



ELSEVIER

Available online at www.sciencedirect.com

Journal of Hospital Infection

journal homepage: www.elsevier.com/locate/jhin



Lowbury Lecture

Chlorhexidine-based decolonization to reduce healthcare-associated infections and multidrug-resistant organisms (MDROs): who, what, where, when, and why?

S.S. Huang

Division of Infectious Diseases and Health Policy Research Institute, University of California Irvine School of Medicine, Irvine, California, USA

ARTICLE INFO

Article history:

Received 29 August 2019
Accepted 29 August 2019
Available online 5 September 2019

Key words:

Chlorhexidine
Decolonization
MRSA
Multidrug-resistant bacteria



SUMMARY

Body surface decolonization with chlorhexidine bathing and nasal mupirocin has become a simple solution for prevention of healthcare-associated infections. The clinical trial evidence for this practice will be reviewed to understand who benefits from this practice, for what reasons, and at what times. The method of bathing and nasal decolonization will also be discussed as proper application is needed for maximal effectiveness. Finally, the conflict between current effectiveness and future potential for fueling resistance is considered.

© 2019 The Healthcare Infection Society. Published by Elsevier Ltd. All rights reserved.

Introduction

From its inception, the medical profession has been steeped in a deep desire to treat, cure, and prevent suffering. A great deal of the diseases that have long occupied us involve contagion. Despite centuries of exceptional progress in thwarting infections, we still strive to conquer the unabating remnant of pathogens that plague us.

In that infinite journey, there have been inspirations so simple that they evoke disbelief prior to being widely adopted as best practice. Once adopted, there is an equally

fervent disbelief that the historical reality was ever perceived as acceptable. For infection prevention, some of these transformative innovations for preventing healthcare-associated infections include hand hygiene to prevent puerperal fever, [1] surgical sterility and skin preparation, [2,3] alcohol-based hand sanitizer, [4] and sealed urinary catheters [5].

Likewise, the simple concept of using antiseptics for full body bathing and showering has been broadly adopted in healthcare for high-risk patient populations to prevent infection. This concept was pioneered by those who saw potential for infection prevention beyond its effective use for hands and pre-operative skin preparation.

E-mail address: susan.huang@uci.edu.

<https://doi.org/10.1016/j.jhin.2019.08.025>

0195-6701/© 2019 The Healthcare Infection Society. Published by Elsevier Ltd. All rights reserved.

While there are several antiseptic products available for body bathing (e.g. bleach, chlorhexidine, tea tree oil, octenidine, and others), this discussion will focus on recent large-scale studies and trials involving full body chlorhexidine (CHG) bathing to reduce healthcare-associated infections (HAIs) and multidrug resistant organisms (MDROs). These studies often include nasal decolonization products, such as 2% mupirocin nasal ointment and 10% povidone-iodine, because of their ability to address the nasal reservoir (and thereby body reservoir) of *Staphylococcus aureus*, while CHG is more broadly active and can reduce body bioburden from a wide variety of human pathogens and commensals.

A brief history of CHG in tribute to Edward Lowbury

CHG was first discovered in the early 1950s by a chemical company in the United Kingdom, and was rapidly commercialized as a broadly active antiseptic in 1954 [6]. Its mechanism of action is based upon cationic properties, which allow disruption of microbial cell surfaces and cell death at concentrations as low as 0.01%. Dr. Edward Joseph Lister Lowbury was the first to perform comparative effectiveness studies of soap, CHG, and other antiseptics for single and repeated hand and focal pre-operative skin disinfection [7–11]. He was also the first to describe their differential effects on removing superficial bacterial skin contamination and removing resident bacteria that surfaces from deeper skin layers [12–14]. His work was later extended by his colleague and successor Graham Ayliffe, who was among the first to study the benefits of CHG for pre-surgical full body bathing [15,16].

Initially, and for many decades, CHG was used in healthcare for focal skin and mucosal cleansing. It was commonly used as a hand antiseptic at concentrations of 4% or less, and also in dilute form for dental hygiene to treat periodontitis [17]. Eventually, published trials codified the superiority of CHG over povidone-iodine for skin disinfection prior to central line placement and surgical incision, both with and without concurrent alcohol [18–22].

The work by Ayliffe and others on full body antiseptic bathing ultimately led to the universal recommendation for full body pre-operative antiseptic bathing in the 1999 US Healthcare Infection Control Practices Advisory Committee (HICPAC) surgical site

infection guidelines [23]. This experience opened the door to pioneering efforts by Robert A. Weinstein who was the first to explore the value of routine daily full body CHG bathing to prevent infections in intensive care units (ICUs) [24–27]. For the purposes here, the term decolonization refers to the use of CHG for full body bathing or showering with or without concomitant nasal products to reduce carriage of *S. aureus*.

Why decolonize in healthcare facilities?

The drive to decolonize as a strategy to reduce infections and MDROs in healthcare arose from public and provider outcry that HAIs unnecessarily occur because of failure to perform preventative steps, some of which are yet to be discovered. The response to this outcry was a genuine quest to achieve the lowest possible levels of HAI – striving for zero cases for greater and greater lengths of time.

Decolonization focuses on bacterial carriage as an endogenous source of infection in highly vulnerable individuals and situations. It is well known that humans extensively shed bacterial pathogens, which then contaminate the environment and provide a series of opportunities for spreading pathogens to others (Figure 1). Several infection prevention activities counter these opportunities, but it is notable that decolonization works upstream of the event cascade, thereby preventing the shedding of pathogens, [27] preventing contamination of the environment and healthcare worker hands, [24] preventing acquisition of multidrug-resistant organisms (MDROs) and other pathogens, and ultimately preventing infection [25,26].

Another important reason to favour decolonization as one of several critical infection prevention strategies is because it is the only strategy that helps those who already harbor MDROs. Most infection prevention strategies (e.g. environmental cleaning, hand hygiene, contact precautions, active screening) are designed to prevent spread of MDROs (or other pathogens) to those who do not yet harbour them. Decolonization provides a universal approach by protecting both MDRO carriers and non-carriers. This is increasingly important in most hospitals, where an increasing proportion of patients asymptotically harbour MDROs over time. In the United States, nearly 15% of hospitalized patients asymptotically harbor an MDRO, [28] with higher estimates in ICUs [29,30]. Admission prevalence of

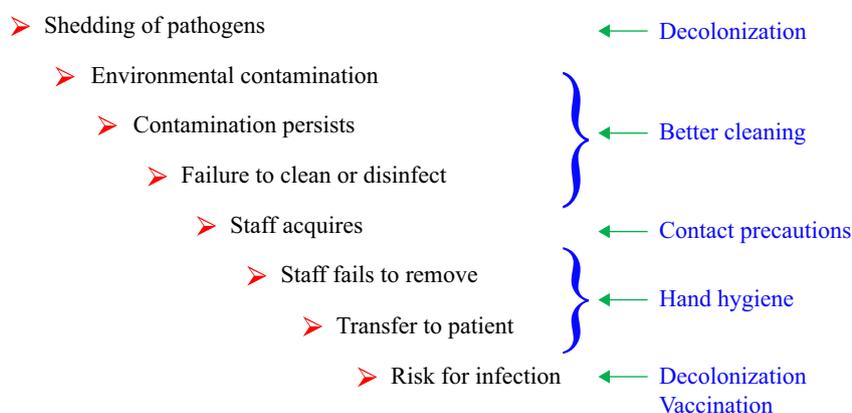


Figure 1. Figure displaying a series of events that enable patient-to-patient transmission of pathogens to occur along with their associated risk of healthcare-associated infection. Also displayed are common infection prevention strategies to mitigate these events.

resistant gram-negative bacteria alone is 10% among German tertiary care centers [31,32]. In nursing homes or care homes for the elderly, estimates range from 10–20% in Belgium, Germany, and Spain [33–35]; 20–30% in the UK and Hong Kong [36,37]; 40–65% in the US [38–41]; and up to 80% in Italy as well as US long-term acute care hospitals [38,42].

Furthermore, decolonization with topical CHG is superior to regular soap not only because of its antiseptic properties, but also because it binds to skin proteins and continues to exert its antiseptic activities on the skin for up to 24 hours [27,43–45]. Hence, the concept of daily CHG bathing is intended to provide continuous protection from HAI during a hospital stay. This is in contrast to alcohol-based hand hygiene or soap and water where lack of residual activity allows contamination to occur when touching objects or people immediately after use.

In the next section, the findings of CHG decolonization trials will be discussed. These trials and other studies show that topical CHG bathing has a legacy of preventing infections due to pathogens for which the skin is not the primary body reservoir. For example, among MDROs, the primary reservoir of methicillin-resistant *S. aureus* (MRSA) is the nose, and the primary reservoir for vancomycin-resistant enterococci (VRE), extended-spectrum beta-lactamase producers (ESBL), and carbapenem-resistant Enterobacteriaceae (CRE) is the gut. Nevertheless, study after study demonstrates that topical decolonization of the skin (with or without addressing the nasal reservoir) can prevent a significant portion of healthcare-associated infections from these and other pathogens in a wide array of patient populations. This emphasizes the importance of skin integrity and cleanliness for preserving health due to endogenous and exogenous pathogen threats.

Who should receive CHG decolonization?

CHG decolonization is a strategy for those at high risk for infection. These include those who are vulnerable because of host characteristics or circumstance. Initial trials for CHG decolonization were focused on **targeted decolonization for secondary prevention** in those with recurrent episodes of *S. aureus* disease [46]. In the past decade, large-scale randomized trials (Table 1) have shifted focus to **targeted decolonization for primary prevention**, and then ultimately to **universal decolonization for primary prevention**.

The targeted CHG decolonization trials for primary prevention of infection focused on *S. aureus* carriers identified through rapid screening of inpatients, most of whom were undergoing surgery. Those harboring *S. aureus* were decolonized with CHG bathing and nasal mupirocin. Bode *et al.* compared decolonization to placebo among 917 *S. aureus* carriers identified by admission screening of predominantly surgical patients. Those receiving decolonization had significantly fewer inpatient *S. aureus* infections, especially deep surgical site infections [47]. These findings suggest that CHG and mupirocin confer benefit when given both pre- and post-operatively, because the surgery could occur anytime during the 5 day decolonization regimen. In contrast, Harbarth *et al.* compared the value of admission MRSA screening to no admission screening in over 10,000 surgical patients in a hospital where decolonization of MRSA carriers was routine. The additional identification of MRSA carriers from screening did not reduce total hospital-associated MRSA infection [48].

In 2013, several universal decolonization trials for primary prevention of MDROs and infection in ICUs were published and expanded the evidence for HAI and MDRO reduction in patient populations at high risk for infection [49–51]. This led to widespread adoption of daily CHG bathing, with and without nasal decolonization, in the US and UK [58–61]. Decolonization trials extending outside ICUs then followed. The ABATE Infection Trial found that universal decolonization in non-critical care units reduced MDROs and bloodstream infections only in the subset of patients with medical devices [53]. This raised natural questions about the targeted role of decolonization for protection of medical devices throughout the continuum of care. Furthermore, targeting MRSA carriers with repeated rounds of CHG and mupirocin decolonization in the CLEAR Trial reduced MRSA infections and hospitalizations following hospital discharge [54]. Finally, recent large scale trials in nursing homes have shown that universal decolonization, but not targeted decolonization, can significantly reduce MDRO prevalence [55–57].

In a brief departure from large-scale randomized controlled trials, the US CDC has been investing in regional prevention of MDROs through decolonization. The SHIELD Orange County Project was a 38 healthcare facility project in Orange County, California that involved CHG bathing and nasal iodophor for universal decolonization in nursing homes and long-term acute care facilities, and targeted decolonization of patients in contact precautions in hospitals [38,62,63]. All 17 participating hospitals were already routinely performing universal ICU decolonization. Across the 25-month SHIELD intervention, MDRO prevalence declined by 24% in long-term care facilities and by 14% among hospitalized patients in contact precautions [62].

What Products Should be Used?

When used for bathing, chlorhexidine concentrations of 2% and 4% are most commonly used. The 4% formulation is generally applied and rinsed off in the shower while 2% CHG is used as a leave-on product for bed bathing. As an applicator, a mesh sponge enables CHG to lather well for the shower since lathering is difficult through hand rubbing alone. Furthermore, non-cotton applicators are important since cotton binds CHG and limits its release to the skin [43].

The 2% leave-on product is favored because it results in higher residual concentrations of CHG on the skin, which then provide germicidal activity for up to 24 hours [43]. The 4% formulation is too drying to be used as a leave-on product, but the 2% concentration is well-tolerated. Safety has been well demonstrated with over a million baths being conducted during clinical trials, with a <1%–2% risk of mild skin reactions that resolve rapidly upon discontinuation [49–51,53,54]. Anaphylaxis is rare, but has been reported in case reports.

Nasal decolonization in combination with CHG has usually involved 2% mupirocin ointment. However, reports of mupirocin resistance in some geographic areas has led to the recent evaluation of 10% povidone-iodine in some clinical trials [52,56]. The Mupirocin-Iodophor Swap Out Trial will directly evaluate the non-inferiority of universal ICU decolonization with CHG-iodophor compared to CHG-mupirocin for the outcomes of ICU-attributable *S. aureus* clinical cultures and all-cause bacteraemia. In the meantime, nasal iodophor has been shown to be effective in reducing MRSA carriage when universally used with

Table 1Large-scale randomized clinical trials evaluating CHG decolonization to reduce infection and MDRO^a

Trial and Target Population	N	Intervention	Impact of Decolonization
Pre-Operative Use			
Bode et al [47]	918	Universal inpatient screening for <i>S. aureus</i> . Carriers randomized to CHG and mupirocin vs routine care	Among <i>S. aureus</i> carriers, 58% less inpatient <i>S. aureus</i> infection, including 79% less deep surgical site infection
Harbarth et al [48]	10,844	Universal inpatient screening for MRSA. Carriers randomized to CHG and mupirocin vs routine care	No difference in overall hospital-associated MRSA infection
Intensive Care Units			
Climo et al [49] 6 Academic medical centers	7727	Universal CHG bathing vs routine care	23% less MRSA/VRE ICU acquisition (as treated)
REDUCE MRSA Trial [50] 43 Community hospitals	74,256	A. Targeted CHG and mupirocin for MRSA carriers B. Universal CHG and mupirocin C. Routine care	37% less MRSA ICU clinical cultures 44% less all-cause ICU bloodstream infection
Pediatric SCRUB Trial [51] 5 Academic medical centers	4947	Universal CHG bathing vs routine care	36% less ICU bloodstream infection (as treated)
Mupirocin Iodophor Swap Out [52] 137 Community hospitals	~250,000	A. Universal CHG and mupirocin B. Universal CHG and iodophor	Results pending
Non-Intensive Care Units			
ABATE Infection Trial [53] 53 Community hospitals	339,902	Universal CHG bathing plus targeted mupirocin for MRSA carriers vs routine care	No difference in MRSA/VRE clinical cultures or bloodstream infection in overall non-ICU population In subset with medical devices: 37% less MRSA/VRE clinical cultures 32% less bloodstream infection (post-hoc analysis)
Post-Discharge			
CLEAR Trial [54]	2121	Targeted education plus 5 day course of CHG bathing, CHG mouthwash, and mupirocin repeated twice a month for six months vs education alone for MRSA carriers	In the year following discharge: 30% less MRSA infection 17% less all-cause infection
Nursing Homes			
Bellini et al [55]	4750	Universal screening for MRSA followed by targeted CHG bathing, CHG mouthwash, nasal mupirocin, and room disinfection for MRSA carriers vs routine care	No difference in one-day MRSA point prevalence
Protect Trial [56,57] 28 Nursing homes	~18,000	Universal CHG bathing plus nasal iodophor vs routine care	29% reduction in MDRO carriage 24% reduction in MRSA carriage 61% reduction in VRE carriage 52% reduction in ESBL carriage Primary trial results on infection and hospitalization: pending

^a MDRO: multidrug-resistant organism; MRSA: methicillin-resistant *Staphylococcus aureus*; VRE: vancomycin-resistant enterococcus; ESBL: extended spectrum beta-lactamase producers.

CHG bathing in nursing homes [57,62]. In both the Swap Out Trial and these other decolonization studies, nasal iodophor was given twice daily for five days, similar to mupirocin, because of evidence that a single dose is only suppressive and daily dosing is inferior to twice daily dosing [64,65].

Where should CHG be used? In what healthcare settings?

To date, clinical trials and other studies have repeatedly demonstrated the value of CHG bathing for infection

prevention in healthcare settings where patients are vulnerable to infection. During hospitalization, this includes patients requiring intensive care and those with medical devices, both inside and outside of intensive care [49–51,53]. This benefit in the most vulnerable populations fuels hope for benefit in oncology and bone marrow transplant units, where data are yet sparse, but accruing [49,66]. Inpatient benefit further extends to surgical patients during the immediate peri-operative period, both before and after surgery [47].

Following discharge, CHG, in combination with nasal decolonization, has been shown to mitigate the high risk of

infection and rehospitalization in MRSA carriers [54]. Similarly, evidence is growing for its benefit in reducing MDRO carriage and infection in long-term care settings [57,67]. This includes both long-term acute care hospitals and nursing homes where MDRO prevalence can be many-fold higher than in hospitals and patients have longstanding health issues that compound acute ones.

In outpatients, use should be commensurate to the vulnerability of infection. As an antiseptic, CHG has been used to mitigate the risk of infection due to chronic dermatologic conditions, as well as for secondary prevention in patients with recurrent *S. aureus* disease. However, vulnerability need not be for oneself, as CHG has been commonly and successfully used to decolonize healthcare workers who harbor MRSA in the setting of a healthcare-associated outbreak.

When should decolonization be used?

In healthcare settings, it is important to perform CHG bathing upon admission. Admitted patients feel unwell and may not have bathed for days. In addition, admission is a key moment where MDROs can be imported into a unit. Cleansing the skin is important for all these reasons and to reduce skin bioburden before surgery or placement of devices occur.

As to frequency of use, daily CHG bathing has been the most well studied, likely because daily bathing is routine in US ICUs

where trials were performed, but also because CHG residue and reduced skin bioburden can last for up to 24 hours [27,43–45]. Thus, bathing daily could protect patients for the full duration of their hospital stay. Nevertheless, some evidence for MDRO and infection reduction with every other day bathing has emerged [64,68]. In addition, single or repeated use of a 5-day decolonization regimen in outpatients has been found to be successful [54,69].

Other considerations for use of decolonization include situations when treatment options are limited for a colonizing highly antibiotic resistant bacteria or a colonized patient is allergic to an extensive array of antibiotics. In these circumstances, body decolonization as an infection prevention strategy can be life-saving, especially during periods of high vulnerability (e.g. hospitalization, operation, open wound, medical devices).

How should decolonization with CHG be performed?

Proper application of CHG is essential [70,71]. While, nurses and nursing assistants have personal experience bathing their own intact skin, it is unreasonable to assume that they would have inherent knowledge on how best to clean breaks in the skin, including abrasions, rashes, wounds, surgical incisions, and medical devices. In fact, the inherent response to these

Table II
Top 10 pearls for appropriate chlorhexidine (CHG) bathing

Key Training Point	Details
1. Application and training matters	Nurses and nursing assistants are not inherently familiar with how best to clean non-intact skin, wounds and devices. Need directed training and need annual refresher training [70,71].
2. Not a topcoat	CHG is not to be applied after a bath; it is the bath itself. Apply with thorough massage to remove dirt, grime, and germs, and to allow CHG to bind to skin proteins and continue to kill germs for up to 24 hours.
3. Commonly missed areas	The neck is germ-ridden like the groin, but not as well cleaned. Other missed areas: back of knee, between fingers and toes [27].
4. Leave-on better than rinse-off	When possible, air dry rather than rinse off to retain germ-killing concentrations of CHG on the skin [43]. CHG only works after it dries (e.g. candidal rashes will improve with CHG application and drying, but if left moist in folds, CHG will not kill germs and moisture can worsen candidal rashes.)
5. Avoid cotton, use mesh sponge	Cotton binds CHG and limits its release to the skin [43]. For bed baths, use non-cotton cloths. For showering, a mesh sponge allows CHG to lather well and applies CHG well to skin.
6. Check compatibility	Contact manufacturers to ensure skin lotions and care products do not inactivate CHG.
7. Clean wounds and devices	Breaks in skin are entry points for germs to invade and cause infection. Clean wounds well unless large or deep, e.g. requires packing. Use a clean CHG cloth to clean the proximal 6 inches of all devices, including lines, tubes, and drains closest to the body, as well as over dressings. For patients who shower, devices are wrapped for waterproofing. After the shower, devices should be cleaned with a clean CHG cloth.
8. Clean the perineum	CHG is safe on vaginal epithelium. Thorough cleaning of the perineum was emphasized in several CHG clinical trials [50,52–54,56]. Cleaning with CHG reduces low-level bacteruria and funguria [74].
9. Clean the face	Clean face well due to common contamination from adjacent nose and mouth. Nose is the major reservoir for <i>S. aureus</i> . Avoid eyes and ear canal since CHG should not directly contact nerves (e.g. eyes, auditory nerves if tympanic membrane ruptured). Cleaning the face with CHG is emphasized in several clinical trials [52–54,56].
10. The nose matters for <i>Staphylococcus aureus</i>	Nasal decolonization is the workhorse for clearing <i>S. aureus</i> . CHG prevents spread, but does not clear carriage.

conditions is to avoid bathing those areas due to fear of causing pain. Nevertheless, those areas are portals of entry for infection that should be well cleaned to prevent infection. In fact, daily bathing coupled with the 24-hour germicidal benefit of CHG is most pertinent to those high risk skin areas to provide continued protection.

Several key training points in response to common errors are found in Table II. Of particular note is the recommendation to use CHG to clean the face, perineum, and all lines, tubes, drains, and other devices for at least 6 inches (15 centimeters) closest to the body. This was standard protocol and safely done for over one million baths in our collective trials. Our detailed protocols, videos, and educational materials are publicly available at several websites [63,72,73].

In addition to training staff, pre-launch activities should include ensuring that other topical products, such as lotions and barrier creams, are compatible with CHG. This is best done by contacting the manufacturer and exchanging incompatible or unknown products with those that will not inactivate CHG. In addition, it is important to note that CHG and bleach chemically interact in the laundry and produce brown stains. Thus, staff should avoid placing CHG saturated cloths directly onto sheets. Fortunately, once CHG is bound to the skin, it will not rub off onto sheets. While hospital laundry is often washed at a sufficiently high temperature to cause CHG to denature, on-site laundry temperatures in nursing homes or care homes is generally not able to prevent brown staining, and a switch from chlorine bleach to peroxide bleach is highly advisable if CHG is used in those settings. Finally, it is advisable to perform a skin check of patients prior to the launch of CHG bathing to avoid misattributing pre-existing skin issues to CHG when staff are unfamiliar with the product.

The adoption of CHG bathing should be considered a major campaign due to the importance of training, validation, and feedback. Training and re-training should be an annual competency for all staff performing bathing due to the importance of proper application. Assessing adherence is critical for success. Feedback about whether CHG bathing was performed and the quality of bathing enables correction and success. Simple skills assessment forms can be found online [75].

The spectre of resistance and future considerations

In an era where case reports of resistance occur shortly after each new antibiotic arrives on the market, it would be foolish to assume invincibility of any systemic or topical germicidal product. Nevertheless, there has been a hope that antiseptics would stave off resistance longer due to their small size and rapid bacteriocidal activity.

The natural diversity of CHG minimum inhibitory concentrations (MIC) (8 µg/mL for *S. aureus* and 32–>300 µg/mL for Gram-negative bacteria) across wild-type bacteria raises the question about inherent mechanisms of resistance. In addition, efflux pumps have been identified that can expel CHG from bacteria. Clinically, two things have been noted. On one hand, elevations in CHG MIC have been reported to emerge while universal CHG bathing is being employed [76,77]. On the other hand, randomized clinical trials have not identified differential emergence of resistance associated with the decolonization group [49,50,54,78]. For MRSA, it may be that the combination

of both CHG bathing and nasal decolonization reduces the emergence of resistance compared to CHG alone.

What is known is that applying 2% or 4% CHG products confers 20,000 µg/mL and 40,000 µg/mL of CHG to the skin, which is in far excess of bacterial MICs. Thus, proper application may be the key to not only achieving benefit, but also preventing resistance, especially if residual skin concentrations exceeding 500 µg/mL are maintained. Continued monitoring is clearly needed as the evidence-base accrues for the benefit of CHG bathing.

In the end, the spectre of resistance should not outweigh the value of CHG protocols in reducing MDRO transmission and infection, device-associated infections, and all-cause bacteremia. CHG bathing remains an astoundingly simple solution that has achieved some of the largest proven gains in modern infection prevention. It should be applied as best practice to protect patients.

Of course, we should not become complacent. Science enables us to innovate and strive for improvement. If there are better alternatives or more effective strategies, we should press onward to find them. If resistance emerges, then necessity should drive the next invention. We must not be wedded to the best things of today, but always seek a future that will find us something more effective, lower in cost, and better able to protect humans from the persistent threat of infection.

Acknowledgements

SSH receives funding from the U.S. government, including the Centers for Disease Control and Prevention, the Agency for Healthcare Research and Quality, and the National Institutes of Health, for research to prevent and address healthcare-associated infections. SSH also conducts studies and trials in which participating hospitals and nursing homes have received contributed antiseptic products from Stryker (Sage Products), Molnlycke, 3M, Xtrium Laboratories, and Medline.

Conflict of interest statement

None declared.

Funding sources

No funds were used for this manuscript.

References

- [1] Manor J, Blum N, Lurie Y. "No good deed goes unpunished": Ignaz Semmelweis and the story of puerperal fever. *Infect Control Hosp Epidemiol* 2016;37:881–7.
- [2] Berríos-Torres SI, Umscheid CA, Bratzler DW, Leas B, Stone EC, Kelz RR, et al., Healthcare Infection Control Practices Advisory Committee. Centers for Disease Control and Prevention Guideline for the Prevention of Surgical Site Infection, 2017. *JAMA Surg* 2017;152:784–91.
- [3] Global Guidelines for the Prevention of Surgical Site Infection World Health Organization. WHO guidelines approved by the guidelines review committee, ISBN 978-92-4-155047-5. <https://www.who.int/infection-prevention/publications/ssi-prevention-guidelines/en/>.
- [4] Allegranzi B, Sax H, Pittet D. Hand hygiene and healthcare system change within multi-modal promotion: a narrative review. *J Hosp Infect* 2013;83(Suppl. 1):S3–10.
- [5] Platt R, Polk BF, Murdock B, Rosner B. Reduction of mortality associated with nosocomial urinary tract infection. *Lancet* 1983;1:893–7.

- [6] Davies GE, Francis J, Martin AR, Rose FL, Swain G. 1:6-di-4'-chlorophenyldiguanidohexane ("hibitane"). laboratory investigation of a new antibacterial agent of high potency. *Br J Pharmacol Chemother* 1954;9:192–6.
- [7] Lowbury EJ. Chlorhexidine. *The Practitioner* 1957;179:489–93.
- [8] Lilly HA, Lowbury EJ. Disinfection of the skin: an assessment of some new preparations. *Br Med J* 1971;3:674–6.
- [9] Lowbury EJ, Lilly HA. Use of 4 per cent chlorhexidine detergent solution (Hibiscrub) and other methods of skin disinfection. *Br Med J* 1973;1:510–5.
- [10] Ayliffe GA, Bridges K, Lilly HA, Lowbury EJ, Varney J, Wilkins MD. Comparison of two methods for assessing the removal of total organisms and pathogens from the skin. *J Hyg (Lond)* 1975;75:259–74.
- [11] Lowbury EJ, Ayliffe GA. Proceedings: Alcoholic solutions and other agents for disinfection of the surgeon's hands. *J Clin Pathol* 1975;28:753–4.
- [12] Lilly HA, Lowbury EJ. Transient skin flora: their removal by cleansing or disinfection in relation to their mode of deposition. *J Clin Pathol* 1978;31:919–22.
- [13] Lilly HA, Lowbury EJ, Wilkins MD. Limits to progressive reduction of resident skin bacteria by disinfection. *J Clin Pathol* 1979;32:382–5.
- [14] Lilly HA, Lowbury EJ, Wilkins MD. Detergents compared with each other and with antiseptics as skin 'degerming' agents. *J Hyg (Lond)* 1979;82:89–93.
- [15] Ayliffe GA, Noy MF, Babb JR, Davies JG, Jackson J. A comparison of pre-operative bathing with chlorhexidine-detergent and non-medicated soap in the prevention of wound infection. *J Hosp Infect* 1983;4:237–44.
- [16] Davies J, Babb JR, Ayliffe GA. The effect on the skin flora of bathing with antiseptic solutions. *J Antimicrob Chemother* 1977;3:473–81.
- [17] Gjermo P. Chlorhexidine in dental practice. *J Clin Periodontol* 1974;1:143–52.
- [18] Maki DG, Ringer M, Alvarado CJ. Prospective randomised trial of povidone-iodine, alcohol, and chlorhexidine for prevention of infection associated with central venous and arterial catheters. *Lancet* 1991;338:339–43.
- [19] Mimoz O, Pieroni L, Lawrence C, Edouard A, Costa Y, Samii K, et al. Prospective, randomized trial of two antiseptic solutions for prevention of central venous or arterial catheter colonization and infection in intensive care unit patients. *Crit Care Med* 1996;24:1818–23.
- [20] Mimoz O, Villeminey S, Ragot S, Dahyot-Fizelie C, Laksiri L, Petitpas F, et al. Chlorhexidine-based antiseptic solution vs alcohol-based povidone-iodine for central venous catheter care. *Arch Intern Med* 2007;167:2066–72.
- [21] Mimoz O, Lucet JC, Kerforne T, Pascal J, Souweine B, Goudet V, et al., CLEAN trial investigators. Skin antisepsis with chlorhexidine-alcohol versus povidone iodine-alcohol, with and without skin scrubbing, for prevention of intravascular-catheter-related infection (CLEAN): an open-label, multicentre, randomised, controlled, two-by-two factorial trial. *Lancet* 2015;386:2069–77.
- [22] Darouiche RO, Wall Jr MJ, Itani KM, Otterson MF, Webb AL, Carrick MM, et al. Chlorhexidine-alcohol versus povidone-iodine for surgical-site antisepsis. *N Engl J Med* 2010;362:18–26.
- [23] Mangram AJ, Horan TC, Pearson ML, Silver LC, Jarvis WR. Guideline for Prevention of Surgical Site Infection, 1999. Centers for Disease Control and Prevention (CDC) Hospital Infection Control Practices Advisory Committee. *Infect Control Hosp Epidemiol* 1999;20:250–78.
- [24] Vernon MO, Hayden MK, Trick WE, Hayes RA, Blom DW, Weinstein RA. Chicago Antimicrobial Resistance Project (CARP). Chlorhexidine gluconate to cleanse patients in a medical intensive care unit: the effectiveness of source control to reduce the bioburden of vancomycin-resistant enterococci. *Arch Intern Med* 2006;166:306–12.
- [25] Bleasdale SC, Trick WE, Gonzalez IM, Lyles RD, Hayden MK, Weinstein RA. Effectiveness of chlorhexidine bathing to reduce catheter-associated bloodstream infections in medical intensive care unit patients. *Arch Intern Med* 2007;167:2073–9.
- [26] Popovich KJ, Hota B, Hayes R, Weinstein RA, Hayden MK. Effectiveness of routine patient cleansing with chlorhexidine gluconate for infection prevention in the medical intensive care unit. *Infect Control Hosp Epidemiol* 2009;30:959–63.
- [27] Popovich KJ, Lyles R, Hayes R, Hota B, Trick W, Weinstein RA, et al. Relationship between chlorhexidine gluconate skin concentration and microbial density on the skin of critically ill patients bathed daily with chlorhexidine gluconate. *Infect Control Hosp Epidemiol* 2012;33:889–96.
- [28] Mody L, Washer LL, Kaye KS, Gibson K, Saint S, Reyes K, et al. Multidrug-resistant Organisms in Hospitals: What Is on Patient Hands and in Their Rooms? *Clin Infect Dis* 2019 Apr 13 [Epub ahead of print].
- [29] Huang SS, Rifas-Shiman SL, Warren DK, Fraser VJ, Climo MW, Wong ES, et al. Centers for Disease Control and Prevention Epicenters Program. Improving methicillin-resistant *Staphylococcus aureus* surveillance and reporting in intensive care units. *J Infect Dis* 2007;195:330–8.
- [30] Huang SS, Rifas-Shiman SL, Pottinger JM, Herwaldt LA, Zembower TR, Noskin GA, et al., Centers for Disease Control and Prevention Epicenters Program. Improving the assessment of vancomycin-resistant enterococci by routine screening. *J Infect Dis* 2007;195:339–46 [Epub 2006 Dec 27].
- [31] Boldt AC, Schwab F, Rohde AM, Kola A, Bui MT, Märtin N, et al. Admission prevalence of colonization with third-generation cephalosporin-resistant Enterobacteriaceae and subsequent infection rates in a German university hospital. *PLoS One* 2018;13:e0201548.
- [32] Hamprecht A, Rohde AM, Behnke M, Feihl S, Gastmeier P, Gebhardt F, et al., DZIF-ATHOS Study Group. Colonization with third-generation cephalosporin-resistant Enterobacteriaceae on hospital admission: prevalence and risk factors. *J Antimicrob Chemother* 2016;71:2957–63.
- [33] Latour K, Huang TD, Jans B, Berhin C, Bogaerts P, Noel A, et al. Prevalence of multidrug-resistant organisms in nursing homes in Belgium in 2015. *PLoS One* 2019;14:e0214327.
- [34] Manzur A, De Gopegui ER, Dominguez M, Mariscal D, Gavalda L, Perez JL, et al., Spanish Network for Research in Infectious Diseases. Clinical significance of methicillin-resistant *Staphylococcus aureus* colonization in residents in community long-term-care facilities in Spain. *Epidemiol Infect* 2012;140:400–6.
- [35] Hogardt M, Proba P, Mischler D, Cuny C, Kempf VA, Heudorf U. Current prevalence of multidrug-resistant organisms in long-term care facilities in the Rhine-Main district, Germany, 2013. *Euro Surveill* 2015;20. pii: 21171.
- [36] Horner C, Parnell P, Hall D, Kearns A, Heritage J, Wilcox M. Methicillin-resistant *Staphylococcus aureus* in elderly residents of care homes: colonization rates and molecular epidemiology. *J Hosp Infect* 2013;83:212–8.
- [37] Chen H, Au KM, Hsu KE, Lai CK, Myint J, Mak YF, et al. Multidrug-resistant organism carriage among residents from residential care homes for the elderly in Hong Kong: a prevalence survey with stratified cluster sampling. *Hong Kong Med J* 2018;24:350–60.
- [38] McKinnell JA, Singh RD, Miller LG, Kleinman K, Gussin G, He J, et al. CDC Safety and Healthcare Epidemiology Prevention Research Development (SHEPherD) Program. The SHIELD Orange County Project -Multi Drug-Resistant Organism (MDRO) Prevalence in 21 Nursing Homes and Long Term Acute Care Facilities in Southern California. *Clin Infect Dis* 2019 Feb 11 [Epub ahead of print].
- [39] Mody L, Foxman B, Bradley S, McNamara S, Lansing B, Gibson K, et al. Longitudinal assessment of multidrug-resistant organisms in newly admitted nursing facility patients: implications for an evolving population. *Clin Infect Dis* 2018;67:837–44.

- [40] McKinnell JA, Miller LG, Singh R, Kleinman K, Peterson EM, Evans KD, et al. Prevalence of and factors associated with Multidrug Resistant Organism (MDRO) colonization in 3 nursing homes. *Infect Control Hosp Epidemiol* 2016;37:1485–8.
- [41] Trick WE, Weinstein RA, DeMarais PL, Kuehnert MJ, Tomaska W, Nathan C, et al. Colonization of skilled-care facility residents with antimicrobial-resistant pathogens. *J Am Geriatr Soc* 2001;49:270–6.
- [42] March A, Aschbacher R, Dhanji H, Livermore DM, Böttcher A, Slegel F, et al. Colonization of residents and staff of a long-term-care facility and adjacent acute-care hospital geriatric unit by multiresistant bacteria. *Clin Microbiol Infect* 2010;16:934–44.
- [43] Rhee Y, Palmer LJ, Okamoto K, Gemunden S, Hammouda K, Kemble SK, et al., Centers for Disease Control And Prevention Epicenter Program. Differential effects of chlorhexidine skin cleansing methods on residual chlorhexidine skin concentrations and bacterial recovery. *Infect Control Hosp Epidemiol* 2018;39:405–11.
- [44] Lin MY, Lolans K, Blom DW, Lyles RD, Weiner S, Poluru KB, et al., Centers for Disease Control and Prevention Epicenter Program. The effectiveness of routine daily chlorhexidine gluconate bathing in reducing *Klebsiella pneumoniae* carbapenemase-producing Enterobacteriaceae skin burden among long-term acute care hospital patients. *Infect Control Hosp Epidemiol* 2014;35:440–2.
- [45] Edmiston Jr CE, Lee CJ, Krepel CJ, Spencer M, Leaper D, Brown KR, et al. Evidence for a standardized preadmission showering regimen to achieve maximal antiseptic skin surface concentrations of chlorhexidine gluconate, 4%, in surgical patients. *JAMA Surg* 2015;150:1027–33.
- [46] Liu C, Bayer A, Cosgrove SE, Daum RS, Fridkin SK, Gorwitz RJ, et al., Infectious Diseases Society of America. Clinical practice guidelines by the Infectious Diseases Society of America for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children. *Clin Infect Dis* 2011;52:e18–55.
- [47] Bode LG, Kluytmans JA, Wertheim HF, Bogaers D, Vandembroucke-Grauls CM, Roosendaal R, et al. Preventing surgical-site infections in nasal carriers of *Staphylococcus aureus*. *N Engl J Med* 2010;362:9–17.
- [48] Harbarth S, Fankhauser C, Schrenzel J, Christenson J, Gervaz P, Bandiera-Clerc C, et al. Universal screening for methicillin-resistant *Staphylococcus aureus* at hospital admission and nosocomial infection in surgical patients. *JAMA* 2008;299:1149–57.
- [49] Climo MW, Yokoe DS, Warren DK, Perl TM, Bolon M, Herwaldt LA, et al. Effect of daily chlorhexidine bathing on hospital-acquired infection. *N Engl J Med* 2013;368:533–42.
- [50] Huang SS, Septimus E, Kleinman K, Moody J, Hickok J, Avery TR, et al., CDC Prevention Epicenters Program. AHRQ DECIDE Network and Healthcare-Associated Infections Program. Targeted versus universal decolonization to prevent ICU infection. *N Engl J Med* 2013;368:2255–65.
- [51] Milstone AM, Elward A, Song X, Zerr DM, Orscheln R, Speck K, et al. Pediatric SCRUB Trial Study Group. Daily chlorhexidine bathing to reduce bacteraemia in critically ill children: a multi-centre, cluster-randomised, crossover trial. *Lancet* 2013;381:1099–106.
- [52] Mupirocin-Iodophor ICU Decolonization Swap Out Trial. Clinicaltrials.gov <https://clinicaltrials.gov/ct2/show/NCT03140423> [last accessed September 8, 2019].
- [53] Huang SS, Septimus E, Kleinman K, Moody J, Hickok J, Heim L, et al., ABATE Infection trial team. Chlorhexidine versus routine bathing to prevent multidrug-resistant organisms and all-cause bloodstream infections in general medical and surgical units (ABATE Infection trial): a cluster-randomised trial. *Lancet* 2019;393:1205–15.
- [54] Huang SS, Singh R, McKinnell JA, Park S, Gombosev A, Eells SJ, et al., Project CLEAR Trial. Decolonization to Reduce Post-discharge Infection Risk among MRSA Carriers. *N Engl J Med* 2019;380:638–50.
- [55] Bellini C, Petignat C, Masserey E, Büla C, Burnand B, Rousson V, et al. Universal screening and decolonization for control of MRSA in nursing homes: a cluster randomized controlled study. *Infect Control Hosp Epidemiol* 2015;36:401–8.
- [56] Protect Trial: Protecting nursing homes from Infections and Hospitalization. Clinicaltrials.gov <https://clinicaltrials.gov/ct2/show/NCT03118232> [last accessed September 8, 2019].
- [57] Miller LG, McKinnell JA, Singh R, Gussin G, Kleinman K, Saavedra R, et al. Universal Decolonization in Nursing Homes: Effect of Chlorhexidine and Nasal Povidone-Iodine on Prevalence of Multi-Drug-Resistant Organisms (MDROs) in the PROTECT Trial. Abstract 680256. IDWeek (Joint Meeting of IDSA, SHEA, HIVMA, and PIDS), October 2–6, 2019 (Washington DC).
- [58] Edgeworth JD. Has decolonization played a central role in the decline in UK methicillin-resistant *Staphylococcus aureus* transmission? A focus on evidence from intensive care. *J Antimicrob Chemother* 2011;66(Suppl. 2):ii41–7.
- [59] Weiner LM, Webb AK, Walters MS, Dudeck MA, Kallen AJ. Policies for Controlling Multidrug-Resistant Organisms in US Healthcare Facilities Reporting to the National Healthcare Safety Network, 2014. *Infect Control Hosp Epidemiol* 2016;37:1105–8.
- [60] Shuman EK, Harpe JM, Calfee DP. Survey of Hospital Practices Regarding Use of Chlorhexidine Gluconate Bathing for Prevention of Healthcare-Associated Infections. IDWeek Abstract; <https://www.shea-online.org/images/docs/Shuman-IDWeekAbstract.pdf> [last accessed September 8, 2019].
- [61] Septimus E, Hickok J, Moody J, Kleinman K, Avery TR, Huang SS, et al. Closing the Translation Gap: toolkit based implementation of universal decolonization in adult intensive care units reduces central line associated bloodstream infections in 95 community hospitals. *Clin Infect Dis* 2016 May;63:172–7. PMID: 27143669.
- [62] McKinnell JA, Singh RD, Miller LG, Gussin G, Kleinman K, Saavedra R, et al. CDC Safety and Healthcare Epidemiology Prevention Research Development (SHEPHERD) Program. The SHIELD Orange County Project: A Decolonization Strategy in 35 Hospitals and Nursing Homes Reduces Multi-Drug Resistant Organism (MDRO) Prevalence in a Southern California Region. IDWeek (Joint Meeting of IDSA, SHEA, HIVMA, and PIDS), October 2-6, 2019 (Washington DC).
- [63] SHIELD: Eliminating Multidrug-Resistant Organisms. University of California Irvine Health. Hospital, Nursing Home, and Long-Term Acute Care Hospital Toolkits for Decolonization. <https://www.ucihealth.org/shield> and <https://www.ucihealth.org/shield/hospital-decolonization-toolkit> [last accessed September 8, 2019].
- [64] Miller LG, McKinnell JA, Singh R, Kleinman K, Gombosev A, Dutciuc T, et al. Reduction of MDRO Colonization in Nursing Home Residents with Routine Use of Chlorhexidine Bathing and Nasal Iodophor (Project PROTECT). IDWeek (5th Annual Joint Meeting of IDSA, SHEA, HIVMA, and PIDS), October 26–30, 2016 (New Orleans, LA).
- [65] Heim L, Dutciuc T, Singh R, Estevez M, Tjoa T, Chang J, et al. Double Swab 5% vs Single Swab 10% Iodophor for Reducing MRSA with Routine Chlorhexidine Bathing (Abstract 8775). Society for Healthcare Epidemiology of America (SHEA) Annual Spring Meeting, March 29-31, 2017, (St. Louis, MO).
- [66] Raulji CM, Clay K, Velasco C, Yu LC. Daily bathing with chlorhexidine and its effects on nosocomial infection rates in pediatric oncology patients. *Pediatr Hematol Oncol* 2015;32:315–21.
- [67] Hayden MK, Lin MY, Lolans K, Weiner S, Blom D, Moore NM, et al., Centers for Disease Control and Prevention Epicenters Program. Prevention of colonization and infection by *Klebsiella pneumoniae* carbapenemase-producing enterobacteriaceae in long-term acute-care hospitals. *Clin Infect Dis* 2015;60:1153–61.
- [68] Swan JT, Ashton CM, Bui LN, Pham VP, Shirkey BA, Blackshear JE, et al. Effect of chlorhexidine bathing every other day on prevention of hospital-acquired infections in the surgical icu: a single-center, randomized controlled trial. *Crit Care Med* 2016;44:1822–32.

- [69] Ammerlaan HS, Kluytmans JA, Berkhout H, Buiting A, de Brauwier EI, van den Broek PJ, et al., MRSA Eradication Study Group. Eradication of carriage with methicillin-resistant *Staphylococcus aureus*: effectiveness of a national guideline. *J Antimicrob Chemother* 2011 Oct;66:2409–17.
- [70] Popovich KJ, Hota B, Hayes R, Weinstein RA, Hayden MK. Daily skin cleansing with chlorhexidine did not reduce the rate of central-line associated bloodstream infection in a surgical intensive care unit. *Intensive Care Med* 2010;36:854–8.
- [71] Supple L, Kumaraswami M, Kundrapu S, Sunkesula V, Cadnum JL, Nerandzic MM, et al. Chlorhexidine only works if applied correctly: use of a simple colorimetric assay to provide monitoring and feedback on effectiveness of chlorhexidine application. *Infect Control Hosp Epidemiol* 2015;36:1095–7.
- [72] Universal ICU Decolonization: an enhanced protocol. Agency for Healthcare Research and Quality. <https://www.ahrq.gov/hai/universal-icu-decolonization/index.html> [last accessed September 8, 2019].
- [73] Active Bathing to Eliminate (ABATE) Infection. NIH Collaboratory Living Textbook. <https://rethinkingclinicaltrials.org/demonstration-projects/uh3-project-active-bathing-to-eliminate-abate-infection/> [last accessed September 8, 2019].
- [74] Huang SS, Septimus E, Hayden MK, Kleinman K, Sturtevant J, Avery TR, et al. Impact of body surface decolonization on bacteriuria and candiduria in a cluster-randomized trial of intensive care units. *Lancet Infect Dis* 2016;16:70–9.
- [75] Staff Skills Assessment: CHG Cloth Observation Checklist. SHIELD: Eliminating Multidrug-Resistant Organisms. University of California Irvine Health. Hospital Toolkit for decolonization. <https://www.ucihealth.org/-/media/files/pdf/shield/hospital/hospital-step-6-chg-cloth-skills-assessment-checklist-pdf.pdf> [last accessed September 8, 2019].
- [76] Batra R, Cooper BS, Whiteley C, Patel AK, Wyncoll D, Edgeworth JD. Efficacy and limitation of a chlorhexidine-based decolonization strategy in preventing transmission of methicillin-resistant *Staphylococcus aureus* in an intensive care unit. *Clin Infect Dis* 2010;50:210–7.
- [77] Chávez-Moreno S, Camacho-Ortiz A. Chlorhexidine bathing every other day still does the trick, but it may come at a cost. *Ann Transl Med* 2016;4:555.
- [78] Hayden MK, Lolans K, Haffenreffer K, Avery TR, Kleinman K, Li H, et al. Chlorhexidine and Mupirocin Susceptibility of Methicillin-Resistant *Staphylococcus aureus* Isolates in the REDUCE-MRSA Trial. *J Clin Microbiol* 2016;54:2735–42.