



## Genetic Meningococcal Antigen Typing System (gMATS): A genotyping tool that predicts 4CMenB strain coverage worldwide <sup>☆</sup>



Alessandro Muzzi <sup>a</sup>, Alessandro Brozzi <sup>a</sup>, Laura Serino <sup>a</sup>, Margherita Bodini <sup>a</sup>, Raquel Abad <sup>b</sup>, Dominique Caugant <sup>c</sup>, Maurizio Comanducci <sup>a,1</sup>, Ana Paula Lemos <sup>d</sup>, Maria Cecilia Gorla <sup>d</sup>, Pavla Křížová <sup>e</sup>, Claudia Mikula <sup>f</sup>, Robert Mulhall <sup>g</sup>, Michael Nissen <sup>h,2</sup>, Hanna Nohynek <sup>i</sup>, Maria João Simões <sup>j</sup>, Anna Skoczyńska <sup>k</sup>, Paola Stefanelli <sup>l</sup>, Muhamed-Kheir Taha <sup>m</sup>, Maija Toropainen <sup>i</sup>, Georgina Tzanakaki <sup>n</sup>, Kumaran Vadivelu-Pechai <sup>o</sup>, Philip Watson <sup>o</sup>, Julio A. Vazquez <sup>b</sup>, Gowrisankar Rajam <sup>p,3</sup>, Rino Rappuoli <sup>a</sup>, Ray Borrow <sup>q</sup>, Duccio Medini <sup>a,\*</sup>

<sup>a</sup> GSK, Siena, Italy

<sup>b</sup> National Centre for Microbiology, Institute of Health Carlos III, Madrid, Spain

<sup>c</sup> Norwegian Institute of Public Health, Oslo, Norway

<sup>d</sup> Instituto Adolfo Lutz, São Paulo, Brazil

<sup>e</sup> National Institute of Public Health, Prague, Czech Republic

<sup>f</sup> Austrian Agency for Health and Food Safety, Institute for Medical Microbiology and Hygiene, Graz, Austria

<sup>g</sup> Irish Meningitis and Sepsis Reference Laboratory (IMSRL), Dublin, Ireland

<sup>h</sup> Queensland Paediatric Infectious Diseases Laboratory, Children's Health Research Centre, University of Queensland, Lady Cilento Children's Hospital South Brisbane, Queensland, Australia

<sup>i</sup> National Institute for Health and Welfare (THL), Helsinki, Finland

<sup>j</sup> National Institute of Health Dr. Ricardo Jorge, Lisbon, Portugal

<sup>k</sup> National Medicines Institute, Warsaw, Poland

<sup>l</sup> Department of Infectious Parasitic and Immune-mediated Diseases, Istituto Superiore di Sanità, Rome, Italy

<sup>m</sup> Institut Pasteur, Paris, France

<sup>n</sup> National Meningitis Reference Laboratory, National School of Public Health, Athens, Greece

<sup>o</sup> GSK, Rockville, USA

<sup>p</sup> Centers for Disease Control and Prevention, Atlanta, GA, USA

<sup>q</sup> Meningococcal Reference Unit, Public Health England, Manchester Royal Infirmary, Manchester, UK

### ARTICLE INFO

#### Article history:

Received 29 August 2018

Received in revised form 17 December 2018

### ABSTRACT

**Background:** The Meningococcal Antigen Typing System (MATS) was developed to identify meningococcus group B strains with a high likelihood of being covered by the 4CMenB vaccine, but is limited by the requirement for viable isolates from culture-confirmed cases. We examined if antigen genotyping could complement MATS in predicting strain coverage by the 4CMenB vaccine.

**Abbreviations:** Ca, Lin's coefficient of accuracy; IDs, identification numbers (protein variants); ELISA, enzyme-linked immunosorbent assays; fHbp, factor H-binding protein; hSBA, serum bactericidal antibody assay with human complement; IMD, invasive meningococcal disease; MATS, Meningococcal Antigen Typing System; NadA, *Neisseria* adhesin A; NIP, national immunization program; NHBA, Neisserial Heparin-Binding Antigen; OMV, outer membrane vesicles; PBT, positive bactericidal threshold; PorA, porin A protein; RP, relative potency; VE, vaccine effectiveness; VR2, variable region 2.

\* **Keypoints:** The Genetic Meningococcal Antigen Typing System (gMATS), based on antigen genotyping, complements MATS in predicting 4CMenB strain coverage without requiring a cultivable isolate. Use of gMATS instead of MATS may simplify 4CMenB strain coverage prediction and accelerate 4CMenB protection surveillance.

\* Corresponding author at: GSK, Via Fiorentina 1, Siena, Italy.

**E-mail addresses:** [alessandro.x.muzzi@gsk.com](mailto:alessandro.x.muzzi@gsk.com) (A. Muzzi), [alessandro.x.brozzi@gsk.com](mailto:alessandro.x.brozzi@gsk.com) (A. Brozzi), [laura.x.serino@gsk.com](mailto:laura.x.serino@gsk.com) (L. Serino), [margherita.x.bodini@gsk.com](mailto:margherita.x.bodini@gsk.com) (M. Bodini), [rabad@isciii.es](mailto:rabad@isciii.es) (R. Abad), [Dominique.Caugant@fhi.no](mailto:Dominique.Caugant@fhi.no) (D. Caugant), [ana.lemos@ial.sp.gov.br](mailto:ana.lemos@ial.sp.gov.br) (A.P. Lemos), [maria.gorla@ial.sp.gov.br](mailto:maria.gorla@ial.sp.gov.br) (M.C. Gorla), [pavla.krizova@szu.cz](mailto:pavla.krizova@szu.cz) (P. Křížová), [claudia.mikula@ages.at](mailto:claudia.mikula@ages.at) (C. Mikula), [Robert.Mulhall@cuh.ie](mailto:Robert.Mulhall@cuh.ie) (R. Mulhall), [michael.d.nissen@gsk.com](mailto:michael.d.nissen@gsk.com) (M. Nissen), [hanna.nohynek@thl.fi](mailto:hanna.nohynek@thl.fi) (H. Nohynek), [M.Joao.Simoes@insa.min-saude.pt](mailto:M.Joao.Simoes@insa.min-saude.pt) (M.J. Simões), [skoczek@cls.edu.pl](mailto:skoczek@cls.edu.pl) (A. Skoczyńska), [paola.stefanelli@iss.it](mailto:paola.stefanelli@iss.it) (P. Stefanelli), [muhamed-kheir.taha@pasteur.fr](mailto:muhamed-kheir.taha@pasteur.fr) (M.-K. Taha), [maija.toropainen@thl.fi](mailto:maija.toropainen@thl.fi) (M. Toropainen), [gtzanakaki@esdy.edu.gr](mailto:gtzanakaki@esdy.edu.gr) (G. Tzanakaki), [kumaran.k.vadivelu-pechai@gsk.com](mailto:kumaran.k.vadivelu-pechai@gsk.com) (K. Vadivelu-Pechai), [philip.s.watson@gsk.com](mailto:philip.s.watson@gsk.com) (P. Watson), [jvazquez@isciii.es](mailto:jvazquez@isciii.es) (J.A. Vazquez), [rino.r.rappuoli@gsk.com](mailto:rino.r.rappuoli@gsk.com) (R. Rappuoli), [Ray.Borrow@phe.gov.uk](mailto:Ray.Borrow@phe.gov.uk) (R. Borrow), [duccio.x.medini@gsk.com](mailto:duccio.x.medini@gsk.com) (D. Medini).

<sup>1</sup> Present Address: Via L. da Vinci 4, 44042 Cento, FE, Italy.

<sup>2</sup> Present Address: Research and Development, GSK Intercontinental, 23 Rochester Park, Singapore.

<sup>3</sup> Present Address: Merck & Co., Inc., PA, USA.

<https://doi.org/10.1016/j.vaccine.2018.12.061>

0264-410X/© 2019 GlaxoSmithKline Biologicals SA. Published by Elsevier Ltd.

This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Accepted 20 December 2018  
Available online 17 January 2019

**Keywords:**

*Neisseria meningitidis* serogroup B  
4CMenB vaccine  
Strain coverage  
gMATS  
Genotyping

**Methods:** From a panel of 3912 MATS-typed invasive meningococcal disease isolates collected in England and Wales in 2007–2008, 2014–2015 and 2015–2016, and in 16 other countries in 2000–2015, 3481 isolates were also characterized by antigen genotyping. Individual associations between antigen genotypes and MATS coverage for each 4CMenB component were used to define a genetic MATS (gMATS). gMATS estimates were compared with England and Wales human complement serum bactericidal assay (hSBA) data and vaccine effectiveness (VE) data from England.

**Results:** Overall, 81% of the strain panel had genetically predictable MATS coverage, with 92% accuracy and highly concordant results across national panels (Lin's accuracy coefficient, 0.98; root-mean-square deviation, 6%). England and Wales strain coverage estimates were 72–73% by genotyping (66–73% by MATS), underestimating hSBA values after four vaccine doses (88%) and VE after two doses (83%). The gMATS predicted strain coverage in other countries was 58–88%.

**Conclusions:** gMATS can replace MATS in predicting 4CMenB strain coverage in four out of five cases, without requiring a cultivable isolate, and is open to further improvement. Both methods underestimated VE in England. Strain coverage predictions in other countries matched or exceeded England and Wales estimates.

An Audio Summary linked to this article that can be found on Figshare [https://figshare.com/articles/gMATS\\_video\\_Vaccine-2019-37\\_7\\_991-1000\\_mp4/9326687](https://figshare.com/articles/gMATS_video_Vaccine-2019-37_7_991-1000_mp4/9326687)

© 2019 GlaxoSmithKline Biologicals SA. Published by Elsevier Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

*Neisseria meningitidis* is the leading cause of bacterial meningitis and sepsis in children and young adults worldwide [1]. With licensure of 4CMenB (Bexsero, GSK), the first broad coverage meningococcus group B (MenB) vaccine, invasive meningococcal disease (IMD) has become a vaccine-preventable disease [2,3]. 4CMenB is a multicomponent vaccine that includes three recombinant protein antigens, *Neisseria* adhesin A (NadA), Neisserial Heparin-Binding Antigen (NHBA), and factor H-binding protein (fHbp), plus detergent-extracted outer membrane vesicles (OMV) obtained from the New Zealand outbreak strain, containing porin A protein (PorA) as main vaccine antigen [4].

Since the incidence of IMD is relatively low, effectiveness of meningococcal vaccines has been evaluated via an accepted surrogate of protection: the serum bactericidal antibody assay with human complement (hSBA) [5]. Assessing protein-based vaccines such as 4CMenB is complicated by diversity in the sequence and expression of MenB surface protein antigens [6]. Predicting efficacy would require hSBA investigation against a large panel of genetically diverse and geographically representative pathogenic strains, which is impractical due to the volumes of serum required [6,7].

The Meningococcal Antigen Typing System (MATS) was developed to predict strain coverage by 4CMenB from large panels of meningococcal isolates, independent of human sera [6,8]. MATS combines genotyping for PorA with three enzyme-linked immunosorbent assays (ELISA) for fHbp, NHBA, and NadA, that quantify in meningococcal strains the relative expression and cross-reactivity of antigenic variants with vaccine-induced antibody. Internationally-standardized MATS [9] was shown to provide a conservative prediction of hSBA results from pooled human sera [10] and indicated high rates of individual seroprotection [11], ranging from 66% to 91% in 14 countries [8,12–20].

The UK was the first country to introduce 4CMenB into its national immunization program (NIP). From September 2015, a reduced two-dose priming schedule was offered to infants aged 2 and 4 months, with a booster at 12 months, along with limited catch-up for those born after April 2015 [21]. In the first 10 months following introduction, two-dose vaccine effectiveness (VE) was 83% (95% confidence interval [95% CI]: 24–95%) against MenB cases, equivalent to 94% effectiveness against all cases predicted to be prevented by the vaccine [22].

A limitation of hSBA unresolved by MATS is the inability to use the assay in non-culture confirmed cases. In some countries, around half of cases are identified solely by PCR [23,24]. Also, MATS requires effort in maintaining the international standardiza-

tion level, as for all immunoassays. A genome-based vaccine strain coverage predictor would provide a more easily accessible, rapid, portable, comprehensive, and cost-effective alternative to both hSBA and MATS assays.

Here, we present 4CMenB vaccine antigen genotyping and MATS data on over 3000 MenB invasive disease isolates, representing all notified meningococcal cases in England and Wales in three epidemiological years together with isolates collected in 16 other countries. We analyzed the correlation between individual antigen genotypes and coverage estimates by MATS to define gMATS, a new genetic Meningococcal Antigen Typing System.

Fig. 1 summarizes the research, clinical relevance and impact on the patient population.

## 2. Methods

### 2.1. Bacterial strains

MATS typing was collected for 3912 invasive MenB isolates from 13 European and 4 non-European countries (Table 1), recovered from the blood or cerebrospinal fluid of patients and representative of national epidemiology in each country at the time of testing. A subset of 3481 isolates was also typed using antigen genotyping.

### 2.2. Strain coverage prediction by MATS

Levels of expression and cross-reactivity of fHbp, NadA, and NHBA were analyzed using the MATS ELISA, according to the method described previously [6]. Vaccine coverage for these three components was predicted by defining a relative potency (RP) of the tested strain versus a reference strain for each antigen and then comparing the RP with a positive bactericidal threshold (PBT), the antigen-specific minimum RP value predictive of killing in the hSBA. If the RP exceeds the PBT for a given antigen, the strain is predicted as covered by that vaccine antigen. Contribution to coverage by the OMV component was estimated by genotyping, by sequencing part of the *porA* gene encoding variable region 2 (VR2) and checking identity to the variant present in the vaccine, i.e. PorA VR2 match with peptide 4 was defined as 'covered' (PorA VR2 = 4) and other cases as 'not covered' [6].

### 2.3. Strain coverage prediction by genotyping

For the 3481 isolates characterized by genotyping (Table 1), *fHbp* and *nhba* genes were PCR amplified and sequenced, or their sequences were extracted from the whole genome sequence when

# Focus on the Patient

## What is the context?

- 4CMenB is a 4-component, meningococcal group B (MenB) vaccine developed to protect against related invasive disease.
- The meningococcal Antigen Typing System (MATS) assay was developed to estimate the proportion of MenB strains against which 4CMenB vaccinees are protected in a given region.
- The MATS assay requires viable isolates and therefore cannot be applied to non-culture confirmed cases

## What is new?

- We defined a genetic MATS (gMATS) for predicting strain coverage by associating antigen genotyping and MATS results, using European and non-EU MenB strain panels.
- In addition to viable isolates, gMATS can be applied to PCR confirmed non-culturable disease samples.
- We observed concordant results across national strain panels with gMATS, although both MATS and gMATS underestimated the effectiveness of the vaccine.

## What is the take-home message?

- Our results suggest that gMATS complement MATS in the estimation of protection provided by 4CMenB vaccination.
- This assay may help simplify estimation of vaccine strain coverage and accelerate surveillance of 4CMenB vaccine protection.

Fig. 1. Focus on the Patient section.

**Table 1**

The global collection of 3912 invasive meningococcal B isolates characterized using the Meningococcal Antigen Typing System (MATS) to estimate 4CMenB vaccine coverage. A subset of 3481 isolates, collected in ten European countries and three non-European countries, was also characterized by antigen genotype.

Country (years)	Number of isolates	Predicted 4CMenB strain coverage by MATS, % (95% CI)	Reference
<i>European countries</i>			
Austria (2008–11)	118	68 (56–73)	Unpublished
Czech Republic (2007–10)	108	74 (58–87)	[12]
Finland (2010–14)	60 <sup>a</sup>	78 (72–88)	[20]
France (2007–08)	200 <sup>a</sup>	85 (69–93)	[12]
(2013–14)	172 <sup>a</sup>	70 (60–80)	Unpublished
Germany (2007–08)	222 <sup>a</sup>	82 (69–92)	[12]
Greece (2008–10)	52 <sup>a</sup>	88 (60–96)	[13]
Ireland (2009–13)	105 <sup>a</sup>	69 (65–85)	Unpublished
Italy (2007–08)	54 <sup>a</sup>	87 (70–93)	[12]
Norway (2007–08)	41 <sup>a</sup>	85 (76–98)	[12]
Poland (2010–11)	196 <sup>a</sup>	84 (79–91)	[14]
Portugal (2011–15)	106	68 (56–81)	[15]
Spain (2009–10)	300 <sup>a</sup>	69 (48–85)	[17]
UK (2007–08)	535 <sup>a</sup>	73 (57–87)	[12]
(2014–15)	251 <sup>a</sup>	66 (52–80)	[18]
(2015–16)	174 <sup>a</sup>	73 (56–83)	Unpublished
<i>Non-European countries</i>			
Australia (2007–11)	520 <sup>a</sup>	75 (61–86)	Unpublished
Brazil (2010)	99	81 (71–95)	[8]
Canada (2006–09)	157 <sup>a</sup>	66 (46–78)	[16]
USA (2000–08)	442 <sup>a</sup>	91 (72–96)	[19]

95% CI, 95% confidence interval.

<sup>a</sup> Isolates also characterized by antigen genotype.

available [25]; the *nadA* gene was PCR-amplified to determine gene presence/absence, but was sequenced only in 5% of cases. Alleles and corresponding peptide identification numbers (IDs; protein variants) were assigned using the PubMLST *Neisseria* multilocus sequence typing database (<https://pubmlst.org/neisseria/>).

Antigen-specific predicted strain coverage by gMATS was defined by identifying peptide IDs significantly associated with MATS coverage/non-coverage for that antigen. For fHbp and NHBA antigens, peptide IDs present in more than five isolates were considered. Peptide IDs for which the percentage of MATS-covered

strains was higher than 60% or lower than 40% were considered predictors of coverage or non-coverage, respectively, if a test of proportions rejected 50% as null hypothesis ( $p < 0.05$  or  $< 0.001$ ). Peptide IDs not fulfilling these criteria were considered 'unpredictable'. The same approach was attempted for NadA, testing the association of *nadA* gene presence/absence and NadA-MATS coverage. Coverage for the OMV vaccine component PorA was genetically defined, as in MATS.

We defined a strain as gMATS 'covered' if one or more antigen-specific gMATS prediction for that strain was 'covered'. If all antigen-specific gMATS predictors were 'not covered', we defined the strain as gMATS 'not covered'. In remaining cases, the strain was defined as gMATS 'unpredictable'.

#### 2.4. gMATS coverage predictions on national strain panels

Overall, 49% of gMATS 'unpredictable' strains were MATS covered. This observation suggests that empirically we could predict that half of the 'unpredictable' strains could be considered as covered even by the gMATS method. Therefore, when we applied the gMATS to estimate the coverage on national strain panels, we used the proportion of 'covered' strains plus half the proportion of 'unpredictable' strains as gMATS final estimation of strain panel coverage. We defined the lower limit of the estimate range as the proportion 'covered', the upper limit as the sum of the proportions 'covered' and 'unpredictable'.

**Table 2**

The genetic Meningococcal Antigen Typing System (gMATS) predictors identified via analysis of associations between antigen genotype and MATS coverage.

4CMenB antigen	gMATS predictors		
	Covered	Not covered	Unpredictable
fHbp	Peptides 1, 2, 4, 14, 15, 37, 89, 90, 110, 144, 224, 232, 245, 249, 252, 510	Peptide 213 and all variant 2 and 3 peptides	All other fHbp variant 1 peptides
NHBA	Peptides 1, 2, 3, 5, 10, 20, 21, 113, 243	Peptides 6, 13, 17, 18, 19, 24, 25, 30, 31, 43, 47, 58, 112, 114, 120, 122, 160, 187, 253	All other NHBA peptides
NadA	Never	Always	Not applicable
OMV	PorA VR2 = 4	PorA VR2 $\neq$ 4	Not applicable

fHbp, factor H binding protein; NHBA, Neisserial Heparin-Binding Antigen; NadA, *Neisseria* adhesin A; OMV, outer membrane vesicles; PorA VR2, porin A variant 2. Peptide numbers correspond to identification numbers in PubMLST *Neisseria* sequence typing database.

**Table 3**

Single antigen genotyping as predictor of 4CMenB strain coverage estimated using the Meningococcal Antigen Typing System (MATS) and combined genetic Meningococcal Antigen Typing System (gMATS). The distribution in  $2 \times 2$  contingency tables of positive/negative antigen genotyping predictions and MATS outcome was always significantly non-random (Fisher exact test  $p$ -value  $< 10^{-16}$ ).

4CMenB antigen (predictor)	Predicted MATS outcome	Prediction of MATS MenB strain coverage by genotyping (%)					
		Isolates that can be predicted	Accuracy	Positive predictive value	Negative predictive value	Sensitivity	Specificity
fHbp peptide	Coverage by fHbp	84.5	93.6	90.7	97.9	98.4	87.9
NHBA peptide	Coverage by NHBA	81.5	82.5	79.9	87.7	92.7	69.1
<i>nadA</i> gene presence	Coverage by NadA	100	98.4	Indet.	98.4	0	100
PorA VR2 match	Coverage by PorA	100	100	100	100	100	100
Combined gMATS	Coverage by any antigen	81.3	92.4	93.7	85.3	97.1	72.4

Indet., indeterminate.

fHbp, factor H binding protein; NHBA, Neisserial Heparin-Binding Antigen; NadA, *Neisseria* adhesin A; PorA VR2, porin A variant 2.

#### 2.5. Comparisons across MenB strain panels

MATS and gMATS predictions for the different strain panels were compared against each other and against hSBA data from England and Wales and effectiveness data from England. The hSBA data were derived from a representative panel of 40 MenB isolates collected in 2007–2008 using sera from infants after the fourth 4CMenB vaccine dose, as described previously [10]. VE data were estimated for vaccine-eligible infants with laboratory-confirmed invasive MenB disease diagnosed in 2015–2016 after two doses administered as part of the NIP [22].

#### 2.6. Statistical methods

Statistical analyses were performed using the *stats* and *Agreement* packages of R 3.3.1 [26,27]. Chi-squared statistical test for proportions was performed by testing the 50% proportion as null hypothesis. Fisher exact test for associations, Pearson correlation, and Poisson regression were performed using default parameters. Lin's coefficient of accuracy ( $C_a$ ) was calculated with error = "const", TDI\_a = 5, target = "random" parameters. MATS coverage 95% CIs were calculated as described previously [9].

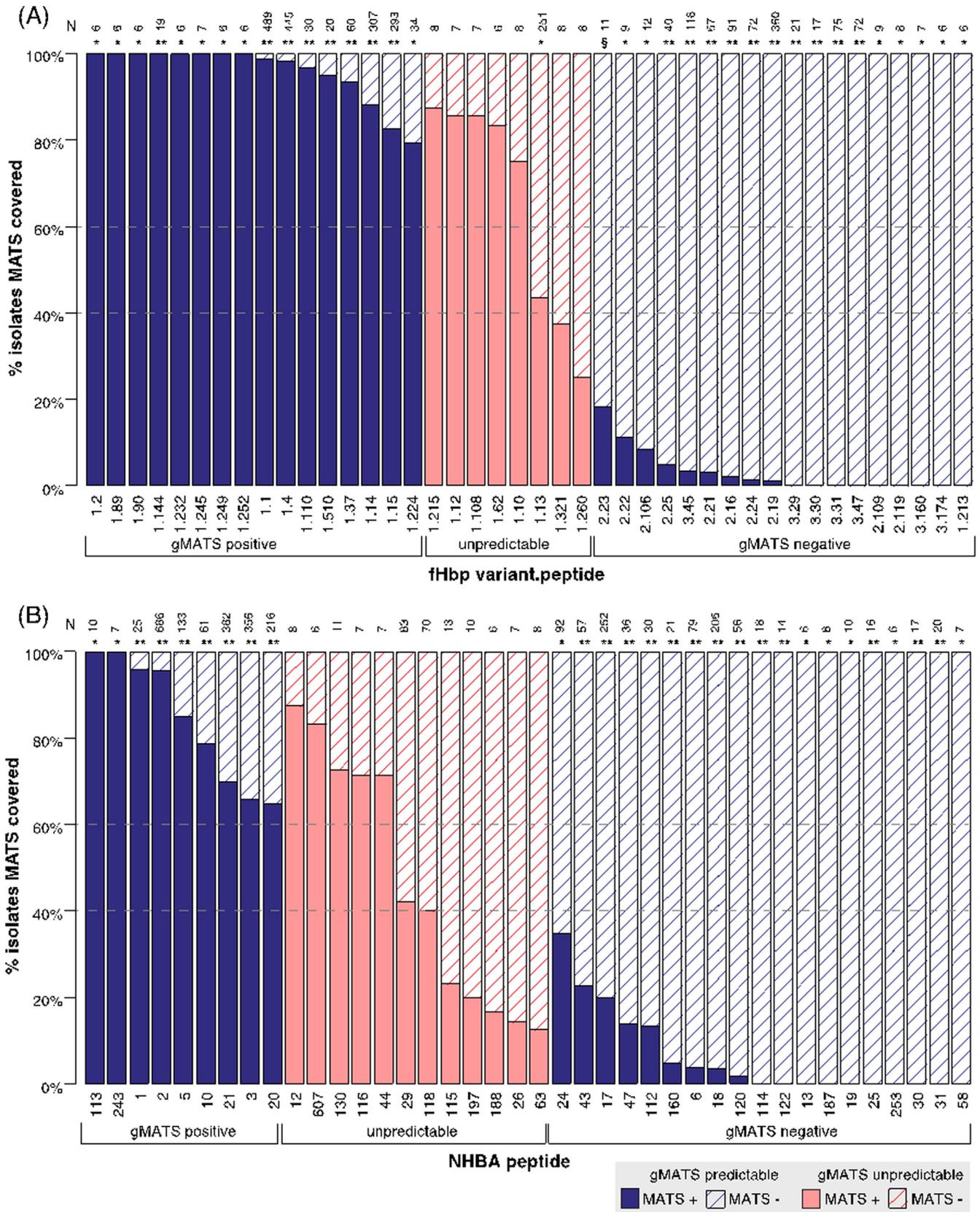
### 3. Results

#### 3.1. Individual 4CMenB antigen genotypes predict antigen-specific MATS coverage in 81–84% of strains analyzed

The association between antigen genotype and MATS coverage was analyzed to identify gMATS predictors. Table 2 reports the antigen-specific gMATS definitions and Table 3, their statistical properties as predictors of strain coverage. Since MATS coverage is already genetically defined for the OMV component, gMATS and MATS for OMV coincide. The NadA antigen was present in 27% of the strain panel, as demonstrated by *nadA* gene PCR, but it is known to be artificially under-expressed under MATS assay conditions [28]. Consequently, only 1.6% of strains ( $N = 57$ ) were NadA-MATS covered. Since the gMATS objective was to reproduce MATS results, the best predictor of NadA-MATS was the *a priori* 'not covered' prediction, independent of gene presence/absence or sequence.

#### 3.1.1. Specific fHbp peptides predict fHbp-MATS coverage for 84% of strain panel

In the strain panel investigated, 285 different fHbp peptides were identified, 42 of which were present in more than five isolates (Supplementary Table 1), representing 88% of isolates. Fig. 2A shows the percentage of strains covered or not covered by fHbp-MATS for each of the 42 fHbp peptides. Sixteen peptides,



**Fig. 2.** Proportion of the 3481 MenB isolates covered by 4CMenB, as estimated using the Meningococcal Antigen Typing System (MATS), stratified by fHbp (A) or NHBA (B) antigen genotypes. Genotypes for which MATS coverage can/cannot be predicted are depicted in blue/pink respectively. The plots show data for fHbp or NHBA peptides present in more than five isolates (peptide numbers are identification numbers in PubMLST *Neisseria* sequence typing database). *Footnotes:* \* p < 0.05; \*\* p < 0.001; † p = 0.07. fHbp, factor H binding protein; NHBA, Neisserial Heparin-Binding Antigen; gMATS, genetic Meningococcal Antigen Typing System.

all belonging to fHbp variant 1, showed coverage of at least 79% and were significantly ( $p < 0.05$ ) or highly significantly ( $p < 0.001$ ) associated with fHbp-MATS coverage. Seventeen peptides belonging to fHbp variants 2 and 3, and variant 1 peptide ID 213, had coverage of 18% or lower and were significantly associated with fHbp-MATS non-coverage ( $p = 0.07$  for ID 23; considered significant and indicative of non-coverage). Eight peptides, all belonging to variant 1, were neither positively nor negatively associated with fHbp-MATS coverage because of small sample size (frequency 6–8), intermediate coverage or both, and were considered ‘unpredictable’, along with peptide IDs with population frequency  $\leq 5$ . Fourteen frameshifted proteins (0.4% of isolates) were considered ‘not covered’. With these definitions, 84% of the strain panel had genetically predictable fHbp-MATS coverage, with 94% predictive accuracy (Table 3).

### 3.1.2. Specific NHBA peptides predict NHBA-MATS coverage for 81% of the strain panel

A total of 288 NHBA peptides were identified, 40 of which were present in more than five isolates (Supplementary Table 2). As shown in Fig. 2B, nine peptides showed coverage of at least 65% and were significantly ( $p < 0.05$ ) or highly significantly ( $p < 0.001$ ) associated with NHBA-MATS coverage. Nineteen peptides had coverage of 35% or lower and were associated with NHBA-MATS non-coverage. Twelve peptide IDs were neither positively nor negatively associated with NHBA-MATS coverage because of small sample size (frequency 6–13) or intermediate coverage. NHBA-gMATS for these and other peptide IDs with population frequency  $\leq 5$  were defined as ‘unpredictable’. Ten frameshifted proteins (0.3% of isolates) were considered ‘not covered’. Overall, 81% of the strain panel had genetically predictable NHBA-MATS coverage, with 82% predictive accuracy (Table 3).

### 3.2. gMATS predicts overall MATS strain coverage for 81% of the strains analyzed

When the four antigen-specific gMATS predictors were combined, 81% of strains in the global panel were predictable by gMATS (covered or not covered), with 92% prediction accuracy ver-

sus MATS (Table 3). Per national panel, 68–88% of isolates were gMATS predictable (Supplementary Table 3). Fig. 3 shows the comparison of MATS and gMATS predictions for gMATS-predictable subsets of each panel. The two methods generated highly correlated results (Pearson correlation coefficient 0.88,  $p < 10^{-5}$ ). The most discordant panels were from Ireland and Spain (10% over- and underestimation by gMATS, respectively). Across all countries, gMATS versus MATS root-mean-square deviation was 5%, indicating strong concordance. The regression analysis showed gMATS slightly exceeded MATS for coverage values  $> 80\%$ , although the 95% dispersion area of the regression model always included the identity, indicating an absence of statistically significant bias.

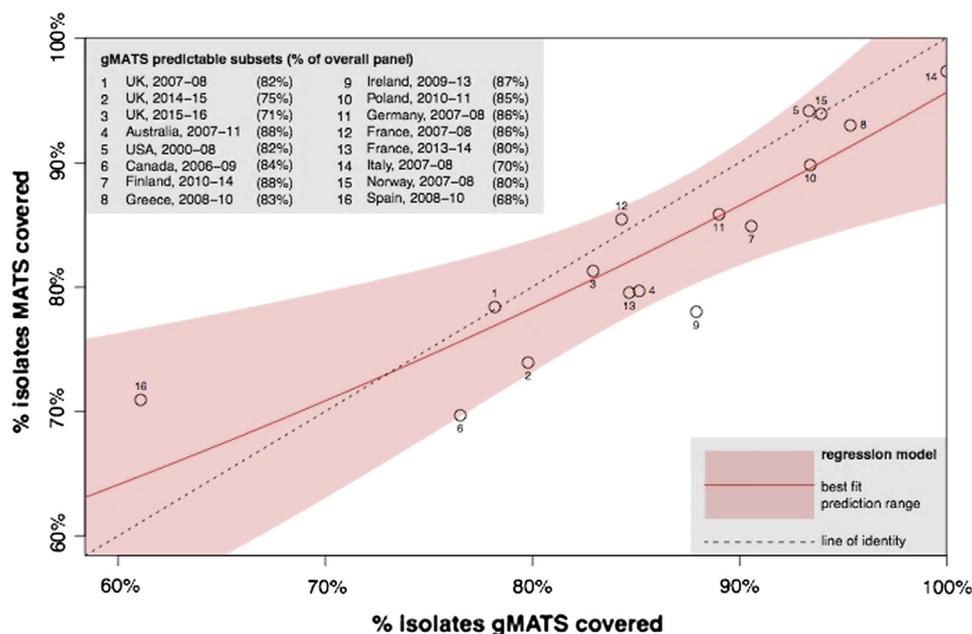
### 3.3. gMATS reproduces MATS strain coverage predictions on national panels

Fig. 4 shows the results of the empirical gMATS strain coverage method applied to the 16 national strain panels, along with corresponding MATS results (see also Supplementary Table 4). gMATS accurately reproduced MATS point estimates across panels (Lin’s accuracy coefficient,  $C_a$ , 0.98; root-mean-square deviation, 6%), with similar uncertainty ranges (average width 25% for MATS and 19% for gMATS) that largely overlapped.

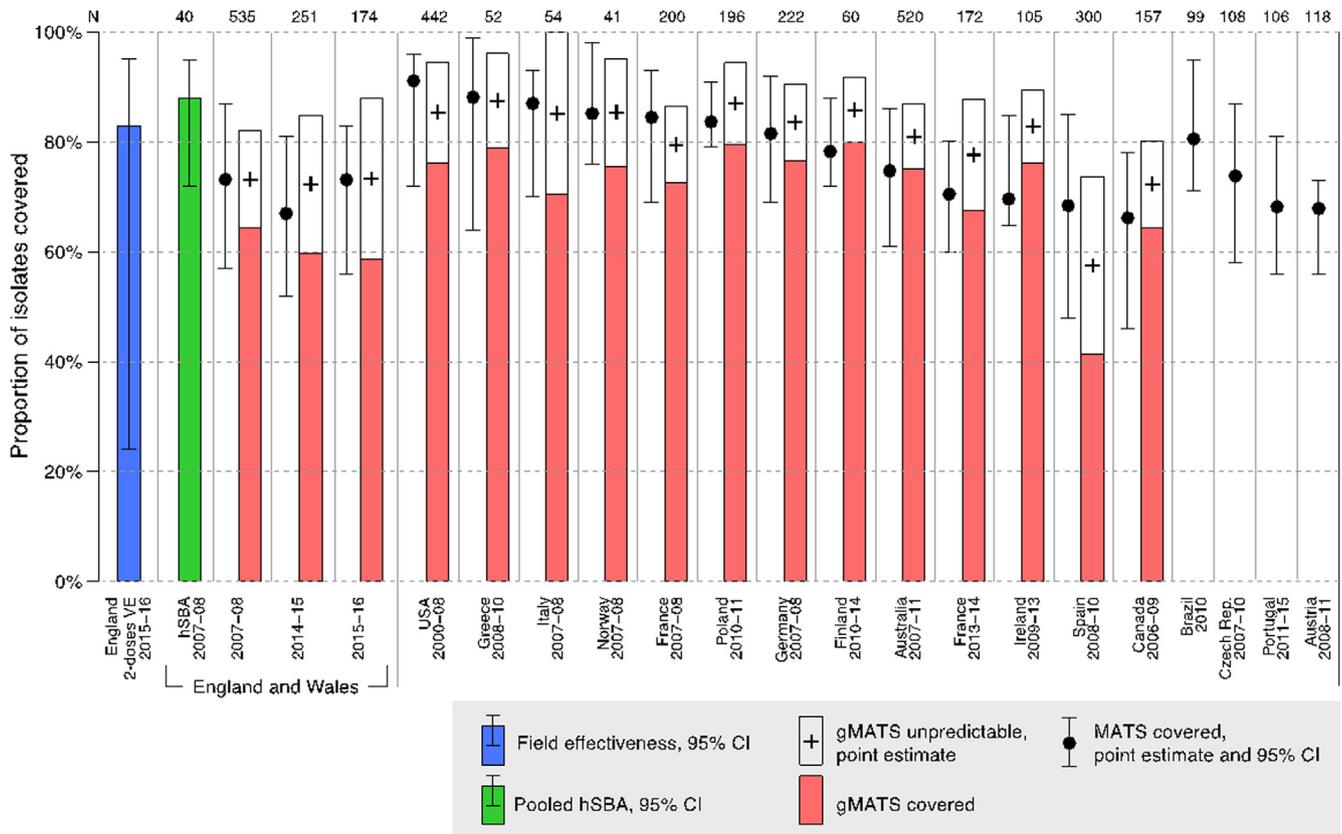
### 3.4. gMATS underestimates England and Wales pooled hSBA results and VE, and predicts similar or higher strain coverage in other countries

Within a dataset of 40 isolates representative of meningococcal disease in England and Wales [10], 100% of gMATS covered strains, 57% of gMATS negative strains, and 75% of gMATS unpredictable strains were killed in hSBA (Table 4), indicating that gMATS substantially underestimates killing in hSBA from infant pooled sera.

Fig. 4 also shows that, as with MATS estimates (66–73%), gMATS (72–73%) underestimated both the hSBA estimate of strain coverage in England and Wales (88%, 95% CI: 72–95%) [10] and VE (83%, 95% CI: 24–95%) in England [22], although with overlapping ranges.



**Fig. 3.** Regression analysis of predicted coverage of 4CMenB by MATS and gMATS on subset of gMATS-predictable strains, in each national strain panel. Predicted gMATS coverage was determined by combining predictions for three vaccine antigens (fHbp, NHBA and PorA). *Footnotes:* MATS, Meningococcal Antigen Typing System; gMATS, genetic Meningococcal Antigen Typing System; fHbp, factor H bonding protein; NHBA, Neisserial Heparin-Binding Antigen; NadA, *Neisseria* adhesin A; PorA, Porin A.



**Fig. 4.** Predicted strain coverage of 4CMenB by MATS and gMATS in England and Wales and 16 other countries. The blue bar represents the England 4CMenB vaccine effectiveness after two doses, 83% (95% CI, 24–95%) [22]. The green bar represents the hSBA coverage estimate 88% (95% CI, 72–95%) for a panel of 40 isolates tested with pooled sera from infant and adolescent vaccines and representative of the England and Wales meningococcal epidemiology [10]. The red and white portions of histograms represent respectively the proportion of gMATS ‘covered’ and ‘unpredictable’ strains for all country-specific panels analyzed. Point estimates of gMATS coverage (‘covered’ + 50% ‘unpredictable’ proportions) are shown as crosses. The proportion of MATS covered strains (point estimates with 95% confidence intervals [CI]) are shown as black circles for all country-specific strain panels analyzed. The panels of 4 countries (Brazil, Czech Republic, Poland and Austria) were not characterized by genotyping of antigens and gMATS prediction was not applicable. *Footnotes:* MATS, Meningococcal Antigen Typing System; gMATS, genetic Meningococcal Antigen Typing System; hSBA, human complement serum bactericidal assay; VE, vaccine effectiveness.

**Table 4**

Comparison of bacterial killing in the human complement serum bactericidal assay (hSBA) with gMATS coverage prediction on 40 representative isolates from England and Wales [10]. gMATS predicts hSBA with 88% accuracy for 80% of isolates for which a prediction can be made. Positive and negative predictive values are, respectively, 100% and 43%.

gMATS coverage prediction	Infant pooled hSBA result	
	Killed	Not killed
Positive	25	0
Negative	4	3
Unpredictable	6	2

gMATS, genetic Meningococcal Antigen Typing System.

Comparison of gMATS and MATS coverage estimates from England and Wales with results from other countries showed percentages ranged from 58% in Spain to 88% in Greece for gMATS, and from 66% in Canada to 91% in USA for MATS (Fig. 4). 4CMenB strain coverage in England and Wales, estimated by gMATS or MATS, is therefore at the lower end of the spectrum of estimates from the countries investigated so far.

#### 4. Discussion

A correlation between MATS and antigen genotype was initially noted in a previous assessment of predicted strain coverage of 4CMenB [12]. Here, we extended the analysis to a global panel of

over 3900 isolates from MenB disease cases to establish the level of correlation between MATS and antigen genotyping and define a new genetic predictor of 4CMenB strain coverage, gMATS.

We found that gMATS could replace MATS in predicting 4CMenB strain coverage in more than 80% of isolates, with 92% accuracy and 0.98 concordance. The least concordant strain sets were from Ireland, where gMATS overestimated MATS by 10% and Spain, where gMATS underestimated MATS by 10%. The reason of this underestimation in Ireland is mainly related to NHBA MATS relative potencies close to, but below, the positive bactericidal threshold for peptides 20 and 21, that are peptides considered ‘covered’ by gMATS. In Spain, the reason is still related to NHBA MATS relative potencies close to, but above, the positive bactericidal threshold for peptide 17, which is ‘not covered’ by gMATS. Also worth noting, Spain had the largest proportion of ‘unpredictable’ strains (32%) due to the specific strain epidemiology in this country [12]. Comparison with hSBA results from England and Wales [10] and VE data from England [22] confirmed that MATS and gMATS are conservative predictors of strain coverage. MATS and gMATS predicted equivalent or higher strain coverage in another 16 countries, highlighting the potential impact of 4CMenB outside of the UK.

A major difference between gMATS and MATS is the ‘unpredictable’ strains category in gMATS, allowing for two uses of the method. When testing a single strain, the gMATS result is either clear (‘covered’/‘not-covered’) or clearly absent, in which case

another method must be used to determine strain coverage, such as MATS or hSBA. When estimating strain coverage at a country level, we considered the proportion of ‘unpredictable’ strains as the uncertainty of the genetic predictor. Supported by the observation that, globally, 49% of unpredictable strains by gMATS were covered by MATS, we defined the gMATS point estimate for national strain panels as the proportion of gMATS ‘covered’ plus half the ‘unpredictable’ proportion. This practical, though simplistic, use of gMATS reproduced MATS predictions at a national level for the 17 countries investigated (accuracy coefficient,  $C_a$ , 0.98; root-mean-square deviation, 6%).

Genotype-phenotype modelling had already shown some promise in analyses of two subsets of the strain panels investigated here, as reported by Brehony and Mowlaboccus [29,30] and summarized in [Supplementary Table 5](#). With a larger sample size and broader geographic representation, overall, gMATS confirms and significantly extends the associations that were identified previously.

Compared to immunoassay-based methods, gMATS has the distinct advantage of not requiring a bacterial isolate and is therefore not limited to culture-positive cases only. gMATS can be performed using antigen PCR or methods that allow whole genome sequencing of *N. meningitidis* directly from clinical specimens [31]. In the England study of 4CMenB effectiveness, 70% of MenB cases in vaccine-eligible infants were confirmed by PCR only [22] indicating that gMATS, although currently applicable to only 80% of isolates, has the potential to duplicate MATS’s basis for strain coverage assessment and accelerate surveillance of 4CMenB VE. Also, gMATS can substantially simplify laboratory operations as technical implementation of MATS requires sufficient numbers of samples to be processed to maintain laboratory proficiency and remain cost-effective [32].

Current implementation of gMATS has limitations. Like MATS, gMATS (i) provides a conservative estimate of killing in the hSBA because it does not consider cooperative effects among antigens that can mediate killing together but not individually [33,34], (ii) ignores the effects of minor constituents of OMV [6], and (iii) underestimates the contribution of NadA antigen to coverage [28]. gMATS also has specific limitations connected with the genotype-phenotype association approach: (iv) low-frequency antigenic variants, including new variants that may appear in the future, cannot be assessed until enough isolates are collected, (v) future changes in regulatory elements or elsewhere in the genome – possibly promoted by immune selection and recombination [35] – may alter the genotype-phenotype association, and (vi) the threshold-based approach to define ‘covered’/‘not covered’ and the heuristic imputation of ‘unpredictable’ variants, albeit practical, is amenable to statistical improvements.

In contrast to immunoassay-based methods, gMATS is open to potentially unlimited improvement with the accumulation of prospective genotypic/phenotypic data, including hSBA or VE estimates from countries implementing 4CMenB. Cooperative effects among antigens could be detected by identifying di-, tri-, or tetra-antigenic profiles associated with killing in hSBA, or disappearing upon mass vaccination. As previously observed [29], the frequency of multi-antigen profiles drops dramatically with the number of antigens included, suggesting that efforts focused on the most frequent combinations, complementary to single-antigen predictors, could have higher chances of achieving statistical significance. Coverage mediated by minor OMV components could be defined using recently developed, OMV-specific typing schemes [36]. The NadA contribution could be completely reassessed using hSBA or VE estimates for NadA-specific isolates. Accumulation of phenotypic data on low-frequency variants will progressively reduce the fraction of ‘unpredictable’ isolates, and the introduction of typing schemes for regulatory elements

[37,38] could clarify the peptide IDs whose phenotype is highly affected by variability in expression levels. Finally, a probabilistic error model applied to the joint probabilities of multiple antigens, both for predictable and unpredictable isolates, would likely further improve the accuracy and robustness of the method. As previously proposed [39], we envision a community effort – possibly supported by a centralized resource for genotypic and phenotypic data [40] – whereby accumulation of prospective data from 4CMenB implementation and surveillance programs worldwide fuels continuous improvements of gMATS, simplifying 4CMenB strain coverage prediction and accelerating surveillance of 4CMenB vaccine protection.

In conclusion, gMATS, a new approach based on antigen genotyping, can accurately complement MATS in predicting 4CMenB strain coverage. Both methods underestimated field VE in England, while strain coverage predictions in 16 other countries matched or exceeded England and Wales coverage estimates. These results suggest that gMATS can become a suitable alternative to MATS for predicting vaccine strain coverage and monitoring vaccine implementation, and point to a positive potential impact of 4CMenB on IMD globally.

#### Trademark statement

*Bexsero* is a trademark owned by the GSK group of companies.

#### Funding

This work was supported by GlaxoSmithKline Biologicals SA, including all costs associated with the development and publishing of the manuscript.

#### Acknowledgements

The authors wish to thank Professor Ulrich Vogel from the University of Würzburg, Germany, for providing access to data on German isolates collected during the period 2007–2008, Dr Raymond Tsang for providing access to data on Canadian isolates collected during the period 2006–2009, and Dr Davide Serruto for critical reading of the manuscript.

Joanne Knowles (independent medical writer c/o GSK) provided writing assistance, Maria Ana de la Grandière (XPE Pharma & Science c/o GSK) provided publication coordination and editorial support, and AMICULUM (c/o GSK) provided support for the development of the associated video.

#### Conflict of interest

MC was and AM, AB, LS, MB, MN, KV, PW, and DM are employees of the GSK groups of companies. MN, PW, RR, and KV hold shares in the GSK group of companies as part of their employee remuneration. RA reports grants from Novartis. APL reports personal fees from Pfizer and Sanofi-Pasteur. HN is the secretary of the Finnish NITAG making recommendations on the use/purchase of vaccines, including MenB, to the Ministry of Health, Finland. HN’s institution received a grant from the GSK group of companies. GR’s institution has a Cooperative Research and Development Agreement (CRADA) with Novartis Vaccine Development and the GSK groups of companies. AS reports grant from the Polish National Science Centre and Novartis, and participated in the National Program for Antibiotic Protection (Ministry of Health) and the Mikrobank Program (Ministry of Science and Higher Education). AS reports personal fees from Baxter and the GSK group of companies, grant from Pfizer and Novartis and, personal fees and non-financial support from Pfizer. PS reports grants from

Novartis. MKT reports grants from Pfizer and the GSK groups of companies. MKT has a patent issued (630133). MT reports grants from the GSK groups of companies. GT reports research and travel grants from the GSK group of companies and Pfizer. JV reports personal fees and grants from the GSK group of companies, Pfizer and Sanofi-Pasteur. RB performs contract research on behalf of Public Health England for GSK, Sanofi Pasteur and Pfizer. DC, MCG, PK, CM, MJS, and RM have nothing to disclose.

### Authors' contributions

All authors had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. All authors reviewed and commented critically drafts of the manuscript for important intellectual content and gave final approval to submit for publication. The corresponding author had final responsibility to submit for publication. Drafts were developed by a professional publication writer according to the recommendations, documentation, and outline provided by the lead author. KV, RB, PW, DM, LS, AM, AB, MKT, JAV, MT, GT, RA, MC, and PS were involved in the conception or the design of the study. All authors (except PW and KV) participated in the collection or generation of the study data. HN, RB, DM, LS, AM, AB, MKT, JAV, AS, MT, GT, RA, CM, MB, MC, GR, APL, MCOG, and PS performed the study. PK, HN, RB, LS, AM, AB, MKT, JAV, AN, MT, RM, GT, RA, MJS, CM, MB, DC, and PS contributed to the study with materials/-analysis tools. KV, HN, RB, PW, DM, LS, AM, AB, MKT, JAV, AS, MT, RM, GT, RA, CM, MB, MC, GR, and PS were involved in the analyses or interpretation of the data.

### Appendix A. Supplementary material

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.vaccine.2018.12.061>.

### References

- [1] Stephens DS, Greenwood B, Brandtzaeg P. Epidemic meningitis, meningococcaemia, and *Neisseria meningitidis*. *Lancet* 2007;369:2196–210.
- [2] Banzhoff A. Multicomponent meningococcal B vaccination (4CMenB) of adolescents and college students in the United States. *Ther Adv Vaccines* 2017;5:3–14.
- [3] Findlow J. Vaccines for the prevention of meningococcal capsular group B disease: what have we recently learned? *Hum Vaccin Immunother* 2016;12:235–8.
- [4] Serruto D, Bottomley MJ, Ram S, Giuliani MM, Rappuoli R. The new multicomponent vaccine against meningococcal serogroup B, 4CMenB: immunological, functional and structural characterization of the antigens. *Vaccine* 2012;30(Suppl 2):B87–97.
- [5] Borrow R, Carlone GM, Rosenstein N, Blake M, Feavers I, Martin D, et al. *Neisseria meningitidis* group B correlates of protection and assay standardization—international meeting report Emory University, Atlanta, Georgia, United States, 16–17 March 2005. *Vaccine* 2006;24:5093–107.
- [6] Donnelly J, Medini D, Boccadifuoco G, Biolchi A, Ward J, Frasch C, et al. Qualitative and quantitative assessment of meningococcal antigens to evaluate the potential strain coverage of protein-based vaccines. *Proc Natl Acad Sci USA* 2010;107:19490–5.
- [7] Findlow J, Borrow R. Does post-implementation vaccine effectiveness data support pre-implementation predictions of 4CMenB utility? *Pathog Dis* 2017;75:ftx025.
- [8] Medini D, Stella M, Wassil J. MATS: Global coverage estimates for 4CMenB, a novel multicomponent meningococcal B vaccine. *Vaccine* 2015;33:2629–36.
- [9] Plikaytis BD, Stella M, Boccadifuoco G, DeTora LM, Agnusdei M, Santini L, et al. Interlaboratory standardization of the sandwich enzyme-linked immunosorbent assay designed for MATS, a rapid, reproducible method for estimating the strain coverage of investigational vaccines. *Clin Vaccine Immunol* 2012;19:1609–17.
- [10] Frosi G, Biolchi A, Lo Sapio M, Rigat F, Gilchrist S, Lucidarme J, et al. Bactericidal antibody against a representative epidemiological meningococcal serogroup B panel confirms that MATS underestimates 4CMenB vaccine strain coverage. *Vaccine* 2013;31:4968–74.
- [11] Budroni S, Kleinschmidt A, Boucher P, Medini D. Pooled-sera hSBA titres predict individual seroprotection in infants and toddlers vaccinated with 4CMenB. *Vaccine* 2016;34:2579–84.
- [12] Vogel U, Taha MK, Vazquez JA, Findlow J, Claus H, Stefanelli P, et al. Predicted strain coverage of a meningococcal multicomponent vaccine (4CMenB) in Europe: a qualitative and quantitative assessment. *Lancet Infect Dis* 2013;13:416–25.
- [13] Tzanakaki G, Hong E, Kesanopoulos K, Xirogianni A, Bambini S, Orlandi L, et al. Diversity of Greek meningococcal serogroup B isolates and estimated coverage of the 4CMenB meningococcal vaccine. *BMC Microbiol* 2014;14:111.
- [14] Waško I, Hong E, De Paola R, Stella M, Moschioni M, Taha MK, et al. High predicted strain coverage by the multicomponent meningococcal serogroup B vaccine (4CMenB) in Poland. *Vaccine* 2016;34:510–5.
- [15] Simões MJ, Bettencourt C, De Paola R, Giuliani M, Pizzi M, Moschioni M, et al. Predicted strain coverage of a meningococcal multicomponent vaccine (4CMenB) in Portugal. *PLoS One* 2017;12:e0176177.
- [16] Bettinger JA, Scheifele DW, Halperin SA, Vaudry W, Findlow J, Borrow R, et al. Diversity of Canadian meningococcal serogroup B isolates and estimated coverage by an investigational meningococcal serogroup B vaccine (4CMenB). *Vaccine* 2013;32:124–30.
- [17] Abad R, Medina V, Stella M, Boccadifuoco G, Comanducci M, Bambini S, et al. Predicted strain coverage of a new meningococcal multicomponent vaccine (4CMenB) in Spain: Analysis of the differences with other European countries. *PLoS One* 2016;11:e0150721.
- [18] Parikh SR, Newbold L, Slater S, Stella M, Moschioni M, Lucidarme J, et al. Meningococcal serogroup B strain coverage of the multicomponent 4CMenB vaccine with corresponding regional distribution and clinical characteristics in England, Wales, and Northern Ireland, 2007–08 and 2014–15: a qualitative and quantitative assessment. *Lancet Infect Dis* 2017;17:754–62.
- [19] Rajam G, Stella M, Kim E, Paulos S, Boccadifuoco G, Serino L, et al. Antigen Typing System (MATS)-based *Neisseria meningitidis* serogroup B coverage prediction for the MenB-4C vaccine in the United States. *mSphere* 2017;2:e00261–e317.
- [20] Toropainen M, Vainio A, Moschioni M, R DP, Stella M, Nohynek H, et al. Predicted strain coverage of four-component meningococcal serogroup B vaccine (4CMenB) in Finland, 2010–2014 [abstract 189]. Program and abstracts of the 20th international pathogenic *Neisseria* conference. Manchester, UK2016. p. 279.
- [21] Ladhani SN, Ramsay M, Borrow R, Riordan A, Watson JM, Pollard AJ. Enter B and W: two new meningococcal vaccine programmes launched. *Arch Dis Child* 2016;101:91–5.
- [22] Parikh SR, Andrews NJ, Beebejaun K, Campbell H, Ribeiro S, Ward C, et al. Effectiveness and impact of a reduced infant schedule of 4CMenB vaccine against group B meningococcal disease in England: a national observational cohort study. *Lancet* 2016;388:2775–82.
- [23] Clark SA, Lucidarme J, Newbold LS, Borrow R. Genotypic analysis of meningococcal factor h-binding protein from non-culture clinical specimens. *PLoS One* 2014;9:e89921.
- [24] Azzari C, Nieddu F, Moriondo M, Indolfi G, Canessa C, Ricci S, et al. Underestimation of invasive meningococcal disease in Italy. *Emerg Infect Dis* 2016;22:469–75.
- [25] Lucidarme J, Comanducci M, Findlow J, Gray SJ, Kaczmarek EB, Guiver M, et al. Characterization of *fHbp*, *nhba* (*gna2132*), *nadA*, *porA*, and sequence type in group B meningococcal case isolates collected in England and Wales during January 2008 and potential coverage of an investigational group B meningococcal vaccine. *Clin Vaccine Immunol* 2010;17:919–29.
- [26] R Core, Team. R: A language and environment for statistical computing. Vienna (Austria): R Foundation for Statistical Computing; 2013.
- [27] Yu Y, Lin L. Agreement: Statistical tools for measuring agreement. R package version 08-1; 2012.
- [28] Fagnocchi L, Biolchi A, Ferlicca F, Boccadifuoco G, Brunelli B, Brier S, et al. Transcriptional regulation of the *nadA* gene in *Neisseria meningitidis* impacts the prediction of coverage of a multicomponent meningococcal serogroup B vaccine. *Infect Immun* 2013;81:560–9.
- [29] Brehony C, Rodrigues CMC, Borrow R, Smith A, Cunney R, Moxon ER, et al. Distribution of Bexsero® Antigen Sequence Types (BASTs) in invasive meningococcal disease isolates: Implications for immunisation. *Vaccine* 2016;34:4690–7.
- [30] Mowlaboccus S, Perkins TT, Smith H, Sloots T, Tozer S, Premph LJ, et al. Temporal changes in BEXSERO® antigen sequence type associated with genetic lineages of *Neisseria meningitidis* over a 15-year period in Western Australia. *PLoS One* 2016;11:e0158315.
- [31] Clark SA, Doyle R, Lucidarme J, Borrow R, Breuer J. Targeted DNA enrichment and whole genome sequencing of *Neisseria meningitidis* directly from clinical specimens. *Int J Med Microbiol* 2018;308:256–62.
- [32] Vogel U, Stefanelli P, Vazquez J, Taha MK, Claus H, Donnelly J. The use of vaccine antigen characterization, for example by MATS, to guide the introduction of meningococcus B vaccines. *Vaccine* 2012;30(Suppl 2):B73–7.
- [33] Giuliani M, Bartolini E, Galli B, Santini L, Lo Surdo P, Buricchi F, et al. Human protective response induced by meningococcus B vaccine is mediated by the synergy of multiple bactericidal epitopes. *Sci Rep* 2018;8:3700.
- [34] Partridge E, Lujan E, Giuntini S, Vu DM, Granoff DM. The role of anti-NHba antibody in bactericidal activity elicited by the meningococcal serogroup B vaccine, MenB-4C. *Vaccine* 2017;35:4236–44.
- [35] Bai X, Borrow R. Genetic shifts of *Neisseria meningitidis* serogroup B antigens and the quest for a broadly cross-protective vaccine. *Expert Rev Vaccines* 2010;9:1203–17.
- [36] Rodrigues C, Chan H, Vipond C, Maiden M. Development of Bexsero® outer membrane vesicle typing system for analysis of meningococcal whole genome

- sequences [abstract O2.05]. Book of abstracts for the 14th Congress of the EMGM. Prague (Czech Republic): European Meningococcal and Haemophilus Disease Society; 2017. p. 17.
- [37] Biagini M, Spinsanti M, De Angelis G, Tomei S, Ferlenghi I, Scarselli M, et al. Expression of factor H binding protein in meningococcal strains can vary at least 15-fold and is genetically determined. *Proc Natl Acad Sci U S A* 2016;113:2714–9.
- [38] Cayrou C, Akinduko AA, Mirkes EM, Lucidarme J, Clark SA, Green LR, et al. Clustered intergenic region sequences as predictors of factor H Binding Protein expression patterns and for assessing *Neisseria meningitidis* strain coverage by meningococcal vaccines. *PLoS One* 2018;13:e0197186.
- [39] Snape MD, Medini D, Halperin SA, DeTora L, Drori J, Moxon ER. The challenge of post-implementation surveillance for novel meningococcal vaccines. *Vaccine* 2012;30(Suppl 2):B67–72.
- [40] Jolley KA, Maiden MC. BIGSdb: Scalable analysis of bacterial genome variation at the population level. *BMC Bioinf* 2010;11:595.