -0.50 (-1.74–0.75)  | 0.435                   | 0.42 (-1.29–2.12)     | 0.633          | 0.39 (-1.31–2.09)     | 0.655                   |
| D-D       | rs217727  | -0.28 (-1.09–0.53)   | 0.501          | -0.30 (-1.11–0.51)   | 0.468                   | -0.54 (-1.56–0.48) | 0.300          | -0.56 (-1.58–0.46)  | 0.282                   | 0.36 (-1.60–2.32)     | 0.717          | 0.31 (-1.65–2.27)     | 0.758                   |
|           | rs4929984 | -0.27 (-1.01–0.47)   | 0.475          | -0.24 (-0.98–0.50)   | 0.531                   | -0.46 (-1.51–0.58) | 0.385          | -0.40 (-1.46–0.65)  | 0.452                   | -0.15 (-1.60–1.30)    | 0.840          | -0.14 (-1.59–1.31)    | 0.849                   |
| FIB       | rs217727  | 0.05 (-0.10–0.20)    | 0.490          | 0.08 (-0.10–0.19)    | 0.534                   | 0.04 (-0.14–0.23)  | 0.649          | 0.04 (-0.15–0.22)   | 0.675                   | 0.15 (-0.21–0.51)     | 0.424          | 0.13 (-0.23–0.49)     | 0.484                   |
|           | rs4929984 | 0.09 (-0.04–0.22)    | 0.184          | 0.10 (-0.04–0.23)    | 0.159                   | 0.16 (-0.03–0.35)  | 0.104          | 0.17 (-0.02–0.36)   | 0.085                   | 0.05 (-0.21–0.31)     | 0.712          | 0.05 (-0.21–0.31)     | 0.691                   |
| INR       | rs217727  | 1.39 (0.25–2.53)     | 0.017          | 1.36 (0.22–2.51)     | 0.019                   | 1.09 (-0.34–2.51)  | 0.137          | 1.06 (-0.37–2.48)   | 0.148                   | 4.18 (1.38–6.97)      | 0.004          | 4.15 (1.35–6.94)      | 0.004                   |
|           | rs4929984 | -0.37 (-1.40–0.66)   | 0.482          | -0.32 (-1.35–0.71)   | 0.544                   | -0.43 (-1.90–1.03) | 0.561          | -0.32 (-1.79–1.15)  | 0.671                   | -0.60 (-2.62–1.42)    | 0.562          | -0.62 (-2.65–1.40)    | 0.545                   |
| PLT       | rs217727  | 5.70 (-4.46–15.86)   | 0.272          | 6.07 (-6.06–16.10)   | 0.237                   | 3.21 (-9.55–15.97) | 0.622          | 3.65 (-8.95–16.26)  | 0.570                   | 21.41 (-3.14–45.96)   | 0.088          | 21.96 (-2.30–46.22)   | 0.077                   |
|           | rs4929984 | 5.94 (-3.20–15.08)   | 0.203          | 5.05 (-3.99–14.10)   | 0.274                   | 4.50 (-8.50–17.50) | 0.498          | 2.24 (-10.66–15.14) | 0.734                   | 14.39 (-3.59–32.37)   | 0.117          | 15.28 (-2.49–33.05)   | 0.092                   |
| PT        | rs217727  | 0.24 (-0.66–1.14)    | 0.607          | 0.22 (-0.68–1.13)    | 0.627                   | 0.43 (-0.70–1.56)  | 0.457          | 0.41 (-0.72–1.54)   | 0.476                   | -0.22 (-242–1.98)     | 0.845          | -0.23 (-2.43–1.97)    | 0.838                   |
|           | rs4929984 | 0.13 (-0.69–0.94)    | 0.761          | 0.16 (-0.66–0.97)    | 0.708                   | 0.38 (-0.78–1.54)  | 0.520          | 0.45 (-0.71–1.61)   | 0.450                   | -0.23 (-1.82–1.37)    | 0.773          | -0.24 (-1.84–1.35)    | 0.764                   |
| PTA       | rs217727  | -3.09 (-5.63to-0.54) | 0.018          | -2.85 (-5.35to-0.35) | 0.026                   | -2.32 (-5.52–0.88) | 0.155          | -2.08 (-5.22–1.05)  | 0.194                   | -9.51 (-15.68to-3.32) | 0.003          | -9.02 (-15.08to-2.96) | 0.004                   |
|           | rs4929984 | 1.54 (-0.76–3.84)    | 0.189          | 1.17 (-1.08–3.44)    | 0.308                   | 3.22 (-0.04–6.49)  | 0.053          | 2.43 (-0.79–5.66)   | 0.139                   | -0.19 (-4.71–4.33)    | 0.935          | -0.07 (-4.50–4.35)    | 0.975                   |
| TT        | rs217727  | -0.42 (-1.84–1.01)   | 0.567          | -0.44 (-1.88–0.98)   | 0.539                   | -0.84 (-2.63–0.95) | 0.355          | -0.88 (-2.67–0.91)  | 0.334                   | 0.71 (-2.77–4.21)     | 0.687          | 0.68 (-2.81–4.17)     | 0.703                   |
|           | rs4929984 | -0.98 (-2.27–0.30)   | 0.135          | -0.93 (-2.21–0.37)   | 0.161                   | -1.56 (-3.39–0.27) | 0.096          | -1.43 (-3.28–0.41)  | 0.128                   | -0.82 (-3.34–1.70)    | 0.524          | -0.84 (-3.36–1.68)    | 0.512                   |

$\beta^a$ : regression coefficient without adjusted;  $\beta^b$ : regression coefficient adjusted by age, sex; *P*<sub>adj</sub>: adjusted by age, sex

**Table 7** Meta-analysis of rs217727 and rs4929984 polymorphisms and the susceptibility to ischemic stroke

| Variables | SNP_ID    | Additive               |           |           | Dominant                |           |           | Recessive                |           |           |
|-----------|-----------|------------------------|-----------|-----------|-------------------------|-----------|-----------|--------------------------|-----------|-----------|
|           |           | $\beta^a$ (95% CI)     | Crude $P$ | $P_{adj}$ | $\beta^a$ (95% CI)      | Crude $P$ | $P_{adj}$ | $\beta^a$ (95% CI)       | Crude $P$ | $P_{adj}$ |
| CRP       | rs217727  | -1.73<br>(-8.91–5.46)  | 0.639     | 0.629     | -1.34<br>(-10.44–7.76)  | 0.774     | 0.763     | -4.89<br>(-21.73–11.94)  | 0.570     | 0.566     |
|           | rs4929984 | -1.65<br>(-8.31–5.02)  | 0.629     | 0.698     | 1.06<br>(-8.36–10.47)   | 0.826     | 0.706     | -8.29<br>(-21.22–4.65)   | 0.212     | 0.198     |
| Hey       | rs217727  | 1.49<br>(-1.06–4.03)   | 0.253     | 0.251     | 0.66<br>(-2.57–3.90)    | 0.688     | 0.683     | 6.13<br>(0.07–12.18)     | 0.048     | 0.048     |
|           | rs4929984 | -1.00<br>(-3.36–1.35)  | 0.403     | 0.437     | -0.96<br>(-4.26–2.35)   | 0.571     | 0.650     | -2.07<br>(-6.76–2.62)    | 0.388     | 0.364     |
| UA        | rs217727  | -6.91<br>(-19.93–7.49) | 0.374     | 0.314     | -4.36<br>(-21.57–12.85) | 0.620     | 0.513     | -20.07<br>(-53.11–12.97) | 0.234     | 0.242     |
|           | rs4929984 | -4.51<br>(-16.96–7.94) | 0.478     | 0.620     | -7.76<br>(-25.33–9.81)  | 0.387     | 0.599     | -2.41<br>(-27.13–22.31)  | 0.849     | 0.806     |

$\beta^a$ : regression coefficient without adjusted;  $\beta^b$ : regression coefficient adjusted by age, sex;  $P_{adj}$ : adjusted by age, sex

## Conclusions

In summary, our findings suggested that the lncRNA H19 may have an effect on the occurrence of IS. LncRNA H19 rs217727 and rs4929984 polymorphisms, which in turn, may affect the level of blood pressure, coagulation function, and homocysteine metabolism of IS patients in the southern Chinese Han population. Functional and mechanism studies with a larger sample size are necessary to elucidate the underlying pathogenesis of lncRNA H19 in stroke.

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## Compliance with ethical standards

**Conflict of interest** There are no financial or other conflicts of interests.

**Ethical approval** All procedures performed in studies involving human participants were in accordance with the ethical standards of the Ethics Committee of the Guangxi University of Chinese Medicine and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

## References

Adkins RM, Somes G, Morrison JC, Hill JB, Watson EM, Magann EF, Krushkal J (2010) Association of birth weight with polymorphisms in the IGF2, H19, and IGF2R genes. *Pediatr Res* 68:429–434. <https://doi.org/10.1203/PDR.0b013e3181f1ca99>

Amouyel P (2012) From genes to stroke subtypes. *Lancet Neurol* 11: 931–933. [https://doi.org/10.1016/S1474-4422\(12\)70235-1](https://doi.org/10.1016/S1474-4422(12)70235-1)

Bao MH, Szeto V, Yang BB, Zhu SZ, Sun HS, Feng ZP (2018) Long non-coding RNAs in ischemic stroke. *Cell Death Dis* 9:281. <https://doi.org/10.1038/s41419-018-0282-x>

Chauhan G, Debette S (2016) Genetic risk factors for ischemic and hemorrhagic stroke. *Curr Cardiol Rep* 18:124. <https://doi.org/10.1007/s11886-016-0804-z>

Chen HH, Almontashiri NA, Antoine D, Stewart AF (2014) Functional genomics of the 9p21.3 locus for atherosclerosis: clarity or confusion? *Curr Cardiol Rep* 16:502. <https://doi.org/10.1007/s11886-014-0502-7>

Chen Z, Jiang B, Ru X, Sun H, Sun D, Liu X, Li Y, Li D, Guo X, Wang W (2017) Mortality of stroke and its subtypes in China: results from a Nationwide population-based survey. *Neuroepidemiology* 48:95–102. <https://doi.org/10.1159/000477494>

Cipolla GA, de Oliveira J, Salviano-Silva A, Lobo-Alves S, Lemos D, Oliveira L, Jucoski T, Mathias C, Pedrosa G, Zambalde E, Gradia D (2018) Long non-coding RNAs in multifactorial diseases: another layer of complexity Non-Coding RNA 4. <https://doi.org/10.3390/nrna4020013>

Cotlarciuc I, Malik R, Holliday EG, Ahmadi KR, Paré G, Psaty BM, Fomage M, Hasan N, Rinne PE, Ikram MA, Markus HS, Rosand J, Mitchell BD, Kittner SJ, Meschia JF, van Meurs J, Uitterlinden AG, Worrall BB, Dichgans M, Sharma P, METASTROKE and the International Stroke

- Genetics Consortium (2014) Effect of genetic variants associated with plasma homocysteine levels on stroke risk. *Stroke* 45:1920–1924. <https://doi.org/10.1161/STROKEAHA.114.005208>
- Cunnington MS, Santibanez Koref M, Mayosi BM, Burn J, Keavney B (2010) Chromosome 9p21 SNPs associated with multiple disease phenotypes correlate with ANRIL expression. *PLoS Genet* 6:e1000899. <https://doi.org/10.1371/journal.pgen.1000899>
- Feigin VL, Krishnamurthi RV, Parmar P, Norrving B, Mensah GA, Bennett DA, Barker-Collo S, Moran AE, Sacco RL, Truelsen T, Davis S, Pandian JD, Naghavi M, Forouzanfar MH, Nguyen G, Johnson CO, Vos T, Meretoja A, Murray CJ, Roth GA, GBD 2013 Writing Group, GBD 2013 Stroke Panel Experts Group (2015) Update on the global burden of ischemic and hemorrhagic stroke in 1990–2013: the GBD 2013 study. *Neuroepidemiology* 45:161–176. <https://doi.org/10.1159/000441085>
- Feng AF, Liu ZH, Zhou SL, Zhao SY, Zhu YX, Wang HX (2017) Effects of AMPD1 gene C34T polymorphism on cardiac index, blood pressure and prognosis in patients with cardiovascular diseases: a meta-analysis. *BMC Cardiovasc Disord* 17:174. <https://doi.org/10.1186/s12872-017-0608-0>
- Gabory A, Jammes H, Dandolo L (2010) The H19 locus: role of an imprinted non-coding RNA in growth and development. *BioEssays* 32:473–480. <https://doi.org/10.1002/bies.200900170>
- Gao W, Zhu M, Wang H, Zhao S, Zhao D, Yang Y, Wang ZM, Wang F, Yang ZJ, Lu X, Wang LS (2015) Association of polymorphisms in long non-coding RNA H19 with coronary artery disease risk in a Chinese population. *Mutat Res* 772:15–22. <https://doi.org/10.1016/j.mrfmmm.2014.12.009>
- Guo QY, Wang H, Wang Y (2017) LncRNA H19 polymorphisms associated with the risk of OSCC in Chinese population. *Eur Rev Med Pharmacol Sci* 21:3770–3774
- Hainsworth AH, Yeo NE, Weekman EM, Wilcock DM (2016) Homocysteine, hyperhomocysteinemia and vascular contributions to cognitive impairment and dementia (VCID). *Biochim Biophys Acta* 1862:1008–1017. <https://doi.org/10.1016/j.bbadis.2015.11.015>
- Han DK, Khaing ZZ, Pollock RA, Haudenschild CC, Liao G (1996) H19, a marker of developmental transition, is reexpressed in human atherosclerotic plaques and is regulated by the insulin family of growth factors in cultured rabbit smooth muscle cells. *J Clin Invest* 97:1276–1285. <https://doi.org/10.1172/JCI118543>
- Hewage AS, Jayanthiny P, Tennekoon KH, Kumarasiri JM, De SWAP, Karunanayake EH (2015) H19 rs217727 genotype and IGF-1/intron -2 dinucleotide CT repeat polymorphism are independently associated with birth weight. *Endocrine* 48:1010–1012. <https://doi.org/10.1007/s12020-014-0402-z>
- Hinds DA, Buil A, Ziemek D, Martinez-Perez A, Malik R, Folkersen L, Germain M, Mälarstig A, Brown A, Soria JM, Dichgans M, Bing N, Franco-Cereceda A, Souto JC, Dermizakis ET, Hamsten A, Worrall BB, Tung JY, METASTROKE Consortium, INVENT Consortium, Sabater-Lleal M (2016) Genome-wide association analysis of self-reported events in 6135 individuals and 252 827 controls identifies 8 loci associated with thrombosis. *Hum Mol Genet* 25:1867–1874. <https://doi.org/10.1093/hmg/ddw037>
- International Consortium for Blood Pressure Genome-Wide Association S et al (2011) Genetic variants in novel pathways influence blood pressure and cardiovascular disease risk. *Nature* 478:103–109. <https://doi.org/10.1038/nature10405>
- Iyer MK, Niknafs YS, Malik R, Singhal U, Sahu A, Hosono Y, Barrette TR, Prensner JR, Evans JR, Zhao S, Poliakov A, Cao X, Dhanasekaran SM, Wu YM, Robinson DR, Beer DG, Feng FY, Iyer HK, Chinnaiyan AM (2015) The landscape of long noncoding RNAs in the human transcriptome. *Nat Genet* 47:199–208. <https://doi.org/10.1038/ng.3192>
- Kearney PM, Whelton M, Reynolds K, Muntner P, Whelton PK, He J (2005) Global burden of hypertension: analysis of worldwide data. *Lancet* 365:217–223. [https://doi.org/10.1016/S0140-6736\(05\)17741-1](https://doi.org/10.1016/S0140-6736(05)17741-1)
- Kim DK, Zhang L, Dzau VJ, Pratt RE (1994) H19, a developmentally regulated gene, is reexpressed in rat vascular smooth muscle cells after injury. *J Clin Invest* 93:355–360. <https://doi.org/10.1172/JCI116967>
- Kim DS, Smith JA, Bielak LF, Wu CY, Sun YV, Sheedy PF, Turner ST, Peyser PA, Kardias SLR (2014) The relationship between diastolic blood pressure and coronary artery calcification is dependent on single nucleotide polymorphisms on chromosome 9p21.3. *BMC Med Genet* 15:89. <https://doi.org/10.1186/s12881-014-0089-2>
- Liu K et al (2013) [Association study between PDE4D gene polymorphism and ischemic stroke] Beijing da xue xue bao Yi xue ban = *Journal of Peking University Health Sciences* 45:359–363
- Luo M, Li Z, Wang W, Zeng Y, Liu Z, Qiu J (2013) Long non-coding RNA H19 increases bladder cancer metastasis by associating with EZH2 and inhibiting E-cadherin expression. *Cancer Lett* 333:213–221. <https://doi.org/10.1016/j.canlet.2013.01.033>
- Matouk IJ, Mezan S, Mizrahi A, Ohana P, Abu-lail R, Fellig Y, deGroot N, Galun E, Hochberg A (2010) The oncofetal H19 RNA connection: hypoxia, p53 and cancer. *Biochim Biophys Acta* 1803:443–451. <https://doi.org/10.1016/j.bbamcr.2010.01.010>
- Mehta SL, Kim T, Vemuganti R (2015) Long noncoding RNA FosDT promotes ischemic brain injury by interacting with REST-associated chromatin-modifying proteins. *J Neurosci* 35:16443–16449. <https://doi.org/10.1523/JNEUROSCI.2943-15.2015>
- Mozaffarian D et al (2016) Heart disease and stroke statistics-2016 update: a report from the American Heart Association. *Circulation* 133:e38–360. <https://doi.org/10.1161/CIR.0000000000000350>
- Ovbiagele B, Goldstein LB, Higashida RT, Howard VJ, Johnston SC, Khavjou OA, Lackland DT, Lichtman JH, Mohl S, Sacco RL, Saver JL, Trogon JG, on behalf of the American Heart Association Advocacy Coordinating Committee and Stroke Council (2013) Forecasting the future of stroke in the United States: a policy statement from the American Heart Association and American Stroke Association. *Stroke* 44:2361–2375. <https://doi.org/10.1161/STR.0b013e31829734f2>
- Pan JX (2017) LncRNA H19 promotes atherosclerosis by regulating MAPK and NF-κB signaling pathway. *Eur Rev Med Pharmacol Sci* 21:322–328
- Puyal J, Clarke PG (2009) Targeting autophagy to prevent neonatal stroke damage. *Autophagy* 5:1060–1061
- Rashid F, Shah A, Shan G (2016) Long non-coding RNAs in the cytoplasm. *Genomics Proteomics Bioinformatics* 14:73–80. <https://doi.org/10.1016/j.gpb.2016.03.005>
- Roy-O'Reilly M, McCullough LD (2014) Sex differences in stroke: the contribution of coagulation. *Exp Neurol* 259:16–27. <https://doi.org/10.1016/j.expneurol.2014.02.011>
- Simion V, Haemmig S, Feinberg MW (2018) LncRNAs in vascular biology and disease. *Vascul Pharmacol*. <https://doi.org/10.1016/j.vph.2018.01.003>
- Soudyab M, Iranpour M, Ghafouri-Fard S (2016) The role of long non-coding RNAs in breast cancer. *Arch Iran Med* 19:508–517
- Sun W, Yang Y, Xu C, Xie Y, Guo J (2016) Roles of long noncoding RNAs in gastric cancer and their clinical applications. *J Cancer Res Clin Oncol* 142:2231–2237. <https://doi.org/10.1007/s00432-016-2183-7>
- Sun W, Yang Y, Xu C, Guo J (2017) Regulatory mechanisms of long noncoding RNAs on gene expression in cancers. *Cancer Genet* 216:217:105–110. <https://doi.org/10.1016/j.cancergen.2017.06.003>
- Tragante V, Barnes MR, Ganesh SK, Lanktree MB, Guo W, Franceschini N, Smith EN, Johnson T, Holmes MV, Padmanabhan S, Karczewski KJ, Almqguera B, Barnard J, Baumert J, Chang YPC, Elbers CC, Farrall M, Fischer ME, Gaunt TR, Gho JM, Gieger C, Goel A, Gong Y, Isaacs A, Kleber ME, Leach IM, McDonough CW, Meijis MFL, Melander O, Nelson CP, Nolte IM, Pankratz N, Price TS,

- Shaffer J, Shah S, Tomaszewski M, van der Most PJ, van Iperen EPA, Vonk JM, Witkowska K, Wong COL, Zhang L, Beitelshes AL, Berenson GS, Bhatt DL, Brown M, Burt A, Cooper-DeHoff RM, Connell JM, Cruickshanks KJ, Curtis SP, Davey-Smith G, Delles C, Gansevoort RT, Guo X, Haiqing S, Hastie CE, Hofker MH, Hovingh GK, Kim DS, Kirkland SA, Klein BE, Klein R, Li YR, Maiwald S, Newton-Cheh C, O'Brien ET, Onland-Moret NC, Palmas W, Parsa A, Penninx BW, Pettinger M, Vasani RS, Ranchalis JE, M Ridker P, Rose LM, Sever P, Shimbo D, Steele L, Stolk RP, Thorand B, Trip MD, van Duijn CM, Verschuren WM, Wijmenga C, Wyatt S, Young JH, Zwinderman AH, Bezzina CR, Boerwinkle E, Casas JP, Caulfield MJ, Chakravarti A, Chasman DI, Davidson KW, Doevendans PA, Dominiczak AF, FitzGerald GA, Gums JG, Fornage M, Hakonarson H, Halder I, Hillege HL, Illig T, Jarvik GP, Johnson JA, Kastelein JJP, Koenig W, Kumari M, März W, Murray SS, O'Connell JR, Oldehinkel AJ, Pankow JS, Rader DJ, Redline S, Reilly MP, Schadt EE, Kottke-Marchant K, Snieder H, Snyder M, Stanton AV, Tobin MD, Uitterlinden AG, van der Harst P, van der Schouw YT, Samani NJ, Watkins H, Johnson AD, Reiner AP, Zhu X, de Bakker PIW, Levy D, Asselbergs FW, Munroe PB, Keating BJ (2014) Gene-centric meta-analysis in 87,736 individuals of European ancestry identifies multiple blood-pressure-related loci. *Am J Hum Genet* 94:349–360. <https://doi.org/10.1016/j.ajhg.2013.12.016>
- Vazquez-Garza E, Jerjes-Sanchez C, Navarrete A, Joya-Harrison J, Rodriguez D (2017) Venous thromboembolism: thrombosis, inflammation, and immunothrombosis for clinicians. *J Thromb Thrombolysis* 44:377–385. <https://doi.org/10.1007/s11239-017-1528-7>
- Verdecchia P, Reboldi G, Angeli F, Trimarco B, Mancina G, Pogue J, Gao P, Sleight P, Teo K, Yusuf S (2015) Systolic and diastolic blood pressure changes in relation with myocardial infarction and stroke in patients with coronary artery disease. *Hypertension* 65:108–114. <https://doi.org/10.1161/HYPERTENSIONAHA.114.04310>
- Wang CY, Chen ZW, Zhang T, Liu J, Chen SH, Liu SY, Han LY, Hui ZH, Chen YM (2014) Elevated plasma homocysteine level is associated with ischemic stroke in Chinese hypertensive patients. *Eur J Intern Med* 25:538–544. <https://doi.org/10.1016/j.ejim.2014.04.011>
- Wang J, Cao B, Han D, Sun M, Feng J (2017a) Long non-coding RNA H19 induces cerebral ischemia reperfusion injury via activation of autophagy. *Aging Dis* 8:71–84. <https://doi.org/10.14336/AD.2016.0530>
- Wang J, Zhao H, Fan Z, Li G, Ma Q, Tao Z, Wang R, Feng J, Luo Y (2017b) Long noncoding RNA H19 promotes neuroinflammation in ischemic stroke by driving histone deacetylase 1-dependent M1 microglial polarization. *Stroke* 48:2211–2221. <https://doi.org/10.1161/STROKEAHA.117.017387>
- Wilkin F, Paquette J, Ledru E, Hamelin C, Pollak M, Deal CL (2000) H19 sense and antisense transgenes modify insulin-like growth factor-II mRNA levels. *Eur J Biochem* 267:4020–4027
- Williams FM, Carter AM, Hysi PG, Surdulescu G, Hodgkiss D, Soranzo N, Traylor M, Bevan S, Dichgans M, Rothwell PM, Sudlow C, Farrall M, Silander K, Kaunisto M, Wagner P, Saarela O, Kuulasmaa K, Virtamo J, Salomaa V, Amouyel P, Arveiler D, Ferrières J, Wiklund PG, Ikram MA, Hofman A, Boncoraglio GB, Parati EA, Helgadottir A, Gretarsdottir S, Thorsteinsdottir U, Thorleifsson G, Stefansson K, Seshadri S, DeStefano A, Gschwendtner A, Psaty B, Longstreth W, Mitchell BD, Cheng YC, Clarke R, Ferrario M, Bis JC, Levi C, Attia J, Holliday EG, Scott RJ, Fornage M, Sharma P, Furie KL, Rosand J, Nalls M, Meschia J, Mosely TH, Evans A, Palotie A, Markus HS, Grant PJ, Spector TD, EuroCLOT Investigators, Wellcome Trust Case Control Consortium 2, MOnica Risk, Genetics, Archiving and Monograph, MetaStroke, International Stroke Genetics Consortium (2013) Ischemic stroke is associated with the ABO locus: the EuroCLOT study. *Ann Neurol* 73:16–31. <https://doi.org/10.1002/ana.23838>
- Yang C, Tang R, Ma X, Wang Y, Luo D, Xu Z, Zhu Y, Yang L (2015) Tag SNPs in long non-coding RNA H19 contribute to susceptibility to gastric cancer in the Chinese Han population. *Oncotarget* 6:15311–15320. <https://doi.org/10.18632/oncotarget.3840>
- Yang J, Gu L, Guo X, Huang J, Chen Z, Huang G, Kang Y, Zhang X, Long J, Su L (2018) LncRNA ANRIL expression and ANRIL gene polymorphisms contribute to the risk of ischemic stroke in the Chinese Han population. *Cell Mol Neurobiol* 38:1253–1269. <https://doi.org/10.1007/s10571-018-0593-6>
- Yuan J, Gao J, Li X, Liu F, Wijmenga C, Chen H, Gilissen LJ (2013) The tip of the “celiac iceberg” in China: a systematic review and meta-analysis. *PLoS One* 8:e81151. <https://doi.org/10.1371/journal.pone.0081151>
- Zhang W, Chen Y, Liu P, Chen J, Song L, Tang Y, Wang Y, Liu J, Hu FB, Hui R (2012) Variants on chromosome 9p21.3 correlated with ANRIL expression contribute to stroke risk and recurrence in a large prospective stroke population. *Stroke* 43:14–21. <https://doi.org/10.1161/STROKEAHA.111.625442>
- Zhang J, Yuan L, Zhang X, Hamblin MH, Zhu T, Meng F, Li Y, Chen YE, Yin KJ (2016) Altered long non-coding RNA transcriptomic profiles in brain microvascular endothelium after cerebral ischemia. *Exp Neurol* 277:162–170. <https://doi.org/10.1016/j.expneurol.2015.12.014>
- Zhang B, Wang D, Ji TF, Shi L, Yu JL (2017) Overexpression of lncRNA ANRIL up-regulates VEGF expression and promotes angiogenesis of diabetes mellitus combined with cerebral infarction by activating NF-kappaB signaling pathway in a rat model. *Oncotarget* 8:17347–17359. <https://doi.org/10.18632/oncotarget.14468>
- Zhao M, Wang X, He M, Qin X, Tang G, Huo Y, Li J, Fu J, Huang X, Cheng X, Wang B, Hou FF, Sun N, Cai Y (2017) Homocysteine and stroke risk: modifying effect of methylenetetrahydrofolate reductase C677T polymorphism and folic acid intervention. *Stroke* 48:1183–1190. <https://doi.org/10.1161/STROKEAHA.116.015324>
- Zhu R, Liu X, He Z (2018) Long non-coding RNA H19 and MALAT1 gene variants in patients with ischemic stroke in a northern Chinese Han population. *Mol Brain* 11:58. <https://doi.org/10.1186/s13041-018-0402-7>

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