

# Fighting Females: Neural and Behavioral Consequences of Social Defeat Stress in Female Mice

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## ABSTRACT

**BACKGROUND:** Despite the twofold higher prevalence of major depressive and posttraumatic stress disorders in women compared with men, most clinical and preclinical studies have focused on male subjects. We used an ethological murine model to study several cardinal symptoms of affective disorders in the female targets of female aggression.

**METHODS:** Intact Swiss Webster (CFW) female resident mice were housed with castrated male mice and tested for aggression toward female intruders. For 10 days, aggressive CFW female residents defeated C57BL/6J (B6) female intruders during 5-minute encounters. Measures of corticosterone, c-Fos activation in hypothalamic and limbic structures, and species-typical behaviors were collected from defeated and control females. Ketamine (20 mg/kg) was tested for its potential to reverse stress-induced social deficits.

**RESULTS:** Housed with a castrated male mouse, most intact resident CFW females readily attacked unfamiliar B6 female intruders, inflicting >40 bites in a 5-minute encounter. Compared with controls, defeated B6 females exhibited elevated plasma corticosterone and increased c-Fos activation in the medial amygdala, ventral lateral septum, ventromedial hypothalamus, and hypothalamic paraventricular nucleus. Chronically defeated females also showed vigilance-like behavior and deficits in social interactions, novel object investigation, and nesting. The duration of social interactions increased 24 hours after chronically defeated female mice received a systemic dose of ketamine.

**CONCLUSIONS:** These findings demonstrate that CFW female mice living with male conspecifics can be used as aggressive residents in an ethological model of female social defeat stress. These novel behavioral methods will encourage further studies of sex-specific neural, physiological, and behavioral adaptations to chronic stress and the biological bases for interfemale aggression.

**Keywords:** Affective disorders, Aggression, Female, Ketamine, Social defeat stress, Social interaction

<https://doi.org/10.1016/j.biopsych.2019.05.005>

The prevalence of affective disorders, including major depressive disorder and posttraumatic stress disorder, is twofold higher in women than in men, and sex can shape the trajectory of these disorders in terms of their onset, duration, and rate of recurrence (1–4). Despite substantial evidence for sex differences in the development, course, and biological underpinnings of affective disorders (5,6), most clinical and preclinical studies focus on male subjects. In preclinical research, the murine chronic social defeat stress protocol employs species-typical male aggression toward submissive conspecifics to induce several cardinal symptoms of stress-related affective disorders in a subset of defeated male mice (7–14). This protocol is highly reproducible between and within laboratories, suggesting that the translational effects of chronic social stress are both robust and reliable. In addition, long-term treatment with tricyclic antidepressants or serotonin reuptake inhibitors or a single ketamine administration can

normalize defeat-associated social deficits and stress-induced molecular adaptations in male mice (9,13,15–18), making this protocol a valuable tool for investigating novel drugs and their mechanisms of action.

In developing a female model of social defeat stress that parallels the existing male protocol, substantial efforts have been made to foster female-directed male aggression by using male odorants transferred onto female mice (19) and through chemogenetic stimulation of the male ventromedial hypothalamus (VMH) (20). A vicarious social defeat stress model in which female mice witness intermale aggression (21) has also been employed. To provide an alternative method that closely mirrors the male chronic social defeat protocol and eliminates the need to generate atypical patterns of male aggression, we identify specific conditions that promote aggression in outbred female mice toward female opponents.

SEE COMMENTARY ON PAGE e31; SEE ALSO VIDEO CONTENT ONLINE

Whereas male mice are aggressive under diverse experimental conditions (22), female aggression is most often studied either during pregnancy [(23–27), but see (28)] or during the first postpartum week while neonatal pups are suckling (29–31). As an adaptive defensive behavior, dams will engage in maternal aggression to prevent postpartum fertilization and to protect their offspring against unfamiliar male or female intruders (29,31–33). Outside of these brief gestational and postpartum windows, female mice are minimally aggressive when subjected to isolation housing (34), a technique often used to induce territorial aggression in outbred male mice (35,36), in female California mice (*Peromyscus californicus*) (37), and in female Syrian golden hamsters (*Mesocricetus auratus*) (38,39). In contrast, we report that most intact female mice (*Mus musculus*) housed with an intact or castrated male mouse will engage in intense aggression when confronted by a female opponent; although isolation-induced territorial competition may not promote interfemale agonistic behavior, a significant subset of female mice will readily fight a rival female intruder, possibly in competition for an available mate.

Upon identifying specific conditions to engender interfemale aggression, studies were conducted to 1) generate an ethological model of female chronic social defeat stress; 2) examine the effects of social defeat stress on plasma corticosterone concentrations; 3) determine if social defeat stress increases c-Fos activation in brain areas, including the medial amygdala (MeA), lateral septum, VMH, and paraventricular nucleus (PVN); 4) characterize defeat-associated deficits in species-typical social and nonsocial behaviors; and 5) increase the duration of social interactions initiated by chronically defeated female mice with acutely administered ketamine.

## METHODS AND MATERIALS

See Supplement for additional details.

### Animals

Twelve-week-old intact ( $n = 74$ ) or ovariectomized ( $n = 27$ ) Swiss Webster (CFW) female mice (Charles River Laboratories, Wilmington, MA) were housed in resident pairs with age-matched intact ( $n = 47$ ) or castrated ( $n = 61$ ) CFW male mice in clear polycarbonate cages ( $18.9 \times 29.7 \times 12.8$  cm) lined with pine shavings. Twelve-week-old intact intruder C57BL/6J (B6) ( $n = 190$ ) (Jackson Laboratories, Bar Harbor, ME) or CFW female mice ( $n = 40$ ) were group-housed in cages ( $25.7 \times 48.3 \times 15.2$  cm;  $n = 10$ /cage) with corncob bedding. Experimental 12-week-old B6 females were housed individually and assigned to control ( $n = 23$ ), acute ( $n = 13$ ), or chronic social defeat ( $n = 28$ ) conditions. Animals were cared for according to the National Research Council *Guide for the Care and Use of Laboratory Animals*, and procedures were approved by the Tufts University Institutional Animal Care and Use Committee.

### Aggression in Outbred Ovariectomized Female Mice

Ovariectomized CFW female mice, housed in resident pairs with intact CFW males, were evaluated for aggression in modified resident-intruder confrontations every other day starting 2 weeks after pair-housing (36). To test the effect of intruder strain and familiarity, male mice were removed and

female intruders were introduced to resident home cages for 2-minute resident-intruder confrontations (Supplemental Figure S2). Attack latency and attack bite frequency were recorded. Following confrontations, female intruders were removed and male mice were returned to their resident home cages.

### Aggression in Intact Outbred Female Mice

Intact female CFW residents, housed with intact male CFWs, were evaluated for aggression toward unfamiliar B6 female intruders. CFW litters were culled on postnatal day 1, at which time CFW female residents were housed singly. After the first postpartum week, there was a substantial decrease in inter-female aggression. To address whether this reduction was related to isolation housing, female residents were housed with castrated male mice, and aggressive confrontations continued 3 days later. After 5 days of living with castrated male mice, most female residents attacked unfamiliar female B6 intruders. Male cohabitation-induced interfemale aggression is referred to as rival aggression to distinguish it from maternal or gestational aggression. In subsequent groups, intact nulliparous CFW female mice were housed exclusively with castrated male CFWs in resident pairs. A significant subset of these female residents were highly aggressive toward unfamiliar female B6 intruders ( $n = 39$  of 61;  $>15$  bites/2-min confrontation); these residents were used as aggressors during the chronic social defeat protocol.

### Sex-Specific Patterns of Aggression

Ten 5-minute resident-intruder confrontations between intact CFW female residents and unfamiliar B6 female intruders were videotaped in the resident home cage for detailed behavioral analyses (Supplemental Video S1). Similar archival videos of intermale confrontations were analyzed to compare the behavioral composition of aggressive encounters in male versus female mice (Supplemental Video S2).

### Testing Aggressive Potential of Female Mice That Do Not Display Rival Aggression

To test whether pregnancy-induced aggression was distinct from rival aggression, consistently nonaggressive female residents that were pair-housed with castrated male mice ( $n = 7$ ) for at least 2 months were assessed for gestational aggression. These female mice were housed with intact male CFWs for 3 days, then returned to their original castrated male partners. Aggression was evaluated every 2 or 4 days during 2-minute resident-intruder confrontations. Pups were culled on postnatal day 1, and female residents were tested for aggression 4 and 7 days postpartum.

### 10-Day Female Chronic Social Defeat Stress

Highly aggressive female CFWs were used as resident stimulus animals for the chronic social defeat stress protocol. Two days before the initial defeat episode, resident CFW pairs composed of intact female and castrated male mice were transferred to large polycarbonate cages ( $25.7 \times 48.3 \times 15.2$  cm) divided in half by perforated, clear polycarbonate partitions [compare with (8,14)]. One day before defeats, female

## Female Aggression and Chronic Defeat Stress

residents were tested for aggression to ensure behavioral reliability under the new housing conditions.

Daily 5-minute defeat episodes occurred in the large divided cages (Supplemental Video S3); male mice were temporarily removed, and experimental female B6 intruders were exposed to unfamiliar aggressive female residents. Following defeats, female B6 mice were housed opposite the female CFW residents that defeated them, and male mice were returned to be pair-housed with CFW female mice. During this 24-hour threat period, cage dividers permitted sensory contact between CFW resident pairs and experimental B6 mice but protected experimental mice from attack. For 10 consecutive days, B6 intruders were defeated by and rehoused adjacent to unfamiliar, aggressive CFW residents. Nondefeated, control B6 females were housed opposite unfamiliar resident CFW pairs daily, but were never physically attacked. Acutely defeated B6 experimental mice were treated as controls until the 10th day, when they were defeated once. Control and defeated mice were weighed every other day and singly housed after defeats concluded on the 10th day. During the 10-day protocol, resident CFW pairs received approximately 15 g of fresh pine shavings every other day, and cages were cleaned on the 5th day following the defeat. Reliably aggressive female CFW residents (i.e., >40 bites/5-min confrontation or >15 bites/2-min confrontation) were used in chronic social defeat stress experiments for 6 to 12 months.

### Estrous Cycling

Vaginal cytology was monitored in experimental mice during the 10-day social defeat stress protocol using the lavage technique (40). Cyclicity was also evaluated in a subset of highly aggressive nulliparous resident CFWs ( $n = 23$ ) housed with castrated male mice to determine if aggression varied according to estrous cycle phase [(41–43), but see (44,45)].

### Corticosterone Measurements

At 2 time points (Supplemental Figure S1A), blood was collected from the submandibular vein of experimental female mice ( $n = 5$  control;  $n = 5$  acute defeat;  $n = 10$  chronic defeat) using sterile 4-mm lancets (Goldenrod Animal Lancet; Medipoint, Inc., Mineola, NY) and centrifuged at 4°C at 3000 rpm for 10 minutes. Plasma (15  $\mu$ L) was collected and stored at –80°C for corticosterone enzyme immunoassay (Arbor Assays, Ann Arbor, MI); standards (7.8125–1000 ng/mL) and samples were run in duplicate.

### Tissue Collection and c-Fos Immunohistochemistry

On the 10th day of chronic social defeat stress, brains were collected from female B6 experimental animals after no defeat or a 5-minute social defeat stress episode followed by an hour-long threat period (Supplemental Figure S1B). Brain slices (50- $\mu$ m) containing the anteromedial bed nucleus of the stria terminalis, ventral lateral septum (LSv), hypothalamic PVN (hPVN), periventricular nucleus, VMH, MeA, and dentate gyrus were selected for c-Fos immunohistochemistry.

### Open Field Social Interaction

After a 2.5-minute habituation period in the social interaction apparatus (84  $\times$  29  $\times$  36 cm) containing an empty wire mesh

stimulus cage (11-cm height, 10.5-cm diameter), control and defeated mice were briefly removed, and an unfamiliar, aggressive CFW female resident was placed in the stimulus cage. Experimental B6 mice were returned to the apparatus and evaluated for social interactions during a 2.5-minute test. Social interaction time was defined as the duration spent within a social interaction zone extending 2.25 cm past the radius of the stimulus cage. Social interaction videos (EthoVision XT version 14; Noldus Information Technology Inc., Leesburg, VA) were scored manually for vigilance-like behavior, defined as time spent oriented toward, but not interacting with, the stimulus animal [compare with (46)]. The open field and stimulus cage were cleaned and dried between mice.

### Novel Object Investigation

Control and defeated female mice were briefly moved to clean holding cages while 4 rubber stoppers (Size No. 5; Thermo Fisher Scientific, Waltham, MA) were placed in their home cages. Mice were returned to their home cages for a 5-minute test. Rectal temperatures were collected immediately before and after tests using a temperature probe (2100 Tele-Thermometer; YSI, Inc., Yellow Springs, OH) lubricated with mineral oil.

### Home Cage Social Interactions and Ketamine Administration

Social contact with a nonaggressive, group-housed B6 female stimulus animal was evaluated during 1.5-minute tests conducted in the home cage. Contact was defined as the duration of social investigation initiated by the experimental female mouse. Alternatively, home cage social interaction tests were conducted using anesthetized stimulus female mice (Supplemental Figure S1A). Rectal temperatures were collected from experimental female mice immediately before and after testing.

Chronically defeated and control female mice received intraperitoneal injections of 0.9% sodium chloride or the *N*-methyl-D-aspartate receptor antagonist ketamine hydrochloride diluted in 0.9% sodium chloride (20 mg/kg; Vedco Inc., St Joseph, MO) (17,21,47,48). Social interactions were evaluated 30 minutes, 24 hours, and 5 days after injection.

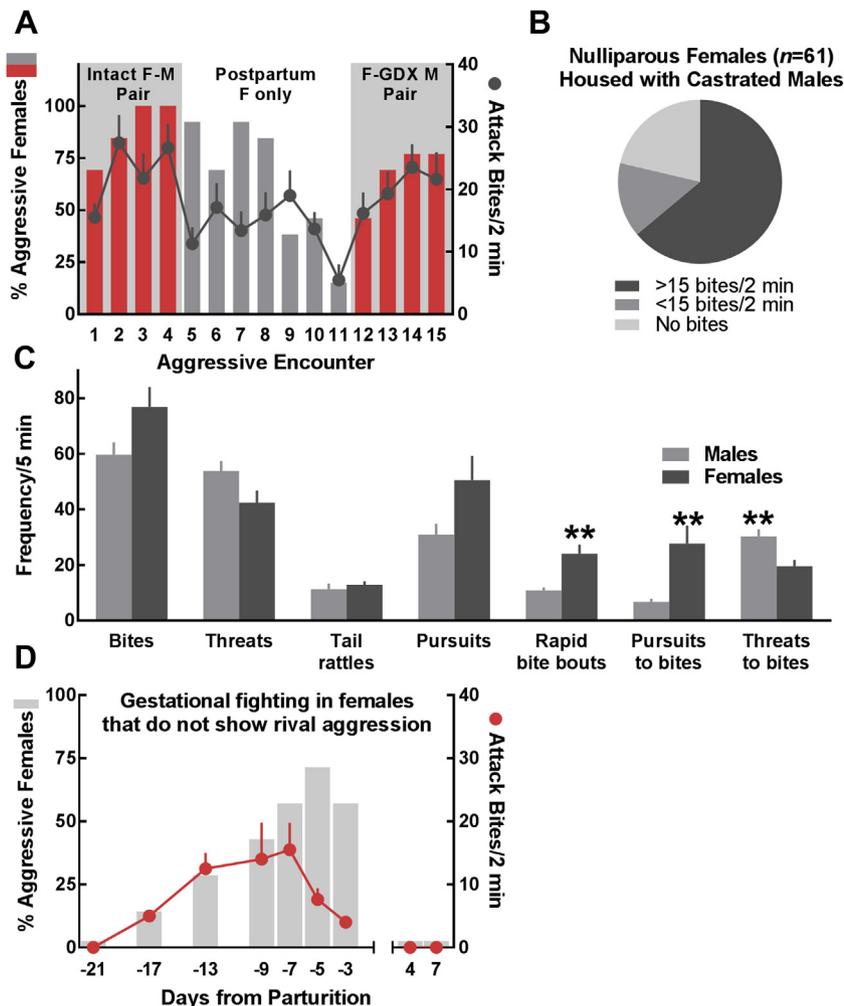
### Nesting

Female mice received 2 g of nesting material (Nestlets; Ancare Corp., Bellmore, NY) at 1330 hours in the home cage. Five days later, nests were scored on a scale of 1 to 5 (49), nest heights and diameters were recorded, and nest images were evaluated for shape (i.e., circularity) in ImageJ (National Institute of Mental Health, Bethesda, MD).

## RESULTS

### Interfemale Rival Aggression

Most intact female CFW residents were aggressive when housed with a male mouse, but not when housed in isolation (Figure 1A) or following ovariectomy (Supplemental Figure S2). By the third aggressive confrontation, >90% of female residents that were housed with intact male CFW mice expressed gestational aggression (mean  $\pm$  SEM attack bites = 26.62  $\pm$



**Figure 1.** Most (A) multiparous and (B) nulliparous outbred female mice living with intact or gonadectomized (GDX), i.e., castrated, male mice developed aggression toward unfamiliar C57BL/6J female intruders. (C) Male and female attack bite frequencies were comparable, but resident female mice ( $n = 10$ ) displayed more sequential bites (rapid bite bouts;  $t_{18} = 3.85$ ), and their bites were more frequently preceded by pursuits (pursuits to bites;  $t_{18} = 3.20$ ), whereas male mice ( $n = 10$ ) exhibited more bites preceded by sideways threats (threats to bites;  $t_{18} = 3.14$ ); data are shown as mean  $\pm$  SEM;  $**p < .01$  male vs. female. (D) Most nonaggressive nulliparous female mice displayed pregnancy-induced aggression that was time-locked with the gestational period. (A, D) Left axes denote the percentage of animals that were aggressive, and right axes depict attack bite frequencies as mean  $\pm$  SEM, calculated from female mice that fought. F, female; M, male.

3.83). However, by the 7th day after litters were culled and intact male mice were removed,  $<20\%$  of female residents were aggressive, and mice that did express aggression showed substantially reduced attack bite frequencies (mean  $\pm$  SEM attack bites =  $5.5 \pm 2.5$ ). Five days after being rehoused with castrated CFWs, most (75%) female residents fought (mean  $\pm$  SEM attack bites =  $21.6 \pm 4.34$ ). Similarly, most intact nulliparous CFW residents housed exclusively with castrated male mice fought, and a significant subset (65%) emerged as highly aggressive toward unfamiliar female B6 intruders (Figure 1B).

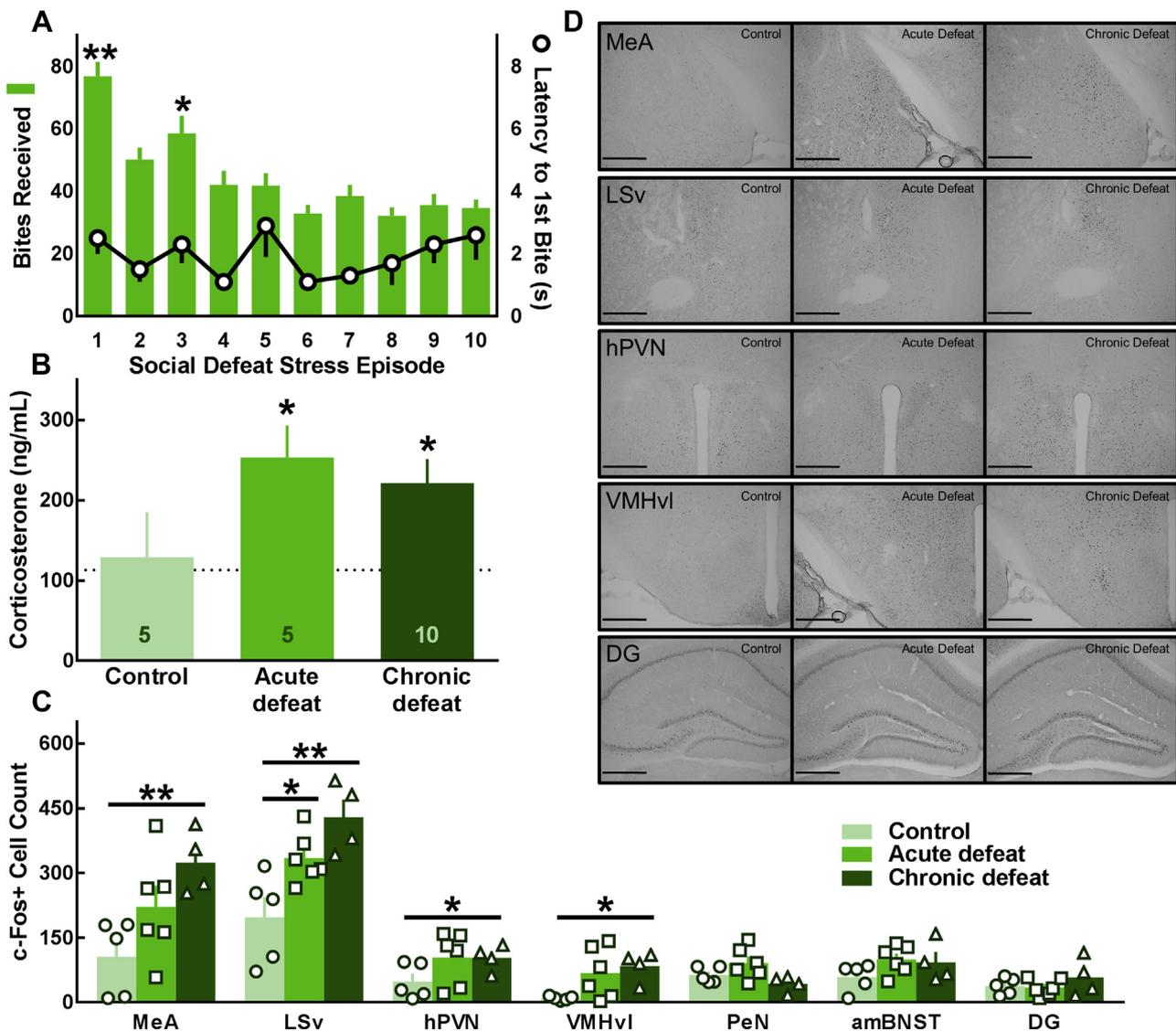
Attack bite frequencies were similar between resident female and male aggressors (Figure 1C). During confrontations with an unfamiliar intruder, female residents exhibited more rearing behavior (Supplemental Figure S4) (50) and rapid bouts of consecutive bites that were often preceded by pursuits (Figure 1C and Supplemental Video S1). In contrast, male attacks were often preceded by sideways threats (Supplemental Video S2). Estrous cycle phase was determined in highly aggressive female residents ( $n = 23$ ). Though an effect of phase on aggression was not apparent (Supplemental

Figure S3), this could be explained by a ceiling effect and low variability in attack bite frequencies among aggressive female mice.

A subset of nulliparous residents housed with castrated male mice was consistently nonaggressive toward female intruders ( $n = 13$  of 61). Half of these residents ( $n = 7$ ) were tested for their sensitivity to the pro-aggressive effects of pregnancy. By late pregnancy, 75% of formerly nonaggressive female mice attacked an unfamiliar female intruder; however, after pups were culled on postnatal day 1, these residents returned to their nonaggressive, prepregnancy baselines (Figure 1D).

### Neural and Physiological Effects of Chronic Female Social Defeat Stress

Acute or 10-day social defeat stress (Figure 2A and Supplemental Video S3) followed by a threat period increased circulating corticosterone more than the threat period alone (Figure 2B). Similar corticosterone concentrations in acutely and chronically defeated individuals suggest that B6 female intruders do not habituate to social defeat stress, much like acutely and repeatedly defeated outbred male intruders (51).



**Figure 2.** Female C57BL/6J mice were defeated by aggressive female CFW residents for 10 consecutive days. **(A)** Attack latencies were <5 seconds (right axis) and the greatest number of attacks were delivered early in the defeat protocol (left axis) ( $F_{9,190} = 13.03$ ,  $p < .0001$ );  $*p < .05$ ,  $**p < .0001$  compared with day 5. **(B)** Elevated concentrations of plasma corticosterone were detected after acute or chronic social defeat stress (time:  $F_{1,17} = 9.6$ ,  $p = .007$ ; defeat:  $F_{2,17} = 4.2$ ,  $p = .033$ );  $*p < .05$  compared with control mice. **(C)** Social defeat stress increased c-Fos activation in the medial amygdala (MeA) ( $F_{2,12} = 6.43$ ,  $p = .013$ ), ventral lateral septum (LSv) ( $F_{2,12} = 9.28$ ,  $p = .004$ ), hypothalamic paraventricular nucleus (hPVN) ( $F_{2,12} = 4.48$ ,  $p = .035$ ), and ventrolateral division of the ventromedial hypothalamus (VMHvl) ( $F_{2,12} = 4.17$ ,  $p = .042$ );  $*p < .05$ ,  $**p < .01$  compared with control mice. **(D)** Representative images of c-Fos in the MeA, LSv, hPVN, VMHvl, and dentate gyrus (DG). Scale bars = 200  $\mu\text{m}$ . Data are shown as mean  $\pm$  SEM. amBNST, anteromedial bed nucleus of the stria terminalis; PeN, periventricular nucleus.

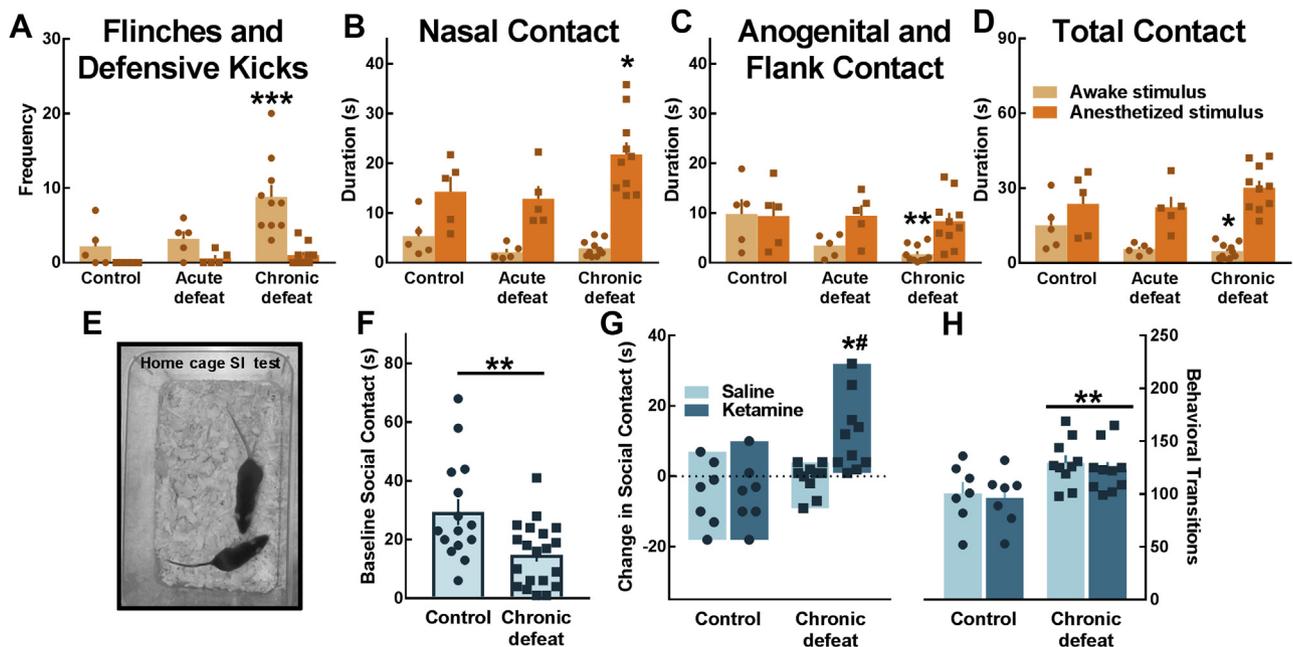
However, neither estrous cycle nor body weight was significantly affected by chronic social defeat (Supplemental Figure S5A–E).

Compared with the control condition, acute or chronic social defeat stress significantly increased c-Fos activation in the LSv, whereas only chronic social defeat significantly increased the number of c-Fos<sup>+</sup> cells in the MeA, hPVN, and ventrolateral division of the VMH (VMHvl) (Figure 2C, D). Exploratory correlational analyses revealed patterns of interregional c-Fos activation. Specifically, there was a positive relationship between c-Fos<sup>+</sup> cell counts in the MeA and anteromedial bed nucleus of the stria terminalis in control mice, whereas an

inverse correlation was observed in defeated mice (Supplemental Table S1). Female mice subjected to chronic social defeat also exhibited a unique pattern of inverse correlations in c-Fos in the periventricular nucleus and anteromedial bed nucleus of the stria terminalis or dentate gyrus.

### Behavioral Effects of Chronic Social Defeat Stress in Female Mice

During social interactions in the home cage (Figure 3E) with a nonaggressive female B6 stimulus animal, chronically defeated mice displayed a greater number of defensive kicks and



**Figure 3.** Chronically defeated female mice exhibited substantial social contact deficits, which were improved with ketamine, 24 hours after injection. **(A)** Defensive flinches and kicks were observed in chronically defeated female mice during social interactions with an awake conspecific (light bars) (stimulus  $\times$  defeat interaction:  $F_{2,17} = 3.67, p = .047$ ; stimulus:  $F_{1,17} = 15.3, p = .001$ ; defeat:  $F_{2,17} = 7.73, p = .004$ ). **(B–D)** Although chronically defeated mice engaged in more nasal contact with anesthetized stimulus animals (dark bars) (stimulus  $\times$  defeat interaction:  $F_{2,17} = 3.8, p = .043$ ; stimulus:  $F_{1,17} = 54.53, p < .0001$ ), they displayed significantly less anogenital/flank contact (stimulus  $\times$  defeat interaction:  $F_{2,17} = 3.87, p = .041$ ; stimulus:  $F_{1,17} = 13.67, p = .002$ ) and total contact (stimulus  $\times$  defeat interaction:  $F_{2,17} = 6.18, p = .0096$ ; stimulus:  $F_{1,17} = 65.11, p < .0001$ ) with awake stimulus mice compared with control animals. **(E, F)** Baseline total social contact with an awake stimulus female mouse in the home cage was suppressed in chronically defeated mice ( $t_{33} = 3.056, p = .0044$ ). **(G)** Ketamine (20 mg/kg) significantly increased social contact in chronically defeated female mice 24 hours after injection (drug  $\times$  defeat interaction:  $F_{1,30} = 4.26, p = .0478$ ; drug:  $F_{1,30} = 4.26, p = .0478$ ; defeat:  $F_{1,30} = 12.52, p = .0013$ ), **(H)** but did not reverse the high rate of behavioral transitions during social interaction tests (defeat:  $F_{1,30} = 11.00, p = .0024$ ). **(A–D, F, H)** Data are shown as mean  $\pm$  SEM. **(G)** Bars depict the maximum and minimum values. The dotted line marks no change in social contact between baseline and postinjection tests; values above the dotted line are increases from baseline, whereas values below the dotted line are decreases from baseline social contact time. Circles and squares represent individuals. \* $p < .05$ , \*\* $p < .01$ , \*\*\* $p < .001$  (**A, C, D**) compared with control mice interacting with an awake social stimulus female mouse, **(B)** compared with control mice interacting with an anesthetized social stimulus female mouse, **(F, H)** compared with control mice, or **(G)** compared with ketamine-treated control mice; # $p < .05$ , compared with saline-treated chronically defeated female mice. SI, social interaction.

flinches compared with control animals (Figure 3A) along with deficits in both anogenital/flank and total social contact (Figure 3C, D). When an anesthetized social stimulus mouse was placed in the home cage, chronically defeated female mice actually engaged in more nasal contact compared with animals in the control group (Figure 3B). Social investigation in the home cage also produced a substantial hyperthermic response, which was greater in chronically defeated versus control mice (Figure 4A); in contrast, there was no group difference in hyperthermia induced by novel object investigation (Figure 4B).

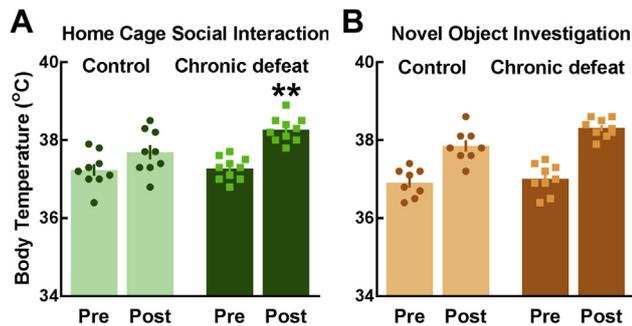
During pretreatment baseline home cage social interaction tests, social contact (i.e., nonaggressive social interaction initiated by the experimental female mouse) (Figure 3E) was significantly shorter among defeated animals (Figure 3F). Twenty-four hours after receiving a dose of ketamine (20 mg/kg), defeated female mice exhibited a significant increase from pretreatment social interactions compared with ketamine-treated control mice and saline-treated defeated mice (Figure 3G and Supplemental Videos S4–S7). This effect of ketamine was not evident 30 minutes or 5 days after injection (Supplemental Figure S6). Importantly, although ketamine increased social contact duration in

animals subjected to chronic defeat, a defeat-associated increase in behavioral transitions during social interactions persisted (Figure 3H).

Experimental mice were also examined in an open field social interaction test (Figure 5C), which is often employed to identify depressive-like phenotypes in chronically defeated male mice (8–10). Female mice exposed to chronic defeat exhibited significantly more vigilance-like behavior compared with animals in the control condition (Figure 5A, C, D and Supplemental Video S8), though the duration of time spent in a predefined social interaction zone was comparable between groups (Figure 5B). Correlational analyses of chronically defeated individuals revealed a significant inverse relationship between time investigating a social partner in the open field and vigilance-like behavior (Supplemental Table S2).

In terms of nonsocial behaviors, chronically defeated animals constructed nests that were significantly less developed than control mice as illustrated by measures of nest peak diameter, height, circularity, and overall nest score (Figure 6A–E). Importantly, there was no group difference in baseline body temperature (Figure 4), suggesting that nest-building deficits in defeated mice were probably not due to stress effects on

## Female Aggression and Chronic Defeat Stress



**Figure 4.** Hyperthermia was observed in response to 1.5-minute social interaction and 5-minute novel object investigation tests. Temperatures were measured immediately before (pre) and after (post) testing. **(A)** Chronically defeated female mice experienced a greater elevation in body temperature after social interactions compared with control animals (defeat  $\times$  time interaction:  $F_{1,17} = 5.91$ ,  $p = .027$ ; time:  $F_{1,17} = 42.21$ ,  $p < .0001$ ). **(B)** All mice showed a similar degree of hyperthermia in response to novel object investigation (time:  $F_{1,15} = 113.6$ ,  $p < .0001$ ). Data are shown as mean  $\pm$  SEM; \*\* $p < .05$ , compared with control posttest temperature.

thermoregulation. A defeat phenotype was also evident when novel objects were placed in the home cage. Defeated female mice spent significantly less time investigating and exhibited a greater number of defensive startle-like behaviors (i.e., flinching and jumping) compared with control individuals (Figure 7B). In contrast, measures of general anxiety-like behavior collected during light/dark box testing were similar between control and chronically defeated mice (Supplemental Figure S8). Interestingly, a greater number of attack bites received during the 10-day chronic social defeat stress protocol predicted reduced home cage social interactions and time spent in the light chamber during light/dark box testing (Supplemental Figure S9).

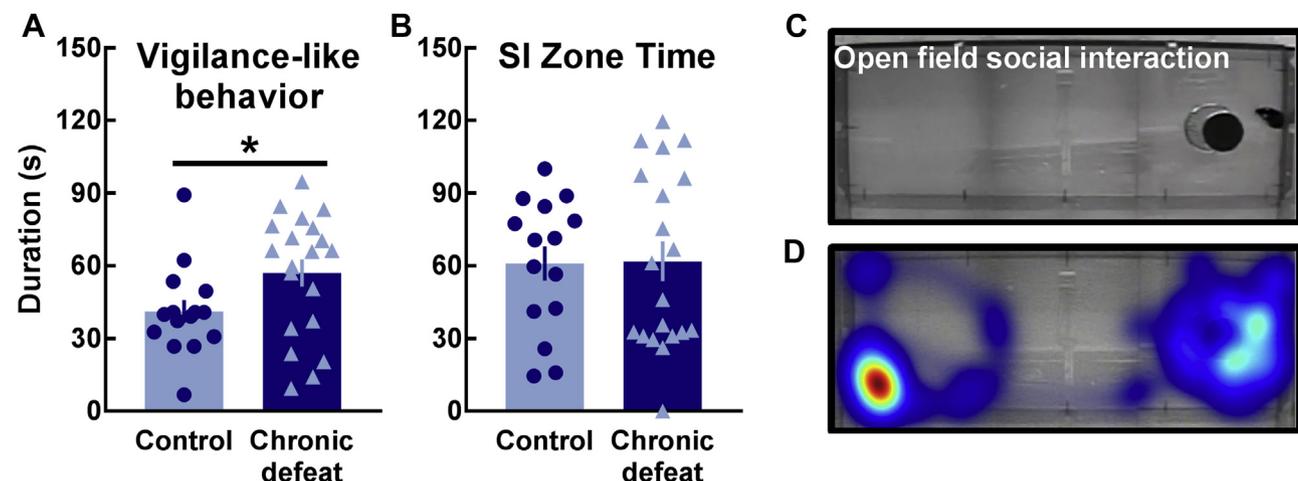
### DISCUSSION

We designed a novel and ethologically relevant model of chronic female social defeat stress that produces a distinct

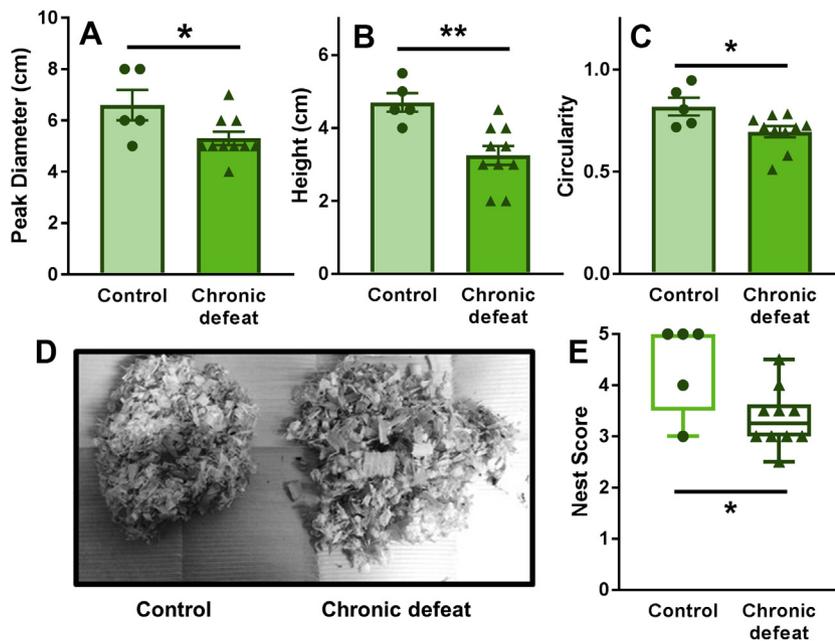
profile of neural and physiological effects along with pronounced depressive-like and anxiety-like behaviors in defeated female mice (Table 1). After 10 days of continuous social stress, female mice exhibited elevated levels of the stress hormone corticosterone and increased c-Fos activation in the MeA, LSv, VMHv, and hPVN. In the days to weeks following social defeat, female mice engaged in atypical behaviors during novel object investigation, nest building, and social interactions. Active investigation of a nonaggressive social partner during social interactions increased in defeated ketamine-treated individuals, indicating that our model of female chronic social defeat stress produces a phenotype that is sensitive to some antidepressant compounds (52,53). Though ketamine increased social contact, defeated female mice that received drug treatment continued to exhibit atypically high rates of behavioral transitioning. Reduced behavioral stability in the presence of a nonaggressive individual may reflect a sensitized social threat response. The behavioral selectivity of ketamine raises the possibility that distinct mechanisms underlie chronic stress-induced deficits in social contact versus vigilance-related impairments. These observations should be considered and extended in future preclinical studies focusing on stress-related psychopathologies that occur at higher rates in women than in men.

### Behavioral Effects of Chronic Female Social Defeat Stress

The pattern of aggressive behaviors recorded during inter-female agonistic encounters differed significantly from attack sequences during intermale fights. Considering the sophisticated exchange of multimodal sensory information between animals during ethological agonistic interactions, chronic social defeat stress procedures that rely on species-typical aggression may increase the translational potential of experimental findings. Similar to the pattern of aggression, significant features of the defeated female phenotype are distinct from the male phenotype, and in female mice, the severity of persistent



**Figure 5.** Chronically defeated female mice expressed more **(A)** vigilance-like behavior than control animals ( $t_{33} = 2.05$ , \* $p = .048$ ) despite both groups spending **(B)** similar durations within the social interaction (SI) zone. **(C)** Representative image of vigilance-like behavior displayed by a chronically defeated female mouse and **(D)** the corresponding heat map of activity during the open field social interaction test.



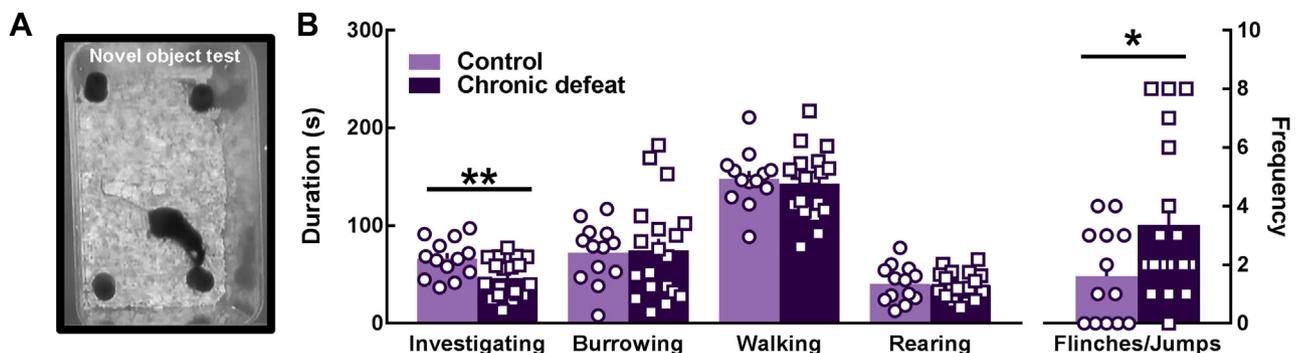
**Figure 6.** Nests constructed by chronically defeated female mice were underdeveloped compared with nests built by control animals, as measured by (A) nest peak diameter ( $t_{13} = 2.347$ ), (B) nest maximal height ( $t_{13} = 3.503$ ), (C, D) circularity (measured on a scale of 0–1, 1 = perfect circle) ( $t_{13} = 2.515$ ), and (E) nest score (Mann-Whitney  $U = 8.5$ ,  $p = .039$ ). Data are shown as (A–C) mean  $\pm$  SEM or as (E) maximum and minimum surrounding the median; \* $p < .05$ , \*\* $p < .01$  compared with control mice.

stress-induced behavioral deficits may depend on the severity of the stress experience. These observations should encourage a sex-specific approach to evaluating the consequences of defeat in male and female mice and the development of strategies tailored to treat specific symptoms (54).

We observed a hypervigilant-like phenotype in female mice subjected to chronic stress (46,55), which may reflect an inability to distinguish threatening from nonthreatening stimuli. Defeated female animals showed exaggerated defensive behaviors, such as whole-body flinches, backwards jumps, and defensive kicks toward nonaggressive social partners. These tests occurred within a familiar, nonthreatening environment, further illustrating impairments in threat assessment. Importantly, social deficits were not readily detected when social interaction zone time, a putative indicator of depressive-like behavior in male mice (9), was used as the dependent

measure. Some defeat-induced behavioral deficits manifest in a sex-specific fashion, highlighting the importance of evaluating novel potential pharmacotherapies with probes that can detect sexually dimorphic adaptations to chronic stress.

Nesting behavior was also impaired in chronically defeated female mice. Measures of nest construction can serve as an overall indicator of rodent health (56–58) and can be inhibited in male mice exposed to social stressors (59,60). As a goal-directed behavior, nesting requires a sequence of intricate actions to ultimately construct a protected, concave nest site (49,61). Among other possibilities, poor nesting may result from decreased concentration on task completion or impaired motivation to engage in potentially rewarding species-typical behaviors [nest material as a reinforcer (61–64)]. Defeated mice constructed incomplete nests, suggesting indecision or issues with concentration, both of which are cardinal symptoms of



**Figure 7.** (A, B) Chronically defeated female mice spent less time investigating novel objects ( $t_{30} = 2.849$ ,  $p = .0078$ ) and exhibited more defensive flinches and jumps ( $t_{30} = 2.126$ ,  $p = .0419$ ) compared with control mice. All data are shown as mean  $\pm$  SEM; \* $p < .05$ , \*\* $p < .01$ , control vs chronic defeat.

**Table 1. Effects of Social Defeat Stress in Female Mice**

	Acute Defeat	Chronic Defeat
Neural Activation (c-Fos)		
MeA	↔	↑
LSv	↑	↑
hPVN	↔	↑
VMHvl	↔	↑
PeN	↔	↔
amBNST	↔	↔
DG	↔	↔
Plasma Corticosterone	↑	↑
Social Interactions (HCSI)	↔	↓
Conspecific-Induced Defense (HCSI)	↔	↑
Social Hyperthermia (HCSI)	NA	↑
Social Vigilance (OFSI)	NA	↑
Social Interaction Zone Time (OFSI)	NA	↔
Novel Object Investigation (HCNO)	NA	↓
Novelty-Induced Defense (HCNO)	NA	↑
Novelty-Induced Hyperthermia (HCNO)	NA	↔
Nest Quality	NA	↓
Anxiety-like Behavior (LDB)	NA	↔

Consequences of acute or chronic social defeat stress in female C57BL/6J mice compared with control animals ( $p < .05$ ).

↔, no difference from control mice; ↑, significantly greater than control mice; ↓, significantly less than control mice; amBNST, anteromedial bed nucleus of the stria terminalis; DG, dentate gyrus; HCNO, home cage novel object test; HCSI, home cage social interaction test; hPVN, hypothalamic paraventricular nucleus; LDB, light/dark box test; LSv, ventral lateral septum; MeA, medial amygdala; NA, not available; OFSI, open field social interaction test; PeN, periventricular nucleus; VMHvl, ventrolateral division of the ventromedial hypothalamus.

posttraumatic stress disorder and major depressive disorder (65). Preferential allocation of attentional resources for threat assessment may impede nest completion in animals that exhibit a hypervigilant-like phenotype. Future studies that evaluate action sequence planning as well as the anticipatory, motivational, and learning processes that drive nesting behaviors could reveal unique circuit-level mechanisms that contribute to the defeat phenotype observed in female mice (61,66).

Estrous cycling was similar between stressed and non-stressed female mice during chronic social defeat. Additional investigations need to determine if cycle phase and circulating hormone concentrations influence specific behavioral end points in female rodents defeated by aggressive conspecifics. Further work is also required to fully examine which stress-induced behavioral impairments are sensitive to one-time versus repeated ketamine administration in animals subjected to ethological female social defeat stress (67). In our sample of defeated female mice, we did not observe a distinct bimodal distribution of susceptible and resilient animals (Supplemental Figure S7A). Large-scale experiments paralleling the studies conducted by Krishnan *et al.* (10) in defeated male mice are necessary to definitively address the possibility of subgroups within the defeated female population. Such work may clarify

the mechanisms that render some individuals more likely to develop affective psychopathologies and may guide the development of personalized treatment options.

### Chronic Social Defeat Stress and Increased c-Fos Activation in Sexually Dimorphic Brain Regions

Chronically defeated female mice exhibited c-Fos activation within several sexually dimorphic brain regions that comprise overlapping social (68–70) and defensive behavioral and threat-processing networks (46,55,71–74), including the MeA, LSv, VMHvl, and hPVN. Whereas these areas are also activated in male rodents (75–80) exposed to repeated social defeat stress, regional sexual dimorphism can contribute to significant sex-dependent behavioral outcomes in response to cues (81–83) and experiences (84–86), illustrating the potential for sex-specific social defeat phenotypes despite similar patterns of c-Fos activation in chronically defeated male and female mice.

Interestingly, estrogen receptor  $\alpha$ -expressing cells in the anterior portion of the VMHvl may control some aspects of active defense during acute social defeat (74). Our findings point to a similar cluster of cells that may be relevant to atypical social behaviors observed in chronically defeated female mice. Persistent stress-induced changes in estrogen signaling (5,87,88) within this cell population could contribute to exaggerated active defense and hypervigilance in female mice subjected to chronic social defeat stress.

### Future Directions: Female Stress and Aggression

Aggression is most often studied in the context of the male behavioral repertoire; yet, male and female rodents, nonhuman primates, and humans will readily engage in aggressive acts under certain conditions (22,89–93). We show that consistent and intense interfemale aggression can be generated in intact but not ovariectomized female mice living with a male conspecific. In addition, distinct aggressive and nonaggressive female subtypes are present within the aggressive subpopulation; some female mice display exclusively gestational aggression, whereas others engage in gestational and rival aggression (Supplemental Figure S10). Comparable studies in rats have shown that intact, nulliparous animals housed with sterile male rats will become highly and persistently aggressive toward unfamiliar conspecifics (94), suggesting that the present murine model could be extended to study female rats under analogous defeat conditions. Additionally, similarities between female mice and rats (94–96) raise the possibility that there may be some adaptive and potentially conserved elements of interfemale rival aggression in rodents as well as other mammalian species.

Chronic social defeat stress produced a pattern of functional activation in brain areas also activated during mating or aggression in female mice, including the MeA (82,97) and VMHvl (98–101). It remains unclear whether these cell populations are functionally, molecularly, and spatially discrete or overlapping (86,102). Interactions between these networks could allow one social experience to modify later behaviors; for example, stress-associated activation of cells in aggression-related brain regions could affect aggressive performance or motivation to engage in future agonistic behaviors. To address this, further behavioral and

molecular studies are necessary to test the potential for rival aggression to motivate operant responding in control and socially defeated female mice [compare with 103–108].

### ACKNOWLEDGMENTS AND DISCLOSURES

This work was supported by the National Institutes of Health (Grant No. F31AA025827 [to ELN], Grant No. R01MH108665 [to KJR], and Grant Nos. R01AA013983 and R01DA031734 [to KAM]). The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

We thank Mr. Thomas J. Sopko for his technical expertise and Tufts University undergraduate research assistants Avni Rajpal and Susannah LaPointe.

The authors report no biomedical financial interests or potential conflicts of interest.

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Received Mar 13, 2019; revised Apr 30, 2019; accepted May 3, 2019.

Supplementary material cited in this article is available online at <https://doi.org/10.1016/j.biopsych.2019.05.005>.

### REFERENCES

- Kessler RC, McGonagle KA, Swartz M, Blazer DG, Nelson CB (1993): Sex and depression in the national comorbidity survey. I: Lifetime prevalence, chronicity and recurrence. *J Affect Disord* 29:85–96.
- Kessler RC, Sonnega A, Bromet E, Hughes M, Nelson CB (1995): Posttraumatic stress disorder in the national comorbidity survey. *Arch Gen Psychiatry* 52:1048–1060.
- Kessler RC (1997): The effects of stressful life events on depression. *Annu Rev Psychol* 48:191–214.
- Kessler RC (2003): Epidemiology of women and depression. *J Affect Disord* 74:5–13.
- Ramkise TS, Ressler KJ (2018): Mechanisms of sex differences in fear and posttraumatic stress disorder. *Biol Psychiatry* 83:876–885.
- Rubinow DR, Schmidt PJ (2019): Sex differences and the neurobiology of affective disorders. *Neuropsychopharmacology* 44:111–128.
- Miczek KA, Thompson ML, Shuster L (1982): Opioid-like analgesia in defeated mice. *Science* 215:1520–1522.
- Kudryavtseva NN, Bakshantovskaya IV, Koryakina LA (1991): Social model of depression in mice of C57BL/6J strain. *Pharmacol Biochem Behav* 38:315–320.
- Berton O, McClung CA, Dileone RJ, Krishnan V, Renthal W, Russo SJ, *et al.* (2006): Essential role of BDNF in the mesolimbic dopamine pathway in social defeat stress. *Science* 311:864–868.
- Krishnan V, Han MH, Graham DL, Berton O, Renthal W, Russo SJ, *et al.* (2007): Molecular adaptations underlying susceptibility and resistance to social defeat in brain reward regions. *Cell* 131:391–404.
- Nestler EJ, Hyman SE (2010): Animal models of neuropsychiatric disorders. *Nat Neurosci* 13:1161–1169.
- Covington HE, Lobo MK, Maze I, Vialou V, Hyman JM, Zaman S, *et al.* (2010): Antidepressant effect of optogenetic stimulation of the medial prefrontal cortex. *J Neurosci* 30:16082–16090.
- Vialou V, Robison AJ, LaPlant QC, Covington HE, Dietz DM, Ohnishi YN, *et al.* (2010): DeltaFosB in brain reward circuits mediates resilience to stress and antidepressant responses. *Nat Neurosci* 13:745–752.
- Golden SA, Covington HE, Berton O, Russo SJ (2011): A standardized protocol for repeated social defeat stress in mice. *Nat Protoc* 6:1183–1191.
- Tsankova NM, Berton O, Renthal W, Kumar A, Neve RL, Nestler EJ (2006): Sustained hippocampal chromatin regulation in a mouse model of depression and antidepressant action. *Nat Neurosci* 9:519–525.
- Covington HE, Maze I, LaPlant QC, Vialou VF, Ohnishi YN, Berton O, *et al.* (2009): Antidepressant actions of histone deacetylase inhibitors. *J Neurosci* 29:11451–11460.
- Donahue RJ, Muschamp JW, Russo SJ, Nestler EJ, Carlezon WA (2014): Effects of striatal DeltaFosB overexpression and ketamine on social defeat stress-induced anhedonia in mice. *Biol Psychiatry* 76:550–558.
- Bagot RC, Cates HM, Purushothaman I, Vialou V, Heller EA, Yieh L, *et al.* (2017): Ketamine and imipramine reverse transcriptional signatures of susceptibility and induce resilience-specific gene expression profiles. *Biol Psychiatry* 81:285–295.
- Harris AZ, Atsak P, Bretton ZH, Holt ES, Alam R, Morton MP, *et al.* (2018): A novel method for chronic social defeat stress in female mice. *Neuropsychopharmacology* 43:1276–1283.
- Takahashi A, Chung JR, Zhang S, Zhang HX, Grossman Y, Aleyasin H, *et al.* (2017): Establishment of a repeated social defeat stress model in female mice. *Sci Rep* 7:12838.
- Iniguez SD, Flores-Ramirez FJ, Riggs LM, Alipio JB, Garcia-Carachure I, Hernandez MA, *et al.* (2018): Vicarious social defeat stress induces depression-related outcomes in female mice. *Biol Psychiatry* 83:9–17.
- Miczek KA, Maxson SC, Fish EW, Faccidomo S (2001): Aggressive behavioral phenotypes in mice. *Behav Brain Res* 125:167–181.
- Noiro E (1969): Interactions between reproductive and territorial behavior in female mice. *Int Ment Health Res News* 11:10–11.
- Noiro E, Goyens J, Buhot MC (1975): Aggressive behavior of pregnant mice toward males. *Horm Behav* 6:9–17.
- Mann MA, Svare B (1982): Factors influencing pregnancy-induced aggression in mice. *Behav Neural Biol* 36:242–258.
- Ogawa S, Makino J (1984): Aggressive behavior in inbred strains of mice during pregnancy. *Behav Neural Biol* 40:195–204.
- Hedricks C, Daniels CE (1981): Agonistic behavior between pregnant mice and male intruders. *Behav Neural Biol* 31:236–241.
- DeBold JF, Miczek KA (1984): Aggression persists after ovariectomy in female rats. *Horm Behav* 18:177–190.
- Svare B, Gandelman R (1973): Postpartum aggression in mice: Experiential and environmental factors. *Horm Behav* 4:323–334.
- Svare B, Gandelman R (1976): Suckling stimulation induces aggression in virgin female mice. *Nature* 260:606–608.
- Haney M, DeBold JF, Miczek KA (1989): Maternal aggression in mice and rats towards male and female conspecifics. *Aggress Behav* 15:443–453.
- Parmigiani S, Brain PF, Mainardi D, Brunoni V (1988): Different patterns of biting attack employed by lactating female mice (*Mus domesticus*) in encounters with male and female conspecific intruders. *J Comp Psychol* 102:287–293.
- Rosenson LM, Asheroff AK (1975): Maternal aggression in CD-1 mice: Influence of the hormonal condition of the intruder. *Behav Biol* 15:219–224.
- Clipperton-Allen AE, Cragg CL, Wood AJ, Pfaff DW, Choleris E (2010): Agonistic behavior in males and females: Effects of an estrogen receptor beta agonist in gonadectomized and gonadally intact mice. *Psychoneuroendocrinology* 35:1008–1022.
- Valzelli L (1973): The “isolation syndrome” in mice. *Psychopharmacology* 31:305–320.
- Miczek KA, O'Donnell JM (1978): Intruder-evoked aggression in isolated and nonisolated mice: Effects of psychomotor stimulants and L-dopa. *Psychopharmacology* 57:47–55.
- Davis ES, Marler CA (2003): The progesterone challenge: Steroid hormone changes following a simulated territorial intrusion in female *Peromyscus californicus*. *Horm Behav* 44:185–198.

## Female Aggression and Chronic Defeat Stress

38. Brain PF (1972): Effects of isolation/grouping on endocrine function and fighting behavior in male and female golden hamsters (*Mesocricetus auratus* Waterhouse). *Behav Biol* 7:349–357.
39. Grell DF, Papson BA, Cole JE, Rowe FA (1974): The influence of caging conditions and hormone treatments on fighting in male and female hamsters. *Horm Behav* 5:355–366.
40. McLean AC, Valenzuela N, Fai S, Bennett SA (2012): Performing vaginal lavage, crystal violet staining, and vaginal cytological evaluation for mouse estrous cycle staging identification. *J Vis Exp* 67:e4389.
41. Been LE, Gibbons AB, Meisel RL (2019): Towards a neurobiology of female aggression. *Neuropharmacology* 156:107451.
42. Floody OR, Pfaff DW (1977): Aggressive behavior in female hamsters: The hormonal basis for fluctuations in female aggressiveness correlated with estrous state. *J Comp Physiol Psychol* 91:443–464.
43. Hyde J, Sawyer TF (1977): Estrous cycle fluctuations in aggressiveness of house mice. *Horm Behav* 9:290–295.
44. de Jong TR, Beiderbeck DI, Neumann ID (2014): Measuring virgin female aggression in the female intruder test (FIT): Effects of oxytocin, estrous cycle, and anxiety. *PLoS One* 9:e91701.
45. More L (2008): Intra-female aggression in the mouse (*Mus musculus domesticus*) is linked to the estrous cycle regularity but not to ovulation. *Aggress Behav* 34:46–50.
46. Duque-Wilckens N, Steinman MQ, Busnelli M, Chini B, Yokoyama S, Pham M, et al. (2018): Oxytocin receptors in the anteromedial bed nucleus of the stria terminalis promote stress-induced social avoidance in female California mice. *Biol Psychiatry* 83:203–213.
47. Zanos P, Moaddel R, Morris PJ, Georgiou P, Fischell J, Elmer GI, et al. (2016): NMDAR inhibition-independent antidepressant actions of ketamine metabolites. *Nature* 533:481–486.
48. Hultman R, Ulrich K, Sachs BD, Blount C, Carlson DE, Ndubuizu N, et al. (2018): Brain-wide electrical spatiotemporal dynamics encode depression vulnerability. *Cell* 173:166–180.e114.
49. Deacon RM (2006): Assessing nest building in mice. *Nat Protoc* 1:1117–1119.
50. Lisicotto CA, DeBold JF, Miczek KA (1990): Sexual differentiation and the effects of alcohol on aggressive behavior in mice. *Pharmacol Biochem Behav* 35:357–362.
51. Norman KJ, Seiden JA, Klickstein JA, Han X, Hwa LS, DeBold JF, et al. (2015): Social stress and escalated drug self-administration in mice: I. Alcohol and corticosterone. *Psychopharmacology* 232:991–1001.
52. Berman RM, Cappiello A, Anand A, Oren DA, Heninger GR, Charney DS, et al. (2000): Antidepressant effects of ketamine in depressed patients. *Biol Psychiatry* 47:351–354.
53. Zarate CA, Singh JB, Carlson PJ, Brutsche NE, Ameli R, Luckenbaugh DA, et al. (2006): A randomized trial of an N-methyl-D-aspartate antagonist in treatment-resistant major depression. *Arch Gen Psychiatry* 63:856–864.
54. Dzirasa K, Covington HE (2012): Increasing the validity of experimental models for depression. *Ann N Y Acad Sci* 1265:36–45.
55. Blanchard DC, Griebel G, Pobbe R, Blanchard RJ (2011): Risk assessment as an evolved threat detection and analysis process. *Neurosci Biobehav Rev* 35:991–998.
56. Van de Weerd HA, Van Loo PL, Van Zutphen LF, Koolhaas JM, Baumans V (1997): Preferences for nesting material as environmental enrichment for laboratory mice. *Lab Anim* 31:133–143.
57. Jirkof P (2014): Burrowing and nest building behavior as indicators of well-being in mice. *J Neurosci Methods* 234:139–146.
58. Gaskill BN, Karas AZ, Garner JP, Pritchett-Corning KR (2013): Nest building as an indicator of health and welfare in laboratory mice. *J Vis Exp* 82:51012.
59. Otabi H, Goto T, Okayama T, Kohari D, Toyoda A (2016): Subchronic and mild social defeat stress alter mouse nest building behavior. *Behav Processes* 122:21–25.
60. Rettich A, Käsemann HP, Pelczar P, Bürki K, Arras M (2006): The physiological and behavioral impact of sensory contact among unfamiliar adult mice in the laboratory. *J Appl Anim Welf Sci* 9:277–288.
61. Roper TJ (1976): Self-sustaining activities and reinforcement in the nest building behaviour of mice. *Behaviour* 59:40–58.
62. Roper TJ (1973): Nesting material as a reinforcer for female mice. *Anim Behav* 21:733–740.
63. Jansen PE, Goodman ED, Jowaisas D, Bunnell BN (1969): Paper as a positive reinforcer for acquisition of bar press response by the golden hamster. *Psychon Sci* 16:113–114.
64. Oley NN, Slotnick BM (1970): Nesting material as a reinforcement for operant behavior in the rat. *Psychon Sci* 21:41–43.
65. American Psychiatric Association (2013): *Diagnostic and Statistical Manual of Mental Disorders*, 5th ed. Arlington, VA: American Psychiatric Press.
66. Der-Avakian A, Markou A (2012): The neurobiology of anhedonia and other reward-related deficits. *Trends Neurosci* 35:68–77.
67. Strong CE, Kabbaj M (2018): On the safety of repeated ketamine infusions for the treatment of depression: Effects of sex and developmental periods. *Neurobiol Stress* 9:166–175.
68. Newman SW (1999): The medial extended amygdala in male reproductive behavior: A node in the mammalian social behavior network. *Ann N Y Acad Sci* 877:242–257.
69. O'Connell LA, Hofmann HA (2011): The vertebrate mesolimbic reward system and social behavior network: A comparative synthesis. *J Comp Neurol* 519:3599–3639.
70. Kim Y, Venkataraju KU, Pradhan K, Mende C, Taranda J, Turaga SC, et al. (2015): Mapping social behavior-induced brain activation at cellular resolution in the mouse. *Cell Rep* 10:292–305.
71. Canteras NS (2002): The medial hypothalamic defensive system: Hodological organization and functional implications. *Pharmacol Biochem Behav* 71:481–491.
72. Steinman MQ, Duque-Wilckens N, Greenberg GD, Hao R, Campi KL, Laredo SA, et al. (2016): Sex-specific effects of stress on oxytocin neurons correspond with responses to intranasal oxytocin. *Biol Psychiatry* 80:406–414.
73. Wang L, Chen IZ, Lin D (2015): Collateral pathways from the ventromedial hypothalamus mediate defensive behaviors. *Neuron* 85:1344–1358.
74. Wang L, Talwar V, Osakada T, Kuang A, Guo Z, Yamaguchi T, et al. (2019): Hypothalamic control of conspecific self-defense. *Cell Rep* 26:1747–1758.e1745.
75. Matsuda S, Peng H, Yoshimura H, Wen TC, Fukuda T, Sakanaka M (1996): Persistent c-fos expression in the brains of mice with chronic social stress. *Neurosci Res* 26:157–170.
76. Wohleb ES, Hanke ML, Corona AW, Powell ND, Stiner LM, Bailey MT, et al. (2011): Beta-adrenergic receptor antagonism prevents anxiety-like behavior and microglial reactivity induced by repeated social defeat. *J Neurosci* 31:6277–6288.
77. Martinez M, Phillips PJ, Herbert J (1998): Adaptation in patterns of c-fos expression in the brain associated with exposure to either single or repeated social stress in male rats. *Eur J Neurosci* 10:20–33.
78. Nikulina EM, Covington HE, Ganschow L, Hammer RP, Miczek KA (2004): Long-term behavioral and neuronal cross-sensitization to amphetamine induced by repeated brief social defeat stress: fos in the ventral tegmental area and amygdala. *Neuroscience* 123:857–865.
79. Kollack-Walker S, Don C, Watson SJ, Akil H (1999): Differential expression of c-fos mRNA within neurocircuits of male hamsters exposed to acute or chronic defeat. *J Neuroendocrinol* 11:547–559.
80. Martinez M, Calvo-Torrent A, Herbert J (2002): Mapping brain response to social stress in rodents with c-fos expression: A review. *Stress* 5:3–13.
81. Xu PS, Lee D, Holy TE (2016): Experience-dependent plasticity drives individual differences in pheromone-sensing neurons. *Neuron* 91:878–892.
82. Ishii KK, Osakada T, Mori H, Miyasaka N, Yoshihara Y, Miyamichi K, et al. (2017): A labeled-line neural circuit for pheromone-mediated sexual behaviors in mice. *Neuron* 95:123–137.e128.
83. Lischinsky JE, Sokolowski K, Li P, Esumi S, Kamal Y, Goodrich M, et al. (2017): Embryonic transcription factor expression in mice

- predicts medial amygdala neuronal identity and sex-specific responses to innate behavioral cues. *Elife* 6.
84. Li Y, Mathis A, Grewe BF, Osterhout JA, Ahanonu B, Schnitzer MJ, *et al.* (2017): Neuronal representation of social information in the medial amygdala of awake behaving mice. *Cell* 171:1176–1190.e1117.
  85. Remedios R, Kennedy A, Zelikowsky M, Grewe BF, Schnitzer MJ, Anderson DJ (2017): Social behaviour shapes hypothalamic neural ensemble representations of conspecific sex. *Nature* 550:388–392.
  86. Moffitt JR, Bambah-Mukku D, Eichhorn SW, Vaughn E, Shekhar K, Perez JD, *et al.* (2018): Molecular, spatial, and functional single-cell profiling of the hypothalamic preoptic region. *Science* 362(6416).
  87. Ressler KJ, Mercer KB, Bradley B, Jovanovic T, Mahan A, Kerley K, *et al.* (2011): Post-traumatic stress disorder is associated with PACAP and the PAC1 receptor. *Nature* 470:492–497.
  88. Mercer KB, Dias B, Shafer D, Maddox SA, Mülle JG, Hu P, *et al.* (2016): Functional evaluation of a PTSD-associated genetic variant: estradiol regulation and ADCYAP1R1. *Transl Psychiatry* 6:e978.
  89. Haug M (1978): Attack by female mice on “strangers”. *Aggress Behav* 4:133–139.
  90. Stockley P, Campbell A (2013): Female competition and aggression: Interdisciplinary perspectives. *Philos Trans R Soc Lond B Biol Sci* 368:20130073.
  91. Reinhardt V, Reinhardt A, Reinhardt C (1987): Evaluating sex differences in aggressiveness in cattle, bison and rhesus monkeys. *Behaviour* 102:58–66.
  92. Duque-Wilckens N, Trainor BC (2017): Behavioral neuroendocrinology of female aggression. *Oxford Research Encyclopedia of Neuroscience* Feb 2017. Available at: <https://oxfordre.com/neuroscience/view/10.1093/acrefore/9780190264086.001.0001/acrefore-9780190264086-e-11?rskey=UuRPG8&result=1>. Accessed September 12, 2019.
  93. Denson TF, O’Dean SM, Blake KR, Beames JR (2018): Aggression in women: Behavior, brain and hormones. *Front Behav Neurosci* 12:81.
  94. Albert DJ, Dyson EM, Petrovic DM, Walsh ML (1988): Activation of aggression in female rats by normal males and by castrated males with testosterone implants. *Physiol Behav* 44:9–13.
  95. Albert DJ, Petrovic DM, Walsh ML (1989): Ovariectomy attenuates aggression by female rats cohabiting with sexually active sterile males. *Physiol Behav* 45:225–228.
  96. Albert DJ, Jonik RH, Watson NV, Moe IV, Walsh ML (1991): Aggression by a female rat cohabiting with a sterile male: Termination of pseudopregnancy does not abolish aggression. *Physiol Behav* 50:519–523.
  97. Unger EK, Burke KJ, Yang CF, Bender KJ, Fuller PM, Shah NM (2015): Medial amygdalar aromatase neurons regulate aggression in both sexes. *Cell Rep* 10:453–462.
  98. Yang CF, Chiang MC, Gray DC, Prabhakaran M, Alvarado M, Juntti SA, *et al.* (2013): Sexually dimorphic neurons in the ventromedial hypothalamus govern mating in both sexes and aggression in males. *Cell* 153:896–909.
  99. Hashikawa K, Hashikawa Y, Tremblay R, Zhang J, Feng JE, Sabol A, *et al.* (2017): *Esr1* + cells in the ventromedial hypothalamus control female aggression. *Nat Neurosci* 20:1580–1590.
  100. Chen P, Hong W (2018): Neural circuit mechanisms of social behavior. *Neuron* 98:16–30.
  101. Hashikawa K, Hashikawa Y, Lischinsky J, Lin D (2018): The neural mechanisms of sexually dimorphic aggressive behaviors. *Trends Genet* 34:755–776.
  102. Sakurai K, Zhao S, Takatoh J, Rodriguez E, Lu J, Leavitt AD, *et al.* (2016): Capturing and manipulating activated neuronal ensembles with CANE delineates a hypothalamic social-fear circuit. *Neuron* 92:739–753.
  103. Covington HE, Newman EL, Tran S, Walton L, Hayek W, Leonard MZ, *et al.* (2018): The urge to fight: Persistent escalation by alcohol and role of NMDA receptors in mice. *Front Behav Neurosci* 12:206.
  104. Golden SA, Heins C, Venniro M, Caprioli D, Zhang M, Epstein DH, *et al.* (2017): Compulsive addiction-like aggressive behavior in mice. *Biol Psychiatry* 82:239–248.
  105. Falkner AL, Grosenick L, Davidson TJ, Deisseroth K, Lin D (2016): Hypothalamic control of male aggression-seeking behavior. *Nat Neurosci* 19:596–604.
  106. Fish EW, De Bold JF, Miczek KA (2002): Aggressive behavior as a reinforcer in mice: Activation by allopregnanolone. *Psychopharmacology* 163:459–466.
  107. Fish EW, McKenzie-Quirk SD, Bannai M, Miczek KA (2008): 5-HT(1B) receptor inhibition of alcohol-heightened aggression in mice: Comparison to drinking and running. *Psychopharmacology* 197:145–156.
  108. Couppis MH, Kennedy CH (2008): The rewarding effect of aggression is reduced by nucleus accumbens dopamine receptor antagonism in mice. *Psychopharmacology* 197:449–456.