
Estimating *CDKN2A* mutation carrier probability among global familial melanoma cases using GenoMELPREDICT



Nicholas J. Taylor, PhD,^a Nandita Mitra, PhD,^b Lu Qian, MS,^c Marie-Françoise Avril, MD,^d D. Timothy Bishop, PhD,^e Brigitte Bressac-de Paillerets, PhD,^f William Bruno, MD, PhD,^g Donato Calista, MD,^h Francisco Cuellar, MD,ⁱ Anne E. Cust, PhD,^{j,k} Florence Demenais, MD,^l David E. Elder, MD,^m Anne-Marie Gerdes, PhD,ⁿ Paola Ghiorzo, PhD,^g Alisa M. Goldstein, PhD,^o Thais C. Grazziotin, MD,^p Nelleke A. Gruis, PhD,^q Johan Hansson, MD, PhD,^r Mark Harland, PhD,^c Nicholas K. Hayward, PhD,^s Marko Hocevar, MD, PhD,^t Veronica Höiom, PhD,^r Elizabeth A. Holland, BSc,^{k,u} Christian Ingvar, MD, PhD,^{v,w} Maria Teresa Landi, MD, PhD,^o Gilles Landman, MD, PhD,^x Alejandra Larre-Borges, MD,^y Graham J. Mann, PhD,^{k,u} Eduardo Nagore, MD, PhD,^z Håkan Olsson, MD, PhD,^{v,w} Jane M. Palmer, RN,^s Barbara Perić, MD, PhD,^t Dace Pjanova, PhD,^{aa} Antonia L. Pritchard, PhD,^s Susana Puig, MD, PhD,^{i,bb} Helen Schmid, MPH,^{k,u} Nienke van der Stoep, PhD,^{cc} Margaret A. Tucker, MD,^o Karin A. W. Wadt, MD, PhD,ⁿ Xiaohong R. Yang, PhD,^o Julia A. Newton-Bishop, MD,^e and Peter A. Kanetsky, PhD,^c on behalf of the GenoMEL Study Group

College Station, Texas; Philadelphia, Pennsylvania; Tampa, Florida; Paris, France; Leeds, United Kingdom; Villejuif, France; Genoa and Cesena, Italy; Sydney and Herston, Australia; Barcelona and Valencia, Spain; Copenhagen, Denmark; Bethesda, Maryland; Porto Alegre and São Paulo, Brazil; Leiden, the Netherlands; Stockholm and Lund, Sweden; Ljubljana, Slovenia; Montevideo, Uruguay; and Riga, Latvia

From the Department of Epidemiology and Biostatistics, Texas A&M University, College Station^a; Department of Biostatistics, Epidemiology, and Informatics, University of Pennsylvania, Philadelphia^b; Department of Cancer Epidemiology, H. Lee Moffitt Cancer Center and Research Institute, Tampa^c; Assistance Publique-Hôpitaux de Paris, Hôpital Cochin et Université Paris Descartes^d; Section of Epidemiology and Biostatistics, Leeds Institute of Medical Research at St. James's, University of Leeds^e; Gustave Roussy, Université Paris-Saclay, Département de Biopathologie and Institut National de la Santé et de la Recherche Médicale U1186, Villejuif^f; Department of Internal Medicine and Medical Specialties, University of Genoa and Istituto de Ricovero e Cura a Carattere Scientifico AOU San Martino-IST^g; Dermatology Unit, Maurizio Bufalini Hospital, Cesena^h; Melanoma Unit, Dermatology Department, Hospital Clinic Barcelona, Institut de Investigacions Biomediques August Pi Sunyer, Universitat de Barcelonaⁱ; Sydney School of Public Health,^j and Melanoma Institute Australia,^k The University of Sydney; Institut National de la Santé et de la Recherche Médicale UMR-946, Genetic Variation and Human Disease Unit, Université Paris Diderot^l; Department of Pathology and Laboratory Medicine, Hospital of the University of Pennsylvania, Philadelphia^m; Department of Clinical Genetics, University Hospital of Copenhagenⁿ; Human Genetics Program, Division of Cancer Epidemiology and Genetics, National Cancer Institute, Bethesda^o; Pontifícia Universidade Católica do Rio Grande do Sul, Porto Alegre^p; Department of Dermatology, Leiden University Medical Centre^q; Department of Oncology-Pathology, Karolinska Institutet, Stockholm^r; QIMR Berghofer Medical Research Institute, Herston^s; Institute of Oncology Ljubljana, Zaloska^t; Westmead Institute for Medical Research, University of Sydney^u; Department of Clinical Sciences^v and Department of Surgery,^w Lund University Hospital; Department of Pathology, Escola Paulista de Medicina, Universidade Federal de São Paulo^x; Unidad de Lesiones Pigmentadas, Cátedra de Dermatología, Hospital de Clínicas, Universidad de la República, Montevideo^y;

Department of Dermatology, Instituto Valenciano de Oncología^z; Latvian Biomedical Research and Study Centre, Riga^{aa}; Centro de Investigación Biomedica en Red de Enfermedades Raras, Instituto de Salud Carlos III, Barcelona^{bb}; and Department of Clinical Genetics, Leiden University Medical Center.^{cc}

Funding sources: Supported by the Cancer Research UK Programme Award (nos. C588/A4994 and C588/A10589); a Cancer Research UK project grant (C8216/A6129); the US National Institutes of Health (CA83115 to Drs Kanetsky, Bishop, and Elder; CA5558 to Dr Landi; Dr Taylor was supported by CA147832 to Dr Kanetsky); the Intramural Research Program of the National Institutes of Health, National Cancer Institute (NCI), Division of Cancer Epidemiology and Genetics; the National Health and Medical Research Council of Australia (NHMRC 107359, 402761, 633004, 566946, 211172); the Cancer Council New South Wales (project grants 77/00, 06/10); the Cancer Institute New South Wales (CINSW 05/TPG/1-01, 10/TPG/1-02); the Cancer Council Victoria and the Cancer Council Queensland (project grant 371); Coordenação de Amparo à Pesquisa do Estado de São Paulo (no. 2007/04313-2); the National Health and Medical Research Council of Australia and the NCI (CA88363); the Cancer Research Foundations of Radiumhemmet and the Swedish Cancer Society; the Paulsson Trust, Lund University; the Swedish Cancer Society and European Research Council Advanced Grant (ERC-2011-294576); the research at the Melanoma Unit in Barcelona is partially funded by Spanish Fondo de Investigaciones Sanitarias (grants P115/00716 and P115/00956); Centros de Investigación Biomédica en Red de Enfermedades Raras of the Instituto de Salud Carlos III, Spain, is co-financed by European Development Regional Fund "A way to achieve Europe"; the Agency for Management of University and Research Grants 2014_SGR_603 of the Catalan government, Spain; the European Commission under the 6th Framework Programme (contract no. LSHC-CT-2006-018702, GenoMEL) and the 7th Framework Programme, Diagnostics;

Background: Although rare in the general population, highly penetrant germline mutations in *CDKN2A* are responsible for 5%–40% of melanoma cases reported in melanoma-prone families. We sought to determine whether MELPREDICT was generalizable to a global series of families with melanoma and whether performance improvements can be achieved.

Methods: In total, 2116 familial melanoma cases were ascertained by the international GenoMEL Consortium. We recapitulated the MELPREDICT model within our data (GenoMELPREDICT) to assess performance improvements by adding phenotypic risk factors and history of pancreatic cancer. We report areas under the curve (AUC) with 95% confidence intervals (CIs) along with net reclassification indices (NRIs) as performance metrics.

Results: MELPREDICT performed well (AUC 0.752, 95% CI 0.730–0.775), and GenoMELPREDICT performance was similar (AUC 0.748, 95% CI 0.726–0.771). Adding a reported history of pancreatic cancer yielded discriminatory improvement ($P < .0001$) in GenoMELPREDICT (AUC 0.772, 95% CI 0.750–0.793, NRI 0.40). Including phenotypic risk factors did not improve performance.

Conclusion: The MELPREDICT model functioned well in a global data set of familial melanoma cases. Adding pancreatic cancer history improved model prediction. GenoMELPREDICT is a simple tool for predicting *CDKN2A* mutational status among melanoma patients from melanoma-prone families and can aid in directing these patients to receive genetic testing or cancer risk counseling. (J Am Acad Dermatol 2019;81:386–94.)

Key words: *CDKN2A*; familial melanoma; GenoMEL; GenoMELPREDICT; mutation prediction.

Inherited mutations in the cyclin-dependent kinase inhibitor 2A (*CDKN2A*) gene are major risk factors for familial melanoma.^{1–3} The frequency of *CDKN2A* mutations in melanoma-prone families varies widely (<5%–40%) with the number of family members with melanoma diagnoses and the number of primary melanomas diagnosed within an individual.^{1,4–6} The penetrance of *CDKN2A* mutations in melanoma-prone families is a function of population incidence rates of melanoma

CAPSULE SUMMARY

- Available prediction tools for cyclin-dependent kinase inhibitor 2A (*CDKN2A*) status were developed among small, homogeneous populations and lack generalizability. GenoMELPREDICT is a globally generalizable and simple clinical tool for predicting *CDKN2A* mutational status among familial melanoma patients.
- GenoMELPREDICT can aid in appropriate patient management, whether that is by recommending genetic testing or cancer risk counselling.

and is modified by environmental factors, melanoma-associated phenotypes, and melanocortin-1-receptor variants.^{3,7} In light of geographic variability in mutation penetrance, a standard guideline for recommending *CDKN2A* genetic testing has not been suitable for heterogeneous populations.⁸ GenoMEL, the International Melanoma Genetics Consortium, supports a qualitative framework to identify candidates for *CDKN2A* mutation testing by using population-based melanoma incidence, the

a grant from Fundació La Marató de TV3 201331-30, Catalonia, Spain; a grant from Fundación Científica de la Asociación Española Contra el Cáncer, Spain (GCB15152978SOEN); Centres de Recerca de Catalunya Programme/Generalitat de Catalunya; the Italian Association for Cancer Research (to Dr Ghorzo); the Italian Ministry of Health-Ricerca Finalizzata 2016 (to IRCCS San Martino-IST, Genoa); the Programme Hospitalier de Recherche Clinique (PHRC-AOM-07-195) awarded to Dr Avril and Dr Demenais; a grant from the Institut National du Cancer (to Dr Bressac de-Paillerets for coordination of Melanoma Oncogenetics in France); the Comisión Honoraria de Lucha Contra el Cáncer, Comisión Sectorial de Investigación Científica, Fundación Manuel Pérez, Montevideo, Uruguay; the Dutch Cancer Society (UL 2012-5489 to Dr Gruis); a scholarship awarded by Consejo Nacional de Ciencia y Tecnología, Mexico (152256/158706 to Dr Cuellar); and a Career Development

Fellowship from the National Health and Medical Research Council (1147843) and Cancer Institute New South Wales (15/CDF/1-14) to Dr Cust. Part of the work was carried out at the Esther Koplowitz Center, Barcelona.

Conflicts of interest: None disclosed.

Accepted for publication January 30, 2019.

Reprint requests: Peter A. Kanetsky, PhD, MPH, H. Lee Moffitt Cancer Center and Research Institute, 12902 Magnolia Dr, MRC Bldg 213, Tampa, FL 33612. E-mail: peter.kanetsky@moffitt.org.

Published online February 5, 2019.

J Am Acad Dermatol 2019;81:386–94.

0190-9622/\$36.00

© 2019 by the American Academy of Dermatology, Inc. Published by Elsevier Inc. All rights reserved.

<https://doi.org/10.1016/j.jaad.2019.01.079>

Abbreviations used:

| | |
|---------|--------------------------------------|
| AUC: | area under the curve |
| CDKN2A: | cyclin-dependent kinase inhibitor 2A |
| CI: | confidence interval |
| ROC: | receiver operating characteristic |

diagnosis of multiple primary melanomas, and a verified family history of melanoma and/or pancreatic cancer.⁸ The rapid identification of familial melanoma patients with low probabilities of germline mutations in *CDKN2A* could help direct patients toward risk counseling and away from inappropriate genetic testing, especially because a negative test result is unlikely to influence risk management or foster a potential conversation about genetic testing for mutations in other known but much rarer high-penetrance melanoma genes.

MELPREDICT is a published logistic regression model to predict *CDKN2A* mutation carrier status.⁹ MELPREDICT performed well (area under the curve [AUC] 0.881) among melanoma patients (n = 116) belonging to melanoma-prone families in Boston, Massachusetts, and similarly (AUC 0.803) among those from melanoma-prone families in Toronto, Canada (n = 143).⁹ We sought to determine whether MELPREDICT was generalizable to a large series of melanoma-prone families from 20 countries participating in GenoMEL. Further, we evaluated whether improvements in model performance can be achieved by adding personal or family history of pancreatic cancer and/or phenotypic risk factors for melanoma.

METHODS

Study population

The GenoMEL Consortium comprises 29 study centers in Australia, Europe, the Middle East, and North and South America. GenoMEL used a common protocol to obtain research data as previously described.¹⁰ Written informed consent was obtained from each participant, and individual GenoMEL centers received study approval from their respective institutional review boards. Consenting participants completed a self-administered questionnaire that solicited information on phenotypic characteristics and personal and family history of melanoma and other cancers.^{10,11}

Study sample

Our study sample includes 2116 melanoma-prone patients with the *CDKN2A* genotype. These participants were from 900 melanoma-prone families defined by the presence of ≥ 3 verified melanoma

patients among blood relatives (individuals who share a common ancestor and are not related by marriage) or 2 verified melanoma patients among first-degree blood relatives recruited at 1 of 20 GenoMEL centers (Table I). There were 359 reports in 122 families of a personal or family history of pancreatic cancer, and pathologic verification was available for 79 (22%) of these reports; the remainder were self-reported.

CDKN2A genotyping

Germline DNA was screened for mutations in *CDKN2A* (including exons 1 α , 1 β , 2, and 3), and mutations were classified as pathogenic (ie, positive) or nonpathogenic (ie, negative) as previously described.^{10,11} Eleven families had ≥ 1 member who was known to carry a mutation in another melanoma high-penetrance gene; these families were included in our analyses.

Statistical analysis

Using the MELPREDICT logistic regression model, we estimated the predictive probability of *CDKN2A* mutation carriage among study participants. In this model, the probability of *CDKN2A* mutation carriage is defined as $e^L / (1 + e^L)$, with $L = 1.99 + [(0.92 \times \text{number of primary melanoma diagnoses}) + (0.74 \times \text{number of additional family members diagnosed with melanoma}) - (2.11 \times \ln [\text{age at first melanoma diagnosis}])]$. The AUC was derived from the set of predictive probabilities.^{9,12} Using data from GenoMEL, we modeled the probability of *CDKN2A* mutation carriage as a function of these 3 variables and considered this our baseline model (GenoMELPREDICT). We used a generalized estimating equation with a logit link function and independence covariance structure with robust standard errors to account for familial clustering. We evaluated changes in baseline model performance associated with the addition of reported personal or family history of pancreatic cancer (yes, no); facial freckling (none, very few, few, some many, very many); proclivity to burn (tan with no burning, mild sun burning, sun burning with peeling, severe sun burning with blistering); proclivity to tan (very tanned, moderate tanning, mild tanning, no tanning); eye color (brown or black, blue, other); hair color (black, brown, blonde or fair, red); and skin type (very fair, fair, olive or brown or black), including all pairwise and triplet combinations of these phenotypic variables.

We used the empirical method of DeLong¹³ to estimate and compare (via a Wald test) paired AUCs of receiver operating characteristic (ROC) curves. For each model, AUCs and 95% confidence intervals (CIs)

Table I. Number of participants and families by ascertainment center

| GenoMEL center | Participants* | Families [†] | Average no. participants per family [‡] | Average no. affected members per family [§] |
|----------------------------|---------------|-----------------------|--|--|
| Barcelona, Spain | 44 | 25 | 1.8 | 2.1 |
| Bethesda, Maryland | 199 | 46 | 4.3 | 4.8 |
| Cesena, Italy | 50 | 24 | 2.1 | 2.1 |
| Copenhagen, Denmark | 47 | 34 | 1.4 | 2.5 |
| Genoa, Italy | 34 | 16 | 2.1 | 2.3 |
| Leeds, Great Britain | 158 | 77 | 2.1 | 2.8 |
| Leiden, the Netherlands | 210 | 60 | 3.5 | 4.6 |
| Ljubljana, Slovenia | 9 | 4 | 2.3 | 2.3 |
| Lund, Sweden | 20 | 7 | 2.9 | 4.4 |
| Montevideo, Uruguay | 8 | 4 | 2.0 | 2.0 |
| Paris, France | 341 | 176 | 1.9 | 2.5 |
| Philadelphia, Pennsylvania | 78 | 36 | 2.2 | 2.4 |
| Porto Alegre, Brazil | 9 | 5 | 1.8 | 2.2 |
| Queensland, Australia | 96 | 21 | 4.6 | 6.2 |
| Riga, Latvia | 5 | 5 | 1.0 | 2.6 |
| Santiago, Chile | 3 | 2 | 1.5 | 2.0 |
| São Paulo, Brazil | 13 | 8 | 1.6 | 2.1 |
| Stockholm, Sweden | 39 | 21 | 1.9 | 2.8 |
| Sydney, Australia | 722 | 305 | 2.4 | 3.4 |
| Tel Aviv, Israel | 21 | 18 | 1.2 | 2.0 |
| Valencia, Spain | 10 | 6 | 1.7 | 2.2 |
| Total | 2116 | 900 | 2.2 | 3.1 |

*Verification of melanoma was available for >99% of participants by pathology report (74%), physician letter or clinical document verifying melanoma diagnosis (23%), cancer registry data (2%), or death certificate (<1%). Excludes affected individuals with noncutaneous melanoma diagnoses or persons who are members of melanoma families by marriage and not ancestry.

[†]Family members with a melanoma of the uveal tract or conjunctiva did not contribute to defining a melanoma family.

[‡]Includes only participants who contribute to prediction modeling.

[§]Includes family members who might not contribute to prediction modeling because of missing data.

were calculated by 10-fold cross-validation to evaluate discrimination between *CDKN2A* mutation carriers and noncarriers, and we used 1-stage cluster sampling to randomly assign all members of a family to the same fold. Optimal discrimination was determined by maximizing sensitivity and specificity. Improvement in model performance was assessed by measuring the difference between paired model AUCs and by event and nonevent net reclassification indices.¹³⁻¹⁵ Models incorporating phenotypic factors were performed on sample sizes that varied according to factor missingness; for each augmented model, we reran our baseline model on the corresponding reduced sample size. Multiple imputation by the fully conditional specification method was used to restore missing values.¹⁶ All analyses were performed with SAS version 9.4 (SAS Institute, Cary, NC) and R (R Core Team; <http://www.R-project.org/>).

RESULTS

CDKN2A genotype was available for 711 (33.6%) mutation carriers and 1405 (66.4%) noncarriers belonging to 900 melanoma-prone families.

CDKN2A mutations identified in GenoMEL families have been previously published.^{10,17} Results of multivariable analyses for our 3-variable baseline and 4-variable GenoMELPREDICT model that included pancreatic cancer are presented in Table II. Age at first melanoma diagnosis, higher numbers of primary melanomas, higher numbers of family members with a melanoma diagnosis, and a personal or family history of pancreatic cancer were independently associated ($P < .0001$) with *CDKN2A* mutation carriage.

Using the published MELPREDICT model parameter coefficients to predict *CDKN2A* mutation carriage in the GenoMEL sample set resulted in an AUC of 0.752 (95% CI 0.730-0.775); the mean estimated probability of *CDKN2A* mutation carriage was 42.7% for mutation carriers, and 13.0% for noncarriers. De novo modeling, ie, GenoMELPREDICT, of age at first melanoma diagnosis, number of primary melanoma diagnoses, and number of additional family members with melanoma diagnoses resulted in an AUC of 0.748 (95% CI 0.726-0.771). For this model, the mean estimated probability of *CDKN2A* mutation carriage was 46.4% for mutation carriers, and 27.2% for

Table II. Distribution of pathogenic *CDKN2A* mutations among GenoMEL cases and model estimates for the baseline and 4-variable GenoMELPREDICT models

| Variable | N (%) with mutation | GenoMELPREDICT | | | |
|---|---------------------|------------------|--------|------------------|--------|
| | | OR (95% CI)* | P | OR (95% CI)* | P |
| ln(age at diagnosis) | | 0.29 (0.22-0.39) | <.0001 | 0.28 (0.22-0.37) | <.0001 |
| Primary melanomas, n | | | | | |
| 1 | 378/1426 (26.5) | 1.20 (1.10-1.31) | <.0001 | 1.20 (1.10-1.32) | <.0001 |
| 2 | 153/380 (40.3) | | | | |
| ≥3 | 180/310 (58.1) | | | | |
| Other family members with melanoma, n | | | | | |
| 1 | 132/669 (19.7) | 1.29 (1.20-1.38) | <.0001 | 1.26 (1.17-1.32) | <.0001 |
| 2 | 146/560 (26.0) | | | | |
| 3 | 91/218 (28.6) | | | | |
| ≥4 | 342/569 (60.1) | | | | |
| Personal or family history of pancreatic cancer | | | | | |
| No | 495/1757 (28.2) | | | | |
| Yes | 216/359 (60.2) | | | 3.05 (1.97-4.74) | <.0001 |

CI, Confidence interval; OR, odds ratio.

*Estimated from a generalized estimating equation model using a logit link function and with adjustment for familial clustering. For reference, age (in years) at first cutaneous melanoma diagnosis is modeled as ln(age at first diagnosis) with range 2.30 (10-year-old) to 4.55 (95-year-old). A ln(age) of 3.0 corresponds to a 20-year-old, an ln(age) of 3.5 to a 33-year-old, and an ln(age) of 4.0 to a 55-year-old.

noncarriers. The difference in AUC values between these models was not statistically significant ($P = .21$; Fig 1, A).

Adding phenotypic risk factors did not result in performance improvements of the 3-variable baseline GenoMELPREDICT model (data not tabulated and available upon request). However, including personal or family history of pancreatic cancer to the 3-variable baseline model significantly ($P < .0001$) augmented its discriminatory performance, yielding an AUC of 0.772 (95% CI 0.750-0.793) (Fig 1, B). The mean estimated probability of *CDKN2A* mutation carriage was 48.4% for mutation carriers and 26.2% for noncarriers. The net reclassification index was 0.404 with noted improvement (79.6%) for reclassification of noncarriers but at the expense of reclassification of carriers (−39.2%). Adding phenotypic variables to the 4-variable model that included personal or family history of pancreatic cancer did not result in further model improvement (data not tabulated and available upon request). Selecting a predicted probability cutoff of 35% for this 4-variable model, similar to the theoretical best cutoff with the Youden Index (34.4%), resulted in a sensitivity of 61%, specificity of 79%, positive predictive value of 60%, and a negative predictive value of 80%. A range of model metrics for the 3-variable baseline and 4-variable GenoMELPREDICT models is available upon request. Consistent with results using observed phenotypic data, adding imputed phenotypic

variables did not result in performance improvement of either the baseline or 4-variable GenoMELPREDICT models (data not tabulated and available upon request).

In subgroup analyses, the AUCs for the 3-variable and 4-variable GenoMELPREDICT models were somewhat higher among Australian participants (0.809, 95% CI 0.773-0.844, for both 3-variable and 4-variable) and similar or slightly higher among participants living in Northern European countries (3-variable AUC 0.760, 95% CI 0.718-0.803, and 4-variable AUC 0.775, 95% CI 0.734-0.816). Model performance was lower among participants from Southern European and South American countries (3-variable AUC 0.625, 95% CI 0.535-0.714, and 4-variable AUC 0.635, 95% CI 0.548-0.722).

Models that excluded families with members who carried a mutation in other known melanoma high-penetrance genes or excluded families without a verified report of personal or family history of pancreatic cancer were consistent with our main results. In models excluding melanoma-prone families from Sydney, which comprised one third of all data used in our analysis, AUCs for the 3-variable baseline (0.772, 95% CI 0.747-0.797) and 4-variable (0.784, 95% CI 0.760-0.808) GenoMELPREDICT models were slightly higher than models using all available GenoMEL data. After excluding participants from the Bethesda and Queensland centers, both of which contributed higher numbers of

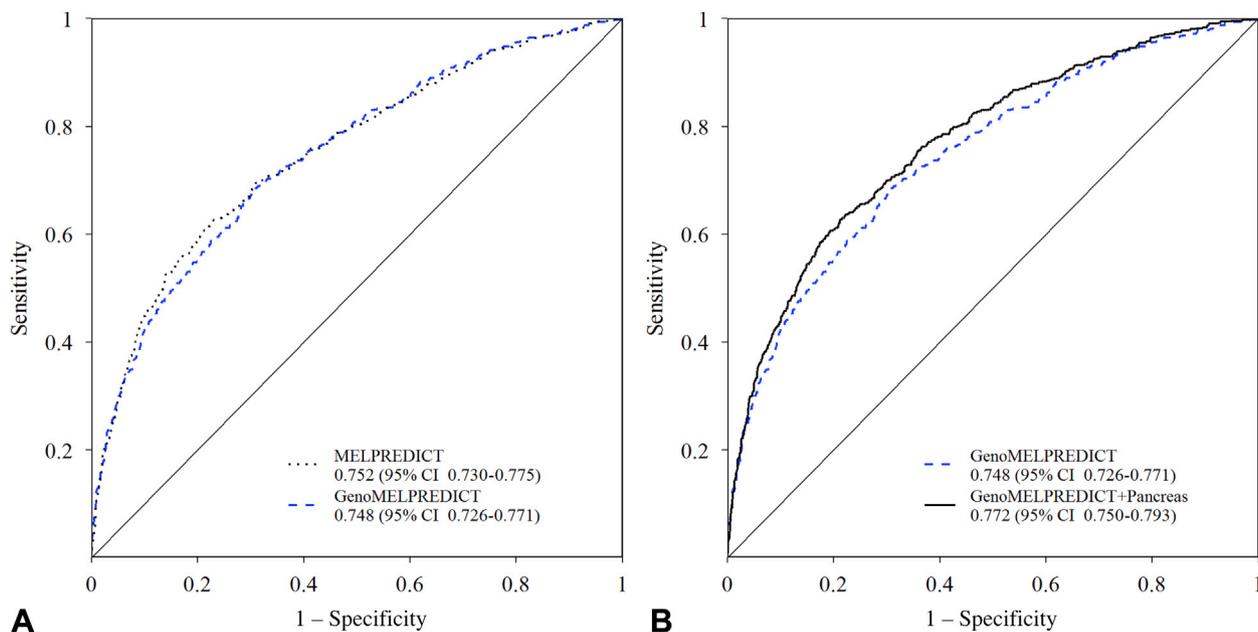


Fig 1. Receiver operator characteristic curves for GenoMELPREDICT models. Comparison of the receiver operator characteristic curves derived from the 3-variable baseline GenoMELPREDICT model and MELPREDICT as reported by Niendorf et al⁹ (**A**) and 3-variable baseline GenoMELPREDICT model and the 4-variable GenoMELPREDICT model (**B**), including any reported personal or family history of pancreatic cancer. Legend results are cross-validated areas under the curve and 95% CIs for GenoMELPREDICT models and areas under the curve and 95% CI for MELPREDICT. *CI*, Confidence interval.

affected members with *CDKN2A* genotype data per family (4.3 and 4.6, respectively), model AUCs were slightly lower than those calculated from all available GenoMEL data (3-variable AUC 0.708, 95% CI 0.681-0.734; 4-variable AUC 0.740, 95% CI 0.714-0.765).

DISCUSSION

We show that the published MELPREDICT model used to predict *CDKN2A* mutational status is generalizable to the global community of melanoma-prone families represented in GenoMEL. We also provide evidence that adding personal and family history of pancreatic cancer to the model, a variable that can be collected with little additional associated cost, leads to some improvement in the ability to predict *CDKN2A* mutational status, and we call this augmented model GenoMELPREDICT. The predictive performance of GenoMELPREDICT is comparable to other clinical tools used to predict *BRCA1* and *BRCA2* mutational status among breast cancer patients.¹⁸⁻²⁰

The diverse global sample of familial melanoma cases recruited by GenoMEL enabled us to detect a broader spectrum of *CDKN2A* mutations compared with the limited number (18 variants) reported by the original MELPREDICT developers.⁹ A total of 85 unique, putatively pathogenic mutations were

identified among GenoMEL cases, allowing for a more representative appraisal of GenoMELPREDICT's performance.

MelaPRO²¹ and CM-Score²² are 2 other published algorithms for *CDKN2A* mutation prediction among melanoma-prone families. MelaPRO incorporates melanoma risk among unaffected family members, uses a Bayesian approach to predict carrier status, and incorporates penetrance estimates for areas of high- and low-baseline incidence and was derived using data from the population-based Genes Environment and Melanoma study.²³ MelaPRO was tested on a patient sample drawn from the same ascertainment center used by Niendorf et al⁹ to test the MELPREDICT algorithm, and it outperformed ($n = 195$, AUC 0.86) MELPREDICT on the prediction of carrier status among the same homogeneous familial cohort. The CM-Score algorithm is a multivariate logistic regression model developed by using a training cohort of 1227 Dutch melanoma-prone families and incorporates 5 clinical features (number of family members with melanoma and with multiple primary melanomas, median age at diagnosis, and presence of pancreatic cancer or upper airway cancer in a family member) to predict germline *CDKN2A* mutational status. CM-Score was validated in a combined Swedish and Dutch cohort of 421

Table III. Candidacy criteria for consideration of genetic testing

| Low melanoma incidence area/population | Moderate-to-high melanoma incidence area/population |
|---|---|
| 2 (synchronous or metachronous) primary melanomas in an individual and/or Families with at least 1 invasive melanoma and ≥ 1 other diagnosis of melanoma and/or pancreatic cancers among first-degree or second-degree relatives on the same side of the family | 3 (synchronous or metachronous) primary melanomas in an individual and/or Families with at least 1 invasive melanoma and ≥ 2 other diagnoses of invasive melanoma and/or pancreatic cancer among first-degree or second-degree relatives on the same side of the family |

This table refers to pathologically confirmed invasive melanoma.

Table adapted from Leachman et al. Selection criteria for genetic assessment of patients with familial melanoma. *J Am Acad Dermatol.* 2009;61(4):677 e1-14.

melanoma-prone families and demonstrated excellent performance characteristics among a homogeneous group of Northern Europeans (AUC 0.94, 95% CI 0.90-0.98), possibly because of the high incidence of specific founder mutations in this population.²²

We opted to assess MELPREDICT rather than MelaPRO or CM-Score. CM-Score was developed among a cohort of Swedish and Dutch melanoma-prone families with a high incidence of specific founder mutations, reducing generalizability. Due to the increased incidence of upper airway cancers observed among carriers of these Swedish and Dutch founder mutations, the CM-Score algorithm incorporates any history of such cancers and might be inappropriate for a heterogeneous population of familial melanoma kindreds.²² In our data set, there were 295 reports of personal or family history of laryngeal, pharyngeal, or oral cavity cancers within 97 families; pathologic verification was available for 30 (10%) of these reports. MelaPRO requires users to specify *CDKN2A* penetrance associated with the population under study, which involves more complex assessments of the source populations from which individual cases arise; this aspect might potentially limit MelaPRO's utility in clinical practice. Because the GenoMEL Consortium includes melanoma-prone families from around the world and simultaneous modeling of multiple *CDKN2A* penetrances was not feasible, our preference was to evaluate generalizability and enhancement of MELPREDICT.

The 3- and 4-variable GenoMELPREDICT models perform best among participants living in Australia. This likely reflects the large influence of these individuals, who comprise nearly 40% of our analytic sample, on overall model estimates. In converse, the 3- and 4-variable GenoMELPREDICT models perform poorest among participants living in Southern European and South American sites. This likely reflects our working definition of a melanoma-prone family, which minimally is 2 verified melanoma cases in a first-degree blood relative. This

definition might be too strict for populations that experience lower incidence of melanoma, for which a definition of ≥ 2 verified melanoma cases among blood relatives might be better suited. Of the 900 families who had at least 1 member who contributed to GenoMELPREDICT modeling, the Southern European and South American sites had, as expected, a lower mean number of affected members per family (2.1) compared with that for the Northern European (3.3) or Australian (3.6) sites.

We have reported on the limitations of the GenoMEL study, which included differences in amount of data collected across centers, possible misclassification of *CDKN2A* mutations, lack of centralized pathology review for reported cases of melanoma, and nonpopulation-based ascertainment and sampling of families at some centers on the basis of known mutation status or number of familial melanoma cases.^{10,17} Although pathologic verification of reported personal or familial cases of pancreatic cancer was low (22%) in GenoMEL, the positive predictive value and sensitivity of self-report of family history for this cancer are both reported to surpass 70%.²⁴

GenoMELPREDICT is an effective predictor of *CDKN2A* mutational status, and statistical performance improvement was made by adding any reported personal or family history of pancreatic cancer. However, only 5%-10% of melanomas can be attributed to high-penetrance germline genes, and thus, only a small proportion of patients with melanoma diagnoses will benefit from genetic testing for *CDKN2A*.²⁵ Despite controversy regarding the genetic testing of individuals in melanoma-prone families,²⁶ there is burgeoning commercial availability of such tests. We have previously published on the challenges of developing a single encompassing worldwide recommendation to best guide health professionals with respect to which patients should be considered for *CDKN2A* genetic testing.⁸ In Table III, we republish our candidacy criteria for consideration of genetic testing.⁸ Complementing

these criteria, GenoMELPREDICT might serve as a quick and robust tool applicable worldwide for directing patients away from unnecessary genetic testing, especially in the event of a low-carrier probability estimate. Moreover, guidance considering the management of patients belonging to melanoma-prone families in the context of genetic testing is available in a Continuing Medical Education article published in the *Journal of the American Academy of Dermatology*.²⁶ A user-friendly web-based interface to calculate the probability of carriage of a *CDKN2A* mutation is available at www.genomel.org.

We acknowledge the contributions of the participants and their families and the many clinicians, geneticists, genetic counsellors, and allied health professionals involved in their management. This work was performed in participation with the following members at the following study locations: Leeds, United Kingdom (Linda Whitaker, Paul Affleck, Jennifer H. Barrett, Jane Harrison, Mark M. Iles, Juliette Randerson-Moor, John C. Taylor, Kairen Kukulizch, Susan Leake, Birute Karpavicius, Sue Haynes, Tricia Mack, May Chan, and Yvonne Taylor); Barcelona, Spain (Paula Aguilera, Llúcia Alós, Celia Badenas, Alicia Barreiro, Neus Calbet, Cristina Carrera, Carlos Conill, Mireia Domínguez, Daniel Gabriel, Pablo Iglesias, Josep Malvehy, M. Eugenia Moliner, Javiera Pérez, Ramon Pigem, Miriam Potrony, Joan Anton Puig Butille, Ramon Rull, Marcelo Sánchez, Gemma Tell-Martí, Sergi Vidal-Sicart, and Oriol Yelamos) Valencia (Zaida García-Casado, Celia Requena, José Bañuls, Virtudes Soriano, José Antonio López-Guerrero, Manuel Moragón, Vicente Oliver); NCI at Cesena, Italy (Paola Minghetti, Laura Fontaine, Katie Beebe, and Giorgio Landi); Genoa, Italy (Giovanna Bianchi-Scarrà, Lorenza Pastorino, Virginia Andreotti, Claudia Martinuzzi, Bruna Dalmasso, Giulia Ciccarese, Francesco Spagnolo, and Paola Queirolo); Riga, Latvia (Kristine Azarjana, Simona Donina, Olita Heisele, Baiba Štreinerte, Aija Ozola and Ludmila Engele); Sydney, Australia (Caroline Watts, Gayathri St. George, Robyn Dalziell, and Kate McBride who assisted with recruitment of study participants; Leo Raudonikis who assisted with data management; and Chantelle Agha-Hamilton and Svetlana Pianova who assisted with bio-specimen management); Montevideo, Uruguay (Virginia Barquet, Javiera Pérez, Miguel Martínez, Jimena Núñez, and Malena Scarone); São Paulo, Brazil (Dirce Maria Carraro, Alexandre Leon Ribeiro de Ávila, Luciana Facure Moredo, Bianca Costa Soares de Sá, Maria Isabel Waddington Achatz, and João Duprat); Porto Alegre, Brazil (Renan Rangel Bonamigo and Maria Carolina Widholzer Rey); Leiden, the Netherlands (Coby Out-Luiting, Clasine van der Drift, Leny van Mourik, Wilma Bergman, Femke de Snoo, Jeanet ter Huurne, and Frans van Nieuwpoort); Queensland, Australia (Nicholas Martin, Grant Montgomery, David Whiteman, Stuart MacGregor, David Duffy and Michael Gattas, along with Judith

Symmons and Harry Beeby who assisted with data management); Stockholm, Sweden (Diana Lindén, RN, for excellent work collecting and entering data into the study data base and Rainer Tuominen for screening of *CDKN2A*); Tel Aviv, Israel (Yael Laitman); Lund, Sweden (Anita Zander, RN, for invaluable help with the data from the Lund Melanoma Study Group and Kari Nielsen, Anna Måsbäck, Katja Harbst, Goran Jonsson, and Åke Borg); and the University of Pennsylvania, Philadelphia, Pennsylvania, USA (Patricia Van Belle, Althea Ruffin, Jillian Knorr and Wenting Zhou). Samples for *CDKN2A* analysis were obtained from the Biobank of the Instituto Valenciano de Oncología. We also wish to thank the French Familial Melanoma Study Group: P. Andry-Benzaquen, B. Bachollet, F. Bérard, P. Berthet, F. Boitier, V. Bonadona, JL. Bonafé, JM. Bonnetblanc, F. Cambazard, O. Caron, F. Caux, J. Chevrant-Breton, A. Chompret (deceased), S. Dalle, L. Demange (deceased), O. Dereure, MX. Doré, MS. Doutre, C. Dugast (deceased), E. Maubec, L. Faivre, F. Grange, Ph. Humbert, P. Joly, D. Kerob, B. Labeille, C. Lasset, MT. Leccia, G. Lenoir, D. Leroux, J. Levang, D. Lipsker, S. Mansard, L. Martin, T. Martin-Denavit, C. Mateus, JL. Michel, P. Morel, L. Olivier-Faivre, JL. Perrot, N. Poulalhon, C. Robert, S. Ronger-Savle, B. Sassolas, P. Souteyrand, D. Stoppa-Lyonnet, L. Thomas, P. Vabres, L. Vincent-Fetita, and E. Wierzbicka. We also thank Hamida Mohamdi for managing the French MELARISK database. We acknowledge the use of the Genetic Counseling Shared Resource at the University of Utah, Salt Lake City, Utah, USA, supported by National Institutes of Health grant P30CA042014 awarded to the Huntsman Cancer Institute. We thank Christophe Blondel at Gustave Roussy for technical assistance in *CDKN2A* genotyping and acknowledge the work of the Gustave Roussy Biobank (BB-0033-00074) in providing DNA resources.

REFERENCES

1. Kefford RF, Newton Bishop JA, Bergman W, et al. Counseling and DNA testing for individuals perceived to be genetically predisposed to melanoma: a consensus statement of the Melanoma Genetics Consortium. *J Clin Oncol*. 1999;17(10):3245-3251.
2. Goldstein AM, Tucker MA. Genetic epidemiology of cutaneous melanoma: a global perspective. *Arch Dermatol*. 2001;137(11):1493-1496.
3. Bishop DT, Demenais F, Goldstein AM, et al. Geographical variation in the penetrance of *CDKN2A* mutations for melanoma. *J Natl Cancer Inst*. 2002;94(12):894-903.
4. Goldstein AM, Tucker MA. Screening for *CDKN2A* mutations in hereditary melanoma. *J Natl Cancer Inst*. 1997;89(10):676-678.
5. Monzon J, Liu L, Brill H, et al. *CDKN2A* mutations in multiple primary melanomas. *N Engl J Med*. 1998;338(13):879-887.
6. Goldstein AM, Chan M, Harland M, et al. Features associated with germline *CDKN2A* mutations: a GenoMEL study of melanoma-prone families from three continents. *J Med Genet*. 2007;44(2):99-106.
7. Demenais F, Mohamdi H, Chaudru V, et al. Association of MC1R variants and host phenotypes with melanoma risk in *CDKN2A* mutation carriers: a GenoMEL study. *J Natl Cancer Inst*. 2010;102(20):1568-1583.

8. Leachman SA, Carucci J, Kohlmann W, et al. Selection criteria for genetic assessment of patients with familial melanoma. *J Am Acad Dermatol*. 2009;61(4):677. e1-14.
9. Niendorf KB, Goggins W, Yang G, et al. MELPREDICT: a logistic regression model to estimate CDKN2A carrier probability. *J Med Genet*. 2006;43(6):501-506.
10. Taylor NJ, Handorf EA, Mitra N, et al. Phenotypic and histopathological tumor characteristics according to CDKN2A mutation status among affected members of melanoma families. *J Invest Dermatol*. 2016;136(5):1066-1069.
11. Harland M, Goldstein AM, Kukulicz K, et al. A comparison of CDKN2A mutation detection within the Melanoma Genetics Consortium (GenoMEL). *Eur J Cancer*. 2008;44(9):1269-1274.
12. Hanley JA, McNeil BJ. The meaning and use of the area under a receiver operating characteristic (ROC) curve. *Radiology*. 1982;143(1):29-36.
13. DeLong ER, DeLong DM, Clarke-Pearson DL. Comparing the areas under two or more correlated receiver operating characteristic curves: a nonparametric approach. *Biometrics*. 1988;44(3):837-845.
14. Pencina MJ, D'Agostino RB, Massaro JM. Understanding increments in model performance metrics. *Lifetime Data Anal*. 2013;19(2):202-218.
15. Leening MJ, Steyerberg EW, Van Calster B, et al. Net reclassification improvement and integrated discrimination improvement require calibrated models: relevance from a marker and model perspective. *Stat Med*. 2014;33(19):3415-3418.
16. Rubin DB. *Multiple Imputation for Nonresponse in Surveys*. New York: John Wiley & Sons Inc; 1987.
17. Taylor NJ, Mitra N, Goldstein AM, et al. Germline variation at CDKN2A and associations with nevus phenotypes among members of melanoma families. *J Invest Dermatol*. 2017;137(12):2606-2612.
18. Lindor NM, Lindor RA, Apicella C, et al. Predicting BRCA1 and BRCA2 gene mutation carriers: comparison of LAMBDA, BRCAPRO, Myriad II, and modified Couch models. *Fam Cancer*. 2007;6(4):473-482.
19. Lindor NM, Johnson KJ, Harvey H, et al. Predicting BRCA1 and BRCA2 gene mutation carriers: comparison of PENN II model to previous study. *Fam Cancer*. 2010;9(4):495-502.
20. Fischer C, Kuchenbacker K, Engel C, et al. Evaluating the performance of the breast cancer genetic risk models BOADICEA, IBIS, BRCAPRO and Claus for predicting BRCA1/2 mutation carrier probabilities: a study based on 7352 families from the German Hereditary Breast and Ovarian Cancer Consortium. *J Med Genet*. 2013;50(6):360-367.
21. Wang W, Niendorf KB, Patel D, et al. Estimating CDKN2A carrier probability and personalizing cancer risk assessments in hereditary melanoma using MelaPRO. *Cancer Res*. 2010;70(2):552-559.
22. Potjer TP, Helgadottir H, Leenheer M, et al. CM-Score: a validated scoring system to predict CDKN2A germline mutations in melanoma families from Northern Europe. *J Med Genet*. 2018. <https://doi.org/10.1136/jmedgenet-2017-105205>.
23. Begg CB, Orlow I, Hummer AJ, et al. Lifetime risk of melanoma in CDKN2A mutation carriers in a population-based sample. *J Natl Cancer Inst*. 2005;97(20):1507-1515.
24. Fiederling J, Shams AZ, Haug U. Validity of self-reported family history of cancer: a systematic literature review on selected cancers. *Int J Cancer*. 2016;139(7):1449-1460.
25. Florell SR, Boucher KM, Garibotti G, et al. Population-based analysis of prognostic factors and survival in familial melanoma. *J Clin Oncol*. 2005;23(28):7168-7177.
26. Soura E, Eliades PJ, Shannon K, et al. Hereditary melanoma: update on syndromes and management: genetics of familial atypical multiple mole melanoma syndrome. *J Am Acad Dermatol*. 2016;74(3):395-407. quiz 408-10.