

Review

Homologous recombination deficiency in triple negative breast cancer

Carmen Belli ^{a, *}, Bruno Achutti Duso ^a, Emanuela Ferraro ^a, Giuseppe Curigliano ^{a, b}^a Division of Early Drug Development for Innovative Therapies, European Institute of Oncology IRCCS, Milan, via Ripamonti 435, 20141, Milan, Italy^b Department of Oncology and Hemato-Oncology, University of Milan, via Festa del Perdono 7, 20122, Milan, Italy

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ABSTRACT

Triple negative breast cancer (TNBC) represents a heterogeneous subtype of breast cancer characterized by an unfavorable prognosis due to its aggressive biology. The median overall survival (OS) for patients with metastatic TNBC is around 9–12 months with conventional cytotoxic agents. Considering this suboptimal outcome, which is induced despite of medical treatment, new therapeutic strategies would be urgently needed. The ultimate goal of precision medicine is to identify specific molecular alterations that permit considering effective targeted drug(s). Germline BRCA mutations occur in 10–20% of TNBC patients while somatic mutations occur in 3–5% of them. Alterations in the homologous recombination (HR) system are typical of BRCA mutant tumors, but can also be identified in tumors that do not carry this mutation, defining a subgroup of patients referred to as BRCAness. In this review, we focus on the role of homologous recombination deficiency (HRD) as both predictive and prognostic factor in different settings of TNBC patients treated with DNA damaging drugs and poly ADP ribose polymerase (PARP) inhibitors.

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1. Introduction

TNBC is an aggressive tumor accounting for 15% of breast cancers and conventionally defined by the absence of estrogen (ER), progesterone (PR) and human epidermal growth factor 2 (HER2) receptors. TNBC includes a heterogeneous group of tumors, typically occurring in very young patients and which metastasizes

commonly by hematogenous spread, having lung and brain as major first sites of metastasis. It usually shows a peak of recurrence during the first three years, with the majority of deaths occurring within the first 5 years [1,2]. Data from The Cancer Genome Atlas (TCGA) and smaller prospective registries identified germline BRCA1/2 deleterious mutations in 10–20% of TNBC patients while somatic mutation in 3–5% of them [3,4].

Tumors with BRCA1/2 mutations typically carry a deficient HR system, cardinal for DNA repair from an insult [5]. DNA is exposed to continuous damage, causing a range of lesions varying from single-strand break (SSB), double-strand break (DSB), bulky

* Corresponding author.

E-mail addresses: carmen.belli@ieo.it, cbelli77@gmail.com (C. Belli).

adducts, base mismatches, insertions, deletions and base alkylation. The type of lesion present on DNA defines the repair mechanism of choice. In presence of DSB, the HR system removes the DSB and uses a homologous DNA template to repair it [6]. This mechanism acts in phase S and G2 of the cell cycle and requires the involvement of proteins encoded by the BRCA1, BRCA2, RAD51 and PALB2 genes. Mutations in one of these key components of DNA repair system are responsible for limited DNA repair capacity [7–11]. Tumors harboring mutations in one of these genes, for example, result more sensible to platinum agents or PARP inhibitors, since these compounds cause DNA inter- and intra-strand crosslinks that can not be recognized and subsequently repaired by HR system, ultimately resulting as defective [12]. Fig. 1 summarizes the key players and their roles in generating the phenotype of HR deficient tumors.

In this review, we focused on the contribution of HRD system as predictive and prognostic factor evaluated in different settings of TNBC patients.

2. BRCA1/2 mutation and homologous recombination (HR) system

Tumors defined as BRCA1/2 deficient show large regions of loss of heterozygosity (LOH), telomeric allelic imbalance (TAI), and

large-scale transition (LST) [13]. These three characteristics combined together define the HRD score, which is useful to define the HR deficient status [14–16]. The test used for HRD determination hereon was based on single nucleotide polymorphism (SNP) analysis using two main assays: Myriad genetics and myChoice HRD [17,18]. The first test was analyzed in a retrospective study in TNBC treated with several neoadjuvant regimes [17]. This analysis showed that HRD positive patients were more likely to present a pathological complete response (pCR) than those without this alteration (44% versus 8%; $p < 0.01$). The second study analyzed the myChoice assay in 48 TNBC patients treated with carboplatin plus nanoparticle albumin-bound paclitaxel with or without vorinostat [18]. Also here the patients with higher HRD score showed a higher pCR rate than HRD low ones (50% versus 7.7%; $p = 0.002$). Although each individual metric is significantly associated with BRCA1/2 status, the combination of the three performed best at distinguishing HRD from non-HRD tumors [13,19]. Alterations in the HR system, while typical of BRCA mutant tumors, can be also identified in tumors not carrying this mutation, defining a subgroup of patients referred to as BRCAness. Telli et al. [20] evaluated in a retrospective trial the combined HRD score, defined as the unweighted numeric sum of LOH, TAI, and LST, and tested the predictive power of a specific HRD threshold. The score was evaluated in a chemonaïve training cohort including 497 TNBC, of

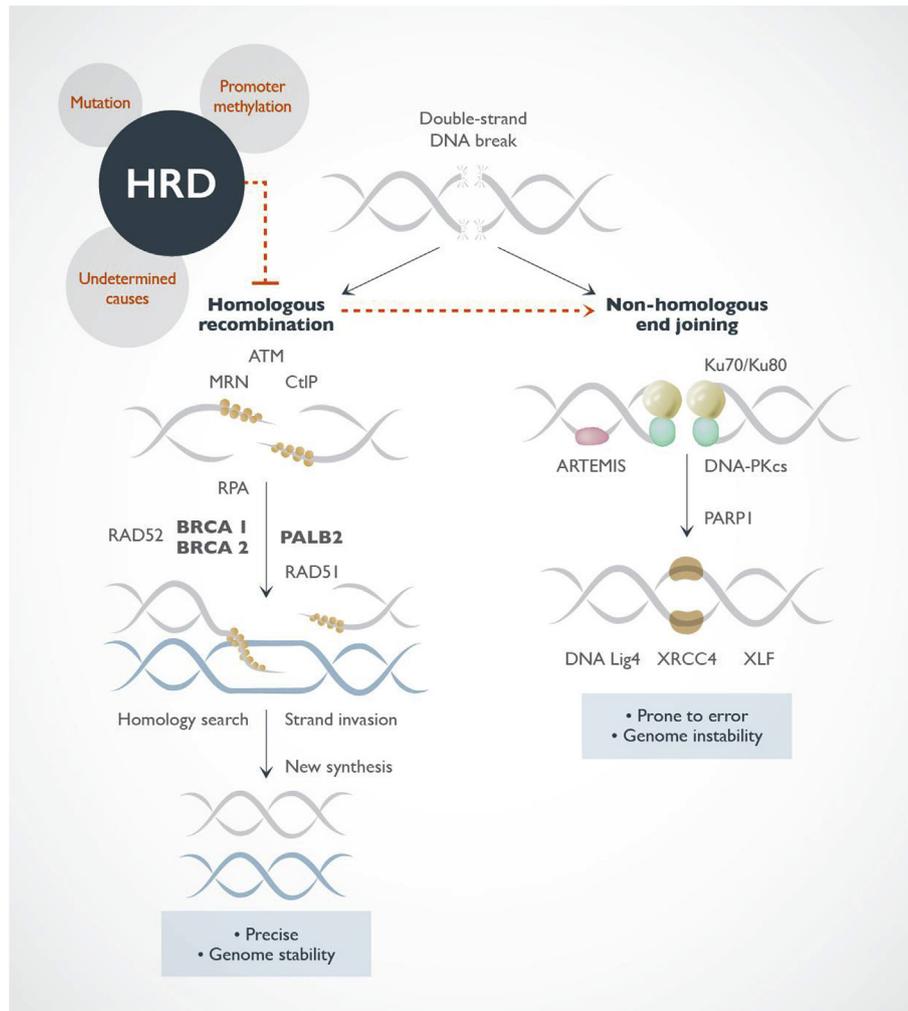


Fig. 1. Representation of how HRD pushes the DNA repair mechanism so as to engage non-homologous pathways, more prone to error and to genome instability.

which 78 had a BRCA1/2 mutation(s); and 561 ovarian tumors, of which 190 were BRCA mutated. BRCA1/2 mutants showed either one deleterious mutation in BRCA1/2 with LOH in the wild-type copy, or two deleterious mutations in the same gene or promoter methylation of BRCA1 with LOH in the wild-type copy. For an optimal accuracy, the threshold for the combined HRD score in the training set of BRCA intact and BRCA deficient subjects was established as ≥ 42 for both breast and ovarian tumors. HRD status was defined as positive by the presence of either BRCA1/2 mutation or a predefined high HRD score (≥ 42). HRD was classified as negative if no mutations in BRCA genes were detected or if a low HRD score (< 42) was calculated. The validation set included patients enrolled in PrECOG 0105, Cisplatin-1 and Cisplatin-2 studies in which HR score and HRD status were correlated to chemotherapy response [21–23].

The PrECOG 0105 trial was a phase II study that enrolled 93 patients affected by stage I-IIIa TNBC and/or BRCA1/2 germline mutants treated with 4 or 6 cycles of neoadjuvant carboplatin, gemcitabine, and iniparib [21]. The Cisplatin-1 and -2 also analyzed this correlation in the neoadjuvant setting and consist of a 28 and a 51 prospective cohorts of patients, respectively, treated with cisplatin as monotherapy for 4 cycles in the first study and cisplatin plus bevacizumab in the latter [22,23]. In these trials, the HR score - previously calculated in training set - and the HRD status were correlated with response to cisplatin by evaluating the residual cancer burden (RCB) index [24]. Four kinds of response to neoadjuvant treatment were described (RCB 0: pCR, RCB I: minimal residual disease, RCB II: moderate residual disease, and RCB III: extensive residual disease). For each variable, two dichotomous measures for tumor response were used (RCB 0/I: present or not present, and pCR: yes versus no). RCB 0/I yes includes tumors with pCR or RCB I. RCB 0/I no corresponds to class RCB II or RCB III. In the PrECOG 0105 study, the HRD score (high/low) was significantly associated with both RCB 0/I ($p = 0.0053$) and pCR ($p = 0.0065$). In the Cisplatin trials, this correspondence was also evidenced by RCB 0/I ($p = 0.0014$) and pCR ($p = 0.0071$). According to the PrECOG 0105, RCB 0/I rates in HRD tumors were 68% versus 30% in non-HRD (OR 4.96 CI 1.61–15.3; $p = 0.0036$). pCR rates were 42% in HR deficient tumors versus 10% in non-HRD (OR = 6.52, 95% CI, 1.36–31.2; $p = 0.0058$). In the Cisplatin-1 and -2 trials, RCB 0/I rate in HRD tumors were 51.7% versus 9.5% in non-HRD ones (OR 10.18, 95% CI 2.00–51.89; $p = 0.0011$). pCR rates were 27.5% in HR deficient and 0% in HR non-deficient tumors (OR 17.00 CI 1.91–2249; $p = 0.0066$). Additionally, in the PrECOG 0105, RCB 0/I rates were higher in BRCA1/2 mutant tumors than wild types (75% versus 48%; $p = 0.0037$), also with higher pCR rates (50% versus 24%; $p = 0.040$). In the cisplatin trials, probably because the number of mutant tumors was small, BRCA mutation was not significantly associated with RCB 0/I (42.9% versus 31.6%; $p = 0.57$) and pCR rates (28.6% vs 13.2%; $p = 0.33$). Rates of RCB 0/I were also evaluated according to the BRCA status in the PrECOG 0105, with 81% versus 47% for BRCA1/2 germline mutants when compared to wild types, respectively.

Using the HRD score as a stratifier tool, RCB 0/I rates were 59% in HRD high (≥ 42) and 32% in HRD low ($p = 0.062$) according to the retrospective analysis from the PrECOG 0105 population; and the pCR was 33% in HRD high versus 11% in HRD low tumors ($p = 0.063$). In the Cisplatin trials, a high HRD score was significantly associated with both higher RCB 0/I (52.6% versus 10.5% $p = 0.0039$) and pCR rates (26.3% vs 0% $p = 0.018$) when compared to the HRD low subgroup. The multivariate logistic analysis performed on all three above mentioned trials confirmed that the HRD was an independent predictor of RCB 0/I and pCR after adjustment for other clinical covariates.

3. HRD in the adjuvant setting

A study conducted by Sharma et al. evaluated the prognostic role of HRD status, HRD score, and BRCA1 promoter methylation in 425 patients affected by TNBC, comprised either by high risk/node-negative or low-risk/node positive disease and treated with two equivalent dose schedules of doxorubicin and cyclophosphamide [25]. HRD status was positive in 67% of patients, from which 27% had BRCA1/2 mutations. Positive HRD status was associated with a prolonged disease free survival (DFS) (HR 0.72, 95% CI 0.51–1.00; $p = 0.049$) and a non-significant trend toward an improvement in OS (HR 0.71, 95% CI 0.48–1.03; $p = 0.073$) after adjustment for treatment arms and nodal status. The positive HRD status correlation was particularly pronounced during the first 5 years. After this time, the impact on DFS and OS was not significant (HR for DFS 1.21, 95% CI 0.65–2.28; $p = 0.55$ and HR for OS 0.85, 95% CI 0.47–1.53; $p = 0.59$) [20]. Among patients with BRCA1/2 wild type tumors, 40% had an HRD score of ≥ 42 and it was associated with a higher DFS (HR 0.64, 95% CI 0.43–0.94; $p = 0.023$) as well as OS (HR 0.65, 95% CI 0.47–1.53; $p = 0.59$) when adjusted for treatment and nodal status. Tumor mutational status per se did not impact on DFS ($p = 0.59$) nor OS ($p = 0.9$) after adjustment for treatment and nodal status, probably due to the modest number of patients carrying this alteration. This study underlines the role of HRD positive status in decision making process of patients affected by TNBC, confirming its prognostic role independently from other patient- or tumor-related features. However, no data are available yet to define if this particular category of patients could benefit from specific cytotoxic agents such as taxanes or platinum salts.

4. HRD in the neoadjuvant setting

The phase III neoadjuvant trial BrightTNess randomized clinical stage II-III TNBC to receive a standard neoadjuvant chemotherapy with paclitaxel followed by anthracyclines and cyclophosphamides with or without carboplatin and with or without veliparib [26]. In an analysis conducted on global population independently from BRCA status, veliparib did not provide any benefit in comparison to chemotherapy. pCR, in fact, was significantly higher among patients receiving paclitaxel plus carboplatin plus veliparib than those treated with paclitaxel alone (53% versus 31%; $p < 0.0001$) but did not differ between patients receiving paclitaxel plus carboplatin plus veliparib versus paclitaxel plus carboplatin (53% versus 58%; $p = 0.36$). In addition, no difference in terms of RCB 0/I were evidenced among these two arms (68% versus 70%; $p = 0.739$). The presence of BRCA1/2 mutation was not predictive for pathological response, but the addition of carboplatin increased the rate of pCR (57% in the paclitaxel plus carboplatin plus veliparib group versus 50% in the paclitaxel plus carboplatin group versus 44% in the paclitaxel alone group) even if this study was not powered to show a significant difference in the attainment of pCR with the addition of carboplatin, with or without veliparib. This study has several limitation such as its retrospective nature, the small sample size and the inclusion of a few number of patients receiving taxanes. However, an important hypothesis arises from this trial that consists of the evaluation of HRD as a new biomarker in predicting the benefit of DNA damaging therapies such as cisplatin, anthracycline, taxane, and cyclophosphamide.

The phase II GeparSixto trial evaluated if the addition of carboplatin to a regimen of paclitaxel, low dose doxorubicin and bevacizumab in TNBC or trastuzumab and lapatinib in HER2 positive disease improved the pCR in stage II-III breast cancer [27]. The results of this trial showed that the addition of carboplatin to a regimen of chemotherapy and targeted therapies significantly increases the proportion of pCR in TNBC (53% versus 37%; $p = 0.005$)

but not in HER2 positive disease (32.8% versus 36.8%; $p = 0.581$) [28]. A subgroup analysis of the same study looked at the BRCA1/2 mutations and HRD status as predictive factors for pCR in patients receiving neoadjuvant chemotherapy. HRD, including the unweighted sum of LOH, TAI, and LST and BRCA1/2 mutations, was discovered on 70.5% of TNBC - 60.3% of tumor samples had HRD high without BRCA mutation. HRD high tumors, in its turn, were more likely to achieve a pCR than HRD low ones (55.9% versus 29.8%; $p = 0.001$). In addition, patients treated with carboplatin with a high HRD score showed significantly higher rates of pCR compared to patients treated without a platinum agent (64.9% versus 45.2%; $p = 0.025$).

Another retrospective analysis evaluated the role of HRD status and BRCA1/2 mutations in 45 patients with TNBC and 2 with hormone receptor-positive disease treated with neoadjuvant chemotherapy [29]. Chemotherapy regimens employed included anthracycline (19%), taxanes (9%), and a combination of these two drugs (70%). The mutation on BRCA genes were the following: BRCA1 in 21%, BRCA2 in 6%, BRCA1 and BRCA2 in 2%. The presence of mutations in either or both of genes was not a significant predictor of pCR (OR 2.06, 95% CI 0.52–8.16; $p = 0.31$) nor of RCB 0/I (OR 1.2, 95% CI 0.34–4.20; $p = 0.78$). Regarding HRD patients, these had higher probability of achieving pCR after neoadjuvant therapy compared to those that do not present this alteration (OR 13.06, 95% CI 1.52–112.41; $p = 0.0028$). Also, RCB 0/I was greater in patients with HRD tumors than in non-HRD (OR 5.10, 95% CI 1.42–18.32; $p = 0.0087$). In addition, HRD score was highly predictive of pCR ($p = 0.011$) and RCB 0/I ($p = 0.0021$). Even if this analysis shows several limitations like the small number of enrolled patients, the retrospective nature and the few patients treated with taxanes without anthracycline, it stresses the role of HRD in predicting the response to neoadjuvant chemotherapy in TNBC.

5. HRD in the metastatic and relapsed settings

The phase II BROCADE trial evaluated the combination of veliparib plus temozolomide (VT) versus veliparib plus carboplatin and paclitaxel (VCP) versus placebo plus carboplatin plus paclitaxel (PCP) in metastatic or locally recurrent TNBC carrying a BRCA mutation [30]. Median PFS was 7.4 months in VT arm, 14.1 months for VCP and 12.3 months for PCP. Median OS was 19.1 months in the first treatment arm, 28.3 in the second and 25.9 in the third. All these data did not reach statistical significance, and the only noteworthy evidence was an increase in overall response rate (ORR) with the combination of VCP versus PCP (77.8% versus 61.3%; $p = 0.027$). In the VT arm, ORR was 28.6%, a sharp reduction compared with the other two arms of treatment. The small sample size could reasonably be the cause of the trend toward PFS and OS increase on both VCP and PCP arms in BRCA mutated TNBC, hence, no definitive conclusions could be drawn. In addition, the same sample size related issue for metastatic disease before platinum treatment also represents a limit for the sensitivity of the analysis. The ongoing phase III of the current study is investigating the efficacy of veliparib added to carboplatin and paclitaxel versus carboplatin plus paclitaxel in BRCA mutated TNBC (BROCADE 3, NCT02163694).

In the phase III study Treating to New Targets (TNT), patients affected by metastatic TNBC or BRCA1/2 mutation-associated breast cancer were randomized to receive carboplatin versus docetaxel [31]. In overall population the response to carboplatin was not different from docetaxel (31.4% versus 35.6%; $p = 0.44$). However, subjects with a BRCA1/2 germline mutation showed a better response when treated with carboplatin compared to docetaxel (68% versus 33%; $p = 0.003$), with a statistically significant interaction between therapy and BRCA mutation status

($p = 0.01$). Also, the median PFS was longer in the platinum group compared to the docetaxel group (6.8 versus 4.4 months). No difference was found in terms of OS. It is important to consider that 56% of patients received carboplatin at progression. BRCA1 methylation as well as low BRCA1 mRNA levels were not associated to a better response to platinum agent [32]. Those with germline BRCA1/2 mutation or BRCA1 methylation showed a high HRD score, being this last condition not associated with a better response to carboplatin compared with docetaxel (38.2% in the carboplatin arm versus 40.4% in the docetaxel arm, a not statistically significant difference [$p = 0.75$]), differently from what was observed in BRCA mutated tumors. This result was also confirmed when the analysis was limited to high HRD scores, including patients irrespective of their BRCA1/2 mutational status. Also in this case a high score was associated to an ORR of 44.7% with carboplatin versus 39.6% with docetaxel and the interaction test evaluating this difference was not statistically significant ($p = 0.67$). No evidence of increase in median PFS was observed in HRD versus non-HRD tumors. These results are similar to those observed in the single-arm, phase II, non randomized TBCRC009 trial in metastatic TNBC, in which patients carrying BRCA1 methylation did not show any response to a platinum agent [33]. The reason could arise from the same condition that the BRCA1 methylation was assessed on primary tissue while the treatment effect was assessed in the metastatic setting. Notwithstanding, this data has a particular value considering that the majority of patients treated in these studies received adjuvant treatment with agents that cause DNA damage, engaging the HR system. Methylation of BRCA1, differently from BRCA mutations, appears to be more revertible. It is postulated that BRCA methylated tumors treated with adjuvant or neoadjuvant chemotherapy actually modify their genetic functionality during treatment since they continue to express the alterations contributing to the HRD score, but drive the tumors towards a soft BRCAness [34,35] phenotype. An example is given by ovarian cancer in which BRCA mutation, but not methylation, was correlated to platinum response, with a biopsy - obtained before and after platinum treatment - showed a reversal of BRCA1 methylation in 31% of tumors [3,36]. The continued presence of methylation was also associated with benefit from a PARP inhibitor [37]. In addition, the Myriad test employed for HRD score definition does not specifically predict response to platinum versus docetaxel in advanced disease [20]. In neoadjuvant TNBC, the high HRD score was associated to platinum response but also in this case these studies do not have a comparison arm for testing the interaction between these biomarkers among specific treatment arms. To overcome these caveats, a new whole-genome sequencing analysis methodology (i.e.: HRDetect) allows to evaluate the presence of multiple pathogenomic mutational signatures and to measure the lifetime HRD due to epigenetic transformations. Studies are ongoing to comprehensively validate this tool [19,20,38–42].

6. Discussion

Hereditary BRCA1/2 mutations are found in 5.3% of all breast cancer and it increases to 10–20% in the TNBC subgroup. Somatic mutations were also reported in 3–5% of TNBC [3,4]. BRCA1/2 and PALB2, among many other genes involved in the mechanism of DNA repair, are critical players in the HR repairs of DSB and are associated with TNBC phenotype [43]. Tumors harboring mutation(s) in one of these genes result more sensible to platinum agents or PARP inhibitors since these compounds cause DNA inter and intra-strand crosslinks that can not be recognized and subsequently repaired by the defective HR system [12]. Although BRCA and PALB2 mutations are sensitive indicators of HRD, other combined biomarkers such as LOH, TAI, and LST are useful to better define the HRD status and to

Table 1

Trials looking at the correlation between HRD status and outcomes among different settings of breast cancer populations. HRD: homologous recombination deficiency; TNBC: triple negative breast cancer; AC: doxorubicin (A) and cyclophosphamide (C); CBDCA: carboplatin; P: paclitaxel; LD: liposomal doxorubicin DFS: disease free survival; OS: overall survival; pCR: pathologic complete response; ns: not significant; T: taxanes; VT: veliparib plus temozolomide; VCP: veliparib + carboplatin + paclitaxel; PCP: placebo + carboplatin + paclitaxel, D: docetaxel; CDDP: cisplatin; ORR: overall response rate.

Study	Population	Treatment	Study Trials	HRD Status Definition	Primary End-Point	Main Results
SWOG S9313 [25]	Stage II-III TNBC	Combined vs sequential adjuvant AC	Retrospective	HRD status ^a	DFS OS	HRD positive status associated with DFS (HR 0.72; p = 0.049). Non significant trend observed with OS (HR 0.71; p = 0.073).
BrightNess [26]	Stage II-III TNBC	Neoadjuvant P followed by AC ± CBDCA ± veliparib	Prospective	BRCA1/2 deleterious mutation	pCR	BRCA 1/2 mutated tumors vs non mutated not predictive for pCR (51% vs 48%; p = ns)
GeparSixto [27]	Stage II-III TNBC Stage II-III HER2+	P + LD ± CBDCA plus bevacizumab in TNBC plus trastuzumab + lapatinib in HER2+	Prospective	HRD status ^a	pCR DFS	HRD positive status associated with increased pCR vs HRD negative (55.9% vs 29.8%; p = 0.001). Adding carboplatin increased pCR in HRD positive but not in HRD negative tumors (64.9% vs 45.2%; p = 0.025)
Telli et al. [29]	Stage II-III TNBC Stage II-III HER2+	A, T, or A + T	Retrospective	HRD status ^a	pCR	HRD positive associated with higher pCR rates after neoadjuvant therapy vs HRD negative (OR 13.06; p = 0.0028)
BROCADE [44]	Stage III-IV TNBC	VT vs VCP vs PCP	Prospective	Germline BRCA1/2 mutations	PFS	No statistical significant increase on PFS in BRCA mutated patients vs non mutated
TNT [31,32]	Stage III-IV TNBC	CBDCA vs D	Prospective	HRD status ^a	ORR	HRD score did not select sensitivity to carboplatin vs docetaxel
TBCRC009 [33]	Stage IV TNBC	CBDCA vs CDDP	Prospective	BRCA1 promoter methylation	ORR	Presence of BRCA1 promoter methylation did not show particular response to platinum agent

^a HRD Status was defined positive as either a deleterious tumor BRCA1/2 (tBRCA) mutation or a pre-defined HRD score ≥ 42 . HRD status was defined negative as either an absence of deleterious tumor BRCA1/2 (tBRCA) mutation or a pre-defined HRD score < 42 .

predict the benefit from platinum agent therapy [13–16]. These alterations, although typical of BRCA mutant tumors, can also be identified in tumors that do not carry this mutation, defining a subgroup of patients referred as BRCAness. A score was identified by a retrospective analysis and defined as the unweighted numeric sum of LOH, TAI, and LST [20]. A score ≥ 42 or the presence of BRCA1/2 mutations were correlated with objective response to cisplatin by an RCB index. This score was evaluated in different settings of breast cancer and Table 1 outlines the main trials regarding them. From these studies, it clearly emerges that HRD has a potential role as a biomarker to predict which patients could benefit the most from a treatment with DNA damaging drugs such as platinum, PARP inhibitors, anthracyclines and/or cyclophosphamide. Two points remain unclear: the role of HRD in predicting response to combination therapy with cisplatin, anthracyclines, cyclophosphamide and taxanes and the role of platinum with or without PARP inhibitors in tumors with a high HRD score.

Alterations of the DNA repairing system have an important implication taken that tumors with genomic instability are responsible for accumulation of multiple genomic aberration in cancer cells, directly contributing to the mutational load [45]. The consequent neoantigen burden correlates with immunotherapy response and survival outcomes in different solid tumors [46–48]. Mismatch repair (MMR) tumors, for example, are characterized by expression of abundant peptides responsible for elicitation of immune response with a high index of infiltrating lymphocytes in the tumor microenvironment [49–51]. MMR deficient (dMMR) tumors, because of their unstable nature, express high levels of PD-1 and PD-L1 proteins that make this subgroup of tumors more responsible to immunotherapy [51,52]. According to these observations, the FDA accelerated the approval of pembrolizumab for pediatric and adult patients with MMR deficient tumors by early 2017 [53,54]. Still, the relationship between the DNA repair system and

immunotherapy response is more complex due to the many DNA lesions types that may elicit different immunological responses. In addition, the DNA repair associated biomarkers were discovered before the next generation sequencing (NGS) era, therefore the employment of this approach could expand the list of features influencing DNA repair system and consequently increase the therapeutic option for patients.

Conflicts of interest

We wish confirm that there are no known conflicts of interest associated with this publication and there has been no significant financial support for this work that could have influenced its outcome. We confirm that the manuscript has been read and approved by all named authors and that there are no other persons who satisfied the criteria for authorship but are not listed. We further confirm that the order of authors listed in the manuscript has been approved by all of us.

We confirm that we have given due consideration to the protection of intellectual property associated with this work and that there are no impediments to publication, including the timing of publication, with respect to intellectual property. In so doing we confirm that we have followed the regulations of our Institutions concerning intellectual property.

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References

- Dent R, Trudeau M, Pritchard KI, et al. Triple-negative breast cancer: clinical features and patterns of recurrence. *Clin Cancer Res* 2007;13(15):4429–34. <https://doi.org/10.1158/1078-0432.CCR-06-3045>.
- Lehmann BD, Shyr Y, Pietenpol JA, et al. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *Nature* 2011;471(7277):250–55. <https://doi.org/10.1038/nature10439>.
- Koboldt DC, Fulton RS, McLellan MD, et al. Comprehensive molecular portraits of human breast tumours. *Nature* 2012;490(7418):61–70. <https://doi.org/10.1038/nature11412>.
- Sharma P, Klemp JR, Kimler BF, et al. Germline BRCA mutation evaluation in a prospective triple-negative breast cancer registry: implications for hereditary breast and/or ovarian cancer syndrome testing. *Breast Cancer Res Treat* 2014;145(3):707–14. <https://doi.org/10.1007/s10549-014-2980-0>.
- Denkert C, Liedtke C, Tutt A, von Minckwitz G. Molecular alterations in triple-negative breast cancer—the road to new treatment strategies. *Lancet* 2017;389(10087):2430–42. [https://doi.org/10.1016/S0140-6736\(16\)32454-0](https://doi.org/10.1016/S0140-6736(16)32454-0).
- Ward JF. Radiation mutagenesis: the initial DNA lesions responsible. *Radiat Res* 1995;142(3):362. <https://doi.org/10.2307/3579145>.
- Cortez D. Requirement of ATM-dependent phosphorylation of Brca1 in the DNA damage response to double-strand breaks. *Science* 1999;286(5442):1162–6. <https://doi.org/10.1126/science.286.5442.1162> (80-).
- Xu B, Kim S -t, Kastan MB. Involvement of Brca1 in S-phase and G2-phase checkpoints after ionizing irradiation. *Mol Cell Biol* 2001;21(10):3445–50. <https://doi.org/10.1128/MCB.21.10.3445-3450.2001>.
- Wang Y, Cortez D, Yazdi P, Neff N, Elledge SJ, Qin J. BASC, a super complex of BRCA1-associated proteins involved in the recognition and repair of aberrant DNA structures. *Genes Dev* 2000;14(8):927–39. <http://www.ncbi.nlm.nih.gov/pubmed/10783165>.
- Pellegrini L, Yu DS, Lo T, et al. Insights into DNA recombination from the structure of a RAD51–BRCA2 complex. *Nature* 2002;420(6913):287–93. <https://doi.org/10.1038/nature01230>.
- Yang H. BRCA2 function in DNA binding and recombination from a BRCA2-DSS1-ssDNA structure. *Science* 2002;297(5588):1837–48. <https://doi.org/10.1126/science.297.5588.1837> (80-).
- Turner N, Tutt A, Ashworth A. Hallmarks of “BRCAness” in sporadic cancers. *Nat Rev Canc* 2004;4(10):814–9. <https://doi.org/10.1038/nrc1457>.
- Marquard AM, Eklund AC, Joshi T, et al. Pan-cancer analysis of genomic scar signatures associated with homologous recombination deficiency suggests novel indications for existing cancer drugs. *Biomark Res* 2015;3(1):9. <https://doi.org/10.1186/s40364-015-0033-4>.
- Abkevich V, Timms KM, Hennessy BT, et al. Patterns of genomic loss of heterozygosity predict homologous recombination repair defects in epithelial ovarian cancer. *Br J J Cancer* 2012;107(10):1776–82. <https://doi.org/10.1038/bjc.2012.451>.
- Birkbak NJ, Wang ZC, Kim J-Y, et al. Telomeric allelic imbalance indicates defective DNA repair and sensitivity to DNA-damaging agents. *Cancer Discov* 2012;2(4):366–75. <https://doi.org/10.1158/2159-8290.CD-11-0206>.
- Popova T, Manie E, Rieunier G, et al. Ploidy and large-scale genomic instability consistently identify basal-like breast carcinomas with BRCA1/2 inactivation. *Cancer Res* 2012;72(21):5454–62. <https://doi.org/10.1158/0008-5472.CAN-12-1470>.
- Telli M, McMillan A, Ford J, et al. Abstract P3-07-12: homologous recombination deficiency (HRD) as a predictive biomarker of response to neoadjuvant platinum-based therapy in patients with triple negative breast cancer (TNBC): a pooled analysis. *Cancer Res* 2016;76(4 Supplement). <https://doi.org/10.1158/1538-7445.SABCS15-P3-07-12>.
- Connolly R, Elkin E, Timms K, et al. Abstract P3-07-13: homologous recombination deficiency (HRD) as a predictive biomarker of response to preoperative systemic therapy (PST) in TBCRC008 comprising a platinum in HER2-negative primary operable breast cancer. *Cancer Res* 2016;76(4 Supplement). <https://doi.org/10.1158/1538-7445.SABCS15-P3-07-13>.
- Timms KM, Abkevich V, Hughes E, et al. Association of BRCA1/2 defects with genomic scores predictive of DNA damage repair deficiency among breast cancer subtypes. *Breast Cancer Res* 2014;16(6):475. <https://doi.org/10.1186/s13058-014-0475-x>.
- Telli ML, Timms K, Reid J, et al. Homologous recombination deficiency (HRD) score predicts response to platinum-containing neoadjuvant chemotherapy in patients with triple negative breast cancer. *Clin Cancer Res* 2016;22(15):3764–73. <https://doi.org/10.1158/1078-0432.CCR-15-2477>.
- Telli ML, Jensen KC, Vinayak S, et al. Phase II study of gemcitabine, carboplatin, and iniparib as neoadjuvant therapy for triple-negative and BRCA1/2 mutation-associated breast cancer with assessment of a tumor-based measure of genomic instability: PrECOG 0105. *J Clin Oncol* 2015;33(17):1895–901. <https://doi.org/10.1200/JCO.2014.57.0085>.
- Silver DP, Richardson AL, Eklund AC, et al. Efficacy of neoadjuvant cisplatin in triple-negative breast cancer. *J Clin Oncol* 2010;28(7):1145–53. <https://doi.org/10.1200/JCO.2009.22.4725>.
- Ryan PD, Tung NM, Isakoff SJ, et al. Neoadjuvant cisplatin and bevacizumab in triple-negative breast cancer (TNBC): safety and efficacy. *J Clin Oncol* 2009;27(15S):551–551. <http://ascopubs.org/doi/abs/10.1200/jco.2009.27.15s.551>.
- Symmans WF, Peintinger F, Hatzis C, et al. Measurement of residual breast cancer burden to predict survival after neoadjuvant chemotherapy. *J Clin Oncol* 2007;25(28):4414–22. <https://doi.org/10.1200/JCO.2007.10.6823>.
- Sharma P, Barlow WE, Godwin AK, et al. Impact of homologous recombination deficiency biomarkers on outcomes in patients with triple-negative breast cancer treated with adjuvant doxorubicin and cyclophosphamide (SWOG S9313). *Ann Oncol* 2018;29(3):654–60. <https://doi.org/10.1093/annonc/mdx821>.
- Loibl S, O’Shaughnessy J, Untch M, et al. Addition of the PARP inhibitor veliparib plus carboplatin or carboplatin alone to standard neoadjuvant chemotherapy in triple-negative breast cancer (BrightNess): a randomised, phase 3 trial. *Lancet Oncol* 2018;1–13. [https://doi.org/10.1016/S1470-2045\(18\)30111-6](https://doi.org/10.1016/S1470-2045(18)30111-6).
- von Minckwitz G, Schneeweiss A, Loibl S, et al. Neoadjuvant carboplatin in patients with triple-negative and HER2-positive early breast cancer (GeparSixto; GBG 66): a randomised phase 2 trial. *Lancet Oncol* 2014;15(7):747–56. [https://doi.org/10.1016/S1470-2045\(14\)70160-3](https://doi.org/10.1016/S1470-2045(14)70160-3).
- von Minckwitz G, Timms K, Untch M, et al. Prediction of pathological complete response (pCR) by Homologous Recombination Deficiency (HRD) after carboplatin-containing neoadjuvant chemotherapy in patients with TNBC: results from GeparSixto. *J Clin Oncol* 2015;33 (15_suppl (May 20 2015)):1004-1004.
- Telli ML, Hellyer J, Audeh W, et al. Homologous recombination deficiency (HRD) status predicts response to standard neoadjuvant chemotherapy in patients with triple-negative or BRCA1/2 mutation-associated breast cancer. *Breast Cancer Res Treat* 2018;168(3):625–30. <https://doi.org/10.1007/s10549-017-4624-7>.
- Han HS, Diéras V, Robson M, et al. Veliparib with temozolomide or carboplatin/paclitaxel versus placebo with carboplatin/paclitaxel in patients with BRCA1/2 locally recurrent/metastatic breast cancer: randomized phase II study. *Ann Oncol* 2018;29(1):154–61. <https://doi.org/10.1093/annonc/mdx505>.
- Tutt A, Ellis P, Kilburn L, et al. Abstract S3-01: the TNT trial: a randomized phase III trial of carboplatin (C) compared with docetaxel (D) for patients with metastatic or recurrent locally advanced triple negative or BRCA1/2 breast cancer (CRUK/07/012). *Cancer Res* 2015;75(9 Supplement). <https://doi.org/10.1158/1538-7445.SABCS14-S3-01>.
- Tutt A, Tovey H, Cheang MCU, et al. Carboplatin in BRCA1/2-mutated and triple-negative breast cancer BRCAness subgroups: the TNT Trial. *Nat Med* 2018;24(5):628–37. <https://doi.org/10.1038/s41591-018-0009-7>.
- Isakoff SJ, Mayer EL, He L, et al. TBCRC009: a multicenter phase II clinical trial of platinum monotherapy with biomarker assessment in metastatic triple-negative breast cancer. *J Clin Oncol* 2015;33(17):1902–9. <https://doi.org/10.1200/JCO.2014.57.6660>.
- Lord CJ, Ashworth A. BRCAness revisited. *Nat Rev Canc* 2016;16(2):110–20. <https://doi.org/10.1038/nrc.2015.21>.
- ter Brugge P, Kristel P, van der Burg E, et al. Mechanisms of therapy resistance in patient-derived xenograft models of BRCA1-deficient breast cancer. *J Natl Cancer Inst* 2016;108(11):djw148. <https://doi.org/10.1093/jnci/djw148>.
- Chiang JW, Karlan BY, Cass L, Baldwin RL. BRCA1 promoter methylation predicts adverse ovarian cancer prognosis. *Gynecol Oncol* 2006;101(3):403–10. <https://doi.org/10.1016/j.ygyno.2005.10.034>.
- Swisher EM, Lin KK, Oza AM, et al. Rucaparib in relapsed, platinum-sensitive high-grade ovarian carcinoma (ARIEL2 Part 1): an international, multicentre, open-label, phase 2 trial. *Lancet Oncol* 2017;18(1):75–87. [https://doi.org/10.1016/S1470-2045\(16\)30559-9](https://doi.org/10.1016/S1470-2045(16)30559-9).
- Davies H, Glodzik D, Morganella S, et al. HRDetect is a predictor of BRCA1 and BRCA2 deficiency based on mutational signatures. *Nat Med* 2017;23(4):517–25. <https://doi.org/10.1038/nm.4292>.
- Mulligan JM, Hill LA, Deharo S, et al. Identification and validation of an anthracycline/cyclophosphamide-based chemotherapy response assay in breast cancer. *JNCI J Natl Cancer Inst* 2014;106(1). <https://doi.org/10.1093/jnci/djt335>.
- Wolf DM, Yau C, Sanil A, et al. Abstract P3-06-05: evaluation of an in vitro derived signature of olaparib response (PARPi-7) as a predictive biomarker of response to veliparib/carboplatin plus standard neoadjuvant therapy in high-risk breast cancer: results from the I-SPY 2 TRIAL. *Cancer Res* 2015;75(9 Supplement). <https://doi.org/10.1158/1538-7445.SABCS14-P3-06-05>.
- Sihto H, Lundin J, Lundin M, et al. Breast cancer biological subtypes and

- protein expression predict for the preferential distant metastasis sites: a nationwide cohort study. *Breast Cancer Res* 2011;13(5):R87. <https://doi.org/10.1186/bcr2944>.
- [42] Lin NU, Claus E, Sohl J, Razzak AR, Arnaout A, Winer EP. Sites of distant recurrence and clinical outcomes in patients with metastatic triple-negative breast cancer. *Cancer* 2008;113(10):2638–45. <https://doi.org/10.1002/cncr.23930>.
- [43] Easton DF, Pharoah PDP, Antoniou AC, et al. Gene-panel sequencing and the prediction of breast-cancer risk. *N Engl J Med* 2015;372(23):2243–57. <https://doi.org/10.1056/NEJMs1501341>.
- [44] Han HS, Diéras V, Robson M, et al. Veliparib with temozolomide or carboplatin/paclitaxel versus placebo with carboplatin/paclitaxel in patients with BRCA1/2 locally recurrent/metastatic breast cancer: randomized phase II study. *Ann Oncol* 2018;29(1):154–61. <https://doi.org/10.1093/annonc/mdx505>.
- [45] Lawrence MS, Stojanov P, Mermel CH, et al. Discovery and saturation analysis of cancer genes across 21 tumour types. *Nature* 2014;505(7484):495–501. <https://doi.org/10.1038/nature12912>.
- [46] Snyder A, Makarov V, Merghoub T, et al. Genetic basis for clinical response to CTLA-4 blockade in melanoma. *N Engl J Med* 2014;371(23):2189–99. <https://doi.org/10.1056/NEJMoa1406498>.
- [47] Van Allen EM, Miao D, Schilling B, et al. Genomic correlates of response to CTLA-4 blockade in metastatic melanoma. *Science* (80-) 2015;350(6257):207–11. <https://doi.org/10.1126/science.aad0095>.
- [48] Rizvi NA, Hellmann MD, Snyder A, et al. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. *Science* (80-) 2015;348(6230):124–8. <https://doi.org/10.1126/science.aaa1348>.
- [49] Chalmers ZR, Connelly CF, Fabrizio D, et al. Analysis of 100,000 human cancer genomes reveals the landscape of tumor mutational burden. *Genome Med* 2017;9(1):34. <https://doi.org/10.1186/s13073-017-0424-2>.
- [50] Jamieson NB, Maker AV. Gene-expression profiling to predict responsiveness to immunotherapy. *Cancer Gene Ther* 2017;24(3):134–40. <https://doi.org/10.1038/cgt.2016.63>.
- [51] Palmieri G, Colombino M, Cossu A, Marchetti A, Botti G, Ascierto PA. Genetic instability and increased mutational load: which diagnostic tool best direct patients with cancer to immunotherapy? *J Transl Med* 2017;15(1):17. <https://doi.org/10.1186/s12967-017-1119-6>.
- [52] Le DT, Uram JN, Wang H, et al. PD-1 blockade in tumors with mismatch-repair deficiency. *N Engl J Med* 2015;372(26):2509–20. <https://doi.org/10.1056/NEJMoa1500596>.
- [53] Dudley JC, Lin M-T, Le DT, Eshleman JR. Microsatellite instability as a biomarker for PD-1 blockade. *Clin Cancer Res* 2016;22(4):813–20. <https://doi.org/10.1158/1078-0432.CCR-15-1678>.
- [54] FDA approves first cancer treatment for any solid tumor with a specific genetic feature. FDA News Release.