



Review article

Gut microbiota in neurodegenerative disorders

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A B S T R A C T

Gut dysbiosis, a primary factor behind various gastrointestinal disorders may also augment lipopolysaccharides, pro-inflammatory cytokines, T helper cells and monocytes causing increased intestinal and BBB permeability via microbiota-gut-brain axis. Consequentially, accumulation of misfolded proteins, axonal damage and neuronal demyelination sets in, thus facilitating the pathogenesis of neurodegenerative disorders like Parkinson's disease, Alzheimer's disease, multiple sclerosis and amyotrophic lateral sclerosis. Studies revealed that intake of probiotics may help in the integrity of intestinal and BBB thus ameliorating the above neurodegenerative disorders. This review summarizes the current understanding of the role of gut microbiota in neurodegenerative disorders and possible intervention strategies.

1. Introduction

Gut microbiota refers to the symbiotic microorganisms like bacteria, archae, viruses and fungi present in human gut (Guarner and Malagelada, 2003; Quigley, 2013). These diverse groups of microorganisms play multiple roles in humans like fermenting undigested carbohydrates, producing short-chain fatty acids (SCFA), synthesizing Vitamin B and K, metabolizing essential substances (bile acids, sterols and drugs) and protecting against outside pathogens. (Quigley, 2013) Generation of microbiota in human gut is itself a unique phenomenon. At birth, the human foetal gut is sterile and devoid of any microorganism. As the child develops, the actual colonization of microbes and its diversity take place (Sekirov et al., 2010). Factors which influence the composition of microbiota are gestational age, mode of delivery, diet, level of sanitation and exposure to antibiotics (Fouhy et al., 2012; Marques et al., 2010). The two most prominent phyla of gut microbiota present in human are *Firmicutes* and *Bacteroides* (Bäckhed, 2011; Eckburg et al., 2005; Qin et al., 2010). The gut flora in human has been differentiated as, essential or beneficial flora (considered as housekeepers of gut) and opportunistic bacteria (which causes infection). *Lactobacteria* species (*L. rhamnosus*, *L. acidophilus*, *L. plantarum* etc), *Bifidobacterium* (*B. bifidum*), *Enterococci*, *Propionobacteria* and *Peptostreptococci* belongs to the first group. The second group comprises of *Bacteriodes*, *Bacilli*, *Clostridia*, *Enterobacteria*, *Actenobacteria*, *Peptococci*, *Staphylococci*, *Streptococci* and *Yeasts* etc. (Joshi et al., 2018). Imbalance between these two groups causes dysbiosis which leads to gastrointestinal disorders like irritable bowel syndrome, ulcerative colitis, crohn's disease, colorectal cancer and metabolic disorders (Frank et al., 2007; Guinane and Cotter, 2013; Ley et al., 2006). Gut has an intimate bidirectional relationship with the central nervous system (CNS). While, gut communicates with CNS via sympathetic and

parasympathetic nervous system (termed as Gut-Brain Axis), CNS communicate via both afferent and efferent autonomic pathways (ANS) with muscle and mucosal layer of gut. Thereby brain modulates gut motility, immunity, permeability and secretion of mucus. In parallel gut also communicates with Hypothalamic–pituitary–adrenal axis (HPA axis) (Belmaker and Agam, 2008; Collins and Bercik, 2009; Forsythe et al., 2009). HPA axis can be activated in response to environmental factors, such as emotion or stress leading to the production of corticotrophin releasing hormone (CRH) from hypothalamus (Belmaker and Agam, 2008). CRH circulates through bloodstream, reaches the anterior pituitary and stimulates the release of adrenocorticotrophic hormone which leads to cortisol secretion from the adrenal glands. The cortisol acts on intestinal targets and disrupts intestinal permeability (Collins and Bercik, 2009; Forsythe et al., 2009; Forsythe et al., 2014; Stilling et al., 2014). Moreover, microbes present in gut effect the growth and regulation of the immune system and therefore may modulate the immune system CNS communication (Belkaid and Hand, 2014). Gut bacteria may also be involved in the synthesis of neuroactive molecules and metabolites which may modulate the pathogenesis of various neurodegenerative disorders like Parkinson's disease, Alzheimer's disease, multiple sclerosis and amyotrophic lateral sclerosis (Girolamo et al., 2017; Lei et al., 2016; Pryde et al., 2002; Quigley, 2017; Vital et al., 2014; Zhang and Davies, 2016).

In this review we present an outline of gut microbiota and its interaction with central nervous system. Also the role of gut microbiota in pathogenesis of various neurodegenerative disorders has been portrayed. Further, we have presented probiotics as potential therapeutic interventions against the above neurodegenerative disorders.

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1.1. Parkinson's disease (PD)

PD is a multicentric neurodegenerative disorder due to progressive alpha-synucleinopathy i.e. deposition of alpha-synuclein (α -syn) in the neuronal cell body. This results in the formation of Lewy bodies which are round lamellated eosinophilic cytoplasmic inclusions (Braak et al., 2003). It causes loss of dopaminergic neurons in the substantia nigra present in the midbrain (Dickson et al., 2009). The motor symptoms include akinesia, muscular rigidity, resting tremor, difficulty in walking and gait disorder (Braak et al., 2003). The loss of dopaminergic neurons and motor symptoms are interrelated (Dickson et al., 2009). Non-motor symptoms include dementia (Tsuang et al., 2013), depression (Parashar and Udayabanu, 2017), loss of smell (Cersosimo et al., 2013) and gastrointestinal (GI) dysfunction such as nausea, dysphagia, abnormal salivation, constipation and defecatory dysfunction (Edwards et al., 1992; Mulak and Bonaz, 2015).

Progression of PD has been frequently associated with the dysbiosis of gut microbiota. Studies have shown that imbalance in gut microbiota plays an important role in aggravating PD (Braak et al., 2006; Kiebertz and Wunderle, 2013; Savica et al., 2009; Shannon et al., 2012). One such study found lower abundance of *Prevotellaceae* species compared to relative abundance of *Enterobacteriaceae* in the stool of PD patients (Scheperjans et al., 2015). *Prevotellaceae* family members are commensals and involved in production of mucin and short chain fatty acids (SCFAs) through the fermentation of dietary fibers (Arumugam et al., 2011). Reduction in *Prevotellaceae* population leads to increase in gut permeability and systemic exposure of bacterial endotoxins which initiates or retains excessive α -syn expression in colon and even supports its misfolding (Forsyth et al., 2011; Niehaus and Lange, 2003). The relative abundance of *Enterobacteriaceae* has been positively related with the severity of postural instability and gait difficulty (Scheperjans et al., 2015). Increase in *Enterobacteriaceae* rises serum lipopolysaccharide (LPS) which is derived from cell wall of these gram negative bacteria (Forsyth et al., 2011). PD patients showed high levels of LPS binding protein in the blood due to high absorption of LPS (Dutta et al., 2008; Tufekci et al., 2011). LPS and other neurotoxins cross the intestinal wall enter into the blood stream and results in disruption of intestinal epithelial barrier (Guo et al., 2013). LPS in the blood stream may induce systemic inflammation by the production of inflammatory cytokines via toll like receptor (TLR) 4 and nuclear factor-kappa B (NF- κ B) pathway (Guo et al., 2013; Tufekci et al., 2011). LPS and inflammatory cytokines like tumour necrosis factor (TNF- α), interleukin (IL)-1 β and IL-6 disrupts blood brain barrier (BBB) and promotes α -synuclein deposition (Block et al., 2007; Chen et al., 2007; Dobbs et al., 1999; Reale et al., 2009; Sui et al., 2014). BBB breakdown has been reported to result in the destruction of dopaminergic neurons in the substantia nigra (Rite et al., 2007). Therefore relative increase of *Enterobacteriaceae* and increase in LPS has been correlated with the progression of PD (Mulak and Bonaz, 2015; Nair et al., 2018). Tight junction proteins like occludins are a necessary for the structural maintenance of intestinal barrier (Forster, 2008). Dysbiosis may decrease occludins and cause increase in intestinal permeability (Clairembault et al., 2015; Forsyth et al., 2011). (Fig. 1) Another study showed increase of pro-inflammatory cytokine producing bacteria such as *Ralstonia*, *Proteobacteria*, *Enterococcaceae* concentration in mucosa of PD patients. A decrease in the butyrate producing bacteria such as *Blautia*, *Coproccoccus* and *Roseburia* and *Faecalibacterium* which are considered as anti-inflammatory, have been reported in the stool samples of PD patients. In addition significant increase in LPS biosynthesis gene was reported in the PD fecal microbiome (Keshavazian et al., 2015). Excessive bacterial growth in small intestine termed as small intestinal bacterial overgrowth (SIBO), has been associated with motor impairment and PD (Fasano et al., 2013; Tan et al., 2014). *Helicobacter pylori* infection has also been considered as a triggering factor in PD pathogenesis (Çamcı and Oğuz, 2016).

According to FAO/WHO Probiotics are defined as “Live

microorganisms which, when administered/ingested in adequate amounts, confer a health benefit to the host.” Probiotics containing *bifidobacteria* and *lactobacilli* has been reported to reverse PD like condition (Pompei et al., 2007). The regular intake of fermented milk beverage containing the probiotic *Lactobacillus casei shirota* have been shown to improve the bowel movement by decreasing the number of fecal *staphylococci* in PD patients (Cassani et al., 2011). Probiotic bacterium *Bacillus* spp. may convert L-tyrosine to L-DOPA which is converted to dopamine in the presence of DOPA decarboxylase (Surwase and Jadhav, 2011).

1.2. Alzheimer's disease (AD)

AD is a neurodegenerative disorder leading to progressive cognitive dysfunction. It is accompanied by the cerebral accumulation of misfolded amyloid- β peptide in senile plaques, a cleavage product of the amyloid- β precursor protein (A β PP) (Alzheimer's Disease Fact Sheet National Institute on Aging and U.S. Department of Health and Human Services, 2013). It is also associated with tau hyperphosphorylation formation of neurofibrillary tangles and loss of cholinergic neurons (Hu et al., 2016; Jiang et al., 2017).

Gut dysbiosis which has been associated with increased gut permeability and inflammation may also results in an increase in circulating levels of microbes like *spirochaete*, *Herpes simplex* virus type 1 (HSV-1), *Chlamydia pneumoniae* and also gut microbiota derived products like β -N-methylamino-L-alanine (BMAA), LPS and microbial amyloids (Heneka et al., 2015; Itzhaki et al., 2016; Ransohoff, 2016). BMAA is one of the neurotoxin produced by gut cyanobacteria (Brenner, 2013) which causes neurodegeneration, cognitive impairment, astrogliosis and accumulation of neurofibrillary tangles in adult rats after neonatal exposure (Karlsson et al., 2011). LPS induces the formation of A β fibrils (Asti and Gioglio, 2014) which is a potent inducer of NF- κ B, known to be involved in neuroinflammation in the AD brain (Lukiw, 2016). Microglia contain Toll-like receptor (TLR) 4 with LPS as its ligand (Lien et al., 2000). TLR4 function depends on the co-receptor CD 14. Abundance of CD 14 has been reported in microglia of AD brain (Doens and Fernández, 2014) (The role of TLR4 and CD14 in AD has been discussed in further details by Liu et al., 2005). Gut microbiota derived amyloid may enhance inflammation in response to cerebral A β ₄₂ by acting upon innate immune system (Friedland, 2015). Moreover aging also causes overstimulation of innate and adaptive immune systems resulting into low-grade, chronic state of inflammation termed as inflammaging (For further details please refer Franceschi et al., 2000; Franceschi, 2007). Inflammaging increases gut and BBB permeability which influences the production of A β in the brain (Elahy et al., 2015; Pistollato et al., 2016; Ulluwishewa et al., 2011; Zhao et al., 2015). Gut microbes, microbiota derived products and inflammatory mediators reach brain, trigger neuroinflammation and also enhance the amyloid accumulation. Relative abundance of *proteobacteria* and decrease in butyrate producing bacteria may result in high level of plasma pro-inflammatory cytokines like IL-6 and IL-8 (Biagi et al., 2010). A study showed an increase in concentration of *Escherichia/shigella* spp. which are characterized as pro-inflammatory and decrease in concentration of *Eubacterium rectale* spp., characterized as anti-inflammatory, in fecal microbiome of AD patients. This dysbiosis may also increase brain amyloid deposition along with peripheral inflammation (Cattaneo et al., 2017). (Fig. 1).

Various animal studies have been carried out to define the role of probiotics on cognitive behavior. Treatment with the combination of probiotics *Lactobacillus rhamnosus* (R0011) and *Lactobacillus helveticus* (R0052) restored the stress-induced memory impairment by dysbiosis caused by *Citrobacter rodentium* infection in mice (Gareau et al., 2011). Another study showed learning and memory improvement in anxious mice by the administration of *Bifidobacterium longum* 1714 (Savignac et al., 2015). A probiotic strain *Lactobacillus fermentum* NS9 improved spatial memory impairment caused by ampicillin associated dysbiosis

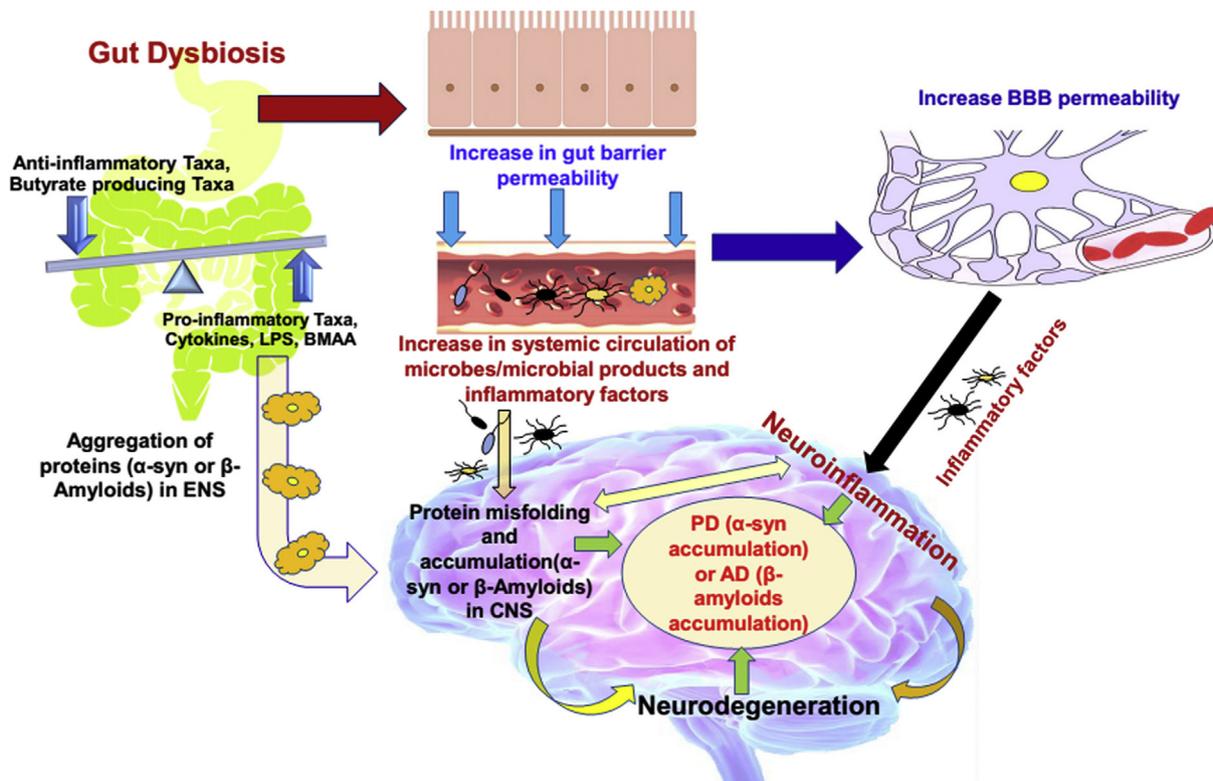


Fig. 1. Gut dysbiosis leading to the pathogenesis of PD and AD.

Imbalance in group of micro-organism producing anti-inflammatory cytokines (Anti-inflammatory Taxa) and micro-organisms producing pro-inflammatory cytokines (Pro-inflammatory Taxa) in gut termed as Gut dysbiosis. Gut dysbiosis generates pro-inflammatory cytokines, LPS and BMAA which in turn leads to disruption of gut permeability and increases systemic circulation of microbes, microbial products, and inflammatory factors. The inflammatory factors disrupt BBB and cause neuroinflammation and neurodegeneration. In addition, aggregation of proteins such as α -syn or β -Amyloids in ENS is enhanced due to gut dysbiosis. This results in the accumulation of proteins in CNS which further results neuroinflammation and neurodegeneration. Also, the microbes and microbial products present in systemic circulation enhance accumulation of proteins in CNS. Accumulation of α -syn in CNS causes PD and accumulation of β -amyloids in CNS causes AD. LPS- Lipopolysaccharide, BMAA- β -N-methylamino-L-alanine, α -Syn- α -Synuclein, BBB-Blood Brain Barrier, ENS-Enteric Nervous System, CNS- Central Nervous System.

(Wang et al., 2015a). One more study reported the improvement of spatial memory due to chronic restraint stress by restoring BDNF levels in hippocampus by the treatment of *Lactobacillus helveticus* NS8 (Liang et al., 2015). VSL#3 restored neurogenesis and increased the Ly6Chi monocyte proportions in the brain, resulting in improved cognition (Möhle et al., 2016). A randomized, double blind controlled clinical trial reported positive effect of *Lactobacillus acidophilus*, *Lactobacillus casei*, *Lactobacillus bifidum*, *Lactobacillus fermentum* probiotic combination on cognitive function in AD patients (Akbari et al., 2016). The administration of *Bifidobacterium* species results in abundance of *Bifidobacterium* in the gut microbiota of elderly leading to lower serum levels of pro-inflammatory cytokines TNF- α , IL-5, IL-6, IL-1 β and IL-8 (Ouweland et al., 2008; Wang et al., 2015b).

1.3. Multiple sclerosis (MS)

MS is an immune mediated chronic neurological disorder. The neuropathological hallmarks of MS are demyelination, axonal damage and neurodegeneration. The clinical manifestations include vertigo, vision loss, dizziness, pain, motor dysfunction, impaired coordination, fatigue, numbness, bladder and bowel dysfunction, depression and cognitive impairment. MS is primarily divided into i) Progressive-relapsing MS (PRMS), ii) Primary Progressive MS (PPMS), iii) Secondary Progressive MS (SPMS), iv) Relapsing-Remitting MS (RRMS) (Noseworthy et al., 2000). The pathogenesis of MS depends upon various genetic and environmental factors (Compston and Coles, 2008).

Gut dysbiosis is considered to be an important environmental factor responsible for the neuropathogenesis of MS (Bhargava and Mowry,

2014; Compston and Coles, 2008; Mowry and Glenn, 2018). RRMS is characterized by decrease in bacteria responsible for producing anti-inflammatory responses by immune cells like T regulatory cells (T_{regs}), $CD4^+$ T cells which produces IL-10, regulatory B cells, tolerogenic dendritic cells and suppressive macrophages and abundance of bacteria responsible for pro-inflammatory reactions produced by $CD4^+$ T cells dendritic cells, monocytes or B cells (Shahi et al., 2017) (For details in inflammatory and regulatory cells please refer Chung and Kasper, 2010). The presence of *Clostridium*, *Bacteroidetes*, (Ríos-Covián et al., 2016) *Prevotella*, *Parabacteroides* and *Lactobacillus* in gut microbiota can induce production of SCFAs (El Kaoutari et al., 2013; Sivieri et al., 2013) and helps in the maintenance of immune cells which produces anti-inflammatory response (Freedman et al., 2018; Shahi et al., 2017). In RRMS patients, decreased abundance of *Parabacteroides distasonis* (Cekanaviciute et al., 2016) and *Prevotella copri* (Miyake et al., 2015) were reported when compared with healthy controls. Within *Firmicutes* spp., concentration of *Dorea* and *Blautia* were increased (Chen et al., 2016) and in few studies depletion of *Clostridium* (Miyake et al., 2015) and *Faecalibacterium* (Cantarel et al., 2015) were observed. However, increased colonization of *Firmicutes* has been reported in pediatric MS patients (Tremlett et al., 2016). Another study reported decrease in relative abundance of *Faecalibacterium* in fecal samples of MS patients (Mowry et al., 2012). *Faecalibacterium* is an important butyrate producing bacteria while butyrate production may increase T_{regs} (Arpaia et al., 2013). The human gut colonization by *Clostridium perfringens* type B has been reported in MS patients (Rumah et al., 2013) resulting in increased levels of epsilon toxin (Mete et al., 2013). This toxin degrades BBB, leading to neuronal and oligodendrocyte damage and triggers

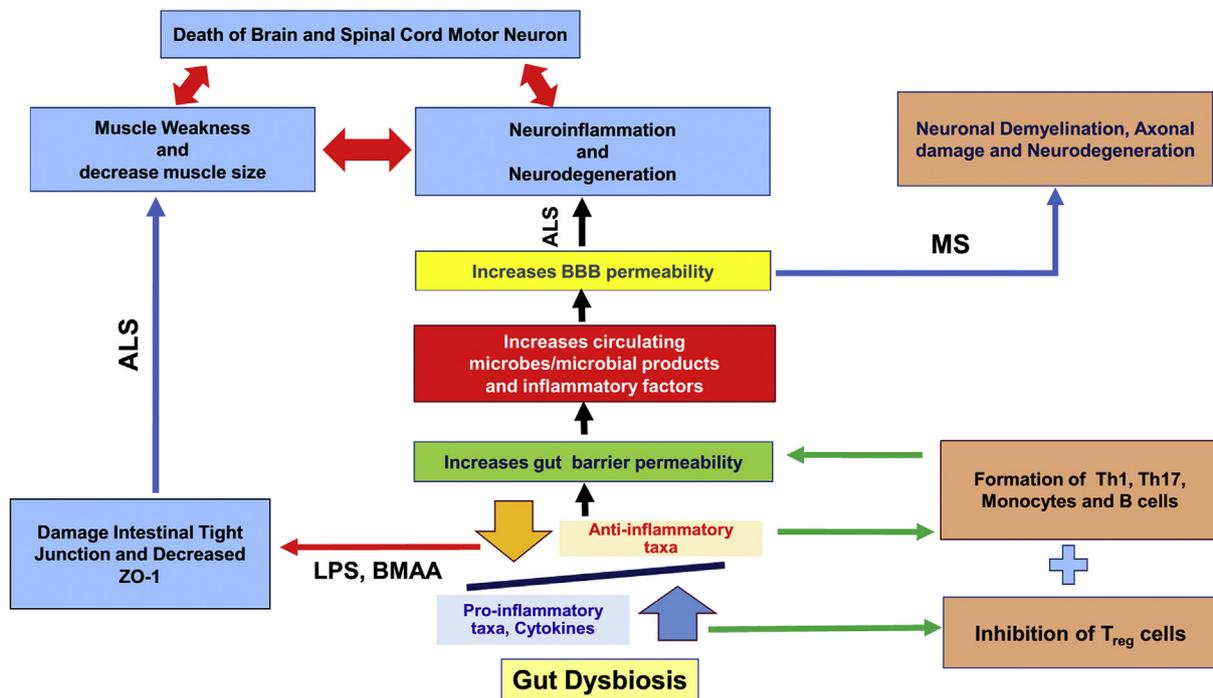


Fig. 2. Gut dysbiosis leading to the pathogenesis of MS and ALS.

In ALS, the pro-inflammatory cytokines produced during dysbiosis leads to disruption of gut permeability and increases systemic circulation of microbes, microbial products, and inflammatory factors. The inflammatory factors disrupt BBB and cause neuroinflammation and neurodegeneration. The dysbiosis product, LPS and BMAA damage intestinal tight junction and junction protein (ZO-1) leads to muscle weakness and decrease muscle size which resultant neuroinflammation, neurodegeneration, and death of brain and spinal cord motor neuron. While in MS, dysbiosis generates Th1, Th17, Monocytes and B cells, and inhibition of T_{reg} cells. These factors disrupt gut permeability and increase systemic circulation of microbes, microbial products, and inflammatory factors. The inflammatory factors disrupt BBB barrier and cause neuronal demyelination, axonal damage, and neurodegeneration.

T_{reg}- T regulatory cells, LPS- Lipopolysaccharide, BMAA-β-N-methylamino-L-alanine, ZO- ZonulaOccludens, Th1- T helper cells, BBB-Blood Brain Barrier, ENS-Enteric Nervous System.

autoimmune demyelinating events (Dorca-Arévalo et al., 2008; Finnie et al., 1999; Lonchamp et al., 2010). Few species of *Dorea* may stimulate IFN γ (Schirmer et al., 2016), metabolize sialic acids and degrade mucin (Hughes et al., 2003). *Dorea* produces gases which are used by *Blautia* resulting in an increased abundance of *Blautia*. (Schirmer et al., 2016) Increased abundance of *Akkermansia* has been reported among MS patients. *Akkermansia muciniphila*. *Blautia* may degrade mucin and enhance the production of pro-inflammatory cytokines (Cekanaviciute et al., 2016; Jangi et al., 2016). A study reported decreased concentration of *Adlercreutzia equolifaciens* and *Collinsella* (Chen et al., 2016) whereas another study reported decrease in the concentrations of *Collinsella* and *Slackia* but no change in *Adlercreutzia equolifaciens* in MS patients (Jangi et al., 2016). Higher abundance of Proteobacteria species like *Mycoplana*, *Pseudomonas*, (Chen et al., 2016) *Bilophila* (Miyake et al., 2015), *Acinetobacter calcoaceticus* (Cekanaviciute et al., 2016) and lower abundance of *Sutterella* (Proteobacteria) were also observed in MS (Jangi et al., 2016). *Acinetobacter calcoaceticus* can stimulate pro-inflammatory Th1 cytokines and depress regulatory CD4 T cells thereby aggravating MS (Cekanaviciute et al., 2016) (Fig. 2).

Different probiotics have been tested in Experimental Autoimmune Encephalomyelitis (EAE) animal model of MS. Oral administration of *Bifidobacterium animalis* to these animals may reduce in EAE symptoms (Ezendam et al., 2008). *Lactobacillus* species enhanced regulatory T cells, producing increased IL-10 and decreased IL-17 and improved EAE clinical score in these animals (Lavasani et al., 2010; Maassen and Claassen, 2008; Takata et al., 2011). Regular oral administration of various combinations of probiotic like *Streptococcus thermophiles*, *Bifidobacterium bifidum* and *Lactobacillus* spp. in both mouse and rat models may improve EAE clinical score, reduce Th1 and Th17 cells along with enhancement of T_{regs} (Kwon et al., 2013).

1.4. Amyotrophic lateral sclerosis (ALS)

ALS is a progressive neurodegenerative disorder pertaining to death of brain, spinal cord and motor neuron. The symptoms include muscle weakness, cramping, problems with coordination, stiff muscles, muscle spasms and muscle twitching. This gradually leads to difficulty in speaking, swallowing and breathing. There are two types of ALS: i) Sporadic ALS, which is the most common form with unknown aetiology and ii) Familial ALS, which is caused by changes to gene (Toepfer et al., 1997).

ALS mouse model which is a G93A SOD1 (superoxide dismutase) transgenic mice are used to study the role of gut microbiota in the disease. The course of ALS in an animal model can be divided into three stages: a) pre-onset- 60 to 70 days, b) onset- 90 to 100 days and c) progressive- 120-130 days (Zhou et al., 2015). During the pre-onset stage the transgenic mice show increased gut permeability and disrupted BBB. Damaged intestinal wall due to decreased zonula occludens (ZO)-1 and E-cadherin has been reported. The SOD1G93A mice feces showed reduced levels of butyrate-producing bacteria like *Butyrivibrio fibrosolvens* and *peptostreptococcus* resulting in elevated level of serum and intestinal pro-inflammatory cytokine IL-17 (Fang, 2015; Wu et al., 2015). Another study reported the delay in ALS progression by administration of 2% Butyrate in drinking water to SOD1 transgenic mice. This may be due to the improvement of intestinal barrier function by relative abundance of *B. fibrosolvens* (Zhang et al., 2017). ALS patients' feces have reduced levels of butyrate producing bacteria like *Oscillibacter*, *Anaerostipes*, and *Lachnospira* and enhanced levels of different species of *Dorea*. *Dorea* may synthesize ethanol as an end product of glucose metabolism and thus considered harmful (Fang et al., 2016). Alterations of LPS and BMAA in plasma levels of ALS individuals have also been reported. These molecules if present in circulation may reach

CNS and contribute to the pathogenesis of ALS through neuroinflammation and deterioration of the BBB function (Bengmark, 2013; Catanzaro et al., 2015; Wright et al., 2018). While, BBB disruption is common in both human and animals with ALS (Garbuzova-Davis and Sanberg, 2014) (Fig. 2). The therapeutic role of probiotics in the improvement of ALS like conditions is yet to be explored.

2. Conclusion

The human intestinal tract contain a huge, active and complex community of micro-organism. The intestinal microbiota plays a vital role in the digestion of food, immunopotentiality, stimulation of development of microvilli, fermentation of dietary fibers and prevention of colonization of the gastrointestinal tract by harmful pathogens. Various observational and animal studies suggest that gut microbiota play a significant role in neuropathogenesis of CNS disorders by altering in Gut-Brain-Axis function. Dysbiosis may increase inflammatory cytokines and bacterial metabolites which may alter the gut and BBB permeability and cause neuroinflammation. This may facilitate the pathogenesis of various neurodegenerative disorders like PD, AD, MS and ALS. Probiotics, by preventing gut dysbiosis may provide protection from the above neurodegenerative disorders.

References

- Akbari, E., Asemi, Z., Daneshvar Kakhaki, R., Bahmani, F., Kouchaki, E., Tamtaji, O.R., Hamidi, G.A., Salami, M., 2016. Effect of probiotic supplementation on cognitive function and metabolic status in Alzheimer's disease: a randomized, double-blind and controlled trial. *Front. Aging Neurosci.* 8, 256.
- Alzheimer's Disease Fact Sheet National Institute on Aging, U.S. Department of Health and Human Services, 2013. <https://www.nia.nih.gov/health/alzheimers-disease-fact-sheet>.
- Arpaia, N., Campbell, C., Fan, X., Dikiy, S., van der Veeke, J., Liu, H., Cross, J.R., Pfeffer, K., Coffey, P.J., Rudensky, A.Y., 2013. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* 504 (7480), 451–455.
- Arumugam, M., Raes, J., Pelletier, E., Le Paslier, D., Yamada, T., Mende, D.R., Fernandes, G.R., Tap, J., Bruls, T., Batto, J.M., Bertalan, M., Borruel, N., Casellas, F., Fernandez, L., Gautier, L., Hansen, T., Hattori, M., Hayashi, T., Kleerebezem, M., Kurokawa, K., Leclerc, M., Levenez, F., Manichanh, C., Nielsen, H.B., Nielsen, T., Pons, N., Poulain, J., Qin, J., Sicheritz-Ponten, T., Tims, S., Torrents, D., Ugarte, E., Zoetendal, E.G., Wang, J., Guarner, F., Pedersen, O., de Vos, W.M., Brunak, S., Doré, J., Antolín, M., Artiguenave, F., Blottiere, H.M., Almeida, M., Brechot, C., Cara, C., Chervaux, C., Cultrone, A., Delorme, C., Denari, G., Dervyn, R., Foerstner, K.U., Friss, C., van de Guchte, M., Guedon, E., Haimet, F., Huber, W., van Hylckama-Vlieg, J., Jamet, A., Juste, C., Kaci, G., Knol, J., Lakhdari, O., Layec, S., Le Roux, K., Maguin, E., Mérioux, A., Melo Minardi, R., M'rim, C., Muller, J., Oozeer, R., Parkhill, J., Renault, P., Rescigno, M., Sanchez, N., Sunagawa, S., Torrejon, A., Turner, K., Vandemeulebrouck, G., Varela, E., Winogradsky, Y., Zeller, G., Weissenbach, J., Ehrlich, S.D., Bork, P., 2011. Enterotypes of the human gut microbiome. *Nature* 473 (7346), 174–180.
- Asti, A., Gioglio, L., 2014. Can a bacterial endotoxin be a key factor in the kinetics of amyloid fibril formation? *J. Alzheimers Dis.* 39 (1), 169–179.
- Bäckhed, F., 2011. Programming of host metabolism by the gut microbiota. *Ann. Nutr. Metab.* 58 (Suppl. 2), 44–52.
- Belkaid, Y., Hand, T.W., 2014. Role of the microbiota in immunity and inflammation. *Cell* 157, 121–141.
- Belmaker, R.H., Agam, G., 2008. Major depressive disorder. *N. Engl. J. Med.* 358, 55–68.
- Bengmark, S., 2013. Gut microbiota, immune development and function. *Pharmacol. Res.* 69 (1), 87–113.
- Bhargava, P., Mowry, E.M., 2014. Gut Microbiome and Multiple Sclerosis. *Curr. Neurol. Neurosci. Rep.* 14, 492.
- Biagi, E., Nylund, L., Candela, M., Ostan, R., Bucchi, L., Pini, E., Nikila, J., Monti, D., Satakori, R., Franceschi, C., Brigidi, P., 2010. Through Ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians. *PLoS One* 5 (5), e10667.
- Block, M.L., Zecca, L., Hong, J.S., 2007. Microglia-mediated neurotoxicity uncovering the molecular mechanisms. *Nat. Rev. Neurosci.* 8 (1), 57–69.
- Braak, H., Del Tredici, K., Rüb, U., de Vos, R.A., Jansen Steur, E.N., Braak, E., 2003. Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol. Aging* 24, 197–211.
- Braak, H., de Vos, R.A., Bohl, J., Del Tredici, K., 2006. Gastric α -synuclein immunoreactive inclusions in Meissner's and Auerbach's plexuses in cases staged for Parkinson's disease-related brain pathology. *Neurosci. Lett.* 396 (1), 67–72.
- Brenner, S.R., 2013. Blue-green algae or cyanobacteria in the intestinal micro-flora may produce neurotoxins such as Beta-N-Methylamino-L-Alanine (BMAA) which may be related to development of amyotrophic lateral sclerosis, Alzheimer's disease and Parkinson-Dementia-complex in humans and Equine Motor Neuron Disease in horses. *Med. Hypotheses* 80 (1), 103.
- Çamcı, G., Oğuz, S., 2016. Association between Parkinson's disease and Helicobacter Pylori. *J. Clin. Neurol.* 12 (2), 147–150.
- Cantarel, B.L., Waubant, E., Chehoud, C., Kuczynski, J., DeSantis, T.Z., Warrington, J., Venkatesan, A., Fraser, C.M., Mowry, E.M., 2015. Gut microbiota in multiple sclerosis: possible influence of immunomodulators. *J. Invest. Med.* 63, 729–734.
- Cassani, E., Privitera, G., Pezzoli, G., Pusani, C., Madio, C., Iorio, L., Barichella, M., 2011. Use of probiotics for the treatment of constipation in Parkinson's disease patients. *Minerva Gastroenterol. Dietol.* 57 (2), 117–121.
- Catanzaro, R., Anzalone, M., Calabrese, F., Milazzo, M., Capuana, M., Italia, A., Occhipinti, S., Marotta, F., 2015. The gut microbiota and its correlations with the central nervous system disorders. *Panminerva Med.* 57 (3), 127–143.
- Cattaneo, A., Cattane, N., Galluzzi, S., Provasi, S., Lopizzo, N., Festari, C., Ferrari, C., Guerra, U.P., Paghera, B., Muscio, C., Bianchetti, A., 2017. Association of brain amyloidosis with pro-inflammatory gut bacterial taxa and peripheral inflammation markers in cognitively impaired elderly. *Neurobiol. Aging* 49, 60–68.
- Cekanaviciute, E., Debelius, J.W., Singh, S., Rúnia, T., Nelson, C., Yoo, B., Kanner, R., Crabtree-Hartman, E., Mazmanian, S., Knight, R., Katz Sand, I., 2016. Gut dysbiosis is a feature of MS and it is characterized by bacteria able to regulate lymphocyte differentiation in vitro. *Mult. Scler.* 22 (S3), 58–59.
- Cersosimo, M.G., Raina, G.B., Pecci, C., Pellene, A., Calandra, C.R., Gutiérrez, C., Micheli, F.E., Benarroch, E.E., 2013. Gastrointestinal manifestations in Parkinson's disease: prevalence and occurrence before motor symptoms. *J. Neurol.* 260 (5), 1332–1338.
- Chen, H., O'Reilly, E.J., Schwarzschild, M.A., Ascherio, A., 2007. Peripheral inflammatory biomarkers and risk of Parkinson's disease. *Am. J. Epidemiol.* 167 (1), 90–95.
- Chen, J., Chia, N., Kalari, K.R., Yao, J.Z., Novotna, M., Soldan, M.M.P., Luckey, D.H., Marietta, E.V., Jeraldo, P.R., Chen, X., Weinschenker, B.G., Rodriguez, M., Kantarci, O.H., Nelson, H., Muray, A.J., Mangalam, A.K., 2016. Multiple sclerosis patients have a distinct gut microbiota compared to healthy controls. *Sci. Rep.* 6, 28484.
- Chung, H., Kasper, D.L., 2010. Microbiota-stimulated immune mechanisms to maintain gut homeostasis. *Curr. Opin. Immunol.* 22, 455–460.
- Clairembault, T., Leclaire-Visonneau, L., Coron, E., Bourreille, A., Le Dily, S., Vavasseur, F., Heymann, M.F., Neunlist, M., Derkinderen, P., 2015. Structural alterations of the intestinal epithelial barrier in Parkinson's disease. *Acta Neuropathol. Commun.* 3 (1), 1.
- Collins, S.M., Bercik, P., 2009. The relationship between intestinal microbiota and the central nervous system in normal gastrointestinal function and disease. *Gastroenterology* 136, 2003–2014.
- Compston, A., Coles, A., 2008. Multiple sclerosis. *Lancet* 372, 1502–1517.
- Dickson, D.W., Fujishiro, H., Orr, C., DelleDonne, A., Josephs, K.A., Frigerio, R., Burnett, M., Parisi, J.E., Klos, K.J., Ahlskog, J.E., 2009. Neuropathology of nonmotor features of Parkinson disease. *Parkinsonism Relat. Disord.* 15, S1–S5.
- Dobbs, R.J., Charlett, A., Purkiss, A.G., Dobbs, S.M., Weller, C., Peterson, D.W., 1999. Association of circulating TNF-alpha and IL-6 with ageing and parkinsonism. *Acta Neurol. Scand.* 100 (1), 34–41.
- Doens, D., Fernández, P.L., 2014. Microglia receptors and their implications in the response to amyloid β for Alzheimer's disease pathogenesis. *J. Neuroinflammation* 11 (1), 48.
- Dorca-Arévalo, J., Soler-Jover, A., Gibert, M., Popoff, M.R., Martín-Satué, M., Blasi, J., 2008. Binding of epsilon-toxin from *Clostridium perfringens* in the nervous system. *Vet. Microbiol.* 131 (1–2), 14–25.
- Dutta, G., Zhang, P., Liu, B., 2008. The lipopolysaccharide Parkinson's disease animal model: mechanistic studies and drug discovery. *Fundam. Clin. Pharmacol.* 22 (5), 453–464.
- Eckburg, P.B., Bik, E.M., Bernstein, C.N., Purdom, E., Dethlefsen, L., Sargent, M., Gill, S.R., Nelson, K.E., Relman, D.A., 2005. Diversity of the human intestinal microbial flora. *Science* 308 (5728), 1635–1638.
- Edwards, L.L., Quigley, E.M., Pfeiffer, R.F., 1992. Gastrointestinal dysfunction in Parkinson's disease: frequency and pathophysiology. *Neurology* 42 (4), 726–732.
- El Kaoutari, A., Armougom, F., Gordon, J.I., Raouf, D., Henrissat, B., 2013. The abundance and variety of carbohydrate active enzymes in the human gut microbiota. *Nat. Rev. Microbiol.* 11, 497–504.
- Elahy, M., Jackaman, C., Mamo, J.C., Lam, V., Dhaliwal, S.S., Giles, C., Nelson, D., Takechi, R., 2015. Blood-brain barrier dysfunction developed during normal aging is associated with inflammation and loss of tight junctions but not with leukocyte recruitment. *Immun. Ageing* 12, 2.
- Ezendam, J., De Klerk, A., Gremmer, E.R., Van Loveren, H., 2008. Effects of *Bifidobacterium animalis* administered during lactation on allergic and autoimmune responses in rodents. *Clin. Exp. Immunol.* 154 (3), 424–431.
- Fang, X., 2015. Potential role of gut microbiota and tissue barriers in Parkinson's disease and amyotrophic lateral sclerosis. *Int. J. Neurosci.* 126 (9), 771–776.
- Fang, X., Wang, X., Yang, S., Meng, F., Wang, X., Wei, H., Chen, T., 2016. Evaluation of the microbial diversity in amyotrophic lateral sclerosis using high-throughput sequencing. *Front. Microbiol.* 7, 1479.
- Fasano, A., Bove, F., Gabrielli, M., Petracca, M., Zocco, M.A., Ragazzoni, E., Barbaro, F., Piano, C., Fortuna, S., Tortora, A., Di Giacopo, R., 2013. The role of small intestinal bacterial overgrowth in Parkinson's disease. *Mov. Disord.* 28 (9), 1241–1249.
- Finnie, J.W., Blumbergs, P.C., Manavis, J., 1999. Neuronal damage produced in rat brains by *Clostridium perfringens* type D epsilon toxin. *J. Comp. Pathol.* 120, 415–420.
- Forster, C., 2008. Tight junctions and the modulation of barrier function in disease. *Histochem. Cell Biol.* 130 (1), 55–70.
- Forsyth, C.B., Shannon, K.M., Kordower, J.H., Voigt, R.M., Shaikh, M., Jaglin, J.A., Estes, J.D., Dodiya, H.B., Keshavarzian, A., 2011. Increased intestinal permeability correlates with sigmoid mucosa alpha-synuclein staining and endotoxin exposure markers in early Parkinson's disease. *PLoS One* 6 (12), e28032.
- Forsythe, P., Sudo, N., Dinan, T., Taylor, V.H., Bienenstock, J., 2009. Mood and gut feelings. *Brain Behav. Immun.* 24 (1), 9–16.

- Forsythe, P., Bienenstock, J., Kunze, W.A., 2014. Vagal pathways for microbiome-brain-gut axis communication. *Adv. Exp. Med. Biol.* 817, 115–133.
- Fouhy, F., Ross, R.P., Fitzgerald, G.F., Stanton, C., Cotter, P.D., 2012. Composition of the early intestinal microbiota: knowledge, knowledge gaps and the use of high-throughput sequencing to address these gaps. *Gut Microbes* 3 (3), 203–220.
- Franceschi, C., 2007. Inflammaging as a major characteristic of old people: can it be prevented or cured? *Nutr. Rev.* 65, S173–S176.
- Franceschi, C., Bonafè, M., Valensin, S., Olivieri, F., De Luca, M., Ottaviani, E., De Benedictis, G., 2000. Inflamm-aging: an evolutionary perspective on immunosenescence. *Ann. N. Y. Acad. Sci.* 908 (1), 244–254.
- Frank, D., St Amand, A., Feldman, R., Boedeker, E., Harpaz, N., Pace, N., 2007. Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. *Proc. Natl. Acad. Sci. U. S. A.* 104, 13780–13785.
- Freedman, S.N., Shahi, S.K., Mangalam, A.K., 2018. The “gut feeling”: breaking down the role of gut microbiome in multiple sclerosis. *Neurotherapeutics* 1–7.
- Friedland, R.P., 2015. Mechanisms of molecular mimicry involving the microbiota in neurodegeneration. *J. Alzheimers Dis.* 45 (2), 349–362.
- Garbuzova-Davis, S., Sanberg, P.R., 2014. Blood-CNS barrier impairment in ALS patients versus an animal model. *Front. Cell. Neurosci.* 8, 21.
- Gareau, M.G., Wine, E., Rodrigues, D.M., Cho, J.H., Whary, M.T., Philpott, D.J., Macqueen, G., Sherman, P.M., 2011. Bacterial infection causes stress-induced memory dysfunction in mice. *Gut* 60, 307–317.
- Girolamo, F., Coppola, C., Ribatti, D., 2017. Immunoregulatory effect of mast cells influenced by microbes in neurodegenerative diseases. *Brain Behav. Immun.* 65, 68–89.
- Guarner, F., Malagelada, J.R., 2003. Gut flora in health and disease. *Lancet* 361 (9356), 512–519.
- Guinane, C.M., Cotter, P.D., 2013. Role of the gut microbiota in health and chronic gastrointestinal disease: understanding a hidden metabolic organ. *Ther. Adv. Gastroenterol.* 6 (4), 295–308.
- Guo, S., Al-Sadi, R., Said, H.M., Ma, T.Y., 2013. Lipopolysaccharide causes an increase in intestinal tight junction permeability in vitro and in vivo by inducing enterocyte membrane expression and localization of TLR-4 and CD14. *Am. J. Pathol.* 182 (2), 375–387.
- Heneka, M.T., Carson, M.J., El Khoury, J., Landreth, G.E., Brosseron, F., Feinstein, D.L., Jacobs, A.H., Wyss-Coray, T., Vitorica, J., Ransohoff, R.M., Herrup, K., 2015. Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* 14 (4), 388–405.
- Hu, X., Wang, T., Jin, F., 2016. Alzheimer's disease and gut microbiota. *Sci. China Life Sci.* 59 (10), 1006–1023.
- Hughes, L.E., Smith, P.A., Bonell, S., Natt, R.S., Wilson, C., Rashid, T., Amor, S., Thompson, E.J., Croker, J., Ebringer, A., 2003. Cross-reactivity between related sequences found in *Acinetobacter* sp., *Pseudomonas aeruginosa*, myelin basic protein and myelin oligodendrocyte glycoprotein in multiple sclerosis. *J. Neuroimmunol.* 144 (1–2), 105–115.
- Ithzaki, R.F., Lathe, R., Balin, B.J., Ball, M.J., Bearer, E.L., Braak, H., Bullido, M.J., Carter, C., Clerici, M., Cosby, S.L., Del Tredici, K., 2016. Microbes and Alzheimer's disease. *J. Alzheimers Dis.* 51 (4), 979–984.
- Jangi, S., Gandhi, R., Cox, L.M., Li, N., Von Glehn, F., Yan, R., Patel, B., Mazzola, M.A., Liu, S., Glanz, B.L., Cook, S., 2016. Alterations of the human gut microbiome in multiple sclerosis. *Nat. Commun.* 7, 12015.
- Jiang, C., Li, G., Huang, P., Liu, Z., Zhao, B., 2017. The gut microbiota and Alzheimer's disease. *J. Alzheimers Dis.* 58 (1), 1–5.
- Joshi, D., Roy, S., Banerjee, S., 2018. Prebiotics: a functional food in health and disease. In: Mandal, S.C., Mandal, V., Konishi, T. (Eds.), *Natural Products & Drug Discovery*. Elsevier, Amsterdam, pp. 507–523.
- Karlsson, O., Roman, E., Berg, A.L., Brittebo, E.B., 2011. Early hippocampal cell death, and late learning and memory deficits in rats exposed to the environmental toxin BMAA (β -N-methylamino-L-alanine) during the neonatal period. *Behav. Brain Res.* 219 (2), 310–320.
- Keshavazian, A., Green, S.J., Engen, P.A., Voigt, R.M., Naqib, A., Forsyth, C.B., Mutlu, E., Shanon, K.M., 2015. Colonic bacterial composition in Parkinson's Disease. *Mov. Disord.* 30 (10), 1351–1360.
- Kiebertz, K., Wunderlich, K.B., 2013. Parkinson's disease: evidence for environmental risk factors. *Mov. Disord.* 28 (1), 8–13.
- Kwon, H.K., Kim, G.C., Kim, Y., Hwang, W., Jash, A., Sahoo, A., Kim, J.E., Nam, J.H., Im, S.H., 2013. Amelioration of experimental autoimmune encephalomyelitis by probiotic mixture is mediated by a shift in T helper cell immune response. *Clin. Immunol.* 146 (3), 217–227.
- Lavasani, S., Dzhambazov, B., Nouri, M., Fåk, F., Buske, S., Molin, G., Thorlacius, H., Alenfall, J., Jeppsson, B., Weström, B., 2010. A novel probiotic mixture exerts a therapeutic effect on experimental autoimmune encephalomyelitis mediated by IL-10 producing regulatory T cells. *PLoS One* 5 (2), e9009.
- Lei, E., Vacy, K., Boon, W.C., 2016. Fatty acids and their therapeutic potential in neurological disorders. *Neurochem. Int.* 95, 75–84.
- Ley, R., Turnbaugh, P., Klein, S., Gordon, J., 2006. Microbial ecology: human gut microbes associated with obesity. *Nature* 444, 1022–1023.
- Liang, S., Wang, T., Hu, X., Luo, J., Li, W., Wu, X., Duan, Y., Jin, F., 2015. Administration of *Lactobacillus helveticus* NS8 improves behavioral, cognitive, and biochemical aberrations caused by chronic restraint stress. *Neuroscience* 310, 561–577.
- Lien, E., Means, T.K., Heine, H., Yoshimura, A., Kusumoto, S., Fukase, K., Fenton, M.J., Oikawa, M., Qureshi, N., Monks, B., Finberg, R.W., 2000. Toll-like receptor 4 imparts ligand-specific recognition of bacterial lipopolysaccharide. *J. Clin. Invest.* 105 (4), 497–504.
- Liu, Y., Walter, S., Stagi, M., Cherny, D., Letiembre, M., Schulz-Schaeffer, W., Heine, H., Penke, B., Neumann, H., Fassbender, K., 2005. LPS receptor (CD14): a receptor for phagocytosis of Alzheimer's amyloid peptide. *Brain* 128 (8), 1778–1789.
- Lonchamp, E., Dupont, J.L., Wioland, L., Courjaret, R., Mbebi-Liegeois, C., Jover, E., Doussau, F., Popoff, M.R., Bossu, J.L., De Barry, J., Poulain, B., 2010. *Clostridium perfringens* epsilon toxin targets granule cells in the mouse cerebellum and stimulates glutamate release. *PLoS One* 5 (9), e13046.
- Lukiw, W.J., 2016. *Bacteroides fragilis* lipopolysaccharide and inflammatory signaling in Alzheimer's disease. *Front. Microbiol.* 7, 1544.
- Maassen, C.B., Claassen, E., 2008. Strain-dependent effects of probiotic lactobacilli on EAE autoimmunity. *Vaccine* 26 (17), 2056–2057.
- Marques, T.M., Wall, R., Ross, R.P., Fitzgerald, G.F., Ryan, C.A., Stanton, C., 2010. Programming infant gut microbiota: influence of dietary and environmental factors. *Curr. Opin. Biotechnol.* 21 (2), 149–156.
- Metz, A., Garcia, J., Ortega, J., Lane, M., Scholes, S., Uzal, F.A., 2013. Brain lesions associated with *Clostridium perfringens* type D epsilon toxin in a Holstein heifer calf. *Vet. Pathol.* 50 (5), 765–768.
- Miyake, S., Kim, S., Suda, W., Oshima, K., Nakamura, M., Matsuoka, T., Chihara, N., Tomita, A., Sato, W., Kim, S.W., Morita, H., Hattori, M., Yamamura, T., 2015. Dysbiosis in the Gut Microbiota of patients with Multiple Sclerosis, with a striking Depletion of Species Belonging to Clostridia XIVa and IV Clusters. *PLoS One* 10 (9), e0137429.
- Möhle, L., Mattei, D., Heimesaat, M.M., Bereswill, S., Fischer, A., Alutis, M., French, T., Hambardzumyan, D., Matzinger, P., Dunay, I.R., Wolf, S.A., 2016. Ly6Chi monocytes provide a link between antibiotic-induced changes in gut microbiota and adult hippocampal neurogenesis. *Cell Rep.* 15 (9), 1945–1956.
- Mowry, E.M., Glenn, J.D., 2018. The dynamics of the gut microbiome in multiple sclerosis in relation to disease. *Neurol. Clin.* 36 (1), 185–196.
- Mowry, E.M., Waubant, E., Chehoud, C., DeSantis, T., Kuczynski, J., Warrington, J., 2012. Gut bacterial populations in multiple sclerosis and in health. *Neurology* 78, P05.106.
- Mulak, A., Bonaz, B., 2015. Brain-gut-microbiota axis in Parkinson's disease. *World J. Gastroenterol.* 21 (37), 10609–10620.
- Nair, A.T., Ramachandran, V., Joghee, N.M., Antony, S., Ramalingam, G., 2018. Gut Microbiota Dysfunction as Reliable Non-invasive early Diagnostic Biomarkers in the Pathophysiology of Parkinson's disease: a critical Review. *J. Neurogastroenterol. Motil.* 24 (1), 30–42.
- Niehaus, I., Lange, J.H., 2003. Endotoxin: is it an environmental factor in the cause of Parkinson's disease? *Occup. Environ. Med.* 60 (5), 378.
- Noseworthy, J.H., Lucchinetti, C., Rodriguez, M., Weinshenker, B.G., 2000. Multiple sclerosis. *N. Engl. J. Med.* 343, 938–952.
- Ouweland, A.C., Bergsma, N., Parhiala, R., Lahtinen, S., Gueimonde, M., Finne-Soveri, H., Strandberg, T., Pitkälä, K., Salminen, S., 2008. Bifidobacterium microbiota and parameters of immune function in elderly subjects. *FEMS Immunol. Med. Microbiol.* 53 (1), 18–25.
- Parashar, A., Udayabanu, M., 2017. Gut microbiota: implications in Parkinson's disease. *Parkinsonism Relat. Disord.* 38, 1–7.
- Pistolato, F., Sumalla Cano, S., Elio, I., Masias Vergara, M., Giampieri, F., Battino, M., 2016. Role of gut microbiota and nutrients in amyloid formation and pathogenesis of Alzheimer disease. *Nutr. Rev.* 74 (10), 624–634.
- Pompei, A., Cordisco, L., Amaretti, A., Zanoni, S., Matteuzzi, D., Rossi, M., 2007. Folate production by bifidobacteria as a potential probiotic property. *Appl. Environ. Microbiol.* 73 (1), 179–185.
- Pryde, S.E., Duncan, S.H., Hold, G.L., Stewart, C.S., Flint, H.J., 2002. The microbiology of butyrate formation in the human colon. *FEMS Microbiol. Lett.* 217 (2), 133–139.
- Qin, J., Li, R., Raes, J., Arumugam, M., Burgdorf, K.S., Manichanh, C., Nielsen, T., Pons, N., Levenez, F., Yamada, T., Mende, D.R., 2010. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* 464 (7285), 59–65.
- Quigley, E.M., 2013. Gut bacteria in health and disease. *Gastroenterol. Hepatol. (NY)* 9 (9), 560–569.
- Quigley, E.M., 2017. Microbiota-brain-gut axis and neurodegenerative diseases. *Curr. Neurol. Neurosci. Rep.* 17 (12), 94.
- Ransohoff, R.M., 2016. How neuroinflammation contributes to neurodegeneration. *Science* 353 (6301), 777–783.
- Reale, M., Iarlori, C., Thomas, A., Gambi, D., Perfetti, B., Di Nicola, M., Onofri, M., 2009. Peripheral cytokines profile in Parkinson's disease. *Brain Behav. Immun.* 23 (1), 55–63.
- Ríos-Covián, D., Ruas-Madiedo, P., Margolles, A., Gueimonde, M., de Los Reyes-Gavilán, C.G., Salazar, N., 2016. Intestinal short chain fatty acids and their link with diet and human health. *Front. Microbiol.* 7, 185.
- Rite, I., Machado, A., Cano, J., Venero, J.L., 2007. Blood-brain barrier disruption induces in vivo degeneration of nigral dopaminergic neurons. *J. Neurochem.* 101 (6), 1567–1582.
- Rumah, K.R., Linden, J., Fischetti, V.A., Vartanian, T., 2013. Isolation of *Clostridium perfringens* type B in an individual at first clinical presentation of multiple sclerosis provides clues for environmental triggers of the disease. *PLoS One* 8 (10), e76359.
- Savica, R., Carlin, J.M., Grossardt, B.R., Bower, J.H., Ahlskog, J.E., Maraganore, D.M., Bharucha, A.E., Rocca, W.A., 2009. Medical records documentation of constipation preceding Parkinson disease a case-control study. *Neurology* 73 (21), 1752–1758.
- Savignac, H.M., Tramullas, M., Kiely, B., Dinan, T.G., Cryan, J.F., 2015. Bifidobacteria modulate cognitive processes in an anxious mouse strain. *Behav. Brain Res.* 287, 59–72.
- Scheperjans, F., Aho, V., Pereira, P.A., Koskinen, K., Paulin, L., Pekkonen, E., Haapaniemi, E., Kaakkola, S., Eerola-Rautio, J., Pohja, M., Kinnunen, E., 2015. Gut microbiota are related to Parkinson's disease and clinical phenotype. *Mov. Disord.* 30 (3), 350–358.
- Schirmer, M., Smeekens, S.P., Vlamakis, H., Jaeger, M., Oosting, M., Franzosa, E.A., Horst, R.T., Jansen, T., Jacobs, L., Bonder, M.J., Kurilshikov, A., 2016. Linking the human gut microbiome to inflammatory cytokine production capacity. *Cell* 167 (4), 1125–1136.
- Sekirov, I., Russell, S.L., Antunes, L.C., Finlay, B.B., 2010. Gut microbiota in health and disease. *Physiol. Rev.* 90 (3), 859–904.

- Shahi, S.K., Freedman, S.N., Mangalam, A.K., 2017. Gut microbiome in multiple sclerosis: the players involved and the roles they play. *Gut Microbes* 8 (6), 607–615.
- Shannon, K.M., Keshavarzian, A., Dodiya, H.B., Jakate, S., Kordower, J.H., 2012. Is alpha-synuclein in the colon a biomarker for premotor Parkinson's disease? Evidence from 3 cases. *Mov. Disord.* 27 (6), 716–719.
- Sivieri, K., Morales, M.L.V., Adorno, M.A.T., Sakamoto, I.K., Saad, S.M.I., Rossi, E.A., 2013. *Lactobacillus acidophilus* CRL 1014 improved "gut health" in the SHIME® reactor. *BMC Gastroenterol.* 13 (1), 100.
- Stilling, R.M., Dinan, T.G., Cryan, J.F., 2014. Microbial genes, brain & behaviour—epigenetic regulation of the gut–brain axis. *Genes Brain Behav.* 13 (1), 69–86.
- Sui, Y.T., Bullock, K.M., Eickson, M.A., Zhang, J., Banks, W.A., 2014. Alpha synuclein is transported into and out of the brain by the blood-brain barrier. *Peptides* 62, 197–202.
- Surwase, S.N., Jadhav, J.P., 2011. Bioconversion of L-tyrosine to L-DOPA by a novel bacterium *Bacillus* sp. *JPJ. Amino Acids* 41 (2), 495–506.
- Takata, K., Kinoshita, M., Okuno, T., Moriya, M., Kohda, T., Honorat, J.A., Sugimoto, T., Kumanogoh, A., Kayama, H., Takeda, K., Sakoda, S., 2011. The lactic acid bacterium *Pediococcus acidilactici* suppresses autoimmune encephalomyelitis by inducing IL-10-producing regulatory T cells. *PLoS One* 6 (11), e27644.
- Tan, A.H., Mahadeva, S., Thalha, A.M., Gibson, P.R., Kiew, C.K., Yeat, C.M., Ng, S.W., Ang, S.P., Chow, S.K., Tan, C.T., Yong, H.S., 2014. Small intestinal bacterial overgrowth in Parkinson's disease. *Parkinsonism Relat. Disord.* 20 (5), 535–540.
- Toepfer, M., Schroeder, M., Klauser, A., Lochmüller, H., Hirschmann, M., Riepl, R.L., Pongratz, D., Müller-Felber, W., 1997. Delayed colonic transit times in amyotrophic lateral sclerosis assessed with radio-opaque markers. *Eur. J. Med. Res.* 2 (11), 473–476.
- Tremlett, H., Fadrosh, D.W., Faruqi, A.A., Hart, J., Roalstad, S., Graves, J., Lynch, S., Waubant, E., 2016. Gut microbiota composition and relapse risk in pediatric MS: a pilot study. *J. Neurol. Sci.* 363, 153–157.
- Tsuang, D., Leverenz, J.B., Lopez, O.L., Hamilton, R.L., Bennett, D.A., Schneider, J.A., Buchman, A.S., Larson, E.B., Crane, P.K., Kaye, J.A., Kramer, P., Woltjer, R., Trojanowski, J.Q., Weintraub, D., Chen-Plotkin, A.S., Irwin, D.J., Rick, J., Schellenberg, G.D., Watson, G.S., Kukull, W., Nelson, P.T., Jicha, G.A., Neltner, J.H., Galasko, D., Masliah, E., Quinn, J.F., Chung, K.A., Yearout, D., Mata, I.F., Wan, J.Y., Edwards, K.L., Montine, T.J., Zabetian, C.P., 2013. APOE ϵ 4 increases risk for dementia in pure synucleinopathies. *JAMA Neurol.* 70 (2), 223–228.
- Tufekci, K.U., Genc, S., Genc, K., 2011. The endotoxin-induced neuroinflammation model of Parkinson's disease. *Parkinsons Dis.* 2011.
- Ulluwishewa, D., Anderson, R.C., McNabb, W.C., Moughan, P.J., Wells, J.M., Roy, N.C., 2011. Regulation of tight junction permeability by intestinal bacteria and dietary components. *J. Nutr.* 141 (5), 769–776.
- Vital, M., Howe, A.C., Tiedje, J.M., 2014. Revealing the bacterial butyrate synthesis pathways by analyzing (meta) genomic data. *MBio* 5 (2), 1–11.
- Wang, T., Hu, X., Liang, S., Li, W., Wu, X., Wang, L., Jin, F., 2015a. *Lactobacillus fermentum* NS9 restores the antibiotic induced physiological and psychological abnormalities in rats. *Benef. Microbes* 6 (5), 707–717.
- Wang, I.K., Wu, Y.Y., Yang, Y.F., Ting, I.W., Lin, C.C., Yen, T.H., Chen, J.H., Wang, C.H., Huang, C.C., Lin, H.C., 2015b. The effect of probiotics on serum levels of cytokine and endotoxin in peritoneal dialysis patients: a randomised, double-blind, placebo-controlled trial. *Benef. Microbes* 6 (4), 423–430.
- Wright, M.L., Fournier, C., Houser, M.C., Tansey, M., Glass, J., Hertzberg, V.S., 2018. Potential role of the gut microbiome in ALS: a systematic review. *Biol. Res. Nurs.* 1–9.
- Wu, S., Yi, J., Zhang, Y.G., Zhou, J., Sun, J., 2015. Leaky intestine and impaired microbiome in an amyotrophic lateral sclerosis mouse model. *Physiol. Rep.* 3 (4).
- Zhang, L.S., Davies, S.S., 2016. Microbial metabolism of dietary components to bioactive metabolites: opportunities for new therapeutic interventions. *Genome Med.* 8 (1), 46.
- Zhang, Y.G., Wu, S., Yi, J., Xia, Y., Jin, D., Zhou, J., Sun, J., 2017. Target intestinal microbiota to alleviate disease progression in amyotrophic lateral sclerosis. *Clin. Ther.* 39 (2), 322–336.
- Zhao, Y., Dua, P., Lukiw, W.J., 2015. Microbial sources of amyloid and relevance to amyloidogenesis and Alzheimer's disease (AD). *J. Alzheimers Dis. Parkinsonism* 5 (1), 177–190.
- Zhou, Y., Lu, Y., Fang, X., Zhang, J., Li, J., Li, S., Deng, X., Yu, Y., Xu, R., 2015. An astrocyte regenerative response from vimentin-containing cells in the spinal cord of amyotrophic lateral sclerosis's disease-like transgenic (G93A SOD1) mice. *Neurodegener. Dis.* 15 (1), 1–12.