



Research paper

A phase 2 trial of the oral smoothed inhibitor glasdegib in refractory myelodysplastic syndromes (MDS)



David A. Sallman^a, Rami S. Komrokji^a, Kendra L. Sweet^a, Qianxing Mo^b, Kathy L. McGraw^a, Vu H. Duong^c, Ling Zhang^d, Lisa Ann Nardelli^a, Eric Padron^a, Alan F. List^a, Jeffrey E. Lancet^{a,*}

^a Department of Malignant Hematology, Moffitt Cancer Center, Tampa, FL, USA

^b Department of Biostatistics and Bioinformatics, Moffitt Cancer Center, Tampa, FL, USA

^c Department of Medicine, University of Maryland School of Medicine, Baltimore, MD, USA

^d Department of Hematopathology and Laboratory Medicine, Moffitt Cancer Center, Tampa, FL, USA

ARTICLE INFO

Keywords:

MDS
Hedgehog
PF-04449913
Glasdegib
Hypomethylating agent failure

ABSTRACT

Hypomethylating agent (HMA) failure myelodysplastic syndrome (MDS) patients have poor outcomes and urgent need for novel therapies. Hedgehog pathway signaling upregulation plays a central role in myeloid neoplasm pathogenesis and leukemia stem cell survival. We evaluated the efficacy and safety of the smoothed inhibitor glasdegib in HMA-failure MDS (n = 35, median age 73 years). According to the International Prognostic Scoring System and the MD Anderson Global Risk Model, 54% and 77% had higher risk disease, respectively. Overall response was 6% (n = 2), and best response was marrow complete remission with hematologic improvement in both patients. Median OS and median follow-up were 10.4 and 42.8 months, respectively. Drug response/stable disease (SD) resulted in better OS than treatment failure (20.6 [95% CI, 10.4–] vs 3.9 months [95% CI, 0.7–9.1]; $P < .0001$). Response/SD was confirmed to be an independent covariate for improved OS ($P < .0001$). Grade 3 or higher infections occurred in 11% of patients (n = 4); non-hematologic toxicities were rare. Early mortality (< 30 days) occurred in 11% of patients (n = 4). Glasdegib was well tolerated among HMA-failure MDS patients, although single-agent activity was limited. SD or better resulted in notably superior OS. These results support further investigation of glasdegib, potentially in novel drug combinations, in MDS patients.

1. Introduction

Myelodysplastic syndromes (MDS) encompass a heterogeneous group of clonal neoplastic hematopoietic stem-cell diseases, which are characterized by ineffective hematopoiesis, bone marrow (BM) dysplasia, and risk of transformation to acute myeloid leukemia (AML). Treatment with the hypomethylating agents (HMA) azacitidine and decitabine are the current standard of care for high-risk MDS patients, and HMA failure in high-risk MDS patients results in a median overall survival (OS) of less than 6 months and a 2-year OS probability of 15% [1]. Furthermore, patients with low-risk MDS who experience HMA failure have a median OS of only 17 months [2]. The low OS rates among both low- and high-risk patients demonstrate the clear need for novel therapeutic approaches to the treatment of MDS patients with HMA failure.

The hedgehog (HH) signaling pathway plays a critical role in hematopoietic stem-cell survival, mediating cell self-renewal and resistance to chemotherapy [3]. In myeloid leukemia models, HH activation is essential for the maintenance of cancer stem cells as well as intrinsic chemoresistance [4–6]. To cue the HH pathway, HH ligands, including the sonic, desert, and Indian varieties, bind to the transmembrane receptor patched, which works to inhibit activity of the smoothed (SMO) protein. The binding of a ligand to patched leads to SMO derepression, resulting in SMO activation and HH signal transduction via zinc finger glioma-associated transcriptional regulators (e.g. *GLI1*, *GLI2*, and *GLI3*) [7–9]. *GLI1* and *GLI2* are activators that play an important role in AML pathogenesis. *GLI1* and *GLI2* expression is often a predictor of inferior event-free survival (EFS) and OS. *GLI2* expression is associated with *FLT3* mutational status in AML while *GLI3* acts as a strong repressor of HH signaling [10,11]. BM stromal cells of MDS

* Corresponding author at: Department of Malignant Hematology, 12902 Magnolia Drive, Tampa, FL 33612, USA.

E-mail addresses: David.Sallman@moffitt.org (D.A. Sallman), Rami.Komrokji@moffitt.org (R.S. Komrokji), Kendra.Sweet@moffitt.org (K.L. Sweet), Qianxing.Mo@moffitt.org (Q. Mo), Kathy.McGraw@moffitt.org (K.L. McGraw), vduong@umm.edu (V.H. Duong), Ling.Zhang@moffitt.org (L. Zhang), Lisa.Nardelli@moffitt.org (L.A. Nardelli), Eric.Padron@moffitt.org (E. Padron), Alan.List@moffitt.org (A.F. List), Jeffrey.Lancet@moffitt.org (J.E. Lancet).

<https://doi.org/10.1016/j.leukres.2019.03.008>

Received 19 February 2019; Received in revised form 26 March 2019; Accepted 28 March 2019

Available online 30 March 2019

0145-2126/© 2019 Published by Elsevier Ltd.

patients show HH ligand upregulation, as well as decreased expression of HH-interacting protein, a negative regulator of HH signaling [12–14]. Moreover, *GLI1* expression directly correlates with increased DNA methyltransferase 1 expression, with targeted knockdown of this pathway inducing cell-cycle arrest and apoptosis in MDS cell lines. [13]

Glasdegib (PF-04449913) is a potent orally bioavailable small-molecule inhibitor of SMO that was investigated in a phase 1 study that involved patients with myeloid malignancies, demonstrating clinical activity in 49% of patients (n = 23). Results included 1 AML patient with complete remission and incomplete hematological recovery and 2 MDS patients with hematologic improvement (HI) [15]. Glasdegib was well tolerated, with the most common treatment-related adverse events being dysgeusia, decreased appetite, and alopecia. Additionally, glasdegib in combination with low dose cytarabine has recently been approved by the FDA for the frontline treatment of AML patients who are ≥ 75 years of age or who are ineligible for intensive chemotherapy due to comorbidities. On the basis of these encouraging results, we designed a phase 2 study to evaluate the efficacy and safety of glasdegib among patients with HMA-refractory MDS or chronic myelomonocytic leukemia (CMML). The recommended dose based on the phase 1 trial was 200 mg or lower once daily.

2. Materials and methods

2.1. Patients

This was a single-center, open-label, 2-stage phase II study of glasdegib among patients who had MDS or CMML, according to World Health Organization (WHO) criteria, or AML with 20%–30% myeloblasts (refractory anemia with excess blasts in transformation), according to French-American-British criteria. Patients must have experienced refractory disease, progression, or relapse following prior HMA therapy. Patients with any risk score, as calculated by the International Prognostic Scoring System (IPSS), were eligible [16]. Inclusion criteria included age ≥ 18 years, an Eastern Cooperative Oncology Group performance status of 0–2, and adequate organ function (eg, serum creatinine ≤ 1.5 times the upper limit of normal [ULN], serum bilirubin < 1.5 times the ULN, and an aspartate aminotransferase/alanine aminotransferase < 2 times the ULN). Patients were excluded if they had a corrected QT interval ≥ 480 ms, a second malignancy requiring active therapy, or concurrent uncontrolled systemic illness.

2.2. Study design

All patients were given a daily oral dose of 100 mg of glasdegib over 4 consecutive weeks (days 1–28) for up to 4 cycles, with an optional continuation phase. There was no dose interruption between cycles. Dose escalation to 200 mg was allowed for patients who did not achieve at least HI following 2 cycles. Dose reduction to 50 mg was permitted for patients who experienced significant toxicity. Responses were assessed after patients completed cycles 2 and 4 (on day 1 of cycles 3 and 5), using International Working Group (IWG) 2006 criteria [17]. Low- (i.e. low or intermediate-1) and high-risk (i.e. intermediate-2 or high) patients were allowed to continue treatment after 4 cycles if they achieved HI or stable disease (SD). SD was defined as failure to achieve response with no evidence of disease progression after > 8 weeks. After completion of study therapy, patients were followed-up monthly for survival. This study was conducted in accordance with the Declaration of Helsinki and was approved by the Moffitt Cancer Center Scientific Review Committee and Institutional Review Board. This trial is registered with ClinicalTrials.gov (NCT01842646).

2.3. Study objectives

The primary objective was to estimate the overall response rate to

glasdegib, as defined by IWG 2006 criteria (i.e. complete remission (CR), marrow CR, partial remission, and/or HI), among patients with MDS and CMML after HMA failure [17]. Secondary objectives included ascertaining duration of response, OS, EFS, safety, and biomarker evaluation of response based on expression of HH targets. Duration of response was measured from the time of initial response until the date of disease progression. OS was defined as the period from the therapy start date until date of death. EFS was defined as the period from the therapy start date until the date of progression or death, whichever occurred first.

2.4. Correlative studies

To assess the relationship between response and expression of HH pathway targets, BM mononuclear cells were isolated using the Ficoll-Paque method. Total RNA was isolated using the RNeasy kit (Qiagen, Germantown, MD). Complementary DNA was synthesized using the High-Capacity cDNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA). Real-time quantitative polymerase chain reaction analysis for each gene was performed using Taqman Gene Expression Arrays (*SMO*, Hs01090242_m1, *PTCH1*, Hs00181117_m1, *GLI1*, Hs00171790_m1; *CCND1*, Hs00765553_m1, *c-MYC*; Hs00153408; Applied Biosystems) and normalized to *GAPDH* (Hs99999905_m1) using the ABI PRISM 7900 HT Sequence Detection System (Applied Biosystems). Relative gene expression was calculated using the $2^{-\Delta\Delta Ct}$ method. Immunohistochemically (IHC) stained paraffin-embedded BM trephine biopsies and clot sections were evaluated for expression of HH targets before and glasdegib treatment, as previously described, using diaminobenzidine immunoperoxidase staining on paraffin sections (EnVision + System-HRP (DAB + substrate); Dako, Santa Clara, CA). Primary antibodies to sonic/Indian HH (IgG rabbit polyclonal; N-19; Santa Cruz Biotechnology, Dallas, TX) at a ratio of 1:50, desert HH (IgG goat polyclonal; H-85; Santa Cruz Biotechnology) at a ratio of 1:250, GLI1 (N-16; Santa Cruz Biotechnology) at a ratio of 1:100, and SMO at a ratio of 1:50 (H-300; Santa Cruz Biotechnology) were incubated overnight with mouse monoclonal BMI1 antibody (Abcam Bmi1 antibody [1.T.21], Cambridge, MA), followed by a 30-minute incubation time with secondary antibodies (EnVision + System-HRP [DAB + substrate]; Dako). Immunodetection was performed with 3,3'-diaminobenzidine (EnVision + System-HRP (DAB + substrate); Dako). Staining intensity was scored by grade category (grade 0 = $< 10\%$; 1 = 10%–20%; 2 = 20%–50%, and 3 = $> 50\%$ positive cells).

2.5. Statistics

A Simon's 2-stage design was used for this phase 2 trial and had an 87% power to evaluate the overall response rate (CR + partial remission + marrow CR + HI), with a null hypothesis of 10% or less versus the alternative hypothesis of an overall response rate of 30% or more. Twenty patients were to be enrolled in the first stage. If ≥ 2 patients achieved a response, the study could proceed to the second stage, at which an additional 15 patients could be enrolled. All time-to-event variables, including OS, EFS, and time to transformation to AML, were analyzed using the Kaplan-Meier method and log-rank test. Toxicity was graded according to Common Terminology Criteria for Adverse Events version 4.0, and data were reported as frequencies and percentages. Exploratory analyses were performed comparing responders to non-responders. Continuous and categorical variables were analyzed using the Wilcoxon Rank Sum and Kruskal-Wallis tests, respectively. All tests were 2 sided and considered statistically significant if $P < .05$. Multivariate Cox regression models were created to adjust for clinical and treatment characteristics. All analyses were performed using Statistical Analysis System 9.4 or R 3.5.1.

Table 1
Baseline patient characteristics of the study cohort.

Characteristic	Patients (N = 35)
Sex, no. (%)	
Male	24 (69)
Female	11 (31)
Age, y	
Median	73
Range	55–88
WHO subtype, no. (%)	
RCMD	9 (26)
RAEB-1	6 (17)
RAEB-2	11 (31)
CMML	5 (16)
AML	4 (13)
Cytopenias	
Grade 2 or 3, no. (%)	27 (77)
ANC, mean (range), $\times 10^9/L$	0.94 (0.06–7.65)
Hemoglobin, mean (range), g/dL	9.3 (7.3–13.1)
Platelets, mean, (range), $\times 10^9/L$	46 (9–270)
IPSS cytogenetic score, no. (%)	
Good	13 (39)
Intermediate	9 (27)
Poor	11 (33)
IPSS Category, no. (%)	
Low/INT-1	15 (43)
INT-2/High	19 (54)
Unknown	1 (3)
MD Anderson Risk Category, no. (%)	
Low/INT-1	7 (20)
INT-2/High	27 (77)
Unknown	1 (3)
Baseline bone marrow blast percentage, %	
Median	5
Range	0.2–23
Prior HMA therapy	
Azacitidine, no. (%)	34 (97)
Median no. of cycles/patient	9
Decitabine, no. (%)	6 (17)
Median no. of cycles/patient	4

Abbreviations: AML, acute myeloid leukemia; ANC, absolute neutrophil count; CMML, chronic myelomonocytic leukemia; HMA, hypomethylating agent; INT, intermediate; IPSS, international prognostic scoring system; RAEB, refractory anemia with excess blasts; RCMD, refractory cytopenia with multilineage dysplasia; WHO, World Health Organization.

3. Results

3.1. Patient characteristics

From August 29, 2013, to September 2, 2015, 35 patients were accrued to the study. Patient characteristics are summarized in Table 1. The median age of the cohort was 73 years and predominantly male (69%). By WHO category, 74% of patients had MDS ($n = 26$), 16% had CMML ($n = 5$), and 13% ($n = 4$) had AML (20%–30% blasts). Of the MDS patients, 65% ($n = 17$) had excess blasts. The median BM blast percentage was 5%, with 33% of patients having poor-risk cytogenetics, according to their IPSS. By IPSS, 54% ($n = 19$) and 43% ($n = 15$) had high- and low-risk disease, respectively. According to the MD Anderson Global Risk Model, [18] 77% of patients had high-risk disease ($n = 27$). Ninety-seven percent of the cohort had received prior azacitidine, with 17% of patients having received prior decitabine. The median numbers of prior cycles of azacitidine and decitabine were 9 and 4, respectively.

3.2. Response to therapy and survival

Thirty-four of the 35 patients were evaluable for response (Table 2). One patient, who received only 1 dose of glasdegib immediately before being admitted to the hospital with cellulitis, declined further therapy, died 1 week later, and was subsequently deemed non-evaluable for

Table 2
Overall Response Rates for Evaluable Patients^a.

Response	Patients, no. (%) (n = 34)
Complete response/CRi	0 (0)
Hematologic improvement ^b	2 (6)
Stable disease	19 (56)
Failure	13 (38)

Abbreviations: CRi, complete response with incomplete blood count recovery.

^a Median no. of cycles received = 3 (range, 0–11). ^bBoth patients also achieved marrow complete remission.

response. The overall response rate was 6% ($n = 2$) with best overall response of marrow CR in 2 patients, both of whom were MDS patients with excess blasts. Additionally, both responding patients had HI and neutrophil response; one had platelet response. Both patients responded at the 100 mg dose level. The median time to response was 81 days (range, 28–134 days). SD was observed in 56% of patients ($n = 19$). Thirty-eight percent ($n = 13$) of patients had treatment failure, including 10 patients with progressive disease and 3 patients who died during study treatment secondary to disease/infection, before their responses could be assessed. The median number of cycles received was 3 (range, 0–11 cycles). One responding patient received 7 cycles of therapy and had a durable response that lasted an additional 9 months, with transfusion independence after drug discontinuation. The other responding patient received 4 cycles of therapy and electively discontinued, without evidence of disease progression. The median overall survival (OS) of the cohort was 10.4 months, with a median follow-up of 42.8 months (range, 0.3–47.7 months; Fig. 1A). The median EFS of the entire cohort was 6.4 months. Based on IPSS category, patients with low-risk disease (i.e. low/intermediate-1) had greater OS than patients with high-risk disease (i.e. intermediate-2/high; median OS 15.0 vs 7.7 months; $P = .03$; Fig. 1B). Patients who achieved drug response/SD had significantly better OS than patients who experienced treatment failure, which was defined as progressive disease or death during treatment (median OS 20.6 vs 3.9 months, respectively; $P < .0001$; Fig. 1C). In the cohort of patients with high-risk disease according to IPSS ($n = 19$), OS was also significantly better among patients who responded/had SD than among those who had treatment failure (median OS 19.5 months vs 3.6 months, respectively; $P = .004$; Fig. 1D). Response/SD was confirmed as an independent covariate for improved OS in Cox regression modeling that included age, gender, WHO, and IPSS risk category (HR 0.13; 95% CI, 0.05–0.34; $P < .0001$).

3.3. Safety

Treatment-emergent grades 1/2 adverse events (AEs) that occurred in $\geq 10\%$ of patients are shown in Table 3. The most common grade 1/2 AEs included pain ($n = 22$ [63%]), dysgeusia ($n = 21$ [60%]), nausea ($n = 11$ [31%]), and anorexia ($n = 11$ [31%]). Other than infections ($n = 4$), grade 3 and higher non-hematologic toxicities were rare and included muscle pain ($n = 1$), rash ($n = 1$), and dyspnea ($n = 1$), of which only muscle pain was attributed to the study drug. Fourteen percent of patients ($n = 5$) had dose reductions to 50 mg because of toxicities after a median of 3 cycles (range, 2–4 cycles). Additionally, 3 patients (9%) had their dose escalated to 200 mg orally daily. Four patients (11%) died within 30 days of treatment initiation.

3.4. HH pathway correlatives

There were no response associated differences in baseline expression levels of HH pathway components or downstream signaling

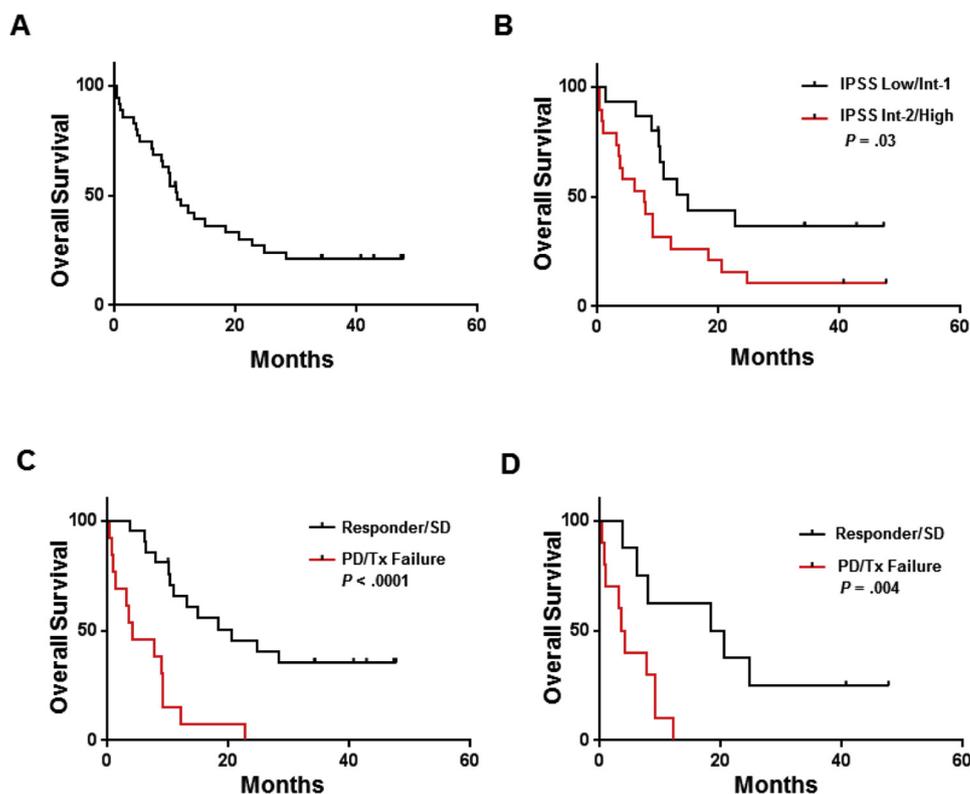


Fig. 1. Overall survival of patients treated with glasdegib. Kaplan-Meier curves of OS (A) in the entire cohort, (B) stratified by IPSS classification, and stratified by response (ie, responder/stable disease versus progressive disease/treatment failure) in (C) the overall cohort and (D) the high-risk cohort. Abbreviations: INT, intermediate; IPSS, international prognostic scoring system; PD, progressive disease; SD, stable disease; Tx, treatment.

Table 3
Treatment-emergent adverse events for all patients treated with glasdegib.

Toxicity	Grades 1-2, no. (%)	Grades 3-5, no. (%)	Total, no. (%)
Pain	22 (63)	1 (3)	23 (66)
Dysgeusia	21 (60)		21 (60)
Nausea	11 (31)		11 (31)
Anorexia	11 (31)		11 (31)
Skin/rash	8 (23)	1 (3)	9 (26)
Oral mucositis	9 (26)		9 (26)
AST elevated	8 (23)		8 (23)
Alopecia	8 (23)		8 (23)
ALT elevated	6 (17)		6 (17)
Dizziness	6 (17)		6 (17)
Dyspnea	5 (14)	1 (3)	6 (17)
Infections	2 (6)	4 (11)	6 (17)
Fatigue	6 (17)		6 (17)
Diarrhea	6 (17)		6 (17)
Constipation	5 (14)		5 (14)
Fever	5 (14)		5 (14)
Hyperkalemia	5 (14)		5 (14)
Hyperglycemia	4 (11)		4 (11)
Thrombocytopenia	2 (6)	2 (6)	4 (11)
Headache	4 (11)		4 (11)

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase.

intermediates that could be observed via messenger RNA (mRNA) expression analysis or IHC (Supplementary Table 1). However, in comparison to baseline expression of HH pathway targets, patients with progressive disease after cycle 2 had a trend for increased *PTCH* expression ($P = .13$; Fig. 2A), with significantly increased mRNA expression of *GLI1* as well as downstream signaling components, including *MYC* and *Cyclin D1* (Fig. 2B-D). However, there was no difference in protein expression on serial assessment by IHC (Supplementary Table 1).

4. Discussion

In a cohort of HMA-refractory MDS, AML, and CMML patients, we demonstrated that glasdegib is safe and well tolerated. The most common treatment-emergent AEs were pain/muscle spasms, dysgeusia, nausea, and anorexia. Treatment-emergent AEs were similar to those observed in previous studies examining this patient population, with the exception of an increased risk of rash, oral mucositis, and transaminitis, which were all grades 1/2, with the exception a grade 3 rash that was experienced by one patient but considered to be unrelated to the study drug [15,19]. Dose reductions to 50 mg daily were uncommon, and the 3 patients whose dose was escalated to 200 mg daily tolerated it well.

There is increasing awareness of the dismal OS, typically less than 6 months, that HMA-refractory, high-risk MDS carries [1]. A recent phase 2 trial (NCT01546038) of glasdegib that compared low-dose cytarabine combination treatment with cytarabine alone showed an increased CR rate (15%) for the combination treatment but, more impressively, a significantly greater OS that was independent of cytogenetic risk [19]. Similarly, the combination of the SMO inhibitor sonidegib with azacitidine has been evaluated in a phase 1/1b study (NCT02129101) that resulted in an SD rate of 76% in relapsed/refractory AML patients, most of whom were HMA refractory [20]. In the current trial of heavily pretreated, primarily high-risk MDS patients, objective response rates were low, with only 6% of patients ($n = 2$) achieving response by IWG criteria; however, 56% of patients had SD. Notably, patients who had SD or better after treatment had a median OS of 20.6 months, which was significantly longer than patients with treatment failure, independent of other clinical variables. Greater OS was also observed in the high-risk cohort of patients who achieved SD/response. Importantly, achievement of SD or better was an independent covariate for improved OS.

Elucidation of potential response biomarkers and evaluation of synergism with additional agents will be critical to the advancement of HH-pathway inhibitors in myeloid malignancies. In this regard, a recent study identified *GLI3* repressor (GLI3R) expression to be paramount to

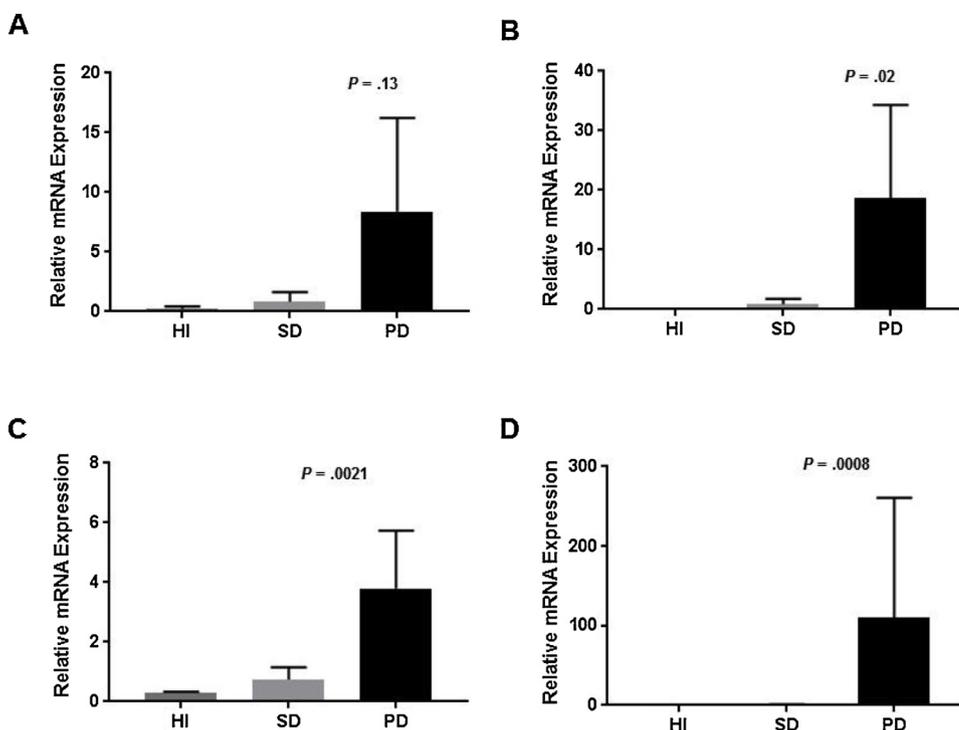


Fig. 2. Serial mRNA expression of Hedgehog pathway components. The relative gene expression of (A) *PTCH*, (B) *GLI1*, (C) *MYC*, and (D) *CCND1* were shown by serial assessment of patients with hematologic improvement, stable disease, and progressive disease. This is a two-column image. Abbreviations: HI, hematologic improvement; PD, progressive disease; SD, stable disease.

SMO inhibitor activity [11]. Specifically, *GLI3R* is required for SMO inhibitor response, which is epigenetically silenced in most of the cases in which low response rates have correlated to treatment with glasdegib as a single agent. Supporting this hypothesis, combination with the HMA decitabine restored *GLI3* expression leading to SMO sensitivity [11]. However, increased response rates of sonidegib with azacitidine were not observed in the aforementioned clinical trial NCT02129101 (CR rate of 23% in frontline setting) [20]. Further evaluation of *GLI3R* expression as a biomarker of response in glasdegib-treated MDS patients would be of interest for future study.

Although we did not observe differential gene expression of HH pathway targets at baseline, patients with progressive disease had significant upregulation of downstream HH pathway targets on serial assessment, suggesting a potential resistance mechanism via SMO independent activation of *GLI*. Specifically, the HH pathway has significant crosstalk with other signaling pathways, including the Ras/Raf/MEK/ERK, PI3K/AKT/mTOR, epidermal growth factor receptor, and transforming growth factor beta pathways [21–23]. Furthermore, combination of SMO inhibitor with *FLT3* inhibitors was found to be efficacious in both *in vitro* and *in vivo* AML models [24]. As these other pathways are actionable with approved therapies, potential combination with glasdegib should be evaluated in future studies.

In conclusion, glasdegib in HMA-refractory MDS and CMML patients was well tolerated with limited single-agent activity. The evidence of prolonged survival in patients who achieved SD was encouraging. As combination studies of glasdegib with low-dose cytarabine have highlighted, improved OS in treatment-naïve patients with MDS/AML is possible, [19] and future investigations should focus on identifying optimal glasdegib combination treatments for patients with myeloid malignancies.

Funding

This work has been supported in part by the Biostatistics Core Facility at the H. Lee Moffitt Cancer Center & Research Institute, an NCI-designated Comprehensive Cancer Center (P30-CA076292).

Conflicts of interest

The authors declare no conflict of interest.

Acknowledgements

We thank Paul Fletcher and Daley Drucker (Moffitt Cancer Center) for editorial support. They were not compensated beyond their regular salaries.

References

- [1] T. Prebet, S.D. Gore, B. Esterni, C. Gardin, R. Itzykson, S. Thepot, et al., Outcome of high-risk myelodysplastic syndrome after azacitidine treatment failure, *J. Clin. Oncol.* 29 (24) (2011) 3322–3327, <https://doi.org/10.1200/jco.2011.35.8135>.
- [2] E.J. Jabbour, G. Garcia-Manero, P. Strati, A. Mishra, N.H. Al Ali, E. Padron, et al., Outcome of patients with low-risk and intermediate-1-risk myelodysplastic syndrome after hypomethylating agent failure: a report on behalf of the MDS Clinical Research Consortium, *Cancer* 121 (6) (2015) 876–882, <https://doi.org/10.1002/cncr.29145>.
- [3] N. Takebe, P.J. Harris, R.Q. Warren, S.P. Ivy, Targeting cancer stem cells by inhibiting Wnt, Notch, and Hedgehog pathways, *Nat. Rev. Clin. Oncol.* 8 (2) (2011) 97–106, <https://doi.org/10.1038/nrclinonc.2010.196>.
- [4] C. Zhao, A. Chen, C.H. Jamieson, M. Fereshteh, A. Abrahamsson, J. Blum, et al., Hedgehog signalling is essential for maintenance of cancer stem cells in myeloid leukaemia, *Nature* 458 (7239) (2009) 776–779, <https://doi.org/10.1038/nature07737>.
- [5] C. Dierks, R. Beigi, G.R. Guo, K. Zirikli, M.R. Stegert, P. Manley, et al., Expansion of Bcr-Abl-positive leukemic stem cells is dependent on Hedgehog pathway activation, *Cancer Cell* 14 (3) (2008) 238–249, <https://doi.org/10.1016/j.ccr.2008.08.003>.
- [6] K.C. Queiroz, R.R. Ruela-de-Sousa, G.M. Fuhler, H.L. Abernson, C.V. Ferreira, M.P. Peppelenbosch, et al., Hedgehog signaling maintains chemoresistance in myeloid leukemic cells, *Oncogene* 29 (48) (2010) 6314–6322, <https://doi.org/10.1038/ncr.2010.375>.
- [7] J. Taipale, P.A. Beachy, The Hedgehog and Wnt signalling pathways in cancer, *Nature* 411 (6835) (2001) 349–354, <https://doi.org/10.1038/35077219>.
- [8] D.A. Irvine, M. Copland, Targeting hedgehog in hematologic malignancy, *Blood* 119 (10) (2012) 2196–2204, <https://doi.org/10.1182/blood-2011-10-383752>.
- [9] D. Kalderon, Transducing the hedgehog signal, *Cell* 103 (3) (2000) 371–374.
- [10] J. Wellbrock, E. Latuske, J. Kohler, K. Wagner, H. Stamm, E. Vettorazzi, et al., Expression of hedgehog pathway mediator *GLI1* represents a negative prognostic marker in human acute myeloid leukemia and its inhibition exerts antileukemic effects, *Clin. Cancer Res.* 21 (10) (2015) 2388–2398, <https://doi.org/10.1158/1078-0432.CCR-14-1059>.
- [11] P. Chaudhry, M. Singh, T.J. Triche, M. Guzman, A.A. Merchant, *GLI3* repressor determines Hedgehog pathway activation and is required for response to SMO

- antagonist glasdegib in AML, *Blood* 129 (26) (2017) 3465–3475, <https://doi.org/10.1182/blood-2016-05-718585>.
- [12] J. Zou, Y. Hong, Y. Tong, J. Wei, Y. Qin, S. Shao, et al., Sonic hedgehog produced by bone marrow-derived mesenchymal stromal cells supports cell survival in myelodysplastic syndrome, *Stem Cells Int.* 2015 (2015) 957502, <https://doi.org/10.1155/2015/957502>.
- [13] J. Zou, Z. Zhou, L. Wan, Y. Tong, Y. Qin, C. Wang, et al., Targeting the sonic Hedgehog-Gli1 pathway as a potential new therapeutic strategy for myelodysplastic syndromes, *PLoS One* 10 (8) (2015) e0136843, <https://doi.org/10.1371/journal.pone.0136843>.
- [14] M. Kobune, S. Iyama, S. Kikuchi, H. Horiguchi, T. Sato, K. Murase, et al., Stromal cells expressing hedgehog-interacting protein regulate the proliferation of myeloid neoplasms, *Blood Cancer J.* 2 (2012) e87, <https://doi.org/10.1038/bcj.2012.36>.
- [15] G. Martinelli, V.G. Oehler, C. Papayannidis, R. Courtney, M.N. Shaik, X. Zhang, et al., Treatment with PF-04449913, an oral smoothened antagonist, in patients with myeloid malignancies: a phase 1 safety and pharmacokinetics study, *Lancet Haematol.* 2 (8) (2015), [https://doi.org/10.1016/S2352-3026\(15\)00096-4](https://doi.org/10.1016/S2352-3026(15)00096-4) e339-346.
- [16] P. Greenberg, C. Cox, M.M. LeBeau, P. Fenaux, P. Morel, G. Sanz, et al., International scoring system for evaluating prognosis in myelodysplastic syndromes, *Blood* 89 (6) (1997) 2079–2088.
- [17] B.D. Cheson, P.L. Greenberg, J.M. Bennett, B. Lowenberg, P.W. Wijermans, S.D. Nimer, et al., Clinical application and proposal for modification of the International Working Group (IWG) response criteria in myelodysplasia, *Blood* 108 (2) (2006) 419–425, <https://doi.org/10.1182/blood-2005-10-4149>.
- [18] H. Kantarjian, S. O'Brien, F. Ravandi, J. Cortes, J. Shan, J.M. Bennett, et al., Proposal for a new risk model in myelodysplastic syndrome that accounts for events not considered in the original International Prognostic Scoring System, *Cancer* 113 (6) (2008) 1351–1361, <https://doi.org/10.1002/cncr.23697>.
- [19] J.E. Cortes, F.H. Heidel, M. Heuser, W. Fiedler, B.D. Smith, T. Robak, et al., A phase 2 randomized study of low dose Ara-C with or without glasdegib (PF-04449913) in untreated patients with acute myeloid leukemia or high-risk myelodysplastic syndrome, *Blood* 128 (22) (2016) 99.
- [20] R. Tibes, H.E. Kosiorek, A. Dueck, J. Palmer, J.L. Slack, E.A. Knight, et al., Phase 1/IB study of Azacitidine and hedgehog pathway inhibition with sonidegib (LDE225) in myeloid malignancies, *Blood* 130 (Suppl 1) (2017) 2629.
- [21] J. Brechbiel, K. Miller-Moslin, A.A. Adjei, Crosstalk between hedgehog and other signaling pathways as a basis for combination therapies in cancer, *Cancer Treat. Rev.* 40 (6) (2014) 750–759, <https://doi.org/10.1016/j.ctrv.2014.02.003>.
- [22] B. Stecca, C. Mas, V. Clement, M. Zbinden, R. Correa, V. Piguet, et al., Melanomas require HEDGEHOG-GLI signaling regulated by interactions between GLI1 and the RAS-MEK/AKT pathways, *Proc. Natl. Acad. Sci. U. S. A* 104 (14) (2007) 5895–5900, <https://doi.org/10.1073/pnas.0700776104>.
- [23] A. Parascandolo, M.O. Laukkanen, N. De Rosa, C. Ugolini, M.C. Cantisani, A.M. Cirafici, et al., A dual mechanism of activation of the Sonic Hedgehog pathway in anaplastic thyroid cancer: crosstalk with RAS-BRAF-MEK pathway and ligand secretion by tumor stroma, *Oncotarget* 9 (4) (2018) doi:10.1016/23388.4496-5.
- [24] Y. Lim, L. Gondek, L. Li, Q. Wang, H. Ma, E. Chang, et al., Integration of Hedgehog and mutant FLT3 signaling in myeloid leukemia, *Sci. Transl. Med.* 7 (291) (2015) 291ra96, <https://doi.org/10.1126/scitranslmed.aaa5731>.