



Staphylococcus aureus nasal carriage among homeless population in Lisbon, Portugal

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Abstract

Methicillin-resistant *Staphylococcus aureus* (MRSA) nasal carriage is a major risk factor for infection, namely among populations in the community with inherent prompting factors, such as the homeless. In Portugal, there are no data on *S. aureus*/MRSA nasal carriage among the homeless community. A total of 84 homeless individuals living in Lisbon (34 with no permanent address and 50 living in shelter) were nasally screened for *S. aureus*/MRSA. All isolates were characterized to determine antimicrobial susceptibility and clonal type. A total of 43 (51.2%) *S. aureus* carriers were identified, including a single individual colonized with MRSA (1.2%). *S. aureus* carriage rate was higher among individuals with no permanent address (58.8% versus 46%), younger (45.7 ± 12.7 versus 52.5 ± 10.8 years), and with diagnosis of asthma (9% versus 0%). The single MRSA belonged to the EMRSA-15 clone (PFGE D, ST15-SCCmec IVh, and *spa* type t790). Almost half of the methicillin-susceptible *S. aureus* (MSSA) isolates (41.9%, $n = 18$) belonged to two major clones, ST398-t1451 ($n = 13$) and ST30-t399/t11980/t12808 associated with PFGE I ($n = 5$). A high proportion of isolates showed non-susceptibility to mupirocin (64%), erythromycin (45%), and fusidic acid (20%) and induced resistance to clindamycin (39%). None of the isolates harboured PVL. Our results suggest that the homeless population of Lisbon does not constitute a reservoir of MRSA in the community, but harbour the highly transmissible ST398-t1451 MSSA lineage.

Keywords MRSA · Nasal carriage · Homeless · Portugal

Introduction

Staphylococcus aureus is not only an ubiquitous colonizer of the human anterior nares and skin, but also a major pathogen,

remaining a leading cause of human infections worldwide, in both hospitals and community largely due to its methicillin-resistant form (MRSA) [1, 2].

Homelessness is one of the main societal problems and an important challenge across many countries. A Social Protection report from the European Union showed that in 2016, 23.5% of Europeans were at risk of poverty or social exclusion [3]. Data from Portugal were similar to the European mean, with 25.1% of people at risk of poverty, namely in metropolitan areas of Lisbon and Oporto [3]. In the capital (Lisbon), it is estimated that 852 individuals live without physical accommodation, and are described as being absolutely homeless [4].

Compared with the general population, homeless people are at higher risk for morbidity and mortality, with higher rates of chronic physical health conditions, including almost all type of infections that frequently lead them to emergency services [5]. Moreover, individuals who are homeless were reported to have an increased risk for MRSA colonization and subsequent infection, since they share crowded and often unsanitary living conditions, typically manifest poor underlying

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health such as HIV infection and tuberculosis or diabetes, and have limited access to hygiene facilities and healthcare services [6–8].

Although the burden caused by MRSA infections is widely recognized and its prevention and control have been identified as public health priorities in the European Union, the situation in Portugal remains troublesome: (i) in 2017, Portugal showed a MRSA nosocomial prevalence of 39.2% that largely exceeds the European mean (16.9%) [9]; (ii) the first European vancomycin-resistant *S. aureus* (VRSA) was isolated in Portugal in 2013 [10]; (iii) prevalent MRSA lineages in Portuguese hospitals have been detected in the community [11, 12]. However, there are no studies on MRSA carriage among homeless people in Portugal, which may constitute a MRSA reservoir in the community. Furthermore, people living in shelters in Lisbon have shown higher significant prevalence of respiratory infections and obstructive lung disease. Homeless shelters may facilitate virus transmission and respiratory infections [13]. The contribution of *S. aureus* carriage to these adverse outcomes is uncertain.

The aim of the present study was to determine MRSA colonization rates among the homeless in Lisbon, determine major risk factors for *S. aureus* carriage, and characterize *S. aureus* clonal population, antibiotic resistance, and virulence.

Methods

Homeless community setting

For the present study, two groups of homeless people were considered accordingly to the European Observatory for Homelessness [14, 15]:

1. individuals living in a roofless condition or in secure housing with no permanent address, in the public space or in a precarious place (from now on referred as “with no permanent address”);
2. people living without a house, being in temporary accommodation on homeless shelters designed for that purpose, which promotes their reintegration (from now on referred as “living in shelters”).

A total of 84 individuals, meeting the eligibility criteria (\geq 18 years old, living without facilities for cooking, and with a nasal mucosa without signs of injury) and representing 10.3% of the entire homeless population identified in Lisbon [4], were included in the study (Table 1). The protocol was approved by the Ethics Committee of Escola Superior de Saúde da Cruz Vermelha Portuguesa and a written informed consent was obtained from all participants.

Nasal screening and bacterial isolates

Sampling was performed by trained doctors and nurses, between November 2016 and January 2018. Samples were taken by swabbing both nares of each individual with a sterile cotton swab, which was stored in Stuart transport medium until processing in the laboratory as previously described [16]. MRSA was confirmed by PCR amplification of the *nuc* and *spa* genes for species identification [17, 18], and the detection of the *mecA* gene for methicillin resistance [19].

Phenotypic and molecular characterization

Antimicrobial susceptibility testing was performed by the disk diffusion method, according to the European Committee on Antimicrobial Susceptibility Testing (EUCAST_ <http://www.eucast.org/>), for ceftiofuran, ciprofloxacin, chloramphenicol, clindamycin, erythromycin, fusidic acid, gentamicin, linezolid, mupirocin, penicillin, quinupristin-dalfopristin, rifampin, teicoplanin, tetracycline, and trimethoprim-sulfamethoxazole (SXT). Strain *S. aureus* ATCC29213 was used as quality control. Vancomycin minimum inhibitory concentration (MIC) was determined by Etest.

The isolates were characterized by pulsed-field gel electrophoresis (PFGE) [20], *spa* typing [18], multilocus sequence typing (MLST) [21], SCC_{mec} typing [22, 23], and by detection of the Panton Valentine leucocidine (PVL) gene [24], as previously described.

Statistical analysis

Categorical variables were compared using the χ^2 or Fisher's exact test when appropriate, while continuous variables were tested using Student's *t* test, through the SPSS Statistics software version 21 (IBM Portugal, Lisbon, Portugal). *p* values of ≤ 0.05 were considered statistically significant.

Results

Prevalence of *S. aureus* and methicillin resistance

Eighty-four homeless individuals (34 with no permanent address and 50 living in shelters) were nasal screened for *S. aureus*/MRSA, out of which 43 (51.2%) were *S. aureus* carriers. A single individual with no permanent address was colonized by an MRSA (1.2%). The prevalence of *S. aureus* carriage in individuals with no permanent address was higher than among people living in shelters ($n = 20$, 58.8% versus $n = 23$, 46%) although the difference was not statistically significant ($p = 0.2739$).

Table 1 Distribution of individuals screened by geographic region and homeless condition

Geographic location	Individuals (No./total no.)	Homeless condition (no.) ^a	
		No permanent address	Shelter
Northeast	9/177	8	1
Centre	5/113	5	–
South	24/82	21	3
Shelter 1	31/210	–	31
Shelter 2	15/67	–	15
Total	84/818	34/431	50/387
%	10.3%	7.9%	12.9%

^a Four individuals, sampled at Northeast (Gare 2, $n = 1$) and South (Gare 1, $n = 3$), have been attending homeless shelters for night accommodation and therefore were considered as shelter attendees, but identification of the attended shelter was not available

For homeless people with no permanent address, the highest *S. aureus* prevalence (66.6%; $n = 14$) was detected among people from Gare 1, followed by city hall (60%, $n = 3$) and by Gare 2 (37.5%, $n = 3$). The MRSA carrier was detected in Gare 1. Similar *S. aureus* carriage rates were detected among people living in the two shelters sampled (Shelter 1, $n = 12$, 38.7% versus Shelter 2, $n = 7$, 46.6%, $p = 0.7515$).

A single individual was colonized with two different *S. aureus* isolates, therefore a total of 44 *S. aureus* isolates (20 from homeless with no permanent address and 24 from shelters) were recovered for further characterization.

Subject demographics and risk factors for MRSA carriage

S. aureus carriage was higher among younger individuals (45.7 ± 12.7 versus 52.5 ± 10.8 years, $p = 0.009$) and people with diagnosis of asthma (9% versus 0%, $p = 0.04$). However, no significant difference was detected neither in *S. aureus* carriage between male and female individuals (50.8%, $n = 32$ and 52.4%, $n = 11$ respectively, $p = 0.90$), nor in other conditions commonly associated with increased MRSA carriage rates (Table 2): smoking ($p = 0.28$), alcohol ($p = 0.12$), and illicit drug use ($p = 0.19$).

Antibiotherapy in the last 6 months preceding the screening showed to be a protective factor for *S. aureus* colonization in this population (0% versus 10%, $p = 0.04$).

Antimicrobial resistance

The *mecA* gene was detected in a single *S. aureus* isolate showing resistance to beta-lactams, rifampicin, erythromycin, and induced resistance to clindamycin.

The MSSA isolates showed a non-multiresistant profile (Table 3), with 29.5% ($n = 13$) of the isolates presenting penicillin resistance only, and 45% ($n = 20$) resistance

to erythromycin, out of which 85% ($n = 17$) showed also induced resistance to clindamycin. Moreover, 20% ($n = 9$) of the isolates showed resistance to fusidic acid and less than 10% showed resistance to ciprofloxacin (7%, $n = 3$), chloramphenicol (5%, $n = 2$), and tetracycline (5%, $n = 2$). Non-susceptibility to mupirocin and to rifampicin was detected in 64% ($n = 28$) and 18% ($n = 8$) of the isolates, respectively. All isolates were susceptible to vancomycin (MIC < 1.5 mg/L).

Antimicrobial resistance profiles detected were similar in both groups, individuals with no permanent address and living in shelter (Table 3), except for mupirocin that was higher among people attending shelters (mupirocin: 45% versus 79%, $p = 0.02$). However, 50% ($n = 6$) of the isolates with resistance to three or more antibiotics were recovered from the population with no permanent address.

S. aureus clonal diversity

The single MRSA isolate belonged to PFGE type D and was characterized by *spa* type t790, sequence type (ST) 22 and carried SCC*mec* cassette type IVh (Table 4). This isolate was recovered from a foreign man from the Middle East, screened at Gare 1, and not having a permanent address.

Almost half of the MSSA isolates (41.9%, $n = 18$) were distributed into two major clones (Table 4): ST398-*spa* type t1451 ($n = 13$) and ST30-*spa* types t399/t11980/t12808 associated with PFGE I ($n = 5$). The remaining MSSA belonged to 10 minor clones out of which four were represented by single isolates only (Table 4). The major MSSA lineage ST398-t1451 was prevalent among both homeless with no permanent address and shelter populations ($p = 1.000$). Noteworthy, seven of the 12 MSSA lineages were exclusively found in one of the groups (Table 4).

None of the isolates carried PVL.

Table 2 Demographic and health associated characteristics of the homeless individuals included in the study

Variable	Total (<i>n</i> = 84) (%)	<i>S. aureus</i> carriers (<i>n</i> = 43) (%)	No <i>S. aureus</i> carriers (<i>n</i> = 41) (%)	<i>p</i> value
Demographic characteristics				
Age (years)	49.0 ± 12.2	45.7 ± 12.7	52.5 ± 10.8	<i>0.009</i>
Gender				
Male	63 (75)	32 (74)	31 (76)	0.900
Female	21 (25)	11 (26)	10 (24)	
Housing status at recruitment				
No permanent address	34 (40)	20 (46)	14 (34)	0.250
Living in shelter	50 (60)	23 (54)	27 (66)	
Has a primary care provider				
Yes	29 (35)	13 (30)	16 (39)	0.400
No	55 (65)	30 (70)	25 (61)	
Emergency room observation (<i><</i> 2 years)	10 (12)	4 (9)	6 (15)	0.450
Recent antibiotic therapy (<i><</i> 6 months)	4 (5)	–	4 (10)	<i>0.040</i>
Addiction	54 (64)	29 (67)	25 (61)	0.540
Smoker	44 (52)	25 (58)	19 (46)	0.280
Alcohol abuse	26 (31)	10 (23)	16 (39)	0.120
Injection drug use	15 (18)	10 (23)	5 (12)	0.190
Vascular disease risk factors	18 (21)	9 (21)	9 (22)	0.910
Hypertension	12 (14)	7 (16)	5 (12)	0.590
Hypercholesterolaemia	5 (6)	1 (2)	4 (10)	0.150
Diabetes	7 (8)	4 (9)	3 (7)	0.740
Respiratory disease	53 (56)	29 (61)	24 (59)	0.400
Chronic bronchitis	53 (63)	29 (67)	24 (59)	0.400
Chronic obstructive pulmonary disease	5 (6)	2 (5)	3 (7)	0.610
Asthma	4 (5)	4 (9)	–	<i>0.040</i>
Pulmonary infection (<i><</i> 2 years)	23 (27)	11 (26)	12 (29)	0.700
Heart disease	8 (10)	4 (9)	4 (9)	0.940
Ischemic cardiac disease	3 (4)	2 (5)	1 (2)	0.580
Infectious disease*	12 (14)	5 (12)	7 (17)	0.480
HIV infection	5 (6)	3 (7)	2 (5)	0.680
HBV infection	2 (2)	–	2 (5)	0.140
HCV infection	5 (5)	2 (5)	3 (7)	0.610
Gastroenterologic disease	24 (29)	13 (30)	11 (27)	0.730
Viral hepatitis	5 (6)	2 (5)	3 (7)	0.610
Alcoholic hepatitis	7 (8)	2 (5)	5 (12)	0.210
Neurologic disease	12 (14)	7 (16)	5 (12)	0.590
Epilepsy	4 (5)	4 (9)	–	0.050
Previous stroke	7 (8)	4 (9)	3 (7)	0.740
Psychiatric disease	20 (24)	10 (23)	10 (24)	0.900
Depression or hopelessness	14 (17)	6 (14)	8 (20)	0.490
Schizophrenia	2 (2)	2 (5)	–	0.160

p values ≤ 0.05 were considered statistically significant and are italicized. *HIV* human immunodeficiency virus, *HBV* hepatitis B virus, *HCV* hepatitis C virus

*HBV and HCV infections were considered under “infectious disease” based on positive serological detection of the virus in individuals that did not developed hepatitis disease. Viral hepatitis that caused diagnosed disease were included in “gastroenterologic disease”

Table 3 Antibiotic susceptibility profiles of the *S. aureus* isolates recovered from nasal colonization among homeless individuals

Antibiotic	Non-susceptible <i>S. aureus</i> , n (%) ^a			<i>p</i> value
	Total (<i>n</i> = 44)	No permanent address (<i>n</i> = 20)	Shelters (<i>n</i> = 24)	
Cefoxitin	1 (2)	1 (5)	0	0.280
Penicillin	32 (73)	16 (80)	16 (67)	0.430
Rifampicin	8 (18)	2 (10)	6 (25)	0.180
Clindamycin	17 (39)	9 (45)	8 (33)	0.320
Erythromycin	20 (45)	9 (45)	11 (46)	0.920
Tetracycline	2 (5)	1 (5)	1 (4)	0.920
Fusidic acid	9 (20)	5 (25)	4 (17)	0.540
Ciprofloxacin	3 (7)	1 (5)	2 (8)	0.640
Mupirocin	28 (64)	9 (45)	19 (79)	<i>0.020</i>
Quinupristin-dalfopristin	4 (9)	1 (5)	3 (13)	0.640
Chloramphenicol	2 (5)	1 (5)	1 (4)	0.920

^a Percentage relative to the total number of *S. aureus* isolated in each condition

p values ≤ 0.050 were considered statistically significant and are italicized. None of the isolates showed resistance to gentamicin, trimethoprim-sulfamethoxazole, teicoplanin, linezolid, and vancomycin

Discussion

We performed the first nasal screening in a population of homeless individuals in Lisbon, Portugal. The *S. aureus* carriage rate (51.2%) detected was higher than rates previously reported in Portuguese healthy populations (20.1 to 46%) [16, 25–27]. However, MRSA was detected in a single individual (1.2%), which is in agreement with the very low MRSA prevalence among the healthy community in Portugal (< 1–1.8%) [16, 26, 27], and also much lower than the rates reported in other homeless communities (2.8 to 25.6%) [6, 28–30].

People living in temporary accommodation on homeless shelters in Lisbon had a significantly greater risk of respiratory infection and diagnosis of asthma and chronic obstructive pulmonary disease (COPD) than people living in a roofless condition [13]. However, our data indicate that *S. aureus* carriage does not contribute to this adverse outcome. Moreover, homeless people colonized with *S. aureus* were younger than individuals not colonized (*p* = 0.009), which could be due to a higher exposition to risk factors such as illicit drug use observed in 31.2% of the individuals with < 40 years; this is in agreement with other European reports [5].

Table 4 Molecular characterization of the *S. aureus* isolates recovered from nasal colonization among homeless individuals

MRSA/ MSSA	PFGE	<i>spa</i> type	ST	SCC <i>mec</i>	Prevalence, <i>n</i> (%)		
					Total (<i>n</i> = 44)	No permanent address (<i>n</i> = 20)	Shelter (<i>n</i> = 24)
MRSA	D	t790	22	IVh	1 (100)	1 (100)	0
MSSA	*	t1451	398	–	13 (30.2)	6 (31.6)	7 (29.2)
	I	t399/t11980/t12808	30	–	5 (11.6)	1 (5.3)	4 (16.7)
	B	t084/t346	582	–	4 (9.3)	3 (15.8)	1 (4.2)
	C	t073/t116/t861/t13186	45	–	4 (9.3)	3 (15.8)	1 (4.2)
	E	t127/t267/t693	1	–	4 (9.3)	2 (10.5)	2 (8.3)
	G	t126/t148	72	–	4 (9.3)	2 (10.5)	2 (8.3)
	L	t002/t5333	5	–	3 (7.0)	0	3 (12.5)
	F	t1309	672	–	2 (4.7)	0	2 (8.3)
	D	t005	22	–	1 (2.3)	1 (5.3)	0
	H	t209	109	–	1 (2.3)	1 (5.3)	0
	J	t493	182	–	1 (2.3)	0	1 (4.2)
	K	t5333	5	–	1 (2.3)	0	1 (4.2)

*No PFGE type was associated with ST398-t1451 isolates. SmaI restriction enzyme does not recognize restriction sites due to DNA methylation, a feature typical of ST398 *S. aureus* lineage

Homeless people have poor access to sanitation facilities and are often subjected to crowded living conditions promoting high person-to-person contact, which could support the increased *S. aureus* carriage rate detected, namely among people attending shelters. However, the very low MRSA rate detected in our study suggests that, despite the high prevalence of MRSA in Portuguese hospitals (39% in 2017) [9], the homeless population in Lisbon does not constitute a MRSA reservoir. Most individuals reported no recent contact with hospitals, which could justify the low MRSA carriage rate detected. Moreover, MSSA colonization has been reported as a protective factor against MRSA carriage and infection [31, 32].

A considerable proportion of isolates showed resistance to antibiotics commonly used to treat skin and soft tissue infections such as erythromycin (45%), clindamycin (39%), and fusidic acid (20%) [33, 34] limiting the treatment options among homeless [35]. Moreover, we found a high mupirocin non-susceptibility (64%), which might be driven by the increasing use of mupirocin in Portuguese hospitals for nasal decolonization protocols, where low-level resistance was reported [36].

The single MRSA recovered in the Lisbon homeless population belonged to EMRSA-15, the major clone circulating in Portuguese hospitals, as well as in the community, public buses and respective passengers in Lisbon and Oporto [11, 12, 37].

The majority of the MSSA isolates belonged to ST398-t1451 clonal type. MRSA-ST398 is the most prevalent *S. aureus* lineage associated with livestock [38]. In Portugal, MRSA-ST398 is extremely prevalent among pigs in different farms [39]. MRSA-ST398 is also found in human colonization but less frequently in disease. Contrarily, MSSA-ST398 was rarely reported in livestock, but is increasingly detected among people in the community, not only in asymptomatic carriage but also in disease [40–45]. MSSA-ST398 lineage is easily transmissible among humans, well adapted to the human host, and showed a high capacity to acquire resistance and pathogenic potential [40, 41, 46–50]. Single MRSA and MSSA ST398-t1451 isolates were already detected in the community in Portugal [11, 51], while the related MSSA-ST398-t571 lineage was found in 11% of nursing students in Lisbon [16] and as a minor clone causing bloodstream infection in Portuguese hospitals [51, 52].

Conclusions

We performed the first nasal screening of the homeless population in Portugal and contributed to overcome the lack of information available in Europe regarding this population. The overall *S. aureus* colonization rate was high (51.2%) while the MRSA nasal carriage was very low (1.2%) despite the broad confidence level used. Our results suggest that the homeless population of Lisbon does not constitute a reservoir of MRSA in the community, but a pool of a highly

transmissible ST398-t1451 MSSA lineage. *S. aureus* non-susceptibility to mupirocin, fusidic acid, and clindamycin among homeless queries the efficacy of decolonization protocols and skin and soft tissue infection antibiotherapy in this vulnerable population.

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Compliance with ethical standards

The protocol was approved by the Ethics Committee of Escola Superior de Saúde da Cruz Vermelha Portuguesa and a written informed consent was obtained from all participants.

Conflict of interest The authors declare that they have no competing interests.

References

- Lakhundi S, Zhang K (2018) Methicillin-resistant *Staphylococcus aureus*: molecular characterization, evolution, and epidemiology. Clin Microbiol Rev 31:e00020–18
- David MZ, Daum RS (2010) Community-associated methicillin-resistant *Staphylococcus aureus*: epidemiology and clinical consequences of an emerging epidemic. Clin Microbiol Rev 23(3):616–687
- European Union (2018) Social protection committee annual report 2018. Publications Office of the European Union, Brussels
- Marrana J, Ferreira P A, Firme J, Gonçalves J, Rosa C, Equipa Intergerações (2015) Do Outro Lado - Programa Intergerações / Intersituações – para o conhecimento das pessoas em situação sem-abrigo em Lisboa. Santa Casa da Misericórdia de Lisboa
- Fazel S, Geddes JR, Kushel M (2014) The health of homeless people in high-income countries: descriptive epidemiology, health consequences, and clinical and policy recommendations. Lancet 384(9953):1529–1540
- Charlebois ED, Bangsberg DR, Moss NJ, Moore MR, Moss AR, Chambers HF, Perdreau-Remington F (2002) Population-based community prevalence of methicillin-resistant *Staphylococcus aureus* in the urban poor of San Francisco. Clin Infect Dis 34(4): 425–433
- Gilbert M, MacDonald J, Gregson D, Siushansian J, Zhang K, Elsayed S, Laupland K, Louie T, Hope K, Mulvey M, Gillespie J, Nielsen D, Wheeler V, Louie M, Honish A, Keays G, Conly J (2006) Outbreak in Alberta of community-acquired (USA300)

- methicillin-resistant *Staphylococcus aureus* in people with a history of drug use, homelessness or incarceration. *CMAJ* 175(2):149–154
8. Farr AM, Aden B, Weiss D, Nash D, Marx MA (2012) Trends in hospitalization for community-associated methicillin-resistant *Staphylococcus aureus* in New York City, 1997–2006: data from New York State’s Statewide Planning and Research Cooperative System. *Infect Control Hosp Epidemiol* 33(7):725–731
 9. European Centre for Disease Control and Prevention (2018) Antimicrobial resistance surveillance in Europe 2017. Annual Report of the European Antimicrobial Resistance Surveillance Network (EARS-Net). European Centre for Disease Control and Prevention (ECDC), Stockholm: ECDC
 10. Melo-Cristino J, Resina C, Manuel V, Lito L, Ramirez M (2013) First case of infection with vancomycin-resistant *Staphylococcus aureus* in Europe. *Lancet* 382(9888):205
 11. Tavares A, Miragaia M, Rolo J, Coelho C, de Lencastre H, CA-MRSA/ MSSA working group (2013) High prevalence of hospital-associated methicillin-resistant *Staphylococcus aureus* in the community in Portugal: evidence for the blurring of community-hospital boundaries. *Eur J Clin Microbiol Infect Dis* 32(10):1269–1283
 12. Simões RR, Aires-de-Sousa M, Conceição T, Antunes F, da Costa PM, de Lencastre H (2011) High prevalence of EMRSA-15 in Portuguese public buses: a worrisome finding. *PLoS One* 6(3): e17630
 13. Martins HFG, Venâncio I, Pinto B, Fernandes RN (2017) Health care of homeless patient in Lisbon: comparison between roofless and houseless people. *Revista Sociedade Portuguesa de Medicina Interna (RSPMI)*. http://revista.spmi.pt/site/artigos_arquivo_consultar.php?id=2017096. Accessed 30 May 2019
 14. Amore K, Baker M, Howden-Chapman P (2011) The ETHOS definition and classification of homelessness: an analysis. *Eur J Homelessness* 5(2):19–37
 15. Instituto da Segurança Social Portugal (2009) Estratégia nacional para a integração de pessoas sem-abrigo. Prevenção, intervenção e acompanhamento, 2009–2015
 16. Conceição T, de Lencastre H, Aires-de-Sousa M (2017) Carriage of *Staphylococcus aureus* among Portuguese nursing students: a longitudinal cohort study over four years of education. *PLoS One* 12(11):e0188855
 17. Poulsen AB, Skov R, Pallesen LV (2003) Detection of methicillin resistance in coagulase-negative staphylococci and in staphylococci directly from simulated blood cultures using the EVIGENE MRSA detection kit. *J Antimicrob Chemother* 51(2):419–421
 18. Aires-de-Sousa M, Boye K, de Lencastre H, Deplano A, Enright MC, Etienne J, Friedrich A, Harmsen D, Holmes A, Huijsdens XW, Kearns AM, Mellmann A, Meugnier H, Rasheed JK, Spalburg E, Strommenger B, Struelens MJ, Tenover FC, Thomas J, Vogel U, Westh H, Xu J, Witte W (2006) High interlaboratory reproducibility of DNA sequence-based typing of bacteria in a multicenter study. *J Clin Microbiol* 44(2):619–621
 19. Okuma K, Iwakawa K, Turnidge JD, Grubb WB, Bell JM, O’Brien FG, Coombs GW, Pearman JW, Tenover FC, Kapi M, Tiensasitorn C, Ito T, Hiramatsu K (2002) Dissemination of new methicillin-resistant *Staphylococcus aureus* clones in the community. *J Clin Microbiol* 40(11):4289–4294
 20. Chung M, de Lencastre H, Matthews P, Tomasz A, Adamsson I, Aires de Sousa M, Camou T, Cocuzza C, Corso A, Couto I, Dominguez A, Gniadkowski M, Goering R, Gomes A, Kikuchi K, Marchese A, Mato R, Melter O, Oliveira D, Palacio R, Sá-Leão R, Santos Sanches I, Song JH, Tassios PT, Villari P (2000) Molecular typing of methicillin-resistant *Staphylococcus aureus* by pulsed-field gel electrophoresis: comparison of results obtained in a multilaboratory effort using identical protocols and MRSA strains. *Microb Drug Resist* 6(3):189–198
 21. Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG (2000) Multilocus sequence typing for characterization of methicillin-resistant and methicillin-susceptible clones of *Staphylococcus aureus*. *J Clin Microbiol* 38(3):1008–1015
 22. Milheirico C, Oliveira DC, de Lencastre H (2007) Update to the multiplex PCR strategy for assignment of *mec* element types in *Staphylococcus aureus*. *Antimicrob Agents Chemother* 51(9): 3374–3377
 23. Milheirico C, Oliveira DC, de Lencastre H (2007) Multiplex PCR strategy for subtyping the staphylococcal cassette chromosome *mec* type IV in methicillin-resistant *Staphylococcus aureus*: ‘SCC*mec* IV multiplex’. *J Antimicrob Chemother* 60(1):42–48
 24. Lina G, Piemont Y, Godail-Gamot F, Bes M, Peter MO, Gauduchon V, Vandenesch F, Etienne J (1999) Involvement of Panton-valentine leukocidin-producing *Staphylococcus aureus* in primary skin infections and pneumonia. *Clin Infect Dis* 29(5):1128–1132
 25. Kluytmans J, van Belkum A, Verbrugh H (1997) Nasal carriage of *Staphylococcus aureus*: epidemiology, underlying mechanisms, and associated risks. *Clin Microbiol Rev* 10(3):505–520
 26. Sá-Leão R, Sanches IS, Couto I, Alves CR, de Lencastre H (2001) Low prevalence of methicillin-resistant strains among *Staphylococcus aureus* colonizing young and healthy members of the community in Portugal. *Microb Drug Resist* 7(3):237–245
 27. Almeida ST, Nunes S, Paulo AC, Faria NA, de Lencastre H, Sa-Leao R (2015) Prevalence, risk factors, and epidemiology of methicillin-resistant *Staphylococcus aureus* carried by adults over 60 years of age. *Eur J Clin Microbiol Infect Dis* 34(3):593–600
 28. Ottomeyer M, Graham CD, Legg AD, Cooper ES, Law CD, Molani M, Matevossian K, Marlin J, Williams C, Newman R, Wasserman JA, Segars LW, Taylor TA (2016) Prevalence of nasal colonization by methicillin-resistant *Staphylococcus aureus* in persons using a homeless shelter in Kansas City. *Front Public Health* 4:234
 29. Landers TF, Harris RE, Wittum TE, Stevenson KB (2009) Colonization with *Staphylococcus aureus* and methicillin-resistant *S. aureus* among a sample of homeless individuals, Ohio. *Infect Control Hosp Epidemiol* 30(8):801–803
 30. Leibler JH, Leon C, Cardoso LJP, Morris JC, Miller NS, Nguyen DD, Gaeta JM (2017) Prevalence and risk factors for MRSA nasal colonization among persons experiencing homelessness in Boston, MA. *J Med Microbiol* 66:1183–1188
 31. Dall’Antonia M, Coen PG, Wilks M, Whiley A, Millar M (2005) Competition between methicillin-sensitive and -resistant *Staphylococcus aureus* in the anterior nares. *J Hosp Infect* 61(1): 62–67
 32. Shrestha NK, Fraser TG, Gordon SM (2019) Methicillin resistance in *Staphylococcus aureus* infections among patients colonized with methicillin-susceptible *Staphylococcus aureus*. *Clin Microbiol Infect* 25(1):71–75
 33. Liu C, Bayer A, Cosgrove SE, Daum RS, Fridkin SK, Gorwitz RJ, Kaplan SL, Karchmer AW, Levine DP, Murray BE, M JR, Talan DA, Chambers HF, Infectious Diseases Society of A (2011) Clinical practice guidelines by the infectious diseases society of america for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children. *Clin Infect Dis* 52(3): e18–e55
 34. Khan A, Wilson B, Gould IM (2018) Current and future treatment options for community-associated MRSA infection. *Expert Opin Pharmacother* 19(5):457–470
 35. Raoult D, Foucault C, Brouqui P (2001) Infections in the homeless. *Lancet Infect Dis* 1(2):77–84
 36. Monteiro M, Read A, Carneiro F, Soares MJ, Soares V (2016) Letter to the editor concerning the evaluation of mupirocin resistance in methicillin-resistant *Staphylococcus aureus* strains. *Acta Medica Port* 29(9):578
 37. Conceição T, Diamantino F, Coelho C, de Lencastre H, Aires-de-Sousa M (2013) Contamination of public buses with MRSA in

- Lisbon, Portugal: a possible transmission route of major MRSA clones within the community. *PLoS One* 8(11):e77812
38. Ballhausen B, Kriegerkorte A, van Alen S, Jung P, Kock R, Peters G, Bischoff M, Becker K (2017) The pathogenicity and host adaptation of livestock-associated MRSA CC398. *Vet Microbiol* 200: 39–45
 39. Conceição T, de Lencastre H, Aires-de-Sousa M (2017) Frequent isolation of methicillin resistant *Staphylococcus aureus* (MRSA) ST398 among healthy pigs in Portugal. *PLoS One* 12(4):e0175340
 40. Uhlemann AC, Porcella SF, Trivedi S, Sullivan SB, Hafer C, Kennedy AD, Barbian KD, McCarthy AJ, Street C, Hirschberg DL, Lipkin WI, Lindsay JA, DeLeo FR, Lowy FD (2012) Identification of a highly transmissible animal-independent *Staphylococcus aureus* ST398 clone with distinct genomic and cell adhesion properties. *mBio* 3:e00027–12
 41. Valentin-Domelier AS, Girard M, Bertrand X, Violette J, Francois P, Donnio PY, Talon D, Quentin R, Schrenzel J, van der Meer-Marquet N, Bloodstream Infection Study Group of the Réseau des Hygienistes du C (2011) Methicillin-susceptible ST398 *Staphylococcus aureus* responsible for bloodstream infections: an emerging human-adapted subclone? *PLoS One* 6(12):e28369
 42. David MZ, Siegel J, Lowy FD, Zychowski D, Taylor A, Lee CJ, Boyle-Vavra S, Daum RS (2013) Asymptomatic carriage of sequence type 398, *spa* type t571 methicillin-susceptible *Staphylococcus aureus* in an urban jail: a newly emerging, transmissible pathogenic strain. *J Clin Microbiol* 51(7):2443–2447
 43. Rasigade JP, Laurent F, Hubert P, Vandenesch F, Etienne J (2010) Lethal necrotizing pneumonia caused by an ST398 *Staphylococcus aureus* strain. *Emerg Infect Dis* 16(8):1330
 44. van Belkum A, Melles DC, Peeters JK, van Leeuwen WB, van Duijkeren E, Huijsdens XW, Spalburg E, de Neeling AJ, Verbrugh HA, Dutch Working Party on S, Research of M-S (2008) Methicillin-resistant and -susceptible sequence type 398 in pigs and humans. *Emerg Infect Dis* 14(3):479–483
 45. Valour F, Tasse J, Trouillet-Assant S, Rasigade JP, Lamy B, Chanard E, Verhoeven P, Decousser JW, Marchandin H, Bes M, Chidiac C, Vandenesch F, Ferry T, Laurent F, Lyon B, Joint Infection study g (2014) Methicillin-susceptible *Staphylococcus aureus* clonal complex 398: high prevalence and geographical heterogeneity in bone and joint infection and nasal carriage. *Clin Microbiol Infect* 20(10):O772–O775
 46. Uhlemann AC, McAdam PR, Sullivan SB, Knox JR, Khiabani H, Rabadan R, Davies PR, Fitzgerald JR, Lowy FD (2017) Evolutionary dynamics of pandemic methicillin-sensitive *Staphylococcus aureus* ST398 and its international spread via routes of human migration. *mBio* 8:e01375–16
 47. Mediavilla JR, Chen L, Uhlemann AC, Hanson BM, Rosenthal M, Stanak K, Koll B, Fries BC, Armellino D, Schilling ME, Weiss D, Smith TC, Lowy FD, Kreiswirth BN (2012) Methicillin-susceptible *Staphylococcus aureus* ST398, New York and New Jersey, USA. *Emerg Infect Dis* 18(4):700–702
 48. Argudin MA, Deplano A, Vandendriessche S, Dodemont M, Nonhoff C, Denis O, Roisin S (2018) CC398 *Staphylococcus aureus* subpopulations in Belgian patients. *Eur J Clin Microbiol Infect Dis* 37(5):911–916
 49. Lozano C, Aspiroz C, Charlez L, Gomez-Sanz E, Toledo M, Zarazaga M, Torres C (2011) Skin lesion by methicillin-resistant *Staphylococcus aureus* ST398-t1451 in a Spanish pig farmer: possible transmission from animals to humans. *Vector Borne Zoonotic Dis* 11(6):605–607
 50. Lozano C, Rezusta A, Gomez P, Gomez-Sanz E, Baez N, Martin-Saco G, Zarazaga M, Torres C (2012) High prevalence of *spa* types associated with the clonal lineage CC398 among tetracycline-resistant methicillin-resistant *Staphylococcus aureus* strains in a Spanish hospital. *J Antimicrob Chemother* 67(2):330–334
 51. Tavares A, Faria NA, de Lencastre H, Miragaia M (2013) Population structure of methicillin-susceptible *Staphylococcus aureus* (MSSA) in Portugal over a 19-year period (1992–2011). *Eur J Clin Microbiol Infect Dis* 33(3):423–432
 52. Faria NA, Miragaia M, de Lencastre H, Multi Laboratory Project C (2013) Massive dissemination of methicillin resistant *Staphylococcus aureus* in bloodstream infections in a high MRSA prevalence country: establishment and diversification of EMRSA-15. *Microb Drug Resist* 19(6):483–490

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