



ELSEVIER



# Chemically induced cell fate reprogramming and the acquisition of plasticity in somatic cells

Yang Zhao

The nature of somatic cell fate has always been considered relatively unchangeable. Only in rare cases, in response to highly specific environmental cues, do differentiated mammalian somatic cells transform into other cell types. However, the fact that cell fate reprogramming can be accomplished by utilizing chemical cocktails, in the absence of any genetic alterations, suggests that the fate determination of somatic cells is much more malleable than previously believed. The use of chemical cocktails to directly alter cell fate sheds light on an important, yet less explored approach to regenerative medicine: the use of chemicals to restore functions to injured, aging or diseased tissues. Here, we review and discuss the recent developments, inspirations, and challenges encountered when modulating cell fate reprogramming with chemicals, and investigate how chemical biology impacts the future of cell fate reprogramming and regenerative medicine.

## Address

State Key Laboratory of Natural and Biomimetic Drugs, MOE Key Laboratory of Cell Proliferation and Differentiation, Beijing Key Laboratory of Cardiometabolic Molecular Medicine, Center for Life Sciences, Institute of Molecular Medicine, Peking University, Beijing, China

Corresponding author: Zhao, Yang ([yangzhao@pku.edu.cn](mailto:yangzhao@pku.edu.cn))

Current Opinion in Chemical Biology 2019, 51:146–153

This review comes from a themed issue on **Chemical genetics and epigenetics**

Edited by Xiaoguang Lei and Xiang David Li

For a complete overview see the [Issue](#) and the [Editorial](#)

Available online 30th May 2019

<https://doi.org/10.1016/j.cbpa.2019.04.025>

1367-5931/© 2019 Elsevier Ltd. All rights reserved.

## Cell fate reprogramming with chemical compounds

The earliest reported cell fate reprogramming by chemicals in mammalian traces back to the conversion of myocytes from fibroblasts by DNA analogue 5-azacytidine in 1979 [1<sup>\*</sup>], almost a decade before the first transgene-mediated reprogramming in 1987 [2<sup>\*</sup>]. The study of chemicals in directing cell fate conversion intensified after the discovery of induced pluripotent stem cells (iPSCs) and transgene-mediated reprogramming across

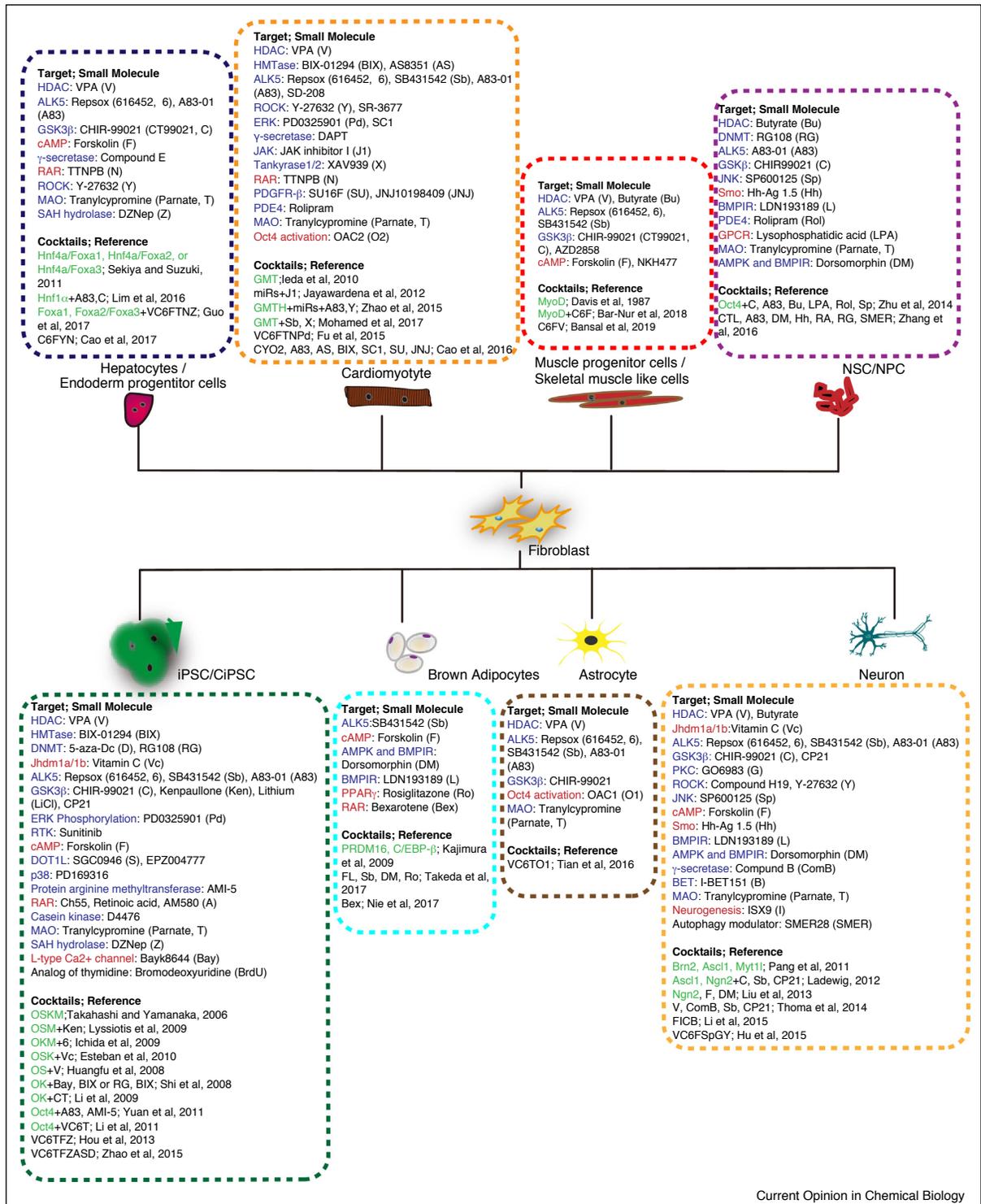
different cell type lineages [3<sup>\*\*</sup>] (Figure 1). Large single and combinatorial screens of hundreds of thousands chemicals identified dozens with the capacity to enhance overall reprogramming efficiency, to replace some of the reprogramming genes during induction, or to benefit the overall quality of iPSCs produced [4–11] (Figure 1).

The discovery of chemical replacers for transgenes and chemical boosters of cell fate reprogramming elicit an intriguing question as to whether reprogramming transgenes can be replaced entirely with chemical compounds. By several rounds of intensive chemical screen, a cocktail of a minimum four chemical compounds, Forskolin, CHIR99021, 616452 and DZNep, has been developed that can reprogram mouse somatic cells into chemically induced pluripotent stem cells (CiPSCs) [12<sup>\*\*</sup>]. In comparison with transgenic methods, chemical approach could be chromosome non-integrative, functionally reversible, cost-efficient, and easier to control and standardize, making it the more pragmatic choice for future applications. Chimeric mice generated from CiPSCs have higher survival rates than mice generated from iPSCs using c-Myc, suggesting that CiPSCs are overall much safer [12<sup>\*\*</sup>]. Besides, CiPSCs bear more epigenetic resemblances to embryonic stem cells than do iPSCs induced with transgenes [13].

Several boosters for chemical reprogramming were further identified, such as TTNPB, Rolipram, UNC0638, and BrdU [12<sup>\*\*</sup>,14]. Chemically induced reprogramming can be fine-tuned through changes in concentration, duration, cocktail composition, and slight changes in structure, allowing the induction efficiency of CiPSCs to be increased [15]. This is evidenced by the dramatically enhanced chemical reprogramming system using an RA agonist, AM580, Dolt11 inhibitors, EPZ004777 and SGC0946, and a Dnmt inhibitor, 5-aza-dC, in a stepwise manner. Moreover, an extraembryonic endoderm (XEN)-like intermediate state was found to mediate chemical reprogramming for somatic cells to CiPSCs [15<sup>\*\*</sup>], even without serum or serum replacement [16]. Chemical reprogramming efficiency can be further increased if the reprogramming steps are divided temporally and controlled more precisely [17].

Not long after the discovery of chemically induced pluripotent stem cells, the chemical reprogramming approach was extended into direct cell lineage reprogramming. Many functional cells such as neurons, neural stem cells, astrocytes, brown adipocytes, skeletal muscle

Figure 1



Current Opinion in Chemical Biology

Representative chemicals or cocktails used in cell fate reprogramming from fibroblasts.

Potential targets of small molecules used in cell fate reprogramming are listed in the dotted box. Red, positive regulation; Blue, negative regulation. Black, downstream effects or potential functions of small molecules. Alias and abbreviations for small molecules are listed in brackets. Shown are the representative chemicals and cocktails used in reprogramming.

progenitors, and cardiomyocytes were reported to be induced from fibroblasts through the use of chemical cocktails without a pluripotent state [18–21,22<sup>••</sup>,23<sup>••</sup>,24–26] (Figure 1). Neural stem cells, gastric epithelial cells, and astrocytes are also used as the initial cell types for chemical reprogramming, either into pluripotent stem cells or directly into differentiated cell types [21,27,28]. Overall, chemical cocktails show their power in generating various ultimate cell types and altering cell fates previously considered stable.

### Molecular dynamics during chemical reprogramming

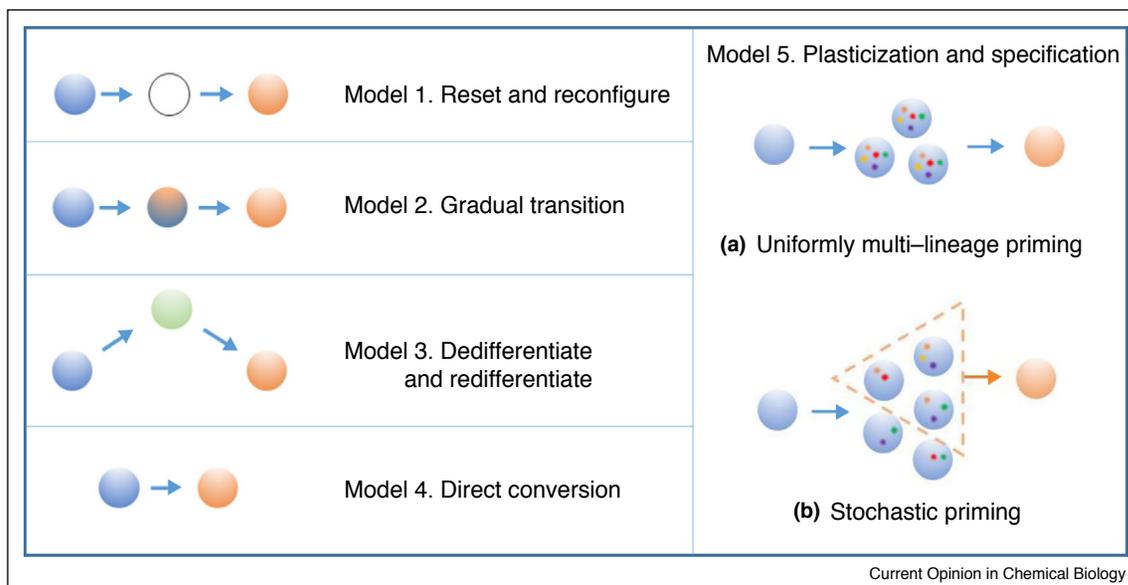
The discovery of the iPSC induction method came as the direct result of the intention to test 24 pluripotency-associated genes. And Yamanaka factors, the final four, are all pluripotency-associated factors that highly expressed in embryonic stem cells and known to be important regulators of pluripotency [29<sup>••</sup>]. Direct cell lineage reprogramming factor cocktails were also developed based on the idea of inducing the target cell fates by the delivery of target cell type-associated master transcription factors. The master transcription factors may act as pioneer factors to directly shape the cell type favorable chromatin accessibility profiles and activates the cell type-specific gene expression program [29–31]. In contrast, the chemical compounds used in chemical reprogramming are not obviously associated with specific

cell fates, and directly target proteins involving signaling transduction pathways or epigenetic factors that play roles in multiple biological processes (Figure 1). This raises the question of how exactly the target cell type is determined by each chemical cocktail, and what exactly happens during chemical reprogramming that determines cell fate.

Study of the cellular dynamics indicated that unlike transgenic reprogramming, chemically induced reprogramming undergo a unique roadmap to achieve pluripotency mediated by an XEN-like state [15<sup>••</sup>]. For XEN specification, the core chemicals in chemical reprogramming, Forskolin, CHIR99021 and 616452, first activate the expression of *Sox17* in an initial plasticity acquisition stage, and further support the endogenous activation of other essential XEN cell-specific transcription factors, such as *Gata4*, *Sall4* and *Foxa2*, thereby established the entire network of XEN cells (Yang *et al.*, unpublished) (Figure 2).

After XEN-like cells are generated by chemical cocktails, DZNep, a SAH hydrolase inhibitor, can initiate the expression of Oct4, probably by suppressing histone and DNA methylation, following the activation of upstream factors *Sall4* and *Sox2* (or *Gata4*) in the first stage of reprogramming [12<sup>••</sup>,15<sup>••</sup>]. Pluripotency factors, such as Oct4, form positive feedback regulatory networks with master genes of embryonic 2-cell (2C) state, bolstering a 2C-like state from XEN-like cells that further

Figure 2



Potential models for cell fate reprogramming.

The blue ball represents initial cell type, and orange ball represents the ultimate cell type during cell fate reprogramming. Different kinds of intermediated cells are proposed as shown. Cell fate reprogramming in model 1 was mediated by an intermediate cell without the feature of initial or ultimate cells. In model 2, reprogramming is mediated by a hybrid cell of initial and ultimate cell type. Model 3 reprogramming is mediated by a stem cell, which is developmentally more naive than initial and ultimate cell type (green ball). Model 4 reprogramming have no intermediate cell type. In model 5, reprogramming was mediated by a priming state, in which multi-lineage factors (colorful dots) are activated for further specification into ultimate cell types. 5a indicates a uniformly primed cell state, while 5b indicates stochastic priming of chemical reprogramming, in which the multi-lineage gene activation is stochastic and dispersed in different cells.

promotes CiPSC induction [17]. In summary, a hierarchical regulation circuitry linking the sequential activation of master transcription factors was revealed throughout chemical reprogramming process (Figure 3).

### Cell plasticity induced with chemicals

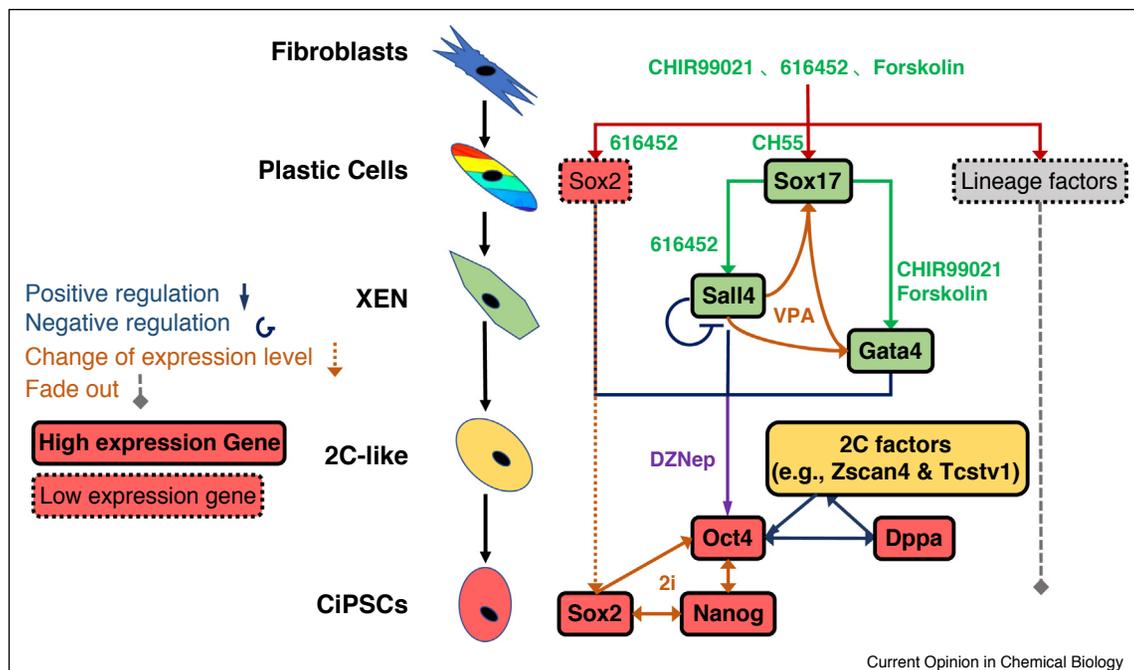
The term cell plasticity has traditionally been used to describe either the potential of cells to change their fates or the ability of cancer cells to transition between epithelial and mesenchymal states. The acquisition of plasticity in a somatic cell is necessary in somatic reprogramming; this may counteract the machinery for the safeguarding of cell identity. Histone chaperons, CAF1 and FACT, and Cbp9-mediated sumoylation modification which stabilize the epigenetic state of a cell identity are barriers in somatic reprogramming [32–34]. The p53-mediated tumor suppressor pathway was reported as a general barrier in different reprogramming systems, which may involve in cell fate stabilization [35,36\*,37–39].

Some chemicals used in reprogramming to obtain different cell types may act in the acquisition of cell plasticity, rather than the choice of ultimate cell fate. For instance, a Brd4 inhibitor, I-BET-151, was initially found to disrupt the cell identity of fibroblasts during reprogramming toward functional neurons [23\*\*]. This was then found to be useful for directing cellular reprogramming into iPSCs [40]. HDAC

inhibitors were also found to be beneficial in a lot of reprogramming systems (Figure 1). Interestingly, the core chemical cocktail that initially discovered in CiPSC generation, CHIR99021, 616452 and Forskolin, are also widely used in various reprogramming systems into different lineages, suggesting their broad roles in inducing cell plasticity rather than determine the cell fate directions (Figure 1).

Detailed investigation of the chemically induced cell fate transition process from fibroblasts to XEN cells uncovered two major steps, plasticity acquisition and cell fate specification (Yang *et al.*, unpublished). In the plasticity acquisition step, a chemically induced multi-lineage priming (CiMP) state initiates, in which the endogenous expression of a wide range of transcription factors not-specific to a certain cell fate is activated. Expression of these transcription factors is stochastically dispersed in different cells, and have overall low correlations with each other, which could be a unique model in cell fate reprogramming (model 5b in Figure 2). Cells in a CiMP state are plastic and can be rewired into different other cell types by fine-tuning of the cocktails and mediums of the subsequent stage, providing a shortcut for obtaining functional desirable cell types in regenerative medicine, bypassing long-term culture in iPSC inductions and directed differentiation mimicking developmental process (Yang *et al.*, unpublished). As these chemicals are all

Figure 3



Transcriptional activation circuitry in chemical reprogramming.

Schematic diagram for transcriptional activation circuitry in chemical reprogramming process. Transcription factors are hierarchically regulated and stimulated through a XEN-like state and 2C-like state. The chemicals function at different stages and play different roles on endogenous gene activation. Multi-lineage factors are activated as the side effects in the initial stage of reprogramming, which can be rewired into other cell fates (Yang *et al.*, unpublished).

commonly known as cell signaling modulators, the question remains as to how these chemicals function to induce such a plastic state on a molecular level.

### Challenges and new thoughts in developing chemical cocktails for reprogramming

Since the traditional strategy to develop chemical compounds such as phenotypic screening is labor intensive and time-consuming, an effective strategy to develop chemical reprogramming cocktails is still needed. In accordance with the fact that, during chemical-induced reprogramming, cells pass through multiple discrete stages, a more effective way to discover chemicals for reprogramming could involve adjusting the small molecule cocktails during cell specification process after chemically inducing cell plasticity. In addition, since the chemical reprogramming process may be mediated by hierarchical activation of transcription factors, it could be valuable to screen for potential chemical candidates by activating the endogenous expression of one or more master transcription factors, based on each specific cell type. In addition, the reprogramming barriers identified in mechanism study on somatic reprogramming could also be potential targets for use of target-based chemical screening. However, because chemical reprogramming always requires a combo of more than 2–3 chemicals, it is still a challenge to systematically ‘design’ specific cocktails but not a single candidate compound for cell fate reprogramming.

### Opportunities and challenges of chemical regeneration *in situ*

One of the ultimate goals of chemical reprogramming is to develop drugs that enable direct reprogramming and regeneration *in vivo* to repair damaged tissues. In recent years, evidence of *in vivo* reprogramming has been shown in mouse models through both genetic manipulation [41<sup>\*</sup>,42–46] and chemical compounds [47<sup>\*</sup>,48]. The micro-environments *in vivo* may provide specific opportunities for cell fate reprogramming. For instances, *in vivo* iPSC generation was found to be promoted by IL-6 secretion from senescent cells in the niche [49]. Injury signaling may play an essential role to direct reprogramming from Muller glia cells to retinal neurons [50]. Direct reprogramming toward pancreatic beta cells, T cells, and hematopoietic stem cells was achieved only *in vivo*, and has not been achieved *in vitro* [41<sup>\*</sup>,51,52]. Moreover, *in vivo* activated astrocyte exhibits more similarities with neural stem cells after injury, and thereby may be more amenable for reprogramming [53]. In addition, cell types *in vivo* may be more heterogeneous, in contrast to cloned cells *in vitro*, which could have more chances to cell fate reprogramming in response to stimuli or chemicals. *In vivo* cell fate adaptive reprogramming naturally occurs in mammals in response to injury [54–56,57], the mechanism of which may be manipulated by chemicals. These are opportunities to induce *in situ* reprogramming.

However, there are also several challenges to develop chemicals for *in situ* reprogramming. For example, chemical cocktails developed *in vitro* may not work *in vivo* because the *in vitro* cultured cell types may not have exact counterparts *in vivo*. Chemicals may lead to unintended off-target effects in certain cell types in the heterogeneous *in vivo* environment. It could be ideal if chemical cocktails can be developed to have activities only in excessively deposited cell types, such as myofibroblasts in the injured region or activated astrocytes, since different cell types may have different responses with use of the same chemicals. Otherwise, targeted deliver methods of chemicals might be required to reduce the risks in clinical applications. Moreover, the partially reprogrammed cells and unintended cell products may be induced *in vivo*, which could be harmful for tissue repair. Further efforts and new strategies are necessary to improve *in vivo* reprogramming efficiency, to efficiently deliver compounds locally for drug sustained release and even to reduce the number of compounds, before *in situ* chemical reprogramming can be widely used in regenerative medicine.

### Conclusions and perspective

Even before the discovery of chemically induced cell reprogramming, small molecules have been used to regulate natural biological processes, such as in stem cell self-renewal and differentiation, even *in vivo*. However, it is only due to the establishment of a method to chemically direct the reprogramming of a cell fate that we now have the means to produce functional cells from more available cell sources both *in vitro* and *in vivo*, even without a relying on the use of genetic modifications. Hopefully this will pave the way for more efficient use of cell fate manipulation and cell fate engineering in furthering future research in drug screens, drug evaluation and in regenerative medicine, by cell transplantation or pharmacological reprogramming (Figure 4).

Moreover, the recent developments on chemical reprogramming make it possible to ask new questions and to further our understanding of cell fate determination and reprogramming. For example, is there any chemical cocktail that functions as a general eraser of a cell fate by breaking down the cell fate keepers? Could chemical cocktails reprogram cell aging, without changing cell type? Could chemical reprogramming be further extended into cancer therapy by altering the fate of cancer cells into a more differentiated one or more normal one, just like reported very recently [58,59,60<sup>\*</sup>,61]? In addition, some traditional concepts may also need to be reconsidered. Because chemically treated somatic cells exhibit plasticity similar to that of stem cells, how can we distinguish the differences between reprogrammable somatic cells and adult stem cells that can be activated to change their fates? [62]

Overall, the use of chemicals to induce cell fate reprogramming has revolutionized our concepts of cell



3. Takahashi K, Yamanaka S: **Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors.** *Cell* 2006, **126**:663-676.

The first report on somatic reprogramming to pluripotency with defined factors.

4. Bar-Nur O, Brumbaugh J, Verheul C, Apostolou E, Pruteanu-Malinici I, Walsh RM, Ramaswamy S, Hochedlinger K: **Small molecules facilitate rapid and synchronous ipsc generation.** *Nat Methods* 2014, **11**:1170-1176.
5. Huangfu D, Osafune K, Maehr R, Guo W, Eijkelenboom A, Chen S, Muhlestein W, Melton DA: **Induction of pluripotent stem cells from primary human fibroblasts with only Oct4 and Sox2.** *Nat Biotechnol* 2008, **26**:1269-1275.
6. Stadtfeld M, Apostolou E, Ferrari F, Choi J, Walsh RM, Chen T, Ooi SS, Kim SY, Bestor TH, Shioda T *et al.*: **Ascorbic acid prevents loss of Dlk1-Dio3 imprinting and facilitates generation of all-IPS cell mice from terminally differentiated B cells.** *Nat Genet* 2012, **44**:398-405 S391-392.
7. Li Y, Zhang Q, Yin X, Yang W, Du Y, Hou P, Ge J, Liu C, Zhang W, Zhang X *et al.*: **Generation of iPSCs from mouse fibroblasts with a single gene, Oct4, and small molecules.** *Cell Res* 2011, **21**:196-204.
8. Li W, Zhou H, Abujarour R, Zhu S, Young Joo J, Lin T, Hao E, Scholer HR, Hayek A, Ding S: **Generation of human-induced pluripotent stem cells in the absence of exogenous Sox2.** *Stem Cells* 2009, **27**:2992-3000.
9. Lyssiotis CA, Foreman RK, Staerk J, Garcia M, Mathur D, Markoulaki S, Hanna J, Lairson LL, Charette BD, Bouchez LC *et al.*: **Reprogramming of murine fibroblasts to induced pluripotent stem cells with chemical complementation of Klf4.** *Proc Natl Acad Sci U S A* 2009, **106**:8912-8917.
10. Shi Y, Do JT, Despons C, Hahm HS, Scholer HR, Ding S: **A combined chemical and genetic approach for the generation of induced pluripotent stem cells.** *Cell Stem Cell* 2008, **2**:525-528.
11. Li X, Xu J, Deng H: **Small molecule-induced cellular fate reprogramming: promising road leading to rome.** *Curr Opin Genet Dev* 2018, **52**:29-35.
12. Hou P, Li Y, Zhang X, Liu C, Guan J, Li H, Zhao T, Ye J, Yang W, Liu K *et al.*: **Pluripotent stem cells induced from mouse somatic cells by small-molecule compounds.** *Science* 2013, **341**:651-654.
- The first somatic reprogramming to pluripotency induced with pure chemicals.
13. Ping W, Hu J, Hu G, Song Y, Xia Q, Yao M, Gong S, Jiang C, Yao H: **Genome-wide DNA methylation analysis reveals that mouse chemical ipscs have closer epigenetic features to mescs than oskm-integrated ipscs.** *Cell Death Dis* 2018, **9**:187.
14. Long Y, Wang M, Gu H, Xie X: **Bromodeoxyuridine promotes full-chemical induction of mouse pluripotent stem cells.** *Cell Res* 2015, **25**:1171-1174.
15. Zhao Y, Zhao T, Guan J, Zhang X, Fu Y, Ye J, Zhu J, Meng G, Ge J, Yang S *et al.*: **A xen-like state bridges somatic cells to pluripotency during chemical reprogramming.** *Cell* 2015, **163**:1678-1691.
- This report proved that chemical reprogramming into pluripotency undergo a fundamentally different molecular pathway through a XEN-like state, in comparison to transgene-induced somatic reprogramming.
16. Cao S, Yu S, Li D, Ye J, Yang X, Li C, Wang X, Mai Y, Qin Y, Wu J *et al.*: **Chromatin accessibility dynamics during chemical induction of pluripotency.** *Cell Stem Cell* 2018, **22**:529-542.e5.
17. Zhao T, Fu Y, Zhu J, Liu Y, Zhang Q, Yi Z, Chen S, Jiao Z, Xu X, Xu J *et al.*: **Single-cell RNA-seq reveals dynamic early embryonic-like programs during chemical reprogramming.** *Cell Stem Cell* 2018, **23**:31-45.e7.
18. Zhang M, Lin YH, Sun YJ, Zhu S, Zheng J, Liu K, Cao N, Li K, Huang Y, Ding S: **Pharmacological reprogramming of fibroblasts into neural stem cells by signaling-directed transcriptional activation.** *Cell Stem Cell* 2016, **18**:653-667.
19. Cheng L, Hu W, Qiu B, Zhao J, Yu Y, Guan W, Wang M, Yang W, Pei G: **Generation of neural progenitor cells by chemical cocktails and hypoxia.** *Cell Res* 2014, **24**:665-679.
20. Tian E, Sun G, Sun G, Chao J, Ye P, Warden C, Riggs AD, Shi Y: **Small-molecule-based lineage reprogramming creates functional astrocytes.** *Cell Rep* 2016, **16**:781-792.
21. Zhang L, Yin JC, Yeh H, Ma NX, Lee G, Chen XA, Wang Y, Lin L, Chen L, Jin P *et al.*: **Small molecules efficiently reprogram human astroglial cells into functional neurons.** *Cell Stem Cell* 2015, **17**:735-747.
22. Hu W, Qiu B, Guan W, Wang Q, Wang M, Li W, Gao L, Shen L, Huang Y, Xie G *et al.*: **Direct conversion of normal and Alzheimer's disease human fibroblasts into neuronal cells by small molecules.** *Cell Stem Cell* 2015, **17**:204-212.
- One of the first reports on chemically induced cell fate direct reprogramming across germ layers, and first chemical reprogramming system in human.
23. Li X, Zuo X, Jing J, Ma Y, Wang J, Liu D, Zhu J, Du X, Xiong L, Du Y *et al.*: **Small-molecule-driven direct reprogramming of mouse fibroblasts into functional neurons.** *Cell Stem Cell* 2015, **17**:195-203.
- One of the first reports on chemically induced cell fate direct reprogramming across germ layers.
24. Nie B, Nie T, Hui X, Gu P, Mao L, Li K, Yuan R, Zheng J, Wang H, Li K *et al.*: **Brown adipogenic reprogramming induced by a small molecule.** *Cell Rep* 2017, **18**:624-635.
25. Takeda Y, Harada Y, Yoshikawa T, Dai P: **Direct conversion of human fibroblasts to brown adipocytes by small chemical compounds.** *Sci Rep* 2017, **7**:4304.
26. Han X, Yu H, Huang D, Xu Y, Saadatpour A, Li X, Wang L, Yu J, Pinello L, Lai S *et al.*: **A molecular roadmap for induced multi-lineage trans-differentiation of fibroblasts by chemical combinations.** *Cell Res* 2017, **27**:843.
27. Wang Y, Qin J, Wang S, Zhang W, Duan J, Zhang J, Wang X, Yan F, Chang M, Liu X *et al.*: **Conversion of human gastric epithelial cells to multipotent endodermal progenitors using defined small molecules.** *Cell Stem Cell* 2016, **19**:449-461.
28. Ye J, Ge J, Zhang X, Cheng L, Zhang Z, He S, Wang Y, Lin H, Yang W, Liu J *et al.*: **Pluripotent stem cells induced from mouse neural stem cells and small intestinal epithelial cells by small molecule compounds.** *Cell Res* 2016, **26**:34-45.
29. Wapinski OL, Vierbuchen T, Qu K, Lee QY, Chanda S, Fuentes DR, Giresi PG, Ng YH, Marro S, Neff NF *et al.*: **Hierarchical mechanisms for direct reprogramming of fibroblasts to neurons.** *Cell* 2013, **155**:621-635.
30. Wapinski OL, Lee QY, Chen AC, Li R, Corces MR, Ang CE, Treutlein B, Xiang C, Baubet V, Suchy FP *et al.*: **Rapid chromatin switch in the direct reprogramming of fibroblasts to neurons.** *Cell Rep* 2017, **20**:3236-3247.
31. Morris SA: **Direct lineage reprogramming via pioneer factors; a detour through developmental gene regulatory networks.** *Development* 2016, **143**:2696-2705.
32. Cheloufi S, Elling U, Hopfgartner B, Jung YL, Murn J, Ninova M, Hubmann M, Badeaux AI, Euong Ang C, Tenen D *et al.*: **The histone chaperone CAF-1 safeguards somatic cell identity.** *Nature* 2015, **528**:218-224.
33. Cossec JC, Theurillat I, Chica C, Bua Aguin S, Gaume X, Andrieux A, Iturbide A, Jouvion G, Li H, Bossis G *et al.*: **SUMO safeguards somatic and pluripotent cell identities by enforcing distinct chromatin states.** *Cell Stem Cell* 2018, **23**:742-757.e8.
34. Kolundzic E, Ofenbauer A, Bulut SI, Uyar B, Baytek G, Sommermeier A, Seelk S, He M, Hirsekorn A, Vucicevic D *et al.*: **Fact sets a barrier for cell fate reprogramming in caenorhabditis elegans and human cells.** *Dev Cell* 2018, **46**:611-626.e12.
35. Jain AK, Barton MC: **P53: emerging roles in stem cells, development and beyond.** *Development* 2018, **145**.

36. Zhao Y, Yin X, Qin H, Zhu F, Liu H, Yang W, Zhang Q, Xiang C, Hou P, Song Z *et al.*: **Two supporting factors greatly improve the efficiency of human iPSC generation.** *Cell Stem Cell* 2008, **3**:475-479.  
 The first report to identify p53 as major barriers in somatic reprogramming to pluripotency.
37. Du Y, Wang J, Jia J, Song N, Xiang C, Xu J, Hou Z, Su X, Liu B, Jiang T *et al.*: **Human hepatocytes with drug metabolic function induced from fibroblasts by lineage reprogramming.** *Cell Stem Cell* 2014, **14**:394-403.
38. Batta K, Florkowska M, Kouskoff V, Lacaud G: **Direct reprogramming of murine fibroblasts to hematopoietic progenitor cells.** *Cell Rep* 2014, **9**:1871-1884.
39. Jiang H, Xu Z, Zhong P, Ren Y, Liang G, Schilling HA, Hu Z, Zhang Y, Wang X, Chen S *et al.*: **Cell cycle and p53 gate the direct conversion of human fibroblasts to dopaminergic neurons.** *Nat Commun* 2015, **6**:10100.
40. Shao Z, Yao C, Khodadadi-Jamayran A, Xu W, Townes TM, Crowley MR, Hu K: **Reprogramming by de-bookmarking the somatic transcriptional program through targeting of bet bromodomains.** *Cell Rep* 2016, **16**:3138-3145.
41. Zhou Q, Brown J, Kanarek A, Rajagopal J, Melton DA: **In vivo reprogramming of adult pancreatic exocrine cells to beta-cells.** *Nature* 2008, **455**:627-632.  
 The first report on direct *in vivo* reprogramming with transgenes.
42. Qian L, Huang Y, Spencer CI, Foley A, Vedantham V, Liu L, Conway SJ, Fu JD, Srivastava D: **In vivo reprogramming of murine cardiac fibroblasts into induced cardiomyocytes.** *Nature* 2012, **485**:593-598.
43. Song K, Nam YJ, Luo X, Qi X, Tan W, Huang GN, Acharya A, Smith CL, Tallquist MD, Neilson EG *et al.*: **Heart repair by reprogramming non-myocytes with cardiac transcription factors.** *Nature* 2012, **485**:599-604.
44. Guo Z, Zhang L, Wu Z, Chen Y, Wang F, Chen G: **In vivo direct reprogramming of reactive glial cells into functional neurons after brain injury and in an Alzheimer's disease model.** *Cell Stem Cell* 2014, **14**:188-202.
45. Song G, Pacher M, Balakrishnan A, Yuan Q, Tsay HC, Yang D, Reetz J, Brandes S, Dai Z, Putzer BM *et al.*: **Direct reprogramming of hepatic myofibroblasts into hepatocytes in vivo attenuates liver fibrosis.** *Cell Stem Cell* 2016, **18**:797-808.
46. Rezvani M, Espanol-Suner R, Malato Y, Dumont L, Grimm AA, Kienle E, Bindman JG, Wiedtke E, Hsu BY, Naqvi SJ *et al.*: **In vivo hepatic reprogramming of myofibroblasts with AAV vectors as a therapeutic strategy for liver fibrosis.** *Cell Stem Cell* 2016, **18**:809-816.
47. Huang C, Tu W, Fu Y, Wang J, Xie X: **Chemical-induced cardiac reprogramming in vivo.** *Cell Res* 2018, **28**:686-689.  
 The first report on direct *in vivo* reprogramming with pure chemicals.
48. Yin JC, Zhang L, Ma NX, Wang Y, Lee G, Hou XY, Lei ZF, Zhang FY, Dong FP, Wu GY *et al.*: **Chemical conversion of human fetal astrocytes into neurons through modulation of multiple signaling pathways.** *Stem Cell Rep* 2019, **12**:488-501.
49. Chiche A, Le Roux I, von Joest M, Sakai H, Aguin SB, Cazin C, Salam R, Fiette L, Alegria O, Flamant P *et al.*: **Injury-induced senescence enables in vivo reprogramming in skeletal muscle.** *Cell Stem Cell* 2017, **20**:407-414.e4.
50. Jorstad NL, Wilken MS, Grimes WN, Wohl SG, VandenBosch LS, Yoshimatsu T, Wong RO, Rieke F, Reh TA: **Stimulation of functional neuronal regeneration from muller glia in adult mice.** *Nature* 2017, **548**:103-107.
51. Zhang M, Dong Y, Hu F, Yang D, Zhao Q, Lv C, Wang Y, Xia C, Weng Q, Liu X *et al.*: **Transcription factor Hoxb5 reprograms B cells into functional T lymphocytes.** *Nat Immunol* 2018, **19**:279-290.
52. Sugimura R, Jha DK, Han A, Soria-Valles C, da Rocha EL, Lu YF, Goettel JA, Serrao E, Rowe RG, Malleshaiah M *et al.*: **Haematopoietic stem and progenitor cells from human pluripotent stem cells.** *Nature* 2017, **545**:432-438.
53. Gotz M, Sirko S, Beckers J, Irmeler M: **Reactive astrocytes as neural stem or progenitor cells: in vivo lineage, in vitro potential, and genome-wide expression analysis.** *Glia* 2015, **63**:1452-1468.
54. Merrell AJ, Stanger BZ: **Adult cell plasticity in vivo: de-differentiation and transdifferentiation are back in style.** *Nat Rev Mol Cell Biol* 2016, **17**:413-425.
55. Deng X, Zhang X, Li W, Feng RX, Li L, Yi GR, Zhang XN, Yin C, Yu HY, Zhang JP *et al.*: **Chronic liver injury induces conversion of biliary epithelial cells into hepatocytes.** *Cell Stem Cell* 2018, **23**:114-122.e3.
56. Jessen KR, Mirsky R, Arthur-Farraj P: **The role of cell plasticity in tissue repair: adaptive cellular reprogramming.** *Dev Cell* 2015, **34**:613-620.
57. Bonfanti P, Claudinot S, Amici AW, Farley A, Blackburn CC, Barrandon Y: **Microenvironmental reprogramming of thymic epithelial cells to skin multipotent stem cells.** *Nature* 2010, **466**:978-982.
58. Cheng Z, He Z, Cai Y, Zhang C, Fu G, Li H, Sun W, Liu C, Cui X, Ning B *et al.*: **Conversion of hepatoma cells to hepatocyte-like cells by defined hepatocyte nuclear factors.** *Cell Res* 2019, **29**:124-135.
59. Lee C, Robinson M, Willerth SM: **Direct reprogramming of glioblastoma cells into neurons using small molecules.** *ACS Chem Neurosci* 2018, **9**:3175-3185.
60. Pattabiraman DR, Bieri B, Kober KI, Thiru P, Krall JA, Zill C, Reinhardt F, Tam WL, Weinberg RA: **Activation of PKA leads to mesenchymal-to-epithelial transition and loss of tumor-initiating ability.** *Science* 2016, **351**:aad3680.  
 Proof-of-principle study for chemically inducing a MET as differentiation therapy for tumor-initiating cells.
61. Park NI, Guilhamon P, Desai K, McAdam RF, Langille E, O'Connor M, Lan X, Whetstone H, Coutinho FJ, Vanner RJ *et al.*: **Ascl1 reorganizes chromatin to direct neuronal fate and suppress tumorigenicity of glioblastoma stem cells.** *Cell Stem Cell* 2017, **21**:411.
62. Raven A, Lu WY, Man TY, Ferreira-Gonzalez S, O'Duibhir E, Dwyer BJ, Thomson JP, Meehan RR, Bogorad R, Kotliansky V *et al.*: **Cholangiocytes act as facultative liver stem cells during impaired hepatocyte regeneration.** *Nature* 2017, **547**:350-354.