

# Genetics of Resilience: Gene-by-Environment Interaction Studies as a Tool to Dissect Mechanisms of Resilience

Immanuel G. Elbau, Cristiana Cruceanu, and Elisabeth B. Binder

## ABSTRACT

The identification and understanding of resilience mechanisms holds potential for the development of mechanistically informed prevention and interventions in psychiatry. However, investigating resilience mechanisms is conceptually and methodologically challenging because resilience does not merely constitute the absence of disease-specific risk but rather reflects active processes that aid in the maintenance of physiological and psychological homeostasis across a broad range of environmental circumstances. In this conceptual review, we argue that the principle used in gene-by-environment interaction studies may help to unravel resilience mechanisms on different investigation levels. We present how this could be achieved by top-down designs that start with gene-by-environment interaction effects on disease phenotypes as well as by bottom-up approaches that start at the molecular level. We also discuss how recent technological advances may improve both top-down and bottom-up strategies.

**Keywords:** Epigenetics, Gene-by-environment interactions, Genetics, Psychiatry, Resilience, Stress

<https://doi.org/10.1016/j.biopsych.2019.04.025>

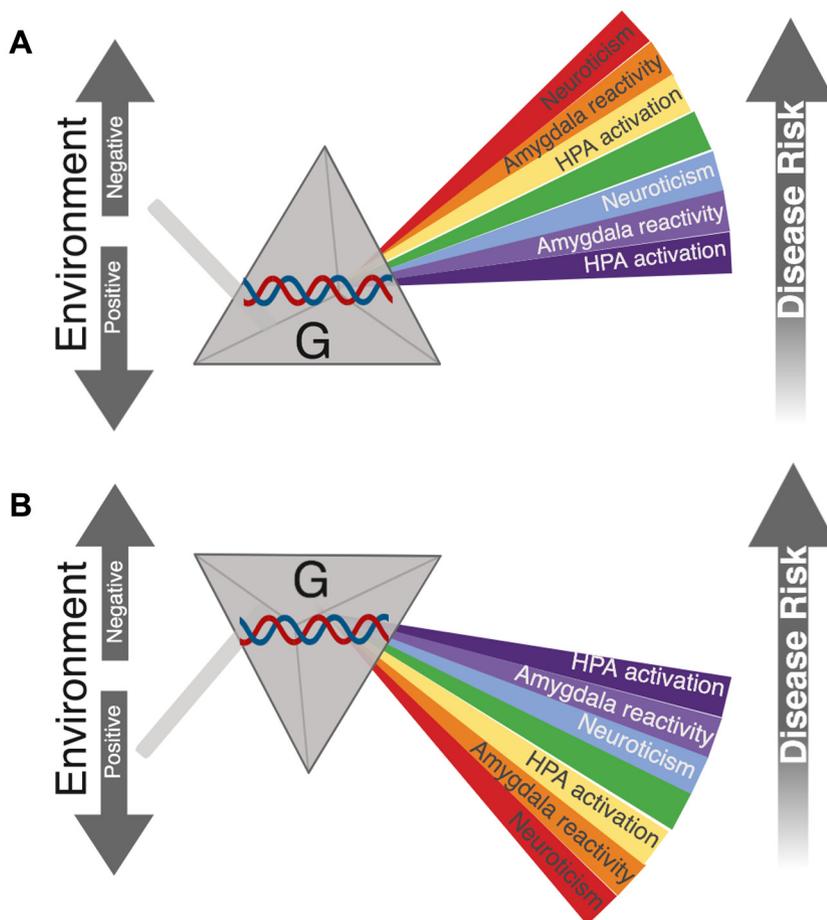
In psychiatric research, the phenomenon of resilience has gained increasing attention over the past decade because it holds potential for mechanistically informed prevention and intervention. Resilience is defined as the ability to maintain normal psychological and physiological functioning despite exposure to stress and adversity. Active processes that contribute to maintaining homeostasis within a broad spectrum of environmental disturbances likely underlie this phenotype (1). These processes may have a genetic basis and contribute to the resilient phenotype through a cascade of molecular and cellular events that subsequently regulate neural, physiological, and endocrine systems; these ultimately converge to the maintenance of what we consider normal (or adaptive) behavior. The identification of resilience mechanisms and their genetic determinants, however, is challenging both practically and conceptually. Environmental risk factors play a major role in shaping long-term outcomes for mental health and behavior (2). Hence, exploring human genetic variations that reduce disease risk in light of adversity may serve as a tool to identify mechanisms and pathways that promote resilience. This would allow the mapping of moderating factors' effects across levels of functioning, that is, from molecules to systems to behaviors. This approach could identify how mechanisms of resilience manifest across these levels and provide insight into novel targets for prevention and intervention at all levels.

The aim of this review was to illustrate how human studies that investigate gene-by-environment interactions (G×Es) may help to unravel resilience mechanisms. We illustrate how both classical top-down designs that start with G×E effects on disease phenotypes and bottom-up approaches that start at

the level of molecular readouts could contribute to our understanding of resilience mechanisms.

## G×E MAPPING TO IDENTIFY COMMON MECHANISMS: DIAGNOSES TO INTERMEDIATE PHENOTYPES

Different conceptual frameworks of how genomic determinants moderate environmental risk have evolved over time. Initially, G×E studies were framed within the diathesis–stress model, which posits that cumulative environmental adversity eventually will lead to dysfunction and that genetic (and other biological) variables determine the individual threshold of adversity necessary for dysfunction to occur. Resilience genotypes within this conceptualization heighten this threshold by affecting adversity-related biological changes. This can happen by reducing long-term negative consequences or by enhancing compensating mechanisms. The diathesis–stress model, however, fails to embed the phenomenon of genetic resilience within a more general theory of environmental sensitivity that is independent of the environment's valence (e.g., stressful vs. nurturing). Relatedly, it fails to explain why susceptibility genotypes have not been selected against during evolution. These shortcomings are addressed in newer, evolutionarily grounded concepts such as the match–mismatch and differential susceptibility frameworks that embed susceptibility and resilience genotypes within a unifying theory of general programming susceptibility to environmental influences, be they adverse or positive (3). Differential sensitivity to environmental programming within these



**Figure 1.** Schematic representation of the concept of resilience and its relationship to gene-by-environment interaction as proposed in this review. **(A)** Situation of high environmental adversity. High adversity is associated with higher disease risk across diagnostic categories. This, however, is not the result of environmental shaping of disease-specific risk factors but rather is mediated through environmental programming of intermediate phenotypes such as hypothalamic-pituitary-adrenal (HPA)-axis functioning, amygdala reactivity, and behavioral dispositions (color beams). The extent of environmental impact on these domains is mediated through genetic determinants such as common gene variants in various genes (G). Here, individuals with less susceptible genotypes (cold colors) show less environmental moderation (or programming) of these domains and hence less increase in disease risk. As such, in this environmental context, they represent a resilient phenotype. The opposite situation is depicted in warm colors. **(B)** Situation of a highly nourishing environment. Here, the less susceptible genotype will lead to more disease risk because the positive environmental influences less shape (or program) the disease risk-conferring intermediate phenotypes toward the low-risk direction. This ambiguous nature of risk and resilience that rests on the notion of more or less environmental programmability is formulated in the framework of differential susceptibility.

conceptualizations can be adaptive or maladaptive depending on the individual's environmental trajectory. Risk versus resilience genotypes may rather reflect high versus low environmental plasticity genotypes (4). This latter, more ecologically valid concept for G×E studies mandates study designs that assess the environment not unidirectionally but rather across the entire spectrum from positive to adverse. Currently, only very few studies have incorporated such multidimensional assessments of the environment, which requires longitudinal designs with complex, multi-informant measures of the environment. Lack of completeness in measurements of the environment across the entire range of development may also contribute to inconsistent findings in G×E studies (5,6). The majority of published studies focus on adversity, often measured at only one or a few developmental time points, which limits our ability to identify whether polymorphisms indeed moderate responses to the complex environment. Despite these limitations, current G×E studies allow us to explore their potential for identifying resilience mechanisms (see Figure 1).

### Candidate Gene Strategies

We first explore how candidate G×E studies may contribute to our understanding of resilience mechanisms. For this, we focus on common variants in three candidate genes that may

moderate the effect of adverse life events on psychiatric disorders and related intermediate phenotypes: the corticotropin-releasing hormone receptor 1 gene (*CRHR1*), the FK506 binding protein 5 gene (*FKBP5*), and the serotonin transporter gene (*SLC6A4*). These three genes were selected based on the wealth of studies across a range of different phenotype levels and the presence of functional studies as well as for the purpose of illustrating the concept of convergence across levels of investigation. They are not meant to be exhaustive of the literature on G×E in the context of risk and resilience, which has recently been reviewed (3,4). *CRHR1* is a G protein-coupled receptor that binds corticotropin-releasing hormone, a peptide that plays a key role in activating signal transduction pathways involved in the stress response (7,8). *FKBP5* encodes a co-chaperone of heat shock protein 90 that moderates stress hormone/glucocorticoid receptor (GR) function (9). *SLC6A4* encodes the serotonin transporter, the target of selective serotonin reuptake inhibitors. For *SLC6A4*, the most commonly studied polymorphism is a repeat polymorphism (the short allele and long allele) in the promoter region (5-HTTLPR polymorphism) [for details, see (10)].

The literature examining G×E effects for *CRHR1*, *FKBP5*, and 5-HTTLPR has recently been comprehensively reviewed in a number of articles (3,4,8,9). The aim of this conceptual review was to illustrate convergence of findings across these

exemplary genes and to illustrate how G×E studies might inform our understanding of resilience mechanisms. A first observation in this regard is that the common variants in all three genes have risk-modifying properties that are not confined to a single psychiatric diagnosis but rather go across diagnostic categories (4). This is in line with the conceptualization of resilience as an active process that aids the maintenance of homeostasis within a broad range of environmental circumstances (1) as opposed to resilience constituting merely the lack of disease-specific risk factors. In fact, the same polymorphisms in these three genes have been shown to moderate the effects of adversity on the development of major depressive disorder (MDD) (9,11–17), posttraumatic stress disorder (18–27), and alcohol use disorder (28–32) as well as other psychiatric disorders, including first-onset psychosis (33,34). This suggests that genetic variation in these genes may influence disease risk by moderating proximal shared resilience mechanisms. This raises the question as to which common physiological processes are modified by these gene variants.

Part of the answer might come from studies that investigated the same G×E effects on measures of intermediate physiological, cognitive, or neural processes that likely underlie risk for psychiatric pathology across specific phenotypic expressions (35). The next section summarizes evidence of convergence of risk-moderating gene variants onto the same intermediate phenotypes, focusing on the stress hormone response as one example, which by all means is not meant to be an exhaustive representation of intermediate phenotypes relevant to resilience.

### Convergence Onto Intermediate Phenotypes

Regulation of the stress hormone system within the hypothalamic–pituitary–adrenal (HPA) axis is central in adversity-related disease risk, with both blunted and exaggerated responses reported following exposure to stressful life events and in disease states (36,37). The set point of this system has been studied with psychological and pharmacological challenge tests (38,39) as well as physiological endocrine measurements (40). All three genes' polymorphisms have been shown to alter these measures, with the so-called risk alleles often associated with an exaggerated or prolonged response of the system (41–59). For *FKBP5*, these findings extend to prolonged endocrine responses to psychosocial stressors (41,47,50), altered responses to pharmacological challenges (53), and physiological HPA axis measures (48,49). Similarly, *CRHR1* risk haplotypes have been shown to modulate responsiveness to psychological challenge tests (51,56), pharmacological challenges (46,58), and diurnal cortisol regulation (57). The 5-HTTLPR polymorphism has been most extensively studied in this regard, with meta-analytical evidence for a heightened endocrine response to psychosocial stress among carriers of the risk/short allele (54), with evidence for a stress response modulatory role across species already during infancy (42–44,52,55). These results support the notion that disease risk-modifying properties of these gene variants may in part be mediated through regulatory effects on HPA axis set points. The polymorphisms may affect different parts of the stress response, from activation to negative feedback, and depending on context, risk alleles may be associated with different HPA axis outcomes and yet overall promote

homeostasis. These findings suggest that different genetic variants might contribute to resilience by moderating the effects of the environment on the same physiological mechanism.

It is worth noting that convergence is evident not just for HPA axis functioning but also for other intermediate phenotypes that are associated with risk and resilience across mental disorders such as neurocircuit measures of threat responsiveness (60–68) and neuroticism as a personality trait (69–74) (Figure 1).

While the presented candidate G×E studies may provide a roadmap for investigating mechanisms of resilience, it is clear that they are likely underpowered, with inconsistent results reported [e.g., in the interaction between the 5-HTTLPR and early life adversity on depression risk (15,71,75–77)] and no support in large cohort studies (6). Similarly, there are also negative findings regarding the moderating effects of the *FKBP5* haplotype on anxiety and depression (17,78,79) and inconsistent findings for *CRHR1* (14,80). Inconsistent findings also exist regarding the discussed findings in intermediate phenotypes, with only few findings having been confirmed in large meta-analyses (54,66,67,71). There are many potential sources for such inconsistencies, the most important being differences in the assessment of environmental risk, with timing, type, and duration likely affecting these interactions and differences between self-reported and objective measures of adversity (5,81–83). Additive and interactive effects of different environments have been reported but are often not considered, and genetic effects have been shown to be diminished (27), but also unmasked (21), with increasing levels of adversity. Other potential sources are heterogeneity of phenotype definition and assessment (84,85) but also insufficiently powered studies. In addition, individual single nucleotide polymorphisms (SNPs) contribute only a very small proportion of the overall variance.

### Genome-wide Strategies

Genome-wide G×E studies (GEWISs) represent a powerful unbiased approach for the identification of gene variants that modify environmental risk and hence can inform about the genetic basis of resilience mechanism. This has proven to be successful in other areas of medicine (86–88). To date, only a few GEWISs have been conducted in psychiatry, the majority of which examined the effect of stressful life events on depression phenotypes (89–93). The largest GEWIS in psychiatry to date investigated the interaction of adult stressful life events with common gene variants on depression symptoms in a UK Biobank sample (~100,000 subjects) and a Generation Scotland sample (~5000 subjects). The study yielded inconsistent evidence for the presence of gene × adult stressful life events on depressive symptoms (89). GEWISs have two main limitations. First, there is limited power to detect the small effect sizes of each individual G×E inherent to the polygenic nature of complex disorders. Second, even more so than in candidate G×E studies, large sample sizes often come at the cost of detailed and in-depth phenotyping. This likely contributes to the inconsistencies in GEWISs.

To circumvent power issues, the use of polygenic scores (PGSs) has become increasingly popular, with the aim of

capturing a larger amount of the trait-associated genetic variance with a single measure. PGSs are in principle weighted additive scores of all alleles that are associated with a trait of interest in a genome-wide association study (GWAS) (94). Such PGSs, based on large GWAS meta-analyses, have been shown to explain a substantial proportion of variance in psychiatric phenotypes, such as schizophrenia (95), and have also been used to explore G×Es. The largest PGS×E study to date applied an MDD PGS from a subsample of the Psychiatric Genomics Consortium wave 2 MDD GWAS cohort to a sample of 5765 subjects, with childhood trauma as the environmental parameter (96). This study showed evidence for additive, but not interactive, effects of this PGS and childhood trauma on MDD. Overall, this study and previous conflicting findings of MDD PGS×E effects on depression (96,97) and a recent study investigating the MDD PGS in children (98) do not support a consistent interaction of this PGS with early adversity.

In addition to its contingency on a sufficiently high-powered GWAS, the PGS×E approach suffers a more inherent conceptual limitation that might contribute to the inconsistent findings. The PGSs in all studies to date are weighted by a main effect on disease risk and not an interaction effect. However, it is possible that common genetic variants showing interaction effects might not have strong main effects on the target trait (99). Here alternative strategies could be of interest.

### **BOTTOM-UP MAPPING OF G×E: FROM MOLECULES TO INTERMEDIATE PHENOTYPES TO DIAGNOSES**

Another avenue toward the identification of resilience mechanisms and intermediate phenotypes could follow a bottom-up approach that starts with mapping regulatory gene variants associated with heightened sensitivity to environmental stimuli on a molecular level.

Exposure to environmental stimuli leads to a cascade of effects that, in the case of adversity, culminates in the activation of stress response systems such as the catecholaminergic system and the stress hormone system. This leads not only to a number of physiological changes but also to induction of changes in gene transcription and likely also epigenetic changes, especially with the activation of the GR. Several studies have indeed shown that GR activation leads to cell-specific transcriptional changes in thousands of genes as well as local epigenetic changes (100,101). A reduction of DNA methylation has been reported for some loci, which makes the transcript more responsive to subsequent GR stimuli (102). Possible mechanisms of resilience could directly relate to genetic moderation of these very proximal responses to stress, that is, greater or lesser impact of the environmental stimulus on the transcriptional or epigenetic level.

### **FKBP5: From Moderation of the Transcriptional Response to Disease Risk**

*FKBP5* was initially reported as a gene that is strongly responsive to glucocorticoids and in turn affects the regulation of the stress hormone system by altering GR sensitivity via direct protein–protein interactions (103). In a study examining genetic variants associated with antidepressant treatment response, we had initially identified that a haplotype in *FKBP5*

that moderates the induction of *FKBP5* messenger RNA by GR activation (104) and hypothesized that such a differential induction at the molecular level could also moderate long-term risk to psychiatric disorders after exposure to early adversity (18). In fact, a number of studies have now reported an interaction of this haplotype with early adversity to predict a range of psychiatric symptoms, as described in the first part of this review. The most likely functional SNP within this haplotype, rs1360780, alters a TATA box binding site close to a glucocorticoid response element in intron 2 of the gene. The allele with high messenger RNA induction that is also the risk allele leads to the presence of such a binding site and an altered three-dimensional conformation, with the glucocorticoid response element in intron 2 coming in proximity to the transcription start site (21). This genetic variant seems to alter direct effects of glucocorticoids not only on *FKBP5* messenger RNA but also on DNA methylation levels in regulatory elements of this gene. In fact, only individuals exposed to early adversity and carrying the high-risk haplotype also show reduced DNA methylation at another intronic glucocorticoid response element. In reporter gene assays, this reduction in DNA methylation led to increased responsiveness of the target gene to glucocorticoids and correlated with GR sensitivity (21). This supports a model in which functional genetic variants and exposure to early adversity lead to prolonged GR activation via increased *FKBP5* levels, which in turn has been shown to induce local DNA demethylation at the site of receptor binding (9). This reduced DNA methylation in an enhancer region increases the response to subsequent exposure to glucocorticoids, so that in exposed individuals carrying the risk haplotype both genetic and epigenetic factors lead to a disinhibited *FKBP5* response. Only these combined effects seem to increase disease risk given that main effects of the *FKBP5* genotypes are not consistently reported. *FKBP5* levels may reach only a certain threshold following this dual disinhibition, leading to altered downstream pathways of glycogen synthase kinase 3β or calcineurin signaling (105), decreased spine density (106), and behavioral effects such as increased anxiety and decreased stress coping (107,108).

### **Mapping Molecular G×E on a Genome-wide Level**

Investigating genetic variants that moderate the impact of environmental exposures on molecular response on a genome-wide level could thus identify novel genetic variants moderating risk and resilience to disorders. Such approaches have been proposed in general for the study of complex disease (109), and examples have been described in toxicology (110). Argos *et al.*, for example, explored the impact of genome-wide SNP × arsenic exposure on arsenic-associated gene expression and DNA methylation in peripheral blood to preselect SNPs that would moderate the extent of skin lesions with exposure (110).

We have adapted such screening approaches to follow up on our finding relating to *FKBP5* that variants that moderate the transcriptional impact of glucocorticoids could be of interest for stress-related psychiatric disorders. For this, we have mapped common regulatory gene variants associated with a differential transcriptional response to the GR agonist

dexamethasone (111), mimicking the activation of this receptor following stress exposure. These genetic variants are located in long-range enhancers with brain and cross-tissue relevance and are significantly more likely to show associations with MDD or schizophrenia in large GWASs than variants moderating baseline gene transcription (111). This suggests that an altered molecular responsivity to glucocorticoids contributes to risk for MDD and schizophrenia, with both disorders showing increased prevalence following exposure to early adversity. By constructing a PGS from the functional GR response SNPs that also associate with MDD, we found that this score that reflects a different molecular sensitivity to stress is also associated with an inappropriate amygdala activation during emotional fear processing (111) and increased cortisol secretion in response to a psychosocial stressor (112). In addition, this PGS also significantly correlated with both baseline values and stress-related changes of the brain's hemodynamic response function (HRF) (112). These HRF changes also correlated with a regulatory gene variant of *KCNJ2*, a gene that mediates glucocorticoid effects on neurovascular coupling in rodents (113). This supports the notion that the observed HRF changes reflect stress-related changes in neurovascular coupling, rendering HRF changes a promising novel intermediate stress phenotype that could help to link the action of stress effectors to pathological findings of chronic stress such as structural changes in the limbic system or prefrontal cortex (114).

In addition to gene expression, epigenetic changes, including DNA methylation, present another interesting intermediate phenotype for such approaches. Environmental perturbations can alter the epigenome, leading to sustained changes in gene transcription and thereby providing a molecular mechanism for altered biological processes following adverse environmental exposures and ultimately altered psychiatric risk later in life (115–119). Epigenetic changes are influenced by not only environmental factors but also genetic variation (21,120), so that epigenetic mechanisms could represent a molecular point of convergence for G×Es. Studies focusing on the developmental impact of adverse environment on the epigenome have shown that the environment alone is rarely the strongest predictor of epigenetic differences. Rather, environmental and genetic factors likely have additive or multiplicative roles in shaping the epigenome and consequently modulate phenotype presentation and disease risk (121). Teh *et al.* investigated the contribution of the prenatal environment as well as genetic variation in *cis* on DNA methylation of umbilical cord tissue in 237 neonates (122). The authors concluded that the large majority of interindividual DNA methylation variability was best explained by the interaction of genotype with different in utero environments rather than the environment or genetic variant independently. More recently, Czamara *et al.* extended these findings by exploring the influence of the prenatal environment and genotype on variably methylated regions in more than 2000 newborns (123). Their study confirmed that interactive and additive effects of genetic variation and prenatal environment were the best predictors of DNA methylation patterns as compared with genetic or environmental predictors alone. Importantly, their study could also show that genetic variants interacting with prenatal environment were enriched for disease-associated

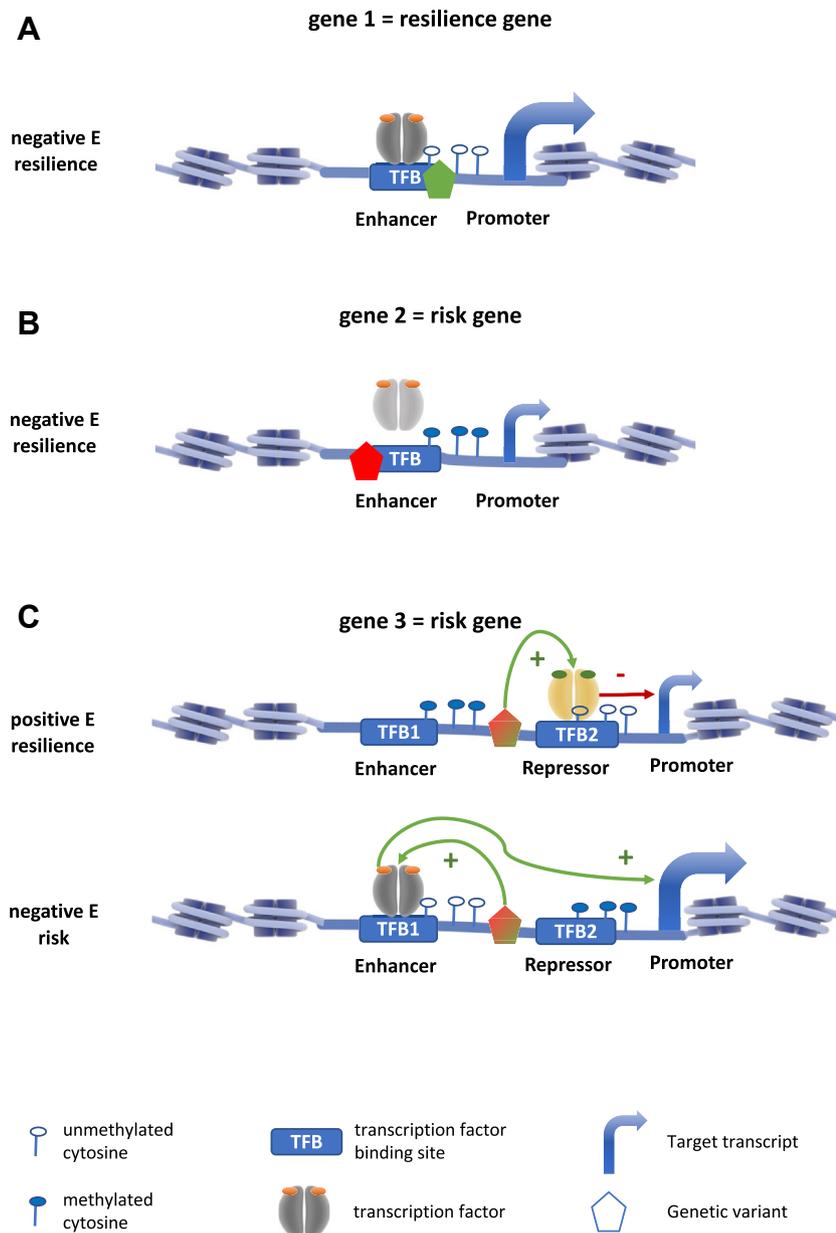
variants from large GWASs, including psychiatric disorders (123). While methylation quantitative trait loci have been previously used to functionally annotate GWAS findings, variants with G×E effects on DNA methylation extend these annotations because they often do not have a strong main genetic effect.

Genetic variants may influence outcome following exposure to adversity by either directly or indirectly altering the molecular consequences of exposure. Transcription factor-mediated epigenetic changes, such as the ones reported for the GR, could represent a possible target mechanism. Genetic variants may directly alter the binding affinity of the factor to the target enhancer site or may indirectly alter the activation of the factor and thus its downstream epigenetic consequences. Both the differential susceptibility hypothesis and diathesis stress hypothesis could be supported at this level with a number of possible different combinatorial mechanisms that are illustrated in Figure 2.

## FUTURE DIRECTIONS

Studying G×Es may provide a tool to dissect mechanisms of resilience. Studies exploring G×E on diagnostic outcomes have had a major conceptual impact on the field (13,81) but are fraught with issues of lack of replication. This relates to power issues, with initial discoveries in small samples, but also inherent problems of our field, with symptom-based and not mechanism-based diagnoses, increasing the heterogeneity within current diagnostic groups. In addition, current studies rarely rely on objective measures of environmental factors, and especially retrospective reports of adversity show only poor agreement with prospective assessments (124). In addition, most studies focus on one or a few environmental factors; however, it is clear that environmental factors influence each other and combine both positive and negative factors. To better address these complex interactions, the concept of an exposome has been brought forward in the study of complex disease (125). In fact, the National Institutes of Health-initiated Environmental influences on Child Health Outcomes program (which includes neurodevelopmental outcomes) has adopted a strategy of integrated exposure analysis, including inorganic and organic toxicants, nutrients, and social stress markers (126). Recent technological developments will facilitate more objective measures of environmental factors by integrating geographic information systems, remote sensing, and geolocation technologies but also portable and personal sensing that include smartphone-based sensors and assessments (127). Especially the latter will allow more reliable assessments of stressful life events. Large longitudinal cohorts, optimally starting at preconception and including such repeated measures, will be necessary for exposome × gene interaction analyses.

Technological advances will also improve our capacities for bottom-up mapping. For now, these approaches have mainly used molecular data derived from peripheral tissues, including blood. While peripheral blood is an important surrogate tissue, with immune cells likely contributing to the risk for a number of psychiatric disorders (128), it would be important to also investigate living cells from the peripheral nervous system at different developmental stages. This is now possible with the



**Figure 2.** Illustration of how genetic variants may promote risk or resilience via affecting epigenetic regulation in gene regulatory elements. **(A)** Situation in which a genetic variant is associated with epigenetic changes (reduction in DNA methylation) that increase binding of a transcription factor that would enhance the transcription of a gene that promotes resilience mechanisms. **(B)** Situation in which a genetic variant is associated with epigenetic changes (increase in DNA methylation) that decrease binding of a transcription factor that would enhance the transcription of a gene that promotes risk mechanisms. Overall, in both scenarios, these variants would promote a decrease in disease risk in the light of adversity. **(C)** Scenario of a genetic variant that is associated with differential susceptibility. In conjunction with a positive environment, the variant is associated with epigenetic silencing of an enhancer of a risk gene and epigenetic activation of a repressor for the same risk gene, thereby decreasing the expression of that risk gene and promoting positive outcomes. The bottom part of **(C)** depicts the situation with exposure to a negative environment. Here, the genetic variant promotes epigenetic silencing of the repressor and activation of the enhancer, thereby overall increasing the expression of the risk gene and promoting risk. E, environment.

advent of induced pluripotent stem cell–derived systems and a number of robust methods to differentiate into progenitors and different neuronal subtypes but also glia cells (129) and derived self-organizing three-dimensional structures (130). Such systems would allow measuring the molecular response in cells of the nervous system to proxies of environmental stimuli in the context of common human genetic variation. While this approach is attractive, it currently has limitations. First, we still need to better understand the relevant downstream effectors of adversity. While glucocorticoids are one potential mediator, there are others that might not be that easily studied *in vitro*. Second, the generation and comparable differentiation of hundreds of induced pluripotent stem cells is currently still a

labor-intensive and very costly endeavor. It is likely that a larger number of individual cell lines will be needed to reliably detect G×E on molecular features because, for example, gene expression profiles in induced pluripotent stem cell–derived motor neurons are more variable than those in primary neurons (131), thereby introducing additional noise. Large-scale perturbation assays, in which the functional effects of thousands of gene variants can be tested in a single cell line, may be an interesting avenue to overcome these issues and map G×E on molecular features (132,133).

Overall, exploring G×E to identify mechanisms of resilience could be a promising avenue, but a number of important challenges are still to be overcome.

## ACKNOWLEDGMENTS AND DISCLOSURES

This work was supported by an Alexander von Humboldt Fellowship and a Banting Postdoctoral Fellowship to CC.

EBB is coinventor of the patent application “FKBP5: A novel target for antidepressant therapy” (European Patent No. EP 1687443 B1). The other authors report no biomedical financial interests or potential conflicts of interest.

## ARTICLE INFORMATION

From the Department of Translational Research in Psychiatry (IGE, CC, EBB), Max Planck Institute of Psychiatry, Munich, Germany; and Department of Psychiatry and Behavioral Sciences (EBB), Emory University School of Medicine, Atlanta, Georgia.

Address correspondence to Elisabeth B. Binder, M.D., Kraepelinstrasse 2-10, 80804 Munich, Germany; E-mail: binder@psych.mpg.de.

Received Jan 16, 2019; revised Apr 8, 2019; accepted Apr 17, 2019.

## REFERENCES

- Russo SJ, Murrough JW, Han MH, Charney DS, Nestler EJ (2012): Neurobiology of resilience. *Nat Neurosci* 15:1475–1484.
- Schmitt A, Malchow B, Hasan A, Falkai P (2014): The impact of environmental factors in severe psychiatric disorders. *Front Neurosci* 8:19.
- Assary E, Vincent JP, Keers R, Pluess M (2018): Gene-environment interaction and psychiatric disorders: Review and future directions. *Semin Cell Dev Biol* 77:133–143.
- Halldorsdottir T, Binder EB (2017): Gene × environment interactions: From molecular mechanisms to behavior. *Annu Rev Psychol* 68:215–241.
- Gerke J, Koenig AM, Conrad D, Doyen-Waldeck C, Pauly M, Gündel H, *et al.* (2018): Childhood maltreatment as risk factor for lifetime depression: The role of different types of experiences and sensitive periods. *Ment Health Prev* 10:56–65.
- Border R, Johnson EC, Evans LM, Smolen A, Berley N, Sullivan PF, Keller MC (2019): No support for historical candidate gene or candidate gene-by-interaction hypotheses for major depression across multiple large samples. *Am J Psychiatry* 176:376–387.
- Koob GF (1999): Corticotropin-releasing factor, norepinephrine, and stress. *Biol Psychiatry* 46:1167–1180.
- Binder EB, Nemeroff CB (2010): The CRF system, stress, depression and anxiety—Insights from human genetic studies. *Mol Psychiatry* 15:574–588.
- Matosin N, Halldorsdottir T, Binder EB (2018): Understanding the molecular mechanisms underpinning gene by environment interactions in psychiatric disorders: The FKBP5 model. *Biol Psychiatry* 83:821–830.
- Heils A, Teufel A, Petri S, Stober G, Riederer P, Bengel D, *et al.* (1996): Allelic variation of human serotonin transporter gene expression. *J Neurochem* 66:2621–2624.
- Appel K, Schwahn C, Mahler J, Schulz A, Spitzer C, Fenske K, *et al.* (2011): Moderation of adult depression by a polymorphism in the FKBP5 gene and childhood physical abuse in the general population. *Neuropsychopharmacology* 36:1982–1991.
- Bradley RG, Binder EB, Epstein MP, Tang Y, Nair HP, Liu W, *et al.* (2008): Influence of child abuse on adult depression—Moderation by the corticotropin-releasing hormone receptor gene. *Arch Gen Psychiatry* 65:190–200.
- Caspi A, Sugden K, Moffitt TE, Taylor A, Craig IW, Harrington H, *et al.* (2003): Influence of life stress on depression: Moderation by a polymorphism in the 5-HTT gene. *Science* 301:386–389.
- Polanczyk G, Caspi A, Williams B, Price TS, Danese A, Sugden K, *et al.* (2009): Protective effect of CRHR1 gene variants on the development of adult depression following childhood maltreatment: Replication and extension. *Arch Gen Psychiatry* 66:978–985.
- Sharpley CF, Palanisamy SK, Glyde NS, Dillingham PW, Agnew LL (2014): An update on the interaction between the serotonin transporter promoter variant (5-HTTLPR), stress and depression, plus an exploration of non-confirming findings. *Behav Brain Res* 273:89–105.
- Wang Q, Shelton RC, Dwivedi Y (2018): Interaction between early-life stress and FKBP5 gene variants in major depressive disorder and post-traumatic stress disorder: A systematic review and meta-analysis. *J Affect Disord* 225:422–428.
- Zimmermann P, Bruckl T, Nocon A, Pfister H, Binder EB, Uhr M, *et al.* (2011): Interaction of FKBP5 gene variants and adverse life events in predicting depression onset: Results from a 10-year prospective community study. *Am J Psychiatry* 168:1107–1116.
- Binder EB, Bradley RG, Liu W, Epstein MP, Deveau TC, Mercer KB, *et al.* (2008): Association of FKBP5 polymorphisms and childhood abuse with risk of posttraumatic stress disorder symptoms in adults. *JAMA* 299:1291–1305.
- Gressier F, Calati R, Balestri M, Marsano A, Alberti S, Antypa N, *et al.* (2013): The 5-HTTLPR polymorphism and posttraumatic stress disorder: A meta-analysis. *J Trauma Stress* 26:645–653.
- Kilpatrick DG, Koenen KC, Ruggiero KJ, Acierno R, Galea S, Resnick HS, *et al.* (2007): The serotonin transporter genotype and social support and moderation of posttraumatic stress disorder and depression in hurricane-exposed adults. *Am J Psychiatry* 164:1693–1699.
- Klengel T, Mehta D, Anacker C, Rex-Haffner M, Pruessner JC, Pariante CM, *et al.* (2013): Allele-specific FKBP5 DNA demethylation mediates gene-childhood trauma interactions. *Nat Neurosci* 16:33–41.
- Koenen KC, Aiello AE, Bakshis E, Amstadter AB, Ruggiero KJ, Acierno R, *et al.* (2009): Modification of the association between serotonin transporter genotype and risk of posttraumatic stress disorder in adults by county-level social environment. *Am J Epidemiol* 169:704–711.
- Liu Y, Garrett ME, Dennis MF, Green KT, Ashley-Koch AE, Hauser MA, *et al.* (2015): An examination of the association between 5-HTTLPR, combat exposure, and PTSD diagnosis among U.S. veterans. *PLoS One* 10:e119998.
- Watkins LE, Han SZ, Harpaz-Rotem I, Mota NP, Southwick SM, Krystal JH, *et al.* (2016): FKBP5 polymorphisms, childhood abuse, and PTSD symptoms: Results from the National Health and Resilience in Veterans Study. *Psychoneuroendocrinology* 69:98–105.
- White S, Acierno R, Ruggiero KJ, Koenen KC, Kilpatrick DG, Galea S, *et al.* (2013): Association of CRHR1 variants and posttraumatic stress symptoms in hurricane exposed adults. *J Anxiety Disord* 27:678–683.
- Xie P, Kranzler HR, Poling J, Stein MB, Anton RF, Farrer LA, *et al.* (2010): Interaction of FKBP5 with childhood adversity on risk for post-traumatic stress disorder. *Neuropsychopharmacology* 35:1684–1692.
- Kolassa IT, Ertl V, Eckart C, Glockner F, Kolassa S, Papassotiropoulos A, *et al.* (2010): Association study of trauma load and SLC6A4 promoter polymorphism in posttraumatic stress disorder: Evidence from survivors of the Rwandan genocide. *J Clin Psychiatry* 71:543–547.
- Kranzler HR, Scott D, Tennen H, Feinn R, Williams C, Armeli S, *et al.* (2012): The 5-HTTLPR polymorphism moderates the effect of stressful life events on drinking behavior in college students of African descent. *Am J Med Genet B Neuropsychiatr Genet* 159B:484–490.
- Lieberman R, Armeli S, Scott DM, Kranzler HR, Tennen H, Covault J (2016): FKBP5 genotype interacts with early life trauma to predict heavy drinking in college students. *Am J Med Genet B Neuropsychiatr Genet* 171:879–887.
- Nylander I, Todkar A, Granholm L, Vrettou M, Bendre M, Boon W, *et al.* (2017): Evidence for a link between Fkbp5/FKBP5, early life social relations and alcohol drinking in young adult rats and humans. *Mol Neurobiol* 54:6225–6234.
- Schmid B, Blomeyer D, Treutlein J, Zimmermann US, Buchmann AF, Schmidt MH, *et al.* (2010): Interacting effects of CRHR1 gene and stressful life events on drinking initiation and progression among 19-year-olds. *Int J Neuropsychopharmacol* 13:703–714.

32. Treutlein J, Kissling C, Frank J, Wiemann S, Dong L, Depner M, *et al.* (2006): Genetic association of the human corticotropin releasing hormone receptor 1 (CRHR1) with binge drinking and alcohol intake patterns in two independent samples. *Mol Psychiatry* 11:594–602.
33. Collip D, Myin-Germeys I, Wichers M, Jacobs N, Derom C, Thiery E, *et al.* (2013): FKBP5 as a possible moderator of the psychosis-inducing effects of childhood trauma. *Br J Psychiatry* 202:261–268.
34. Bernardo M, Bioque M, Cabrera B, Lobo A, Gonzalez-Pinto A, Pina L, *et al.* (2017): Modelling gene-environment interaction in first episodes of psychosis. *Schizophr Res* 189:181–189.
35. Hornung OP, Heim CM (2014): Gene-environment interactions and intermediate phenotypes: Early trauma and depression. *Front Endocrinol* 5:14.
36. Bunea IM, Szentagotai-Tatar A, Miu AC (2017): Early-life adversity and cortisol response to social stress: A meta-analysis. *Transl Psychiatry* 7:1274.
37. De Kloet ER, Joëls M, Holsboer F (2005): Stress and the brain: From adaptation to disease. *Nat Rev Neurosci* 6:463–475.
38. Heuser I, Yassouridis A, Holsboer F (1994): The combined dexamethasone/CRH test: A refined laboratory test for psychiatric disorders. *J Psychiatr Res* 28:341–356.
39. Kirschbaum C, Pirke KM, Hellhammer DH (1993): The “Trier Social Stress Test”—A tool for investigating psychobiological stress responses in a laboratory setting. *Neuropsychobiology* 28:76–81.
40. Steptoe A, Serwinski B (2016): Cortisol awakening response. In: Fink G, editor. *Stress: Concepts, Cognition, Emotion, and Behavior*. San Diego: Academic Press, 277–283.
41. Buchmann AF, Holz N, Boecker R, Blomeyer D, Rietschel M, Witt SH, *et al.* (2014): Moderating role of FKBP5 genotype in the impact of childhood adversity on cortisol stress response during adulthood. *Eur Neuropsychopharmacol* 24:837–845.
42. Chen MC, Joormann J, Hallmayer J, Gotlib IH (2009): Serotonin transporter polymorphism predicts waking cortisol in young girls. *Psychoneuroendocrinology* 34:681–686.
43. Cicchetti D, Rogosch FA, Oshri A (2011): Interactive effects of corticotropin releasing hormone receptor 1, serotonin transporter linked polymorphic region, and child maltreatment on diurnal cortisol regulation and internalizing symptomatology. *Dev Psychopathol* 23:1125–1138.
44. Frigerio A, Ceppi E, Rusconi M, Giorda R, Raggi ME, Fearon P (2009): The role played by the interaction between genetic factors and attachment in the stress response in infancy. *J Child Psychol Psychiatry* 50:1513–1522.
45. Grad I, Picard D (2007): The glucocorticoid responses are shaped by molecular chaperones. *Mol Cell Endocrinol* 275:2–12.
46. Heim C, Bradley B, Mletzko TC, Deveau TC, Musselman DL, Nemeroff CB, *et al.* (2009): Effect of childhood trauma on adult depression and neuroendocrine function: Sex-specific moderation by CRH receptor 1 gene. *Front Behav Neurosci* 3:41.
47. Ising M, Depping AM, Siebertz A, Lucae S, Unschuld PG, Kloiber S, *et al.* (2008): Polymorphisms in the FKBP5 gene region modulate recovery from psychosocial stress in healthy controls. *Eur J Neurosci* 28:389–398.
48. Koenig AM, Ramo-Fernandez L, Boeck C, Umlauf M, Pauly M, Binder EB, *et al.* (2018): Intergenerational gene × environment interaction of FKBP5 and childhood maltreatment on hair steroids. *Psychoneuroendocrinology* 92:103–112.
49. Kohrt BA, Worthman CM, Ressler KJ, Mercer KB, Upadhyaya N, Koirala S, *et al.* (2015): Cross-cultural gene-environment interactions in depression, post-traumatic stress disorder, and the cortisol awakening response: FKBP5 polymorphisms and childhood trauma in South Asia. *Int Rev Psychiatry* 27:180–196.
50. Luijk MPCM, Velders FP, Thamer A, van IJzendoorn MH, Bakermans-Kranenburg MJ, Jaddoe VVW, *et al.* (2010): FKBP5 and resistant attachment predict cortisol reactivity in infants: Gene-environment interaction. *Psychoneuroendocrinology* 35:1454–1461.
51. Mahon PB, Zandi PP, Potash JB, Nestadt G, Wand GS (2013): Genetic association of FKBP5 and CRHR1 with cortisol response to acute psychosocial stress in healthy adults. *Psychopharmacology* 227:231–241.
52. McCormack K, Newman TK, Higley JD, Maestriepieri D, Sanchez MM (2009): Serotonin transporter gene variation, infant abuse, and responsiveness to stress in rhesus macaque mothers and infants. *Horm Behav* 55:538–547.
53. Mehta D, Gonik M, Klengel T, Rex-Haffner M, Menke A, Rubel J, *et al.* (2011): Using polymorphisms in FKBP5 to define biologically distinct subtypes of posttraumatic stress disorder: Evidence from endocrine and gene expression studies. *Arch Gen Psychiatry* 68:901–910.
54. Miller R, Wankerl M, Stalder T, Kirschbaum C, Alexander N (2013): The serotonin transporter gene-linked polymorphic region (5-HTTLPR) and cortisol stress reactivity: A meta-analysis. *Mol Psychiatry* 18:1018–1024.
55. Mueller A, Brocke B, Fries E, Lesch KP, Kirschbaum C (2010): The role of the serotonin transporter polymorphism for the endocrine stress response in newborns. *Psychoneuroendocrinology* 35:289–296.
56. Sheikh HI, Kryski KR, Smith HJ, Hayden EP, Singh SM (2013): Corticotropin-releasing hormone system polymorphisms are associated with children’s cortisol reactivity. *Neuroscience* 229:1–11.
57. Sumner JA, McLaughlin KA, Walsh K, Sheridan MA, Koenen KC (2014): CRHR1 genotype and history of maltreatment predict cortisol reactivity to stress in adolescents. *Psychoneuroendocrinology* 43:71–80.
58. Tyrka AR, Price LH, Gelernter J, Schepker C, Anderson GM, Carpenter LL (2009): Interaction of childhood maltreatment with the corticotropin-releasing hormone receptor gene: Effects on hypothalamic-pituitary-adrenal axis reactivity. *Biol Psychiatry* 66:681–685.
59. Vermeer H, Hendriks-Stegeman BI, van der Burg B, van Buul-Offers SC, Jansen M (2003): Glucocorticoid-induced increase in lymphocytic FKBP51 messenger ribonucleic acid expression: A potential marker for glucocorticoid sensitivity, potency, and bioavailability. *J Clin Endocrinol Metab* 88:277–284.
60. Shin LM, Liberzon I (2010): The neurocircuitry of fear, stress, and anxiety disorders. *Neuropsychopharmacology* 35:169–191.
61. Hamilton JP, Etkin A, Furman DJ, Lemus MG, Johnson RF, Gotlib IH (2012): Functional neuroimaging of major depressive disorder: A meta-analysis and new integration of baseline activation and neural response data. *Am J Psychiatry* 169:693–703.
62. Etkin A, Wager TD (2007): Functional neuroimaging of anxiety: A meta-analysis of emotional processing in PTSD, social anxiety disorder, and specific phobia. *Am J Psychiatry* 164:1476–1488.
63. Hulvershorn LA, Karne H, Gunn AD, Hartwick SL, Wang Y, Hummer TA, *et al.* (2012): Neural activation during facial emotion processing in unmedicated bipolar depression, euthymia, and mania. *Biol Psychiatry* 71:603–610.
64. Demers CH, Drabant Conley E, Bogdan R, Hariri AR (2016): Interactions between anandamide and corticotropin-releasing factor signaling modulate human amygdala function and risk for anxiety disorders: An imaging genetics strategy for modeling molecular interactions. *Biol Psychiatry* 80:356–362.
65. Holz NE, Buchmann AF, Boecker R, Blomeyer D, Baumeister S, Wolf I, *et al.* (2015): Role of FKBP5 in emotion processing: Results on amygdala activity, connectivity and volume. *Brain Struct Funct* 220:1355–1368.
66. Munafò MR, Brown SM, Hariri AR (2008): Serotonin transporter (5-HTTLPR) genotype and amygdala activation: A meta-analysis. *Biol Psychiatry* 63:852–857.
67. Murphy SE, Norbury R, Godlewska BR, Cowen PJ, Mannie ZM, Harmer CJ, *et al.* (2013): The effect of the serotonin transporter polymorphism (5-HTTLPR) on amygdala function: A meta-analysis. *Mol Psychiatry* 18:512–520.
68. White MG, Bogdan R, Fisher PM, Munoz KE, Williamson DE, Hariri AR (2012): FKBP5 and emotional neglect interact to predict individual differences in amygdala reactivity. *Genes Brain Behav* 11:869–878.

69. Kotov R, Gamez W, Schmidt F, Watson D (2010): Linking “big” personality traits to anxiety, depressive, and substance use disorders: A meta-analysis. *Psychol Bull* 136:768–821.
70. Ormel J, Jeronimus BF, Kotov R, Riese H, Bos EH, Hankin B, *et al.* (2013): Neuroticism and common mental disorders: Meaning and utility of a complex relationship. *Clin Psychol Rev* 33:686–697.
71. Munafo MR, Freimer NB, Ng W, Ophoff R, Veijola J, Miettunen J, *et al.* (2009): 5-HTTLPR genotype and anxiety-related personality traits: A meta-analysis and new data. *Am J Med Genet B Neuropsychiatr Genet* 150B:271–281.
72. Terracciano A, Tanaka T, Sutin AR, Deiana B, Balaci L, Sanna S, *et al.* (2010): BDNF Val66Met is associated with introversion and interacts with 5-HTTLPR to influence neuroticism. *Neuropsychopharmacology* 35:1083–1089.
73. DeYoung CG, Cicchetti D, Rogosch FA (2011): Moderation of the association between childhood maltreatment and neuroticism by the corticotropin-releasing hormone receptor 1 gene. *J Child Psychol Psychiatry* 52:898–906.
74. Perez-Perez B, Cristobal-Narvaez P, Sheinbaum T, Kwapił TR, Ballesteri S, Pena E, *et al.* (2018): Interaction between FKBP5 variability and recent life events in the anxiety spectrum: Evidence for the differential susceptibility model. *PLoS One* 13:e193044.
75. Culverhouse RC, Saccone NL, Horton AC, Ma Y, Anstey KJ, Banaschewski T, *et al.* (2018): Collaborative meta-analysis finds no evidence of a strong interaction between stress and 5-HTTLPR genotype contributing to the development of depression. *Mol Psychiatry* 23:133–142.
76. Risch N, Herrell R, Lehner T, Liang KY, Eaves L, Hoh J, *et al.* (2009): Interaction between the serotonin transporter gene (5-HTTLPR), stressful life events, and risk of depression: A meta-analysis. *JAMA* 301:2462–2471.
77. Karg K, Burmeister M, Shedden K, Sen S (2011): The serotonin transporter promoter variant (5-HTTLPR), stress, and depression meta-analysis revisited: Evidence of genetic moderation. *Arch Gen Psychiatry* 68:444–454.
78. Bustamante AC, Aiello AE, Guffanti G, Galea S, Wildman DE, Uddin M (2018): FKBP5 DNA methylation does not mediate the association between childhood maltreatment and depression symptom severity in the Detroit Neighborhood Health Study. *J Psychiatr Res* 96:39–48.
79. Isaksson J, Comasco E, Aslund C, Rehn M, Tuvblad C, Andershed H, *et al.* (2016): Associations between the FKBP5 haplotype, exposure to violence and anxiety in females. *Psychoneuroendocrinology* 72:196–204.
80. Kranzler HR, Feinn R, Nelson EC, Covault J, Anton RF, Farrer L, *et al.* (2011): A CRHR1 haplotype moderates the effect of adverse childhood experiences on lifetime risk of major depressive episode in African-American women. *Am J Med Genet B Neuropsychiatr Genet* 156B:960–968.
81. Caspi A, Hariri AR, Holmes A, Uher R, Moffitt TE (2010): Genetic sensitivity to the environment: The case of the serotonin transporter gene and its implications for studying complex diseases and traits. *Am J Psychiatry* 167:509–527.
82. Uher R, McGuffin P (2008): The moderation by the serotonin transporter gene of environmental adversity in the aetiology of mental illness: Review and methodological analysis. *Mol Psychiatry* 13:131–146.
83. Kolassa IT, Ertl V, Eckart C, Onyut LP, Kolassa S, Elbert T (2010): Spontaneous remission from PTSD depends on the number of traumatic event types experienced. *Psychol Trauma* 2:169–174.
84. Conrad D, Wilker S, Pfeiffer A, Lingenfelder B, Ebalu T, Lanzinger H, *et al.* (2017): Does trauma event type matter in the assessment of traumatic load? *Eur J Psychotraumatol* 8:1344079.
85. Wilker S, Pfeiffer A, Kolassa S, Koslowski D, Elbert T, Kolassa IT (2015): How to quantify exposure to traumatic stress? Reliability and predictive validity of measures for cumulative trauma exposure in a post-conflict population. *Eur J Psychotraumatol* 6:28306.
86. Aschard H, Lutz S, Maus B, Duell EJ, Fingerlin TE, Chatterjee N, *et al.* (2012): Challenges and opportunities in genome-wide environmental interaction (GWEI) studies. *Hum Genet* 131:1591–1613.
87. Aschard H, Tobin MD, Hancock DB, Skurnik D, Sood A, James A, *et al.* (2017): Evidence for large-scale gene-by-smoking interaction effects on pulmonary function. *Int J Epidemiol* 46:894–904.
88. Molfino NA, Coyle AJ (2008): Gene-environment interactions in chronic obstructive pulmonary disease. *Int J Chron Obstruct Pulmon Dis* 3:491–497.
89. Arnau-Soler A, Macdonald-Dunlop E, Adams MJ, Clarke T, MacIntyre DJ, Milburn K, *et al.* (2019): Genome-wide by environment interaction studies (GWEIS) of depressive symptoms and psychosocial stress in UK Biobank and Generation Scotland. *Transl Psychiatry* 9:14.
90. Coleman JRI, Peyrot WJ, Purves KL, Davis KAS, Rayner C, Choi SW, *et al.* (2018): Genome-wide gene-environment analyses of major depressive disorder and reported lifetime traumatic experiences in UK Biobank [published online ahead of print Nov 1]. *BioRxiv*.
91. Dunn EC, Wiste A, Radmanesh F, Almli LM, Gogarten SM, Sofer T, *et al.* (2016): Genome-wide association study (GWAS) and genome-wide by environment interaction study (GWEIS) of depressive symptoms in African American and Hispanic/Latina women. *Depress Anxiety* 33:265–280.
92. Ikeda M, Shimasaki A, Takahashi A, Kondo K, Saito T, Kawase K, *et al.* (2016): Genome-wide environment interaction between depressive state and stressful life events. *J Clin Psychiatry* 77:e29–e30.
93. Otowa T, Kawamura Y, Tsutsumi A, Kawakami N, Kan C, Shimada T, *et al.* (2016): The first pilot genome-wide gene-environment study of depression in the Japanese population. *PLoS One* 11:e160823.
94. Maier RM, Visscher PM, Robinson MR, Wray NR (2018): Embracing polygenicity: A review of methods and tools for psychiatric genetics research. *Psychol Med* 48:1055–1067.
95. Schizophrenia Working Group of the Psychiatric Genomics Consortium (2014): Biological insights from 108 schizophrenia-associated genetic loci. *Nature* 511:421–427.
96. Peyrot WJ, Van der Auwera S, Milaneschi Y, Dolan CV, Madden PAF, Sullivan PF, *et al.* (2018): Does childhood trauma moderate polygenic risk for depression? A meta-analysis of 5765 subjects from the Psychiatric Genomics Consortium. *Biol Psychiatry* 84:138–147.
97. Mullins N, Power RA, Fisher HL, Hanscombe KB, Euesden J, Iniesta R, *et al.* (2016): Polygenic interactions with environmental adversity in the aetiology of major depressive disorder. *Psychol Med* 46:759–770.
98. Halldorsdottir T, Piechaczek C, Soares de Matos AP, Czamara D, Pehl V, Wagenbuechler P, *et al.* (2019): Polygenic risk: Predicting depression outcomes in clinical and epidemiological cohorts of youths. *Am J Psychiatry* 176:615–625.
99. Engelman CD, Baurley JW, Chiu YF, Joubert BR, Lewinger JP, Maenner MJ, *et al.* (2009): Detecting gene-environment interactions in genome-wide association data. *Genet Epidemiol* 33(suppl 1):S68–S73.
100. Gray JD, Kogan JF, Marrocco J, McEwen BS (2017): Genomic and epigenomic mechanisms of glucocorticoids in the brain. *Nat Rev Endocrinol* 13:661–673.
101. Grontved L, John S, Baek S, Liu Y, Buckley JR, Vinson C, *et al.* (2013): C/EBP maintains chromatin accessibility in liver and facilitates glucocorticoid receptor recruitment to steroid response elements. *EMBO J* 32:1568–1583.
102. Kress C, Thomassin H, Grange T (2006): Active cytosine demethylation triggered by a nuclear receptor involves DNA strand breaks. *Proc Natl Acad Sci U S A* 103:11112–11117.
103. Jaaskelainen T, Makkonen H, Palvimo JJ (2011): Steroid up-regulation of FKBP51 and its role in hormone signaling. *Curr Opin Pharmacol* 11:326–331.
104. Binder EB, Salyakina D, Lichtner P, Wochnik GM, Ising M, Putz B, *et al.* (2004): Polymorphisms in FKBP5 are associated with increased recurrence of depressive episodes and rapid response to antidepressant treatment. *Nat Genet* 36:1319–1325.
105. Gassen NC, Hartmann J, Zannas AS, Kretschmar A, Zschocke J, Maccarone G, *et al.* (2016): FKBP51 inhibits GSK3 $\beta$  and augments the effects of distinct psychotropic medications. *Mol Psychiatry* 21:277–289.

106. Young KA, Thompson PM, Cruz DA, Williamson DE, Selemon LD (2015): BA11 FKBP5 expression levels correlate with dendritic spine density in postmortem PTSD and controls. *Neurobiol Stress* 2:67–72.
107. Hartmann J, Wagner KV, Liebl C, Scharf SH, Wang X-D, Wolf M, *et al.* (2012): The involvement of FK506-binding protein 51 (FKBP5) in the behavioral and neuroendocrine effects of chronic social defeat stress. *Neuropharmacology* 62:332–339.
108. Touma C, Gassen NC, Herrmann L, Cheung-Flynn J, Bull DR, Ionescu IA, *et al.* (2011): FK506 binding protein 5 shapes stress responsiveness: Modulation of neuroendocrine reactivity and coping behavior. *Biol Psychiatry* 70:928–936.
109. McAllister K, Mechanic LE, Amos C, Aschard H, Blair IA, Chatterjee N, *et al.* (2017): Current challenges and new opportunities for gene-environment interaction studies of complex diseases. *Am J Epidemiol* 186:753–761.
110. Argos M, Tong L, Roy S, Sabarinathan M, Ahmed A, Islam MT, *et al.* (2018): Screening for gene-environment (G×E) interaction using omics data from exposed individuals: An application to gene-arsenic interaction. *Mamm Genome* 29:101–111.
111. Arloth J, Bogdan R, Weber P, Frishman G, Menke A, Wagner Klaus V, *et al.* (2015): Genetic differences in the immediate transcriptome response to stress predict risk-related brain function and psychiatric disorders. *Neuron* 86:1189–1202.
112. Elbau IG, Bruckmeier B, Uhr M, Arloth J, Czamara D, Spoomaker VI, *et al.* (2018): The brain's hemodynamic response function rapidly changes under acute psychosocial stress in association with genetic and endocrine stress response markers. *Proc Natl Acad Sci U S A* 115:E10206–E10215.
113. Longden TA, Dabertrand F, Hill-Eubanks DC, Hammack SE, Nelson MT (2014): Stress-induced glucocorticoid signaling remodels neurovascular coupling through impairment of cerebrovascular inwardly rectifying K<sup>+</sup> channel function. *Proc Natl Acad Sci U S A* 111:7462–7467.
114. Goodkind M, Eickhoff SB, Oathes DJ, Jiang Y, Chang A, Jones-Hagata LB, *et al.* (2015): Identification of a common neurobiological substrate for mental illness. *JAMA Psychiatry* 72:305–315.
115. Bale TL (2015): Epigenetic and transgenerational reprogramming of brain development. *Nat Rev Neurosci* 16:332–344.
116. Binder EB (2017): Dissecting the molecular mechanisms of gene × environment interactions: Implications for diagnosis and treatment of stress-related psychiatric disorders. *Eur J Psychotraumatol* 8:1412745.
117. Demetriou CA, van Veldhoven K, Relton C, Stringhini S, Kyriacou K, Vineis P (2015): Biological embedding of early-life exposures and disease risk in humans: A role for DNA methylation. *Eur J Clin Invest* 45:303–332.
118. Klengel T, Binder EB (2015): Epigenetics of stress-related psychiatric disorders and gene × environment interactions. *Neuron* 86:1343–1357.
119. Zannas AS, West AE (2014): Epigenetics and the regulation of stress vulnerability and resilience. *Neuroscience* 264:157–170.
120. Klengel T, Binder EB (2013): Allele-specific epigenetic modification: A molecular mechanism for gene-environment interactions in stress-related psychiatric disorders? *Epigenomics* 5:109–112.
121. Gluckman PD, Hanson MA, Cooper C, Thornburg KL (2008): Effect of in utero and early-life conditions on adult health and disease. *N Engl J Med* 359:61–73.
122. Teh AL, Pan H, Chen L, Ong ML, Dogra S, Wong J, *et al.* (2014): The effect of genotype and in utero environment on interindividual variation in neonate DNA methylomes. *Genome Res* 24:1064–1074.
123. Czamara D, Eraslan G, Lahti J, Page CM, Lahti-Pulkkinen M, Hämäläinen E, *et al.* (2018): Variably methylated regions in the newborn epigenome: Environmental, genetic and combined influences [published online ahead of print Oct 17]. *BioRxiv*.
124. Baldwin JR, Reuben A, Newbury JB, Danese A (2019): Agreement between prospective and retrospective measures of childhood maltreatment: A systematic review and meta-analysis. *JAMA Psychiatry* 76:584–593.
125. Wild CP (2012): The exposome: From concept to utility. *Int J Epidemiol* 41:24–32.
126. Wright RO, Teitelbaum S, Thompson C, Balshaw D (2018): The Child Health Exposure Analysis Resource as a vehicle to measure environment in the Environmental Influences on Child Health Outcomes program. *Curr Opin Pediatr* 30:285–291.
127. Turner MC, Nieuwenhuijsen M, Anderson K, Balshaw D, Cui Y, Dunton G, *et al.* (2017): Assessing the exposome with external measures: Commentary on the state of the science and research recommendations. *Annu Rev Public Health* 38:215–239.
128. Network and Pathway Analysis Subgroup of Psychiatric Genomics Consortium (2015): Psychiatric genome-wide association study analyses implicate neuronal, immune and histone pathways. *Nat Neurosci* 18:199–209.
129. Brennan KJ, Simone A, Tran N, Gage FH (2012): Modeling psychiatric disorders at the cellular and network levels. *Mol Psychiatry* 17:1239–1253.
130. Pasca SP (2018): The rise of three-dimensional human brain cultures. *Nature* 553:437–445.
131. Schwartztruber J, Foskolou S, Kilpinen H, Rodrigues J, Alasoo K, Knights AJ, *et al.* (2018): Molecular and functional variation in iPSC-derived sensory neurons. *Nat Genet* 50:54–61.
132. Sharon E, Chen SA, Khosla NM, Smith JD, Pritchard JK, Fraser HB (2018): Functional genetic variants revealed by massively parallel precise genome editing. *Cell* 175:544–557.
133. Ma L, Boucher JI, Paulsen J, Matuszewski S, Eide CA, Ou J, *et al.* (2017): CRISPR-Cas9-mediated saturated mutagenesis screen predicts clinical drug resistance with improved accuracy. *Proc Natl Acad Sci U S A* 114:11751–11756.