



Short report

Combining detergent/disinfectant with microfibre material provides a better control of microbial contaminants on surfaces than the use of water alone

A. Robertson, M. Barrell, J-Y. Maillard*

School of Pharmacy and Pharmaceutical Sciences, Cardiff University, Cardiff, UK

ARTICLE INFO

Article history:

Received 2 April 2019

Accepted 14 May 2019

Available online 18 May 2019

Keywords:

Microfibre

Detergent

Disinfection

Staphylococcus aureus

Acinetobacter baumannii

Clostridium difficile spores

SUMMARY

The use of microfibre cloths with either water, detergent or disinfectant is currently recommended for hospital cleaning. This study explored the efficacy of a microfibre cloth with either water or detergent/disinfectant or sporicidal products using the ASTM2967-15 standard against *Staphylococcus aureus*, *Acinetobacter baumannii* and *Clostridium difficile* spores. The use of detergent/disinfectant or sporicidal products had a significantly (analysis of variance (ANOVA), $P < 0.001$) better activity than water alone in reducing bacteria and spores' viability, and in reducing the transfer microorganisms between surfaces. The use of water alone with a microfibre cloth is less effective and should not replace the use of biocidal products.

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Introduction

Efficient cleaning and disinfection are an integral part of infection-control regimens currently used in healthcare facilities [1] and can result in a reduction in healthcare-associated infections (HCAIs), [2] and have an impact of infection outbreaks [3]. Such reduction in HCAIs will provide, in turn, a significant financial benefit to healthcare systems [4]. For surface decontamination, formulated products or water are used in combination with various materials [5] although the use

of formulated wipes might be more efficacious [6]. The type of materials used will impact on the concentration of formulation (notably quaternary ammonium compounds) delivered [7]. Microfibres, which are commonly used for surface decontamination [8], have a higher density of strands, when compared with cotton cloths and nonwoven materials, increasing the surface area of the cloth [9]. The cleaning efficacy of microfibre cloths has been proven to be so effective that UK infection-control polices advocate their use with water [10].

The term 'cleaning' describes the physical removal of soil, dirt or dust from surfaces [1], but in the process it may also remove microorganisms from surfaces. Indeed, the use of materials in combination with various detergents including quaternary ammonium compounds for cleaning purposes has been shown to impact not only on the removal of pathogens

* Corresponding author. Address: School of Pharmacy and Pharmaceutical Sciences, Cardiff University Redwood Building, King Edward VII avenue, Cardiff CF10 3NB, UK. Tel.: +44 2920 879088.

E-mail address: maillardj@cardiff.ac.uk (J-Y. Maillard).

from inanimate surfaces but on their transmission to other surfaces [11]. Thus, combining formulated solutions, whether detergent or disinfectant, with materials should be evaluated for their impact in removing and transferring microorganisms from and between surfaces. With this in mind, the impact of using water alone in combination with microfibre materials to remove or prevent pathogen transfer between surfaces has not been widely reported. Here, the impact of using water vs quaternary ammonium compounds (QAC)-based detergent/disinfectant or sporicidal products in combination with a microfibre material was tested using the ASTM2967-15 standard to measure wipe products' efficacy.

Methods

Staphylococcus aureus (ATCC 6538) and *Acinetobacter baumannii* (ATCC 19568) and spores of *Clostridium difficile* (NCTC 11209) were used. Test bacteria inocula were resuspended in a buffer (tryptone 1 g/L; sodium chloride 8.5 g/L; TSC) following overnight propagation at 37°C in tryptone soya broth (TSB; Oxoid) [11]. *C. difficile* spores were resuspended in sterile distilled water following propagation and purification based on the Clospore method [12]. Test bacteria/spores (1×10^9 cfu-spores/mL; final concentration) were added to bovine serum albumin (BSA) at a final concentration of 0.3 g/L (clean condition) or to BSA 3 g/L and sheep erythrocytes 3 ml/L (final concentration; dirty condition). The ASTM 2967-15 [13] was used to measure bacteria/spores removal from, and transfer between surfaces.

A 10-s wiping time with 300 g weight was used with the detergent/disinfectant and sporicidal products as it reflects the conditions of use in practice. With the detergent/disinfectant, surfaces were neutralized immediately after wiping. With the sporicidal product, surfaces were left 15 min before neutralization was performed in accordance with the manufacturer's instructions. For the transfer experiment, the used wipe was used to wipe a clean surface (10 s, 300 g) immediately after the initial wiping. Bacteria deposited on the clean surface were enumerated after neutralization as described below. In addition, considering the ability of spores to survive well in the environment, the growth at 24 h, at 25°C and 40% relative humidity after wiping, following use of the sporicidal product was also investigated.

The conditions for use of water with the microfibre reflected the use of the detergent/disinfectant or sporicidal product. Two surfaces were used: stainless steel (AISI Type 430; 1 cm diameter and 0.7 mm thickness), and polyvinylchloride (PVC with a PUR coating, 1 cm diameter and 0.7 mm thickness; Armstrong, Stuttgarter Str. 75, 74321 Bietigheim Bissingen). Stainless-steel disks were cleaned and sterilized by autoclave. The PVC disks were cleaned then disinfected in 3% peracetic acid. Prior to wiping, surfaces were contaminated with 0.01 mL of test suspension (1×10^8 cfu/mL) and left to dry in a biological safety cabinet until the disks were visibly dry. Surfaces were then tested against a microfibre wipe (Decitex), soaked in sterile water and a wipe soaked in a solution of 0.25% detergent/disinfectant product (containing N-(3-aminopropyl)-N-dodecylpropane-1,3-diamine (5.1%) and didecyldimethyl ammonium chloride (2.5%)) or 0.5% sporicidal product (containing peracetic acid (750 ppm) and N-alkyl(C12-14)-N-benzyl-N,N-dimethylammonium chloride (0.012%)).

The microfibre wipes were then wrung lightly until they were no longer dripping and were used only once. The sporicidal product was only used against *C. difficile* spores. To quench the activity of the detergent/disinfectant and sporicidal products, the following neutralizer was used: 30 g/L polysorbate 80; 30 g/L saponin; 5 g/L sodium thiosulphate; 3 g/L azolectin; 1 g/L histidine; dissolved in TSC. Neutralized suspensions were diluted in TSC, plated on TSA and incubated for 24 h at 37°C for bacteria. Recovered spores were plated on BHI agar containing 0.1% sodium taurocholate for spores and incubated for 48 h, anaerobically (MG500 anaerobic workstation, Don Whitley) at 37°C. Statistical analyses (analysis of variance (ANOVA)) was performed using the R-program [14].

Results and discussion

There was a significant difference (ANOVA, $P < 0.001$) in the number of bacteria removed from surfaces following wiping between the use of water alone and the detergent/disinfectant product, regardless of the type of surface (Table I). Although the performance of combining the microfibre cloth with water reduced bacterial counts mostly by 1–2 \log_{10} (Table I), bacterial transfer from the microfibre to a different surface following wiping was significant (3–4 \log_{10} bacterial transfer) (Table II). In comparison the use of detergent/disinfectant significantly (ANOVA, $P < 0.001$) prevented the transfer of bacteria. The level of organic load did not affect the efficacy of the test product and material performance.

The use of the sporicidal product significantly (ANOVA, $P < 0.001$) reduced the concentration of *C. difficile* spores comparing to the use of water regardless of the type of surfaces and organic load (Table I). Following a 24-h recovery period post-wiping, the sporicidal product performed significantly better (ANOVA, $P < 0.001$) than the use of water. Of practical significance, the use of the sporicidal product prevented the transfer of *C. difficile* spores between surfaces, regardless of the type of surfaces or level of organic load (Table II). The use of water was associated with significant spore transfer 15 min post-wiping or 24 h after wiping.

Although it has been previously suggested that microfibre cloths can reduce the transfer of spores [15], our results clearly indicate that the water-damp microfibre cloth was able to transfer high levels of spores. This suggests that the spores are not retained within the material and are at risk to being re-deposited on to clean surfaces during wiping/mopping. The sporicidal product did not transfer spores and was sporicidal following 10 s wiping time and 15 min surface contact time as recommended by the manufacturer.

Overall the type of surface used did not have a significant effect of the removal of bacteria/spores (ANOVA, $P = 0.754$), or the transfer of bacteria/spores (ANOVA, $P = 0.642$). Likewise soiling had no significant effect on the removal of bacteria/spores (ANOVA, $P = 0.915$) or the transfer of bacteria/spores (ANOVA, $P = 0.424$). Our results also highlighted no significant differences in removal (ANOVA with Tukey post hoc test; $P = 0.959$), or transfer ($P = 0.815$) between vegetative bacteria.

Overall, this in vitro study justified the use of detergent/disinfectant or sporicidal products for the control of microorganisms or spores on surfaces and it does not favour the use of water only. Hamilton and colleagues [16] reported on the performance of ultra-microfibre cloths and mops moistened

Table I

Bacteria/spores removal from surfaces following wiping in clean and dirty conditions

Clean conditions		Log ₁₀ removal (±SD) from surfaces		
Stainless steel	Sampling time ¹	Water	Detergent/disinfectant	Sporicide
<i>Staphylococcus aureus</i>	0	2.10 (0.19)	4.23 (0.25)	
<i>Acinetobacter baumannii</i>	0	2.53 (0.27)	5.21 (1.20)	
<i>Clostridium difficile</i> ²	0	1.38 (0.39)		5.67 (0.06)
	24	1.24 (0.35)		5.99 (0.21)
PVC				
<i>S. aureus</i>	0	2.14 (0.51)	3.19 (0.40)	
<i>A. baumannii</i>	0	2.72 (0.68)	3.86 (0.97)	
<i>C. difficile</i> ²	0	1.88 (0.23)		4.16 (0.18)
	24	1.63 (0.22)		6.14 (0.12)
Dirty conditions		Log ₁₀ removal from surfaces		
Stainless steel	Sampling time ¹	Water	Detergent/disinfectant	Sporicide
<i>S. aureus</i>	0	2.23 (0.18)	4.67 (0.58)	
<i>A. baumannii</i>	0	2.05 (0.39)	4.50 (0.97)	
<i>C. difficile</i> ²	0	1.55 (0.56)		5.89 (0.04)
	24	1.72 (0.21)		6.07 (0.52)
PVC				
<i>S. aureus</i>	0	2.60 (0.79)	4.12 (0.77)	
<i>A. baumannii</i>	0	2.82 (0.36)	5.01 (0.83)	
<i>C. difficile</i> ²	0	1.12 (0.50)		4.28 (0.19)
	24	1.84 (0.34)		5.90 (0.30)

PVC, polyvinylchloride; SD, standard deviation.

¹ Sampling time. 0: surfaces were neutralized immediately after wiping; 24: surfaces were left for 24 h at 25°C and 40% relative humidity before neutralization and processing.² Spores of *C. difficile*.

Table II

Bacteria/spores transfer between surfaces following wiping in clean and dirty conditions

Clean conditions		Log ₁₀ transfer (±SD) between surfaces		
Stainless steel	Sampling time ¹	Water	Detergent/disinfectant	Sporicide
<i>Staphylococcus aureus</i>	0	4.66 (0.53)	0.89 (0.43)	
<i>Acinetobacter baumannii</i>	0	3.52 (1.58)	0.40 (0.00)	
<i>Clostridium difficile</i> ²	0	4.73 (0.44)		0.40 (0.00)
	24	3.69 (0.37)		0.40 (0.00)
PVC				
<i>S. aureus</i>	0	5.09 (0.67)	1.15 (1.05)	
<i>A. baumannii</i>	0	4.55 (0.74)	0.66 (0.45)	
<i>C. difficile</i> ²	0	4.46 (0.46)		0.76 (0.32)
	24	2.52 (0.51)		0.40 (0.00)
Dirty conditions		Log ₁₀ transfer (±SD) between surfaces		
Stainless steel	Sampling time ¹	Water	Detergent	Disinfectant
<i>S. aureus</i>	0	2.92 (0.14)	0.40 (0.00)	
<i>A. baumannii</i>	0	2.39 (0.26)	0.40 (0.00)	
<i>C. difficile</i> ²	0	4.18 (0.55)		0.40 (0.00)
	24	3.12 (0.27)		0.50 (0.17)
PVC				
<i>S. aureus</i>	0	4.64 (0.76)	0.40 (0.00)	
<i>A. baumannii</i>	0	4.30 (0.36)	0.40 (0.00)	
<i>C. difficile</i> ²	0	4.72 (0.59)		0.56 (0.28)
	24	2.61 (0.30)		0.40 (0.00)

PVC, polyvinylchloride; SD, standard deviation.

¹ Sampling time. 0: surfaces were neutralized immediately after wiping; 24: surfaces were left for 24 h at 25°C and 40% relative humidity before neutralization and processing.² Spores of *C. difficile*.

with water or a copper-based biocide in a cross-over trial over a seven-week period in an in situ study. Although this trial did not investigate microbial transfer, the authors reported that the use of the biocide significantly enhanced the efficacy of the microfibre in reducing total viable count. Some hospitals have advocated the use of microfibre materials with water alone with no other interventions. Such practice might need to be reconsidered because the use of a detergent/disinfectant or sporicidal-based product in combination with the microfibre cloth provides the assurance that the potentially harmful bacteria or spore are not only eliminated from surfaces but also prevented from transfer to any other surfaces during wiping/mopping. As published 10 years ago [17], wipes should be used on 'one surface, one direction' before being disposed of; this however may be more difficult to apply to mops. Staff training is essential to minimize the spread of pathogens when using such materials [18].

Conflict of interest statement

The authors have no conflicts of interest to declare.

Funding sources

This project was funded by a grant from the Laboratoires Anios (France).

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