



# A Highly Selective “Turn-on” Fluorescent Probe for Detection of Fe<sup>3+</sup> in Cells

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## Abstract

A new “turn-on” fluorescent probe **Py** based on rhodamine and piperonaldehyde was designed and synthesized for detecting Fe<sup>3+</sup> in cells. The free probe **Py** was non-fluorescent. While only upon addition of Fe<sup>3+</sup>, the significant increase of the fluorescence and color were observed which could be visible directly by “naked-eye”. The probe **Py** shows high selectivity and sensitivity for Fe<sup>3+</sup> over other common metal ions in EtOH-H<sub>2</sub>O (3/2, v/v) mixed solution. The association constant and the detection limit were calculated to be  $4.81 \times 10^4 \text{ M}^{-1}$  and  $1.18 \times 10^{-8} \text{ mol/L}$  respectively. The introduction of piperonaldehyde unit could increase probe rigidity which could enhance its optical properties. Meanwhile, the binding mode between **Py** and Fe<sup>3+</sup> was found to be a 1:1 complex formation. The density functional theory (DFT) calculations were performed which would further confirm the recognition mechanism between probe **Py** and Fe<sup>3+</sup>. In addition, the probe has been proved to be reversible for detecting Fe<sup>3+</sup>. Moreover, the probe **Py** was used to detect Fe<sup>3+</sup> in cells successfully.

**Keywords** Fluorescent probe · Rhodamine · Fe<sup>3+</sup> · Cell imaging · DFT calculations

## Introduction

The normal life activities of the human body are inseparable from trace elements. Iron as one of the most essential trace elements, it exists in ferrous (Fe<sup>2+</sup>) and ferric (Fe<sup>3+</sup>) forms in living organism. The iron ions play an important role in the fundamental physiological processes of living systems. It is related to the biological cellular metabolism, gene transcription, oxygen metabolism, electron transit, protein synthesis, enzyme catalysis, DNA and RNA synthesis and so on [1–4]. Whether the iron ion content in the organism is excessive or deficient will induce various diseases such as heart failure, neurodegenerative, anemia, liver and kidney damage and

diabetes [5–8]. No that Fe<sup>2+</sup> can be easily converted to Fe<sup>3+</sup>, discrimination of Fe<sup>3+</sup> from Fe<sup>2+</sup> is important to understand the biological processes such as iron metabolism. So it is very important to design a good method which could detect Fe<sup>3+</sup> in biological organism. Over the past years, many fluorescent sensors for Fe<sup>3+</sup> have been reported because of its simple operation, high sensitivity, selectivity and response speed, as well as low detection limit [9, 10]. In the early days, most of them are “turn-off” types, because of the paramagnetic nature of Fe<sup>3+</sup> which could quench the fluorescence of probe [11, 12]. Now many “turn-on” fluorescent sensors based on Rhodamine family derivatives have been reported due to their long absorption and emission wavelengths, high fluorescence quantum yield, large absorption coefficient and the property of spirocyclic closed ring (non-fluorescent and colorless) and ring-opened (fluorescent and strong absorption) of the rhodamine family compounds which could efficiently respond to metal ions [13–25]. A solvent-dependent fluorescent probe based on rhodamine B derivative has been reported because of its high selectivity and low detection limit [26]. A optical–electrochemical multichannel chemosensor has been reported which could detect Fe<sup>3+</sup> through multiple physical responses [27]. A reversible probe based on Rhodamine B also has been reported but it does not show the reversibility well [28]. A fluorescent probe based on rhodamine6G for detecting Fe<sup>3+</sup>

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in water has been reported, but it is hard to distinguish  $\text{Fe}^{3+}$  from  $\text{Hg}^{2+}$ ,  $\text{Cr}^{3+}$  directly in water [29]. Furthermore, there are still some deficiencies for most probes, such as very poor solubility in water [30] and interfered a little to other ions such as  $\text{Cr}^{3+}$ ,  $\text{Cu}^{2+}$  [31, 32]. Other than this, some probes are cell impermeable or need long response time [33]. In addition, strict working condition and high excited energy could limit the biological applications of them [34, 35]. What's more, only a few probes having been reported are reversible due to their reaction-based nature [36]. Therefore, designing a reversible and highly selective probe with better biological properties is still full of challenges.

In this work, we synthesized a fluorescent probe **Py** (Scheme 1) based on rhodamine and piperonaldehyde, which was characterized by IR, NMR and HRMS. The introduction of piperonaldehyde unit could increase the rigidity of the probe which could enhance its optical performance. In addition, the imide structure can coordinate well with metal ions. The probe **Py** exhibits high selectivity and sensitivity for  $\text{Fe}^{3+}$  detection in EtOH- $\text{H}_2\text{O}$  (3/2, v/v) over other commonly coexistent metal ions. In addition, the probe **Py** can be used as a “naked-eye” sensor to detect  $\text{Fe}^{3+}$  by significant color change. The limit of probe **Py** detection for  $\text{Fe}^{3+}$  was found to be as low as  $1.18 \times 10^{-8}$  mol/L which was lower than most researches [36]. The DFT calculations further better explained the recognition mechanism between probe **Py** and  $\text{Fe}^{3+}$ . We also confirmed the reversibility of the probe for detection  $\text{Fe}^{3+}$  which can reduce the cost of its application. The most important is that the biological imaging of the probe **Py** was used in SGC7901 cells successfully and had been tested to have low biological toxicity.

## Experimental

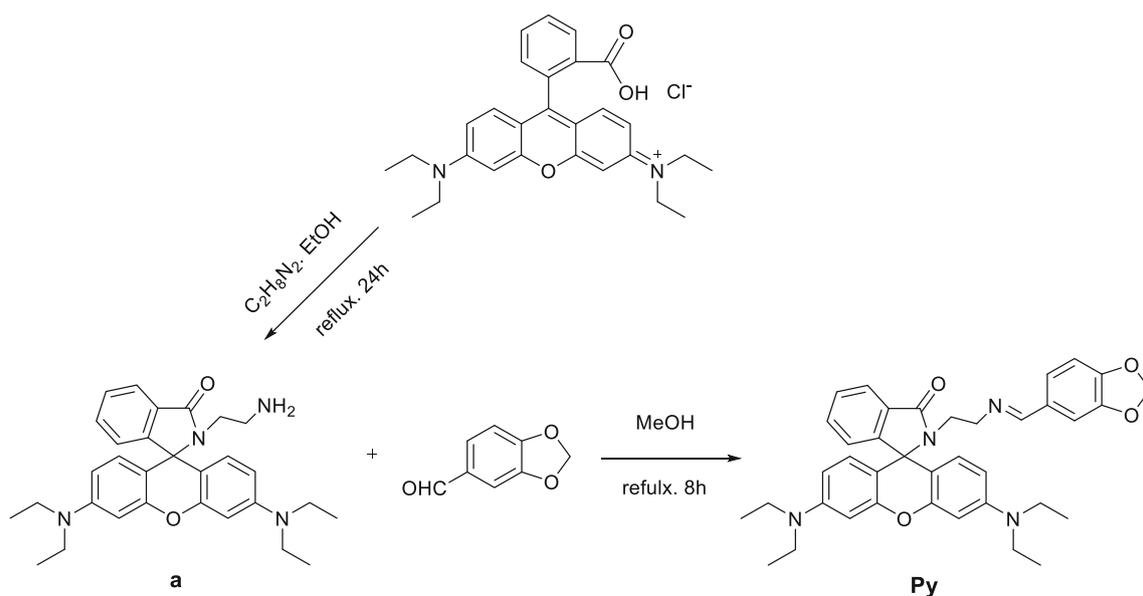
### General Information

Mass spectra was performed with Bruker micro TOF-QII.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded on a Varian unity INOVA-400 MHz spectrometer, using the solvent the TMS signal as an internal standard. The fluorescence spectra were measured on HITACHI F-4500 fluorescence spectrophotometer. IR spectra were taken on a Bruker EQUINOX-55 spectrometer with KBr pellets. The absorbance spectra were recorded on a Shimadzu UV-1700 spectrophotometer. Melting points were obtained on a Laboratory Devices XT4B melting apparatus and uncorrected. All the solvents and reagents (analytical grade) were purchased from commercial suppliers and used without further purification. Double distilled water was used throughout the experiment. The solutions of metal ions were prepared from corresponding nitrate and chloride salts.

Fluorescence quantum yield ( $\varphi_f$ ) is the ratio of the number of photons emitted after absorption of energy by the fluorescent material to the number of photons absorbed. In this paper, 0.05 mol/L rhodamine B solution is used as reference solution, and then we calculated the fluorescence quantum yield of the sample under test [Eq. (1)] [37]:

$$\varphi_f = \varphi_s \times \frac{(F_f \times A_s)}{(F_s \times A_f)}$$

$\varphi_f$  and  $\varphi_s$  in the formula are the fluorescence quantum yield of the sample to be measured and the reference sample, respectively;  $A_f$  and  $A_s$  are the absorbance of the excitation light



**Scheme 1** The synthetic route to **Py**

of the sample and the reference sample respectively. The fluorescence quantum yield of the compound **Py** was measured to be 0.31 (At the same excitation wavelength and in the same solvent).

### Preparation of the Test Solution

A stock solution of probe **Py** ( $2 \times 10^{-4}$  mol/L): in a 25 mL volumetric flask, 154.1 mg of **Py** was dissolved in dichloromethane and then diluted to the mark with ethanol. 0.5 mL of the solution was then transferred to a 25 mL volumetric flask and diluted to the mark with ethanol.

Stock solutions of the metal ions were prepared in double distilled water with the metal salts (KCl, NaCl, LiCl, MgCl<sub>2</sub>, BaCl<sub>2</sub>, AgNO<sub>3</sub>, CaCl<sub>2</sub>, ZnCl<sub>2</sub>, CdCl<sub>2</sub>, Cu(NO<sub>3</sub>)<sub>2</sub>, MnCl<sub>2</sub>, CoCl<sub>2</sub>, NiCl<sub>2</sub>, PbCl<sub>2</sub>, Al(NO<sub>3</sub>)<sub>3</sub>, FeCl<sub>3</sub>, HgCl<sub>2</sub>, CrCl<sub>3</sub>, FeCl<sub>2</sub>).

### Syntheses

Rhodamine B ethylenediamide (**a**) was synthesized from rhodamine B and ethylenediamine used the literature procedure [38]. yield 73.6%. ESI-HRMS Calcd for (C<sub>30</sub>H<sub>36</sub>N<sub>4</sub>O<sub>2</sub> + H<sup>+</sup>) m/z = 485.2916. Found: 485.2623 (M + H<sup>+</sup>).

**Synthesis of probe Py:** In a single necked round-bottomed flask, compound **a** (0.49 g, 1 mmol) and pepper aldehyde (0.18 g, 1 mmol) were mixed into 20 mL methanol solution. The mixture was heated to reflux. The process of reaction was monitored by TLC. After cooled to room temperature, the precipitate was separated by filtration and further purified by recrystallizing from ethanol to obtained pink solid product. Yield 53.5%, m.p.183–184 °C. IR (KBr)  $\nu_{\max}/\text{cm}^{-1}$ : 3432, 3376, 3085, 3043, 2973, 2896, 1697, 1617, 1550, 1517, 1448, 1376, 1349, 1299, 1249, 1228, 1118, 1039, 929, 813, 752, 698, 601, 536. <sup>1</sup>H NMR (400 MHz; CDCl<sub>3</sub>;Me<sub>4</sub>Si),  $\delta$  (ppm): 7.93 (s, 1H, ArH), 7.91–7.89 (m, 1H,ArH), 7.42–7.40 (m, 2H, ArH), 7.20 (d,  $J = 1.5$  Hz, 1H,ArH), 7.07–7.05 (m, 1H, ArH), 6.99 (dd,  $J_1 = 8.0$ ,  $J_2 = 1.5$  Hz, 1H, ArH), 6.75

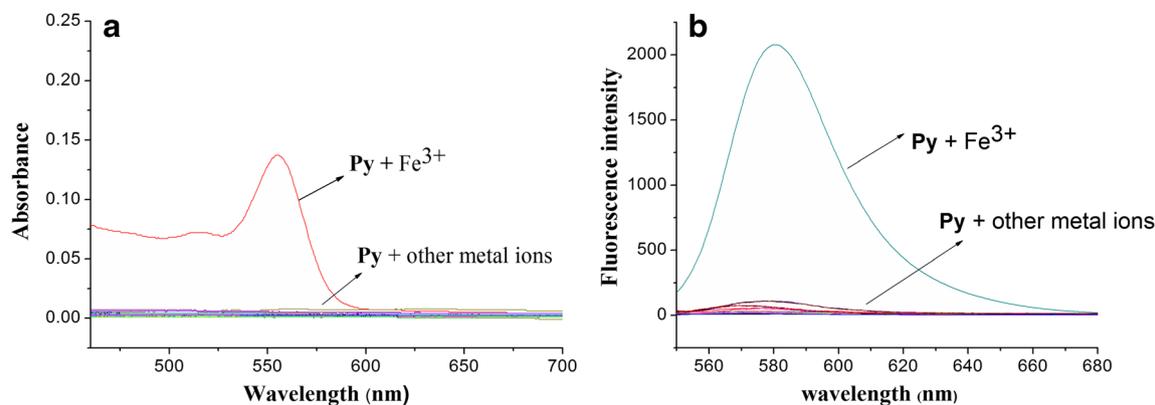
(d,  $J = 8.0$  Hz, 1H,ArH), 6.44 (d,  $J = 8.8$  Hz, 2H, ArH), 6.38 (d,  $J = 2.6$  Hz, 2H, ArH), 6.25 (dd,  $J_1 = 6.4$ ,  $J_2 = 2.6$  Hz, 2H, ArH), 5.95 (s, 2H, OCH<sub>2</sub>O), 3.41 (s, 4H, NCH<sub>2</sub>CH<sub>2</sub>N), 3.32 (q,  $J = 7.1$  Hz, 8H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>4</sub>), 1.16 (t,  $J = 7.0$  Hz, 12H, N(CH<sub>2</sub>CH<sub>3</sub>)<sub>4</sub>). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>),  $\delta$  (ppm): 168.6, 161.9, 154.1, 153.5, 149.7, 149.05, 147.8, 132.1, 131.1, 129.2, 128.2, 124.4, 123.7, 122.7, 106.8, 105.6, 101.2, 97.9, 64.96, 58.9, 44.3, 41.3, 12.7. ESI-HRMS calcd for (C<sub>38</sub>H<sub>40</sub>N<sub>4</sub>O<sub>4</sub> + H<sup>+</sup>) m/z = 617.3128. Found: 617.3125 (M + H<sup>+</sup>).

### Cells Imaging Study

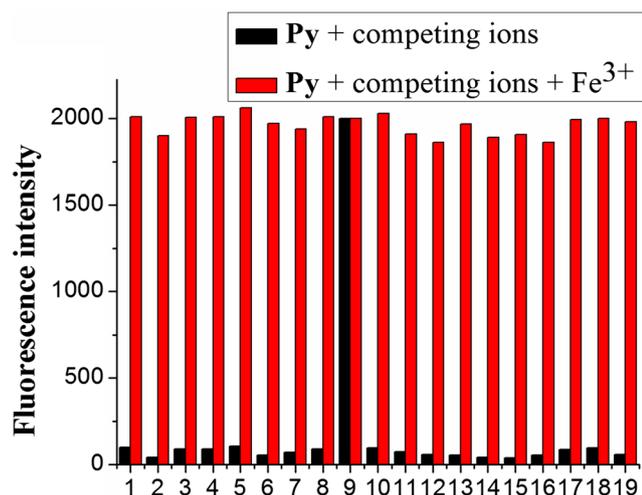
The probe was dissolved in a small amount of DMSO and then dissolved it in water to form a stock solution. The cell lines were then added with **Py** (in aqueous DMSO, culture medium) to obtain a final concentration of 10  $\mu\text{M}$  **Py** and incubated under reoxygenation (95% air, 5% CO<sub>2</sub>) for 30 min at 37 °C. After the incubation, the cell lines were washed with PBS to remove excess probe and monitored with confocal microscope. Finally, these cells were treated with Fe<sup>3+</sup> ion solution (10  $\mu\text{M}$ ) and fresh DMEM culture medium for 1 h at 37 °C and then washed with PBS for three times. Cells were observed using confocal fluorescence microscopy at  $\lambda_{\text{ex}}/\lambda_{\text{em}} = 550 \text{ nm}/560\text{--}680 \text{ nm}$ .

### Cells Toxicity Study

Cells toxicity and survival rate study were tested by CCK-8 assay. Cells were seeded into 96-well plates and cultured at 37 °C (5% CO<sub>2</sub>) for 24 h, different concentrations of probe **Py** (0, 6.25, 12.5, 25, 50 and 100  $\mu\text{M}$ ) were then added to the wells. After incubation for 24 h, CCK-8 (10% in culture medium) was added to each well, and the plate was incubated for another 1 h. Absorbance was measured at 450 nm. All experiments were repeated three times, and the data were presented as the percentage of control cells.



**Fig. 1** **a** The UV-vis spectra of **Py** solution in the presence of several different metal ions. **b** Fluorescence spectra of **Py** (20  $\mu\text{M}$ ) in the presence of several different metal ions (20  $\mu\text{M}$ ) in EtOH-H<sub>2</sub>O (3/2, v/v) solution,  $\lambda_{\text{ex}} = 550 \text{ nm}$



**Fig. 2** Fluorescence intensity changes of **Py** (20  $\mu\text{M}$ ) upon the addition of various metal ions (20  $\mu\text{M}$ ) in the absence and presence of  $\text{Fe}^{3+}$  (20  $\mu\text{M}$ ) in EtOH- $\text{H}_2\text{O}$  (3/2, v/v) solution. 1  $\text{Ag}^+$ , 2  $\text{Al}^{3+}$ , 3  $\text{Ba}^{2+}$ , 4  $\text{Ca}^{2+}$ , 5  $\text{Cd}^{2+}$ , 6  $\text{Co}^{2+}$ , 7  $\text{Cr}^{3+}$ , 8  $\text{Cu}^{2+}$ , 9  $\text{Fe}^{3+}$ , 10  $\text{Hg}^{2+}$ , 11  $\text{K}^+$ , 12  $\text{Li}^+$ , 13  $\text{Mg}^{2+}$ , 14  $\text{Mn}^{2+}$ , 15  $\text{Na}^+$ , 16  $\text{Ni}^{2+}$ , 17  $\text{Pd}^{2+}$ , 18  $\text{Zn}^{2+}$ , 19  $\text{Fe}^{2+}$  ( $\lambda_{\text{ex}} = 550 \text{ nm}/\lambda_{\text{em}} = 580 \text{ nm}$ )

## Results and Discussion

### Synthesis of Probe **Py**

The synthesis of **Py** was shown in Scheme 1. It was characterized by  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, IR and HRMS, the corresponding spectra were shown in the Supporting Information.

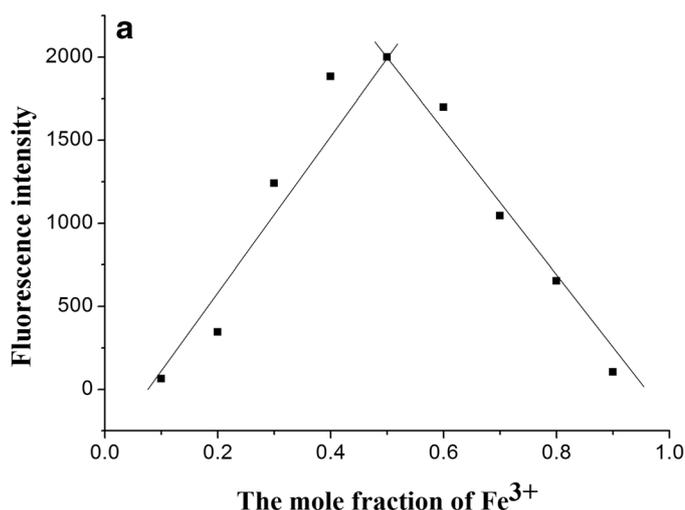
### Selective Study

The UV–vis absorption and fluorescence spectral behavior of **Py** to a series of metal ions were investigated, including  $\text{Ag}^+$ ,

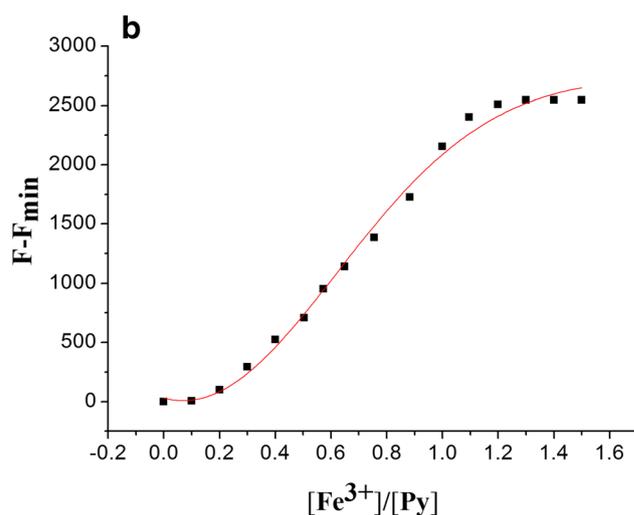
$\text{Al}^{3+}$ ,  $\text{Ba}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Cr}^{3+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Hg}^{2+}$ ,  $\text{K}^+$ ,  $\text{Li}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Ni}^{2+}$ ,  $\text{Pd}^{2+}$ ,  $\text{Zn}^{2+}$  and  $\text{Fe}^{2+}$  respectively. As shown in Fig. 1a, a remarkable enhancement in absorption spectra at 550 nm was observed when  $\text{Fe}^{3+}$  was added while all other metal ions did not make apparent absorbance enhancement. Moreover, upon the addition of  $\text{Fe}^{3+}$ , the solution of probe **Py** immediately yield a pink color (Fig. S1). Under the identical condition, no obvious response could be observed with the addition of other metal ions. As shown in Fig. 1b, the same results in fluorescence spectra were observed at 580 nm. When added other metal ions, the probe **Py** would not combine these ions, so it keep five-membered spirolactam structure. However, when  $\text{Fe}^{3+}$  was added, the probe **Py** lactam ring was opened, resulting in strong fluorescence and color changes. These results demonstrated that probe **Py** had a high selectivity for  $\text{Fe}^{3+}$  over other ions and it could serve as a fluorescent and “naked eye” probe for detecting  $\text{Fe}^{3+}$  in EtOH- $\text{H}_2\text{O}$  mixture.

### Interference Study

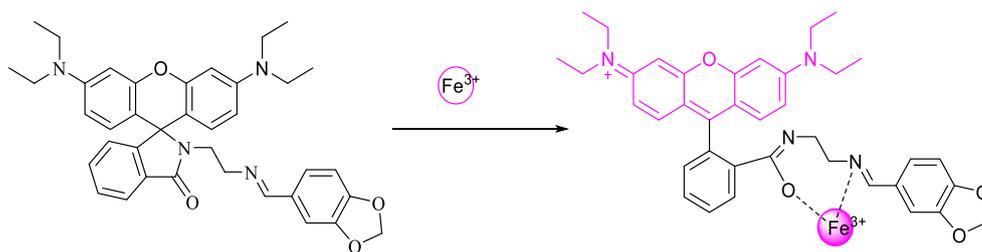
Besides  $\text{Fe}^{3+}$ , there are many other metal ions in the living body. Therefore, in order to further test the selectivity of **Py** to  $\text{Fe}^{3+}$  from other ions, the selectivity of **Py** (20  $\mu\text{M}$ ) for  $\text{Fe}^{3+}$  over other metal ions (1.0 equiv) were explored by the competition assay in the presence of other various metal ions. As shown in Fig. 2, there was almost no effect on the fluorescence of  $\text{Fe}^{3+}$ -**Py** in the presence of various interference metal ions. Selectivity and competition experiments indicated that probe **Py** could specifically detect  $\text{Fe}^{3+}$  with little interference from other commonly coexistent ions.



**Fig. 3** a The Job's plot of probe **Py** and  $\text{Fe}^{3+}$  (the total concentration was 40  $\mu\text{M}$ ) in EtOH- $\text{H}_2\text{O}$  (3/2, v/v) solution; (b) Non-linear plot of probe **Py** (20  $\mu\text{M}$ ) assuming a 1:1 stoichiometry for association between probe



**Py** and  $\text{Fe}^{3+}$  in Ethanol/ $\text{H}_2\text{O}$  (v/v, 3/2) solution by fluorescence spectroscopy.  $\lambda_{\text{em}} = 550 \text{ nm}/\lambda_{\text{em}} = 580 \text{ nm}$

**Scheme 2** The binding mechanism between **Py** and  $\text{Fe}^{3+}$ 

### Effect of $\text{Fe}^{3+}$ Concentration Study

Fluorescence and UV-vis titration experiments were also carried out. Under neutral conditions, rhodamine-type probes exist in a spiro ring structure which makes **Py** colorless and non-fluorescent. The intensity of fluorescence at 580 nm and UV absorption at 550 nm are increasing with the concentration of  $\text{Fe}^{3+}$  (0–1.25 equiv) increased (Fig. S2 and S3). It is attributed to the delocalization effects in the xanthene moiety of the rhodamine. These results clearly indicated that  $\text{Fe}^{3+}$  could bind to the probe **Py**, thus the probe **Py** lactam ring was opened and formed a highly delocalized p- $\pi$  conjugated

structure. Therefore, significant enhancement of absorbance and fluorescence were observed.

According to the Job's plots, the fluorescence intensity reached maximum when the molar fraction of  $\text{Fe}^{3+}$  was 0.5 (Fig. 3a). This showed that the binding stoichiometry of  $\text{Fe}^{3+}$  to the probe **Py** molecule was found to be 1:1. Conceivably,  $\text{Fe}^{3+}$  with probe **Py** formed a 1:1 metal complex which induced the spirane structure of probe **Py** to be opened. The binding model was proposed and shown in Scheme 2.

The fluorescence spectrum association constants of  $\text{Fe}^{3+}$  were calculated by nonlinear fitting (Fig. 3b) using the following formula [Eq. (2)] [39]:

$$\Delta F = \frac{\Delta\beta \left( [H]_0 + [G]_0 + 1/K_a \right) \pm \sqrt{\Delta\beta^2 \left( [H]_0 + [G]_0 + 1/K_a \right)^2 - 4\Delta\beta^2 [H]_0 [G]_0}}{2}$$

$\Delta F$  is the change in the fluorescence intensity of the **Py** upon gradual addition of the  $\text{Fe}^{3+}$ , and  $\Delta\beta$  refers to the different constant of the free host and the interaction complex. The total concentrations of host and guest are denoted by  $[H]_0$  and  $[G]_0$ , respectively. The association constant of **Py** with  $\text{Fe}^{3+}$  was accordingly calculated to be  $4.81 \times 10^4 \text{ M}^{-1}$  ( $R = 0.9935$ ). The limit of detection for  $\text{Fe}^{3+}$  was calculated to be  $1.18 \times 10^{-8} \text{ mol/L}$ . These results indicated that probe **Py** holds great potential for using in the development of sensor materials for  $\text{Fe}^{3+}$ .

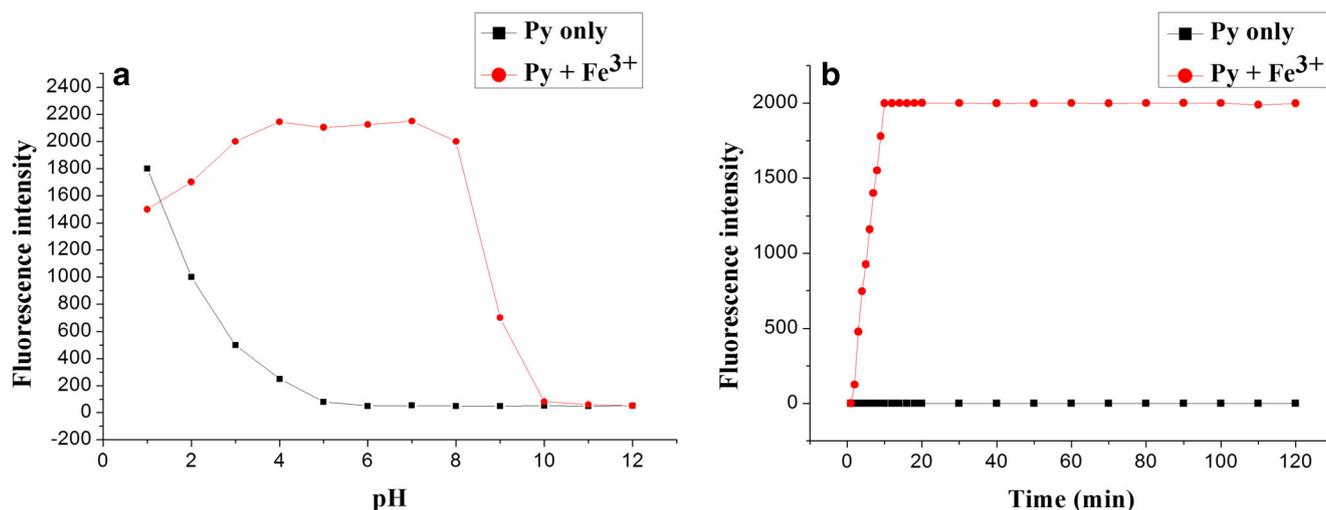
### Effect of pH and Response Time Study

Normally, the spironolactone ring of the rhodamine derivatives would be opened in acidic media. Therefore, it is necessary to evaluate the effect of pH on the fluorescence of probe **Py**. The effects of pH on probe **Py** experiment was evaluated in the different pH media (Fig. 4a). No obvious enhancement of fluorescence at 580 nm was observed in the pH range of 5.0–12.0, suggesting that it is insusceptible to the change of acid–base solution. However, in the presence of  $\text{Fe}^{3+}$ , a remarkable fluorescence emission band at 580 nm was formed under different pH conditions. It showed that the pH corresponded to the highest response approximately in 4.0–8.0, which revealed that the probe **Py** for  $\text{Fe}^{3+}$  could work well

in approximate physiological conditions and with very low background response. When the pH exceed 10.0, the fluorescence of probe **Py** would vanish, this is because the high concentration of  $\text{OH}^-$  would combine with  $\text{Fe}^{3+}$  to form  $\text{Fe}(\text{OH})_3$ . As shown in Fig. 4b, with the increase of the reaction time, the fluorescence intensity of probe **Py** with  $\text{Fe}^{3+}$  increased and reached equilibrium within 10 min. These indicated that probe **Py** could serve as an efficient probe for detection of  $\text{Fe}^{3+}$  in a neutral medium in a short time.

### Cell Imaging and Cytotoxicity Study

To test the potential biological application of probe **Py** for detecting  $\text{Fe}^{3+}$  in living cells, the fluorescence imaging was recorded using confocal fluorescence microscopy. The human cancer cells SGC7901 (stomach cells) were cultured in high glucose type DMEM culture medium and then further incubated with the probe **Py** under reoxygenation (95% air, 5%  $\text{CO}_2$ ) for 30 min at 37 °C. After incubation, excess unbound probe were washed with PBS buffer. No intracellular fluorescence was observed inside the cells (Fig. 5a) but the bright field image of cells was seen clearly in Fig. 5b which proved that the cell could remain in good condition after incubated with probe. These cells were treated with  $\text{Fe}^{3+}$  (10  $\mu\text{M}$ ) for 1 h at 37 °C and then washed with PBS buffer to remove excess



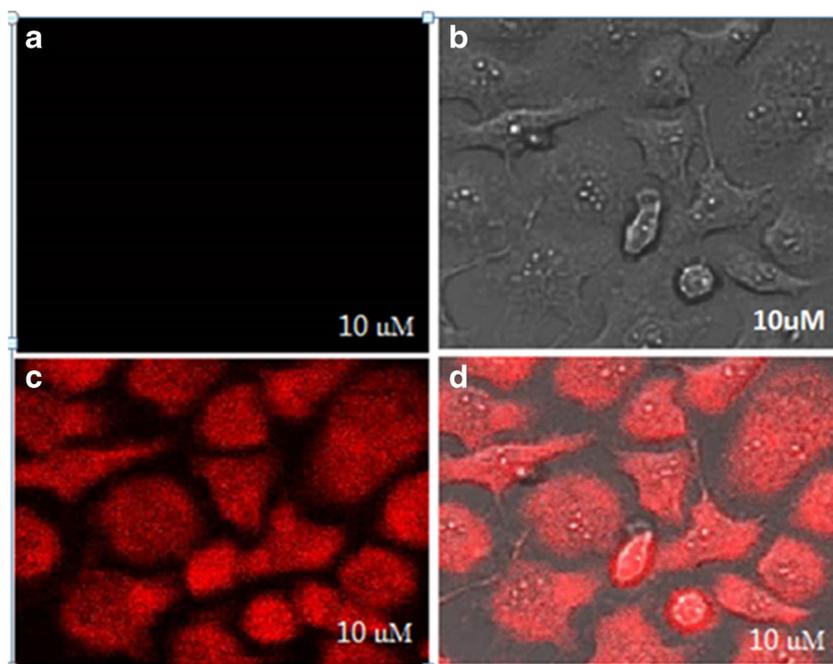
**Fig. 4** **a** Effect of pH on the fluorescence intensity of **Py** (black line) and **Py-Fe<sup>3+</sup>** (red line). **b** Effect of time on the fluorescence intensity of **Py** (black line) and **Py-Fe<sup>3+</sup>** (red line),  $\lambda_{\text{ex}} = 550 \text{ nm}/\lambda_{\text{em}} = 580 \text{ nm}$

$\text{Fe}^{3+}$ . Obviously, a significant fluorescence from the intracellular area was observed (Fig. 5c), this indicated that probe **Py** has the fine cell membrane permeability and could be used to detect  $\text{Fe}^{3+}$  in cells. The overlay image (Fig. 5d) of bright field and fluorescence image further confirmed that the fluorescent signals were localized in cells. As shown in **Table S1** (Supporting Information), the cells survival rate was as high as 82.43% after they were incubated with 100  $\mu\text{M}$  probe **Py** for 24 h. These data demonstrated that probe **Py** has superior biocompatibility and low cytotoxicity. Therefore it has the potential value for detecting  $\text{Fe}^{3+}$  in the biological system due to its ideal chemical, biological and spectroscopic properties.

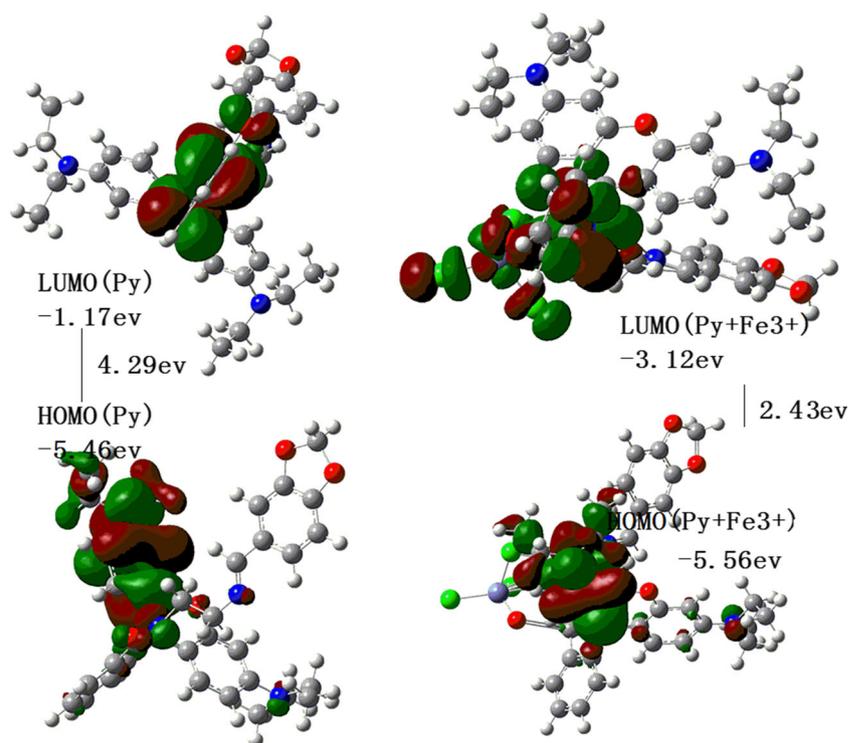
### Theoretical Calculations

In order to better understand the reaction mechanism between probe **Py** and  $\text{Fe}^{3+}$ , the DFT calculations of **Py** and **Py-Fe<sup>3+</sup>** complex were performed and the basis set was B3LYP/6-31G\*\* [23, 40, 41]. The optimized geometries of **Py** and **Py-Fe<sup>3+</sup>** complex were shown in **Fig. S4**. As shown in **Fig. S4b**, the binding sites with  $\text{Fe}^{3+}$  are N and O atoms. According to NBO analysis [40] of the lone pair orbitals of N and O atoms in the probe **Py**, the Fukui function of O (0.036), N<sub>2</sub> (0.014), and N<sub>1</sub> (-0.007) were calculated respectively. The *f* value of the N<sub>2</sub> and O atoms were shown to be larger than that of the N<sub>1</sub> atom. These indicated that the O and

**Fig. 5** Confocal dark-field fluorescence image of SGC7901 cells incubated with **Py** (10  $\mu\text{M}$ ) for 30 min. **a** Dark-field image **(b)** Bright-field image. **c** Fluorescence image of the cells in **(b)** further incubated with  $\text{Fe}^{3+}$  (10  $\mu\text{M}$ ) for 1.0 h. **d** Overlay image of **(b)** and **(c)**



**Fig. 6** HOMO and LUMO orbitals of probe **Py** and the **Py-Fe<sup>3+</sup>** complex



$N_2$  are better active sites than  $N_1$  atom which facilitates its contact with  $Fe^{3+}$ . Therefore, compared with the  $N_1$  atom, the combination of the  $Fe^{3+}$  with the  $N_2$  and O atoms should be more energetically favorable. As shown in Scheme 2, the spirocyclic form of probe **Py** was opened by the transfer of electrons, which induce the changes in fluorescence. In summary,  $Fe^{3+}$  is more likely to complex with  $N_2$  and O atom than  $N_1$ , thus a 1:1 **Py-Fe<sup>3+</sup>** complex was formed. We also calculated HOMO and LUMO orbitals of probe **Py** and the **Py-Fe<sup>3+</sup>** complex. As shown in Fig. 6, in probe **Py**, the HOMO are spread out on the xanthenes of the rhodamineB moiety while the LUMO are centered on the spirocycle of the rhodamineB moiety. In **Py-Fe<sup>3+</sup>** complex, maximum electron concentrated on the rhodamineB moiety at HOMO, while the major electron distribution was on metal centre at LUMO. The energy gap ( $\Delta E$ ) exhibited that the binding of  $Fe^{3+}$  to **Py** lowered the HOMO-LUMO energy gap of the complex and stabilized the system. In **Py-Fe<sup>3+</sup>** complex, the distance of  $Fe^{3+}$  to O and  $N_2$  are calculated to be 2.02 and 1.95 Å respectively, while it is 2.63 Å in “ $Fe^{3+}$ - $N_1$ ” bond. These results further explained the model of reaction mechanism between probe **Py** and  $Fe^{3+}$ .

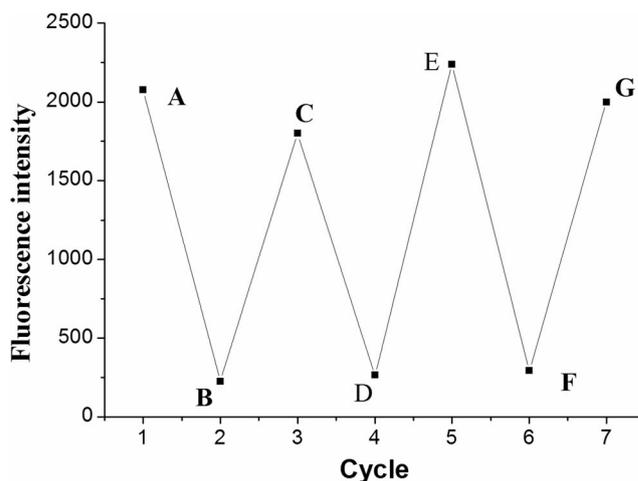
### Reversibility Study

To further study the reversibility of the probe **Py** for detecting  $Fe^{3+}$ , we added ethylenediamine (ED) (20  $\mu$ M) to the solution of **Py-Fe<sup>3+</sup>** (20  $\mu$ M), as show in Fig. 7, the fluorescence intensity changed to weak (A  $\rightarrow$  B). When  $Fe^{3+}$  (20  $\mu$ M) were

added to the system again, however, the fluorescence intensity enhanced again (B  $\rightarrow$  C). The above process can be repeated several cycles and still remained the same orderliness without significant changes in the fluorescence spectrum. This proved that the probe detects  $Fe^{3+}$  is reversible and can be reused.

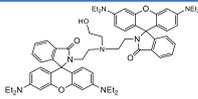
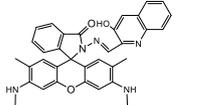
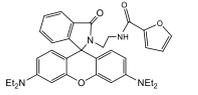
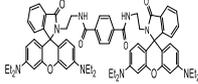
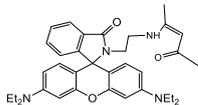
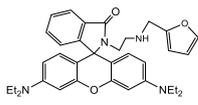
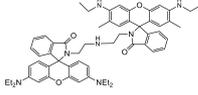
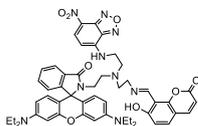
### Comparison with Other Probes

We also compared the probe **Py** with some other previously reported  $Fe^{3+}$  fluorescent probes. As shown in Table 1, most



**Fig. 7** The fluorescence intensity for cycles of the recognition process of **Py** (20  $\mu$ M) toward  $Fe^{3+}$  (20  $\mu$ M) in EtOH-H<sub>2</sub>O (3/2, v/v) solution,  $\lambda_{ex}$  = 550 nm/ $\lambda_{em}$  = 580 nm

**Table 1** The comparison of **Py** with some other rhodamine derivative probes for Fe<sup>3+</sup>.

Sensor	Solution (V/V)	LOD ( $\mu\text{M}$ )	Interference	Reference
	MeOH/H <sub>2</sub> O = 1:1,	0.107	Al <sup>3+</sup> , Cr <sup>3+</sup>	[4]
	aqueous medium	0.033	Fe <sup>2+</sup>	[32]
	MeOH/H <sub>2</sub> O = 1:1,	0.437	Al <sup>3+</sup> , Cu <sup>2+</sup>	[33]
	THF-H <sub>2</sub> O = 4/6,	0.0231	Al <sup>3+</sup> , Hg <sup>2+</sup>	[36]
	Water (PH = 5.5)	1.0	Cu <sup>2+</sup>	[42]
	EtOH/H <sub>2</sub> O = 1:1	0.025	Al <sup>3+</sup>	[43]
	EtOH/H <sub>2</sub> O = 1:1	0.16	Fe <sup>2+</sup> , Hg <sup>2+</sup>	[44]
	CH <sub>3</sub> CN/H <sub>2</sub> O = (97/3)	0.074	/	[25]
<b>Py</b>	EtOH/H <sub>2</sub> O = 3:2	0.0118	/	This work

of them are not only interfered to other metal ions but also have higher detection limit. However, probe **Py** shows higher selectivity and lower detection limit than most of them. Therefore, it has some certain advantages in application.

## Conclusion

In summary, we synthesized a highly selective “turn-on” fluorescent probe **Py** for detecting Fe<sup>3+</sup>. In addition, it can be

recognized by the “naked eye”. The probe **Py** shows high selectivity, sensitivity, reversibility and low cytotoxicity. The reaction mechanism between probe **Py** and  $\text{Fe}^{3+}$  were performed using DFT calculation. Importantly, the probe **Py** was used to sense  $\text{Fe}^{3+}$  in cells successfully. These features will make it more potential for biological applications.

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