



## Mental retardation in Down syndrome: Two ways to treat

Pierre P. Kamoun

Biochemistry and Molecular Biology, Paris-Descartes University, 26 rue de Chartres, 92200 Neuilly, France



### ABSTRACT

Mental retardation is a progressive condition in Down syndrome: intelligence starts to decline linearly within the first year. This phenomenon could be related to the overproduction of a toxic compound, hydrogen sulfide. Indeed, a gene located on chromosome 21 controls the production of cystathionine- $\beta$ -synthase, an enzyme involved in hydrogen sulfide production in the central nervous system. It has recently been demonstrated that excess cystathionine- $\beta$ -synthase levels are needed and sufficient to induce cognitive phenotypes in mouse models of Down syndrome. Thus, two therapeutic options might be used in Down syndrome patients: the use of a specific cystathionine  $\beta$ -synthase inhibitor and the use of an effective antidote to reduce hydrogen sulfide toxicity. Prenatal treatment of Down syndrome fetuses is also suggested.

### Introduction

Apart from profound hypotonia, the behavior of newborns with Down syndrome (DS) may appear reasonably normal. Developmental delay generally becomes obvious during the first few months of life. The achievement of developmental landmarks is usually increasingly delayed over time. Thus, the average delay may be of about two months for the very early landmarks (e.g. rolling over, transferring objects) but it lengthens gradually to reach one to two years for functions that normally appear around the age of two years [1]. Many studies assessing development during the first decade of life have shown that, even in DS children raised at home, there is a progressive, virtually linear decline in intelligence quotient (IQ) which starts within the first year of life [2]. By analogy with the toxic effect of phenylalanine in phenylketonuria, mental retardation in DS may also have a metabolic origin. It has been suggested that the metabolism of sulfur amino acids in the brain could lead to the production of a toxic compound, hydrogen sulfide [3].

Cystathionine  $\beta$ -synthase (CBS or EC 4.2.1.22) is an enzyme encoded by a gene located on chromosome 21 (21q23). CBS activity is increased by about 150% in fibroblasts from DS patients compared to normal subjects [4]. Also, CBS expression is 12 times higher in myeloblasts from DS children compared to normal subjects [5]. A polymorphism of the CBS allele is significantly under-represented in children with high IQ, suggesting that the level of CBS may influence cognitive functions [6]. In the brain of 34-week old DS patients, the level of CBS is about 3 times higher than in age-matched control subjects [7]. CBS is an enzyme involved in hydrogen sulfide production in the central nervous system (CNS) [8]. The role of hydrogen sulfide in different human models of cognitive defects has recently been described [9] as well as the role of polysulfides [10,11]. In rats, hydrogen sulfide

is mainly synthesized by cystathionine  $\gamma$ -lyase (cystathionase) in the liver, kidney, enterocytes and vascular smooth muscle cells and by 3-mercaptopyruvate sulfotransferase in heart tissues [12]. It has recently been found that the brain of CBS-knockout mice produces hydrogen sulfide, suggesting the presence of another hydrogen sulfide-producing enzyme. This enzyme has been identified as the 3-mercaptosulfurtransferase (in combination with cysteine aminotransferase). Both enzymes are located in mitochondria [13,14].

Thiosulfate is the main product of hydrogen sulfide metabolism [12]. Due to its stability, thiosulfate has been used as an index for hydrogen sulfide poisoning [15]. Thus, the endogenous production of hydrogen sulfide can be estimated by monitoring thiosulfate excretion in urine (about 31  $\mu$ moles/day) in control subjects [16]. Following hydrogen sulfide poisoning, the urinary thiosulfate level significantly increases [17]. In a study assessing urinary compounds in 17 DS patients compared to 17 normal subjects, it has been shown that urinary thiosulfate excretion was two times higher in DS patients. In contrast, no differences have been observed for the other urinary sulfur compounds (cystine, taurine and inorganic sulfate) [18,19]. The volume of the cerebellum is significantly smaller in DS patients than in matched controls, even after adjustment for the total brain volume or total intracranial volume [20]. Interestingly, the chronic exposure of pregnant dams to low concentrations of hydrogen sulfide leads to abnormal growth of developing cerebellar Purkinje cells of pups [21]. However, the cerebellum of CBS-knockout mice is also smaller than that of wild-type mice [22]. Mouse models of CBS deficiency have been shown to be good, although not perfect, models for human CBS deficiency [23]. There are many discrepancies between mouse models and the disease in humans: (a) homozygous null mice, unlike humans, show a high degree of neonatal lethality due to liver failure; (b) homozygous null mice show a normal or subnormal methionine plasma concentration while

E-mail address: [ppkamoun@wanadoo.fr](mailto:ppkamoun@wanadoo.fr).

<https://doi.org/10.1016/j.mehy.2019.109289>

Received 2 April 2019; Received in revised form 18 June 2019; Accepted 25 June 2019  
0306-9877/ © 2019 Elsevier Ltd. All rights reserved.

hypermethioninemia is found in humans; (c) a lack of cerebellum abnormalities is found in homocystinuria [24].

Clinical and biological findings have suggested a relationship between DS and chronic hydrogen sulfide poisoning. In a recently published study [25], the major role of CBS in the cognitive defects observed in DS has been suggested. In a mouse model of DS, i.e. Dp (17Abcg1-cbs)1Yah also referred to as Dp1Yah, mice are trisomic for CBS and 11 protein-encoding genes. The region also encompasses 6 non-coding genes. These mice show a deficit in the novel object recognition test (NORT). To decipher the role of CBS in DS cognitive phenotypes, the authors have generated and characterized constitutive and conditional changes in CBS levels in the CNS of various mouse models. Thus, they have demonstrated that three copies of CBS are necessary to induce cognitive impairment in Dp1Yah mice and that an excess CBS level is sufficient to induce cognitive phenotypes in mouse models of DS.

### Potential therapeutic options for mental retardation in Down syndrome

Two methods may be used to reduce hydrogen sulfide toxicity on the CNS: (a) inhibiting hydrogen sulfide production with specific CBS inhibitors, and (b) using hydrogen sulfide scavengers. In both cases, the chemical compounds must be able to pass the blood-brain barrier in order to target hydrogen sulfite levels in the brain.

#### Use of CBS inhibitors

CBS inhibitors have been widely investigated because CBS has recently been identified as a drug-target in several types of cancer [26]. Aminoxyacetic acid (AOAA) is the most widely used CBS inhibitor, but it should be noted that it also acts as a GABA-transaminase inhibitor. AOAA has been administered to infants and children who were resistant to usual anticonvulsant medications [27]. It has been used at a dose of 200 or 300 mg/day without significant adverse events but its efficacy was not uniform. Indeed, AOAA is a general inhibitor of aminotransferases and it has been shown to inhibit CBS and cystathionine  $\gamma$ -lyase (cystathionase) ( $IC_{50}$  at about 8 and 1  $\mu$ mole/L, respectively). AOAA improved learning and memory capacities in a chronic alcoholism rat model, and may be associated with reduced hippocampal hydrogen sulfide levels [28]. Thus, the use of AOAA as a CBS inhibitor seems safe and effective.

Benserazide is a decarboxylase inhibitor approved by the FDA for the adjuvant treatment of Parkinson's disease associated with a very low toxicity. It has been combined with L-DOPA (Madopar<sup>®</sup>) for its action on the DOPA decarboxylase. Benserazide is a less potent CBS inhibitor than the reference compound, AOAA ( $IC_{50}$  at about 30 and 1  $\mu$ M, respectively) [29]. An *in vivo* study has shown that the intraperitoneal injection of 300 and 600 mg/kg of benserazide inhibited cancer growth in tumor-bearing mice and no toxicity has been observed. Despite the low dose used in Parkinson's patients (about 50 mg/day), a slight increase in plasma homocysteine levels has been reported, confirming the inhibition of CBS [30]. However, benserazide cannot inhibit hydrogen sulfide formation in the brain because it is not able to pass the blood-brain barrier, although it is a very good inhibitor of CBS produced in extra-cerebral tissues (liver, kidney and others).

#### Use of hydrogen sulfide scavengers

Drugs counteracting the phenotypical consequences of CBS overexpression might also be used in DS. They have been developed to suppress the toxic effect of hydrogen sulfide in the brain. Sodium nitrite is the first compound developed to suppress hydrogen sulfide toxicity [19] and it has been used to treat acute hydrogen sulfide poisoning. Its action is mediated through the oxidation of hemoglobin to methemoglobin (metHb), a compound able to form a complex with hydrogen

sulfide resulting in sulfhemoglobin [31,32]. Another mechanism can be effective. Nitrite releases NO which reacts with hydrogen sulfide to produce polysulfides [33]. The administration of 4 mg/kg of sodium nitrite to human volunteers led to the formation of up to 7% of metHb. A significant difference has been observed in thiosulfate urinary excretion between DS patients and controls (21 diet-matched subjects):  $5.36 \pm 0.76$  versus  $2.23 \pm 0.42$  mmole/mole of creatinine ( $p < 0.0001$ ) [19]. Thus, the daily thiosulfate production has been estimated at 74.5  $\mu$ moles in DS patients compared to 31.0  $\mu$ moles in normal subjects. Furthermore, a recent study [34] has shown that the reaction between hydrogen sulfide and metHb leads to the formation of a metHb-SH complex in intact red blood cells. This study has shown that the metHb-SH complex was stable in the long term, and that its slow decomposition leads to the formation of reduced oxyHb, thiosulfate and/or polysulfides as final products. Interestingly, nitrite-induced methemoglobinemia remains one of the best antidotes available for the treatment of hydrogen sulfide poisoning [35]. Although the involvement of other mechanisms has been suggested in nitrite-induced hydrogen sulfide detoxification (including the effect of nitrite on mitochondrial enzymes), another mechanism might be the enhanced oxidative inactivation of hydrogen sulfide due to increased metHb levels in red blood cells. The safety use of oral sodium nitrite has been discussed [36] after its administration at a dose of 80 mg/day (1.1 mmole) in patients with diabetes mellitus and active or healed foot ulcers where only a few adverse events (headache, nausea) have been observed. Thus, sodium nitrite may be used to reduce chronic hydrogen sulfide intoxication.

Disulfiram, a potent inhibitor of mitochondrial aldehyde dehydrogenase, has been used in human CBS transgenic mice at a dose of 10 mg/kg/day before testing them for the NORT [25]. The NORT paradigm was restored in disulfiram-treated transgenic mice but not in non-treated mice. However, a better understanding of its mode of action and identifying disulfiram targets involved in the NORT are needed. The authors have suggested that this molecule may not directly inhibit CBS activity but probably rather act on the consequences of CBS overexpression [25]. Upon its absorption, disulfiram is rapidly reduced to diethyldithiocarbamate which then reacts with thiol groups. Disulfiram and its catabolite are potent copper chelators, so they might affect the activity of many copper-dependent enzymes. Disulfiram is a relatively non-toxic substance except for some cases of liver toxicity.

Cobinamide is the penultimate precursor of hydroxocobalamin (vitamin B12) produced by microorganisms. It has recently been shown that it readily reacts with hydrogen sulfide, by neutralizing two moles of sulfide [37]. The effects of three different types of ligand of cobinamide to reverse hydrogen sulfide toxicity have been investigated in a lethal rabbit model [38]. Rabbits received a continuous infusion of hydrogen sulfide donors (NaSH). Dinitrocobinamide was among the most effective compounds able to reverse hydrogen sulfide toxic effects [38]. Cobinamide or various types of ligand must be administered intravenously or intramuscularly in case of acute hydrogen sulfide toxicity but it has been suggested that it could also be used in prophylaxis [38]. The significant superiority of cobinamide over hydroxocobalamin in both *in vitro* and *in vivo* studies could be due to the fact that cobinamide has a higher affinity for hydrogen sulfide and/or to the ability of cobinamide to neutralize reactive oxygen species [39]. The administration of cobinamide 2 min after mice exposure to hydrogen sulfide has been shown to significantly and dose-dependently reduce lethality [40]. On the other hand, cobinamide-treated mice experienced significantly fewer seizures and knockdowns compared to the hydrogen sulfide-exposed group. Moreover, cobinamide has also been shown to reverse hydrogen sulfide-induced weight loss, behavioral deficits, neurochemical changes, cytochrome *c* oxidase inhibition and neurodegeneration in a dose- and time-dependent manner. Also, cobinamide increases survival and is neuroprotective in case of hydrogen sulfide-induced neurological sequelae. Thus, cobinamide could be a perfect drug candidate to be used in DS. Unfortunately, cobinamide is not yet

marketed but it should be the upcoming years [41].

## Conclusions

The best therapeutic option to treat mental retardation in DS might be to use a CBS inhibitor in combination with a hydrogen sulfide scavenger. Benserazide is of particular interest because it may decrease blood levels of hydrogen sulfide through its inhibitory effect on CBS overexpression in the liver, kidneys and in other peripheral tissues. DS patients treated with benserazide should present blood levels of hydrogen sulfide passing to the brain similar to those of normal subjects. To determine the dose needed to achieve it, the dose of benserazide could be progressively increased from 50 mg/day while monitoring thiosulfate blood levels [15]. It should be noted that the cost of this therapy is less than 16 USD for 200 mg/day of benserazide (Sigma-Aldrich-Merck). Sodium nitrite should be used at a dose allowing achieving a metHb concentration of 5%. MetHb and sulfmetHb blood levels may be used to determine the efficacy of treatment [42]. The cost is very low, less than 1 USD per week.

Ultimately, prenatal treatment of DS fetuses remains an issue. Hydroxocobalamin reacts with hydrogen sulfide according to a two-step detoxification: first, it forms a complex with hydrogen sulfide which reduces  $\text{Co}^{3+}$  in the hydroxocobalamin core to  $\text{Co}^{+}$ -Hydroxocobalamin, which then catalyzes the oxidation of hydrogen sulfide to sulfate. Hydroxocobalamin (vitamin B12) is the sole non-toxic and safe hydrogen sulfide scavenger currently available. A dose of 6 mg/day could be used [43] in pregnant women as a prenatal treatment of DS fetuses.

Further studies are needed to confirm these assumptions and validate the use of the drugs in the pre- and postnatal treatment of DS.

## Declaration of Competing Interest

The author, Pierre Kamoun, has no conflict of interest to declare.

## Acknowledgments

This work has been supported by Fondation Jerome Lejeune, Paris, France.

## References

- [1] Share JB, Veale AM. *Developmental Landmarks for children with Down syndrome*. Dunedin: University of Otago Press; 1974.
- [2] Morgan SB. Development and distribution of intellectual and adaptive skills in Down syndrome children: implications for early intervention. *Mental Retard* 1979;17:247–9.
- [3] Kamoun P. Mental retardation in Down syndrome: a hydrogen sulfide hypothesis. *Med Hypotheses* 2001;57:389–92.
- [4] Chadeaux B, Rethore MO, Raoul O, et al. Cystathionine beta synthase: gene dosage effect in trisomie 21. *Biochem Biophys Res Commun* 1985;128:40–4.
- [5] Taub JW, Huang X, Matherly LH, et al. Expression of chromosome 21-localized genes in acute myeloid leukemia: differences between Down and non-Down syndrome blast cells and relationship to in vitro sensitivity to cytosine arabinoside and daunorubicin. *Blood* 1999;94:1393–400.
- [6] Barbaux S, Plomin R, Whitehead AS. Polymorphism of genes controlling homocysteine/folate metabolism and cognitive function. *NeuroReport* 2000;11:1133–6.
- [7] Ichinohe A, Kanaumi T, Takashima S, Enokido Y, Nagai Y, Kimura H. Cystathionine beta-synthase is enriched in the brains of Down's patients. *Biochem Biophys Res Commun* 2005;338:1547–50.
- [8] Kimura H. Hydrogen sulfide as a neuromodulator. *Mol Neurobiol* 2002;26:13–9.
- [9] He JT, Li H, Yang L, Mao CY. Role of hydrogen sulfide in cognitive deficits: evidences and mechanisms. *Eur J Pharmacol* 2019. (in press).
- [10] Kimura H. Physiological roles of hydrogen sulfide and polysulfides. *Handb Exp Pharmacol* 2015;230:61–81.
- [11] Kimura H. Signaling by hydrogen sulfide and polysulfides via protein S-sulfuration. *Br J Pharmacol*. 2019. (in press).
- [12] Kamoun P. Endogenous production of hydrogen sulfide in mammals. *Amino Acids* 2004;26:243–54.
- [13] Shibuya N, Tanaka M, Yoshida M, et al. 3-mercaptopyruvate sulfoxyltransferase produces hydrogen sulfide and bound sulfane sulfur in the brain. *Antioxid Redox Signal* 2009;11:709–14.
- [14] Kimura Y, Toyofuku Y, Koike, et al. Identification of  $\text{H}_2\text{S}_3$  and  $\text{H}_2\text{S}$  produced by 3-mercaptopyruvate sulfoxyltransferase in the brain. *Sci Rep* 2015;5:4774.
- [15] Maseda C, Hayakawa A, Okuda K, et al. Liquid chromatography-tandem mass spectrometry method for the determination of thiosulfate in human blood and urine as an indicator of hydrogen sulfide poisoning. *Leg Med* 2017;24:67–74.
- [16] Sorbo B, Ohman S. Determination of thiosulphate in urine. *Scand J Clin Lab Invest* 1978;38:521–7.
- [17] Kangas J, Savolainen H. Urinary thiosulphate as an indicator of exposure to hydrogen sulphide vapor. *Clin Chim Acta* 1987;164:7–10.
- [18] Bellardinelli MC, Chabli A, Chadeaux-Vekemans B, Kamoun P. Urinary sulfur compounds in Down syndrome. *Clin Chem* 2001;47:1500–1.
- [19] Kamoun P, Bellardinelli MC, Chabli A, Lallouchi K, Chadeaux-Vekemans B. Endogenous hydrogen sulfide overproduction in Down syndrome. *Am J Med Genet A* 2003;116A:310–1.
- [20] Aylward EH, Habbak R, Warren AC, et al. Cerebellar volume in adults with Down syndrome. *Arch Neurol* 1997;54:209–12.
- [21] Hannah RS, Roth SH. Chronic exposure to low concentrations of hydrogen sulfide produces abnormal growth in developing cerebellar Purkinje cells. *Neurosci Lett* 1991;122:225–8.
- [22] Enokido Y, Suzuki E, Iwasawa K, Namekata K, Okasawa H, Kimura H. Cystathionine beta-synthase, a key enzyme for homocysteine metabolism, is preferentially expressed in the radial glia/astrocyte lineage of developing mouse CNS. *FASEB J* 2005;19:1854–6.
- [23] Kruger WD. Cystathionine beta synthase deficiency: og mice and man. *Mol Genet Metab* 2017;121:199–205.
- [24] Mudd SH, Levy HL, Skovby F. Disorders of transsulfuration. In: Scriver CR, Beaudet AL, Sly WS, Valle D (editors). *The metabolic and molecular bases of inherited disease*. Vol. 1 (7th ed.) New York: McGraw-Hill (pub) 1995. p. 1279–1327.
- [25] Marechal D, Brault V, Leon A, et al. Cbs overdosage is necessary and sufficient to induce cognitive phenotypes in mouse models of Down syndrome and interacts genetically with Dyrk1 a. *Hum Mol Genet* 2019. (in press).
- [26] Zhu H, Blake S, Chan KT, Pearson RB, Kang J. Cystathionine beta synthase in Physiology and Cancer. *Biomed Res Int* 2018. (in press).
- [27] Tibbles JA, McGreal DA. Trial of amino-oxyacetic, an anticonvulsant. *Can Med Assoc J* 1963;88:881–6.
- [28] Du AL, Qin HZ, Jiang HB, Py Fu, Lou K, Xu YM. Aminoxyacetic acid improves learning and memory in a rat model of chronic alcoholism. *Neural Regen Res* 2018;13:1568–74.
- [29] Druzhyna N, Szczesny B, Olah G, et al. Screening of a composite library of clinically used drugs and well-characterized pharmacological compounds for cystathionine beta synthase inhibition identifies benserazide as a drug potentially suitable for repurposing for the experimental therapy of colon cancer. *Pharmacol Res* 2016;113(Pt A):18–37.
- [30] Guo G, Xu S, Cao LD, Wu QY. The effect of levodopa benserazide hydrochloride levels in patients with Parkinson's disease and treatment of hyperhomocysteinemia. *Eur Rev Med Pharmacol Sci* 2016;20:2409–12.
- [31] Kiese M, Weger N. Formation of ferrihaemoglobin with aminophenols in the human for the treatment of cyanide poisoning. 1969;7:97–105.
- [32] Nichol AW, Hendry I, Morell DB. Mechanism of formation of sulphhaemoglobin. *Biochim Biophys Acta* 1968;156:97–108.
- [33] Miyamoto R, Koike S, Takano Y, et al. Polysulfides ( $\text{H}_2\text{S}_n$ ) produced from the interaction of hydrogen sulfide ( $\text{H}_2\text{S}$ ) and nitric oxide (NO) activate TRPA1 channels. *Sci Rep* 2017;7:45995.
- [34] Bianco CL, Savitsky A, Feelisch M, Cortese-Krott MM. Investigations on the role of hemoglobin in sulfide metabolism by intact red blood cells. *Biochem Pharmacol* 2018;149:163–73.
- [35] Butler AR, Feelisch M. Therapeutic uses of inorganic nitrite and nitrate: from the past to the future. *Circulation* 2008;117:2151–9.
- [36] Greenway FL, Predmore BL, Flanagan DR, et al. Single-dose pharmacokinetics of different oral sodium nitrite formulations in diabetes patients. *Diabetes Technol Ther* 2012;14. 552–530.
- [37] Salnikov DS, Makarov, van Eldik R, Kucherenko PN, Boss GR. Kinetics and mechanism of the reaction of hydrogen sulfide with diaquacobinamide in aqueous solution. *Eur J Inorg Chem* 2014 2014:4123–33.
- [38] Brenner M, Benavides S, Mahon SB, et al. The vitamin B12 analog cobinamide is an effective hydrogen sulfide antidote in a lethal rabbit model. *Clin Toxicol* 2014;52:490–7.
- [39] Jiang J, Chan A, Ali S, et al. Hydrogen sulfide-Mechanism of toxicity and development of an antidote. *Sci Rep* 2016;6:20831.
- [40] Anantharam P, Whitley EM, Mahama B, et al. Cobinamide is effective for treatment of hydrogen sulfide-induced neurological sequelae in a mouse model. *Ann N Y Acad Sci* 2017;1408:61–78.
- [41] Boss GR. (personal communication) February 2019.
- [42] Van Leeuwen SR, Baranovski GVG, Kimmel BW. Three-wavelength method for the optical differentiation of methemoglobin and sulfhemoglobin in oxygenated blood. *Conf Proc IEEE Eng Med Biol Soc* 2017;2017:4570–3.
- [43] Truong DH, Mihajlovic, Guinness P, Hindmarsh W, O'Brien PJ. Prevention of hydrogen sulfide ( $\text{H}_2\text{S}$ )-induced mouse lethality and cytotoxicity by hydroxocobalamin (vitamin B12a). *Toxicology* 2007;242:16–22.