

Review

Circulating microRNAs to predict heart failure after acute myocardial infarction in women

Torkia Lalem, Yvan Devaux*

Cardiovascular Research Unit, Luxembourg Institute of Health, Luxembourg



ARTICLE INFO

Keywords:

Heart failure
Acute myocardial infarction
Left ventricular remodeling
Sex
Biomarker
microRNA

ABSTRACT

Left ventricular remodeling after acute myocardial infarction affects cardiac function and increases the risk of developing heart failure. Despite the emergence of biomarkers associated with remodeling, the ideal biomarker to accurately predict the risk of developing heart failure after acute myocardial infarction is still to be discovered. Female and male hearts cope differently with ischemic stress, leading to different consequences on cardiac morphology and function. As biomarkers reflect the pathogenesis of remodeling, utilization of sex-specific biomarkers might improve risk stratification. Expressed in cardiac and inflammatory cells, microRNAs regulate several biological pathways triggering the remodeling process. In addition, circulating microRNAs are associated with the risk of developing heart failure after acute myocardial infarction, hence their biomarker potential. Interestingly, multiple microRNAs display sex-specific expression profiles as they can be modulated by sexual hormones and escape X-inactivation, for those located on the X-chromosome. This review article aims to discuss the potential of circulating microRNAs to predict heart failure after acute myocardial infarction in a sex-specific manner.

1. Introduction

Cardiovascular disease (CVD) is the first cause of mortality Worldwide [1]. Half of deaths are imputable to coronary artery disease (CAD) which kills as many men as women [1,2]. Among CAD, acute myocardial infarction (AMI) plays a major role and is associated with a high risk of developing left ventricular (LV) remodeling (LVR) leading to cardiac dysfunction and heart failure (HF). Identification of patients at high risk of LVR represents an unmet clinical need and could aid reducing the incidence of post-AMI HF. Indeed, being able to stratify patients according to their risk of developing adverse outcomes after AMI would allow tailoring healthcare to each individual, optimizing the use of imaging technologies, novel drugs, and planning a close follow-up with frequent re-hospitalization in high-risk patients. Novel biomarkers of outcome after AMI are required to aid in moving towards implementation of personalized medicine.

Multiple specificities linked to sex have been shown in AMI risk factors, presentation features and prognosis [3–5]. Male and female hearts do not equally respond to ischemic stress leading to sex-related

differences in the consequences of AMI, notably related to the changes in cardiac morphology and function [6]. As biomarkers reflect the underlying disease process, sex-specific biomarkers of LVR have the potential to improve risk stratification of patients and allow a gender-based personalized medicine.

Non-coding RNAs (ncRNAs) among which microRNAs (miRNAs) emerged as putative regulators and potential biomarkers of LVR [7]. Interestingly, miRNAs show sex-specific expression profiles due either to their escape of X-inactivation for those located on the X-chromosome or to modulation by sexual hormones [8,9]. This review presents the sex-disparities in AMI and LVR and discusses the potential of miRNAs as sex-specific predictors of LVR development following AMI.

2. Sex-disparities in AMI

Analysis of patient cohorts and in vivo experimental studies revealed multiple sex-disparities in AMI patients related to risk factors, pathophysiology, diagnosis and prognosis. Women with AMI are older than men and present more comorbidities and risk factors, particularly

Abbreviations: CVD, cardiovascular disease; CAD, coronary artery disease; AMI, acute myocardial infarction; LV, left ventricular; LVR, left ventricular remodeling; ncRNA, non-coding RNA; miRNA, microRNA; HF, heart failure; ICM, ischemic cardiomyopathy; BNP, Brain natriuretic peptide; NT-proBNP, N terminal prohormone of brain natriuretic peptide; AUC, area under the receiver operating characteristic curve; AIC, Akaike information criterion; lncRNA, long non-coding RNA; IL, interleukin; ECs, endothelial cells; ER, estrogen receptor

* Corresponding author at: Cardiovascular Research Unit, Luxembourg Health Institute, L1445, Luxembourg.

E-mail address: yvan.devaux@lih.lu (Y. Devaux).

<https://doi.org/10.1016/j.clinbiochem.2019.05.011>

Received 26 February 2019; Received in revised form 22 May 2019; Accepted 23 May 2019

Available online 24 May 2019

0009-9120/ © 2019 The Canadian Society of Clinical Chemists. Published by Elsevier Inc. All rights reserved.

diabetes and hypertension [5,10]. Furthermore, angiographic studies showed that female patients present less extensive CAD [11], more plaque erosion [12] and non-occlusive CAD (15% versus 8%) than their male counterparts [13]. Although chest pain is the predominant symptom of AMI in both sexes, atypical symptoms are more common among women [3,4,14]. Moreover, women are less often diagnosed with ST-segment elevation myocardial infarction than men (27% versus 37%) [15]. Women have longer ischemic time (i.e. delay between chest pain onset and reperfusion) and treatment times than men, especially among those aged < 60 years old [16]. The higher mortality rate in women after AMI has been mainly assigned to an older age at event time and accumulation of comorbidities [15,17]. Nevertheless, despite a matching for age and risk factors in *ISAR-RISK* and *ART* studies, women still had higher 1-year mortality [18]. The *VALLANT* study including > 10,000 patients with myocardial infarction and diagnosed for HF or asymptomatic systolic dysfunction showed higher risk of HF and combined outcome of cardiovascular death or HF hospitalization among women, although this risk appeared to be independent from differences in LVR between women and men [5]. Thus, whether different outcomes after AMI between women and men are direct consequences of different LVR is unclear and most probably include other important parameters such as ischemic time and comorbidities.

3. Sex-disparities in LVR

Animal models showed that male and female hearts do not evenly respond to ischemic stress. Females had smaller infarct size, less necrosis and apoptosis, better contractility and better LV function recovery [19]. In addition, coronary occlusion in hypertensive female rats led to concentric hypertrophy without thinning and dilation of the infarct zone, whereas males were more prone to eccentric LV hypertrophy with thinning and dilation of the infarct zone [20]. On the other hand, males had a delayed healing process, which predisposes to ventricular rupture and exaggerated LVR [21].

In human, and depending on the study cohorts, between fifteen to 30% of AMI patients develop LVR, with comparable incidence among men and women [22,23]. Post-mortem analyses revealed that men had 10 times higher apoptosis in the infarct border zone compared to women [24]. In another study addressing the association between sex, reperfusion and LVR, angioplasty was more efficient in women who had a better myocardial recovery, a smaller infarct size and less microvascular deterioration [25]. In a group of patients with end-stage HF, sex-disparities in LVR were dependent on the aetiology of HF [6]. Indeed, while no gender-biased differences were observed in patients with idiopathic cardiomyopathy, heart weight and LV mass index was greater in men than women with ischemic cardiomyopathy (ICM). This difference was assigned to fundamental cellular remodeling, with men having 2-fold more cardiomyocytes hypertrophy [6]. Thus, LVR differs between women and men and this may be reflected in biomarker levels.

4. Sex-disparities in biomarkers of LV remodeling

Brain natriuretic peptide (BNP) and its N-terminal fraction NT-proBNP have been widely studied for their ability to diagnose HF and predict LVR after AMI, yet their potential to act in a sex-specific manner is still unclear. Sex affects the circulating levels of these peptides, with women having higher levels independently of age [26–28]. This is mainly due to stimulatory effects of estrogens and lower hemoglobin levels in women [28,29]. Cardiac troponins are secreted at lower levels in women compared to men [30], presumably because of a lower mass of the female left ventricle [31]. Nevertheless, a recent study showed that higher cardiac troponin levels in men persisted even after adjustment with LV mass [32]. Cardiac troponins were shown to be associated with LVR in several patient cohorts, although the influence of sex on this association was not thoroughly addressed [33,34].

Among other potential biomarkers of LVR, both galectin 3 and the

interleukin (IL) 33 receptor ST2 (suppression of tumorigenicity 2) showed sex-biased levels. Community studies showed that women had higher levels of galectin 3 leading to the utilization of sex-specific cut-offs for the analyses [35,36]. Conversely, women have been reported to have less ST2 than men and this difference was attributed to female sex-hormones [37]. Despite these differences, the association between these biomarkers and LVR has not been addressed separately in men and women. Recently, a novel female-specific biomarker of LVR after AMI has been reported. This biomarker is a messenger RNA encoding the cyclin dependent kinase inhibitor 1C, a regulator of cell proliferation [38]. This finding supports the potential of RNA molecules as sex-specific biomarkers of outcome after AMI.

5. miRNAs as biomarkers of LVR

The RNA family can be dichotomized into protein-coding RNAs and ncRNAs. Non-coding RNAs have been further classified into small (< 200 nucleotides) and long (> 200 nucleotides) RNAs. Small RNAs include the well-known miRNAs which are widely expressed in the cardiovascular system and are involved in numerous pathophysiological processes through down-regulation of gene expression [7,39]. Circulating miRNAs are mostly present in exosomes and are highly stable, hence their potential as cardiovascular biomarkers [7,40]. Circulating profiles of cardiac-enriched miRNAs vary after AMI, yet an added diagnostic value over cardiac troponins could not be demonstrated [41].

Several studies reported associations between circulating miRNA levels and LVR after AMI (Table 1). Levels of cardiac-enriched miR-208 and miR-499 were correlated with LV function after AMI [42,43], although miR-1 and miR-133a did not show significant associations [44,45]. In addition, a few miRNAs not known to be enriched in the heart (e.g. miR-34a/-21/-146/-150/-194) have been shown to be associated with LVR and HF after AMI [46–48]. Interestingly, some studies reported the incremental value of panels of miRNAs compared to single miRNAs to predict LVR after AMI [43,48,49]. Whether these miRNAs can be used as sex-specific predictors of LVR has not been properly addressed, notably due to low number of women in AMI cohorts limiting the statistical power of the analyses.

The potential of miRNAs as sex-specific biomarkers of LVR could originate from the capacity of X-linked miRNAs to escape the X chromosome inactivation as well as their modulation by sexual hormones. In mouse, the largest proportions of somatic sex-biased miRNAs have been found in heart and liver [51]. Sexual dimorphisms in miRNAs have been studied in both human and murine hearts with and without ICM [52]. In human, 3 miRNAs were sexually dimorphic in ICM and 15 in normal heart. No miRNA showed sexual dimorphism both in normal and ICM hearts suggesting that the profile of sexual dimorphic miRNAs changes under ischemic stress condition. ICM-and sex-associated miRNAs were predicted to be involved in pathways important for cardiac homeostasis such as cardiac muscle growth, angiogenesis, apoptosis, and cation transport [52].

6. X chromosome –linked miRNAs

X-chromosome encodes more miRNAs than most of the autosomes. Based on miRBase miRNA archive [www.mirbase.org 2018], 118 miRNAs are annotated on the human X-chromosome and only 4 on the Y chromosome. The function of the majority of these miRNAs remain to be elucidated. It is known that 15% and 3% of the X-linked genes escape X inactivation respectively in human and mouse [53]. In diploid cells, females have two copies of the X chromosome and males have one copy. In order to compensate sex-biased differences in gene dosage, one of the two X chromosomes in females is randomly inactivated and the genes from this inactive X chromosome are silenced [54]. This inactivation involves the long non-coding RNA (lncRNA) Xist which coats the X chromosome and leads to DNA methylation [54]. Among the X-

Table 1
Association of miRNAs with LVR after AMI.

miRNA	Cardiac enrichment	Time of measurement (post-AMI)	Number of patients	Assessment of LVR (month post-AMI)	Association with LVR	Predictive value and correlation	Ref
miR-208b	Yes	Admission	359	6	Positive	AUC 0.780	[43]
miR-34a	No	Admission	359	6	Positive	AUC 0.738	[43]
miR-208b-34a combination		Admission	359	6	Positive	AUC 0.81	[43]
miR-208b	Yes	Admission	362	4	Negative	AUC 0.69	[42]
miR-499	Yes	Admission	362	4	Negative	AUC 0.64	[42]
miR-150	No	3–4 days	90	4–6	Negative	AUC 0.74	[47]
miR-146a	No	5 days	198	12	Positive	AUC 0.818	[48]
miR-21	No	5 days	198	12	Positive	AUC 0.719	[48]
miR-16/27a/101/150 combination	No	Prior to discharge	150	6	Negative miR-150/101 Positive miR-16/27a	Δ AIC (versus clinical model) 6.83 (p = .005)	[49]
miR-194	No	Median 18 days	58	12	Positive	Correlation coefficient = 0.33	[46]
miR-155	No	5 days	20	6	Positive	Correlation coefficient = 0.49	[50]

AUC, area under the receiver operating characteristic curve; AIC, Akaike information criterion.

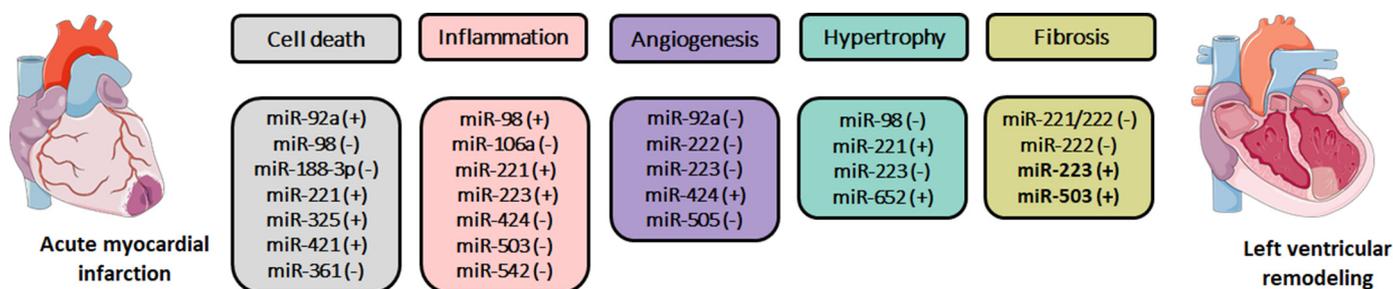


Fig. 1. Implication of X-linked miRNAs in processes triggering LVR after AMI. + signs indicate a stimulatory effect of miRNAs on biological processes; – signs indicate an inhibitory effect.

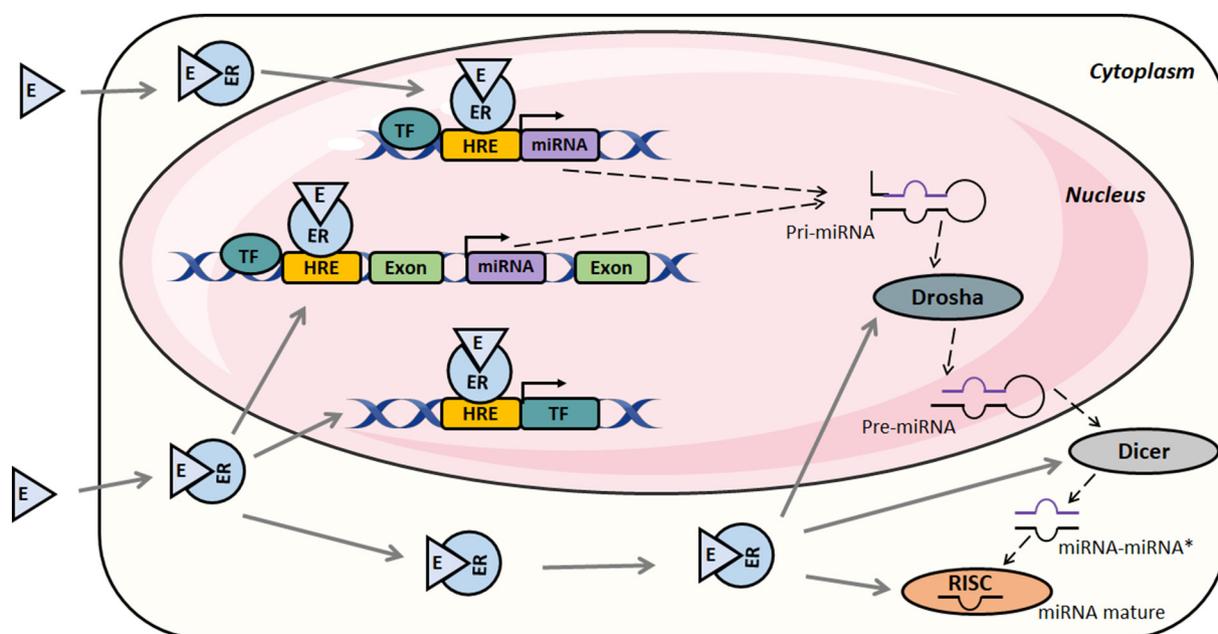


Fig. 2. Modulation of miRNAs by estrogen. E, estrogen; ER, estrogen receptor; HRE, hormone response element; RISC, RNA-induced silencing complex; TF, transcription factor.

linked miRNAs, many were found to escape X chromosome inactivation [8]. These miRNAs could therefore be responsible for sex-disparities in physiological and disease conditions. X-linked miRNAs have been shown to be involved in the different biological processes triggering LVR such as cell death, inflammation, angiogenesis, cardiomyocyte

hypertrophy, and fibrosis (Fig. 1).

6.1. Cell death

Inhibition of X-linked miR-92a in pigs undergoing myocardial

Table 2
Association of miRNAs with different biological pathways leading to LVR after AMI.

miRNA	Mechanism	Potential target	Cell type/animal model	Reference
<i>miR-92a</i>	Induction of cardiomyocytes apoptosis	–	Porcine I/R model	[55]
	Inhibition of angiogenesis	Integrin subunit $\alpha 5$	Human endothelial cells	[72]
<i>miR-361</i>	Induction of mitochondrial fission	Prohibitin (PHB1)	Murine limb ischemia and MI model	[56]
			Murine cardiomyocytes	
<i>miR-421</i>	Induction of mitochondrial fission	Pink1	Murine MI model	[57]
			Murine cardiomyocytes	
<i>miR-188</i>	Inhibition of autophagy	ATG7	Murine I/R model	[58]
			Murine cardiomyocytes subjected to anoxia/reperfusion	
<i>miR-325</i>	Induction of autophagy	Apoptosis repressor with caspase recruiting domain (ARC)	Murine I/R model	[59]
			Murine cardiomyocytes subjected to anoxia/reperfusion	
<i>miR-221</i>	Inhibition of autophagy	P27-CDK2-mTOR axis	Murine I/R model	[60]
			H9c2 cardiomyocytes cell line	
	Dendritic cells apoptosis, differentiation and maturation	P27 ^{kip1} protein	Neonatal rat cardiomyocytes	[64]
	Induction of hypertrophy	P27	Transgenic mice	
<i>miR-98</i>	Inhibition of apoptosis	Fas and caspase-3	TAC mice	[73]
			Cultured cardiomyocytes	
	Inhibition of hypertrophy	TGF- β signaling genes (JNK1, TGF β R, ETS-1)	Rat cardiac fibroblasts	[77]
<i>miR-223</i>	Inhibition of apoptosis	Fas and caspase-3	Angiotensin II-mediated pressure overload mice	[61]
			Neonatal rat cardiomyocytes treated with H ₂ O ₂	
	Inhibition of hypertrophy	Cyclin D2	Murine MI model	[76]
			Angiotensin II-mediated pressure overload mice	
	Inhibition of IL-10 production and endotoxin tolerance in macrophages	IL-10 mRNA	Neonatal rat cardiomyocytes	[65]
			HEK-293cell line	
Immune cells activation, differentiation, proliferation and survival	Granzyme B, IKK α , STAT3	RAW.264.7 cell line	[62]	
		In vitro/in vivo models		
Macrophage polarization	$\beta 1$ integrin	Human umbilical vein endothelial cells	[69]	
		Murine hindlimb ischemia model		
Cytokine production	Cardiac troponin I interacting kinase (TNNI3K)	Neonatal rat cardiomyocytes	[75]	
		TAC model		
Inhibition of angiogenesis	RASA-1	Rat cardiac fibroblasts	[78]	
		Rat MI model		
<i>miR-155</i>	Induction of dendritic cells apoptosis	P27 ^{kip1} protein	Human and murine dendritic cells	[64]
<i>miR-424</i>	Differentiation of monocytes to dendritic cells	Culin-2	Human endothelial cells subjected to hypoxia	[68]
			Murine ischemia models	
<i>miR-505</i>	Induction of endothelial cells migration and tube formation	Fibroblast growth factor-18 (FGF-18)	Endothelial cells	[70]
			Endothelial cells	
<i>miR-222</i>	Inhibition of endothelial cells proliferation and migration	Signal transducer and activator of transcription 5A (STAT5A)	Endothelial cells	[71]
			Endothelial cells	
<i>miR-652</i>	Inhibition of fibrosis	TGF- β signaling genes (JNK1, TGF β R, ETS-1)	Cardiac rat fibroblasts	[77]
			Angiotensin II-mediated pressure overload mice	
<i>miR-503</i>	Induction of hypertrophy	Jagged-1	TAC mice	[74]
			Neonatal murine cardiac fibroblasts	
<i>miR-126-3p</i>	Induction of cellular proliferation and collagen production in fibroblasts	Apelin-13	TAC mice	[79]
			Neonatal murine cardiac fibroblasts	
<i>miR-106a/-106b/-24/-27a/-27b</i>	Anti-atherogenic	Spred-1/VCAM-1	ApoE ^{-/-} mice	[83]
			Human umbilical vein endothelial cells	
<i>miR-23a</i>	Induction of myocardial apoptosis	Caspase-3	Rat I/R model	[84]
			Rat fibroblasts	
<i>miR-106a/-106b/-24/-27a/-27b</i>	Induction of fibrosis	MAPK/ERK1/2 pathway	H9c2 cell line	[85]
			TAC model	
<i>miR-23a</i>	Induction of concentric remodeling	PGC-1 α	Cardiomyocytes cell line	[87]
			Induction of mitochondrial structural and respiratory damage	
			Cultured cardiomyocytes	

I/R, ischemia/reperfusion; MI, myocardial infarction; TAC, transverse aortic constriction.

ischemia/reperfusion reduced infarct size and apoptosis, and improved cardiac function [55]. MiR-421 and miR-361 had opposite effects on cardiomyocyte apoptosis upon myocardial ischemia by modulating mitochondrial fission [56,57]. Overexpression of miR-188-3p in an ischemia-reperfusion model attenuated cardiomyocytes death through autophagy [58]. This effect involved a lncRNA named autophagy promoting factor which was found to interact directly with miR-188-3p and modulate its activity [58]. MiR-325 was upregulated upon ischemia/reperfusion and its overexpression in cardiomyocytes increased autophagic death and infarct size [59]. Moreover, cardiac-specific overexpression of miR-221 led to HF in mice which exhibited hypertrophic heart with excessive interstitial fibrosis, apoptosis and altered cardiac function [60]. This effect implicated an inhibition of cardiomyocyte autophagy through activation of mTOR pathway [60]. Moreover, miR-98 was found to exert an anti-apoptotic effect after AMI through modulation of Fas/Caspase 3 apoptotic pathway [61]. Taken together, these data show that X-linked miRNAs play a key role in modulating cell death after AMI and could account for the difference in infarct size between women and men.

6.2. Inflammation

The X-linked miR-223 has been extensively studied and plays a prominent role in inflammation, as well as in diabetes, insulin-resistance and stem cell differentiation [62]. MiR-221 regulates dendritic cells and T lymphocytes proliferation through the inhibition of cyclin inhibitor proteins [63,64]. MiR-98 is upregulated in activated macrophages and inhibits IL-10 production [65]. MiR-106a, miR-424, miR-503, and miR-542 are involved in monocytes differentiation [66]. Data from HUNT-study addressing the ability of miRNAs to predict the risk of 10 years fatal AMI in a healthy population identified a combination of 5 miRNAs among which the 2 X-linked miR-106a and miR-424 [67]. Interestingly, in this population, miR-424 was associated with the endpoint specifically in men [67].

6.3. Angiogenesis

Hypoxia induced the expression of the X-linked miR-424 in endothelial cells, leading to an increase in vascular endothelial growth factor expression, cell migration and vessel formation [68]. On the opposite, overexpression of miR-223 abrogated endothelial cells (ECs) proliferation, migration and sprouting through preventing growth factor signaling [69]. MiR-505, which is upregulated in the plasma of hypertensive patients, inhibits ECs migration and vessel formation by targeting the proangiogenic fibroblast growth factor 18 [70]. MiR-222, which was downregulated in ECs upon treatment with the inflammatory proangiogenic mediators IL-3 and basic fibroblast growth factor, reduced ECs proliferation and migration through down-regulation of the transcription factor signal transducer and activator of transcription 5A [71]. MiR-92a controls angiogenesis in mice and pigs subjected to ischemia [55,72].

6.4. Hypertrophy

X-linked miR-221 promoted cardiac hypertrophy by down-regulating the hypertrophic suppressor p27 [73]. Inhibition of miR-652 in mice subjected to pressure overload reduced hypertrophy, apoptosis, fibrosis, and preserved angiogenesis, effects mediated by modulation of the Notch1 ligand Jagged1 [74]. MiR-223 was downregulated in both hypertrophic neonatal rat cardiomyocytes and hypertrophic murine heart [75]. It was observed that miR-223 directly targets the cardiac troponin I-interacting kinase to modulate cardiomyocytes hypertrophy [75]. In another study, knocking down miR-98 increased angiotensin II-induced cardiac hypertrophy, an effect partially attributed to cyclin D2 downregulation of [76].

6.5. Fibrosis

Expression of the X-linked miRNA cluster 221/222 negatively correlated with the extent of fibrosis in myocardial biopsies from patients [77]. Inhibition of both miRNAs in mice subjected to angiotensin II-mediated pressure overload increased fibrosis and induced LV dysfunction, an effect involving the transforming growth factor β signaling [77]. MiR-223 regulates fibrosis after AMI through stimulation of cardiac fibroblasts proliferation, migration and differentiation [78]. In a transverse aortic constriction model as well as in angiotensin II-treated cardiac fibroblasts, miR-503 promoted fibrosis through inhibition of apelin13, a crucial protein for cardiac fibroblasts activation [79].

7. Estrogen-modulated miRNAs

Estrogen binds to estrogen receptors (ER) ER α and ER β and the recently discovered G-protein coupled ER (GPER) [19]. Cells of the cardiovascular system such as ECs, vascular smooth muscle cells and cardiomyocytes express ERs which suggests the involvement of estrogen in cardiovascular pathophysiology. Estrogen plays a pivotal role in cardioprotection against ischemic stress: decreased infarct size, decreased cardiomyocyte death, improved neovascularization, regulation of inflammation [19]. Estrogen affects miRNAs biogenesis through downregulation of *AGO2* and upregulation of *DICER1* and *DROSHA* [80,81]. At the transcriptional level, estradiol seems to modulate the expression of miRNAs through either direct binding of ER α on the regulatory regions of the miRNA, or by modulating the expression of the mRNA-encoding genes that harbor intronically miRNAs, or lastly by modulating the expression of the transcription factors that control the expression of miRNAs [9] (Fig. 2). Estrogen-bound ER α binds to estrogen response elements on DNA and induce the expression of miR-21, miR-23a, and miR-221/222 [9,82].

Few data show the involvement of miRNAs in the cardioprotective effects of estrogen. MiR-126-3p is modulated through the cycle phases in menstruating females suggesting a regulation by estrogens [83]. Furthermore, the anti-thrombotic effect of estrogen is abrogated after inhibition of miR-126-3p in ApoE $^{-/-}$ mice [83]. This effect was proposed to involve a regulation of ECs proliferation and migration and inhibition of monocytes infiltration by miR-126-3p [83]. In another study, miR-126-3p enhanced apoptosis in female rats undergoing myocardial ischemia-reperfusion injury, and its inhibition reduced infarct size [84]. Following pressure overload in mice, miR-106a/-106b/-24/-27a/-27b were found to be upregulated only in male hearts and contribute to fibrosis through MAPK/ERK1/2 signaling pathway regulation [85]. This sex-biased modulation of fibrosis was mediated by estrogen through ER β activation [85]. Moreover, miR-23a is involved in ageing-associated concentric remodeling which is more common among women [86,87]. Estrogen deficiency during menopause leads to mitochondrial respiration inefficiency, which is mediated by the downregulation of peroxisome proliferator-activated receptor- γ co-activator 1- α by miR-23a [87].

8. Conclusion and future directions

Personalizing healthcare after AMI is required to decrease the risk of developing LVR and HF and to improve life quality. Novel biomarkers might be useful in this regard. However, LVR and outcome after AMI show important differences between women and men, implying the discovery of sex-specific biomarkers. Since they escape X-chromosome inactivation and are modulated by sex hormones, miRNAs appear to be a reservoir of such sex-specific biomarkers.

Although the miRNAs summarized in this article and in Table 2 have been shown to be associated with different biological pathways leading to adverse remodeling after AMI (e.g. cell death, inflammation, angiogenesis, hypertrophy, and fibrosis), whether targeting one or several of them is a suitable strategy to prevent the development of LVR

still needs to be demonstrated. In addition, since miRNAs are known to act both intracellularly and through paracrine mechanisms, it will be insightful to determine whether they act at a local or systemic level on pathways leading to LVR. This could inform about the desired mode of administration of miRNA-targeting drugs. The identification of sex-specific miRNAs associated with outcome after AMI motivates the search for novel sex-specific therapeutic strategies based on miRNAs.

Evaluation of the additive prognostic impact on novel biomarkers such as miRNAs on classical markers is paramount and requires specific statistical methods that have been deployed in some but not all studies. Care should be taken to avoid model overfitting due to the multiplication of explanatory variables in prediction models. The issue of sex-specificity raised in this review article has to be carefully taken into consideration and requires proper sample size calculations to reach a sufficient study power.

Long non-coding RNAs represent the majority of ncRNAs. They are involved in cardiac development and disease [88,89], have been associated with post AMI outcome [90,91], can escape X-inactivation [92], and are regulated by estrogen in cancer cells [93]. Therefore, studying the sex-biased expression of lncRNAs after AMI and their functional role in LVR may lead to the discovery of novel sex-specific biomarkers of LVR.

Acknowledgements

The authors are members of the Cardioline™ network (www.cardiolinc.org).

Sources of funding

This work was supported by the Ministry of Higher Education and Research and the Society for Research on Cardiovascular Diseases of Luxembourg.

Declarations of interest

None.

References

- http://www.who.int/cardiovascular_diseases/world-heart-day-2017/en/Cd-WHO.Cardiovascular_diseases.
- Cardiovascular disease in Europe 2016: an epidemiological update, *Eur. Heart J.* 37 (2016) 3182–3183.
- S. Dey, M.D. Flather, G. Devlin, et al., Sex-related differences in the presentation, treatment and outcomes among patients with acute coronary syndromes: the Global Registry of Acute Coronary Events, *Heart* 95 (2009) 20–26.
- J.G. Canto, W.J. Rogers, R.J. Goldberg, et al., Association of age and sex with myocardial infarction symptom presentation and in-hospital mortality, *Jama* 307 (2012) 813–822.
- C.S. Lam, M. McEntegart, B. Claggett, et al., Sex differences in clinical characteristics and outcomes after myocardial infarction: insights from the Valsartan in Acute Myocardial Infarction Trial (VALIANT), *Eur. J. Heart Fail.* 17 (2015) 301–312.
- D.L. Crabbe, K. Dipla, S. Ambati, et al., Gender differences in post-infarction hypertrophy in end-stage failing hearts, *J. Am. Coll. Cardiol.* 41 (2003) 300–306.
- E. Goretti, D.R. Wagner, Y. Devaux, miRNAs as biomarkers of myocardial infarction: a step forward towards personalized medicine? *Trends Mol. Med.* 20 (2014) 716–725.
- R. Song, S. Ro, J.D. Michaels, C. Park, J.R. McCarrey, W. Yan, Many X-linked microRNAs escape meiotic sex chromosome inactivation, *Nat. Genet.* 41 (2009) 488–493.
- P. Bhat-Nakshatri, G. Wang, N.R. Collins, et al., Estradiol-regulated microRNAs control estradiol response in breast cancer cells, *Nucleic Acids Res.* 37 (2009) 4850–4861.
- G. Albrektsen, I. Heuch, M.L. Lochen, et al., Lifelong gender gap in risk of incident myocardial infarction: the Tromso Study, *JAMA Intern. Med.* 176 (2016) 1673–1679.
- A.J. Lansky, V.G. Ng, A. Maehara, et al., Gender and the extent of coronary atherosclerosis, plaque composition, and clinical outcomes in acute coronary syndromes, *JACC Cardiovasc. Imaging* 5 (2012) S62–S72.
- S.J. White, A.C. Newby, T.W. Johnson, Endothelial erosion of plaques as a substrate for coronary thrombosis, *Thromb. Haemost.* 115 (2016) 509–519.
- K.H. Humphries, M. Izadnegahdar, T. Sedlak, et al., Sex differences in cardiovascular disease - impact on care and outcomes, *Front. Neuroendocrinol.* 46 (2017) 46–70.
- N.A. Khan, S.S. Daskalopoulou, I. Karp, et al., Sex differences in acute coronary syndrome symptom presentation in young patients, *JAMA Intern. Med.* 173 (2013) 1863–1871.
- J.S. Hochman, J.E. Tamis, T.D. Thompson, et al., Sex, clinical presentation, and outcome in patients with acute coronary syndromes. Global use of strategies to open occluded coronary arteries in acute coronary syndromes IIb investigators, *N. Engl. J. Med.* 341 (1999) 226–232.
- P. Kaul, P.W. Armstrong, S. Sookram, B.K. Leung, N. Brass, R.C. Welsh, Temporal trends in patient and treatment delay among men and women presenting with ST-elevation myocardial infarction, *Am. Heart J.* 161 (2011) 91–97.
- M.J. Valero-Masa, J. Velasquez-Rodriguez, F. Diez-Delgado, et al., Sex differences in acute myocardial infarction: is it only the age? *Int. J. Cardiol.* 231 (2017) 36–41.
- R. Ubrich, P. Barthel, B. Haller, et al., Sex differences in long-term mortality among acute myocardial infarction patients: results from the ISAR-RISK and ART studies, *PLoS One* 12 (2017) e0186783.
- V. Regitz-Zagrosek, G. Kararigas, Mechanistic pathways of sex differences in cardiovascular disease, *Physiol. Rev.* 97 (2017) 1–37.
- M. Jain, R. Liao, B.K. Podesser, S. Ngoy, C.S. Apstein, F.R. Eberli, Influence of gender on the response to hemodynamic overload after myocardial infarction, *Am. J. Physiol. Heart Circ. Physiol.* 283 (2002) H2544–H2550.
- M.A. Cavasin, Z.Y. Tao, A.L. Yu, X.P. Yang, Testosterone enhances early cardiac remodeling after myocardial infarction, causing rupture and degrading cardiac function, *Am. J. Physiol. Heart Circ. Physiol.* 290 (2006) H2043–H2050.
- M. Vausort, A. Salgado-Somoza, L. Zhang, et al., Myocardial infarction-associated circular RNA predicting left ventricular dysfunction, *J. Am. Coll. Cardiol.* 68 (2016) 1247–1248.
- M. Piro, R. Della Bona, A. Abbate, L.M. Biasucci, F. Crea, Sex-related differences in myocardial remodeling, *J. Am. Coll. Cardiol.* 55 (2010) 1057–1065.
- G.G. Biondi-Zoccai, A. Abate, R. Bussani, et al., Reduced post-infarction myocardial apoptosis in women: a clue to their different clinical course? *Heart* 91 (2005) 99–101.
- E. Canali, P. Masci, J. Bogaert, et al., Impact of gender differences on myocardial salvage and post-ischaemic left ventricular remodeling after primary coronary angioplasty: new insights from cardiovascular magnetic resonance, *Eur. Heart J. Cardiovasc. Imaging* 13 (2012) 948–953.
- R.S. Vasan, E.J. Benjamin, M.G. Larson, et al., Plasma natriuretic peptides for community screening for left ventricular hypertrophy and systolic dysfunction: the Framingham heart study, *Jama* 288 (2002) 1252–1259.
- M.M. Redfield, R.J. Rodeheffer, S.J. Jacobsen, D.W. Mahoney, K.R. Bailey, J.C. Burnett Jr., Plasma brain natriuretic peptide concentration: impact of age and gender, *J. Am. Coll. Cardiol.* 40 (2002) 976–982.
- M. Hamada, Y. Shigematsu, M. Takezaki, S. Ikeda, A. Ogomoto, Plasma levels of atrial and brain natriuretic peptides in apparently healthy subjects: effects of sex, age, and hemoglobin concentration, *Int. J. Cardiol.* 228 (2017) 599–604.
- C.S. Lam, S. Cheng, K. Choong, et al., Influence of sex and hormone status on circulating natriuretic peptides, *J. Am. Coll. Cardiol.* 58 (2011) 618–626.
- S.D. Wiviott, C.P. Cannon, D.A. Morrow, et al., Differential expression of cardiac biomarkers by gender in patients with unstable angina/non-ST-elevation myocardial infarction: a TACTICS-TIMI 18 (Treat Angina with Aggrastat and determine Cost of Therapy with an Invasive or Conservative Strategy-Thrombolysis In Myocardial Infarction 18) substudy, *Circulation* 109 (2004) 580–586.
- High sensitivity cardiac troponin and the under-diagnosis of myocardial infarction in women: prospective cohort study, *BMJ* 350 (2015) h626.
- J. Lew, M. Sanghavi, C.R. Ayers, et al., Sex-based differences in cardiometabolic biomarkers, *Circulation* 135 (2017) 544–555.
- M. Fertin, B. Hennache, M. Hamon, et al., Usefulness of serial assessment of B-type natriuretic peptide, troponin I, and C-reactive protein to predict left ventricular remodeling after acute myocardial infarction (from the REVE-2 study), *Am. J. Cardiol.* 106 (2010) 1410–1416.
- A. Mayr, J. Mair, G. Klug, et al., Cardiac troponin T and creatine kinase predict mid-term infarct size and left ventricular function after acute myocardial infarction: a cardiac MR study, *J. Magn. Reson. Imaging* 33 (2011) 847–854.
- J.E. Ho, C. Liu, A. Lyass, et al., Galectin-3, a marker of cardiac fibrosis, predicts incident heart failure in the community, *J. Am. Coll. Cardiol.* 60 (2012) 1249–1256.
- L.B. Daniels, P. Clopton, G.A. Laughlin, A.S. Maisel, E. Barrett-Connor, Galectin-3 is independently associated with cardiovascular mortality in community-dwelling older adults without known cardiovascular disease: the Rancho Bernardo Study, *Am. Heart J.* 167 (2014) 674–682 (e1).
- E.E. Coglianese, M.G. Larson, R.S. Vasan, et al., Distribution and clinical correlates of the interleukin receptor family member soluble ST2 in the Framingham Heart Study, *Clin. Chem.* 58 (2012) 1673–1681.
- T. Lalem, L. Zhang, M. Scholz, et al., Cyclin dependent kinase inhibitor 1 C is a female-specific marker of left ventricular function after acute myocardial infarction, *Int. J. Cardiol.* 274 (2019) 319–325.
- J. Beermann, M.T. Piccoli, J. Viereck, T. Thum, Non-coding RNAs in development and disease: background, mechanisms, and therapeutic approaches, *Physiol. Rev.* 96 (2016) 1297–1325.
- J. Viereck, T. Thum, Circulating noncoding RNAs as biomarkers of cardiovascular disease and injury, *Circ. Res.* 120 (2017) 381–399.
- Y. Devaux, M. Mueller, P. Haaf, et al., Diagnostic and prognostic value of circulating microRNAs in patients with acute chest pain, *J. Intern. Med.* 277 (2015) 260–271.
- Y. Devaux, M. Vausort, E. Goretti, et al., Use of circulating microRNAs to diagnose acute myocardial infarction, *Clin. Chem.* 58 (2012) 559–567.
- P. Lv, M. Zhou, J. He, et al., Circulating miR-208b and miR-34a are associated with

- left ventricular remodeling after acute myocardial infarction, *Int. J. Mol. Sci.* 15 (2014) 5774–5788.
- [44] U. Grabmaier, S. Claus, L. Gross, et al., Diagnostic and prognostic value of miR-1 and miR-29b on adverse ventricular remodeling after acute myocardial infarction - the SITAGRAMI-miR analysis, *Int. J. Cardiol.* 244 (2017) 30–36.
- [45] C. Bauters, R. Kumarswamy, A. Holzmann, et al., Circulating miR-133a and miR-423-5p fail as biomarkers for left ventricular remodeling after myocardial infarction, *Int. J. Cardiol.* 168 (2013) 1837–1840.
- [46] S. Matsumoto, Y. Sakata, S. Suna, et al., Circulating p53-responsive microRNAs are predictive indicators of heart failure after acute myocardial infarction, *Circ. Res.* 113 (2013) 322–326.
- [47] Y. Devaux, M. Vausort, G.P. McCann, et al., MicroRNA-150: a novel marker of left ventricular remodeling after acute myocardial infarction, *Circ. Cardiovasc. Genet.* 6 (2013) 290–298.
- [48] X. Liu, Y. Dong, S. Chen, et al., Circulating MicroRNA-146a and MicroRNA-21 predict left ventricular remodeling after ST-elevation myocardial infarction, *Cardiology* 132 (2015) 233–241.
- [49] Y. Devaux, M. Vausort, G.P. McCann, et al., A panel of 4 microRNAs facilitates the prediction of left ventricular contractility after acute myocardial infarction, *PLoS One* 8 (2013) e70644.
- [50] S.C. Latet, P.L. Van Herck, M.J. Claeys, et al., Failed downregulation of circulating MicroRNA-155 in the early phase after ST elevation myocardial infarction is associated with adverse left ventricular remodeling, *Cardiology* 138 (2017) 91–96.
- [51] M. Warnefors, K. Mossinger, J. Halbert, et al., Sex-biased microRNA expression in mammals and birds reveals underlying regulatory mechanisms and a role in dosage compensation, *Genome Res.* 27 (2017) 1961–1973.
- [52] M. Tsuji, T. Kawasaki, T. Matsuda, T. Arai, S. Gojo, J.K. Takeuchi, Sexual dimorphisms of mRNA and miRNA in human/murine heart disease, *PLoS One* 12 (2017) e0177988.
- [53] J.B. Berletch, F. Yang, J. Xu, L. Carrel, C.M. Disteche, Genes that escape from X inactivation, *Hum. Genet.* 130 (2011) 237–245.
- [54] J.T. Lee, M.S. Bartolomei, X-inactivation, imprinting, and long noncoding RNAs in health and disease, *Cell* 152 (2013) 1308–1323.
- [55] R. Hinkel, D. Penzkofer, S. Zuhlke, et al., Inhibition of microRNA-92a protects against ischemia/reperfusion injury in a large-animal model, *Circulation* 128 (2013) 1066–1075.
- [56] K. Wang, C.Y. Liu, X.J. Zhang, et al., miR-361-regulated prohibitin inhibits mitochondrial fission and apoptosis and protects heart from ischemia injury, *Cell Death Differ.* 22 (2015) 1058–1068.
- [57] K. Wang, L.Y. Zhou, J.X. Wang, et al., E2F1-dependent miR-421 regulates mitochondrial fragmentation and myocardial infarction by targeting Pink1, *Nat. Commun.* 6 (2015) 7619.
- [58] K. Wang, C.Y. Liu, L.Y. Zhou, et al., APF lncRNA regulates autophagy and myocardial infarction by targeting miR-188-3p, *Nat. Commun.* 6 (2015) 6779.
- [59] L. Bo, D. Su-Ling, L. Fang, et al., Autophagic program is regulated by miR-325, *Cell Death Differ.* 21 (2014) 967–977.
- [60] M. Su, J. Wang, C. Wang, et al., MicroRNA-221 inhibits autophagy and promotes heart failure by modulating the p27/CDK2/mTOR axis, *Cell Death Differ.* 22 (2015) 986–999.
- [61] C. Sun, H. Liu, J. Guo, et al., MicroRNA-98 negatively regulates myocardial infarction-induced apoptosis by down-regulating Fas and caspase-3, *Sci. Rep.* 7 (2017) 7460.
- [62] M. Haneklaus, M. Gerlic, L.A. O'Neill, S.L. Masters, miR-223: infection, inflammation and cancer, *J. Intern. Med.* 274 (2013) 215–226.
- [63] Y.A. Grigoryev, S.M. Kurian, T. Hart, et al., MicroRNA regulation of molecular networks mapped by global microRNA, mRNA, and protein expression in activated T lymphocytes, *J. Immunol.* 187 (2011) 2233–2243.
- [64] C. Lu, X. Huang, X. Zhang, et al., miR-221 and miR-155 regulate human dendritic cell development, apoptosis, and IL-12 production through targeting of p27kip1, KPC1, and SOCS-1, *Blood* 117 (2011) 4293–4303.
- [65] Y. Liu, Q. Chen, Y. Song, et al., MicroRNA-98 negatively regulates IL-10 production and endotoxin tolerance in macrophages after LPS stimulation, *FEBS Lett.* 585 (2011) 1963–1968.
- [66] I. Pinheiro, L. Dejager, C. Libert, X-chromosome-located microRNAs in immunity: might they explain male/female differences? The X chromosome-genomic context may affect X-located miRNAs and downstream signaling, thereby contributing to the enhanced immune response of females, *Bioessays* 33 (2011) 791–802.
- [67] A. Bye, H. Rosjo, J. Nauman, et al., Circulating microRNAs predict future fatal myocardial infarction in healthy individuals - the HUNT study, *J. Mol. Cell. Cardiol.* 97 (2016) 162–168.
- [68] G. Ghosh, I.V. Subramanian, N. Adhikari, et al., Hypoxia-induced microRNA-424 expression in human endothelial cells regulates HIF-alpha isoforms and promotes angiogenesis, *J. Clin. Invest.* 120 (2010) 4141–4154.
- [69] L. Shi, B. Fisslthaler, N. Zippel, et al., MicroRNA-223 antagonizes angiogenesis by targeting beta1 integrin and preventing growth factor signaling in endothelial cells, *Circ. Res.* 113 (2013) 1320–1330.
- [70] Q. Yang, C. Jia, P. Wang, et al., MicroRNA-505 identified from patients with essential hypertension impairs endothelial cell migration and tube formation, *Int. J. Cardiol.* 177 (2014) 925–934.
- [71] P. Dentelli, A. Rosso, F. Orso, C. Olgasi, D. Taverna, M.F. Brizzi, microRNA-222 controls neovascularization by regulating signal transducer and activator of transcription 5A expression, *Arterioscler. Thromb. Vasc. Biol.* 30 (2010) 1562–1568.
- [72] A. Bonauer, G. Carmona, M. Iwasaki, et al., MicroRNA-92a controls angiogenesis and functional recovery of ischemic tissues in mice, *Science* 324 (2009) 1710–1713.
- [73] C. Wang, S. Wang, P. Zhao, et al., MiR-221 promotes cardiac hypertrophy in vitro through the modulation of p27 expression, *J. Cell. Biochem.* 113 (2012) 2040–2046.
- [74] B.C. Bernardo, S.S. Nguyen, C.E. Winbanks, et al., Therapeutic silencing of miR-652 restores heart function and attenuates adverse remodeling in a setting of established pathological hypertrophy, *FASEB J.* 28 (2014) 5097–5110.
- [75] Y.S. Wang, J. Zhou, K. Hong, X.S. Cheng, Y.G. Li, MicroRNA-223 displays a protective role against cardiomyocyte hypertrophy by targeting cardiac troponin I-interacting kinase, *Cell. Physiol. Biochem.* 35 (2015) 1546–1556.
- [76] Y. Yang, T. Ago, P. Zhai, M. Abdellatif, J. Sadoshima, Thioredoxin 1 negatively regulates angiotensin II-induced cardiac hypertrophy through upregulation of miR-98/let-7, *Circ. Res.* 108 (2011) 305–313.
- [77] R. Verjans, T. Peters, F.J. Beaumont, et al., MicroRNA-221/222 family counteracts myocardial fibrosis in pressure overload-induced heart failure, *Hypertension* 71 (2018) 280–288.
- [78] X. Liu, Y. Xu, Y. Deng, H. Li, MicroRNA-223 regulates cardiac fibrosis after myocardial infarction by targeting RASA1, *Cell. Physiol. Biochem.* 46 (2018) 1439–1454.
- [79] Y. Zhou, L. Deng, D. Zhao, et al., MicroRNA-503 promotes angiotensin II-induced cardiac fibrosis by targeting Apelin-13, *J. Cell. Mol. Med.* 20 (2016) 495–505.
- [80] C. Cheng, X. Fu, P. Alves, M. Gerstein, mRNA expression profiles show differential regulatory effects of microRNAs between estrogen receptor-positive and estrogen receptor-negative breast cancer, *Genome Biol.* 10 (2009) R90.
- [81] D. Perez-Cremades, A. Mompeon, X. Vidal-Gomez, C. Hermenegildo, S. Novella, miRNA as a new regulatory mechanism of estrogen vascular action, *Int. J. Mol. Sci.* 19 (2018).
- [82] G. Di Leva, P. Gasparini, C. Piovani, et al., MicroRNA cluster 221-222 and estrogen receptor alpha interactions in breast cancer, *J. Natl. Cancer Inst.* 102 (2010) 706–721.
- [83] P. Li, J. Wei, X. Li, et al., 17beta-estradiol enhances vascular endothelial Ets-1/miR-126-3p expression: the possible mechanism for attenuation of atherosclerosis, *J. Clin. Endocrinol. Metab.* 102 (2017) 594–603.
- [84] B. Li, Y. Tao, Q. Huang, Effect and mechanism of miR-126 in myocardial ischemia reperfusion, *Genet. Mol. Res.* 14 (2015) 18990–18998.
- [85] A.M. Queiros, C. Eschen, D. Flegner, et al., Sex- and estrogen-dependent regulation of a miRNA network in the healthy and hypertrophied heart, *Int. J. Cardiol.* 169 (2013) 331–338.
- [86] S. Cheng, V. Xanthakis, L.M. Sullivan, et al., Correlates of echocardiographic indices of cardiac remodeling over the adult life course: longitudinal observations from the Framingham Heart Study, *Circulation* 122 (2010) 570–578.
- [87] L.Y. Sun, N. Wang, T. Ban, et al., MicroRNA-23a mediates mitochondrial compromise in estrogen deficiency-induced concentric remodeling via targeting PGC-1alpha, *J. Mol. Cell. Cardiol.* 75 (2014) 1–11.
- [88] Y. Devaux, J. Zangrando, B. Schroen, et al., Long noncoding RNAs in cardiac development and ageing, *Nat. Rev. Cardiol.* 12 (2015) 415–425.
- [89] J. Viereck, T. Thum, Circulating noncoding RNAs as biomarkers of cardiovascular disease and injury, *Circ. Res.* 120 (2017) 381–399.
- [90] R. Kumarswamy, C. Bauters, I. Volkman, et al., Circulating long noncoding RNA, LIPCAR, predicts survival in patients with heart failure, *Circ. Res.* 114 (2014) 1569–1575.
- [91] M. Vausort, D.R. Wagner, Y. Devaux, Long noncoding RNAs in patients with acute myocardial infarction, *Circ. Res.* 115 (2014) 668–677.
- [92] B. Reinius, C. Shi, L. Hengshuo, et al., Female-biased expression of long non-coding RNAs in domains that escape X-inactivation in mouse, *BMC Genomics* 11 (2010) 614.
- [93] A. Aiello, L. Bacci, A. Re, et al., MALAT1 and HOTAIR long non-coding RNAs play opposite role in estrogen-mediated transcriptional regulation in prostate cancer cells, *Sci. Rep.* 6 (2016) 38414.