



Mini-review

HOX cluster-embedded antisense long non-coding RNAs in lung cancerLianlian Li^{a,*}, Yong Wang^b, Guoqiang Song^c, Xiaoyu Zhang^a, Shan Gao^a, Hongyan Liu^{a,**}^a Institute of Basic Medicine, Shandong Academy of Medical Sciences, Jinan, 250062, China^b Shandong Xinchuang Biotechnology Co., LTD, Jinan, 250102, China^c Shandong University, Jinan, 250012, China

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ABSTRACT

Homeobox (HOX) genes play vital roles in embryonic development and oncogenesis. In humans, there are 39 *HOX* genes found in four clusters that are located on different chromosomes. The *HOX* clusters also contain numerous non-protein-coding RNAs, including some lncRNAs. The *HOX* cluster-embedded lncRNAs (*HOX*-lncRNAs), most notably, *HOTTIP* and *HOTAIR* play a major role in the regulation of their adjacent coding genes. Recently, most *HOX*-lncRNAs have been shown to impact tumorigenesis and cancer progression. Several *HOX*-lncRNAs, including *HOTTIP*, *HOXA11-AS*, *HOTAIRM1*, *HOXA-AS3*, *HOXA10-AS*, *HOTAIR*, and *HAGLR*, are dysregulated in lung cancer. Moreover, their expression levels are correlated with the clinical features of this disease. These *HOX*-lncRNAs regulate the proliferation, invasion, migration, and chemo-resistance of lung cancer cells through various molecular mechanisms. Although lncRNAs have received much attention lately, the functions of some *HOX*-lncRNAs in the development of cancer are unclear. Thus, *HOX*-embedded lncRNAs should be widely investigated in cancer. Here, we review the functions of *HOX*-lncRNAs in lung cancer.

1. Introduction

Long non-coding RNAs (lncRNAs) are generally defined as RNA products that more than 200 nucleotides in length produced from non-protein-coding RNA genes [1]. lncRNAs can be divided into sense, antisense, intronic, bidirectional, and intergenic forms according to their relationship with adjacent protein-coding genes [2]. They regulate fundamental biochemical and cellular processes via multiple different mechanisms, including but not limited to, direct interaction with mRNAs, micro-RNAs, or proteins [3]. Recently, an increasing number of studies have shown that lncRNAs regulate the initiation and progression of human cancers, as well as the response of these diseases to therapy [4–6]. Lung cancer has the highest death rate among all human cancers [7]. Morphologically, lung cancer is classified into small cell lung cancer (SCLC) and non-small cell lung cancer (NSCLC) sub-types, and NSCLC accounts for about 85% of total cases and can be further subdivided into lung adenocarcinoma (LUAD), lung squamous cell carcinoma (LUSC), and large cell carcinoma [8,9]. Continued research has discovered the roles of some lncRNAs in the proliferation, invasion, and survival of lung cancer cells, as well as demonstrating the correlation in lncRNA expression to metastasis, advanced pathological stages, and prognosis in lung cancer patients [10].

Homeobox (HOX) genes, which have essential functions in

embryonic development and tumorigenesis, are characterized by the presence of highly conserved homeodomains. In humans, 39 *HOX* genes are divided into 4 clusters (*HOXA*, *HOXB*, *HOXC*, and *HOXD*), located on different chromosomes (7p15.2, 17q21.32, 12q13.13 and 2q31.1, respectively) [11]. In addition to their roles in normal development and in the determination of cell fate, they also control other cellular processes, which has been demonstrated by descriptions of the congenital [12], metabolic [13], and neoplastic alterations of these genes [14,15]. Based on studies demonstrating the differences in *HOX* gene expression between normal and neoplastic tissues, *HOX* genes may play roles in promoting cancer. Abnormalities of *homeobox* gene expression have been identified in many primary tumors [16], including those of the kidney, colon and even SCLC [17–19]. There is also considerable evidence that the deregulation of *HOX* expression is associated with human acute myeloid leukemia [20–22]. Several *HOX* genes are the targets of chromosomal translocations in leukemia and are thought to be potential oncogenes [23–25]. Thus, the dysregulation of *HOX* genes might be directly related to their abnormal functions in cancers.

Human *HOX* gene clusters harbor non-protein coding genes, the transcription of which yields, besides evolutionarily conserved micro-RNAs [26], numerous lncRNAs [27]. The non-protein-coding RNAs within the *HOX* clusters play important roles in tumor pathogenesis and progression through regulating the *HOX* gene network [28]. In humans,

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Table 1
HOX cluster-embedded antisense lncRNA genes and relevant cancers/tumors reported in previous studies.

| HOX cluster | Antisense RNA gene | HGNC ID | Aliases | Affected cancers/tumors |
|-----------------|--------------------|-----------------|---|--|
| HOXA | <i>HOTAIRM1</i> | 37117 | <i>HOXA-AS1, HOXA1-AS1, NCRNA00179</i> | Acute myeloid leukemia [31–34], colorectal cancer [35], head and neck tumors [36], hepatocellular carcinoma [37], lung cancer [30] |
| | <i>HOXA-AS2</i> | 43745 | <i>HOXA3-AS</i> | Breast, colorectal, gallbladder carcinoma, gastric and pancreatic cancers, hepatocellular carcinoma, malignant glioma, and promyelocytic leukemia (reviewed in Ref. [95]) |
| | <i>HOXA-AS3</i> | 43748 | <i>HOXA6-AS</i> | Glioma [38], lung adenocarcinoma [39] |
| | <i>HOXA10-AS</i> | 40281 | <i>HOXA-AS4</i> | Lung adenocarcinoma [40] |
| | <i>HOXA11-AS</i> | 24957 | <i>HOXA-AS5, NCRNA00076</i> | Breast, cervical, colorectal, gastric and ovarian cancers, glioblastoma, glioma, hepatocellular carcinoma, osteosarcoma, uveal melanoma, and NSCLC (reviewed in Ref. [41]), laryngeal squamous cell carcinoma [42], renal cancer [43] |
| HOXB | <i>HOTTIP</i> | 37461 | <i>HOXA-AS6, HOXA13-AS1, NCRNA00213</i> | Colorectal, gastric, lung, pancreatic and prostate cancers, hepatocellular carcinoma, and osteosarcoma (reviewed in Ref. [49]), breast cancer [50,51], endometrial cancer [52], esophageal squamous carcinoma [53,54], glioma [55,56], periamputary region tumors [57], renal cell carcinoma [58], thyroid carcinoma [59], tongue squamous cell carcinoma [60] |
| | <i>HOXB-AS1</i> | 43744 | <i>HOXB3-AS</i> | Unknown |
| | <i>HOXB-AS2</i> | 40284 | | Unknown |
| | <i>HOXB-AS3</i> | 40283 | | Colon Cancer [96] |
| | <i>HOXB-AS4</i> | 40285 | | Unknown |
| | <i>PRAC2</i> | 30143 | <i>C17orf93, HOXB-AS5, HOXB13-AS1, NARNA00253</i> | Breast cancer [97], prostate cancer [98] |
| | HOXC | <i>HOXC-AS1</i> | 43749 | |
| <i>HOXC-AS2</i> | | 43750 | | Unknown |
| <i>HOXC-AS3</i> | | 43751 | | Unknown |
| <i>HOTAIR</i> | | 33510 | <i>HOXAS, HOXC-AS4, HOXC11-AS1, NCRNA00072</i> | Breast, cervical, colorectal, gastric, lung, head and neck, ovarian and prostate cancers, endometrial carcinoma, esophageal and papillary thyroid carcinoma, glioblastoma, hematologic malignancies, hepatocellular carcinoma, osteosarcoma, and renal carcinoma (reviewed in Ref. [66]) |
| HOXD | <i>HOXC13-AS</i> | 43753 | <i>HOXC-AS5</i> | Unknown |
| | <i>HAGLR</i> | 43755 | <i>HOXD-AS1, Mdgt</i> | Bladder, cervical, colorectal, gastric, liver, ovarian and prostate cancers, glioma, melanoma, neuroblastoma, osteosarcoma, and NSCLC (reviewed in Ref. [80]) |
| | <i>HOXD-AS2</i> | 43756 | | Unknown |

HGNC, HUGO Gene Nomenclature Committee (<https://www.genenames.org/>); NSCLC, non-small cell lung cancer.

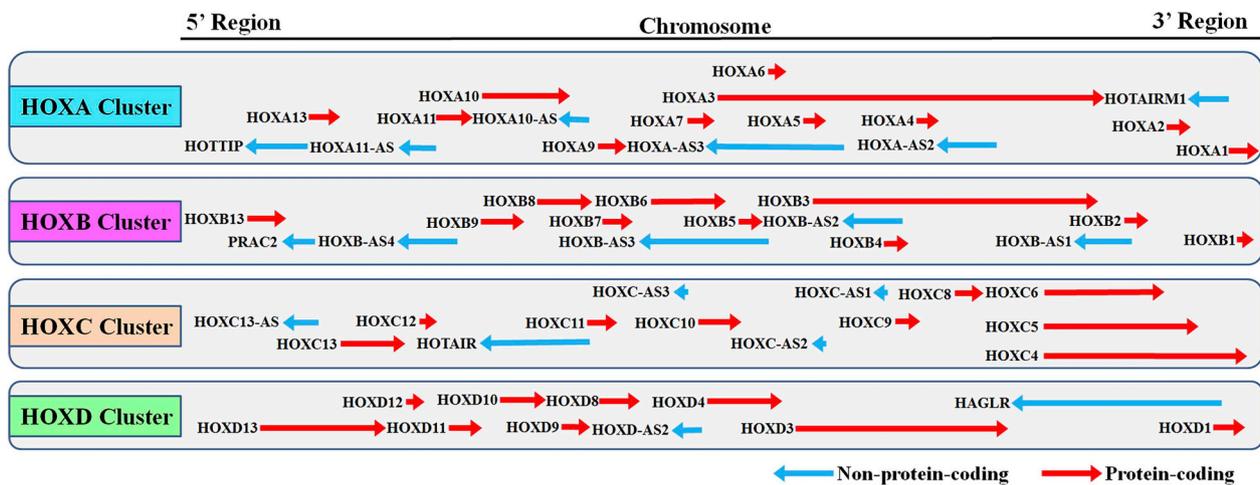


Fig. 1. Diagrams of the locations of antisense lncRNAs and *HOX* genes in the *HOX* clusters.

18 antisense RNA genes are found in the NCBI GeneBank database within the 4 *HOX* gene clusters. The *HOXA* region contains six *HOXA* antisense RNA genes, the *HOXB* and *HOXC* regions have five each, and the *HOXD* region holds two (Table 1). The positions of the antisense RNA genes in *HOX* clusters and their adjacent *HOX* genes are shown in Fig. 1. Half of these antisense RNAs reported to be associated with the development of different cancers (Table 1). Herein, we review the role of *HOX* cluster-embedded antisense lncRNAs (*HOX*-lncRNAs) in lung cancer, focusing on the cellular biological functions, molecular mechanisms of action, and the aberrant expression of these RNAs, as well as delving into the clinical features accompanying the abnormal expression of these lncRNAs in lung cancer patients (Table 2). We also summarize the chemo-resistance related pathways and modification complex network of these lncRNAs in other types of cancers (Tables S1 and S2).

2. *HOX* cluster-embedded antisense lncRNAs in lung cancer

2.1. *HOTAIRM1*

The human *HOXA* transcript antisense RNA myeloid-specific 1 (*HOTAIRM1*) gene is located between *HOXA1* and *HOXA2* in the 3' region of the *HOXA* cluster. The lncRNA *HOTAIRM1* has been studied more in leukemia than in solid tumors, possibly because initial findings with it were associated with myelopoiesis [29]. Recently, Tian et al. reported that the expression of *HOTAIRM1* in myeloid-derived suppressor cells (MDSCs) from lung cancer tissues was significantly decreased compared with that from adjacent normal tissues and that the overexpression of *HOTAIRM1* inhibited the development of MDSCs [30]. Additionally, *HOTAIRM1* levels were determined to be lower in the peripheral blood of lung cancer patients than in those of control patients. *HOTAIRM1* was also found to be expressed chiefly in LUAD as compared to LUSC or SCLC. This study established that *HOTAIRM1* delayed tumor progression and promoted antitumor immune responses in lung cancer by downregulating the immunosuppressive activity of MDSCs via targeting of *HOXA1*. These results demonstrated that the *HOTAIRM1*/*HOXA1* axis downregulates the immunosuppressive function of MDSCs and may be a potential therapeutic target in lung cancers. In addition to lung cancer, *HOTAIRM1* has also been reported to be involved in the carcinogenesis and progression of acute myeloid leukemia [31–34], colorectal cancer [35], head and neck tumors [36], and hepatocellular carcinoma [37]. As a novel lncRNA, *HOTAIRM1* should be investigated in a larger cancer cohort, due to its prominent role in preventing tumor progression.

2.2. *HOXA-AS3*

HOXA cluster antisense RNA 3 (*HOXA-AS3*) is a novel lncRNA transcribed from the *HOXA* cluster; studies on this lncRNA are limited. Wu et al. reported that *HOXA-AS3* expression was upregulated in glioma tissues and in cell lines, and a high *HOXA-AS3* expression level was associated with a poor prognosis for patients with glioma [38]. In this study, *HOXA-AS3* was found to regulate cell-cycle progression, proliferation, apoptosis, and the migration of glioma cells. In the case of lung cancer, it has also been shown that *HOXA-AS3* is significantly overexpressed in LUAD tissues and in A549 cells [39]. Moreover, knockdown of *HOXA-AS3* has been found to inhibit cancer cell proliferation, migration, and invasion. This work also examined the mechanism of *HOXA-AS3* action. The authors found that it promoted the stability of *HOXA6* mRNA and became bound to NF110, which determined its sub-cellular location. Furthermore, upregulation of *HOXA-AS3* in A549 cells was related to histone acetylation. This study demonstrated the important role of *HOXA-AS3* in LUAD progression and indicated that *HOXA-AS3* might serve as a potential prognostic biomarker for LUAD.

2.3. *HOXA10-AS*

Another novel antisense lncRNA is *HOXA10-AS*, which has its gene located adjacent to *HOXA10* in the *HOXA* cluster. Only a single article thus far has described a role for *HOXA10-AS* in cancer, which was in LUAD [40]. That paper demonstrated that *HOXA10-AS* was upregulated in LUAD tissues and cells, and that the upregulation of its expression was correlated with poor prognosis in LUAD patients. Moreover, knockdown of *HOXA10-AS* inhibited cell proliferation and migration, and enhanced apoptosis in LUAD cells. Furthermore, it was shown that *HOXA10-AS* expression promoted LUAD progression by activating the Wnt/ β -Catenin signaling pathway. Therefore, these results indicate that *HOXA10-AS* might be a novel prognostic biomarker for LUAD, although further research is necessary to justify this.

2.4. *HOXA11-AS*

The *HOXA11* antisense RNA (*HOXA11-AS*) gene is located between *HOXA11* and *HOXA13* in the human genome, and plays a critical role in tumorigenesis and tumor progression [41–43]. Bioinformatics analysis has shown that *HOXA11-AS* may play pivotal roles in NSCLC development and progression by regulating the expression of various genes and signaling pathways [44]. Several studies have explored the mechanisms of *HOXA11-AS* action using functional experiments. Zhang et al. reported that the expression of *HOXA11-AS* was markedly

Table 2
Lung cancer-associated HOX cluster-embedded antisense lncRNAs.

| lncRNAs (HGNC ID) | Cancer subtypes | Expression in cancer | Associated clinical features | Cellular biological functions | Related molecules and pathways | REF |
|-------------------|-----------------|----------------------|--|--|--|---------|
| HOTAIRM1 (37117) | Lung cancer | Downregulated | Smoking history, lymph node metastasis, TNM stage, and histological tumor type | Inhibits the development of MDSCs and downregulates the immunosuppressive activity of MDSCs in lung cancer | HOXA1 | [30] |
| HOXA-AS3 (43748) | LUAD | Upregulated | | Promotes cell proliferation, migration, and invasion | HOXA6, NF110 and H3K9ac | [39] |
| HOXA10-AS (40281) | LUAD | Upregulated | Poor prognosis | Promotes cell proliferation and metastasis, and inhibits cell apoptosis | ELK1 and Wnt/ β -catenin signaling | [40] |
| HOXA11-AS (24957) | NSCLC | Upregulated | Tumor size, lymph node metastasis, advanced clinical stage, TNM stage, and poor prognosis | Promotes cell proliferation, migration, and invasion, tumorigenic and angiogenic ability and inhibits cell apoptosis | miR-124/Sp1; EZH2 and DNMT1/miR-200b; EMT signaling pathway; miR-642b-3p | [45–48] |
| HOTTIP (37461) | SCLC | Upregulated | Clinical stage, chemotherapy response, and poor prognosis of SCLC patients | Promotes cell proliferation, migration, cell cycle, drug-resistance and inhibits cell apoptosis | miR-574-5p/EZH1; HOXA13; miR-216a/BCL-2; AKT signaling pathway | [61–64] |
| HOTAIR* (33510) | SCLC | Upregulated | Histology subtype, TNM stage, clinical stage, lymphatic metastasis, and poor prognosis | Promotes cell proliferation, migration, invasion, and drug-resistance and inhibits cell apoptosis | p53 and H3K27me3, Cav-1; Bax/Caspase-3 and TGF- α /EGFR signaling; miR-613; 14-3-3 σ ; miR-326/Phox2a; miR-326/SP1 | [67–78] |
| HAGLR (43755) | NSCLC | Upregulated | Tumor size, stage, recurrence, and TNM stage, lymph node metastasis, and poor overall survival | Promotes cell proliferation, migration, invasion, and cell cycle progression, and suppresses cell apoptosis | p21 and MMP9; miR-133b/MMP9; miR-147a/pRB | [81–83] |

HGNC, HUGO Gene Nomenclature Committee (<https://www.genenames.org/>); LUAD, lung adenocarcinoma; MDSCs, myeloid-derived suppressor cells; NSCLC, non-small cell lung cancer; REF, references; SCLC, small cell lung cancer; TNM, tumor lymph node metastasis. *Data from recent studies.

increased in NSCLC tissues as well as in cell lines and that its high expression was correlated with an advanced clinical stage in NSCLC patients [45]. Moreover, knockdown of HOXA11-AS suppressed the proliferation, migration, invasion, tumorigenic, and angiogenic capabilities of NSCLC cells, as well as inducing apoptosis in these cells. Furthermore, inhibition of HOXA11-AS also led to a cell cycle arrest at the G0/G1 transition or at the G2/M phase checkpoint. In addition, Chen et al. found that high expression of HOXA11-AS in NSCLC tissues showed a correlation with lymph node metastasis, tumor node metastasis (TNM) stage, and poor prognosis [46]. Their results in NSCLC cells further indicated that the lncRNA HOXA11-AS promoted cell invasion, in addition to an epithelial to mesenchymal transition (EMT) process by suppressing miR-200b expression through an interaction with EZH2 and DNMT1. Following knockdown of HOXA11-AS in NSCLC cells, miR-642b-3p was also significantly downregulated [47]. Yu et al. confirmed that HOXA11-AS expression was upregulated in NSCLC tissues and in cell lines and that high expression levels of HOXA11-AS in patients were associated with larger tumor size and lymph node metastasis [48]. Analysis of its biological function demonstrated that HOXA11-AS promoted the proliferation and invasion of NSCLC cells by sequestering miR-124, effectively regulating SP1 expression. The lncRNA HOXA11-AS thus plays a pivotal role in NSCLC, and it may be a promising biomarker for the early detection and prognostic evaluation of NSCLC.

2.5. HOTTIP

The *HOXA transcript at the distal tip (HOTTIP)* gene is located at the 5' end of the HOXA cluster in the human genome and is transcribed into a functional lncRNA. The lncRNA HOTTIP has been reported to be dysregulated in various cancers, including lung cancer [49–60]. In SCLC, HOTTIP is upregulated in tissues and cells, and its expression is correlated with the clinical stage, survival and chemotherapy response of SCLC patients [61,62]. HOTTIP overexpression promotes SCLC cell proliferation and the progression of the cell cycle through the regulation of miR-574-5p/EZH1 [61]. In addition, HOTTIP regulates apoptosis and chemo-resistance of SCLC cells by binding to miR-216a, thereby enhancing the expression of BCL-2, which suppresses apoptosis [62]. In addition to SCLC, HOTTIP is also highly expressed in NSCLC tissues and cells, and overexpression of HOTTIP significantly promotes cell proliferation and migration. HOTTIP overexpression also inhibits cell apoptosis by suppressing the expression of HOXA13 [63]. Moreover, overexpression of HOTTIP promotes the drug resistance of LUAD by regulating the AKT signaling pathway [64]. These findings suggest that the lncRNA HOTTIP also may be a valuable biomarker for lung cancer progression.

2.6. HOTAIR

One of the most studied HOX cluster-embedded lncRNAs is the HOX transcript antisense RNA (HOTAIR); its gene resides between the *HOXC11* and *HOXC12* genes in the HOXC cluster on human chromosome 12q13.13 [65]. HOTAIR has been reported to play oncogenic roles in various cancers, including lung cancer [66]. Indeed, the functions of the lncRNA HOTAIR have been widely studied in lung cancer; a review [10] has summarized the results from those studies. HOTAIR expression was significantly upregulated in cell lines, tissues and even the plasma of NSCLC patients [67–72], and this upregulation of HOTAIR showed a correlation with histology subtype, high TNM and clinical stage, lymphatic metastasis, and poor prognosis in NSCLC patients [69,73]. Functionally, HOTAIR affected tumorigenesis and progression in NSCLC by enhancing the proliferation, migration, and invasion of NSCLC cells, and inhibiting apoptosis in NSCLC cells [68,70,73]. Mechanistically, HOTAIR promoted the expression of 14-3-3 σ [71] and negatively regulated the expression of p53 through enhancing H3K27me3 levels at the p53 promoter [67] in NSCLC.

Moreover, the expression of HOTAIR was increased by CAV-1 [68], and knockdown of HOTAIR expression increased the expression of miR-613 [70] and miR-326/Phox2a [74] to induce oncogenesis and tumor progression. The ratio of FOXA1 to FOXA2 was also increased, which at a moderate level is involved in carcinogenesis and disease evolution [73]. Thus, HOTAIR can be considered a promising biomarker for the diagnosis and monitoring of NSCLC.

In addition, aberrant expression of HOTAIR is also associated with drug-resistance in SCLC and NSCLC patients [72,75–78]. It has been reported that HOTAIR may mediate the chemo-resistance of SCLC cells by regulating the methylation of the *HOXA1* gene by affecting DNMT1 and DNMT3b protein expression [77]. In drug-resistant NSCLC patients, the expression of HOTAIR was also elevated, and overexpression of HOTAIR increased the resistance to Cisplatin in NSCLC cells [75]. The study demonstrated that the drug resistance induced by elevated HOTAIR expression might be caused by an upregulation in the expression of *Klf4*, a tumor stem cell biomarker. Liu et al. reported that the knockdown of HOTAIR restored Gefitinib sensitivity through activation of Bax/Caspase3 and suppression of TGF α /EGFR signaling in LUAD [78]. Silencing of HOTAIR expression was shown to decrease the resistance of NSCLC cells to Crizotinib by inactivating autophagy through suppression of the activation of the ULK1 pathway [72]. Silencing of the *HOTAIR* gene enhanced the sensitivity of LUAD cells to Cisplatin by increasing miR-326 expression and synergistically reducing SP1 expression [76]. In summary, the lncRNA HOTAIR may be a potential therapeutic target for chemo-resistance in SCLC and NSCLC.

2.7. HAGLR

The *HAGLR* (*HOXD antisense growth-associated long noncoding RNA*, also known as *HOXD-AS1* and *Mdgt*) gene is located in the intergenic region between *HOXD1* and *HOXD3*. It has been reported to play essential roles in gut development [79] and also in various cancers [80]. In NSCLC, HAGLR was upregulated in tissues and cells, and this upregulation was associated with large tumor size, advanced tumor stage, recurrence, lymph node metastasis, high TNM stage, and reduced overall survival of patients [81–83]. Moreover, HAGLR promoted NSCLC cell proliferation and cell cycle progression while suppressing cell apoptosis [81]; HAGLR stimulated NSCLC cell growth by targeting miR-147a/pRB. Lu et al. have reported that the knockdown of HAGLR suppressed cell invasion and decreased levels of fatty acid synthase in NSCLC cells [82]. They also showed that the free fatty acid content in cancer cells was decreased and the expression levels of the tumor suppressor p21 and a matrix metalloproteinase (MMP9) were dysregulated following inhibition of HAGLR. Additionally, it was demonstrated that HAGLR promoted NSCLC cell migration and invasion by regulating the miR-133b/MMP9 pathway [83]. In conclusion, these data suggested that HAGLR may be a novel prognostic biomarker and a promising therapeutic target for treating NSCLC.

3. Conclusions and future perspectives

LncRNAs, including the *HOX* cluster-embedded antisense lncRNAs, play a vital role in tumorigenesis and tumor progression by affecting multiple pathways. They are dysregulated in many different types of cancers, including solid tumors and hematologic malignancies. In lung cancer, the *HOX* cluster-embedded antisense lncRNAs participate in the initiation and progression of tumor formation and development of drug-resistance by regulating the expression of *HOX* genes or other genes (Fig. 2). In summary, HOTTIP regulates the growth of SCLC cells by modulating miR-574-5p/EZH1 and mediates chemo-resistance through miR-216a/BCL-2. HOTTIP promotes the proliferation and migration of NSCLC cells by suppressing *HOXA13* expression and is involved in drug resistance by regulating the AKT signaling pathway in NSCLC.

HOTAIRM1 enhances *HOXA1* expression to regulate the immunosuppressive activity of MDSCs in lung cancer. HOTAIR promotes tumorigenesis and metastasis in NSCLC by reducing the expression of miR-613. HOTAIR also regulates the growth and chemo-resistance of lung cancer cells by sponging miR-326. *HOXA10-AS* promotes lung cancer progression by increasing Wnt/ β -catenin signaling pathway. *HOXA11-AS* promotes proliferation and invasion of NSCLC cells by targeting miR-124/Sp1. *HOXA11-AS* promotes lung cancer progression by recruiting EZH2 and DNMT1 to suppress miR-200b expression and regulating EMT signaling pathway. *HOXA-AS3* promotes lung cancer cells proliferation through binding with NF110 to increase *HOXA6* expression. HAGLR promotes NSCLC progression by regulating miR-133b/MMP9 and miR-147a/pRB. Therefore, most *HOX* cluster-embedded antisense lncRNAs may serve as potential biomarkers and therapeutic targets for lung cancer.

In addition to lung cancer, HOTTIP, *HOXA11-AS*, HOTAIR, and HAGLR have been reported in many other common cancers. Fu et al. have reported that HOTTIP is upregulated in pancreatic cancer stem cells (PCSCs) [84], the same finding as in lung cancer. HOTTIP alterations affect the stem cell characteristics of PCSCs by modulating *HOXA9* to enhance the Wnt/ β -catenin pathway. In prostate cancer, TWIST1-WDR5-HOTTIP increases H3K4me3 chromatin at the *HOXA9* promoter to facilitate metastasis [85]. HOTTIP also regulates tumorigenesis of gastric cancer and esophageal squamous carcinoma through regulating *HOXA13* at the transcriptional and posttranscriptional level [53,86]. *HOXA11-AS* promotes the progression of hepatocellular carcinoma through repressing miR-214-3p [87]. In gastric cancer, *HOXA11-AS* promotes proliferation and invasion by scaffolding the chromatin modification factors PRC2, LSD1, and DNMT1 [88]. Moreover, overexpression of *HOXA11-AS* promotes cell cycle progression and metastasis in gastric cancer by interacting with WDR5 and promoting β -catenin transcription, and through binding with EZH2 and repressing p21 transcription, and *HOXA11-AS* induces *KLF2* mRNA degradation via interacting with STAU1 [89]. HOTAIR expression is high in ovarian cancer, and it regulates the expression of CCND1 and CCND2 by sponging miR-206 in cancer cells [90]. HOTAIR is a prognostic biomarker for the proliferation and chemo-resistance of colorectal cancer through miR-203a-3p-mediated Wnt/ β -catenin pathway [91]. The STAT3/HOTAIR signaling axis regulates head and neck squamous cell cancer growth with an EZH2-dependent manner [92]. HAGLR expression is increased in bladder cancer tissues and cells, and it regulates the progression of this cancer [93]. STAT3-mediated upregulation of HAGLR facilitates liver cancer metastasis through binding with miR-130a-3p and regulation of *SOX4* [94]. *HOXA-AS2* has been studied in many kinds of tumors, but not in lung cancer [95]. HOTAIRM1, *HOXA-AS3*, *HOXA10-AS*, *HOXB-AS3* [96] and *PRAC2* [97,98] have only been explored in a limited number of cancer types. Furthermore, the roles of about half of the *HOX*-lncRNAs in cancer are unclear, namely, *HOXB-AS1*, *HOXB-AS2*, *HOXB-AS4*, *HOXC-AS1*, *HOXC-AS2*, *HOXC-AS3*, *HOXC13-AS*, and *HOXD-AS2*. Therefore, an investigation of *HOX* cluster-embedded antisense lncRNAs in cancer is a promising area for future research.

Conflicts of interest

The authors declare that they do not have any conflicts of interest for this article.

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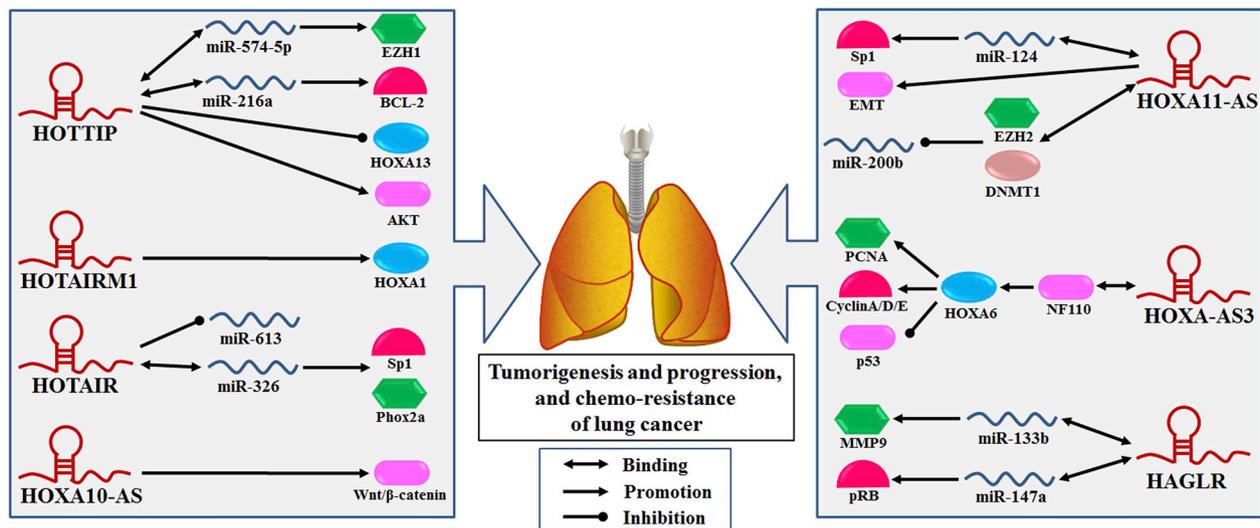


Fig. 2. The network of *HOX*-lncRNAs and other regulatory factors in lung cancer.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.canlet.2019.02.036>.

References

- J.L. Rinn, H.Y. Chang, Genome regulation by long noncoding RNAs, *Annu. Rev. Biochem.* 81 (2012) 145–166.
- K.C. Pang, M.C. Frith, J.S. Mattick, Rapid evolution of noncoding RNAs: lack of conservation does not mean lack of function, *Trends Genet.* 22 (2006) 1–5.
- K.C. Wang, H.Y. Chang, Molecular mechanisms of long noncoding RNAs, *Mol. Cell.* 43 (2011) 904–914.
- H. Zhang, Z. Chen, X. Wang, Z. Huang, Z. He, Y. Chen, Long non-coding RNA: a new player in cancer, *J. Hematol. Oncol.* 6 (2013) 37–43.
- J.R. Prensner, A.M. Chinnaiyan, The emergence of lncRNAs in cancer biology, *Cancer Discov.* 1 (2011) 391–407.
- G. Yang, X. Lu, L. Yuan, lncRNA: a link between RNA and cancer, *Biochim. Biophys. Acta* 1839 (2014) 1097–1109.
- R.L. Siegel, K.D. Miller, A. Jemal, Cancer statistics, *Ca - Cancer J. Clin.* 68 (2018) 7–30.
- R.S. Herbst, J.V. Heymach, S.M. Lippman, Lung cancer, *N. Engl. J. Med.* 359 (2008) 1367–1380.
- W.D. Travis, Pathology of lung cancer, *Clin. Chest Med.* 23 (2002) 65–81.
- G. Loewen, J. Jayawickramarajah, Y. Zhuo, B. Shan, Functions of lncRNA HOTAIR in lung cancer, *J. Hematol. Oncol.* 7 (2014) 90–99.
- F. Apiou, D. Flagiello, C. Cillo, B. Malfoy, M.F. Poupon, B. Dutrillaux, Fine mapping of human HOX gene clusters, *Cytogenet. Cell Genet.* 73 (1996) 114–115.
- D.P. Mortlock, L.C. Post, J.W. Innis, The molecular basis of hypodactyly (Hd): a deletion in Hoxa 13 leads to arrest of digital arch formation, *Nat. Genet.* 13 (1996) 284–289.
- A. Procino, C. Cillo, The HOX genes network in metabolic diseases, *Cell Biol. Int.* 37 (2013) 1145–1148.
- M. Cantile, R. Franco, A. Tschan, D. Baumhoer, I. Zlobec, G. Schiavo, I. Forte, M. Bihl, G. Liguori, G. Botti, L. Tornillo, E. Karamitopoulou-Diamantis, L. Terracciano, C. Cillo, HOX D13 expression across 79 tumor tissue types, *Int. J. Cancer* 125 (2009) 1532–1541.
- S. Bhatlekar, J.Z. Fields, B.M. Boman, HOX genes and their role in the development of human cancers, *J. Mol. Med. (Berl.)* 92 (2014) 811–823.
- D.G. Grier, A. Thompson, A. Kwasniewska, G.J. McGonigle, H.L. Halliday, T.R. Lappin, The pathophysiology of HOX genes and their role in cancer, *J. Pathol.* 205 (2005) 154–171.
- C. Cillo, P. Barba, G. Freschi, G. Bucciarelli, M.C. Magli, E. Boncinelli, HOX gene expression in normal and neoplastic human kidney, *Int. J. Cancer* 51 (1992) 892–897.
- G. De Vita, P. Barba, N. Odartchenko, J.C. Givel, G. Freschi, G. Bucciarelli, M.C. Magli, E. Boncinelli, C. Cillo, Expression of homeobox-containing genes in primary and metastatic colorectal cancer, *Eur. J. Cancer* 29a (1993) 887–893.
- C. Tiberio, P. Barba, M.C. Magli, F. Arvelo, T. Le Chevalier, M.F. Poupon, C. Cillo, HOX gene expression in human small-cell lung cancers xenografted into nude mice, *Int. J. Cancer* 58 (1994) 608–615.
- H.A. Drabkin, C. Parsy, K. Ferguson, F. Guilhot, L. Lacotte, L. Roy, C. Zeng, A. Baron, S.P. Hunger, M. Varella-Garcia, R. Gemmill, F. Brizard, A. Brizard, J. Roche, Quantitative HOX expression in chromosomally defined subsets of acute myelogenous leukemia, *Leukemia* 16 (2002) 186–195.
- S. Debernardi, D.M. Lillington, T. Chaplin, S. Tomlinson, J. Amess, A. Rohatiner, T.A. Lister, B.D. Young, Genome-wide analysis of acute myeloid leukemia with normal karyotype reveals a unique pattern of homeobox gene expression distinct from those with translocation-mediated fusion events, *Genes Chromosomes Cancer* 37 (2003) 149–158.
- V.P. Rawat, S. Thoene, V.M. Naidu, N. Arseni, B. Heilmeyer, K. Metzler, K. Petropoulos, A. Deshpande, L. Quintanilla-Martinez, S.K. Bohlander, K. Spiekermann, W. Hiddemann, M. Feuring-Buske, C. Buske, Overexpression of CDX2 perturbs HOX gene expression in murine progenitors depending on its N-terminal domain and is closely correlated with deregulated HOX gene expression in human acute myeloid leukemia, *Blood* 111 (2008) 309–319.
- S.Z. Raza-Egilmez, S.N. Jani-Sait, M. Grossi, M.J. Higgins, T.B. Shows, P.D. Aplan, NUP98-HOXD13 gene fusion in therapy-related acute myelogenous leukemia, *Cancer Res.* 58 (1998) 4269–4273.
- T. Nakamura, Y. Yamazaki, Y. Hatano, I. Miura, NUP98 is fused to PMX1 homeobox gene in human acute myelogenous leukemia with chromosome translocation t(1;11)(q23;p15), *Blood* 94 (1999) 741–747.
- J. Borrow, A.M. Shearman, V.P. Stanton Jr., R. Becher, T. Collins, A.J. Williams, I. Dube, F. Katz, Y.L. Kwong, C. Morris, K. Ohyashiki, K. Toyama, J. Rowley, D.E. Housman, The t(7;11)(p15;p15) translocation in acute myeloid leukaemia fuses the gene for nucleoporin NUP98 and class I homeoprotein HOXA9, *Nat. Genet.* 12 (1996) 159–167.
- S. Fantini, V. Salsi, V. Zappavigna, HOX cluster-embedded micro-RNAs and cancer, *Biochim. Biophys. Acta* 1869 (2018) 230–247.
- J.L. Rinn, M. Kertesz, J.K. Wang, S.L. Squazzo, X. Xu, S.A. Brugmann, L.H. Goodnough, J.A. Helms, P.J. Farnham, E. Segal, H.Y. Chang, Functional demarcation of active and silent chromatin domains in human HOX loci by noncoding RNAs, *Cell* 129 (2007) 1311–1323.
- G. Botti, A. De Chiara, M. Di Bonito, M. Cerrone, M.G. Malzone, F. Collina, M. Cantile, Noncoding RNAs within the HOX gene network in tumor pathogenesis and progression, *234 (2018) 395–413.*
- X. Zhang, Z. Lian, C. Padden, M.B. Gerstein, J. Rozowsky, M. Snyder, T.R. Gingeras, P. Kapranov, S.M. Weissman, P.E. Newburger, A myelopoiesis-associated regulatory intergenic noncoding RNA transcript within the human HOXA cluster, *Blood* 113 (2009) 2526–2534.
- X. Tian, J. Ma, T. Wang, J. Tian, Y. Zhang, L. Mao, H. Xu, S. Wang, Long non-coding RNA HOXA transcript antisense RNA myeloid-specific 1-HOXA1 Axis down-regulates the immunosuppressive activity of myeloid-derived suppressor cells in lung cancer, *Front. Immunol.* 9 (2018) 473–484.
- Z.H. Chen, W.T. Wang, W. Huang, K. Fang, Y.M. Sun, S.R. Liu, X.Q. Luo, Y.Q. Chen, The lncRNA HOTAIRM1 regulates the degradation of PML-RARA oncoprotein and myeloid cell differentiation by enhancing the autophagy pathway, *Cell Death Differ.* 24 (2017) 212–224.
- S. Wei, M. Zhao, X. Wang, Y. Li, K. Wang, PU.1 controls the expression of long noncoding RNA HOTAIRM1 during granulocytic differentiation, *J. Hematol. Oncol.* 9 (2016) 44–52.
- M. Diaz-Beya, S. Brunet, J. Nomdedeu, M. Pratcorona, A. Cordeiro, D. Gallardo, L. Escoda, M. Tormo, I. Heras, J.M. Ribera, R. Duarte, M.P. de Llano, J. Bargay, A. Sampol, M. Nomdedeu, R.M. Risueno, M. Hoyos, J. Sierra, M. Monzo, A. Navarro, J. Esteve, The lincRNA HOTAIRM1, located in the HOXA genomic region, is expressed in acute myeloid leukemia, impacts prognosis in patients in the intermediate-risk cytogenetic category, and is associated with a distinctive microRNA signature, *Oncotarget* 6 (2015) 31613–31627.
- X. Zhang, S.M. Weissman, P.E. Newburger, Long intergenic non-coding RNA HOTAIRM1 regulates cell cycle progression during myeloid maturation in NB4 human promyelocytic leukemia cells, *RNA Biol.* 11 (2014) 777–787.
- L. Wan, J. Kong, J. Tang, Y. Wu, E. Xu, M. Lai, H. Zhang, HOTAIRM1 as a potential

- biomarker for diagnosis of colorectal cancer functions the role in the tumour suppressor, *J. Cell Mol. Med.* 20 (2016) 2036–2044.
- [36] M. Zheng, X. Liu, Q. Zhou, G. Liu, HOTAIRM1 competed endogenously with miR-148a to regulate DLGAP1 in head and neck tumor cells, *J. Cell. Physiol.* 154 (2018) 3143–3156.
- [37] Y. Zhang, L. Mi, Y. Xuan, C. Gao, Y.H. Wang, H.X. Ming, J. Liu, LncRNA HOTAIRM1 inhibits the progression of hepatocellular carcinoma by inhibiting the Wnt signaling pathway, *Eur. Rev. Med. Pharmacol. Sci.* 22 (2018) 4861–4868.
- [38] F. Wu, C. Zhang, J. Cai, F. Yang, T. Liang, X. Yan, H. Wang, W. Wang, J. Chen, T. Jiang, Upregulation of long noncoding RNA HOXA-AS3 promotes tumor progression and predicts poor prognosis in glioma, *Oncotarget* 8 (2017) 53110–53123.
- [39] H. Zhang, Y. Liu, L. Yan, M. Zhang, X. Yu, W. Du, S. Wang, Q. Li, H. Chen, Y. Zhang, H. Sun, Z. Tang, D. Zhu, Increased levels of the long noncoding RNA, HOXA-AS3, promote proliferation of A549 cells, *Cell Death Dis.* 9 (2018) 707–720.
- [40] K. Sheng, J. Lu, H. Zhao, ELK1-induced upregulation of lncRNA HOXA10-AS promotes lung adenocarcinoma progression by increasing Wnt/beta-catenin signaling, *Biochem. Biophys. Res. Commun.* 501 (2018) 612–618.
- [41] C.W. Lu, D.D. Zhou, T. Xie, J.L. Hao, O.P. Pant, C.B. Lu, X.F. Liu, HOXA11 antisense long noncoding RNA (HOXA11-AS): a promising lncRNA in human cancers, *Cancer Med* 7 (2018) 3792–3799.
- [42] L. Qu, M. Jin, L. Yang, C. Sun, P. Wang, Y. Li, L. Tian, M. Liu, Y. Sun, Expression of long non-coding RNA HOXA11-AS is correlated with progression of laryngeal squamous cell carcinoma, *Am J Transl Res* 10 (2018) 573–580.
- [43] F.Q. Zhang, J.Q. Zhang, J.J. Jin, C.Y. Yang, W.J. Zhang, H.M. Zhang, J.H. Zheng, Z.M. Weng, HOXA11-AS promotes the growth and invasion of renal cancer by sponging miR-146b-5p to upregulate MMP16 expression, *J. Cell. Physiol.* 233 (2018) 9611–9619.
- [44] Y. Zhang, R.Q. He, Y.W. Dang, X.L. Zhang, X. Wang, S.N. Huang, W.T. Huang, M.T. Jiang, X.N. Gan, Y. Xie, P. Li, D.Z. Luo, G. Chen, T.Q. Gan, Comprehensive analysis of the long noncoding RNA HOXA11-AS gene interaction regulatory network in NSCLC cells, *Cancer Cell Int.* 16 (2016) 89–108.
- [45] Y. Zhang, W.J. Chen, T.Q. Gan, X.L. Zhang, Z.C. Xie, Z.H. Ye, Y. Deng, Z.F. Wang, K.T. Cai, S.K. Li, D.Z. Luo, G. Chen, Clinical significance and effect of lncRNA HOXA11-AS in NSCLC: a study based on bioinformatics, *in vitro and in vivo verification*, *Sci. Rep.* 7 (2017) 5567–5584.
- [46] J.H. Chen, L.Y. Zhou, S. Xu, Y.L. Zheng, Y.F. Wan, C.P. Hu, Overexpression of lncRNA HOXA11-AS promotes cell epithelial-mesenchymal transition by repressing miR-200b in non-small cell lung cancer, *Cancer Cell Int.* 17 (2017) 64–74.
- [47] Y. Zhang, J. Luo, X. Wang, H.L. Wang, X.L. Zhang, T.Q. Gan, G. Chen, D.Z. Luo, A comprehensive analysis of the predicted targets of miR-642b-3p associated with the long non-coding RNA HOXA11-AS in NSCLC cells, *Oncol. Lett.* 15 (2018) 6147–6160.
- [48] W. Yu, W. Peng, H. Jiang, H. Sha, J. Li, LncRNA HOXA11-AS promotes proliferation and invasion by targeting miR-124 in human non-small cell lung cancer cells, *Tumour Biol* 39 (2017) 1–8.
- [49] Y. Lian, Z. Cai, H. Gong, S. Xue, D. Wu, K. Wang, HOTTIP: a critical oncogenic long non-coding RNA in human cancers, *Mol. Biosyst.* 12 (2016) 3247–3253.
- [50] Y. Sun, C. Zeng, S. Gan, H. Li, Y. Cheng, D. Chen, R. Li, W. Zhu, LncRNA HOTTIP-mediated HOXA11 expression promotes cell growth, migration and inhibits cell apoptosis in breast cancer, *Int. J. Mol. Sci.* 19 (2018) 472–483.
- [51] W. Gao, X.L. Wu, D.Z. Li, H.D. Liu, HOTTIP participates in mammary cancer by promoting cell proliferation via PI3K/AKT pathway, *Eur. Rev. Med. Pharmacol. Sci.* 22 (2018) 4181–4187.
- [52] Q. Guan, Q. Zhang, C. Zhang, Q. Liu, Q.L. Ren, HOTTIP regulates progression of endometrial cancer via activating PI3K/AKT pathway, *Eur. Rev. Med. Pharmacol. Sci.* 22 (2018) 3727–3733.
- [53] C. Lin, Y. Wang, Y. Wang, S. Zhang, L. Yu, C. Guo, H. Xu, Transcriptional and posttranscriptional regulation of HOXA13 by lncRNA HOTTIP facilitates tumorigenesis and metastasis in esophageal squamous carcinoma cells, *Oncogene* 36 (2017) 5392–5406.
- [54] X. Chen, H. Han, Y. Li, Q. Zhang, K. Mo, S. Chen, Upregulation of long noncoding RNA HOTTIP promotes metastasis of esophageal squamous cell carcinoma via induction of EMT, *Oncotarget* 7 (2016) 84480–84485.
- [55] S. Zhang, W. Wang, G. Liu, S. Xie, Q. Li, Y. Li, Z. Lin, Long non-coding RNA HOTTIP promotes hypoxia-induced epithelial-mesenchymal transition of malignant glioma by regulating the miR-101/ZEB1 axis, *Biomed. Pharmacother.* 95 (2017) 711–720.
- [56] L.M. Xu, L. Chen, F. Li, R. Zhang, Z.Y. Li, F.F. Chen, X.D. Jiang, Over-expression of the long non-coding RNA HOTTIP inhibits glioma cell growth by BRE, *J. Exp. Clin. Oncol. Res.* 35 (2016) 162–176.
- [57] O. Balcin, S. Ak Aksoy, B. Tunca, E. Kaya, U. Egeli, G. Tezcan, N. Ugras, G. Cecener, O. Isik, H.Z. Dunder, O. Yerci, Overexpression of the long noncoding RNA HomeoboxA transcript at the distal tip predicts poor prognosis in a KRAS-independent manner in periampullary region tumors, *Pancreas* 47 (2018) 213–220.
- [58] F. Peng, X. Shi, Y. Meng, B. Dong, G. Xu, T. Hou, Y. Shi, T. Liu, Long non-coding RNA HOTTIP is upregulated in renal cell carcinoma and regulates cell growth and apoptosis by epigenetically silencing of LATS2, *Biomed. Pharmacother.* 105 (2018) 1133–1140.
- [59] Q. Yuan, Y. Liu, Y. Fan, Z. Liu, X. Wang, M. Jia, Z. Geng, J. Zhang, X. Lu, LncRNA HOTTIP promotes papillary thyroid carcinoma cell proliferation, invasion and migration by regulating miR-637, *Int. J. Biochem. Cell Biol.* 98 (2018) 1–9.
- [60] H. Zhang, L. Zhao, Y.X. Wang, M. Xi, S.L. Liu, L.L. Luo, Long non-coding RNA HOTTIP is correlated with progression and prognosis in tongue squamous cell carcinoma, *Tumour Biol* 36 (2015) 8805–8809.
- [61] Y. Sun, Y. Zhou, Y. Bai, Q. Wang, J. Bao, Y. Luo, Y. Guo, L. Guo, A long non-coding RNA HOTTIP expression is associated with disease progression and predicts outcome in small cell lung cancer patients, *Mol. Canc.* 16 (2017) 162–176.
- [62] Y. Sun, B. Hu, Q. Wang, M. Ye, Q. Qiu, Y. Zhou, F. Zeng, X. Zhang, Y. Guo, L. Guo, Long non-coding RNA HOTTIP promotes BCL-2 expression and induces chemoresistance in small cell lung cancer by sponging miR-216a, *Int. J. Mol. Sci.* 9 (2018) 85–101.
- [63] Y. Sang, F. Zhou, D. Wang, X. Bi, X. Liu, Z. Hao, Q. Li, W. Zhang, Up-regulation of long non-coding HOTTIP functions as an oncogene by regulating HOXA13 in non-small cell lung cancer, *Am J Transl Res* 8 (2016) 2022–2032.
- [64] G.J. Zhang, W. Song, Y. Song, Overexpression of HOTTIP promotes proliferation and drug resistance of lung adenocarcinoma by regulating AKT signaling pathway, *Eur. Rev. Med. Pharmacol. Sci.* 21 (2017) 5683–5690.
- [65] C.J. Woo, R.E. Kingston, HOTAIR lifts noncoding RNAs to new levels, *Cell* 129 (2007) 1257–1259.
- [66] Q. Tang, S.S. Hann, HOTAIR: an oncogenic long non-coding RNA in human cancer, *Cell. Physiol. Biochem.* 47 (2018) 893–913.
- [67] N. Zhai, Y. Xia, R. Yin, J. Liu, F. Gao, A negative regulation loop of long noncoding RNA HOTAIR and p53 in non-small-cell lung cancer, *OncoTargets Ther.* 9 (2016) 5713–5720.
- [68] W. Liu, N.C. Yin, H. Liu, K.J. Nan, Cav-1 promote lung cancer cell proliferation and invasion through lncRNA HOTAIR, *Gene* 641 (2018) 335–340.
- [69] N. Li, Y. Wang, X. Liu, P. Luo, W. Jing, M. Zhu, J. Tu, Identification of circulating long noncoding RNA HOTAIR as a novel biomarker for diagnosis and monitoring of non-small cell lung cancer, *Technol. Canc. Res. Treat.* 16 (2017) 1060–1066.
- [70] C. Jiang, Y. Yang, Y. Yang, L. Guo, J. Huang, X. Liu, C. Wu, J. Zou, Long noncoding RNA (lncRNA) HOTAIR affects tumorigenesis and metastasis of non-small cell lung cancer by upregulating miR-613, *Oncol. Res.* 26 (2018) 725–734.
- [71] R. Wang, B. Yan, Z. Li, Y. Jiang, C. Mao, X. Wang, X. Zhou, Long non-coding RNA HOX transcript antisense RNA promotes expression of 14-3-3sigma in non-small cell lung cancer, *Exp. Ther. Med.* 14 (2017) 4503–4508.
- [72] Y. Yang, C. Jiang, Y. Yang, L. Guo, J. Huang, X. Liu, C. Wu, J. Zou, Silencing of lncRNA-HOTAIR decreases drug resistance of Non-Small Cell Lung Cancer cells by inactivating autophagy via suppressing the phosphorylation of ULK1, *Biochem. Biophys. Res. Commun.* 497 (2018) 1003–1010.
- [73] R. Wang, Y. Shi, L. Chen, Y. Jiang, C. Mao, B. Yan, S. Liu, B. Shan, Y. Tao, X. Wang, The ratio of FoxA1 to FoxA2 in lung adenocarcinoma is regulated by lncRNA HOTAIR and chromatin remodeling factor LSH, *Sci. Rep.* 5 (2015) 17826–17836.
- [74] R. Wang, X. Chen, T. Xu, R. Xia, L. Han, W. Chen, W. De, Y. Shu, MiR-326 regulates cell proliferation and migration in lung cancer by targeting phox2a and is regulated by HOTAIR, *Am. J. Cancer Res.* 6 (2016) 173–186.
- [75] M.Y. Liu, X.Q. Li, T.H. Gao, Y. Cui, N. Ma, Y. Zhou, G.J. Zhang, Elevated HOTAIR expression associated with cisplatin resistance in non-small cell lung cancer patients, *J. Thorac. Dis.* 8 (2016) 3314–3322.
- [76] J. Li, S. Li, Z. Chen, J. Wang, Y. Chen, Z. Xu, M. Jin, W. Yu, miR-326 reverses chemoresistance in human lung adenocarcinoma cells by targeting specificity protein 1, *Tumour Biol* 37 (2016) 13287–13294.
- [77] S. Fang, H. Gao, Y. Tong, J. Yang, R. Tang, Y. Niu, M. Li, L. Guo, Long noncoding RNA-HOTAIR affects chemoresistance by regulating HOXA1 methylation in small cell lung cancer cells, *Lab. Invest.* 96 (2016) 60–68.
- [78] Y. Liu, H. Jiang, H. Zhou, X. Ying, Z. Wang, Y. Wang, W. Xu, X. He, Y. Li, Lentivirus-mediated silencing of HOTAIR lncRNA restores gefitinib sensitivity by activating Bax/Caspase-3 and suppressing TGF-alpha/EGFR signaling in lung adenocarcinoma, *Oncol. Lett.* 15 (2018) 2829–2838.
- [79] J. Zakany, F. Darbellay, B. Mascrez, A. Necsculea, D. Duboule, Control of growth and gut maturation by HoxD genes and the associated lncRNA Haglr, *Proc. Natl. Acad. Sci. U. S. A.* 114 (2017) E9290–E9299.
- [80] L. Li, Y. Wang, X. Zhang, Q. Huang, Y. Diao, H. Yin, H. Liu, Long non-coding RNA HOXD-AS1 in cancer, *Clin. Chim. Acta* 487 (2018) 197–201.
- [81] Q. Wang, S. Jiang, A. Song, S. Hou, Q. Wu, L. Qi, X. Gao, HOXD-AS1 functions as an oncogenic ceRNA to promote NSCLC cell progression by sequestering miR-147a, *OncoTargets Ther.* 10 (2017) 4753–4763.
- [82] C. Lu, J. Ma, D. Cai, Increased HAGLR expression promotes non-small cell lung cancer proliferation and invasion via enhanced de novo lipogenesis, *Tumour Biol* 39 (2017) 1–9.
- [83] H. Xia, H. Jing, Y. Li, X. Lv, Long noncoding RNA HOXD-AS1 promotes non-small cell lung cancer migration and invasion through regulating miR-133b/MMP9 axis, *Biomed. Pharmacother.* 106 (2018) 156–162.
- [84] Z. Fu, C. Chen, Q. Zhou, Y. Wang, Y. Zhao, X. Zhao, W. Li, S. Zheng, H. Ye, L. Wang, Z. He, Q. Lin, Z. Li, R. Chen, LncRNA HOTTIP modulates cancer stem cell properties in human pancreatic cancer by regulating HOXA9, *Cancer Lett.* 410 (2017) 68–81.
- [85] R. Malek, R.P. Gajula, R.D. Williams, B. Nghiem, B.W. Simons, K. Nugent, H. Wang, K. Taparra, G. Lemtiri-Chlieh, A.R. Yoon, L. True, S.S. An, T.L. DeWeese, A.E. Ross, E.M. Schaeffer, K.J. Pienta, P.J. Hurley, C. Morrissey, P.T. Tran, TWIST1-WDR5-Hottip regulates Hoxa9 chromatin to facilitate prostate cancer metastasis, *Cancer Res.* 77 (2017) 3181–3193.
- [86] D.C. Wu, S.S.W. Wang, C.J. Liu, K. Wuputra, K. Kato, Y.L. Lee, Y.C. Lin, M.H. Tsai, C.C. Ku, W.H. Lin, S.W. Wang, S. Kishikawa, M. Noguchi, C.C. Wu, Y.T. Chen, C.Y. Chai, C.S. Lin, K.K. Kuo, Y.H. Yang, H. Miyoshi, Y. Nakamura, S. Saito, K. Nagata, C.S. Lin, K.K. Yokoyama, Reprogramming antagonizes the oncogenicity of HOXA13-long noncoding RNA HOTTIP, *Axis in Gastric Cancer Cells* 35 (2017) 2115–2128.
- [87] M. Zhan, K. He, J. Xiao, F. Liu, H. Wang, Z. Xia, X. Duan, R. Huang, Y. Li, X. He, H. Yin, G. Xiang, LncRNA HOXA11-AS promotes hepatocellular carcinoma progression by repressing miR-214-3p, *J. Cell Mol. Med.* 22 (2018) 3758–3767.
- [88] M. Sun, F. Nie, Y. Wang, Z. Zhang, J. Hou, D. He, M. Xie, L. Xu, W. De, Z. Wang, J. Wang, LncRNA HOXA11-AS promotes proliferation and invasion of gastric cancer by scaffolding the chromatin modification factors PRC2, LSD1, and DNMT1, *Cancer Res.* 76 (2016) 6299–6310.
- [89] Z. Liu, Z. Chen, R. Fan, B. Jiang, X. Chen, Q. Chen, F. Nie, K. Lu, M. Sun, Over-

- expressed long noncoding RNA HOXA11-AS promotes cell cycle progression and metastasis in gastric cancer, *Mol. Canc.* 16 (2017) 82–90.
- [90] L. Chang, R. Guo, Z. Yuan, H. Shi, D. Zhang, LncRNA HOTAIR regulates CCND1 and CCND2 expression by sponging miR-206 in ovarian cancer, *Cell. Physiol. Biochem.* 49 (2018) 1289–1303.
- [91] Z. Xiao, Z. Qu, Z. Chen, Z. Fang, K. Zhou, Z. Huang, X. Guo, Y. Zhang, LncRNA HOTAIR is a prognostic biomarker for the proliferation and chemoresistance of colorectal cancer via MiR-203a-3p-mediated Wnt/ss-catenin signaling pathway, *Cell. Physiol. Biochem.* 46 (2018) 1275–1285.
- [92] S. Sun, Y. Wu, W. Guo, F. Yu, L. Kong, Y. Ren, Y. Wang, X. Yao, C. Jing, C. Zhang, M. Liu, Y. Zhang, M. Zhao, Z. Li, C. Wu, Y. Qiao, J. Yang, STAT3/HOTAIR signaling axis regulates HNSCC growth in an EZH2-dependent manner, 24 (2018) 2665–2677.
- [93] J. Li, C. Zhuang, Y. Liu, M. Chen, Y. Chen, Z. Chen, A. He, J. Lin, Y. Zhan, L. Liu, W. Xu, G. Zhao, Y. Guo, H. Wu, Z. Cai, W. Huang, Synthetic tetracycline-controllable shRNA targeting long non-coding RNA HOXD-AS1 inhibits the progression of bladder cancer, *J. Exp. Clin. Canc. Res.* 35 (2016) 99–109.
- [94] H. Wang, X. Huo, X.R. Yang, J. He, L. Cheng, N. Wang, X. Deng, H. Jin, N. Wang, C. Wang, F. Zhao, J. Fang, M. Yao, J. Fan, W. Qin, STAT3-mediated upregulation of lncRNA HOXD-AS1 as a ceRNA facilitates liver cancer metastasis by regulating SOX4, *Mol. Canc.* 16 (2017) 136–150.
- [95] J. Wang, Z. Su, S. Lu, W. Fu, Z. Liu, X. Jiang, S. Tai, LncRNA HOXA-AS2 and its molecular mechanisms in human cancer, *Clin. Chim. Acta* 485 (2018) 229–233.
- [96] J.Z. Huang, M. Chen, Chen, X.C. Gao, S. Zhu, H. Huang, M. Hu, H. Zhu, G.R. Yan, A peptide encoded by a putative lncRNA HOXB-AS3 suppresses colon cancer growth, *Mol. Cell.* 68 (2017) 171–184.
- [97] J. Rui, Z. Chunming, G. Binbin, S. Na, W. Shengxi, S. Wei, IL-22 promotes the progression of breast cancer through regulating HOXB-AS5, *Oncotarget* 8 (2017) 103601–103612.
- [98] P. Olsson, A. Motegi, T.K. Bera, B. Lee, I. Pastan, PRAC2: a new gene expressed in human prostate and prostate cancer, *Prostate* 56 (2003) 123–130.