

WHAT'S NEW IN INTENSIVE CARE



Fecal microbiota transplantation in the ICU: perspectives on future implementations

Laura Alagna¹, Bastiaan W. Haak² and Andrea Gori^{1,3*}

© 2019 Springer-Verlag GmbH Germany, part of Springer Nature

Introduction

An increasing amount of evidence has shown that both the intestinal ecosystem, as well as the microbiota that inhabits these niches, can change dramatically during the course of an ICU stay [1–3]. Yet, few data exist on the consequences of these changes on patients' clinical outcome. Recent preclinical studies suggest that severe disruption of the microbiota, otherwise described as dysbiosis, could be associated with a profound state of immunosuppression, as well as an increased risk of organ failure and mortality [4–6]. In addition, it is thought that disruption of beneficial obligate anaerobes in the gut opens the door to colonization with multi-drug-resistant (MDR) pathogens, such as vancomycin-resistant *Enterococcus* (VRE), and extended-spectrum β -lactamase-producing *Enterobacteriaceae* (ESBL-E) [7, 8]. Therefore, mitigating dysbiosis in the ICU is a novel field to be explored for its eventual impact on outcome for critically ill patient. Faecal microbiota transplantation (FMT) consists of the infusion of faeces from a healthy donor into the gastrointestinal tract of a patient, aimed at reconstituting the intestinal microbiota and treat disorders associated with dysbiosis. While evidence-based consensus exists that the treatment is extremely effective against recurrent *Clostridium difficile* infection (CDI), the use of FMT for this indication on the ICU remains controversial [9, 10]. A recent retrospective study of 111 patients in France showed that early FMT significantly improved survival in patients with severe CDI, thereby opening the door for further evaluation of FMT in ICU settings [11]. Beyond CDI, studies investigating the role in FMT

to improve outcomes of critical illness are largely unexplored. However, we do foresee a potential role for this treatment approach in the restoration of ICU-associated dysbiosis, as well as a future tool to decolonize the gut of MDR organisms.

Only five cases have been described in which FMT has been employed to address disruption of the microbiota in the ICU. All these cases showed that treatment with FMT led to a successful reversal of dysbiosis, with subsequent improvement of outcome. In addition, some cases marked a steep reduction in inflammatory mediators, and normalized Th1/Th2 and Th1/Th17 ratios following FMT. (Supplementary Table 1). Apart from difficulties with extrapolating the data derived from these case reports to the general ICU population, we are far from obtaining conclusive evidence that restoration of dysbiosis in critical illness is beneficial. For example, patients in the relevant studies showed signs of recovery prior to the initiation of FMT, which implies that these patients could have recovered without receiving this experimental therapy. The use of FMT to decolonize the gut with MDR organisms has been studied in several trials in non-ICU settings. (Supplementary Table 2a). The results of the largest and most strictly controlled study did not support the routine use of FMT for purposes; instead, it indicated that larger trials with improved faecal preparation protocols are still warranted [12]. Of interest, other studies examining this procedure are currently ongoing, with one phase 2 open label trial focusing on eradicating MDR bacteria in 10 ICU patients (NCT03350178) (Supplementary Table 2b).

*Correspondence: andrea.gori@unimi.it

¹ Infectious Diseases Unit, Department of Internal Medicine, Fondazione IRCCS Ca' Granda, Ospedale Maggiore Policlinico, Milan, Italy
Full author information is available at the end of the article

Practical considerations for implementing FMT in the ICU

Apart from the aforementioned trial, no other studies are currently investigating the safety and feasibility of the use of FMT in the ICU. It is understandable that researcher clinicians have so far refrained from engaging on this subject, as the potential impact of administering live bacteria to a critically ill and potentially immunosuppressed patient has been described as potentially perilous [5]. In addition, numerous practical and technical challenges exist, such as heterogeneity in the selection of patients and donors, modes of faeces preparation, route and type of administration [8, 9, Table 1]. Selection of patients for a potential FMT trial is particularly cumbersome in the ICU, as the majority of critically ill patients receive antibiotics at some point during their ICU stay, potentially nullifying the proposed effect of this specific therapeutic approach. Therefore, investigators are faced with the dilemma to either account for this intervention in initial study design and, therefore, investigate a very narrow subset of ICU patients, or make a daunting commitment to stop any antibiotics during the course of the study, which many clinicians would not be comfortable undertaking.

In addition to the issues with patient selection, appropriate donor screening is equally cumbersome, as transmission of potentially transmittable diseases could be detrimental to an already vulnerable critically ill host [8]. In addition, studies treating inflammatory bowel disorders have shown that failure of FMT can sometimes be attributed to reduced immunomodulatory properties of the donor faeces, leading to the suggestion that donor faeces needs to contain a certain amount of

Take home message

FMT has the potential to recover dysbiosis associated with critical illness. Future studies should identify the target population, optimal timing, mode of administration and the ultimate effect on patients' outcome in the ICU setting.

butyrate-producing anaerobic bacteria, such as *Faecalibacterium prausnitzii*, *Roseburia hominis*, and *Coprococcus eutactus*, in order to be effective [13]. Finally, consensus regarding the best route of administration of FMT is currently lacking. Recent evidence states that critically ill recipients who received FMT via a nasogastric tube experienced higher rates of serious adverse events, such as fever and aspiration pneumonia, compared to administration via enema or colonoscopy [8]. Potential new administration routes for FMT, such as the use of faecal capsules, lyophilized stool, or faecal filtrate transfer (FFT) could circumvent the current challenges of administration with potentially fewer side effects; however, evidence regarding their efficacy has yet to be established [14, 15]. Table 1 depicts general considerations for FMT in critically ill patients, which, if addressed in future studies, could help the assessment of safety, feasibility and effectiveness of FMT in the ICU setting.

In conclusion, FMT is an interesting but importantly understudied approach that could potentially be effective in improving morbidity and mortality in certain categories of ICU patients. Clinical trials that systematically address the aforementioned issues are needed to provide more insight in the potential safety, feasibility and benefit of this treatment approach in critical illness.

Table 1 Practical considerations for the adoption of FMT in ICU patients. Adapted from Cammarota and colleagues [9]

Facility implemen- tations		Quality of evidence	Strength of recommen- dation
Clinical requirements and facilities	Referral FMT centres should be implemented in hospitals with appropriate expertise and facilities	Moderate	Strong
	A clinical governance dealing with administrative issues of FMT (e.g., reimbursements, authorisations) is recommended	Low	Strong
	Specific training on FMT processes are encouraged: clinical training (donor and patient selection, patient management after FMT), delivery training (learning of different routes of delivery) and microbiological training (preparation of fresh and frozen faecal material)	Low	Strong
	The availability of the endoscopy service is required. A multidisciplinary team (including gastroenterologists, microbiologists and infectious disease physicians) is encouraged	Low	Strong
Microbiological requirements and facilities	Safe processing of human samples (biosafety level 2: aliquoting, storage and preparation of faeces) is required. Stool banking is encouraged	Low	Strong
	Storage of FMT procedure and donor screening for at least 10 years is required	Low	Strong
Regulatory require- ments	Appropriate FMT registries should be implemented, in order to collect data concerning indications, procedure, effectiveness and safety profiles	Low	Strong
	Specific national rules for the classification of FMT should be followed to implement an FMT centre	Low	Strong
Pre-procedure			
Donor selection	Potential donors for FMT have to undergo a medical interview: i) at the beginning of the selection process, and ii) on the same day of the donation. Key issues in the collection of medical history include: risk factors of transmittable diseases, travel history, gastrointestinal/metabolic/neurological disorders, exposure to drugs that can impair gut microbiota composition	Low	Strong
	Potential donors for FMT should undergo blood and stool testing for transmittable diseases at most 4 weeks before donation	Low	Strong
Preparation of fae- cal material	Fresh stool should be used within 6 h after defecation	Moderate	Strong
	To protect anaerobic bacteria, the storage and preparation should be as brief as possible	Moderate	Strong
	Anaerobic storage and processing should be applied if possible		
	Until further processing, the stool sample can be stored at ambient temperature (20–30 °C)		
	A minimum amount of 30 g of faeces should be used		
	Faecal material should be suspended in saline using a blender or manual effort and sieved in order to avoid the clogging of infusion syringes and tubes		
	A dedicated space, disinfected using measures that are effective against sporulating bacteria, should be used		
	Frozen faecal material preparation has to undergo a minimum set of general steps		
	At least 30 g of donor faeces and 150 mL of saline solution should be used		
	Before freezing, glycerol should be added up to a final concentration of 10%		
The final suspension should be clearly labelled and traceable, and stored at – 80 °C			
On the day of faecal infusion, faecal suspension should be thawed in a warm (37 °C) water bath and infused within 6 h from thawing			
After thawing, saline solution can be added to obtain a desired suspension volume			
Repetitive thawing and freezing should be avoided			
During and post-procedure			
Faecal delivery	Recipients should be prepared with bowel lavage by polyethylene glycol before procedure when FMT is performed by upper route or by colonoscopy. For delivery via enema, no preparation instructions have been developed	Low	Weak
	Patients should be able to hold the infused material for at least 30 min. Some study protocols implicate repeated enemas to increase clinical success	Low	Strong
	FMT appears to be safe in immunocompromised and critically ill patients regardless the route of delivery. In case of critically ill patients, faecal infusion by enema(s) should be preferred. the rate of serious adverse events in recipients who received FMT via upper GI tract seems to be higher (aspiration pneumonia due to nausea and vomiting by nasogastric or nasoduodenal tube)		
Clinical manage- ment	Short-term monitoring of patients for adverse events (AEs). The most commonly described AEs after FMT for CDI are: diarrhoea, abdominal cramps, belching, constipation, fever, Gram-negative bacteraemia and perforation	Low	Weak
	Long-term monitoring of patients for AEs. Periodicity and length of follow-up for long-term AEs are not determined. Follow-up should include both clinical and analytical data	Low	Weak

Electronic supplementary material

The online version of this article (<https://doi.org/10.1007/s00134-019-05645-7>) contains supplementary material, which is available to authorized users.

Author details

¹ Infectious Diseases Unit, Department of Internal Medicine, Fondazione IRCCS Ca' Granda, Ospedale Maggiore Policlinico, Milan, Italy. ² Center for Experimental and Molecular Medicine, Amsterdam UMC, Amsterdam Infection and Immunity Institute, University of Amsterdam, Amsterdam, The Netherlands. ³ Department of Pathophysiology and Transplantation, School of Medicine and Surgery, University of Milan, Milan, Italy.

Acknowledgements

We thank Professor Alessandra Bandera (University of Milan) for the insightful discussions during the course of writing this manuscript.

Compliance with ethical standards

Conflicts of interest

LA has nothing to disclose. BWH has nothing to disclose. AG discloses the receipt of grants/research supports; receipt of honoraria or consultation fees; participation in a company sponsored speaker's bureau; travel grant/supports: Abbvie, Astellas, BMS, Boehringer, Gilead, Janssen, MSD, Novartis, Pfizer, Roche, Viiv.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 14 February 2019 Accepted: 8 May 2019

Published online: 24 May 2019

References

- Zaborin A, Smith D, Garfield K et al (2014) Membership and behavior of ultra-low-diversity pathogen communities present in the gut of humans during prolonged critical illness. *MBio* 5:e01361-14
- McDonald D, Ackermann G, Khailova L et al (2016) Extreme dysbiosis of the microbiome in critical illness. *mSphere*. <https://doi.org/10.1128/mSphere.00199-16>
- Lankelma JM, van Vught LA, Belzer C, Schultz MJ, van der Poll T, de Vos WM, Wiersinga WJ (2017) Critically ill patients demonstrate large inter-personal variation in intestinal microbiota dysregulation: a pilot study. *Intensive Care Med* 43(1):59–68
- Krezalek MA, DeFazio J, Zaborina O, Zaborin A, Alverdy JC (2016) The shift of an intestinal "microbiome" to a "pathobiome" governs the course and outcome of sepsis following surgical injury. *Shock* 45:475–482
- Klingensmith NJ, Coopersmith CM (2016) Fecal microbiota transplantation for multiple organ dysfunction syndrome. *Crit Care* 20:398
- Shimizu K, Ogura H, Hamasaki T, Goto M, Tasaki O, Asahara T, Nomoto K, Morotomi M, Matsushima A, Kuwagata Y, Sugimoto H (2011) Altered gut flora are associated with septic complications and death in critically ill patients with systemic inflammatory response syndrome. *Dig Dis Sci* 56(4):1171–1177
- Zilahi G, Artigas A, Martin-Loeches I (2016) What's new in multidrug-resistant pathogens in the ICU? *Ann Intensive Care* 6:96
- Taur Y, Xavier JB, Lipuma L, Ubeda C, Goldberg J, Gobourne A, Lee YJ, Dubin KA, Socci ND, Viale A, Perales MA, Jenq RR, van den Brink MR, Pamer EG (2012) Intestinal domination and the risk of bacteremia in patients undergoing allogeneic hematopoietic stem cell transplantation. *Clin Infect Dis* 55(7):905–914
- Cammarota G, Ianiro G, Tilg H et al (2017) European FMT Working Group. European consensus conference on faecal microbiota transplantation in clinical practice. *Gut* 66:569–580
- McDonald LC, Gerding DN, Johnson S et al (2018) Clinical practice guidelines for *Clostridium difficile* infection in adults and children: 2017 update by the Infectious Diseases Society of America (IDSA) and Society for Healthcare Epidemiology of America (SHEA). *Clin Infect Dis* 66:987–994
- Hocquart M, Lagier JC, Cassir N et al (2018) Early fecal microbiota transplantation improves survival in severe *Clostridium difficile* infections. *Clin Infect Dis* 66:645–650
- Huttner BD, de Lastours V, Wassenberg M et al (2019) A 5-day course of oral antibiotics followed by faecal transplantation to eradicate carriage of multidrug-resistant Enterobacteriaceae: a randomized clinical trial. *Clin Microbiol Infect*. <https://doi.org/10.1016/j.cmi.2018.12.009>
- Rossen NG, Fuentes S, van der Spek MJ, Tijssen JG, Hartman JH, Duflou A, Löwenberg M, van den Brink GR, Mathus-Vliegen EM, de Vos WM, Zoetendal EG, D'Haens GR, Ponsioen CY (2015) Findings from a randomized controlled trial of fecal transplantation for patients with ulcerative colitis. *Gastroenterology* 149(1):110–118.e4. <https://doi.org/10.1053/j.gastro.2015.03.045> (Epub 2015 Mar 30)
- Ott SJ, Waetzig GH, Rehman A (2017) Efficacy of sterile fecal filtrate transfer for treating patients with *Clostridium difficile* infection. *Gastroenterology* 152(4):799.e7–811.e7
- Ramai D, Zakhia K, Ofosu A, Ofori E, Reddy M (2019) Fecal microbiota transplantation: donor relation, fresh or frozen, delivery methods, cost effectiveness. *Ann Gastroenterol* 32(1):30–38