

Learning from experience: cellular and molecular bases for improved outcome in subsequent pregnancies



Debra Goldman-Wohl, PhD; Moriya Gamliel, PhD; Ofer Mandelboim, PhD; Simcha Yagel, MD

Consider the following scenario: gravida 2 para 1 vs gravida 1 para 0: Two patients of similar age and socio-demographic status visit a prenatal clinic. Both are apparently healthy non-smokers who do not report any significant history. Patients are 6 weeks by dates with confirmatory ultrasound demonstrating fetal viability. The first patient is gravida 2 para 1, following an uncomplicated first pregnancy that resulted in the normal vaginal delivery of a healthy term infant, now 2.5 years old. The next patient is gravida 1 para 0. As these 2 patients carry different risks for pregnancy complications, consider the different degrees of caution and circumspection that will come into play as you follow these 2 gravidae through the coming months.

There is general agreement that pregnancy outcomes are more favorable for parous as compared with nulliparous women.¹ Differences between first vs subsequent pregnancies have been widely investigated through multiple

The frequencies of preeclampsia, fetal growth restriction, fetal demise, and low birth-weight are lower in subsequent pregnancies. Enhanced maternal cardiovascular adaptation, shorter first and second stages of labor, and more robust lactation also have been observed in subsequent as compared with first pregnancies. We sought to investigate the cellular and molecular bases for better outcomes in subsequent pregnancies. Based on the knowledge that specialized immune cells at the maternal–fetal interface, decidual natural killer cells, promote development of the placental bed and conversion of the spiral arteries by secreting a myriad of angiogenic and growth factors, we asked whether decidual natural killer cells differ in subsequent as compared with first pregnancies. This idea stemmed from recent studies suggesting that natural killer cells, although part of the innate immune system, possess some features of adaptive immunity, including a certain type of immune cell memory, termed trained immunity. We found that decidual natural killer cells from parous women “remember pregnancy” and differ from decidual natural killer cells of primigravidae. Compared with the decidual natural killer cells of first pregnancy, these cells, that we termed pregnancy-trained decidual natural killer cells, express greater levels of the natural killer receptors NKG2C and leukocyte immunoglobulin-like receptor B1, which interact with ligands expressed on invasive trophoblasts. Furthermore, they secrete greater levels of several growth factors, including vascular endothelial growth factor α as well as interferon- γ , augmenting remodeling of the placental bed. We propose that this pregnancy-trained memory dwells in the epigenome, where memory of stimuli is known to persist even when the stimulus is no longer present. This epigenetic memory apparently resides in endometrial natural killer cells between pregnancies. We suggest that this trained memory, which we coined pregnancy-trained decidual natural killer cells, may be the missing link in the immune basis for enhanced subsequent pregnancy. Epigenetic memory (chromatin modification) also may afford a global explanation for additional findings of enhanced maternal cardiovascular adaptation, shorter first and second stages of labor, and more robust lactation. Understanding the molecular and cellular bases of improved outcomes of subsequent pregnancy may lead to the development of treatment modalities designed for women at high risk for pregnancy disorders originating at the maternal–fetal interface.

Key words: decidual NK cells, DNA methylation, epigenetics, fetal growth restriction, FGR, IFN- γ , multigravida, NK cells, preeclampsia, primigravida, trained memory, VEGF

From the Magda and Richard Hoffman Center for Human Placenta Research, Department of Obstetrics and Gynecology (Drs Goldman-Wohl and Yagel), and The Concern Foundation Laboratories at the Lautenberg Centre for Immunology and Cancer Research, IMRIC, Faculty of Medicine (Drs Gamliel and Mandelboim), Hebrew University Hadassah Medical Center, Jerusalem, Israel.

Received Nov. 22, 2018; revised Feb. 7, 2019; accepted Feb. 18, 2019.

The authors report no conflict of interest.

O.M. is a Crown Professor of Molecular Immunology. M.G. is supported by the Hoffman Leadership and Responsibility Fund at The Hebrew University.

Corresponding author: Simcha Yagel, MD. simcha.yagel@gmail.com

0002-9378/\$36.00

© 2019 Elsevier Inc. All rights reserved.

<https://doi.org/10.1016/j.ajog.2019.02.037>

approaches: from population-based epidemiologic studies to histologic analysis of the maternal–fetal interface, to molecular biology–based examination of epigenetic memory. First pregnancies are associated with a 2.5-fold greater stillbirth risk,^{2,3} and placental and newborn weights are greater in parous as compared with primiparous women.^{4,5–7} Indeed, first deliveries are associated with lower birthweight and

greater rates of small for gestational age.^{8–10} Further examples of these observations would be the reduction in preeclampsia risk in subsequent pregnancies as a result of upgraded implantation/placentation,¹¹ enhanced maternal cardiovascular adaptation^{12–14} observed in increased end-diastolic volume, stroke volume, and cardiac output, and decreased vascular resistance;¹⁵ decreased mean arterial pressure and

reduced arterial stiffness;¹⁴ as well as better outcomes of first and second stages of labor and delivery;¹⁵ and more successful lactation, characterized by an improved nursing experience and increased milk supply¹⁶ in repeat pregnancy.¹⁷ Here we review the evidence of better outcomes of subsequent pregnancies by showing that the natural killer (NK) cells of the decidua remember pregnancy, enhancing development of the placental bed with epigenetic memory residing in the NK cells of the endometrium, between pregnancies.¹⁸

Many of the “great obstetrical syndromes” have their biological basis in defects of deep placentation and development of the placental bed,¹¹ and some are more prevalent in nulliparous women. There is evidence for augmented deep placentation in subsequent pregnancies. It has been shown that the proportion of decidual vessels undergoing trophoblast invasion is greater in first-trimester decidua isolated from parous as compared with nulliparous women.¹⁹ Khong et al showed that an “experienced” parous uterus differs from the nulliparous, lending histologic support to the notion that the uterus does not return to its nulliparous state following delivery,²⁰ while suggesting that the uteri of parous women are “primed” for future pregnancies. They demonstrated that the elastic tissue and musculature surrounding the uterine arteries in the nulliparous uterus can be distinguished from that of the altered uterine architecture found in the parous uterus²⁰ and that the parous uterus does not revert to the nulliparous tissue architecture. Enhanced placentation is further exhibited by differing levels of circulating angiogenic factors (greater angiopoietin-2 at 8 weeks) and more rapid placental development, as measured by ultrasound scan, in the early first trimester of parous as compared with nulliparous women.⁴

An unexpected finding, that parity negatively affects the success rate of first-trimester medical abortion, is described in several studies.^{21–24} Investigators examined different end points to determine successful medical abortion, such as β -human chorionic gonadotropin

levels, ultrasound-based imaging, and the need for surgical intervention, and no specific mechanism has been offered to elucidate this observation. However, Bartley et al comment that this finding is suggestive of a difference in the placental bed in parous vs nulliparous women, ie, the placental bed of parous women may be sounder early in pregnancy, resulting in less efficacious medical abortion.²¹

One of the most striking examples of contrasting risks in first and subsequent pregnancies is the increased risk for the development of preeclampsia.²⁵ Primiparity differs from the myriad identified risk factors for preeclampsia, which implicate fetal, maternal²⁶ and environmental factors²⁷ in the development of the syndrome. The rate of preeclampsia in primiparae is more than double that observed in parous women.²⁸ Yet while some of these risk factors seem to have obvious connections with the disorder (such as metabolic syndrome, for example) first pregnancy seems to defy most logical explanations for its association with preeclampsia, unless we look for some underlying immune component. First pregnancy as a risk factor for preeclampsia was recognized as early as the 17th century by the Frenchman Francois Mauriceau, who was the first to systematically describe eclampsia and to note that primigravidae were at greater risk for convulsions as compared with multigravidae^{29,30} (reported by Mcmillen 2003).³¹ Preeclampsia is thought to be initiated early in pregnancy with ineffective spiral artery conversion mediated by shallow trophoblast invasion, leading to inefficient utero placental blood flow.^{32–34}

To further categorize preeclampsia, attempts have been made, with some success, to connect specific histopathology of the placenta and gene expression profiles.³⁵ Burton et al have extensively studied and modeled blood flow to the intervillous space, including when the spiral arteries are narrow and have not undergone appropriate conversion. They reason that oxygenation and reoxygenation malperfusion resulting in oxidative stress and damage to syncytiotrophoblast would be a leading factor

in the development of preeclampsia.^{36–38} This defective placentation, coupled with an imbalance between antiangiogenic and angiogenic factors expression^{39–41} and a maternal inflammatory stress response,^{42,43} subsequently lead to endothelial cell dysfunction^{44,45} and the maternal cascade of the syndrome.⁴⁶

These observations of remembered development of the fetal–maternal interface in first pregnancy, persevering to enhance subsequent pregnancies, led us to seek a mechanism on a molecular and cellular level retaining the memory of pregnancy, thereby leading to more efficient or more rapid development of the placental bed and hence better pregnancy outcomes in parous women. We predict that this example of enhanced subsequent pregnancy, for which we demonstrate a molecular epigenetic model of NK cells at the maternal–fetal interface, will provide the basis for investigation into other, similar mechanisms leading to improved pregnancy, labor, and delivery outcomes and more robust lactation, in other cell types and tissues, as this field develops.

In this review, we will focus on the possible cellular and molecular bases for sounder subsequent pregnancies as compared with first pregnancies. We will discuss the evidence for enhanced adaptation to pregnancy through cellular epigenetic memory in decidual natural cell (dNK)-trained memory.¹⁸

Epigenetic Memory

When we discuss memory and the experience of pregnancy and childbirth, we can paraphrase the Japanese writer Haruki Murakami, who said that no matter how much suffering you went through, you never want to let go of those memories.

Epigenetics is defined as modification of chromatin but not the DNA code itself (reviewed in Dor et al⁴⁷). This epigenetic modification is in part facilitated through methylation of cytosine nucleotides usually found within a framework of CpG (cytosine phosphate guanine) in the chromosomal DNA. The process is controlled in part by enzymes, DNA methyltransferases, responsible for

adding a methyl group to cytosine residues in cytosine/guanine-rich regions of DNA (called “CpG islands”). The methylated DNA is generally more tightly wrapped around nucleosomes that pack the DNA with histone proteins and restrict accessibility of the transcriptional machinery inhibiting gene expression, whereas unmethylated DNA is more accessible to the cellular proteins responsible for activation of gene expression,⁴⁷ commonly referred to as “open” DNA.

Epigenetic modification of chromosomal DNA affords an explanation of how organs develop and maintain their identity, albeit with the same DNA sequence in each cell of the body.⁴⁸ Based on the understanding that a specific pattern of methyl groups added to chromosomal DNA may affect gene activity, hypomethylated regions of chromosomal DNA are thought to be more accessible to transcriptional machinery. This stable, mitotically inheritable pattern of “open” and “closed” DNA able to activate or silence transcription, has implications to cancer biology, developmental biology, and aging and could explain how an experienced cell lineage may be more readily activated when encountering a repeat of previous stimuli. This system affords a molecular mechanism for long-term memory of transcriptional events when the stimulus is no longer present.⁴⁷ We will discuss how this epigenetic mechanism functions as memory of previous pregnancy at the maternal–fetal interface.

dNK Cells Benefit the Establishment of a Healthy Placental Bed

dNK cells are in direct contact with trophoblast cells that have anchored the placenta and invaded the uterus and spiral arteries. This is a unique challenge to the maternal immune system: although it is essential to protect the fetus from infectious agents, there must be a mechanism that allows the semi-allogenic trophoblast cells to invade the maternal tissues without being eliminated by the maternal immune cells. NK cells are part of the innate immune system, accounting for 5%–15% of total lymphocytes in the peripheral blood

(pbNK). Since their first characterization more than 30 years ago, these innate immune lymphocytes have been found to serve as a first line of defense against a variety of infections.⁴⁹ The NK cells that are recruited to or proliferate in the decidua, the dNK, function not in the typical NK cytotoxic fashion but rather secrete factors that facilitate the remodeling of the placental bed.⁵⁰ This field of investigation will likely rapidly advance with the application of the technology of single-cell RNA sequencing to analyze RNA expression at the maternal–fetal interface of individual cell types.^{51–53}

One of the possible differences between the parous and nulliparous uterine environments could rest in the dNK cells found in the uterus during pregnancy. These cells enrich the maternal–fetal environmental interface as they produce chemokines, growth factors and angiogenic factors, including isoforms of vascular endothelial growth factor (VEGF), placental growth factor, and NKG5, an alternative splicing product of the granulysin gene that stimulates mitogenicity of endothelial cells that promote spiral artery remodeling and development of the placental bed.⁵⁰ Furthermore, specific chemokines produced by dNKs, notably interleukin-8 and interferon-inducible protein-10, can attract trophoblasts expressing the appropriate chemokine receptors to the maternal–fetal interface.^{50,54} Concomitantly, extravillous trophoblasts expressing the chemokine CXCL12 can attract dNKs expressing the appropriate chemokine receptor, CXCR4, to the maternal fetal interface.⁵⁵ Phenotypically, dNK cells express a different repertoire of cell-surface receptors from the vast majority of NK cells in the peripheral blood (ie, pbNK).⁵⁶

The pioneering work of Croy and colleagues, employing alymphoid mice, revealed a role for uterine NK cells in remodeling the uterine vascular environment (Ashkar et al,⁵⁷ reviewed by Ratsep et al⁵⁸), with interferon- γ (IFN- γ) enlarging the diameter of the lumen of the uterine arteries.

Highlighting the importance of NK cells at the fetal–maternal interface, recent investigations using rat models of

NK cell deficiency and of hypertension of pregnancy address a dual role for NK cells in both promoting and inhibiting excessive trophoblast invasion, and in the development of vasculopathy.^{59,60}

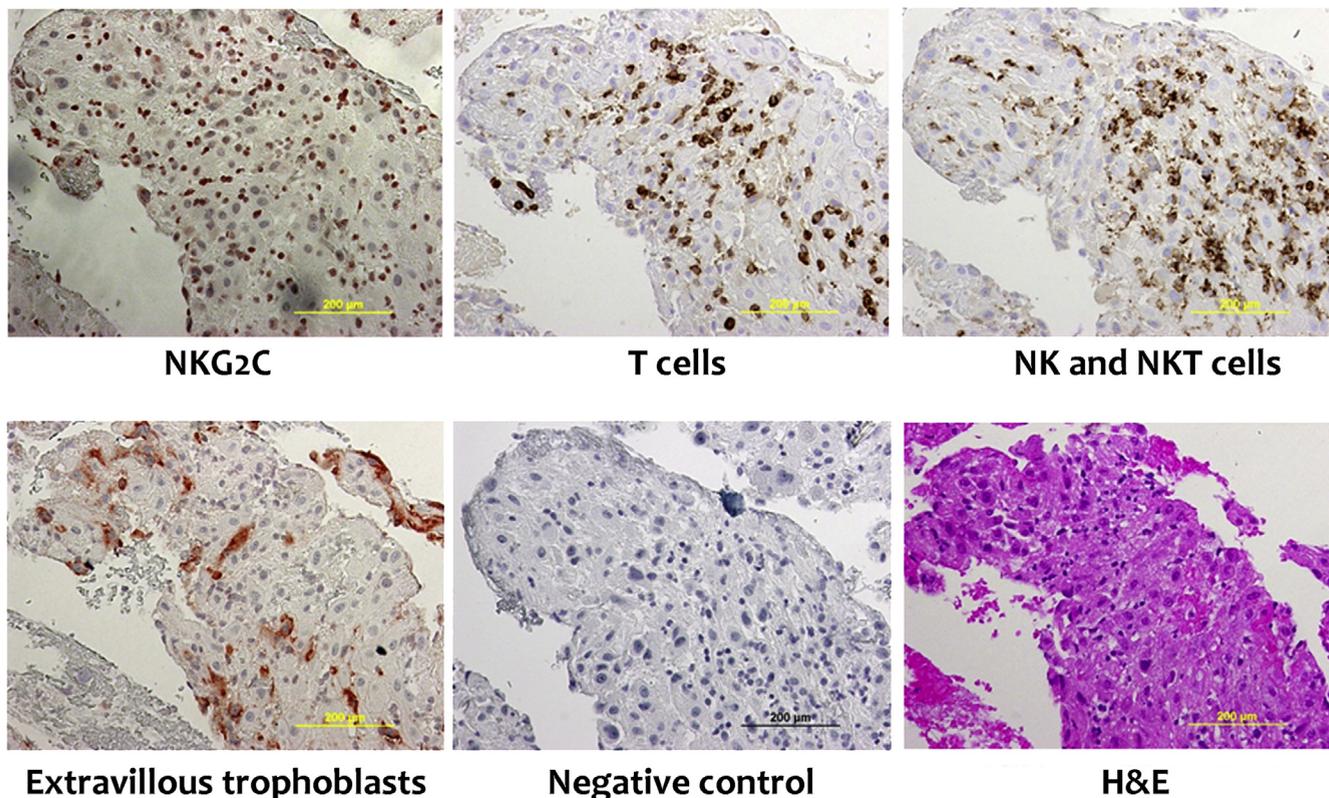
In summary, dNK cells are major players in placental bed remodeling, through secretion of growth factors, angiogenic factors, and chemokines and cytokines. Furthermore, dNKs are in active cross talk with trophoblasts migrating into the decidua and spiral arteries⁵⁵ but do not kill these invaders.^{50,61} Rather, they attract the extravillous trophoblasts essential for remodeling and conversion of the spiral arteries.⁵⁵ For these reasons, we propose that dNK cells may act as a reservoir of memory of pregnancy.

Memory-Like Properties of NK Cells

NK cells in mice and humans may possess memory-like properties. As NK cells are part of the innate rather than the adaptive immune system, finding NK cell memory was in some ways revolutionary. Memory in the immune system is composed of several steps: antigen specificity, clonal expansion, generation of long-lived memory cells, and the mounting of recall responses.^{62,63} In mice, these stages of memory have been demonstrated in peripheral blood NK cells in several experimental systems, including exposure to viral infections.^{62,64} Antigen-specific memory of NK cells to murine cytomegalovirus infection and expansion of these NK cells in subsequent murine cytomegalovirus exposure, along with mounting more efficient secondary and tertiary recall responses, have been demonstrated.⁶⁵ Thus, the criteria for immune memory have been met in the circulating pbNK cells in the murine system.

In humans, the evidence of memory in NK cells is derived mainly from the finding that a subset of human NK cells expressing the CD94/NKG2C receptor expands in response to human cytomegalovirus (CMV) antigens^{66,67} (reviewed in Rolle et al⁶²). In these studies of pbNK cells, expansion of the CD94/NKG2C NK cells receptor population may indicate a first step in the generation of NK cell memory. Evidence

FIGURE 2
Decidual lymphocyte immunohistochemistry profile



Immunohistochemistry of human decidual formalin-fixed, paraffin-embedded serial sections. Individual panels show immunohistochemistry for NKG2C expression, and identification of T, NK, NKT cells, and trophoblasts. A negative (no primary antibody) control and H&E staining are included. Immunohistochemistry was performed as routinely performed in our laboratory.⁵⁰ Antibodies used were antiCD56 and anti-CD3 (Zymed, South San Francisco, CA; Ventana automated stainer performed by our Pathology Department, Hadassah University Medical Center, Ein Kerem). Antibodies for trophoblast detection: HLA-G (a kind gift from Professor Mike McMaster, UCSF, clone 4H84), and for NKG2C bs 2416R (Bioss Antibodies, Woburn, MA). Immunohistochemistry detection was either with AEC (reddish-colored stain) or DAB substrate (brownish-colored stain) (Zytomed Systems, Berlin, Germany). AEC, 3-amino-9-ethylcarbazole; DAB, 3,3' diaminobenzidine; H&E, hematoxylin and eosin; NK, natural killer; NKT, natural killer T.

Goldman-Wohl. Learning from experience: cellular and molecular bases for improved outcome in subsequent pregnancies. *Am J Obstet Gynecol* 2019.

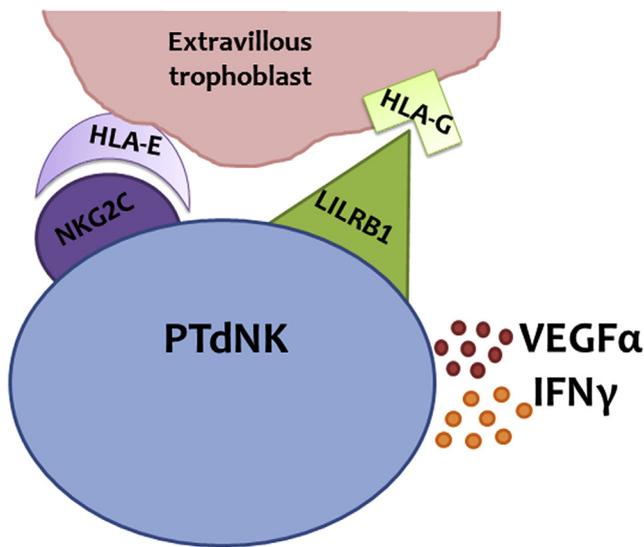
HLA-G,^{71–74} also was found to be upregulated in parous samples.

Because dNKs produce abundant chemokines, cytokines, and growth factors,⁵⁰ we explored whether growth factors were differentially secreted in dNKs of parous as compared with nulliparous women. As trophoblasts express the ligand HLA-E that engages with the dNK receptor, NKG2C, as well as the ligand HLA-G that engages with the dNK receptor, LILRB1, we asked whether the dNK with upregulated receptor expression (as in parous women) also secreted more growth factors when incubated with cells expressing their

ligands (NKG2C/HLA-E engagement and/or LILRB1/HLA-G engagement) (Figure 3). We found that secretion of VEGF and IFN- γ is induced on interaction of the extravillous trophoblast expressing ligands, HLA-E and HLA-G, with NKG2C and LILRB1 receptors of pregnancy-trained decidual natural killer (PTdNK), respectively. Indeed, there was significantly more secretion of the angiogenic factor VEGF α as well as IFN- γ in the cell population with the upregulated dNK receptors (as in parous women) (Figure 4¹⁸). These factors have been shown to be essential for angiogenesis and to possess properties

necessary for appropriate placental bed formation.⁷⁵ On finding that the dNK of parous women, as compared with primigravidae, express greater levels of NKG2C and LILRB1, possess a unique transcriptome and produce and secrete increased VEGF α and IFN- γ , we termed these “pregnancy-trained decidual natural killer” (ie, PTdNK) cells. Functionally, the supernatants derived from dNK cells of parous (with high levels of VEGF α) as compared with primiparous women promoted angiogenesis in both aortic ring outgrowth and tumor development assays.¹⁸ These experiments demonstrate,

FIGURE 3
EVT interacting with PTdNK induces secretion of growth factors



When EVTs expressing the ligands HLA-E and HLA-G interact with the receptors NKG2C and LILRB1, respectively, on PTdNK, both IFN- γ and VEGF α are secreted.

EVT, extravillous trophoblast; IFN- γ , interferon-gamma; PTdNK, pregnancy-trained decidual natural killer; VEGF α , vascular endothelial growth factor α .

Goldman-Wohl. Learning from experience: cellular and molecular bases for improved outcome in subsequent pregnancies. *Am J Obstet Gynecol* 2019.

for the first time, enhanced function in normal physiological memory of dNK cells in humans.

Epigenetic Memory of PTdNK Cells

To determine how this pregnancy-trained memory could be maintained, we proceeded to investigate the epigenome of PTdNKs as compared with primigravid dNKs, using the ATAC-seq (Assay for Transposase-Accessible Chromatin sequencing) technique, which allows one to identify regions of chromatin accessibility. In this assay, sequencing of DNA is performed on areas of chromatin that are accessible and exposed to enzymatic cleavage because the region is nucleosome free. These sequenced areas are thought to be available for active RNA transcription. The number of times a specific area of chromatin is sequenced is an indicator of its accessibility to proteins activating transcription, thus reflecting potential gene expression. The assay employs a

mutated hyperactive transposase enzyme that cuts DNA at transcriptionally active areas equivalent to DNase 1 hypersensitive sites. We used this technology on DNA isolated from the 2 classes of dNKs: those with high levels of NKG2C vs those negative for NKG2C expression. Overall, the PTdNKs were found to have a unique epigenetic profile. More specifically, focusing on IFNG and VEGF α , the NKG2C^{high} cells were shown to have a more “open” chromatin configuration,¹⁸ allowing for active transcription. These data corroborate the uniqueness of the PTdNK cell population and the observation of increased cytokine secretion from these cells as compared to primigravid dNKs (Figure 5).

PTdNK Cells Appear to Reside in the Endometrium Between Pregnancies

In light of these findings, we explored where PTdNKs might reside between pregnancies. Initially we considered

whether the reservoir of PTdNKs was to be found in the pbNK cells of women who had been pregnant as compared with pbNKs of nulligravidae women, but this was not observed to be significant. Knowing that NK cells reside in the endometrium (eNK),⁷⁶ we investigated whether the PTdNKs were present in the uterus between pregnancies. Menstrual blood was collected and the eNK cells from women who had been pregnant in the past and from nulligravidae were compared. Both NKG2C and LILRB1, receptors that were elevated in the decidua of parous women, were significantly increased in the eNKs isolated from the menstrual blood of parous as compared with nulligravidae women. Thus, the 2 receptors whose expansion we found elevated in the eNK of women between pregnancies, as compared with low levels in the nulliparous women. These results suggest that the memory of PTdNK cells likely resides in eNK cells.

To characterize these cells on the epigenetic level and determine chromatin areas “open” for transcription, we performed ATACseq on the NKG2C^{high} cells isolated from (menstrual) eNK cells (nulligravidae vs parous) and dNK cells (primigravidae vs parous). We were taken aback when we noted that the eNK epigenetic profile of parous women was strikingly similar to dNKs and quite dissimilar to eNK of nulligravidae. Thus, the PTdNK memory is found in the epigenome of NK cells of the endometrium, as depicted in Figure 5.

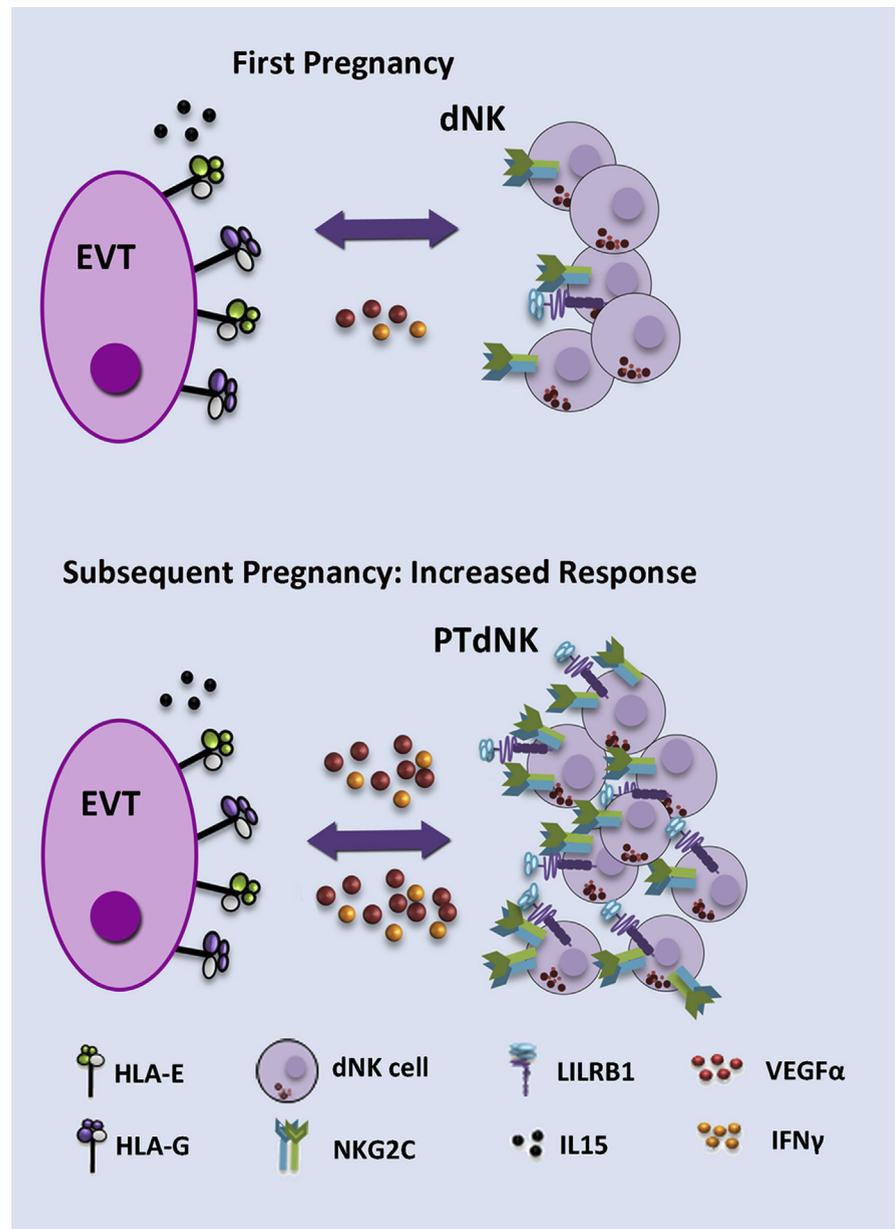
Linking Memory of dNK Cells with Reduced Preeclampsia Risk

Our findings indicate that dNK cells are primed and established in the first trimester of first pregnancy and subsequently possess properties of trained memory, hence PTdNK cells. These dNK cells provide beneficial factors necessary for the development of the placental bed and the maintenance of pregnancy.^{50,57,77} These PTdNKs, residing in the endometrium between pregnancies, have memory that allows them to expand more easily or earlier

than in first pregnancy, thus enhancing their functions (Figures 4 and 5). Placentation would be augmented and spiral artery conversion would be more easily achieved in parous women; consequently, pregnancy disorders with an origin in the placental bed would be less likely in these women. We propose that memory of pregnancy generated by dNK cells may be the missing link joining the immunological underpinning of preeclampsia with the additional risk of preeclampsia in first pregnancy.^{78–80} We suggest that on interaction with the decidual environment, including but not limited to trophoblasts, the PTdNKs, by producing greater levels of factors known to be essential for successful remodeling of the maternal–fetal interface (chemokines, cytokines, growth and angiogenic factors), enhance placentation/implantation and thus reduce the risk of preeclampsia.

A caveat to these findings is the lack of data concerning paternity in subsequent pregnancies. Given that there is a primipaternity model, as compared with primigravidity as the risk factor for the preeclampsia, it would be of interest to investigate PTdNK from pregnancies with a change in paternity or with a long interval between pregnancies.^{81–83} Perhaps the extravillous trophoblast interaction with PTdNK, which results in enhanced expression of proteins related to the placental bed, may be influenced by a change in paternity. An additional avenue of future investigation may be to characterize the eNK in menstrual blood in women suffering from recurrent spontaneous abortion. It is possible that their eNK “do not remember” pregnancy and hence their subsequent pregnancies are not at an advantage for placentation and success. Translational research may be along the lines of finding a way to prime the eNK of these women toward eNK of the type observed between pregnancies. Furthermore, given that particular combinations of maternal KIR receptors on dNK and fetal expression of HLA-C influence the risk for preeclampsia,⁸⁴ it will be interesting to continue this investigation, as a subset of PTdNK expresses KIR2DL2-3.¹⁸

FIGURE 4
Proposed model



HLA-E and HLA-G are expressed on the EVT of the placenta. During first pregnancy, we assume that a priming process is initiated. NK cells of first pregnancies express low levels of NKG2C and LILRB1. In subsequent pregnancies, the decidual cells are already primed, thus rapidly expand expression of NK receptors NKG2C and LILRB1 and secrete greater amounts of VEGF α and IFN- γ , which enhance vascularization and proliferation. Modified from Gamliel et al.¹⁸

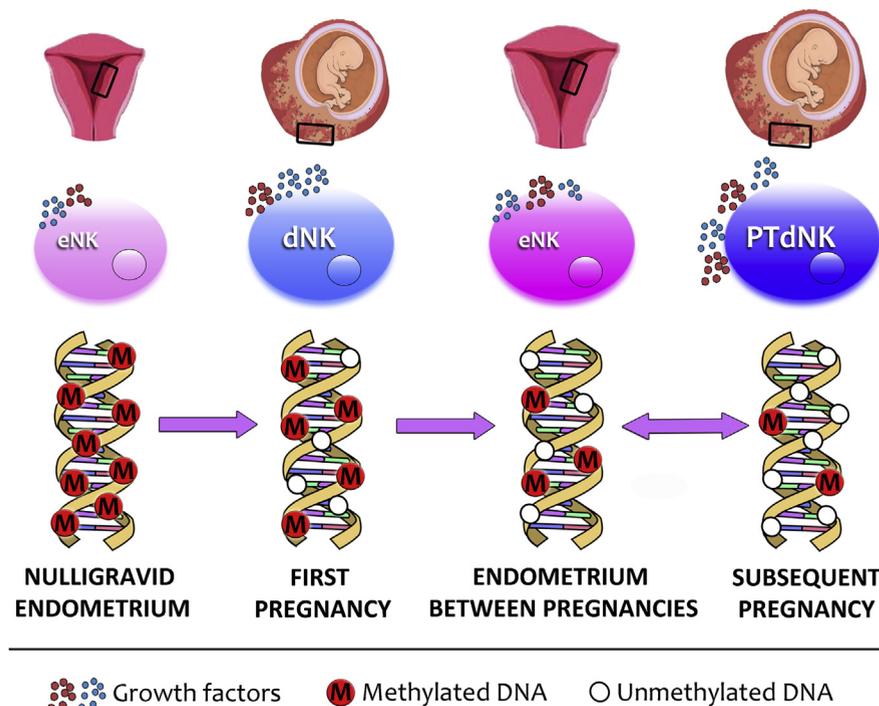
EVT, extravillous trophoblast; IFN- γ , interferon-gamma; NK, natural killer; VEGF α , vascular endothelial growth factor.

Goldman-Wohl. Learning from experience: cellular and molecular bases for improved outcome in subsequent pregnancies. *Am J Obstet Gynecol* 2019.

We can also speculate that as abnormal invasive placentation is associated with pregnancies following uterine surgery and damaged or deficient

decidua, lack of sufficient PTdNK interaction with trophoblasts may be part of the puzzle of abnormal invasive trophoblast.

FIGURE 5
Summary model



Nulligravid women have a relatively “closed” epigenetic profile of specific genes in their eNK cells (M in red circle, methylation). In first pregnancy, there is enhanced expression of dNK cell receptors and growth factors (colored red and blue dots) as well as a more “open” (white circles on chromatin) epigenetic configuration. Some of this epigenetic memory of hypomethylation remains in eNK between pregnancies. In subsequent pregnancies, the educated PTdNK produce significantly greater amounts of growth factors, thereby enhancing development of the placental bed with the epigenetic signature found in a more “open” configuration.

dNK, decidual natural killer; eNK, endometrial natural killer; PTdNK, pregnancy-trained decidual natural killer.

Goldman-Wohl. Learning from experience: cellular and molecular bases for improved outcome in subsequent pregnancies. *Am J Obstet Gynecol* 2019.

Epigenetic Memory May Explain a Wide Range of Observed Improved Outcomes in Subsequent Pregnancy and Lactation

We have provided cellular and molecular epigenetic evidence supporting the notion that the immune cells of the fetal–maternal interface recall passed pregnancies and maintain characteristics that enhance future pregnancies. This may have bearing on other, often observed instances of better outcomes in subsequent pregnancies.

Maternal cardiovascular system adaptations to pregnancy also have been investigated. Clapp and Capeless showed that measurable changes to cardiac functional parameters: increased end-diastolic volume, stroke volume, and

cardiac output and decreased vascular resistance are observed in first pregnancies, persist post-partum, and are enhanced in subsequent pregnancies.¹³

More recently, Morris et al showed similar persistent changes to vascular compliance: decreased mean arterial pressure and reduced arterial stiffness during and following first pregnancy.¹⁴ Cardiac adaptation to subsequent pregnancy was compared in women who were preeclamptic in their first pregnancy.¹² The investigators found that women in whom preeclampsia recurred had lower left ventricular mass and stroke volume than those in whom preeclampsia did not recur, during the interpregnancy interval; however, adaptation to pregnancy was similar in the 2

groups.¹² Although these studies focused on physiologic response to pregnancy without corroboration from molecular epigenetic investigation, it is tempting to speculate whether the memory of pregnancy is recalled by myocardial or vascular endothelial cells or others.

Another example would be the well-known differences between the length of the first and second stages of labor in first vs subsequent deliveries. Evidence for epigenetic memory in the uterus or cervix that could provide a molecular basis for the observed greater efficiency of labor and delivery remains to be described. However, Seaborne et al observed muscle memory of exercise on both the epigenetic and consequently the transcriptional level in humans. They showed that induced muscle hypertrophy invoked a “signature” of epigenetic memory by hypomethylation of specific genes in human skeletal muscle.⁸⁵

Although the study by Seaborne et al refers to skeletal muscle and not uterine muscle, we may conjecture that shorter first and second stages of labor and better outcome in subsequent deliveries may be related to epigenetic memory in uterine and cervical muscle.

Lactation is yet another aspect of reproduction in which subsequent pregnancies have an advantage over first. Significantly more milk is produced in second as compared with first pregnancies.¹⁶ Although in humans epigenetic memory for more robust lactation in subsequent pregnancies has not been found, such epigenetic memory was found in the mouse mammary gland.¹⁷ In the parous mouse, a more-robust response of the mammary glands to subsequent pregnancy has been shown.¹⁷ The physiological response involves both the expansion of ductal structures and synthesis of milk proteins earlier in pregnancy, in parous as compared with primiparous mice. The rapid response of gene expression is conferred though epigenetic memory of pregnancy by master regulators of mammary gland transcription. Hypomethylated regions of transcriptional regulation persist as stable changes. They are first induced in the mammary epithelium by pregnancy primed genes

and produce greater responses to hormone exposure. This may in turn result in a mammary gland that functions more effectively during subsequent pregnancies. These targeted experiments demonstrated that genes primed by parity-associated epigenomic changes are poised for more rapid reactivation in a subsequent pregnancy, supporting the existence of an epigenetic memory of past pregnancies.¹⁷

In conclusion, we have delineated some of the many aspects of pregnancy, childbirth, and lactation wherein subsequent pregnancies have been shown to be more robust than first. Our investigations have shown that NK cells at the maternal–fetal interface act as builders of the maternal–fetal interface, promoting angiogenesis, attracting trophoblasts and producing chemokines, cytokines, and growth factors essential for conversion of the spiral arteries. Our most recent work has further shown that “primed” PTdNK cells recall pregnancy on an epigenetic level, which may contribute to their more rapidly and successfully remodeling the placental bed. This offers an explanation as to why first pregnancy, where dNK cells are possibly not yet primed in their functions, is a risk factor for the development of preeclampsia, and others of the “great obstetrical syndromes.” Molecular epigenetic evidence in humans for other examples of enhanced pregnancy and lactation outcomes remain to be investigated. For now, we speculate that investigating the activation of dNK may yield treatment modalities for women at risk for disorders of primigravidity originating at the placental bed. ■

ACKNOWLEDGMENTS

The authors thank the past and present members of The Magda and Richard Hoffman Center for Human Placenta Research and the Mandelboim Laboratory for helpful insights and discussions. We thank Professor Ilana Ariel, Caryn Greenfield, and Galina Skarzynski, PhD, of the Department of Pathology, Hadassah-Hebrew University Medical Center, Mt. Scopus (I.A., G.S.) and Magda and Richard Hoffman Center for Human Placenta Research (C.G.), for Figure 2. We thank Sarah M. Cohen, MPH, of the Department of Obstetrics and Gynecology, Hadassah-Hebrew University Medical Center,

Mt. Scopus, for preparing and modifying the figures and for expert editing of the manuscript.

REFERENCES

- Kramer MS. Determinants of low birth weight: methodological assessment and meta-analysis. *Bull World Health Organ* 1987;65:663–737.
- Stillbirth Collaborative Research Network Writing Group. Association between stillbirth and risk factors known at pregnancy confirmation. *JAMA* 2011;306:2469–79.
- Iams JD, Lynch CD. Stillbirth and lessons for pregnancy care. *JAMA* 2011;306:2506–7.
- Ballering G, Leijnse J, Eijkelkamp N, Peeters L, de Heus R. First-trimester placental vascular development in multiparous women differs from that in nulliparous women. *J Matern Fetal Neonatal Med* 2018;31:209–15.
- Roland MC, Friis CM, Godang K, Bollerslev J, Haugen G, Henriksen T. Maternal factors associated with fetal growth and birthweight are independent determinants of placental weight and exhibit differential effects by fetal sex. *PLoS ONE* 2014;9:e87303.
- Wallace JM, Bhattacharya S, Horgan GW. Gestational age, gender and parity specific centile charts for placental weight for singleton deliveries in Aberdeen, UK. *Placenta* 2013;34:269–74.
- Hinkle SN, Albert PS, Mendola P, et al. The association between parity and birthweight in a longitudinal consecutive pregnancy cohort. *Paediatr Perinat Epidemiol* 2014;28:106–15.
- Shah PS, Knowledge Synthesis Group on Determinants of LBW/PT births. Parity and low birth weight and preterm birth: a systematic review and meta-analyses. *Acta Obstet Gynecol Scand* 2010;89:862–75.
- McCowan L, Horgan RP. Risk factors for small for gestational age infants. *Best Pract Res Clin Obstet Gynaecol* 2009;23:779–93.
- Thompson JM, Clark PM, Robinson E, et al. Risk factors for small-for-gestational-age babies: the Auckland Birthweight Collaborative Study. *J Paediatr Child Health* 2001;37:369–75.
- Brosens I, Pijnenborg R, Vercruyse L, Romero R. The “great obstetrical syndromes” are associated with disorders of deep placentation. *Am J Obstet Gynecol* 2011;204:193–201.
- Ghossein-Doha C, Spaanderman ME, Al Doulah R, Van Kuijk SM, Peeters LL. Maternal cardiac adaptation to subsequent pregnancy in formerly pre-eclamptic women according to recurrence of pre-eclampsia. *Ultrasound Obstet Gynecol* 2016;47:96–103.
- Clapp JF, 3rd, Capeless E. Cardiovascular function before, during, and after the first and subsequent pregnancies. *Am J Cardiol* 1997;80:1469–73.
- Morris EA, Hale SA, Badger GJ, Magness RR, Bernstein IM. Pregnancy induces persistent changes in vascular compliance in primiparous women. *Am J Obstet Gynecol* 2015;212:633 e631–6.
- Kilpatrick SG, Garrison E. Normal labor and delivery. In: Gabbe SG, ed. *Obstetrics: normal and problem pregnancies*, 7th ed. Philadelphia: Elsevier; 2017. p. 246–70.
- Ingram J, Woolridge M, Greenwood R. Breastfeeding: it is worth trying with the second baby. *Lancet* 2001;358:986–7.
- Dos Santos CO, Dolzhenko E, Hodges E, Smith AD, Hannon GJ. An epigenetic memory of pregnancy in the mouse mammary gland. *Cell Rep* 2015;11:1102–9.
- Gamliel M, Goldman-Wohl D, Isaacson B, et al. Trained memory of human uterine NK cells enhances their function in subsequent pregnancies. *Immunity* 2018;48:951–62 e955.
- Prefumo F, Ganapathy R, Thilaganathan B, Sebire NJ. Influence of parity on first trimester endovascular trophoblast invasion. *Fertil Steril* 2006;85:1032–6.
- Khong TY, Adema ED, Erwich JJ. On an anatomical basis for the increase in birth weight in second and subsequent born children. *Placenta* 2003;24:348–53.
- Bartley J, Tong S, Everington D, Baird DT. Parity is a major determinant of success rate in medical abortion: a retrospective analysis of 3161 consecutive cases of early medical abortion treated with reduced doses of mifepristone and vaginal gemeprost. *Contraception* 2000;62:297–303.
- Haimov-Kochman R, Arbel R, Sciaky-Tamir Y, Brzezinski A, Laufer N, Yagel S. Risk factors for unsuccessful medical abortion with mifepristone and misoprostol. *Acta Obstet Gynecol Scand* 2007;86:462–6.
- Odeh M, Tendler R, Sosnovsky V, Kais M, Ophir E, Bornstein J. The effect of parity and gravidity on the outcome of medical termination of pregnancy. *Isr Med Assoc J* 2010;12:606–8.
- Lefebvre P, Cotte M, Monniez N, Norel G. The role of parity in medical abortion up to 49 days of amenorrhoea. *Eur J Contracept Reprod Health Care* 2008;13:404–11.
- Luo ZC, An N, Xu HR, Larante A, Audibert F, Fraser WD. The effects and mechanisms of primiparity on the risk of pre-eclampsia: a systematic review. *Paediatr Perinat Epidemiol* 2007;21(suppl 1):36–45.
- Goldman-Wohl DS, Yagel S. Examination of distinct fetal and maternal molecular pathways suggests a mechanism for the development of preeclampsia. *J Reprod Immunol* 2007;76:54–60.
- Karumanchi SA, Maynard SE, Stillman IE, Epstein FH, Sukhatme VP. Preeclampsia: a renal perspective. *Kidney Int* 2005;67:2101–13.
- Sole KB, Staff AC, Laine K. The association of maternal country of birth and education with hypertensive disorders of pregnancy: a population-based study of 960 516 deliveries in Norway. *Acta Obstet Gynecol Scand* 2018;97:1237–47.
- Bell MJ. A historical overview of pre-eclampsia-eclampsia. *J Obstet Gynecol Neonatal Nurs* 2010;39:510–8.

30. Chamberlen H, translator. The Fourth Edition Corrected, and Augmented with Several New Figures. 1710; Available at: http://find.galegroup.com/ecco/infomark.do?&contentSet=ECCOArticles&type=multipage&tabID=T001&prodId=ECCO&docId=CW3309817161&source=gale&userGroupName=upitt_main&version=1.0&docLevel=FASCIMILE, 2018. Accessed June 3, 2018.
31. McMillen S. Eclampsia. In: Kiple KF, ed. The Cambridge Historical Dictionary of Disease. New York, NY: Cambridge University Press; 2003. p. 110–2.
32. Labarrere CA, DiCarlo HL, Bammerlin E, et al. Failure of physiologic transformation of spiral arteries, endothelial and trophoblast cell activation, and acute atherosclerosis in the basal plate of the placenta. *Am J Obstet Gynecol* 2017;216:287 e281–7 e216.
33. Roberts JM. The perplexing pregnancy disorder preeclampsia: what next? *Physiol Genomics* 2018;50:459–67.
34. Pijnenborg R, Anthony J, Davey DA, et al. Placental bed spiral arteries in the hypertensive disorders of pregnancy. *Br J Obstet Gynaecol* 1991;98:648–55.
35. Benton SJ, Leavey K, Grynspan D, Cox BJ, Bainbridge SA. The clinical heterogeneity of preeclampsia is related to both placental gene expression and placental histopathology. *Am J Obstet Gynecol* 2018;219:604 e601–4 e625.
36. Jauniaux E, Poston L, Burton GJ. Placental-related diseases of pregnancy: involvement of oxidative stress and implications in human evolution. *Hum Reprod Update* 2006;12:747–55.
37. Hung TH, Burton GJ. Hypoxia and reoxygenation: a possible mechanism for placental oxidative stress in preeclampsia. *Taiwanese J Obstet Gynecol* 2006;45:189–200.
38. Burton GJ, Jauniaux E. Pathophysiology of placental-derived fetal growth restriction. *Am J Obstet Gynecol* 2018;218:S745–61.
39. Maynard SE, Min JY, Merchan J, et al. Excess placental soluble fms-like tyrosine kinase 1 (Sflt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. *J Clin Invest* 2003;111:649–58.
40. Levine RJ, Maynard SE, Qian C, et al. Circulating angiogenic factors and the risk of preeclampsia. *N Engl J Med* 2004;350:672–83.
41. Levine RJ, Lam C, Qian C, et al. Soluble endoglin and other circulating antiangiogenic factors in preeclampsia. *N Engl J Med* 2006;355:992–1005.
42. Redman CW, Sargent IL. Latest advances in understanding preeclampsia. *Science* 2005;308:1592–4.
43. Redman CW, Sacks GP, Sargent IL. Preeclampsia: an excessive maternal inflammatory response to pregnancy. *Am J Obstet Gynecol* 1999;180:499–506.
44. Roberts JM, Hubel CA. The two stage model of preeclampsia: variations on the theme. *Placenta* 2009;30:S32–7.
45. Roberts JM. Endothelial dysfunction in preeclampsia. *Semin Reprod Endocrinol* 1998;16:5–15.
46. Redman CW, Sargent IL. Placental stress and pre-eclampsia: a revised view. *Placenta* 2009;30(suppl A):S38–42.
47. Dor Y, Cedar H. Principles of DNA methylation and their implications for biology and medicine. *Lancet* 2018;392:777–86.
48. D'Urso A, Brickner JH. Mechanisms of epigenetic memory. *Trends Genet* 2014;30:230–6.
49. Jonjic S, Babic M, Polic B, Krmpotic A. Immune evasion of natural killer cells by viruses. *Curr Opin Immunol* 2008;20:30–8.
50. Hanna J, Goldman-Wohl D, Hamani Y, et al. Decidual Nk cells regulate key developmental processes at the human fetal-maternal interface. *Nat Med* 2006;12:1065–74.
51. Vento-Tormo R, Efremova M, Botting RA, et al. Single-cell reconstruction of the early maternal-fetal interface in humans. *Nature* 2018;563:347–53.
52. Suryawanshi H, Morozov P, Straus A, et al. A single-cell survey of the human first-trimester placenta and decidua. *Sci Adv* 2018;4:eau4788.
53. Liu Y, Fan X, Wang R, et al. Single-cell RNA-Seq reveals the diversity of trophoblast subtypes and patterns of differentiation in the human placenta. *Cell Res* 2018;28:819–32.
54. Saito S, Kasahara T, Sakakura S, et al. Interleukin-8 production by Cd16–Cd56bright natural killer cells in the human early pregnancy decidua. *Biochem Biophys Res Commun* 1994;200:378–83.
55. Hanna J, Wald O, Goldman-Wohl D, et al. Cxcl12 expression by invasive trophoblasts induces the specific migration of Cd16– human natural killer cells. *Blood* 2003;102:1569–77.
56. Koopman LA, Kopicow HD, Rybalov B, et al. Human decidua natural killer cells are a unique Nk cell subset with immunomodulatory potential. *J Exp Med* 2003;198:1201–12.
57. Ashkar AA, Croy BA. Functions of uterine natural killer cells are mediated by interferon gamma production during murine pregnancy. *Semin Immunol* 2001;13:235–41.
58. Ratsep MT, Felker AM, Kay VR, Toluoso L, Hofmann AP, Croy BA. Uterine natural killer cells: supervisors of vasculature construction in early decidua basalis. *Reproduction* 2015;149:R91–102.
59. Renaud SJ, Scott RL, Chakraborty D, Rumi MA, Soares MJ. Natural killer-cell deficiency alters placental development in rats. *Biol Reprod* 2017;96:145–58.
60. Golic M, Haase N, Herse F, et al. Natural killer cell reduction and uteroplacental vasculopathy. *Hypertension* 2016;68:964–73.
61. El Costa H, Tabiasco J, Berrebi A, et al. Effector functions of human decidua nk cells in healthy early pregnancy are dependent on the specific engagement of natural cytotoxicity receptors. *J Reprod Immunol* 2009;82:142–7.
62. Rolle A, Pollmann J, Cerwenka A. Memory of infections: an emerging role for natural killer cells. *PLoS Pathog* 2013;9:e1003548.
63. Sun JC, Ugolini S, Vivier E. Immunological memory within the innate immune system. *EMBO J* 2014;33:1295–303.
64. Paust S, Gill HS, Wang BZ, et al. Critical role for the chemokine receptor Cxcr6 in Nk cell-mediated antigen-specific memory of haptens and viruses. *Nat Immunol* 2010;11:1127–35.
65. Sun JC, Beilke JN, Lanier LL. Adaptive immune features of natural killer cells. *Nature* 2009;457:557–61.
66. Guma M, Angulo A, Vilches C, Gomez-Lozano N, Malats N, Lopez-Botet M. Imprint of human cytomegalovirus infection on the nk cell receptor repertoire. *Blood* 2004;104:3664–71.
67. Guma M, Budt M, Saez A, et al. Expansion of Cd94/Nkg2c+ Nk cells in response to human cytomegalovirus-infected fibroblasts. *Blood* 2006;107:3624–31.
68. Foley B, Cooley S, Vemeris MR, et al. Human cytomegalovirus (Cmv)-induced memory-like Nkg2c(+) Nk cells are transplantable and expand in vivo in response to recipient cmv antigen. *J Immunol* 2012;189:5082–8.
69. Hammer Q, Ruckert T, Borst EM, et al. Peptide-specific recognition of human cytomegalovirus strains controls adaptive natural killer cells. *Nat Immunol* 2018;19:453–63.
70. Siewiera J, El Costa H, Tabiasco J, et al. Human cytomegalovirus infection elicits new decidua natural killer cell effector functions. *PLoS Pathog* 2013;9:e1003257.
71. Gonen-Gross T, Achdout H, Gazit R, et al. Complexes of Hla-G protein on the cell surface are important for leukocyte Ig-like receptor-1 function. *J Immunol* 2003;171:1343–51.
72. Gonen-Gross T, Gazit R, Achdout H, et al. Special organization of the Hla-G protein on the cell surface. *Hum Immunol* 2003;64:1011–6.
73. Gonen-Gross T, Achdout H, Arnon TI, et al. The Cd85j/leukocyte inhibitory receptor-1 distinguishes between conformed and beta 2-microglobulin-Free Hla-G molecules. *J Immunol* 2005;175:4866–74.
74. Navarro F, Liano M, Bellon T, Colonna M, Geraghty DE, Lopez-Botet M. The Ilt2(Lir1) and Cd94/Nkg2a Nk cell receptors respectively recognize Hla-G1 and Hla-E molecules co-expressed on target cells. *Eur J Immunol* 1999;29:277–83.
75. Ashkar AA, Croy BA. Interferon-gamma contributes to the normalcy of murine pregnancy. *Biol Reprod* 1999;61:493–502.
76. Manaster I, Mizrahi S, Goldman-Wohl D, et al. Endometrial Nk Cells are special immature cells that await pregnancy. *J Immunol* 2008;181:1869–76.
77. Lash GE, Schiessl B, Kirkley M, et al. Expression of angiogenic growth factors by uterine natural killer cells during early pregnancy. *J Leukoc Biol* 2006;80:572–80.
78. Borzychowski AM, Croy BA, Chan WL, Redman CW, Sargent IL. Changes in systemic type 1 and type 2 immunity in normal pregnancy

and pre-eclampsia may be mediated by natural killer cells. *Eur J Immunol* 2005;35:3054–63.

79. Robillard PY, Dekker G, Chaouat G, Hulse TC, Saftlas A. Epidemiological studies on primipaternity and immunology in preeclampsia—a statement after twelve years of workshops. *J Reprod Immunol* 2011;89:104–17.

80. Redman CW, Sargent IL. Immunology of pre-eclampsia. *Am J Reprod Immunol* 2010;63:534–43.

81. Robillard PY, Dekker GA, Hulse TC. Revisiting the epidemiological standard of preeclampsia: primigravidity or primipaternity? *Eur J Obstet Gynecol Reprod Biol* 1999;84:37–41.

82. Skjaerven R, Wilcox AJ, Lie RT. The interval between pregnancies and the risk of pre-eclampsia. *N Engl J Med* 2002;346:33–8.

83. Hercus A, Dekker G, Leemaqz S. Primipaternity and birth interval; independent risk

factors for preeclampsia. *J Matern Fetal Neonatal Med* 2019 [Epub ahead of print].

84. Hiby SE, Walker JJ, O’Shaughnessy KM, et al. Combinations of maternal Kir and fetal HLA-C genes influence the risk of preeclampsia and reproductive success. *J Exp Med* 2004;200:957–65.

85. Seaborne RA, Strauss J, Cocks M, et al. Human skeletal muscle possesses an epigenetic memory of hypertrophy. *Sci Rep* 2018;8:1898.