

Regardless of the designation employed for this neoplasm, we believe reporting this case can be helpful in calling this issue to the attention of others, with the hope that additional cases will be reported and perhaps lead to a consensus for the optimal terminology for cases such as that described in this report.

Acknowledgements: We acknowledge that Dr Guilin Tang of the University of Texas MD Anderson Cancer Center kindly helped us perform and interpret the FISH testing for *DUSP22-IRF4* rearrangement.

Conflicts of interest and sources of funding: No research funding was obtained for this study. The authors state that there are no conflicts of interest to disclose.

Brenda Mai¹, Wei Wang², L. Jeffrey Medeiros², Hilary Y. Ma³, Zhihong Hu¹

¹Department of Pathology, The University of Texas Health Center at Houston, Houston, TX, United States; ²Department of Hematopathology, The University of Texas MD Anderson Cancer Center, Houston, TX, United States; ³Department of General Oncology, The University of Texas MD Anderson Cancer Center, TX, United States

Contact Zhihong Hu, MD, PhD.

E-mail: zhihong.hu@uth.tmc.edu

1. Swerdlow SH, Campo E, Seto M, *et al.* Follicular lymphoma. In: Swerdlow SH, Campo E, Harris NL, *et al.*, editors. *World Health Organization Classification of Tumours of Haematopoietic and Lymphoid Tissues*. Revised 4th ed. Lyon: IARC Press, 2017; 266–90.
2. Lee JT, Innes Jr DJ, Williams ME. Sequential bcl-2 and c-myc oncogene rearrangements associated with the clinical transformation of non-Hodgkin's lymphoma. *J Clin Invest* 1989; 84: 1454–9.
3. Horn H, Schmelter C, Leich E, *et al.* Follicular lymphoma grade 3B is a distinct neoplasm according to cytogenetic and immunohistochemical profiles. *Haematologica* 2011; 96: 1327–34.
4. Wagner-Johnston ND, Link BK, Byrtek M, *et al.* Outcomes of transformed follicular lymphoma in the modern era: a report from the National LymphoCare Study (NCLS). *Blood* 2015; 126: 851–7.
5. Beral B, Peterman T, Berkelman R, *et al.* AIDS-associated non-Hodgkin lymphoma. *Lancet* 1991; 337: 805–9.
6. Raphael M, Borisch B, Jaffe E. Lymphomas associated with infection by the human immune deficiency virus (HIV). In: Jaffe E, Harris N, Stein H, *et al.*, editors. *World Health Organization Classification of Tumours. Pathology and Genetics of Tumours of Haematopoietic and Lymphoid Tissues*. Lyon: IARC Press, 2001; 260–3.
7. Agarwal AM, Agarwal N, Glenn MJ. Blastic transformation of low-grade follicular lymphoma. *J Clin Oncol* 2007; 25: 2326–8.
8. Swerdlow SH, Campo E, Seto M, *et al.* High-grade B-cell lymphoma. Revised. In: Swerdlow SH, Campo E, Harris NL, *et al.*, editors. *World Health Organization Classification of Tumours of Haematopoietic and Lymphoid Tissues*. 4th ed. Lyon: IARC Press, 2017; 335–42.
9. Landsburg DJ, Petrich AM, Abramson JS, *et al.* Impact of oncogene rearrangement patterns on outcomes in patients with double-hit non-Hodgkin lymphoma. *Cancer* 2016; 122: 559–64.
10. Uchida A, Isobe Y, Uemura Y, *et al.* De novo acute lymphoblastic leukemia-like disease of high grade B-cell lymphoma with MYC and BCL2 and/or BCL6 rearrangements: a case report and literature review. *BMC Clin Pathol* 2017; 17: 21–9.
11. Geyer JT, Subramaniam S, Jiang Y, *et al.* Lymphoblastic transformation of follicular lymphoma: a clinicopathologic and molecular analysis of 7 patients. *Hum Pathol* 2015; 46: 260–71.
12. Ok CY, Medeiros LJ, Thakral B, *et al.* High-grade B-cell lymphomas with Tdt expression: a diagnostic and classification dilemma. *Mod Pathol* 2019; 32: 48–58.

DOI: <https://doi.org/10.1016/j.pathol.2019.08.009>

Anti-Xa levels with low molecular weight heparin calibrator can be used to exclude significant apixaban effect



Sir,

Apixaban levels can be measured using an anti-Xa assay with apixaban-specific calibrator. However, as apixaban levels are infrequently required and the specific calibration and control material has limited stability at room temperature, the commitment of resources to perform an apixaban level may not be deemed justifiable at some laboratories. Therefore, they may not offer the assay, or limit availability to restricted hours of operation. In contrast anti-Xa levels for low molecular weight heparin (LMWH) are more commonly requested, and easier to perform after hours. We sought to correlate apixaban levels with anti-Xa-LMWH, with the aim of identifying a threshold anti-Xa-LMWH level that would allow exclusion of therapeutic apixaban levels in emergency situations.

We collected blood from 28 patients who had been taking apixaban for at least 3 days, and performed anti-Xa using STA-Liquid Anti-Xa kit with STA-Multihep calibrator and STA-apixaban calibrator (Stago, France). The demographic data are shown in [Table 1](#), and the correlation curve is shown in [Fig. 1](#).

At lower levels of apixaban, there is a near-linear relationship between apixaban level and anti-Xa-LMWH. These findings are consistent with previous studies demonstrating a linear relationship between anti-Xa-LMWH activity and apixaban levels in this range measured using either liquid chromatography tandem mass spectrometry (LC-MS/MS) or an apixaban-calibrated anti-Xa assay.^{1–3} The 'on target' range for apixaban is quoted as being 20–100 ng/mL for 2.5 mg twice daily dosing and 30–412 ng/mL for 5 mg dosing twice daily.^{4–6} According to our results, 20 ng/mL correlates with an anti-Xa-LMWH of 0.40 IU/mL. Hence, an anti-Xa-

Table 1 Demographics

Apixaban dose	2.5 mg BD	5 mg BD	All samples
<i>n</i>	17 (61%)	11 (39%)	28 (100%)
Gender, <i>n</i>			
Male	11 (65%)	5 (45%)	16 (57%)
Female	6 (35%)	6 (55%)	12 (43%)
Age, years			
Median	66	59	65
Range	47–83	18–87	18–87
Weight, kg			
Median	88	76	81
Range	55–128	50–130	50–130
Creatinine clearance, mL/min			
Median	84.1	100.4	93.2
Range	39.8–115.5	50–136.1	39.8–136.1
Time since ingestion, hours			
Median	7.25	7.42	7.25
Range	4.4–8.75	5–10.17	4.4–10.17
Apixaban level, ng/mL			
Median	48.5	100.9	70.3
Range	21–143.3	30.8–202.5	21–202.5

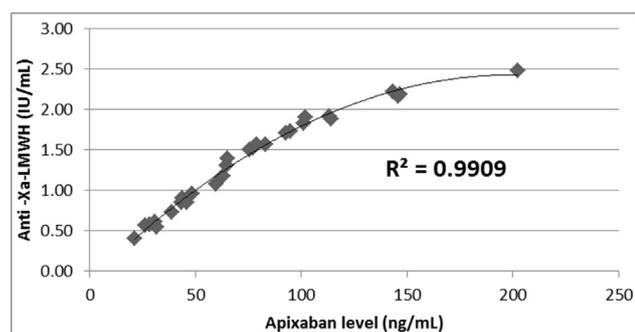


Fig. 1 Anti-Xa-LMWH versus apixaban level.

LMWH result of less than 0.40 IU/mL, for example in a patient going for emergency surgery, can be reassuring that the apixaban level is very low. Our findings are consistent with those of a recently published study.⁷ Therefore, when no apixaban level is available it would be reasonable to use an anti-Xa-LMWH assay to exclude significant apixaban effect in emergency situations.

A similar approach has also been proposed for rivaroxaban.⁸ As illustrated in the correlation curve, the relationship between anti-Xa-LMWH and apixaban levels becomes non-linear as apixaban concentration increases, and therefore anti-Xa-LMWH activity is not a suitable alternative for reliably quantitating apixaban. Furthermore, there is variability between absolute anti-Xa-LMWH activity reported with different commercial reagents and studies.^{8,9} Ideally laboratories should develop their own cut-offs with their own reagents if using anti-Xa-LMWH to exclude apixaban. Importantly, it should also be confirmed that the patient has been receiving apixaban rather than rivaroxaban, as rivaroxaban will also be detected but may have a different cut-off point for exclusion using anti-Xa-LMWH. Conversely, both apixaban (Fig. 1) and rivaroxaban have greater anti-Xa activity than that seen with a similar therapeutic dose of LMWH,^{1–3} so even a small amount of apixaban or rivaroxaban significantly interferes with attempts to measure enoxaparin activity by anti-Xa assay.

Conflicts of interest and sources of funding: This work was funded by Bristol Myers Squibb (BMS)/Pfizer. BMS/Pfizer were not involved in the design or conduct of the study.

J. Singh¹, D. M. Ong¹, A. Wallis¹, G. Kelsey^{1,2}, H. Tran^{1,3}

¹Laboratory Haematology, Alfred Hospital, Melbourne, Vic, Australia; ²Laboratory Haematology, Royal Melbourne Hospital, Melbourne, Vic, Australia; ³Australian Centre for Bleeding Disorders, Monash University, Melbourne, Vic, Australia

Contact Dr Jasmine Singh.
E-mail: snghjsmn07@gmail.com

1. Becker RC, Yang H, Barrett Y, *et al.* Chromogenic laboratory assays to measure the factor Xa-inhibiting properties of apixaban—an oral, direct and selective factor Xa inhibitor. *J Thromb Thrombolysis* 2011; 32: 183–7.
2. Beyer J, Trujillo T, Fisher S, *et al.* Evaluation of a heparin-calibrated antifactor Xa assay for measuring the anticoagulant effect of oral direct Xa inhibitors. *Clin Appl Thromb Hemost* 2016; 22: 423–8.

3. Bonar R, Favaloro E, Mohammed S, *et al.* The effect of the direct factor Xa inhibitors apixaban and rivaroxaban on haemostasis tests: a comprehensive assessment using in vitro and ex vivo samples. *Pathology* 2016; 48: 60–71.
4. Dale BJ, Chan NC, Eikelboom JW. Laboratory measurement of the direct oral anticoagulants. *Br J Haematol* 2016; 172: 315–36.
5. Gosselin RC, Adcock M. The laboratory's 2015 perspective on direct oral anticoagulant testing. *J Thromb Haemost* 2016; 14: 886–93.
6. Samuelson BT, Cuker A, Siegal DM, *et al.* Laboratory assessment of the anticoagulant activity of direct oral anticoagulants: a systematic review. *Chest* 2017; 151: 127–38.
7. Billoir P, Barbay V, Joly LM, *et al.* Anti-Xa oral anticoagulant plasma concentration assay in real life: rivaroxaban and apixaban quantification in emergency with LMWH calibrator. *Ann Pharmacother* 2019; 53: 341–7.
8. Gosselin RC, Francart SJ, Hawes EM, *et al.* Heparin-calibrated chromogenic anti-Xa activity measurements in patients receiving rivaroxaban: can this test be used to quantify drug level? *Ann Pharmacother* 2015; 49: 777–83.
9. Sabor L, Raphaël M, Dogné JM, *et al.* Heparin-calibrated chromogenic anti-Xa assays are not suitable to assess the presence of significant direct factor Xa inhibitors levels. *Thromb Res* 2017; 156: 36–8.

DOI: <https://doi.org/10.1016/j.pathol.2019.07.012>

ACTH measurements in Cushing's syndrome: the need for caution and communication



Sir,

Adrenocorticotrophic hormone (ACTH) concentration levels are key in determining the cause of Cushing's syndrome. However, the assay may be vulnerable to interference. This report discusses the case of a 40-year-old woman with cortisol excess and unilateral adrenal lesion but elevated ACTH concentration suggestive of ACTH-dependent Cushing's syndrome. Through close collaboration with the chemical pathology team, it was determined that assay interference was likely leading to a falsely elevated ACTH concentration. The patient underwent a successful unilateral adrenalectomy and avoided further unnecessary testing.

A 40-year-old Chinese woman was referred to the endocrinology outpatient clinic for investigation of an incidentally discovered left adrenal mass and elevated random serum cortisol. On questioning, she reported 2 years of lethargy, central adiposity, and easy bruising as well as amenorrhoea for the previous 3 months. She had no headaches or visual changes. She had suffered from low back pain. She had no relevant family history and took no prescribed or alternative medications. She was a life-long non-smoker and worked in an administrative role.

On clinical examination, her weight was 67 kg and she was normotensive (BP 120/80 mmHg). She had the classical Cushingoid appearance of moon facies, buffalo hump and central adiposity and when compared to photos from 5 years prior, the physical changes were marked. There was no significant abdominal striae or thin skin.

Repeat computed tomography (CT) of her adrenal glands demonstrated a stable left 31 × 24 mm adrenal adenoma with heterogeneous contrast enhancement (non-contrast 22 and post-contrast 126 Hounsfield units) unchanged compared to imaging 18 months ago. Functional testing for primary hyperaldosteronism and pheochromocytoma was negative.