

# Exercise as Adjunct Therapy in Cancer

Kathleen A. Ashcraft, PhD,<sup>\*,1</sup> Allison Betof Warner, MD, PhD,<sup>†,1</sup> Lee W. Jones, PhD,<sup>‡,‡,2</sup>  
and Mark W. Dewhirst, DVM, PD<sup>\*,2</sup>



Data from observational studies indicate that both physical activity as well as exercise (ie, structured physical activity) is associated with reductions in the risk of recurrence and cancer mortality after a diagnosis of certain forms of cancer. Emerging evidence from pre-clinical studies indicates that physical activity/exercise paradigms regulate intratumoral vascular maturity and perfusion, hypoxia, and metabolism and augments the antitumor immune response. Such responses may, in turn, enhance response to standard anticancer treatments. For instance, exercise improves efficacy of chemotherapeutic agents, and there is rationale to believe that it will also improve radiotherapy response. This review overviews the current preclinical as well as clinical evidence supporting exercise modulation of therapeutic response and postulated biological mechanisms underpinning such effects. We also examine the implications for tumor response to radiation, chemotherapy, and immunotherapy.

Semin Radiat Oncol 29:16–24 © 2018 Elsevier Inc. All rights reserved.

## Introduction

Over the past 2 decades, increased research and clinical attention has focused on the efficacy of exercise therapy as an adjunct strategy following a cancer diagnosis.<sup>1,2</sup> Randomized trials demonstrate structured exercise therapy, is a feasible adjunct strategy associated with significant improvements in symptom-related outcomes including exercise tolerance<sup>3,4</sup> as well as multiple patient-reported end points such as fatigue, quality of life, and physical functioning both during conventional adjuvant therapy. We have previously reviewed the preclinical literature to assess the role of exercise in tumor incidence, progression, and metastasis.<sup>5</sup> A critical corollary is whether exercise impacts the antitumor efficacy of cancer treatment. Such a notion is biologically plausible, as emerging evidence suggests exercise modulates several factors inherent in cancer treatment sensitivity, including radiotherapy. Arguably, some of the most relevant are alterations in the tumor microenvironment (TME)

include tumor hypoxia, perfusion, tumor cell metabolism, and the antitumor immune phenotype.

In this review, we outline how exercise-mediated changes in the TME and antitumor immune response may influence the radiation response. We further explore the impact of exercise on additional treatment modalities such as chemotherapy and immunotherapy.

Reduction in systemic levels of oxidative stress may play a role in improved neurocognitive function after chemotherapy. Several studies have shown increases in levels of circulating inflammatory markers in women with breast cancer, who underwent adjuvant chemotherapy.<sup>6,7</sup> Changes in systemic levels of several inflammatory markers are associated with reduced neurocognitive performance. Importantly, patients who undergo exercise in conjunction with chemotherapy exhibit reduced levels of inflammatory biomarkers and retain neurocognitive function. However, cancer patients participating in a thrice-weekly exercise program demonstrated reduced blood 8-OHdG levels,<sup>8</sup> a significant (41%) increase in systemic antioxidant capacity, and a significant (36%) decrease in protein oxidation; these changes correlated with reduced cancer-related fatigue.<sup>9</sup> It is not known whether the systemic changes in oxidative stress caused by exercise extend to effects on tumor. We previously reported that physical activity improves perfusion and reduces hypoxia in the 4T1 breast tumor model.<sup>10</sup> Not reported was our observation that levels of oxidative stress in the tumor, as depicted by 8-OHdG levels within tumor, were reduced by nearly 3-fold in tumors of mice that engaged in physical activity (running wheel) vs sedentary controls (Fig. 1). It is

<sup>\*</sup>Departments of Radiation Oncology, Duke University School of Medicine, Durham, NC.

<sup>†</sup>Department of Medicine, Memorial Sloan Kettering Cancer Center, New York, NY.

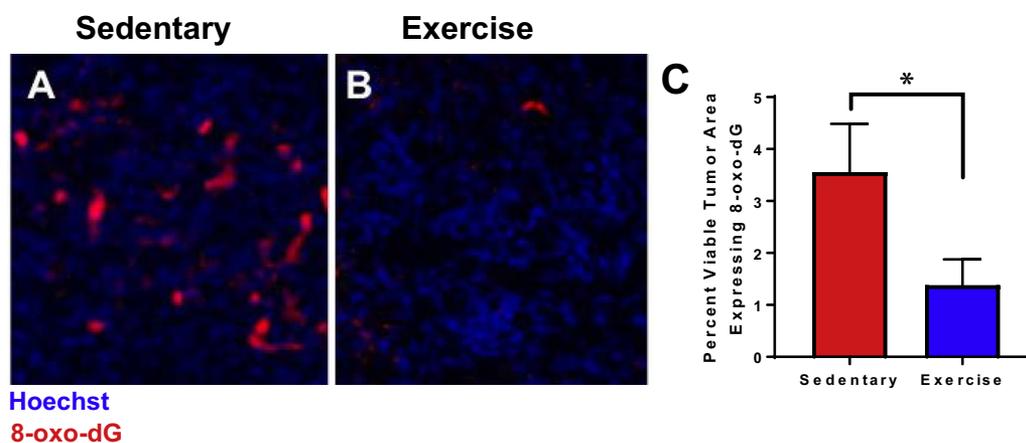
<sup>‡</sup>Weill Cornell Medical College, New York, NY.

Address reprint requests to Mark W. Dewhirst, DVM, PD, Department of Radiation Oncology, Duke University Medical Center, Box 3455, Medical Sciences Research Building 1, Room 201, Durham, NC 27710

E-mail: [mark.dewhirst@duke.edu](mailto:mark.dewhirst@duke.edu)

<sup>1</sup> These authors contributed equally.

<sup>2</sup> These authors contributed equally.



**Figure 1** 4T1 tumor sections were immunostained for 8-oxo-dG (red), a marker of oxidative damage to DNA. Cellular nuclei were stained with Hoechst 33,342 (blue). (A) and (B) show representative tumors taken from sedentary (A) or running (B) mice (5× objective, magnification 100%). Panel C shows quantification of 8-oxo-dG + pixels per square millimeter of viable tumor area. Error bars represent SEM, N = 23, 20, \* indicates  $p < 0.05$ , Mann-Whitney U test (Color version of figure is available online.)

not known whether changes in oxidative stress within the tumor were a consequence of alteration in systemic levels of oxidative stress or whether they were secondary to local alterations in the TME induced by physical activity. Furthermore, the consequences of reduced oxidative stress associated with exercise on treatment response are not defined. Additionally, in order to fully understand how exercise affects oxidative stress, a more comprehensive set of studies with more in-depth assessment of the oxidative stress environment within the host and the tumor would need to be conducted.<sup>11</sup>

## Exercise Regulation of the Host Milieu

The direct effects of exercise on the respiratory, cardiovascular, and musculoskeletal systems are well-described.<sup>12</sup> In response to repeated bouts of exercise, multiple organ systems adapt, resulting in improved oxygen delivery and utilization,  $VO_2$  max, and mitochondrial biogenesis.

These exercise-induced physiologic adaptations (ie, changes in glucose metabolism, circulating insulin levels, mitochondrial biogenesis, angiogenesis signaling pathways, and cytokine release) are not confined to the skeletal muscle. They have broad-reaching systemic implications that affect the overall health of the host. It is now becoming apparent that systemic exercise-induced changes influence growing tumor tissue as well, and have the potential to profoundly impact the TME and treatment response, as will be detailed below.

## Tumor Angiogenesis and Vascular Function

Solid tumor growth is dependent on angiogenesis, supported by optimized levels of vascular endothelial growth factor (VEGF) and other proangiogenic cytokines in the

TME. VEGF expression is increased under hypoxic conditions, driven by signaling through the hypoxia-inducible factor-1 (HIF-1) pathway.<sup>13</sup> However, the pressure to create new blood vessels able to serve an ever-evolving mass results in chaotic and immature vessel structure. Consequently, most tumors feature tortuous and leaky vasculature, characterized by shunts, low microvessel density, and poor pericyte coverage.<sup>14</sup> Aberrant tumor vasculature causes mismatch between oxygen supply and demand, resulting in pockets of hypoxia. Hypoxic regions are intensified by a decreased microvessel density, which result in regions too far from a vessel to receive adequate oxygen.<sup>15</sup> Finally, poor pericyte coverage is associated with leaky tumor vessels and contributes to invasion and metastasis.<sup>16</sup>

Tumor angiogenesis and vascular normalization are targets of antitumor therapeutics.<sup>17</sup> “Normalization” of tumor vasculature refers to several steps, including improvement in vascular maturity (reduced permeability, increased pericyte coverage, and reduced microvessel diameter), combined with pruning of redundant, or nonfunctional microvessels. The normalization process, by definition, therefore, results in a lowered vascular density. However, efficient normalization increases oxygen and drug delivery because of improvement in the orientation and network structure of the resulting normalized vascular network.

Initial trials with antiangiogenic agents were promising, but clinical use of antiangiogenic agents became problematic in many tumor types due to treatment resistance, vascular rarefaction, and re-emergence of hypoxia following prolonged use of such agents.<sup>18,19</sup> Some of the negative effects of angiogenesis inhibitors have been avoided by using lower doses of antiangiogenic agents.<sup>18</sup> This approach prolongs the period of normalization, which could prove beneficial for optimizing drug delivery or enhancing radiotherapy. Nevertheless, alternative approaches to altering tumor vascularity/angiogenesis may have clinical promise—exercise is one such potential strategy.

Vascular changes are perhaps the most well-documented effects of exercise on solid tumor physiology in preclinical studies. Several reports have shown that exercise increases tumor VEGF levels,<sup>10,20,21</sup> vessel density, and perfusion.<sup>10,22</sup> In addition to VEGF, platelet-derived growth factor receptor-beta is involved in endothelial cell recruitment during angiogenesis.<sup>23</sup> Interestingly, 1 report showed that exercise increased tumor VEGF expression occurred in conjunction with a reduction in platelet-derived growth factor receptor-beta expression<sup>10</sup> leading to an overall increase in tumor angiogenesis, which dramatically reduces tumor hypoxia through increased microvessel density and perfusion.

The increased tumor microvessel density<sup>10</sup> and perfusion<sup>22</sup> may have implications for drug delivery and are associated with reduced tumor hypoxia (discussed in more detail below). Importantly, we have also shown that exercise increases HIF-1 expression, which is regulated by oxidative stress.<sup>22,24</sup> Therefore, the observed differences in microvessel density and perfusion may reflect how exercise either reduces oxidative stress or improves the tumors' ability to deal with oxidative stress.

Functional vasculature that can effectively deliver blood, oxygen, and systemic therapies requires mature, long vessels with visible lumens, and few nonfunctional sprouts. Emerging data suggest that exercise may improve vascular function/maturity. Increased shear stress during exercise results in vascular remodeling, as defined by increased number of visible vessel lumens and longer average vessel length.<sup>25</sup> Exercise-induced shear stress activates the transcription factor and nuclear factor of activated T-cells, which in turn increases transcription of thrombospondin 1, thereby promoting vascular maturity. These changes have important implications for delivery and potential efficacy of systemic anticancer agents. For instance, 2 studies have demonstrated improved efficacy of chemotherapy in combination with exercise. Our group demonstrated that voluntary running improved response to cyclophosphamide in mice bearing syngeneic mammary tumors.<sup>10</sup> Three doses of cyclophosphamide significantly slowed tumor growth compared in exercising mice compared to sedentary control mice. Similarly, Schadler et al found that gemcitabine was significantly more efficacious at slowing tumor growth in a mouse pancreatic ductal adenocarcinoma model, when administered to mice running at 60%-70% of exercise capacity using a forced exercise paradigm, compared to sedentary mice.<sup>25</sup> This effect was abrogated in thrombospondin-1 knockout mice, suggesting that exercise-mediated vascular normalization is an important mechanism underpinning the exercise-chemotherapy efficacy relationship. Schadler et al quantified tumor doxorubicin concentrations 20 minutes post-treatment and found that when doxorubicin administration was preceded by 2 weeks of treadmill running (45 minutes of 12 m/min, 5 d/wk) doxorubicin concentrations increased in murine PDAC allografts nearly 2-fold.<sup>25</sup> Further, doxorubicin was more efficacious at slowing B16F10 melanoma growth in exercising vs sedentary mice.

The underlying mechanisms for improved tumor growth delay with the combination of exercise and chemotherapy is

likely multifold. First, and perhaps the most logical, is that the increase in functional tumor microvessels increases chemotherapy exposure to a higher portion of tumor cells. Normalization of the tumor vasculature also may promote homogeneous blood flow throughout the tumor, eliminating shunts that leave certain regions unexposed to drug. Using magnetic resonance perfusion imaging, Betof et al showed that exercise-mediated increases in uniformity of blood flow.<sup>10</sup> Additionally, by improving vascular maturity, exercise may decrease interstitial fluid pressure (IFP). Decreasing IFP could improve macromolecular and nanoparticle drug transport (> 1000 MW), because transport of these sized drugs is dominated by the pressure gradient across the vessel wall. The pressure gradient is a consequence of the extent of vascular permeability.<sup>26</sup> Elevated IFP directly impedes large drug and nanoparticle transport from tumor blood vessels into the tumor tissue.

Changes in tumor angiogenesis may influence alternative mechanisms of tumor control, including antitumor immunity. Tumors use a variety of mechanisms to diminish T-cell infiltration and recognition, thereby attempting to evade immune surveillance and contributing to decreased antitumor immunity.<sup>27,28</sup> Interleukin-6 (IL6) can overcome this checkpoint, to facilitate T-cell trafficking into tumors.<sup>28</sup> Lack of pericyte coverage facilitates myeloid-derived suppressor cell (MDSC) trafficking.<sup>29</sup> This contributes further to an immune suppressed phenotype within the tumor microenvironment.

Both local and systemic effects of exercise may contribute to enhanced T-cell infiltration into tumors. First, exercise increases circulating levels of IL6, which can promote upregulation of adhesion molecules on tumor vascular endothelium, thereby promoting T-cell trafficking. Further, exercise causes redistribution of NK cells in an epinephrine and IL-6 dependent manner, and these mature effector NK cells have cytotoxic activity against cancer cells in vitro.<sup>30,31</sup> "Exercise-induced leukocytosis" is the phenomenon by which a single bout of exercise mobilizes vascular, pulmonary, hepatic, and splenic white blood cells into peripheral circulation.<sup>32</sup> A 45 to 60 minute bout of vigorous exercise increases NK cell concentrations 10-fold and CD8<sup>+</sup> T cells approximately 2.5-fold.<sup>33</sup> Dynamic changes in blood pressure, shear force, and epinephrine-mediated stimulation of beta-2-adrenergic receptors on the surface of lymphocytes collaborate to cause leukocyte demargination and circulation.<sup>34-36</sup> This intensity-dependent mobilization occurs in proportion to the expression of beta-2-adrenergic receptors on lymphocytes, with NK cells and CD8<sup>+</sup> T cells responding more strongly than B cells and CD4<sup>+</sup> T cells.<sup>33,37,38</sup>

On a systemic level, immune cells exhibit a characteristic bi-phasic response to exercise, in which acute leukocytosis (during or immediately following exercise treatment) is followed by leukopenia within 1-2 hours after cessation.<sup>37,39,40</sup> Stromberg et al reported that an hour of exercise transiently increased ICAM-1 and VCAM-1 in skeletal muscle vasculature,<sup>41</sup> while Santos et al reported increased neutrophil ICAM-1 and L-selectin expression in marathon runners following the race.<sup>42</sup> Lymphocyte counts

generally nadir below baseline and then normalize within 24 hours.<sup>37,39,40</sup> This has led to speculation of an “open-window” following vigorous exercise in which the individual is particularly susceptible to infection due to immunosuppression.<sup>43</sup> However, recent evidence has forced re-evaluation of this hypothesis. In a rodent model, Kruger et al demonstrated that the leukopenia following exercise reflects a redistribution of T lymphocytes to peripheral tissues (ie, the lung and Peyer’s patches).<sup>36</sup> This is now known as the acute stress/exercise immune-enhancement hypothesis.<sup>44,45</sup> It is unclear at this time how these time-dependent systemic changes in circulating immune cells affect local infiltration of the tumor.

## Tumor Hypoxia

Tumor hypoxia is the result of a discrepancy between oxygen supply and demand. Although proliferating tumor cells have high oxygen requirements, immature tumor vasculature and low microvessel density results in hypoxia.<sup>46</sup> Vaupel et al reviewed 125 clinical studies and determined that tumor tissue generally is poorly oxygenated compared to normal tissue, and that all types of solid tumors include regions with clinically relevant hypoxia ( $< 10$  mmHg  $O_2$ ).<sup>47</sup> Tumor hypoxia is associated with poor radiotherapy outcome<sup>48</sup> as well as an increased propensity toward metastasis. The impact of hypoxia on prognosis has been reported in multiple trials. For example, a multi-center study stratified head and neck cancer patients based on median pretreatment tumor oxygen tension ( $pO_2$ ) and fraction of tumors with  $pO_2$  of  $\leq 2.5$  mmHg ( $HP_{2.5}$ ) and reported significantly worse survival in patients with “more hypoxic” ( $HP_{2.5} > 19\%$ ) tumors compared to those with “less hypoxic” ( $HP_{2.5} \leq 19\%$ ) tumors, regardless of treatment approach.

McCullough et al found that a single bout of aerobic treadmill training increased oxygen delivery 3-fold in rat prostate tumors by decreasing vascular resistance and increasing blood flow to the tumor.<sup>49</sup> Wiggins et al postulated that although exercise redirects blood flow from splanchnic organs to the active skeletal muscles, tumor vessels are uniquely unable to respond to vasoconstrictive signals,<sup>49</sup> and therefore benefit from exercise-induced increases in cardiac output.<sup>50</sup> These physiological effects of exercise reduce tumor hypoxia, as has been shown by multiple researchers.<sup>10,49,51</sup> Betof et al reported that voluntary running throughout tumor development reduced the hypoxic fraction by nearly 50%, and McCullough et al reported that a single bout of treadmill running halved the hypoxic tumor fraction.

Hypoxic cells are 3-fold more resistant to radiation, compared with aerobic cells.<sup>52</sup> Nearly all solid cancers contain some hypoxic cells, and the severity and extent of hypoxia is associated with poor prognosis.<sup>47</sup> A number of different strategies have been attempted to alleviate the impact of hypoxia, including increasing oxygen delivery or reducing oxygen consumption rate. Despite some successes, there is no accepted standard of care for reducing hypoxia.<sup>53</sup>

Against this background, several groups have shown that exercise reduces tumor hypoxia.<sup>10,49,51,54</sup> The chronic effects of exercise in reducing hypoxia may make it an attractive means of increasing tumor radiosensitivity. Our group sought to test the hypothesis that exercise would improve the tumor response to radiation, in preclinical murine models.<sup>51</sup> Indeed, in unpublished data from our group mice bearing either 4T1 mammary or MC38 colorectal carcinomas, exercise prior to and during radiation improved tumor response, characterized by slowed tumor growth and delayed metastasis.<sup>51</sup> The addition of voluntary exercise to fractionated radiation treatment increased time to 4T1 tumor volume quintupling from 5.4 days in sedentary/RT mice to 11.6 days in exercising/RT mice. Importantly, we also showed that at the time that RT was applied, the tumor hypoxic fraction averaged 8.8% in sedentary mice, compared to 2.8% in tumors from exercising mice.

Hypoxia is also a well-known mediator of chemoresistance. The same mechanisms that cause hypoxia, such as low microvessel density and vascular shunts limit the delivery of chemotherapy to the tumor.<sup>26</sup> Another key mechanism of drug resistance is upregulation of the transcription factor HIF-1 and its downstream targets, including key protein and miRNA mediators of proliferation, drug efflux, metabolism, and autophagy.<sup>55</sup> Notably, HIF-1 activation increases expression of *MDR1*, which confers multidrug resistance by increasing drug efflux. For example, targeted downregulation of HIF-1 $\alpha$  showed a dose-dependent increase in cisplatin-mediated apoptosis in oral squamous cell carcinoma cells.<sup>56</sup> Hypoxia may also abrogate the apoptotic effects of chemotherapy.<sup>57</sup> Thus, it is possible that the reduction of tumor hypoxia associated with exercise will be accompanied by improved response to chemotherapy.

An exercise-induced decrease in tumor hypoxia may affect the efficacy of immunotherapy. Infiltration of various immune cells is affected by hypoxic conditions. Hatfield et al showed that CD8<sup>+</sup> tumor infiltrating lymphocytes (TILs) have a decreased presence in hypoxic tumor regions.<sup>58</sup> Hypoxic microenvironments also limit the efficacy of TILs by inhibiting their activation by dendritic cells<sup>59</sup> and increasing tumor expression of PD-L1, a key modulator of the immune checkpoint mechanisms.<sup>60,61</sup> Housing mice in hyperoxic chambers (60% oxygen) increased the numbers of CD8<sup>+</sup> TILs more than 3-fold.<sup>58</sup> The potential clinical implications of this finding are staggering, as Adams et al, postulated that for every 10% increase in CD8<sup>+</sup> TILs would translate to a 19% decrease in patient mortality.<sup>62</sup> Yet the sobering reality is that Hatfield’s findings have limited clinical scope, as patients cannot be sequestered in 60% oxygen environments at the time of tumor development. However, exercise may be an effective way to increase TIL numbers, in part by creating a more favorable, normoxic tumor.

The effects of hypoxia extend beyond TILs. Dendritic cells are suppressed by hypoxia-inducible genes including VEGF.<sup>55</sup> Additionally, tumor-associated macrophages respond to the hypoxia-induced cytokines IL-4 and IL-10 by differentiating into an immunosuppressive M2 phenotype.<sup>63</sup> Indoleamine 2,3-dioxygenase (IDO), a tryptophan

metabolism enzyme expressed by most tumors, is garnering interest for its immunomodulatory role in T cell suppression and tumor tolerance.<sup>64,65</sup> IDO expression correlates with reduced TILs and poor prognosis in numerous cancer types.<sup>66-70</sup> Notably, IDO production by dendritic cells increases when cultured in a hypoxic environment.<sup>71</sup> Thus far, no one has reported the effects of exercise on IDO levels in tumors. However, it is a logical hypothesis that as exercise reduces tumor hypoxia, IDO production may be reduced, thereby reducing immunosuppression. Expanding this hypothesis, exercise may improve tumor response to immunotherapy by removing one of the roadblocks to an effective antitumor immune response.

The combination of radiation and immunotherapy creates the perfect storm of augmenting the immune response's potential and increasing tumor cells' susceptibility to immune cell killing.<sup>72</sup> Immunogenic cell death (ICD) occurs when an injured cell increases presentation of damage associated molecular patterns such as calreticulin and high-mobility group box 1 protein.<sup>73</sup> These changes promote dendritic cell activation and facilitate an antitumor T-cell response. Radiation therapy is a means of inducing ICD in tumor cells. In addition, radiation increases tumor cell expression of MHC Class I<sup>74</sup> and Fas.<sup>75</sup> Together, these changes have been viewed as using the tumor to create an *in situ* vaccine<sup>76</sup> and contribute to abscopal effects, in which the immune response extends beyond the primary irradiated tumor to target distant metastases. However, although the reported radiation-mediated "abscopal effects" against metastatic lesions<sup>77-79</sup> have sparked much excitement, abscopal effects in humans remain rare.<sup>80</sup> Nevertheless, it is worth considering how exercise may further prime the antitumor immune response for an abscopal effect. As detailed above, many of these changes will stem from exercise-mediated modulations in hypoxia (increased T cell infiltration<sup>58</sup>) or changes to tumor vascular maturity (decreased MDSC trafficking.<sup>29</sup>) Hypoxia also reduces tumor MHC class I expression<sup>81</sup>; by removing this blockade, antigen recognition, and ICD may be enhanced.

## Tumor Cell Metabolism

One of the emerging "hallmarks of cancer" is deregulated cellular energetics, or altered metabolism,<sup>82</sup> which influences radiosensitivity.<sup>83-85</sup> Many cancer cells increase glucose consumption rate and preferentially utilize glycolysis over aerobic respiration; this may be due to hypoxia or a high rate of cell proliferation and oxidative stress.<sup>86,87</sup> A downstream consequence of glycolysis is increased lactate concentration within tumors via the Pasteur effect—increased lactate stabilizes HIF-1,<sup>88</sup> which promotes VEGF expression,<sup>88</sup> increases metastasis, and correlates with worse prognosis.<sup>89,90</sup>

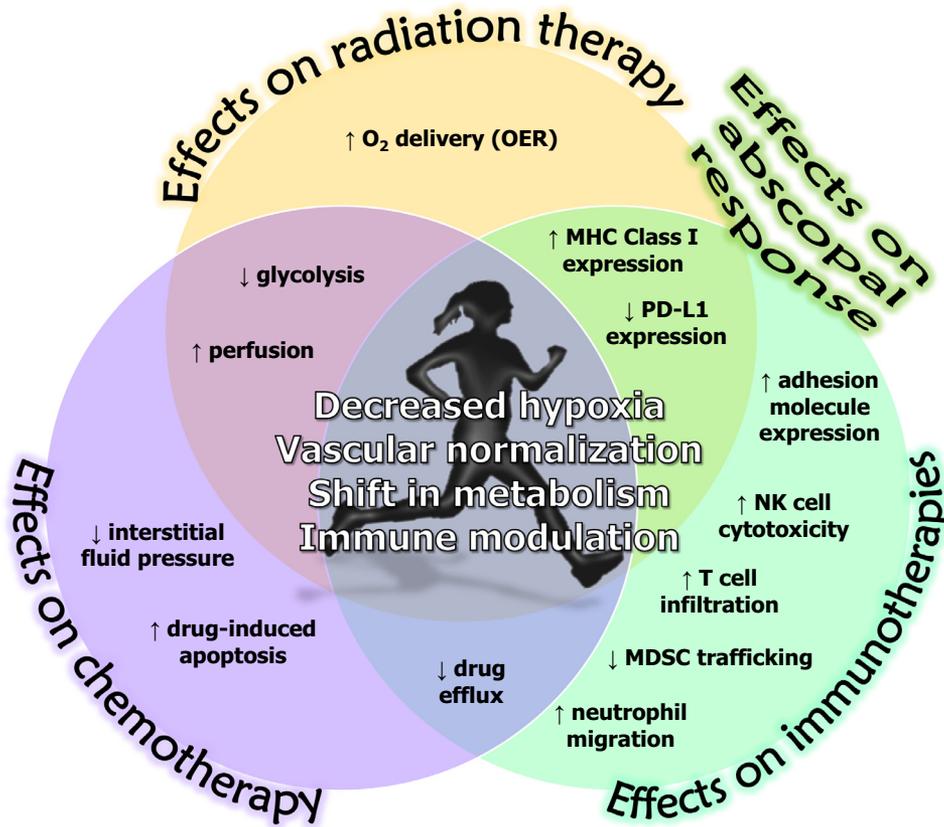
Exercise may regulate metabolic reprogramming of tumor cells. For example, Bacurau et al reported that treadmill running (at 60%  $\text{VO}_2$  max) decreased carcinoma glucose consumption and decreased lactate production.<sup>91,92</sup> Lu et al showed that metabolic responses to exercise were associated

with tumor response to exercise (unpublished data).<sup>93</sup> Specifically, comparison of exercise-responsive and nonresponsive patient-derived colorectal xenograft (PDX) models revealed that phosphocreatinine metabolism was predictive of tumor growth delay. Tumors that showed growth delay in response to exercise had significantly lower phosphocreatinine compared to sedentary controls, whereas there was no difference in phosphocreatinine levels between sedentary or exercising mice with tumors, whose growth was not affected by exercise. Metabolomics data reported by Glass et al showed that nucleotide metabolism, which correlates with increased cell proliferation, was increased in a murine tumor cell line that showed accelerated tumor growth following exercise, compared to an exercise-responsive tumor that showed growth delay following exercise.<sup>94</sup>

Our group also found preliminary evidence that exercise increased long chain acyl carnitines, consistent with fatty acid metabolism, elongation and oxidation (Ashcraft et al, unpublished data). The effects of fatty acid oxidation (FAO) vs glycolysis on oxygen consumption rate relate to the respiratory exchange ratio (RER), or the ratio of ATP molecules gleaned per molecule  $\text{O}_2$  consumed. While the RER for glycolysis is 6.3 ATP/ $\text{O}_2$ , the RER for FAO is 5.6 ATP/ $\text{O}_2$ , indicating that equivalent energy is produced by FAO, with ~10% less oxygen consumption. Mathematical modeling by Secomb et al predicted that a 30% decrease in oxygen consumption rate in the R3230Ac mammary carcinoma would completely abolish tumor hypoxia<sup>95</sup>; thus even a modest decrease in oxygen consumption rate induced by exercise could prove important in dictating radiotherapy response. It is also important to note that hypoxia suppresses FAO by regulating medium and long-chain acyl carnitines through the hypoxia-inducible transcription factor, HIF-1,<sup>96</sup> meaning that the cascade of events leading to improved tumor oxygenation is likely bidirectional. Tumors that increase HIF-1 expression following exercise also demonstrate accelerated tumor growth compared to tumors that do not express HIF-1.<sup>94</sup> The results with metabolism suggest that exercise can exert a diversity of effects on tumor metabolism and radiosensitivity. If exercise pushes metabolism toward aerobic metabolism, radiosensitivity may be improved.<sup>97,98</sup> If the opposite occurs, namely a push toward anaerobic metabolism radiosensitivity will decrease.

The observed exercise-mediated improvement in chemotherapy efficacy<sup>10,25</sup> may have metabolic underpinnings, possibly, by reducing the tumors' reliance on glycolysis.<sup>91,92</sup> If exercise does reduce tumor glucose reliance, resultant metabolic changes could improve efficacy of cytotoxic agents. Pharmacologically inhibiting glycolysis (via either 2-deoxyglucose or 3-bromopyruvate) improved etoposide efficacy against lymphoma,<sup>99</sup> rapamycin efficacy against neuroblastoma,<sup>100</sup> and tamoxifen efficacy against mammary carcinoma.<sup>101</sup> Cantelmo et al showed that glycolysis inhibition within tumor vessel endothelial cells normalized tumor vasculature and improved perfusion, thereby increasing cisplatin delivery causing inhibition of tumor growth.<sup>102</sup>

Finally, we can link shifts in tumor metabolism to potential improvement in immunotherapy. Immune responses



**Figure 2** Exercise oncology studies have demonstrated that exercise modulates both the tumor microenvironment and the robustness of the anti-tumor immune response. Preclinical studies have already reported that in these changes are sufficient for exercise to play an anti-tumor role in tumor incidence, progression and metastasis when administered as a monotherapy. In this paper, we have described how the exercise-mediated changes may potentiate tumor sensitivity to radiation, chemotherapy or immunotherapy. Furthermore, we hypothesize that some changes may increase potency of abscopal responses, when radiation and immunotherapy are applied concurrently.

become suboptimal, when they develop in acidic microenvironments (reviewed in<sup>103</sup>). Deficient immune responses begin with poor immune cell infiltration into acidic tumor environments. In vitro lymphocyte<sup>104</sup> and neutrophil<sup>105</sup> migration are reduced as their environment becomes increasingly acidic. In addition, cells that are able to gain entry into the tumor are less effective. Increased lactate concentrations impede NK cell function in 2 ways<sup>106</sup>. First, NK cells cytotoxic potential is decreased under acidic conditions. Second, exogenous lactate promotes *ex vivo* differentiation of MDSCs. Lactate increases tumor cell PD-L1 expression.<sup>107</sup> Inhibiting the lactate/PD-L1 cascade within the tumor augments cytotoxic T cell numbers.<sup>108</sup> Currently, there are no reports on how exercise affects tumor pH. However, given that lactate levels are decreased with exercise, it is likely that pH levels are reduced.

## Concluding Remarks

Oxidative stress is present in untreated tumors, and may increase following some treatment modalities. Oxidative stress, along with closely associated tumor hypoxia and acidic microenvironments, contributes to tumor

aggressiveness and cancer fatigue. While pharmaceutical means of reducing oxidative stress have been pursued, exercise may provide a nonpharmacological therapy of regulating oxidative stress thereby alleviating these factors. Additionally, emerging evidence shows that exercise exerts other effects on tumor physiology including alterations in hypoxia, vascular normalization, metabolic reprogramming, and immune cell mobilization (summarized in Fig. 2). We and others have begun to show that these changes translate to improved response to tumor therapy. Clinical studies that use functional imaging to monitor tumor hypoxia and perfusion in exercising vs sedentary patients would be helpful in whether changes might influence response to therapy. Although, additional work is needed to optimize exercise prescriptions (ie, frequency, duration, and intensity), current studies suggest that human trials should be considered. For example, exercise could be evaluated as a means to improve treatment response to established cancer treatment modalities.

## Conflict of Interest

The authors declare no potential conflicts of interest.

## References

- Friedenreich CM, Orenstein MR. Physical activity and cancer prevention: Etiologic evidence and biological mechanisms. *J Nutr* 132(11 Suppl):3456S-3464S, 2002
- Friedenreich CM, Woolcott CG, McTiernan A, et al: Alberta physical activity and breast cancer prevention trial: Sex hormone changes in a year-long exercise intervention among postmenopausal women. *J Clin Oncol* 28:1458-1466, 2010
- Scott JM, Zabor EC, Schwitzer E, et al: Efficacy of exercise therapy on cardiorespiratory fitness in patients with cancer: A systematic review and meta-analysis. *J Clin Oncol* 36:2297-2305, 2018
- Loughney L, West MA, Kemp GJ, et al: Exercise intervention in people with cancer undergoing neoadjuvant cancer treatment and surgery: A systematic review. *Eur J Surg Oncol* 42:28-38, 2016
- Ashcraft KA, Peace RM, Betof AS, et al: Efficacy and mechanisms of aerobic exercise on cancer initiation, progression, and metastasis: A critical systematic review of in vivo preclinical data. *Cancer Res* 76:4032-4050, 2016
- Lyon DE, Cohen R, Chen H, et al: Relationship of systemic cytokine concentrations to cognitive function over two years in women with early stage breast cancer. *J Neuroimmunol* 301:74-82, 2016
- Starkweather A, Kelly DL, Thacker L, et al: Relationships among psychoneurological symptoms and levels of C-reactive protein over 2 years in women with early-stage breast cancer. *Support Care Cancer* 25:167-176, 2017
- Repka CP, Hayward R. Oxidative stress and fitness changes in cancer patients after exercise training. *Med Sci Sports Exerc* 48:607-614, 2016
- Repka CP, Hayward R. Effects of an exercise intervention on cancer-related fatigue and its relationship to markers of oxidative stress. *Integr Cancer Ther* 17:503-510, 2018
- Betof AS, Lascola CD, Weitzel D, et al: Modulation of murine breast tumor vascularity, hypoxia and chemotherapeutic response by exercise. *J Natl Cancer Inst* 2015, 107(5).
- Du J, Cieslak JA, Welsh JL, et al: Pharmacological ascorbate radiosensitizes pancreatic cancer. *Cancer Res* 75:3314-3326, 2015
- Krzesiak A, Delpech N, Sebille S. Structural, contractile and electrophysiological adaptations of cardiomyocytes to chronic exercise. *Adv Exp Med Biol* 999:75-90, 2017
- Zhang L, Conejo-Garcia JR, Yang N, et al: Different effects of glucose starvation on expression and stability of VEGF mRNA isoforms in murine ovarian cancer cells. *Biochem Biophys Res Commun* 292:860-868, 2002
- Goel S, Wong AH, Jain RK. Vascular normalization as a therapeutic strategy for malignant and nonmalignant disease. *Cold Spring Harb Perspect Med* 2012, 2(3):a006486
- Dewhirst MW, Cao Y, Moeller B. Cycling hypoxia and free radicals regulate angiogenesis and radiotherapy response. *Nat Rev Cancer* 8:425-437, 2008
- Cooke VG, LeBleu VS, Keskin D, et al: Pericyte depletion results in hypoxia-associated epithelial-to-mesenchymal transition and metastasis mediated by met signaling pathway. *Cancer Cell* 21:66-81, 2012
- Vasudev NS, Reynolds AR. Anti-angiogenic therapy for cancer: current progress, unresolved questions and future directions. *Angiogenesis* 17:471-494, 2014
- Jain RK. Antiangiogenesis strategies revisited: From starving tumors to alleviating hypoxia. *Cancer Cell* 26:605-622, 2014
- Paez-Ribes M, Allen E, Hudock J, et al: Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. *Cancer Cell* 15:220-231, 2009
- Shalamzari SA, Agha-Alinejad H, Alizadeh S, et al: The effect of exercise training on the level of tissue IL-6 and vascular endothelial growth factor in breast cancer bearing mice. *Iranian J Basic Med Sci* 17:231-258, 2014
- Zhu Z, Jiang W, Sells JL, et al: Effect of nonmotorized wheel running on mammary carcinogenesis: circulating biomarkers, cellular processes, and molecular mechanisms in rats. *Cancer Epidemiol Biomarkers Prev* 17:1920-1929, 2008
- Jones LW, Antonelli J, Masko EM, et al: Exercise modulation of the host-tumor interaction in an orthotopic model of murine prostate cancer. *J Appl Physiol* 113:263-272, 2012
- Lin B, Song X, Yang D, et al: Anlotinib inhibits angiogenesis via suppressing the activation of VEGFR2, PDGFRbeta and FGFR1. *Gene* 654:77-86, 2018
- Jones LW, Viglianti BL, Tashjian JA, et al: Effect of aerobic exercise on tumor physiology in an animal model of human breast cancer. *J Appl Physiol* 108:343-348, 2010
- Schadler KL, Thomas NJ, Galie PA, et al: Tumor vessel normalization after aerobic exercise enhances chemotherapeutic efficacy. *Oncotarget* 7:65429-65440, 2016
- Dewhirst MW, Secomb TW. Transport of drugs from blood vessels to tumour tissue. *Nat Rev Cancer* 17:738-750, 2017
- Wu NZ, Klitzman B, Dodge R, et al: Diminished leukocyte-endothelium interaction in tumor microvessels. *Cancer Res* 52:4265-4268, 1992
- Fisher DT, Chen Q, Skitzki JJ, et al: IL-6 trans-signaling licenses mouse and human tumor microvascular gateways for trafficking of cytotoxic T cells. *J Clin Invest* 121:3846-3859, 2011
- Hong J, Tobin NP, Rundqvist H, et al: Role of tumor pericytes in the recruitment of myeloid-derived suppressor cells. *J Natl Cancer Inst* 2015, 107(10).
- Bigley AB, Rezvani K, Chew C, et al: Acute exercise preferentially redeploys NK-cells with a highly-differentiated phenotype and augments cytotoxicity against lymphoma and multiple myeloma target cells. *Brain Behav Immun* 39:160-171, 2014
- Pedersen L, Idorn M, Olofsson GH, et al: Voluntary running suppresses tumor growth through epinephrine- and IL-6-dependent NK cell mobilization and redistribution. *Cell Metab* 23:554-562, 2016
- Simpson RJ, Kunz H, Agha N, et al: Exercise and the regulation of immune functions. *Prog Mol Biol Transl Sci* 135:355-380, 2015
- Campbell JP, Riddell NE, Burns VE, et al: Acute exercise mobilises CD8+ T lymphocytes exhibiting an effector-memory phenotype. *Brain Behav Immun* 23:767-775, 2009
- Benschop RJ, Nijkamp FP, Ballieux RE, et al: The effects of beta-adrenoceptor stimulation on adhesion of human natural killer cells to cultured endothelium. *BJ Pharmacol* 113:1311-1316, 1994
- Dimitrov S, Lange T, Born J. Selective mobilization of cytotoxic leukocytes by epinephrine. *J Immunol* 184:503-511, 2010
- Kruger K, Lechtermann A, Fobker M, et al: Exercise-induced redistribution of T lymphocytes is regulated by adrenergic mechanisms. *Brain Behavior Immun* 22:324-338, 2008
- Kruger K, Alack K, Ringseis R, et al: Apoptosis of T-cell subsets after acute high-intensity interval exercise. *Med Sci Sports Exerc* 48:2021-2029, 2016
- Turner JE, Spielmann G, Wadley AJ, et al: Exercise-induced B cell mobilisation: Preliminary evidence for an influx of immature cells into the bloodstream. *Physiol Behav* 164:376-382, 2016
- Shek PN, Sabiston BH, Buguet A, et al: Strenuous exercise and immunological changes: a multiple-time-point analysis of leukocyte subsets, CD4/CD8 ratio, immunoglobulin production and NK cell response. *Int J Sports Med* 16:466-474, 1995
- Shinkai S, Shore S, Shek PN, et al: Acute exercise and immune function. Relationship between lymphocyte activity and changes in subset counts. *Int J Sports Med* 13:452-461, 1992
- Stromberg A, Rullman E, Jansson E, et al: Exercise-induced upregulation of endothelial adhesion molecules in human skeletal muscle and number of circulating cells with remodeling properties. *J Appl Physiol* 122:1145-1154, 2017
- Santos VC, Sierra AP, Oliveira R, et al: Marathon race affects neutrophil surface molecules: Role of Inflammatory Mediator. *PLoS one* 2016, 11(12):e0166687
- Peake JM, Neubauer O, Walsh NP, et al: Recovery of the immune system after exercise. *J Appl Physiol* 122:1077-1087, 2017
- Campbell JP, Turner JE. Debunking the myth of exercise-induced immune suppression: Redefining the impact of exercise on immunological health across the lifespan. *Front Immunol* 9:648, 2018
- Dhabhar FS. Effects of stress on immune function: The good, the bad, and the beautiful. *Immunol Res* 58:193-210, 2014
- Dewhirst MW. Relationships between cycling hypoxia, HIF-1, angiogenesis and oxidative stress. *Radiat Res* 172:653-665, 2009

47. Vaupel P, Hockel M, Mayer A. Detection and characterization of tumor hypoxia using pO<sub>2</sub> histography. *Antioxid Redox Signal* 9:1221-1235, 2007
48. Nordmark M, Bentzen SM, Rudat V, et al: Prognostic value of tumor oxygenation in 397 head and neck tumors after primary radiation therapy. An international multi-center study. *Radiother Oncol* 77:18-24, 2005
49. McCullough DJ, Stabley JN, Siemann DW, et al: Modulation of blood flow, hypoxia, and vascular function in orthotopic prostate tumors during exercise. *J Natl Cancer Inst* 2014, 106(4): dju036
50. Wiggins JM, Opoku-Acheampong AB, Baumfalk DR, et al: Exercise and the tumor microenvironment: Potential therapeutic implications. *Exerc Sport Sci Rev* 46:56-64, 2018
51. Ashcraft K.A., Roy Choudhury K., Zessin A.S., et al: Physical activity increases tumor response to radiation and reduces spontaneous metastasis manuscript in preparation.
52. Brizel DM, Dodge RK, Clough RW, et al: Oxygenation of head and neck cancer: changes during radiotherapy and impact on treatment outcome. *Radiother Oncol* 53:113-117, 1999
53. Overgaard J, Horsman MR. Modification of hypoxia-induced radioresistance in tumors by the use of oxygen and sensitizers. *Semin Radiation Oncol* 6:10-21, 1996
54. Buss LA, Dachs GU. Voluntary exercise slows breast tumor establishment and reduces tumor hypoxia in Apo(E-/-) Mice. *J Appl Physiol* 124:938-949, 2018
55. Manoochehri Khoshinani H, Afshar S, et al: Hypoxia: A double-edged sword in cancer therapy. *Cancer Invest* 34:536-545, 2016
56. Sasabe E, Zhou X, Li DC, et al: The involvement of hypoxia-inducible factor-1 alpha in the susceptibility to gamma-rays and chemotherapeutic drugs of oral squamous cell carcinoma cells. *Int J Cancer* 120:268-277, 2007
57. Rohwer N, Cramer T. Hypoxia-mediated drug resistance: novel insights on the functional interaction of HIFs and cell death pathways. *Drug Resist* 14:191-201, 2011
58. Hatfield SM, Kjaergaard J, Lukashev D, et al: Immunological mechanisms of the antitumor effects of supplemental oxygenation. *Sc Transl Med* 2015, 7(277):277ra230
59. Elia AR, Cappello P, Puppo M. Human dendritic cells differentiated in hypoxia down-modulate antigen uptake and change their chemokine expression profile. *J Leukoc Biol* 84:1472-1482, 2008
60. Barsoum IB, Smallwood CA, Siemens DR, et al: A mechanism of hypoxia-mediated escape from adaptive immunity in cancer cells. *Cancer Res* 74:665-674, 2014
61. Ruf M, Moch H, Schraml P. PD-L1 expression is regulated by hypoxia inducible factor in clear cell renal cell carcinoma. *Int J Cancer* 139:396-403, 2016
62. Adams S, Novik Y, Oratz R, et al: Clinical trial evidence of the antitumor activity of topical imiquimod for breast cancer skin metastases. *J Clin Oncol* 32:3204-3205, 2014
63. Murdoch C, Giannoudis A, Lewis CE. Mechanisms regulating the recruitment of macrophages into hypoxic areas of tumors and other ischemic tissues. *Blood* 104:2224-2234, 2004
64. Friberg M, Jennings R, Alsarraj M, et al: Indoleamine 2,3-dioxygenase contributes to tumor cell evasion of T cell-mediated rejection. *Int J Cancer* 101:151-155, 2002
65. Uyttenhove C, Pilotte L, Theate I, et al: Evidence for a tumoral immune resistance mechanism based on tryptophan degradation by indoleamine 2,3-dioxygenase. *Nat Med* 9:1269-1274, 2003
66. Brandacher G, Perathoner A, Ladurner R, et al: Prognostic value of indoleamine 2,3-dioxygenase expression in colorectal cancer: effect on tumor-infiltrating T cells. *Clin Cancer Res* 12:1144-1151, 2006
67. Kiyozumi Y, Baba Y, Okadome K, et al: IDO1 expression is associated with immune tolerance and poor prognosis in patients with surgically resected esophageal cancer. *Ann Surg* 2018
68. Wainwright DA, Balyasnikova IV, Chang AL, et al: IDO expression in brain tumors increases the recruitment of regulatory T cells and negatively impacts survival. *Clin Cancer Res* 18:6110-6121, 2012
69. Zhang G, Liu WL, Zhang L, et al: Involvement of indoleamine 2,3-dioxygenase in impairing tumor-infiltrating CD8 T-cell functions in esophageal squamous cell carcinoma. *Clin Dev Immunol* 2011:384726, 2011
70. Yu J, Sun J, Wang SE, et al: Upregulated expression of indoleamine 2,3-dioxygenase in primary breast cancer correlates with increase of infiltrated regulatory T cells in situ and lymph node metastasis. *Clin Dev Immunol* 2011. 469135, 2011
71. Song X, Zhang Y, Zhang L, et al: Hypoxia enhances indoleamine 2,3-dioxygenase production in dendritic cells. *Oncotarget* 9:11572-11580, 2018
72. Vanpouille-Box C, Pilonis KA, Wennerberg E, et al: In situ vaccination by radiotherapy to improve responses to anti-CTLA-4 treatment. *Vaccine* 33:7415-7422, 2015
73. Wu CY, Yang LH, Yang HY, et al: Enhanced cancer radiotherapy through immunosuppressive stromal cell destruction in tumors. *Clin Cancer Res* 20:644-657, 2014
74. Wang X, Schoenhals JE, Li A, et al: Suppression of type I IFN signaling in tumors mediates resistance to anti-PD-1 treatment that can be overcome by radiotherapy. *Cancer Res* 77:839-850, 2017
75. Chakraborty M, Abrams SI, Camphausen K, et al: Irradiation of tumor cells up-regulates Fas and enhances CTL lytic activity and CTL adoptive immunotherapy. *J Immunol* 170:6338-6347, 2003
76. Formenti SC, Demaria S. Combining radiotherapy and cancer immunotherapy: a paradigm shift. *J Natl Cancer Inst* 105:256-265, 2013
77. Demaria S, Ng B, Devitt ML, et al: Ionizing radiation inhibition of distant untreated tumors (abscopal effect) is immune mediated. *Int J Radiat Oncol Biol Phys* 58:862-870, 2004
78. Dewan MZ, Galloway AE, Kawashima N, et al: Fractionated but not single-dose radiotherapy induces an immune-mediated abscopal effect when combined with anti-CTLA-4 antibody. *Clin Cancer Res* 15:5379-5388, 2009
79. Postow MA, Callahan MK, Barker CA, et al: Immunologic correlates of the abscopal effect in a patient with melanoma. *N Engl J Med* 366:925-931, 2012
80. Brix N, Tiefenthaler A, Anders H, et al: Abscopal, immunological effects of radiotherapy: Narrowing the gap between clinical and pre-clinical experiences. *Immunol Rev* 280:249-279, 2017
81. Sethumadhavan S, Silva M, Philbrook P, et al: Hypoxia and hypoxia-inducible factor (HIF) downregulate antigen-presenting MHC class I molecules limiting tumor cell recognition by T cells. *PLoS One* 2017, 12(11):e0187314
82. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. *Cell* 144:646-674, 2011
83. Allen KT, Chin-Sinex H, DeLuca T, et al: Dichloroacetate alters Warburg metabolism, inhibits cell growth, and increases the X-ray sensitivity of human A549 and H1299 NSC lung cancer cells. *Free Radic Biol Med* 89:263-273, 2015
84. Bol V, Bol A, Bouzin C, et al: Reprogramming of tumor metabolism by targeting mitochondria improves tumor response to irradiation. *Acta Oncol* 54:266-274, 2015
85. Mims J, Bansal N, Bharadwaj MS, et al: Energy metabolism in a matched model of radiation resistance for head and neck squamous cell cancer. *Radiat Res* 183:291-304, 2015
86. Dang CV, Semenza GL. Oncogenic alterations of metabolism. *Trends Biochem Sci* 24:68-72, 1999
87. Gatenby RA, Gillies RJ. Why do cancers have high aerobic glycolysis. *Nat Rev Cancer* 4:891-899, 2004
88. Sonveaux P, Copetti T, De Saedeleer CJ, et al: Targeting the lactate transporter MCT1 in endothelial cells inhibits lactate-induced HIF-1 activation and tumor angiogenesis. *PLoS One* 7:e33418, 2012
89. Walenta S, Wetterling M, Lehrke M, et al: High lactate levels predict likelihood of metastases, tumor recurrence, and restricted patient survival in human cervical cancers. *Cancer Res* 60:916-921, 2000
90. Brizel DM, Schroeder T, Scher RL, et al: Elevated tumor lactate concentrations predict for an increased risk of metastases in head-and-neck cancer. *Int J Radiat Oncol Biol Phys* 51:349-353, 2001
91. Bacurau RF, Belmonte MA, Seelaender MC, et al: Effect of a moderate intensity exercise training protocol on the metabolism of macrophages and lymphocytes of tumour-bearing rats. *Cell Biochem Funct* 18:249-258, 2000
92. Bacurau AV, Belmonte MA, Navarro F, et al: Effect of a high-intensity exercise training on the metabolism and function of macrophages and

- lymphocytes of walker 256 tumor bearing rats. *Exp Biol Med (Maywood)* 232:1289-1299, 2007
93. Lu M., Sanderson S.M., Zessin A.S., et al: Manuscript under review.
  94. Glass OK, Inman BA, Broadwater G, et al: Effect of aerobic training on the host systemic milieu in patients with solid tumours: an exploratory correlative study. *Br J Cancer* 112:825-831, 2015
  95. Secomb TW, Hsu R, Ong ET, et al: Analysis of the effects of oxygen supply and demand on hypoxic fraction in tumors. *Acta Oncol* 34:313-316, 1995
  96. Huang D, Li T, Li X, et al: HIF-1-mediated suppression of acyl-CoA dehydrogenases and fatty acid oxidation is critical for cancer progression. *Cell Rep* 8:1930-1942, 2014
  97. Klement RJ. Fasting, fats, and physics: Combining ketogenic and radiation therapy against cancer. *Complement Med Res* 25:102-113, 2018
  98. Li L, Liu H, Du L, et al: MiR-449a suppresses LDHA-mediated glycolysis to enhance the sensitivity of non-small cell lung cancer cells to ionizing radiation. *Oncol Res* 2017
  99. Beneteau M, Zunino B, Jacquin MA, et al: Combination of glycolysis inhibition with chemotherapy results in an antitumor immune response. *Proc Natl Acad Sci U S A* 109:20071-20076, 2012
  100. Levy AG, Zage PE, Akers LJ, et al: The combination of the novel glycolysis inhibitor 3-BrOP and rapamycin is effective against neuroblastoma. *Invest New Drugs* 30:191-199, 2012
  101. Attia YM, El-Abhar HS, Al Marzabani MM, et al: Targeting glycolysis by 3-bromopyruvate improves tamoxifen cytotoxicity of breast cancer cell lines. *BMC Cancer* 15:838, 2015
  102. Cantelmo AR, Conradi LC, Brajic A, et al: Inhibition of the glycolytic activator PFKFB3 in endothelium induces tumor vessel normalization, impairs metastasis, and improves chemotherapy. *Cancer Cell* 30:968-985, 2016
  103. Lardner A. The effects of extracellular pH on immune function. *J Leukocyte Biol* 69:522-530, 2001
  104. Ratner S. Motility of IL-2-stimulated lymphocytes in neutral and acidified extracellular matrix. *Cell Immunol* 139:399-410, 1992
  105. Rotstein OD, Fiegel VD, Simmons RL, et al: The deleterious effect of reduced pH and hypoxia on neutrophil migration in vitro. *J Surg Res* 45:298-303, 1988
  106. Husain Z, Huang Y, Seth P, et al: Tumor-derived lactate modifies antitumor immune response: effect on myeloid-derived suppressor cells and NK cells. *J Immunol* 191:1486-1495, 2013
  107. Feng J, Yang H, Zhang Y, et al: Tumor cell-derived lactate induces TAZ-dependent upregulation of PD-L1 through GPR81 in human lung cancer cells. *Oncogene* 36:5829-5839, 2017
  108. Seth P, Csizmadia E, Hedblom A, et al: Deletion of lactate dehydrogenase-A in myeloid cells triggers antitumor immunity. *Cancer Res* 77:3632-3643, 2017