

Osteoarthritis and Cartilage

Review

Gut microbiota and obesity-associated osteoarthritis

Y. Liu † ||, W. Ding ‡ ||, H.L. Wang §, L.L. Dai ||, W.H. Zong ||, Y.Z. Wang ||, J. Bi ||, W. Han ||, G.J. Dong † || *

† School of Kinesiology, Shanghai University of Sport, Shanghai, China

‡ Department of Construction and Real Estate Management, Laiyang, China

§ College of Physical Education, Henan Normal University, Xinxiang, China

|| Shandong Sport University, Jinan, China



ARTICLE INFO

Article history:

Received 15 December 2018

Accepted 18 May 2019

Keywords:

Gut microbiota

Obesity

Osteoarthritis

SUMMARY

Obesity is a well-known primary risk factor for osteoarthritis (OA). In recent decades, the biomechanics-based theoretical paradigm for the pathogenesis of obesity-associated OA has been gradually but fundamentally modified. This modification is a result of accumulating evidence that biological factors also contribute to the etiology of the disease. The gut microbiota is a complicated ecosystem that profoundly influences the health of the host and can be modulated by the combined effects of environmental stimuli and genetic factors. Recently, enteric dysbacteriosis has been identified as a causal factor in the initiation and propagation of obesity-associated OA in animal models. Gut microbes and their components, microbe-associated lipid metabolites, and OA interact at both systemic and local levels through mechanisms that involve interplay with the innate immune system. However, the demonstration of causality in humans will require further studies. Nonetheless, probiotics, prebiotics, dietary habits and exercise, which aid the restoration of a healthy microbial community, are potential therapeutic approaches in the treatment of obesity-associated OA.

© 2019 Osteoarthritis Research Society International. Published by Elsevier Ltd. All rights reserved.

Introduction

Osteoarthritis (OA), which is the most prevalent arthritic disease and one of the leading causes of disability, is no longer considered as a degenerative illness of the cartilage but as a disorder of the whole joint with a heterogeneous and multifaceted etiology¹. Obesity is a predisposing risk factor in OA initiation and perpetuation^{2,3}. The pathophysiological mechanisms that underpin this close link have not been fully elucidated, but microbial causes of gut origin have been proposed.

The human body is not a self-sufficient and isolated unit, but a diverse and dynamic ecosystem. The intestinal canal is an excellent culture medium harboring approximately 10¹⁴ microorganisms, which contains microbial genetic information that represents an extension of the host's metagenomes and, thereby, influences the

individual's health status^{4,5}. In turn, the gut microbiome can be shaped by the combined effects of environmental stimuli and genetic factors⁶. The crosstalk among the environment, the gut microbiota and the host is a fundamental determinant of metabolic and immune physiology or pathology^{7,8}. Interference of the delicate balance of homeostatic mechanisms caused by nutrient overload, microbial agents or immune dysfunction underlies many diseases^{9,10}. Links have been demonstrated between dysbiosis of the gut microbiota and rheumatoid arthritis (RA) in animal models and humans¹¹. Furthermore, the involvement of the gut microbiota in obesity-associated OA is being elucidated.

Hence, we review the key findings on the association between obesity and OA in a historical context and summarize the evidence that gut microbes, their components, and microbe-associated lipid metabolites contribute mechanistically to the etiology and pathogenesis of obesity-associated OA (Fig. 1). Additionally, we review the potential therapeutic approaches for gut microbial ecosystem reshaping in the prevention and treatment of OA.

Obesity and OA

The concept that OA is a wear-and-tear form of arthritis resulting from years of long, hard use has been proposed for

Abbreviations: OA, osteoarthritis; RA, rheumatoid arthritis; MetS, metabolic syndrome; HFD, high-fat diet; LPS, lipopolysaccharides; PGN, peptidoglycan; FFAs, free fatty acids; PRRs, pattern recognition receptors; TLRs, Toll-like receptors; NLRs, NOD-like receptors; PGN-PS, peptidoglycan-polysaccharide.

* Address correspondence and reprint requests to: G.J. Dong, Shandong Sport University, 10600 Century Avenue, Jinan, 250102, China. Tel: 86-531-89655208; Fax: 86-531-89655208.

E-mail address: donggj@163.com (G.J. Dong).

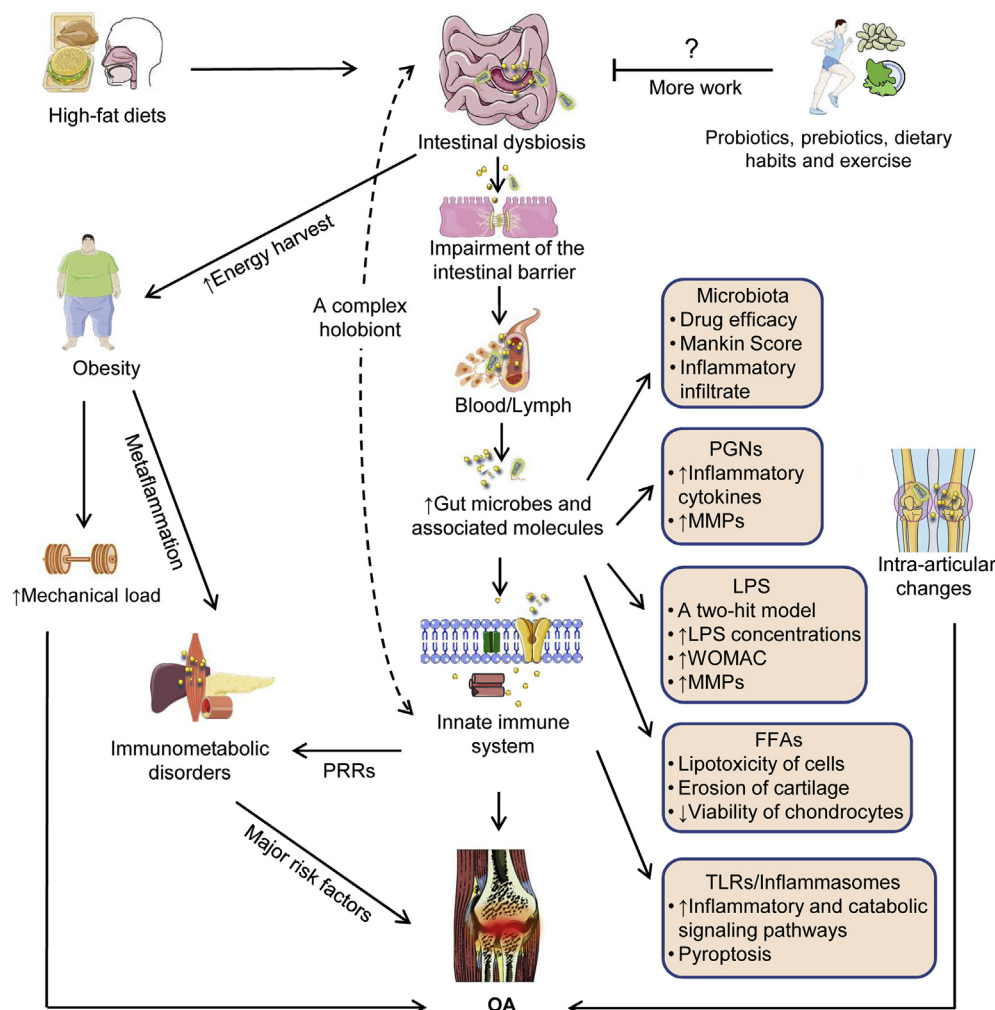


Fig. 1. A schematic illustration of the proposed mechanisms underlying the gut microbiome–osteoarthritis (OA) axis, and potential therapeutic approaches involving altering the microbiota composition. Abbreviations: LPS, lipopolysaccharides; PGNs, peptidoglycans; FFAs, free fatty acids; PRRs, pattern recognition receptors; TLRs, Toll-like receptors; MMPs, matrix metalloproteases; WOMAC, Western Ontario and McMaster Universities Osteoarthritis Index.

decades¹². However, obesity significantly increases the risk of developing OA in nonweight-bearing joints, such as hand and temporomandibular joints, thereby indicating that obesity influences OA in a nonmechanical manner^{2,3}. However, metabolic and mechanical factors are inextricably intertwined in weight-bearing joints. Thus, demonstrating the net effects of these factors without reciprocal interference *in vivo* is difficult. Nonetheless, some animal studies with ingenious designs demonstrated that body fat instead of body mass was highly correlated with the radiographic and symptomatic severities of knee OA (KOA) caused by a high-fat diet (HFD) or sucrose diet^{13–15}. Metabolic disturbance due to nutrient overload is a comorbidity factor in OA-related osteophyte formation and cartilage degeneration that is independent of weight¹⁶. In addition to systemic factors, the locally hypertrophic infrapatellar fat pad is involved in cartilage degradation by secreting proinflammatory cytokines in obese mice¹⁷. However, a study performed by superimposing energy-dense diets on a tail-suspension mouse model found that infrapatellar fat pad inflammation and chondrocyte apoptosis were the results of metabolic stresses caused by an HFD alone, and the surface irregularity of articular cartilage and osteophyte formation resulted from both metabolic stresses and mechanical load¹⁸. An animal study using an *in vivo* tibial loading model indicated that severe adiposity and

systemic inflammation aggravated cartilage damage induced by mechanical load¹⁹. Collectively, these results support the determinative role of these biological contributions in OA etiology. Nonetheless, mechanical effects may be important in some cases.

The delineation of OA into distinguishable phenotypes on the basis of etiological underpinnings has attracted considerable interest. An OA subtype called metabolic OA displays a unique OA trajectory characterized by its major causative metabolic factors¹. The convergence of research into OA and metabolic syndrome (MetS) has yielded considerable information about their bidirectional effects. Epidemiological and experimental studies indicated the increased incidence of OA in patients with MetS^{13,20}. A large-scale cohort study provided solid evidence that the accumulation of MetS components (e.g., central obesity, hypertension, high triglyceride levels and impaired fasting glucose) and their clustering significantly increased the risk of KOA occurrence and progression²¹. A study of obese mice further suggested that improved glucose tolerance and the disrupted coexpression of inflammatory cytokines delayed the progression of structural damage in OA without lowering the inflammatory cytokine levels¹⁵. Moreover, OA is associated with the increased prevalence of MetS and its components, particularly in young individuals²². Regardless of some controversial findings^{23,24}, the bona fide association between MetS

and OA phenotypic outcomes is generally accepted, as this link persists after the body weight or body mass index is adjusted^{22,25}. Metabolic OA has even been proposed as a new criterion for the definition of MetS, which is supported by shared pathogenic mechanisms, such as inflammation, in the etiologies of MetS and OA²⁶.

Persistent inflammation is less pronounced in OA than in prototypical RA²⁷. This situation is reminiscent of the low-level sub-acute inflammation that occurs in obesity. Accumulating evidence suggests that obesity provides a permissive inflammatory setting in OA pathophysiology. The altered plasma levels of human C-reactive protein, which is a well-established biomarker for systemic inflammation, can be evoked by metabolic stresses and used as a prognostic biomarker for the estimation of individual susceptibility to developing OA later in life²⁸. Obesity also triggers a locally dynamic inflammatory response guided by predominant infiltration of proinflammatory M1-polarized macrophages in the OA synovium prior to cartilage degradation²⁹. Some proinflammatory cytokines and adipokines produced by adipocytes are highly involved in OA pathogenesis^{30,31}. A preclinical study found that improved metabolic inflammation substantially mitigated OA deterioration elicited by an HFD²⁸. All of these data indicate the causative role of metabolic inflammation.

To summarize briefly, the link between obesity and OA onset and progression is well-established, and MetS and metabolic inflammation are considered as major hidden risk factors that provide unifying mechanisms underlying this association.

Gut microbiota and immunometabolic disorder regulation

The interfaces between metabolism and immunity exist at multiple levels within a complex network and have important implications for physiological homeostasis^{7–10}. Gut microbes and microbe-associated molecules may be present in the intricate network involved in the pathogenesis and potentiation of immunometabolic disorders, such as MetS and metabolic inflammation.

Gut microbiota

Large-scale alterations in the gut microbiota, especially in the proportional representation of *Bacteroidetes* and *Firmicutes*, in leptin-deficient obese mice³² and the reverse changes during weight loss in obese individuals³³ suggest that microbiome configurations are correlated with host energy balance. A seminal study found that the colonization of germ-free mice with gut microbiota harvested from the cecum of conventionally raised donors induced a significant increase in epididymal fat weight and total body fat content even with decreased chow consumption³⁴. The overgrowth of a specific taxa, such as endotoxin-producing *Enterobacter*, has been identified as an etiological factor rather than an epiphenomenon in obesity³⁵.

A series of studies found that a Western diet-mediated gut microbiota was an instigator of MetS by activating multiple mechanisms or pathways in various organs and tissues³⁶. The modulation of MetS-associated phenotypes of the gut microbiota attenuates metabolic deterioration in mice fed an HFD³⁷. Specific taxonomic changes, such as the decreased abundance of *Akkermansia muciniphila*, in a holistic bacterial community result in adipose tissue inflammation and systemic dysmetabolism³⁸. A study of dietary intervention in obese human subjects highlighted the importance of restoring the gut microbiota in the amelioration of chronic inflammation³⁹.

These data provide evidence that an altered gut microbiota plays a robust causal role in the development of obesity and immunometabolic disorders.

Components from gut microbes and microbe-associated lipid metabolites

A number of leading theories about the role of the components of gut microbes and microbe-associated lipid metabolites as key molecules in relaying and transferring signals from the gut to systemic tissues have been proposed^{40–42}. These molecules are likely involved in the emergence and persistence of immunometabolic disorders.

Lipopolysaccharide (LPS), which is a major component of the outer membranes of Gram-negative bacteria, can provoke vigorous and generalized proinflammatory responses in infected hosts^{40,43}. The increased metabolic concentrations of plasma LPS (by two-to three-fold) due to microbiota dysbiosis and impairment of the intestinal epithelium barrier caused by an HFD are defined as metabolic endotoxemia, which can serve as a sufficient molecular mechanism for the initiation of obesity and hyperinsulinemia via an inflammation-mediated pathway⁴⁰. Meanwhile, the process of uptake and trafficking of gut-derived LPS is a coherent mechanism for the hypertrophy of adipose tissues because it can facilitate lipoprotein transcytosis through the endothelial barrier and endocytosis in adipocytes⁴⁴. Gut-derived LPS can augment adipose macrophage accumulation and skew the polarization of alternatively activated M2 macrophages toward proinflammatory M1 phenotypes⁴³.

Peptidoglycan (PGN) is also a component of bacterial cell walls. Bacterial PGNs derived from the intestinal microbiota can penetrate internal sites and induce immune and inflammatory responses⁴⁵. Specific PGN motifs can enhance the cytotoxicity of adipocytes by stimulating cell-autonomous lipolysis, which drives hyperlipidemia and systemic inflammation⁴¹. In addition to adipocytes, other types of metabolic cells, such as hepatocytes, can be activated by the mimetics of bacterial PGN and cause considerable whole-body insulin resistance⁴⁶.

Free fatty acids (FFAs) are released from adipocytes during triglyceride lipolysis and can be utilized by various tissues. An expanded adipose tissue mass can accelerate FFA efflux into the bloodstream⁴². FFAs are also bioactive molecules that link obesity and metabolic diseases by mechanisms involving the activation of oxidative stress and innate immune pathways⁴². Imbalanced microbial populations are associated with the considerably increased expression of key genes associated with FFA synthesis, FFA transport and immune regulation in the liver⁴⁷. Altered serum FFA profiles are related to specific gut microbial signatures, including imbalanced populations of *Akkermansia* and *Lactobacillus*⁴⁸.

Innate immunity

The effects of the gut microbiota on host infection/inflammation mainly rely on pattern recognition receptors (PRRs), particularly the Toll-like receptors (TLRs) and NOD-like receptors (NLRs)^{7,8}. In addition to their classical role in detecting pathogens and inducing innate immune responses, PRRs are emerging as key players in host metabolism programming. TLR4, a well-characterized membrane receptor, can be activated by both classical microorganism-derived toxins, such as LPS and PGNs, and endogenous metabolic cues, such as FFAs^{49,50}. This receptor also plays a major role in linking increased dietary consumption of lipids, body fat accumulation and intestinal dysbacteriosis with metabolic inflammation and insulin resistance^{49,50}. NLR proteins, which are cytosolic PRRs, are essential components of the inflammasome. This large, multimolecular immune complex assembles upon the recognition of infection or pathogens^{51,52}. Recently, the NLRP3 inflammasome has been identified as a remote control for MetS by modulating the prevalence of

colitogenic species of intestinal microflora and the influx of bacterial TLR agonists into portal circulation^{53,54}.

The gut microbiota and the innate immune system can work as a complex holobiont; the innate immune system can shape the gut microbiota, and the gut microbiota has immune-activating functions^{55,56}. Most perturbations in the microbiome or innate immune system can result in a new equilibrium for the pathogenesis of metabolic diseases⁵⁶. This idea was further validated by the finding that genetic modification of the NLRP3 inflammasome or TLR5 induced impaired glucose metabolism, which was closely related to an altered microbiota structure^{53,57,58}. Alterations in microbial community structure, elevation of TLR ligand levels and over-expression of TLRs were also observed in mice and humans with obesity or metabolic disorders⁵⁸.

Dysbacteriosis and OA

The concept that gut microbiota correlates with musculoskeletal health has been recognized for >60 years⁵⁹. Previous studies on animal models and humans revealed the role of the microbiota in the onset, severity and progression of RA⁶⁰. Although the importance of intestinal dysbiosis in either triggering or driving autoimmunity related to RA has been proposed¹¹, the mechanism underlying the gut microbiome–joint axis remains unclear. Evidence from animal studies indicates that the gut microbiota is also involved in the etiology of OA. Work on the gut microbiome–OA connection will provide a basis for the discovery of novel therapeutics for patients with chronic inflammatory arthritis.

Gut microbiota

The evidence for the efficacy of oral chondroitin sulfate in OA treatment is contentious and inconsistent⁶¹. A hypothesis paper proposed that the relative abundance of *A. muciniphila*, a commensal gut bacterium, might determine the contribution of chondroitin sulfate to the amelioration or aggravation of OA⁶². An animal study indicated that the altered abundance of the gut microbes *Lactobacillus* spp. and *Methanobrevibacter* spp. was associated with the Mankin scores of articular cartilage¹⁴. *Lactobacillus casei* can enhance the mitigative effects of type II collagen/glucosamine on OA treatment by downregulating the inflammatory responses of articular tissues in mice⁶³. In revolutionary initial research, Schott *et al.* provided direct evidence that obesity-associated OA was an inflammatory ailment driven by obesity-related gut dysbiosis⁶⁴. They found that reversal of a proinflammatory shift in the obese murine gut microbiota, through the prebiotic manipulation of specific microbial species, particularly *Bifidobacterium pseudolongum*, could reduce inflammatory profiles in the colon and circulation, thereby preserving the articular cartilage and essentially rescuing OA⁶⁴. Similar to a previous study showing that the intestinal epigenome and even gene expression were reprogrammed by an obesity-associated gut microbiome⁶⁵, this study indicated that alterations in the obesogenic diet-conditioned microbiota changed the colonic gene expression patterns associated with the support of intestinal cell types related to epithelial proliferation and barrier function; thus, endotoxin leakage and macrophage infiltration into the synovium were reduced⁶⁴. The therapeutic modification of dysbalanced microbial populations characterized by a MetS-like phenotype in TLR5 knockout mice can alleviate the OA cartilage pathology and subchondral bone morphology¹⁹. This finding further indicates the role of a person's enterotype in the development of metabolic OA.

A considerable number of both intracellular and extracellular bacteria were observed in the synovial tissues of patients with OA, which was associated with a substantial quantity of inflammatory

infiltrate⁶⁶. However, the exact source of these organisms is difficult to identify because many species are normal commensal residents both in the intestinal tract and skin⁶⁶. It seems reasonable to propose the gut as the source of these organisms because impaired gut permeability and mucosal competence have been observed in patients with arthritis^{66,67}, and some living bacteria could be transferred from the gut to the joints/enthesis through the circulatory or lymphatic system⁶⁸.

Components from gut microbes and microbe-associated lipid metabolites

A hypothesis article proposed the pivotal role of metabolic endotoxemia in cross-sectional associations among gut dysbiosis, systemic inflammation and articular damage in obesity-induced OA⁶⁹. However, another hypothesis article proposed that an increased serum endotoxin load was insufficient for OA onset and progression but exerted adjuvant effects on the process. A two-hit model of OA pathogenesis was formulated, wherein an initial proinflammatory innate immune response triggered by LPS was the first hit. Structural damage in a joint resulted from the second hit, which involved coexisting complementary mechanisms, such as inflammasome activation or assembly by the fragmented matrices of joint tissues that synergistically activated innate immunity⁷⁰. Reliably quantifying LPS has been a challenge for years because some LPS inhibitors combine with LPS and form complexes that mask detection in biospecimens^{71,72}. Nonetheless, an increased serum LPS concentration was previously detected in obese mice with OA¹⁴. A study also optimized the methodology of LPS quantification, thereby confirming that LPS placed a burden in both synovial fluid and serum samples of patients with OA⁷². Synovial LPS concentrations were correlated with macrophage activation, osteophyte severity, joint space narrowing severity and the total Western Ontario and McMaster Universities OA Index score in the knee⁷². LPS can also induce the secretion of matrix metalloproteases and components of the innate immune response in cultured chondrocytes⁷³ and accelerate the matrix degradation of articular cartilage explants⁷⁴. These studies strongly support the pathogenic role of LPS in the perpetuation of OA.

Although not specific to patients with OA, peptidoglycan–polysaccharide (PGN-PS) complexes were detected in cells in the synovial sublining layer of OA-afflicted joints⁷⁵. PGN-PS complexes were also isolated successfully from human feces and ileostomy fluids. The arthritogenic properties of PGN-PS complexes were further proven by the adjuvant-induced arthritis model⁷⁶. Bacterial PGNs were present in some immune cells, including macrophages and dendritic cells, in the synovia of patients with OA. The different cellular phenotypes coexpressed costimulatory molecules that were involved in antigen presentation and produced cytokines contributing to the inflammatory internal environments in joints⁷⁷. Several *in vitro* studies indicated the direct stimulatory effects of bacterial PGNs on synovial fibroblasts obtained from patients with OA and showed that bacterial PGNs induced the expression of matrix metalloproteases and proinflammatory cytokines⁷⁸. All of these results suggest that the intra-articular PGNs are involved in triggering or exacerbating OA progression.

The serum and synovial fluid lipidomic profiles are sensitive biomarkers of the radiographic stage of obesity-associated OA⁷⁹. Some metabolites related to FFA metabolism in the synovial fluid, such as myristic acid, oleic acid and lanosterol, increased with the structural deterioration of OA⁸⁰. Pathological concentrations of oleate could reduce the viability of articular chondrocytes through apoptosis⁸¹. The accumulation of ectopic lipids was observed in articular cartilage in joints afflicted with OA, and total fatty acids in general and arachidonic acid in particular closely correlated with

the erosion severity of the cartilage surface⁸². The lipotoxic effects induced by dyslipidemia in dominant cells within synovial tissues, including adipocytes and macrophages, worsened synovitis in patients with OA and MetS⁸³. The exertion of FFA-induced lipotoxicity in chondrocytes correlated with the amount of cellular FFAs, which were originally sequestered within lipid droplets⁸¹. Thus, FFAs may play an important role in OA pathophysiology.

Innate immunity

The complex and intricate interplay between the innate immune system and the intestinal microbiota plays a role in immunometabolic disorders^{8,56,58} and has been recognized as a causal factor in the biological basis of metabolic OA^{19,26,28,29}. In addition, overexpression of TLR2 and TLR4 has been observed in the lesions of OA articular cartilage⁸⁴. The treatment of chondrocytes obtained from OA patients with PGN and LPS activated TLR2-mediated inflammatory and catabolic signaling pathways⁸⁴. These results indicate the direct pathogenic role of non-immunocytes, which operate via the same mechanisms as canonical innate immune responses in OA. The activated synovial fibroblasts in patients with OA are also part of the innate immune system due to the activation of their outer membrane protein, TLR2, with PGN stimulation⁷⁸. The oral administration of resveratrol partly rescued the joint architecture disorganization in obesity-related OA by suppressing the TLR4-mediated inflammatory signaling pathways in the cartilage⁸⁵. However, MetS-like disorders elicited by TLR5 deletion did not aggravate load-induced cartilage damage in mice¹⁹. There are a number of possibilities to explain this. In particular, the extent of metabolic changes is insufficient to cause damage, the effects of metabolic factors are time-dependent and require time to manifest, and TLR5 itself may be involved in OA progression.

Increasing evidence has indicated that the NLRP3 inflammasome is a novel biomarker for OA diagnosis and monitoring⁸⁶. The high correlation between NLRP3 protein expression and pro-oxidant enzyme levels in the synovial membranes of patients with KOA suggested the possibility that the inflammasome was activated by oxidative stress in OA progression⁸⁷. Inhibition of the expression of the NLRP3 inflammasome or its components, such as pro-caspase-1, contributed to the amelioration of OA progression^{51,88}. In addition to the NLRP3 inflammasome, the NLRP1 inflammasome showed upregulated expression in the synovia of the joints of patients with OA^{51,87}. NLRP1 and NLRP3 inflammasomes also mediated inflammation and pyroptosis induced by LPS in fibroblast-like synoviocytes obtained from patients with OA⁵¹. These data strongly indicate the important role of inflammasomes in OA pathology and progression. Intriguingly, some exogenous and endogenous stimuli, including LPS and oxidative stress, which activate inflammasomes in OA progression, overlap with common pathogenic factors in obesity. However, whether these inflammasomes can be activated in the context of obesity contributing to OA development needs further investigation.

Therapeutics

Considering the continuously increasing global prevalence of obesity, novel preventive and therapeutic approaches for realizing symptomatic and structural relief of obesity-associated OA are greatly needed. Probiotics, prebiotics, nutritional supplements, dietary habits and exercise may influence the gut microbiome–OA interlinkage. Thus, these methods are considered as potential therapeutic approaches in the treatment of obesity-associated OA.

Probiotics and prebiotics, which are safe and effective dietary substances that can be used for elaborating microbial communities⁸⁹, are plausible therapeutic options for obesity-associated

OA. A randomized double-blind clinical trial proposed that commercial probiotic *L. casei* Shirota exhibited beneficial effects on the treatment outcomes of knee joints afflicted with OA⁹⁰. Supplementation with prebiotic fiber in the form of oligofructose could reverse the detrimental effects of fat-enriched diets on gut microbial communities by increasing the abundance of beneficial *Bifidobacteria* at the expense of numerous proinflammatory microbes, while inducing a considerable and consistent reduction in metabolic inflammation. It protected the joints of patients with OA against articular cartilage degeneration⁶⁴. Green-lipped mussel extract, a dietary supplement, is efficient in reducing OA symptoms, and its therapeutic efficacy is likely correlated with a decrease in *Clostridia* spp., which are potent modulators of colonic Th17 and CD4⁺ regulatory T cells, in the microbiota⁹¹.

The structures and functionalities of microbial communities are strongly related to diet^{36,37}. Some data have verified that dietary fats, especially saturated fatty acids, exert toxic effects on the etiologies of OA and the proinflammatory profiles of microbial communities might be a hidden causal factor underlying these effects^{64,92}. Conversely, high intake of dietary fiber and polyunsaturated fatty acids or a high ratio between polyunsaturated fatty acids and saturated fatty acids was associated with the alleviation of the symptoms and joint structures of obesity-associated OA and reduced the risk of the illness^{93–95}. Although the role of the gut microbiota still needs to be further explored, dietary habits provide a potential path toward the regulation of OA progression.

Although evidence from clinical trials on the effectiveness of exercise or physical activity for OA in individuals with obesity is limited, some intervention studies, which were not restricted to patients with obesity, had many recruited subjects that were overweight or obese, and the results confirmed the beneficial therapeutic effects of exercise on obesity-associated OA to some extent^{96–98}. An experimental study in obese mice showed that wheel running exerted protective rather than damaging effects on OA¹⁵. The integrated therapeutic approaches of exercise and dietary weight loss induce considerable improvements in knee pain and mobility performance in adults with obesity, compared with a single intervention^{99,100}. In addition, exercise considerably contributes to the modification of obese gut microbiota^{101–103}. Several specific microbiota taxa, such as *Lactobacillus* and *Bifidobacterium*, which can be positively regulated by exercise^{102,104}, also have potential value in OA treatment^{63,64}. Metabolic endotoxemia is also lower in active individuals than in those with sedentary lifestyles¹⁰⁵. Nonetheless, given the varying degrees of obesity in patients with OA and the pleiotropic effects of exercise involving mechanical and biological effects, the context-dependent roles of exercise should be further explored. Whether exercise presents favorable effects on OA by regulating the gut microbiota should be clarified.

Conclusion and future perspectives

The association between obesity and OA has been well established. Researchers have only recently begun to appreciate aberrant intestinal microbiota as a causal factor in the genesis and perpetuation of obesity-associated OA in animal models. However, the demonstration of the causality in humans will require further research. Microbiota-associated molecules affect the etiopathology of OA at both systemic and local levels by mechanisms involving innate immune activation. Given that many factors potentially contribute to immunometabolic disorders, at what point and to what extent microbiota-mediated factors play a major role in OA pathogenesis remains to be elucidated. In addition, further mechanistic studies are still required to provide a deep understanding of the interactions between the microbiota and the host either locally in the intestine or peripherally in the joint tissues. Probiotics and

prebiotics, dietary habits, and exercise may influence the gut microbiome–OA interlinkage and are considered as potential therapeutic approaches in the treatment of obesity-associated OA. Some oral nutritional supplements, such as green-lipped mussel extract, which have symptom-modulating effects in OA, although no consensus has been reached about the mechanism of action, may act by modulating the gut microbiome. The exploration of approaches for restoring a healthy microbiota, especially increasing the amount of specific commensal microbiota that antagonize proinflammatory microbes and maintain the intestinal mucosa barrier, is an important future direction for OA treatment.

Author contributions

All authors contributed substantially to drafting the article or revising it critically for important intellectual content. All authors approved the final version for submission.

Conflict of interest

None of the authors have competing interests to disclose. No benefits in any form have been received or will be received from a commercial party related directly or indirectly to the subject of this article.

Acknowledgments

This work was supported by National Natural Science Foundation (31701042) the Natural Science Foundation of Shandong Province (ZR2017LC012; ZR2017MC059).

References

1. Bijlsma JW, Berenbaum F, Lafeber FP. Osteoarthritis: an update with relevance for clinical practice. *Lancet* 2011;377: 2115–26.
2. Visser AW, Ioanfacsinay A, Mutsert RD, Widya RL, Loeff M, Roos AD, et al. Adiposity and hand osteoarthritis: The Netherlands epidemiology of obesity study. *Arthritis Res Ther* 2014;16:R19.
3. Oliveria SA, Felson DT, Cirillo PA, Reed JI, Walker AM. Body weight, body mass index, and incident symptomatic osteoarthritis of the hand, hip, and knee. *Epidemiology* 1999;10: 161–6.
4. Gill SR, Pop M, Deboy RT, Eckburg PB, Turnbaugh PJ, Samuel BS, et al. Metagenomic analysis of the human distal gut microbiome. *Science* 2006;312:1355–9.
5. Kundu P, Blacher E, Elinav E, Pettersson S. Our Gut Microbiome: the evolving inner self. *Cell* 2017;171:1481–93.
6. Benson AK, Kelly SA, Legge R, Ma F, Low SJ, Kim J, et al. Individuality in gut microbiota composition is a complex polygenic trait shaped by multiple environmental and host genetic factors. *Proc Natl Acad Sci U S A* 2010;107:18933–8.
7. Nagpal R, Kumar M, Yadav AK, Hemalatha R, Yadav H, Marotta F, et al. Gut microbiota in health and disease: an overview focused on metabolic inflammation. *Benef Microbes* 2016;7:181–94.
8. McPhee JB, Schertzer JD. Immunometabolism of obesity and diabetes: microbiota link compartmentalized immunity in the gut to metabolic tissue inflammation. *Clin Sci* 2015;129: 1083–96.
9. Hotamisligil GS. Inflammation, metaflammation and immunometabolic disorders. *Nature* 2017;542:177–85.
10. Hotamisligil GS, Erbay E. Nutrient sensing and inflammation in metabolic diseases. *Nat Rev Immunol* 2008;8:923–34.
11. Horta-Baas G, Romero-Figueroa MDS, Montiel-Jarquín AJ, Pizano-Zárate ML, García-Mena J, Ramírez-Durán N. Intestinal dysbiosis and rheumatoid arthritis: a link between gut microbiota and the pathogenesis of rheumatoid arthritis. *J Immunol Res* 2017;2017:1–13.
12. Radin EL, Paul IL, Rose RM. Role of mechanical factors in pathogenesis of primary osteoarthritis. *Lancet* 1972;299: 519–22.
13. Collins KH, Reimer RA, Seerattan RA, Leonard TR, Herzog W. Using diet-induced obesity to understand a metabolic subtype of osteoarthritis in rats. *Osteoarthritis Cartilage* 2015;23:957–65.
14. Collins KH, Paul HA, Reimer RA, Seerattan RA, Hart DA, Herzog W. Relationship between inflammation, the gut microbiota, and metabolic osteoarthritis development: studies in a rat model. *Osteoarthritis Cartilage* 2015;23: 1989–98.
15. Griffin TM, Huebner JL, Kraus VB, Yan Z, Guilak F. Induction of osteoarthritis and metabolic inflammation by a very high-fat diet in mice: effects of short-term exercise. *Arthritis Rheum* 2014;64:443–53.
16. Mooney RA, Sampson ER, Lerea J, Rosier RN, Zuscik MJ. High-fat diet accelerates progression of osteoarthritis after meniscal/ligamentous injury. *Arthritis Res Ther* 2011;13: R198.
17. Iwata M, Ochi H, Hara Y, Tagawa M, Koga D, Okawa A, et al. Initial responses of articular tissues in a murine high-fat diet-induced osteoarthritis model: pivotal role of the IPFP as a cytokine fountain. *PLoS One* 2013;8, e60706.
18. Asou Y, Iwata M, Ochi H, Ailixiding M, Aibibula Z, Piao J, et al. Pleiotropic functions of high fat diet in the etiology of osteoarthritis. *PLoS One* 2016;11, e0162794.
19. Guss JD, Ziemian SN, Luna M, Sandoval TN, Holyoak DT, Guisado GG, et al. The effects of metabolic syndrome, obesity, and the gut microbiome on load-induced osteoarthritis. *Osteoarthritis Cartilage* 2019;27:129–39.
20. Xie D, Wei J, Zeng C, Yang T, Li H, Wang Y, et al. Association between metabolic syndrome and knee osteoarthritis: a cross-sectional study. *BMC Musculoskelet Disord* 2017;18: 533.
21. Yoshimura N, Muraki S, Oka H, Tanaka S, Kawaguchi H, Nakamura K, et al. Accumulation of metabolic risk factors such as overweight, hypertension, dyslipidaemia, and impaired glucose tolerance raises the risk of occurrence and progression of knee osteoarthritis: a 3-year follow-up of the ROAD study. *Osteoarthritis Cartilage* 2012;20:1217–26.
22. Puenpatom RA, Victor TW. Increased prevalence of metabolic syndrome in individuals with osteoarthritis: an analysis of NHANES III data. *Postgrad Med* 2009;121:9–20.
23. Niu J, Clancy M, Aliabadi P, Vasan R, Feoson DT. The metabolic syndrome, its components and knee osteoarthritis (OA): the framingham OA study. *Arthritis Rheum* 2017;69:1194–203.
24. Shin D. Association between metabolic syndrome, radiographic knee osteoarthritis, and intensity of knee pain: results of a national survey. *J Clin Endocrinol Metab* 2014;99: 3177–83.
25. Monira Hussain SM, Wang Y, Cicuttini FM, Simpson JA, Giles GG, Graves S, et al. Incidence of total knee and hip replacement for osteoarthritis in relation to the metabolic syndrome and its components: a prospective cohort study. *Semin Arthritis Rheum* 2014;7:429–36.
26. Zhuo Q, Yang W, Chen J, Wang Y. Metabolic syndrome meets osteoarthritis. *Nat Rev Rheumatol* 2012;8:729–37.
27. Robinson WH, Lepus CM, Wang Q, Raghu H, Mao R, Lindstrom TM, et al. Low-grade inflammation as a key mediator of the pathogenesis of osteoarthritis. *Nat Rev Rheumatol* 2016;12:580–92.

28. Gierman LM, van der Ham F, Koudijs A, Wielinga PY, Kleemann R, Kooistra T, *et al.* Metabolic stress-induced inflammation plays a major role in the development of osteoarthritis in mice. *Arthritis Rheum* 2014;64:1172–81.
29. Sun AR, Panchal SK, Friis T, Sekar S, Crawford R, Brown L, *et al.* Obesity-associated metabolic syndrome spontaneously induces infiltration of pro-inflammatory macrophage in synovium and promotes osteoarthritis. *PLoS One* 2017;12, e0183693.
30. Conde J, Scotece M, Gómez R, Lopez V, Gómezreino JJ, Gualillo O. Adipokines and osteoarthritis: novel molecules involved in the pathogenesis and progression of disease. *Arthritis* 2011;2011:203901.
31. Griffin TM, Fermor B, Huebner JL, Kraus VB, Rodriguiz RM, Wetsel WC, *et al.* Diet-induced obesity differentially regulates behavioral, biomechanical, and molecular risk factors for osteoarthritis in mice. *Arthritis Res Ther* 2010;12: R130.
32. Ley RE, Bäckhed F, Turnbaugh P, Lozupone CA, Knight RD, Gordon JI. Obesity alters gut microbial ecology. *Proc Natl Acad Sci U S A* 2005;102:11070–5.
33. Ley R, Turnbaugh P, Klein S, Gordon J. Microbial ecology: human gut microbes associated with obesity. *Nature* 2006;444:1022–3.
34. Bäckhed F, Ding H, Wang T, Hooper LV, Gou YK, Nagy A, *et al.* The gut microbiota as an environmental factor that regulates fat storage. *Proc Natl Acad Sci USA* 2004;101:15718–23.
35. Fei N, Zhao L, Fei N, Zhao LP. An opportunistic pathogen isolated from the gut of an obese human causes obesity in germfree mice. *ISME J* 2012;7:880–4.
36. Velasquez MT. Altered gut microbiota: a link between diet and the metabolic syndrome. *Metab Syndr Disord* 2018;16:321–8.
37. Wang J, Tang H, Zhang C, Zhao Y, Derrien M, Rocher E, *et al.* Modulation of gut microbiota during probiotic-mediated attenuation of metabolic syndrome in high fat diet-fed mice. *ISME J* 2015;9:1–15.
38. Schneeberger M, Everard A, Gómez-Valadés AG, Matamoros S, Ramírez S, Delzenne NM, *et al.* Akkermansia muciniphila inversely correlates with the onset of inflammation, altered adipose tissue metabolism and metabolic disorders during obesity in mice. *Sci Rep* 2015;5:16643.
39. Xiao S, Fei N, Shen J, Wang L, Zhang B, *et al.* A gut microbiota-targeted dietary intervention for amelioration of chronic inflammation underlying metabolic syndrome. *FEMS Microbiol Ecol* 2014;87:357–67.
40. Cani PD, Amar J, Iglesias MA, Poggi M, Knauf C, Bastelica D, *et al.* Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes* 2007;56:1761–72.
41. Chi W, Dao D, Lau TC, Henriksbo BD, Cavallari JF, Foley KP, *et al.* Bacterial peptidoglycan stimulates adipocyte lipolysis via NOD1. *PLoS One* 2014;9, e97675.
42. Boden G. Obesity and free fatty acids (FFA). *Endocrinol Metab Clin N Am* 2008;37:635–43.
43. Robert C, Reigstad CS, Kling BH, Christoph R, Maria K, Gunnel ÖL, *et al.* Gut-derived lipopolysaccharide augments adipose macrophage accumulation but is not essential for impaired glucose or insulin tolerance in mice. *Gut* 2012;61: 1701–7.
44. Hersoug LG, Møller P, Loft S. Gut microbiota-derived lipopolysaccharide uptake and trafficking to adipose tissue: implications for inflammation and obesity. *Obes Rev* 2016;17: 297–312.
45. Wolf AJ, Underhill DM. Peptidoglycan recognition by the innate immune system. *Nat Rev Immunol* 2018;18:243–54.
46. Schertzer JD, Tamrakar AK, Magalhães JG, Pereira S, Bilan PJ, Fullerton MD, *et al.* NOD1 activators link innate immunity to insulin resistance. *Diabetes* 2011;60:2206–15.
47. Jin Y, Wu Y, Zeng Z, Jin C, Wu S, Wang Y, *et al.* From the cover: exposure to oral antibiotics induces gut microbiota dysbiosis associated with lipid metabolism dysfunction and low-grade inflammation in mice. *Toxicol Sci* 2016;154:140–52.
48. Rodríguezcarrio J, Salazar N, Margolles A, González S, Gueimonde M, De RG, *et al.* Free fatty acids profiles are related to gut microbiota signatures and short-chain fatty acids. *Front Immunol* 2017;8:823–36.
49. Shi H, Kokoeva MV, Inouye K, Tzamelis I, Yin H, Flier JS. TLR4 links innate immunity and fatty acid-induced insulin resistance. *J Clin Invest* 2006;116:3015–25.
50. Velloso LA, Folli F, Saad MJ. TLR4 at the crossroads of nutrients, gut microbiota, and metabolic inflammation. *Endocr Rev* 2015;36:245–71.
51. Zhao LR, Xing RL, Wang PM, Zhang NS, Yin SJ, Li XC, *et al.* NLRP1 and NLRP3 inflammasomes mediate LPS/ATP-induced pyroptosis in knee osteoarthritis. *Mol Med Rep* 2018;17: 5463–9.
52. Horng T, Hotamisligil GS. Linking the inflammasome to obesity-related disease. *Nat Med* 2011;17:164–5.
53. Henao-Mejia J, Elinav E, Jin C, Hao L, Mehal WZ, Strowig T, *et al.* Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity. *Nature* 2012;482:179–85.
54. Lamkanfi M, Kanneganti TD. The inflammasome: a remote control for metabolic syndrome. *Cell Res* 2012;22:1095–8.
55. Vijaykumar M, Aitken JD, Carvalho FA, Cullender TC, Mwangi S, Srinivasan S, *et al.* Metabolic syndrome and altered gut microbiota in mice lacking Toll-like receptor 5. *Science* 2010;328:228–31.
56. Chassaing B, Gewirtz AT. Gut microbiota, low-grade inflammation, and metabolic syndrome. *Toxicol Pathol* 2014;42: 49–53.
57. Cheng J, Xue F, Zhang M, Cheng C, Qiao L, Ma J, *et al.* TRIM31 deficiency is associated with impaired glucose metabolism and disrupted gut microbiota in mice. *Front Physiol* 2018;9: 24–34.
58. Miura K, Ishioka M, Iijima K. The roles of the gut Microbiota and Toll-like receptors in obesity and nonalcoholic fatty liver disease. *Int J Obes Relat Metab Disord* 2017;26:86–96.
59. Jukes TH, Williams WL. Nutritional effects of antibiotics. *Pharmacol Rev* 1953;5:381–420.
60. Kang Y, Cai Y, Zhang X, Kong X, Su J. Altered gut microbiota in RA: implications for treatment. *Z Rheumatol* 2017;76:451–7.
61. Vasiladis HS, Tsikopoulos K. Glucosamine and chondroitin for the treatment of osteoarthritis. *World J Orthoped* 2017;8: 1–11.
62. Wang Q, Huang SQ, Li CQ, Xu Q, Zeng QP. Akkermansia muciniphilamay determine chondroitin sulfate ameliorating or aggravating osteoarthritis. *Front Microbiol* 2017;8:1955.
63. So JS, Song MK, Kwon HK, Lee CG, Chae CS, Sahoo A, *et al.* Lactobacillus casei enhances type II collagen/glucosamine-mediated suppression of inflammatory responses in experimental osteoarthritis. *Life Sci* 2011;88:358–66.
64. Schott EM, Farnsworth CW, Grier A, Lillis JA, Soniwal S, Dadourian GH, *et al.* Targeting the gut microbiome to treat the osteoarthritis of obesity. *JCI insight* 2018;3(8), e95997.
65. Qin Y, Roberts JD, Grimm SA, Lih FB, Deterding LJ, Li R, *et al.* An obesity-associated gut microbiome reprograms the intestinal epigenome and leads to altered colonic gene expression. *Genome Biol* 2018;19:7–21.
66. Kempell KE, Cox CJ, Hurle M, Wong A, Wilkie S, Zanders ED, *et al.* Reverse transcriptase-PCR analysis of bacterial rRNA for

- detection and characterization of bacterial species in arthritis synovial tissue. *Infect Immun* 2000;68:6012–26.
67. Katz KD, Hollander D. Intestinal mucosal permeability and rheumatological diseases. *Bailliere's Clin Rheumatol* 1989;3: 271–84.
 68. Berthelot JM, Claudepierre P. Trafficking of antigens from gut to sacroiliac joints and spine in reactive arthritis and spondyloarthropathies: mainly through lymphatics? *Jt Bone Spine* 2016;83:485–90.
 69. Metcalfe D, Harte AL, Aletrari MO, Al Daghri NM, Al Disi D, Tripathi G, et al. Does endotoxaemia contribute to osteoarthritis in obese patients? *Clin Sci* 2012;123:627–34.
 70. Huang Z, Kraus VB. Does lipopolysaccharide-mediated inflammation have a role in OA? *Nat Rev Rheumatol* 2016;12:123–9.
 71. Hurley JC, Tosolini FA, Louis WJ. Quantitative limulus lysate assay for endotoxin and the effect of plasma. *J Clin Pathol* 1991;44:849–54.
 72. Huang ZY, Stabler T, Pei FX, Kraus VB. Both systemic and local lipopolysaccharide (LPS) burden are associated with knee OA severity and inflammation. *Osteoarthritis Cartilage* 2016;24: 1769–75.
 73. Haglund L, Bernier SM, Onnerfjord P, Recklies AD. Proteomic analysis of the LPS-induced stress response in rat chondrocytes reveals induction of innate immune response components in articular cartilage. *Matrix Biol* 2008;27:107–18.
 74. Macdonald MH, Stover SM, Willits NH, Benton HP. Effect of bacterial lipopolysaccharides on sulfated glycosaminoglycan metabolism and prostaglandin E2 synthesis in equine cartilage explant cultures. *Am J Vet Res* 1994;55:1127–38.
 75. van der Heijden IM, Wilbrink B, Tchetverikov I, Schrijver IA, Schouls LM, Hazenberg MP, et al. Presence of bacterial DNA and bacterial peptidoglycans in joints of patients with rheumatoid arthritis and other arthritides. *Arthritis Rheum* 2000;43:593–8.
 76. Kool J, Ruseler-van Embden JG, van Lieshout LM, de Visser H, Gerrits-Boeye MY, van den Berg WB, et al. Induction of arthritis in rats by soluble peptidoglycan-polysaccharide complexes produced by human intestinal flora. *Arthritis Rheum* 1991;34:1611–6.
 77. Schrijver IA, Melief MJ, Tak PP, Hazenberg MP, Laman JD. Antigen-presenting cells containing bacterial peptidoglycan in synovial tissues of rheumatoid arthritis patients coexpress costimulatory molecules and cytokines. *Arthritis Rheum* 2000;43:2160–8.
 78. Kyburz D, Rethage J, Seibl R, Lauener R, Gay RE, Carson DA, et al. Bacterial peptidoglycans but not CpG oligodeoxynucleotides activate synovial fibroblasts by Toll-like receptor signaling. *Arthritis Rheum* 2003;48:642–50.
 79. Wu CL, Kimmerling KA, Little D, Guilak F. Serum and synovial fluid lipidomic profiles predict obesity-associated osteoarthritis, synovitis, and wound repair. *Sci Rep* 2017;7:44315.
 80. Kim S, Hwang J, Kim J, Ahn JK, Cha HS, Kim KH. Metabolite profiles of synovial fluid change with the radiographic severity of knee osteoarthritis. *Jt Bone Spine* 2017;84: 605–10.
 81. Lee SW, Rho JH, Sang YL, Chung WT, Oh YJ, Kim JH, et al. Dietary fat-associated osteoarthritic chondrocytes gain resistance to lipotoxicity through PKCK2/STAMP2/FSP27. *Bone Res* 2018;6:20.
 82. Lippiello L, Walsh T, Fienhold M. The association of lipid abnormalities with tissue pathology in human osteoarthritic articular cartilage. *Metabolism* 1991;40:571–6.
 83. Larrañaga-Vera A, Lamuedra A, Pérez-Baos S, Prieto-Potin I, Peña L, Herrero-Beaumont G, et al. Increased synovial lipodystrophy induced by high fat diet aggravates synovitis in experimental osteoarthritis. *Arthritis Res Ther* 2017;19:R264.
 84. Kim HA, Cho ML, Choi HY, Yoon CS, Jhun JY, Oh HJ, et al. The catabolic pathway mediated by Toll-like receptors in human osteoarthritic chondrocytes. *Arthritis Rheum* 2010;54: 2152–63.
 85. Jiang M, Li X, Yu X, Liu X, Xu X, He J, et al. Oral administration of resveratrol alleviates osteoarthritis pathology in C57BL/6J mice model induced by a high-fat diet. *Mediat Inflamm* 2017;2017:7659023.
 86. Mcallister MJ, Chemaly M, Eakin AJ, Gibson DS, McGilligan VE. NLRP3 as a potentially novel biomarker for the management of osteoarthritis. *Osteoarthritis Cartilage* 2018;26:612–9.
 87. Clavijo-Cornejo D, Martínez-Flores K, Silva-Luna K, Gabriela, Martínez-Nava A, Fernández-Torres J, et al. The over-expression of NALP3 inflammasome in knee osteoarthritis is associated to synovial membrane prolidase and NADPH oxidase 2. *Oxid Med Cell Longev* 2016;2016:1–7.
 88. Sun Y, Liu W, Zhang H, Li H, Liu J, Zhang F, et al. Curcumin prevents osteoarthritis by inhibiting the activation of inflammasome NLRP3. *J Interferon Cytokine Res* 2017;37: 449–55.
 89. Dahiya DK, Renuka, Puniya M, Shandilya UK, Dhewa T, Kumar N, et al. Gut microbiota modulation and its relationship with obesity using prebiotic fibers and probiotics: a review. *Front Microbiol* 2017;8:563.
 90. Lei M, Guo C, Wang D, Zhang C, Hua L. The effect of probiotic *Lactobacillus casei* Shirota on knee osteoarthritis: a randomised double-blind, placebo-controlled clinical trial. *Benef Microbes* 2017;8:697–703.
 91. Coulson S, Butt H, Vecchio P, Gramotnev H, Vitetta L. Green-lipped mussel extract (*Perna canaliculus*) and glucosamine sulphate in patients with knee osteoarthritis: therapeutic efficacy and effects on gastrointestinal microbiota profiles. *Inflammopharmacology* 2013;21(1):79–90.
 92. Sekar S, Shafie SR, Prasadani I, Crawford R, Panchal SK, Brown L, et al. Saturated fatty acids induce development of both metabolic syndrome and osteoarthritis in rats. *Sci Rep* 2017;7:46457.
 93. Dai Z, Niu J, Zhang Y, Jacques P, Felson DT. Dietary intake of fibre and risk of knee osteoarthritis in two US prospective cohorts. *Ann Rheum Dis* 2017;76:1411–9.
 94. Lu B, Driban JB, Xu C, Lapane KL, McAlindon TE, Eaton CB. Dietary fat intake and radiographic progression of knee osteoarthritis: data from the osteoarthritis initiative. *Arthritis Care Res* 2017;69:368–75.
 95. Wu CL, Jain D, McNeill JN, Little D, Anderson JA, Huebner JL, et al. Dietary fatty acid content regulates wound repair and the pathogenesis of osteoarthritis following joint injury. *Ann Rheum Dis* 2015;74:2076–83.
 96. Ettinger WH, Burns R, Messier SP, Applegate W, Rejeski WJ, Morgan T, et al. A randomized trial comparing aerobic exercise and resistance exercise with a health education program in older adults with knee osteoarthritis. The Fitness Arthritis and Seniors Trial (FAST). *J Am Med Assoc* 1997;277:25–31.
 97. Waller B, Munukka M, Rantalainen T, Lamentusta E, Nieminen MT, Kiviranta I, et al. Effects of high intensity resistance aquatic training on body composition and walking speed in women with mild knee osteoarthritis: a 4-month RCT with 12-month follow-up. *Osteoarthritis Cartilage* 2017;25:1238–46.
 98. Farr JN, Going SB, Mcknight PE, Kastle S, Cussler EC, Cornett M. Progressive resistance training improves overall physical activity levels in patients with early osteoarthritis of the knee: a randomized controlled trial. *Phys Ther* 2010;90:356–66.

99. Messier SP, Loeser RF, Miller GD, Morgan TM, Rejeski WJ, Sevick MA, *et al.* Exercise and dietary weight loss in overweight and obese older adults with knee osteoarthritis: the arthritis, diet, and activity promotion trial. *Arthritis Rheum* 2004;50:1501–10.
100. Messier SP, Mihalko SL, Legault C, Miller GD, Nicklas BJ, Devita P, *et al.* Effects of intensive diet and exercise on knee joint loads, inflammation, and clinical outcomes among overweight and obese adults with knee osteoarthritis: the IDEA randomized clinical trial. *J Am Med Assoc* 2013;310:1263–73.
101. Kang SS, Jeraldo PR, Kurti A, Miller MEB, Cook MD, Whitlock K, *et al.* Diet and exercise orthogonally alter the gut microbiome and reveal independent associations with anxiety and cognition. *Mol Neurodegener* 2014;9:36–48.
102. Petrizz BA, Castro AP, Almeida JA, Gomes CP, Fernandes GR, Kruger RH, *et al.* Exercise induction of gut microbiota modifications in obese, non-obese and hypertensive rats. *BMC Genomics* 2014;15:511–24.
103. Clarke SF, Murphy EF, O'Sullivan O, Lucey AJ, Humphreys M, Hogan A, *et al.* Exercise and associated dietary extremes impact on gut microbial diversity. *Gut* 2014;63:1913–20.
104. Yin HL, Niu AJ. Effect of "eight-trigram boxing" exercise on blood oxidative status and intestine bifidobacterium and lactobacillus count in practitioners. *Afr J Microbiol Res* 2010;4:2434–8.
105. Lira FS, Rosa JC, Pimentel GD, Souza HA, Caperuto EC, Carnevali LC, *et al.* Endotoxin levels correlate positively with a sedentary lifestyle and negatively with highly trained subjects. *Lipids Health Dis* 2010;9:82–7.