



# High-Mobility Group Box 1 Neutralization Prevents Chronic Cerebral Hypoperfusion-Induced Optic Tract Injuries in the White Matter Associated with Down-regulation of Inflammatory Responses

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## Abstract

Chronic cerebral hypoperfusion (CCH)-induced white matter lesions (WMLs) are region-specific with the optic tract (OT) displaying the most severe damages and leading to visual-based behavioral impairment. Previously we have demonstrated that anti-high-mobility group box 1 (HMGB1) neutralizing antibody (Ab) prevents CCH-induced hippocampal damages via inhibition of neuroinflammation. Here we tested the protective role of the Ab on CCH-induced OT injuries. Rats were treated with permanent occlusion of common carotid arteries (2-VO) or a sham surgery, and then administered with PBS, anti-HMGB1 Ab, or paired control Ab. Pupillary light reflex examination, visual water maze, and tapered beam-walking were performed 28 days post-surgery to investigate the behavioral deficits. Meanwhile, WMLs were measured by Klüver-Barrera (KB) and H&E staining, and glial activation was further assessed to evaluate inflammatory responses in OT. Results revealed that anti-HMGB1 Ab ameliorated the morphological damages (grade scores, vacuoles, and thickness) in OT area and preserved visual abilities. Additionally, the increased levels of inflammatory responses and expressions of TLR4 and NF- $\kappa$ B p65 and phosphorylated NF- $\kappa$ B p65 (p-p65) in OT area were partly down-regulated after anti-HMGB1 treatment. Taken together, these findings suggested that HMGB1 neutralization could ease OT injuries and visual-guided behavioral deficits via suppressing inflammatory responses.

**Keywords** HMGB1 neutralization · Chronic cerebral hypoperfusion · Optic tract · Glial activation · NF- $\kappa$ B

## Introduction

Chronic cerebral hypoperfusion (CCH) could cause long-lasting reduction in cerebral blood flow (CBF) and result in white matter lesions (WMLs), visual dysfunction, glial activation, and cognitive impairments (Davidson et al. 2000; Farkas et al. 2007; Back et al. 2017). Permanent occlusion of common carotid arteries (2-VO) in rats has now been widely accepted as a suitable approach to unravel the pathological

changes and drug candidates for the treatment of WMLs in CCH and associated neurodegenerative diseases, such as vascular dementia (VaD), Alzheimer's disease (AD), and aging (Farkas and Luiten 2001; Akinyemi et al. 2013). In particular, substantial evidence has shown that CCH-induced WMLs are preferentially presented in regions, and the area of optic tract (OT), as a part of the visual system, displays much severer rarefaction (reduction of thickness), vacuolation, and sustained glial activation than other white matter regions (like corpus callosum, internal capsule, and caudoputamen) (Takizawa et al. 2003; Lee et al. 2006; Tian et al. 2014; Lee et al. 2015; Edrissi et al. 2016). However, previous studies have not pay sufficient attention to the pathological changes and the related functional outcomes (like visual-dependent spatial learning and motor deficits) in this area during CCH.

High-mobility group box 1 (HMGB1) can bind to receptor for advanced glycation end-products (RAGE) and toll-like receptor 4 (TLR4) and activate inflammatory responses once released into extracellular space under pathological

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conditions (Hamada et al. 2008; Pisetsky 2014). However, the role of HMGB1 in white matter injury remains controversial. On one hand, it has been reported that the activation of HMGB1 signaling results in persistent damages in the white matter in animal models of systemic inflammation and traumatic brain injury (Cardoso et al. 2015; Braun et al. 2017); on the other hand, some studies have showed that HMGB1 from astrocytes promotes the function of progenitor cells and plays as a autocrine factor in WMLs in acute ischemic models (Hayakawa et al. 2013; Choi et al. 2017). As a matter of fact, most of these studies focused on HMGB1 signaling in the acute WMLs models. However, CCH-induced chronic WMLs are usually accompanied with prolonged hyper-activation of glial cells (Choi et al. 2016), and the anti-inflammation therapies in the chronic phase of CCH via down-regulation of TLR4 and RAGE proved to be beneficial to the CCH-induced WMLs (Kim et al. 2015; Lee et al. 2015; Kim et al. 2016; Saggu et al. 2016). As the HMGB1 and its down-stream signaling TLR4/NF- $\kappa$ B were persistently activated in the chronic phase of CCH (Lee et al. 2015; Ashok et al. 2016; Hei et al. 2018), it is deducible that HMGB1 signaling plays a role in the pathogenesis of chronic WMLs and OT damages induced by CCH.

Previously, we have demonstrated that anti-HMGB1 neutralizing antibody (Ab) treatment suppresses glial activation, pro-inflammatory cytokine production, and oxidative stress in a rat model of CCH, which finally leads to long-time beneficial effects on the neuronal survival and cognitive ability in the chronic phase (Hei et al. 2018). Here we aimed to present the evidence that anti-HMGB1 neutralizing Ab is capable of easing OT injuries in the white matter and improving the visual-guided behavioral abilities via suppressing inflammatory responses in the chronic phase of CCH in rats.

## Materials and Methods

### Animals

A total of 91 male Wistar rats weighing 220–230 g were used. National Institutes of Health Ethic Committee gave permission to the animal experiments, which were conducted in accordance with the National Experimental Animals Guidelines. Rats were obtained from Vital River and kept in Animal Center of the Fourth Military Medical University (Xi'an, China). Wistar rats present much severe damages in the visual system in CCH model in comparison to SD rats and are frequently used to investigate 2VO-induced WMLs (Kim et al. 2008; Lee et al. 2015; Ueno et al. 2015; Back et al. 2017). Apart from the randomly selected 18 rats in the sham group [treated with a sham surgery and phosphate-buffered saline (PBS, pH 7.4)] and six rats in the sham + anti-HMGB1 group (treated with a sham surgery

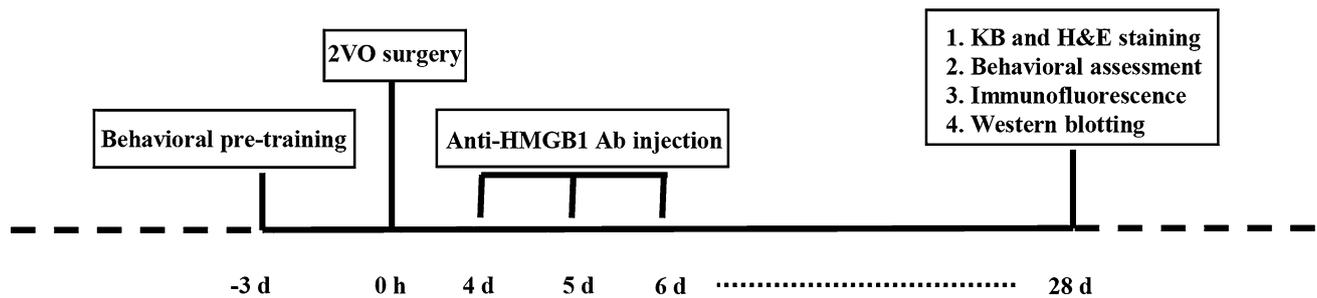
and anti-HMGB1 neutralizing antibody (Ab) and used for histological assessment of WMLs only), other 67 rats were randomly selected and underwent 2VO surgery.

### Model Establishment and Experimental Design

Rats underwent the surgery of permanent bilateral common carotid artery occlusion (2VO) to establish CCH as previously described (Tian et al. 2014; Back et al. 2017). Briefly, the bilateral common carotid arteries were exposed after a midline cervical incision. The bilateral nerve fibers should be gently separated. Then the bilateral arteries were permanently ligated with silk suture. Temperature was maintained at 37.5 °C using a heat pad during the surgery. Sham operation was done without ligation of the arteries. A total of 16 rats died during or after the 2VO surgery and the remained 51 rats were randomly divided into vehicle group (treated with PBS), anti-HMGB1 group (treated with anti-HMGB1 Ab) and control Ab group (treated with paired control Ab) ( $n = 17$  in either group). The anti-HMGB1 Ab or the paired control Ab (1 mg/kg, dissolved in 0.9% PBS, Shino-test, Kanagawa, Japan) were injected via the tail vein after 2VO surgery for thrice. The detailed information of dose study, efficacy, and method of tail vein injection was described in our previous research (Hei et al. 2018). Specifically, as anti-HMGB1 therapy turned out good in acute WMLs (Hayakawa et al. 2013; Choi et al. 2017), we delayed the time of injection [Day 4–Day6, which is in the sub-acute phase of CCH (Farkas et al. 2007; Soria et al. 2013)] to see anti-HMGB1's effects in the chronic phase of CCH (28 days after surgery) without interfering with HMGB1's beneficial functions in the acute phase. The experimental schedule was presented in the Fig. 1a.

### Klüver-Barrera (KB) and Hematoxylin–Eosin (H&E) Staining

All animals ( $N = 6$  in each group) that assigned to histological analyses of CCH-induced white matter damages were intraperitoneally anesthetized with chloral hydrate (10%, 3 ml/kg) and transcardially perfused with 4% paraformaldehyde in PBS. The rat brains were processed into coronal sections (5  $\mu$ m, from bregma  $-2.04$  to  $-3.36$  mm) (Wang et al. 2010) and mounted on polarized glasses. The KB staining was conducted to evaluate the general changes in white matter regions including the corpus callosum (CC), internal capsule (IC), and optic tract (OT) (assessing the grade scores and the percentage of thickness) (Tachibana et al. 2017). The tissue sections were incubated with 0.1% Luxol fast blue (Sigma, MO, USA) at 56 °C overnight and the severity of damages were graded as normal (grade 0), disarrangement of the nerve fibers (grade 1), formation of marked vacuoles (grade 2), and the disappearance of myelinated fibers (grade



**Fig. 1** The experimental schedule. Rats were pre-trained for behavioral tests 3 days before 2VO surgery and pupillary light reflex (PLR), behavioral tests, histological assessment and western blotting were

carried out on day 28 (which is in the chronic phase of CCH) after anti-HMGB1 treatment (day 4–day 6, which is in the sub-acute phase of CCH)

3) (Wakita et al. 1994, 2002; Tachibana et al. 2017). And the (%) of thickness within a predetermined region-of-interest (ROI) was measured by mode methods (Takizawa et al. 2003; Edrissi et al. 2016; Tachibana et al. 2017) and the value in the sham group was defined as 100% (thus the value in the sham group was presented without any error bars). Furthermore, the hematoxylin–eosin (H&E) staining was conducted to validate the vacuolation of the OT area. Briefly, sections were washed (in xylol for 10 min, ethanol (100%) for 5 min, ethanol (80%) for 5 min), incubated in hematoxylin solution (0.1%, Beyotime, Shanghai, China), washed in water for 5 min, and incubated in alcohol eosin solution (0.5%, Beyotime) and dehydrated. The % area of vacuoles was calculated in the same way as the % of thickness.

### Behavioral Assessment

Rats ( $N=6$  in the sham group;  $N=5$  in the vehicle group, the anti-HMGB1 group and the control Ab group) underwent behavioral assessment including the pupillary light reflex (PLR) examination (at 9:00 a.m. before the beam-walking test), tapered beam-walking test (after the PLR), and visual water maze (at 3:00 p.m.) 28 days after 2VO surgery.

First of all, the PLR was examined by two investigators that are blind to the groups (Lavinsky et al. 2006). Briefly, rats were allowed to adapt to darkness for 1 min prior to the examination. Then a light from an otoscope was directed to the right eye for evaluating the direct PLR. Subsequently, the beam quickly moved to the left eye to assess the indirect reflex. The same procedure was conducted on the other eye with time interval of 1 min. PLR loss was finally defined as the failure of pupillary constriction within a 10 s light exposure. For tapered beam-walking (Edrissi et al. 2016), the test was performed in a quiet room by two investigators that are blind to the groups.

The beam was 1.65 m in length, tapering from 6 cm to 1.5 cm in width with a ledge extending beneath the beam; 1 m above the floor. The time to traverse the beam and the number of foot faults (slip off the beam) were counted in

2 m-distance under a monitor (Smart, Shanghai, China). Afterwards, visual water maze test with a visible platform was performed. The platform was put 1 cm above the surface of the water (could be seen clearly) located in quadrant I. Rats were given maximum 60 s to locate the platform and repeated for 3 times. After finding the platform, the rats were allowed to stay on the platform for additional 20 s. Rats in the four groups were trained for the tapered beam-walking test and visual water maze test 3-day before the 2VO surgery for three times.

### Immunofluorescence

The preparation of the brain sections from rats ( $N=6$  in each group) were the same as above. After washing with PBS, sections were incubated with Triton x-100 (0.1%) and 5% BSA for 30 min. Then the primary antibodies were added including rabbit anti-Iba1 (1:100, Abcam, CA, USA) and mouse anti-GFAP (1:100, Abcam, CA, USA) at 4 °C overnight. The secondary antibodies (1:1000, Invitrogen, CA, USA) were added accordingly and incubated for 4 h at room temperature. Besides, 0.0001% 4',6-diamidino-2-phenylindole (DAPI) staining (Beyotime) was conducted to stain cell nuclei for 10 min. The images were obtained using confocal laser scanning (Olympus, Japan) (magnification,  $\times 200$ ) and analyzed in gray scale by Image pro 6.0 with a fixed exposure.

### Western Blotting

Following the behavioral tests, rats ( $N=6$  in each group) were decapitated after anesthesia (10% chloral hydrate, 3 ml/kg, i.p.). Tissues of the OT area were carefully separated from the rat brain on ice under microscope and the protein were extracted using BioRad protein assay kit (Hercules, DE, USA). But for the detection of p65 (nuclear expression) and p-p65, the nuclear and cytosolic proteins were obtained using Nuclear & Cytoplasmic Protein Extraction Kit (Beyotime). As previously described (Zhang et al. 2011; Ouyang

et al. 2016), the homogenized protein samples (50 µg) were added to SDS-PAGE. Then the bands of the proteins were transferred to 0.22 µm NC membranes (Whatman, Germany). The primary antibodies of TLR4 (1:1000, Abcam, CO, USA),  $\beta$ -actin and Histone H3 antibody (1:2000, Abcam, CO, USA), and NF- $\kappa$ B p65 and phosphorylated NF- $\kappa$ B p65 (p-p65) (1:1000; Cell Signaling Technology, MA, USA) (known as the important pro-inflammatory molecular signaling that could be regulated by HMGB1), were added to the membranes in blocking solution. After incubation overnight at 4 °C, the membranes were washed and incubated with appropriate peroxidase-conjugated secondary antibody (Santa Cruz Biotechnology, CA, USA) for 1.5 h and the final electrochemiluminescence was conducted with an IPP 6.0 system. Densities were normalized to  $\beta$ -actin and the value of the sham group was defined as 100%.

### Statistical Analysis

Data are expressed as mean  $\pm$  s.e.m. Multiple comparisons in behavioral tests and morphological changes in the OT area were analyzed using one-way ANOVA followed by post hoc Bonferroni's test using SPSS 22.0.0 and GraphPad Prism 7.0.1 (GraphPad software). The  $P < 0.05$  were considered to be statistically significant.

## Results

### Anti-HMGB1 Ab Ameliorated the CCH-Induced WMLs Especially in the OT Subarea in the Chronic Phase

As the KB staining was frequently used to show the WMLs (expressed as grade scores and thickness), firstly we performed such experiment 28 days after 2VO surgery to show the damages in the CC, IC, and OT regions (Fig. 2). Compared with the vehicle group, the anti-HMGB1 group showed a decrease in the grade scores ( $F_{(4,25)} = 73.51$ ,  $P < 0.0001$ ) and an increase of thickness ( $F_{(4,25)} = 96.77$ ,  $P < 0.001$ ) in the OT area, and showed only a decrease of grade scores ( $F_{(4,25)} = 67.8$ ,  $P < 0.0001$ ) in the CC area, and showed no significant difference in the IC area ( $P > 0.05$ ). It should be noted that the sham + anti-HMGB1 group showed no significant alteration either in the grade scores or thickness in the KB staining in comparison to the sham group ( $P > 0.05$ ), indicating that the anti-HMGB1 treatment alone has no visible influence on white matter. We therefore focused on the investigation of CCH-induced OT injuries within the other four groups and further used H&E staining to verify the results (Fig. 3). We found that there were significant increases in the grade scores ( $F_{(3,20)} = 56.06$ ,  $P < 0.0001$ ) and vacuoles ( $F_{(3,20)} = 49.75$ ,  $P < 0.0001$ ) and there was a decrease of thickness ( $F_{(3,20)} = 87.35$ ,  $P < 0.001$ )

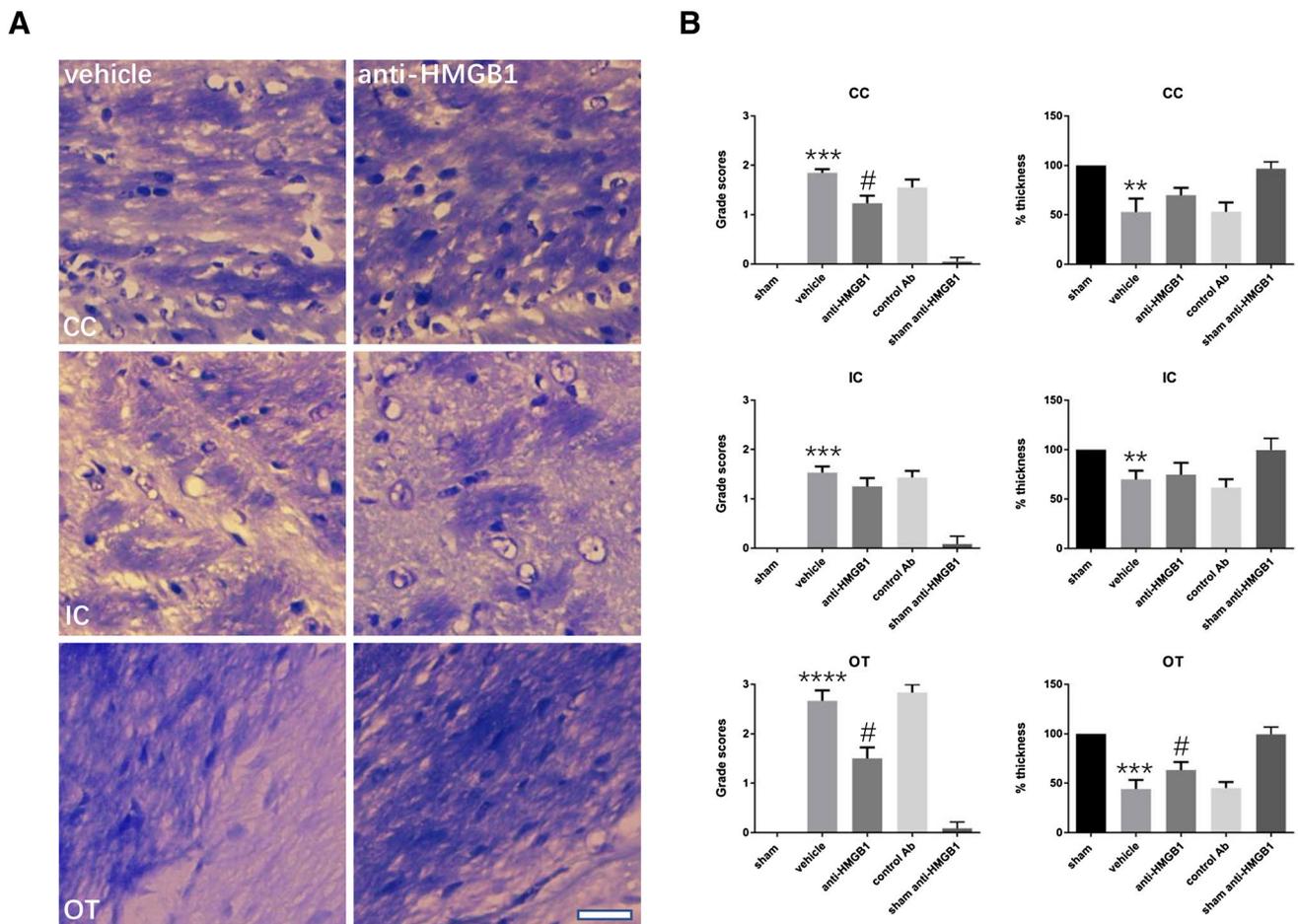
in the vehicle group in comparison with the sham group. Anti-HMGB1 group presented dramatically reduced damages (grade score:  $P < 0.05$ ; thickness:  $P < 0.05$ ; vacuoles:  $P < 0.001$  vs. the vehicle group), indicating that anti-HMGB1 Ab significantly ameliorated the OT injuries in the chronic phase of CCH. Meanwhile, no significant difference in the above parameters was found between the vehicle group and the control Ab group.

### Anti-HMGB1 Ab Improved Behavioral Performances in the Visual Water Maze and Tapered Beam-Walking Tests in the 2VO Model

First of all, the pupillary light reflex (PLR) examination was conducted (Fig. 4a). Nearly 50% rats in the vehicle group lost the PLR ( $F_{(3,17)} = 64.12$ ,  $P < 0.001$  vs. the sham group), but the anti-HMGB1 treatment reduced the PLR loss ( $P < 0.05$ ) in comparison with the vehicle group. In the visual water maze tests (Fig. 4b), the vehicle group presented reduced percentage of platform findings compared with the sham group ( $F_{(3,17)} = 49.72$ ,  $P < 0.01$ ). And again, anti-HMGB1 group showed better performances compared to the vehicle group ( $P < 0.05$ ). Besides, we further investigated the visual-based motor ability using the beam-walking test (Fig. 4c, d). Similarly, the vehicle group showed longer time latencies ( $F_{(3,17)} = 19.52$ ,  $P < 0.001$ ) and foot faults ( $F_{(3,17)} = 11.66$ ,  $P < 0.001$ ) in comparison to the sham group and rats in the anti-HMGB1 group performed better compared to the vehicle group (time latency:  $P < 0.01$ ; foot faults:  $P < 0.05$ ). In addition, rats in the control Ab group performed worse than the ones in the sham group, but no significant difference in the above parameters was found between the vehicle group and the control Ab group.

### Anti-HMGB1 Ab Suppressed Inflammatory Responses Characterized by the Activation of Glial Cells and TLR4/NF- $\kappa$ B Signaling

Meanwhile, to further evaluate the level of glial activation in the OT area, we performed Iba1 and GFAP immunostaining (Fig. 5). The results showed that the intensity of Iba1 and GFAP staining was significantly up-regulated in the vehicle group in comparison to the sham group. However, after anti-HMGB1 treatment, these two parameters were relatively down-regulated compared with the vehicle group. To further clarify the molecular mechanisms associated with inflammatory responses in the OT region, western blotting analysis was performed 28 days after surgery. As shown in Fig. 6, the relative protein expressions of TLR4 ( $F_{(3,20)} = 20.11$ ,  $P < 0.05$ ), p65 ( $F_{(3,20)} = 35.12$ ,  $P < 0.05$ ), and p-p65 ( $F_{(3,20)} = 22.41$ ,  $P < 0.05$ ) significantly increased in comparison with the sham group. However, anti-HMGB1 treatment partly suppressed the activation of the TLR4



**Fig. 2** The influence of anti-HMGB1 Ab on the CCH-induced white matter injuries. **a** Representative images of the KB staining in white matter areas including the corpus callosum (CC), internal capsule (IC), and optic tract (OT) in the vehicle group (left) and anti-HMGB1 group (right). **b** The quantification of the injuries (including the grade

score and the percentage (%) of thickness) among the five groups in different subareas in the white matter. Bar=50  $\mu$ m. Data are expressed as mean  $\pm$  s.e.m. \*\* $P < 0.01$ , \*\*\* $P < 0.001$ , \*\*\*\* $P < 0.0001$  versus sham group; # $P < 0.05$  versus vehicle group

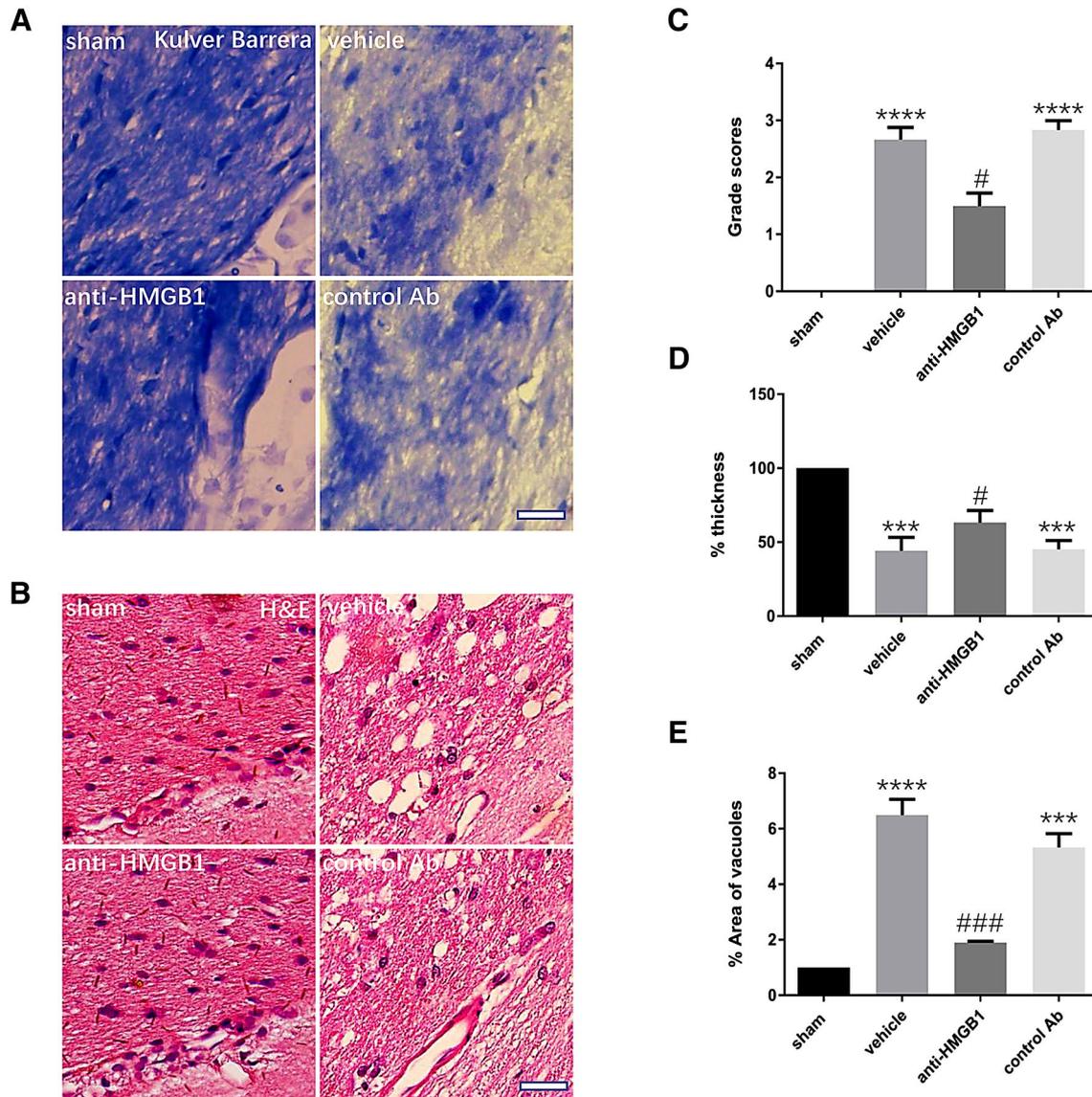
( $P < 0.05$ ), p65 ( $P < 0.05$ ), p-p65 ( $P < 0.05$ ) as compared to the vehicle group. Besides, no significant difference was found between the vehicle group and the control Ab group, indicating that the control Ab has no visible influence on the expressions of TLR4/NF- $\kappa$ B signaling.

## Discussion

CCH causes progressive WMLs, which appear 5 days after 2VO surgery and are persistent in the chronic phase (Davidson et al. 2000; Tian et al. 2014). Specifically, the white matter, which is predominantly consisted of glia and myelinated axons, presents remarkable demyelination, vacuolation, axonal damage, and glial activation during CCH, and the OT region is selectively damaged; leading to visually guided behavioral deficits (Wakita et al. 1994; Edrissi et al. 2016). Previously we have used anti-HMGB1 neutralizing

antibody to treat CCH-induced hippocampal damages via inhibition of neuroinflammation. However, it is still lack of reports about the therapeutic role of HMGB1 neutralization on the chronic OT injuries in CCH.

The white matter areas display varying degrees of vulnerabilities to CCH, and given the fact that the OT region is dependent on the direct blood supply of the internal carotid artery, damage in this specific optic structure comes heavier in comparison with other white matter regions during CCH (Kim et al. 2009; Zhang et al. 2011). Consistently, in the present study we found increased level of WMLs (especially in the OT area) and the anti-HMGB1 treatment seems to gain much more benefits in this area (using KB and H&E staining). Besides, we also tested the effects of the Ab on sham-operated rats (sham + anti-HMGB1 group) and found that the Ab did not alter the morphological characteristics in the CCH model, which was also in line with others (Festoff et al. 2016; Kigerl et al. 2017). Thus, we focused on the OT



**Fig. 3** The influence of anti-HMGB1 Ab on the CCH-induced OT injuries assessed by KB and H&E staining. **a, c, d** The KB staining in OT area and the quantification of the injuries (including the grade score and the percentage (%) of thickness). **b, e** The H&E

staining and the quantification of the percentage (%) area of vacuoles. Bar=50  $\mu$ m. Data are expressed as mean  $\pm$  s.e.m. \*\*\*\* $P$ <0.0001, \*\*\* $P$ <0.001 versus sham group; # $P$ <0.05, ### $P$ <0.001 versus vehicle group

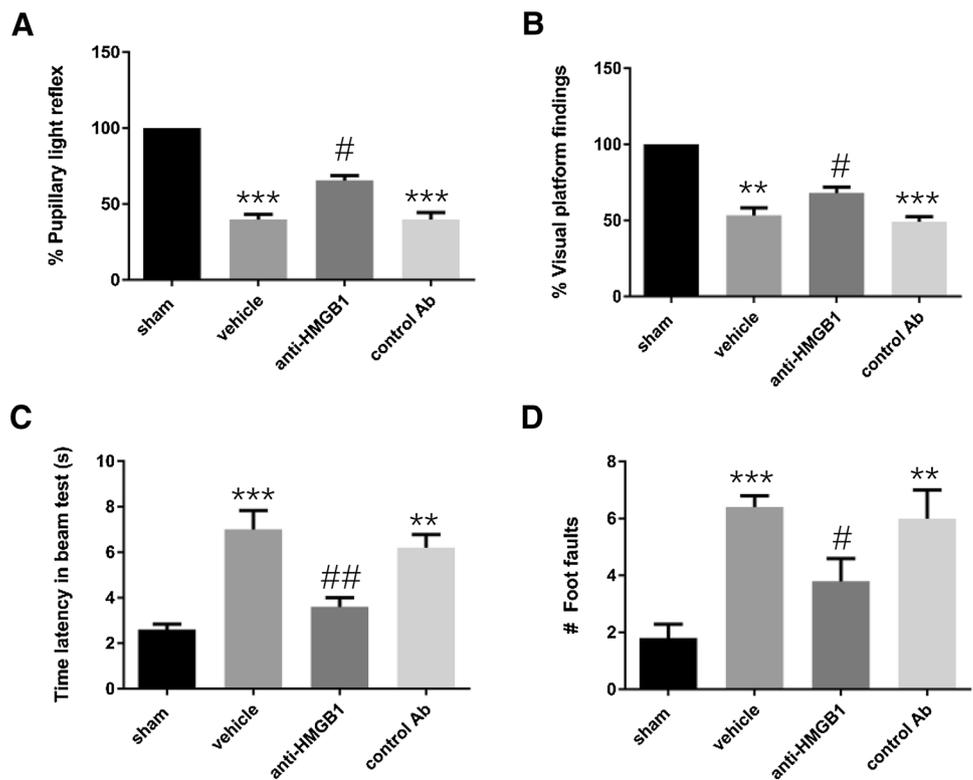
damages to perform the further study using the four groups (sham group, vehicle group, anti-HMGB1 group, and control Ab group).

The presence of dilated pupils and impaired performances in visually guided tasks sufficiently suggested the existence of damages in the visual system. In addition, we further found significant correlations between the existence of PLR and the behavioral ability (both for visual water maze and beam-walking tests) (data not shown). Interestingly, previous studies have shown that while 2VO surgery affects PLR within 5 days, the morphological changes in retinas cannot be assessed until 3 months later (Davidson et al. 2000;

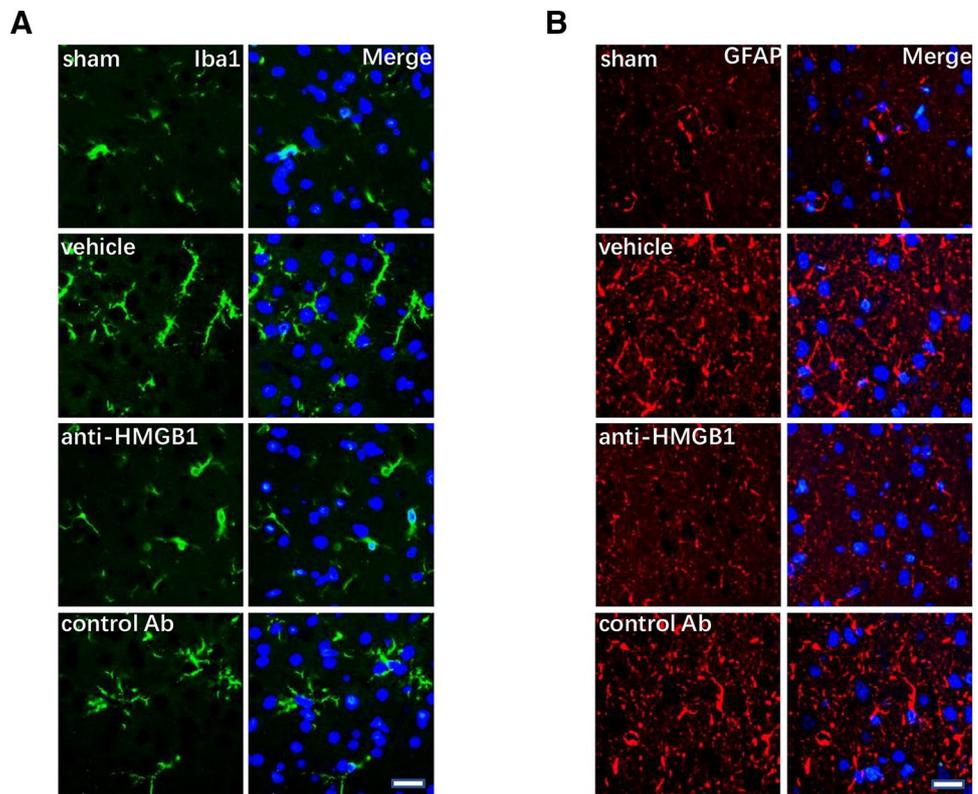
Stevens et al. 2002). Therefore, CCH-induced OT injuries come earlier and heavier in the visual system, indicating that 2VO surgery rapidly damages the optic nerve and causes loss of PLR.

Substantial evidence has demonstrated that inflammatory responses in 2VO rats has spatial preference in the white matter with OT area presenting most severe and prolonged damages during the course of disease (Tian et al. 2014; Kim et al. 2015). In accordance with these findings, we have also found excessive glial activation (including activated microglia and astrocytes) in OT area together with increased degenerative changes, such as rarefaction and vacuolation in

**Fig. 4** The influence of anti-HMGB1 Ab on visual-based behavioral tests. **a** The percentage (%) of PLR. **b** The percentage (%) of visual platform findings in the visual water maze tests. **c, d** Time latency and the number of foot faults in the tapered beam-walking tests. Data are expressed as mean  $\pm$  s.e.m.  $**P < 0.01$ ,  $***P < 0.001$  versus sham group;  $\#P < 0.05$ ,  $\#\#P < 0.01$  versus vehicle group

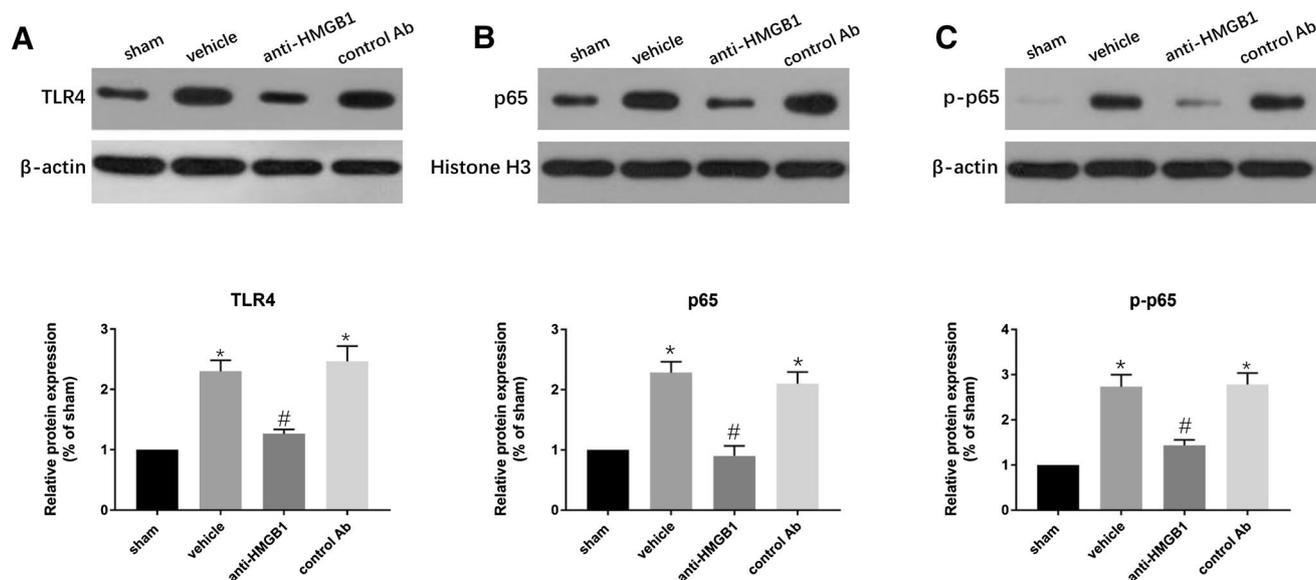


**Fig. 5** The influence of anti-HMGB1 Ab on the CCH-induced glial activation in the OT area. **a** Representative images of Iba1 immunostaining. **b** Representative images of GFAP immunostaining. Bar = 50  $\mu$ m



the chronic phase of CCH. Some studies have demonstrated that in the early stage of CCH the activated astrocytes could

release neurotrophic factors (which help axonal regrowth) and to the contrary, the excessive gliosis hinders axonal



**Fig. 6** The influence of anti-HMGB1 Ab on suppression of the activation of TLR4/NF- $\kappa$ B signaling in the OT area. **a, c** The protein bands and the corresponding quantification of TLR4 (**a**), NF- $\kappa$ B p65 (**b**) and phosphorylated NF- $\kappa$ B p65 (p-p65) (**c**). The value

of the sham group was defined as 100%. Data are expressed as mean  $\pm$  s.e.m. \* $P$  < 0.05 versus sham group; # $P$  < 0.05 versus vehicle group

growth and starts its glial-scar formation process in the chronic phase of CCH and in other neurodegenerative diseases (Maragakis and Rothstein 2006; Phatnani and Maniatis 2015). Thus, it stands a good chance that inflammatory responses plays a core role in the chronic phase of CCH-induced WMLs (rather than in the acute phase), the anti-HMGB1 treatment was done early in the sub-acute phase in consideration of the BBB penetration of the neutralizing Ab and the early activation of HMGB1 signaling during CCH (Lee et al. 2015; Hei et al. 2018). After all, these findings strongly suggested that inhibition of CCH-induced inflammatory responses probably lead to beneficial outcomes in the chronic phase.

HMGB1, one damage-associated molecular patterns (DAMPs) molecule, has been well-established to exert its pro-inflammatory cytokine-like effects once activated in various diseases (mainly through the pro-inflammatory receptors RAGE and TLR4) (Liu et al. 2007; Terrando et al. 2016; Wang et al. 2017). Both we and others have shown the therapeutic role of anti-HMGB1 Ab in behavioral and biochemical improvement in various inflammatory conditions in CNS (Festoff et al. 2016; Kigerl et al. 2017). Although recent studies have shown that HMGB1 is capable of promoting oligodendrocytes survival, re-myelination of the axons and the migration of EPCs after acute WMLs (Hayakawa et al. 2013; Choi et al. 2017), the chronic WMLs show difference in the pattern of glial activation (as described above) and hypoxia-related gene expressions (Bang et al. 2013; Yang et al. 2013). Specifically, hypoxia

inducible factor (HIF)-1 $\alpha$  and the associated pro-survival factors (like glucose transport 1 and endothelial growth factor), which could induce HMGB1 relocation (Andrassy et al. 2008), significantly decreased in the chronic phase of CCH (Bang et al. 2013; Yang et al. 2013). Therefore, the function of HMGB1 signaling (both the pattern of HMGB1 mobilization and the final effects on glial activation) probably changes accordingly. In agreement with these findings, we revealed that HMGB1 neutralization is capable of suppressing inflammatory responses in the chronic phase of CCH, suggesting that excessive HMGB1 signaling in the OT area probably leads to detrimental results. Previous studies have also used anti-inflammatory methods to treat CCH-induced WMLs and demonstrated that the inhibition of HMGB1 signaling (such as TLR4/NF- $\kappa$ B) results in various beneficial effects such as the up-regulation of myelin basic protein (MBP) and cholinergic transmission and down-regulation of glial activation and pro-inflammatory cytokine production (Kim et al. 2015; Lee et al. 2015; Kim et al. 2016; Saggiu et al. 2016; Hei et al. 2018), which are in line with ours.

In conclusion, this study demonstrated that anti-HMGB1 neutralizing Ab treatment eases CCH-induced optic tract injuries in the white matter associated with down-regulation of inflammatory responses characterized by the activation of glial cells and HMGB1's down-stream TLR4/NF- $\kappa$ B signaling. This study further elucidates the significance of inhibition of HMGB1 signaling and glial activation in the chronic phase of CCH, which may provide new targets for

development of molecular therapies to treat CCH-induced WMLs in related vascular disorders and neurodegenerative diseases.

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**Author Contributions** YH and XZ performed the experiments, RC and YZ wrote the manuscript, DG and WL designed the experiments, and WL is responsible for the final version. All authors read and approved the final manuscript.

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**Data Availability** All data generated or analyzed during this study are included in this published article.

## Compliance with Ethical Standards

**Conflict of interest** No conflict of interest was declared.

**Ethical Approval** National Institutes of Health Ethic Committee gave permission to the animal experiments, which were conducted in accordance with the National Experimental Animals Guidelines.

## References

- Akinyemi RO, Mukaetova-Ladinska EB, Attems J, Ihara M, Kalaria RN (2013) Vascular risk factors and neurodegeneration in ageing related dementias: Alzheimer's disease and vascular dementia. *Curr Alzheimer Res* 10:642–653. <https://doi.org/10.2174/15672050113109990037>
- Andrassy M, Volz HC, Igwe JC, Funke B, Eichberger SN, Kaya Z et al (2008) High-mobility group box-1 in ischemia-reperfusion injury of the heart. *Circulation* 117:3216–3226. <https://doi.org/10.1161/circulation.108.769331>
- Ashok A, Rai NK, Raza W, Pandey R, Bandyopadhyay S (2016) Chronic cerebral hypoperfusion-induced impairment of Abeta clearance requires HB-EGF-dependent sequential activation of HIF1alpha and MMP9. *Neurobiol Dis* 95:179–193. <https://doi.org/10.1016/j.nbd.2016.07.013>
- Back DB, Kwon KJ, Choi DH, Shin CY, Lee J, Han SH et al (2017) Chronic cerebral hypoperfusion induces post-stroke dementia following acute ischemic stroke in rats. *J Neuroinflamm* 14:216. <https://doi.org/10.1186/s12974-017-0992-5>
- Bang J, Jeon WK, Lee IS, Han JS, Kim BY (2013) Biphasic functional regulation in hippocampus of rat with chronic cerebral hypoperfusion induced by permanent occlusion of bilateral common carotid artery. *PLoS ONE* 8:e70093. <https://doi.org/10.1371/journal.pone.0070093>
- Braun M, Vaibhav K, Saad NM, Fatima S, Vender JR, Baban B et al (2017) White matter damage after traumatic brain injury: a role for damage associated molecular patterns. *Biochim Biophys Acta Mol Basis Dis* 1863:2614–2626. <https://doi.org/10.1016/j.bbdis.2017.05.020>
- Cardoso FL, Herz J, Fernandes A, Rocha J, Sepodes B, Brito MA et al (2015) Systemic inflammation in early neonatal mice induces transient and lasting neurodegenerative effects. *J Neuroinflamm* 12:82. <https://doi.org/10.1186/s12974-015-0299-3>
- Choi B, Kim D, Back DB, Kang CH, Moon W, Han J et al (2016) Characterization of white matter injury in a rat model of chronic cerebral hypoperfusion. *Stroke* 47:542–547. <https://doi.org/10.1161/stroke.115.011679>
- Choi JY, Cui Y, Chowdhury ST, Kim BG (2017) High-mobility group box-1 as an autocrine trophic factor in white matter stroke. *Proc Natl Acad Sci USA* 114:E4987–E4995. <https://doi.org/10.1073/pnas.1702035114>
- Davidson CM, Pappas BA, Stevens WD, Fortin T, Bennett SAL (2000) Chronic cerebral hypoperfusion: loss of pupillary reflex, visual impairment and retinal neurodegeneration. *Brain Res* 859:96–103. [https://doi.org/10.1016/S0006-8993\(00\)01937-5](https://doi.org/10.1016/S0006-8993(00)01937-5)
- Edrissi H, Schock SC, Cadonic R, Hakim AM, Thompson CS (2016) Cilostazol reduces blood brain barrier dysfunction, white matter lesion formation and motor deficits following chronic cerebral hypoperfusion. *Brain Res* 1646:494–503. <https://doi.org/10.1016/j.brainres.2016.06.036>
- Farkas E, Luiten PG (2001) Cerebral microvascular pathology in aging and Alzheimer's disease. *Prog Neurobiol* 64:575–611
- Farkas E, Luiten PGM, Bari F (2007) Permanent, bilateral common carotid artery occlusion in the rat: a model for chronic cerebral hypoperfusion-related neurodegenerative diseases. *Brain Res Rev* 54:162–180. <https://doi.org/10.1016/j.brainresrev.2007.01.003>
- Festoff BW, Sajja RK, van Dreden P, Cucullo L (2016) HMGB1 and thrombin mediate the blood-brain barrier dysfunction acting as biomarkers of neuroinflammation and progression to neurodegeneration in Alzheimer's disease. *J Neuroinflamm* 13:194. <https://doi.org/10.1186/s12974-016-0670-z>
- Hamada T, Torikai M, Kuwazuru A, Tanaka M, Horai N, Fukuda T et al (2008) Extracellular high mobility group box chromosomal protein 1 is a coupling factor for hypoxia and inflammation in arthritis. *Arthritis Rheum* 58:2675–2685. <https://doi.org/10.1002/art.23729>
- Hayakawa K, Miyamoto N, Seo JH, Pham LD, Kim KW, Lo EH et al (2013) High-mobility group box 1 from reactive astrocytes enhances the accumulation of endothelial progenitor cells in damaged white matter. *J Neurochem* 125:273–280. <https://doi.org/10.1111/jnc.12120>
- Hei Y, Chen R, Yi X, Long Q, Gao D, Liu W (2018) HMGB1 neutralization attenuates hippocampal neuronal death and cognitive impairment in rats with chronic cerebral hypoperfusion via suppressing inflammatory responses and oxidative stress. *Neuroscience* 383:150–159. <https://doi.org/10.1016/j.neuroscience.2018.05.010>
- Kigerl KA, Lai W, Wallace LM, Yang H, Popovich PG (2017) High mobility group box-1 (HMGB1) is increased in injured mouse spinal cord and can elicit neurotoxic inflammation. *Brain Behav Immun* 72:22–33. <https://doi.org/10.1016/j.bbi.2017.11.018>
- Kim SK, Cho KO, Kim SY (2008) White matter damage and hippocampal neurodegeneration induced by permanent bilateral occlusion of common carotid artery in the rat: comparison between Wistar and Sprague-Dawley strain. *Korean J Physiol Pharmacol* 12:89–94. <https://doi.org/10.4196/kjpp.2008.12.3.89>
- Kim SK, Cho KO, Kim SY (2009) The plasticity of posterior communicating artery influences on the outcome of white matter injury induced by chronic cerebral hypoperfusion in rats. *Neurol Res* 31:245–250. <https://doi.org/10.1179/174313209X382278>
- Kim MS, Bang JH, Lee J, Kim HW, Sung SH, Han JS et al (2015) Salvia miltiorrhiza extract protects white matter and the hippocampus from damage induced by chronic cerebral hypoperfusion in rats. *BMC Complement Altern Med* 15:415. <https://doi.org/10.1186/s12906-015-0943-6>
- Kim MS, Bang JH, Lee J, Han JS, Baik TG, Jeon WK (2016) *Ginkgo biloba* L. extract protects against chronic cerebral hypoperfusion by modulating neuroinflammation and the cholinergic system.

- Phytomedicine 23:1356–1364. <https://doi.org/10.1016/j.phymed.2016.07.013>
- Lavinsky D, Arterni NS, Achaval M, Netto CA (2006) Chronic bilateral common carotid artery occlusion: a model for ocular ischemic syndrome in the rat. *Graefes Arch Clin Exp Ophthalmol* 244:199–204. <https://doi.org/10.1007/s00417-005-0006-7>
- Lee JH, Park SY, Shin YW, Hong KW, Kim CD, Sung SM et al (2006) Neuroprotection by cilostazol, a phosphodiesterase type 3 inhibitor, against apoptotic white matter changes in rat after chronic cerebral hypoperfusion. *Brain Res* 1082:182–191. <https://doi.org/10.1016/j.brainres.2006.01.088>
- Lee KM, Bang J, Kim BY, Lee IS, Han JS, Hwang BY et al (2015) Fructus mume alleviates chronic cerebral hypoperfusion-induced white matter and hippocampal damage via inhibition of inflammation and downregulation of TLR4 and p38 MAPK signaling. *BMC Complement Altern Med* 15:125. <https://doi.org/10.1186/s12906-015-0652-1>
- Liu K, Mori S, Takahashi HK, Tomono Y, Wake H, Kanke T et al (2007) Anti-high mobility group box 1 monoclonal antibody ameliorates brain infarction induced by transient ischemia in rats. *FASEB J* 21:3904–3916. <https://doi.org/10.1096/fj.07-8770com>
- Maragakis NJ, Rothstein JD (2006) Mechanisms of disease: astrocytes in neurodegenerative disease. *Nat Clin Pract Neurol* 2:679–689. <https://doi.org/10.1038/ncpneuro0355>
- Ouyang F, Huang H, Zhang M, Chen M, Huang H, Huang F et al (2016) HMGB1 induces apoptosis and EMT in association with increased autophagy following H/R injury in cardiomyocytes. *Int J Mol Med* 37:679–689. <https://doi.org/10.3892/ijmm.2016.2474>
- Phatnani H, Maniatis T (2015) Astrocytes in neurodegenerative disease. *Cold Spring Harb Perspect Biol*. <https://doi.org/10.1101/cshperspect.a020628>
- Pisetsky DS (2014) The translocation of nuclear molecules during inflammation and cell death. *Antioxid Redox Sign* 20:1117–1125. <https://doi.org/10.1089/ars.2012.5143>
- Saggu R, Schumacher T, Gerich F, Rakers C, Tai K, Delekate A et al (2016) Astroglial NF- $\kappa$ B contributes to white matter damage and cognitive impairment in a mouse model of vascular dementia. *Acta Neuropathol Commun* 4:76. <https://doi.org/10.1186/s40478-016-0350-3>
- Soria G, Tudela R, Márquez-Martín A, Camón L, Batalle D, Muñoz-Moreno E et al (2013) The ins and outs of the BCCAo model for chronic hypoperfusion: a Multimodal and longitudinal MRI approach. *PLoS ONE* 8:e74631. <https://doi.org/10.1371/journal.pone.0074631>
- Stevens WD, Fortin T, Pappas BA (2002) Retinal and optic nerve degeneration after chronic carotid ligation: time course and role of light exposure. *Stroke* 33:1107–1112
- Tachibana M, Ago T, Wakisaka Y, Kuroda J, Shijo M, Yoshikawa Y et al (2017) Early reperfusion after brain ischemia has beneficial effects beyond rescuing neurons. *Stroke* 48:2222–2230. <https://doi.org/10.1161/stroke.117.016689>
- Takizawa S, Fukuyama N, Hirabayashi H, Kohara S, Kazahari S, Shinohara Y et al (2003) Quercetin, a natural flavonoid, attenuates vacuolar formation in the optic tract in rat chronic cerebral hypoperfusion model. *Brain Res* 980:156–160. [https://doi.org/10.1016/S0006-8993\(03\)03009-9](https://doi.org/10.1016/S0006-8993(03)03009-9)
- Terrando N, Yang T, Wang X, Fang J, Cao M, Andersson U et al (2016) Systemic HMGB1 neutralization prevents postoperative neurocognitive dysfunction in aged rats. *Front Immunol* 7:441. <https://doi.org/10.3389/fimmu.2016.00441>
- Tian XS, Guo XJ, Ruan Z, Lei Y, Chen YT, Zhang HY (2014) Long-term vision and non-vision dominant behavioral deficits in the 2-VO rats are accompanied by time and regional glial activation in the white matter. *PLoS ONE* 9:e101120. <https://doi.org/10.1371/journal.pone.0101120>
- Ueno Y, Koike M, Shimada Y, Shimura H, Hira K, Tanaka R et al (2015) L-Carnitine enhances axonal plasticity and improves white-matter lesions after chronic hypoperfusion in rat brain. *J Cereb Blood Flow Metab* 35:382–391. <https://doi.org/10.1038/jcbfm.2014.210>
- Wakita H, Tomimoto H, Akiguchi I, Kimura J (1994) Glial activation and white matter changes in the rat brain induced by chronic cerebral hypoperfusion: an immunohistochemical study. *Acta Neuropathol* 87:484–492. <https://doi.org/10.1007/bf00294175>
- Wakita H, Tomimoto H, Akiguchi I, Matsuo A, Lin J, Ihara M et al (2002) Axonal damage and demyelination in the white matter after chronic cerebral hypoperfusion in the rat. *Brain Res* 924:63–70. [https://doi.org/10.1016/S0006-8993\(01\)03223-1](https://doi.org/10.1016/S0006-8993(01)03223-1)
- Wang J, Zhang HY, Tang XC (2010) Huperzine improves chronic inflammation and cognitive decline in rats with cerebral hypoperfusion. *J Neurosci Res* 88:807–815. <https://doi.org/10.1002/jnr.22237>
- Wang D, Liu K, Wake H, Teshigawara K, Mori S, Nishibori M (2017) Anti-high mobility group box-1 (HMGB1) antibody inhibits hemorrhage-induced brain injury and improved neurological deficits in rats. *Sci. Rep* 7:46243. <https://doi.org/10.1038/srep46243>
- Yang Y, Zhang J, Liu H, Wang J, Xin J, Deng M (2013) Changes in levels of hypoxia-induced mediators in rat hippocampus during chronic cerebral hypoperfusion. *Neurochem Res* 38:2433–2439. <https://doi.org/10.1007/s11064-013-1158-1>
- Zhang G, Zhao Z, Gao L, Deng J, Wang B, Xu D et al (2011) Gypenoside attenuates white matter lesions induced by chronic cerebral hypoperfusion in rats. *Pharmacol Biochem Be.* 99:42–51. <https://doi.org/10.1016/j.pbb.2011.03.019>

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