



Gout in males: a possible role for *COMT* hypomethylation

Xiuru Ying¹ · Yanfei Chen¹ · Zhonghua Zheng¹ · Shiwei Duan¹

Received: 10 December 2018 / Revised: 27 April 2019 / Accepted: 15 May 2019 / Published online: 4 June 2019
© International League of Associations for Rheumatology (ILAR) 2019

Abstract

Objective Gout is a common inflammatory disease, and the prevalence of gout in men is significantly higher than in women. Catechol-O-methyltransferase (*COMT*) regulates dopamine activity and metabolism, thereby participating in the uric acid metabolism, which in turn affects the occurrence of gout. Our study aimed to investigate the association between *COMT* methylation and gout in men.

Methods This study involved 57 male gout patients and 103 age-matched healthy men. We used quantitative methylation-specific polymerase chain reaction (qMSP) to determine DNA methylation levels in the blood. The *COMT* methylation level was represented by the percentage of methylation reference (PMR).

Results Our results showed that *COMT* methylation levels were significantly lower in gout patients than in the control group (median PMR 9.50 vs 31.34, $p = 3E-5$). The area under the curve (AUC) was 0.701 (95% CI 0.611–0.790, $p = 2.7E-5$) with a sensitivity of 68% and a specificity of 68.4%.

Conclusion Our study found that there was a significant correlation between *COMT* hypomethylation and the risk of gout in males, and this provides an epigenetic mechanism of *COMT* in gout. *COMT* hypomethylation might be used as a potential diagnostic biomarker for gout in the future.

Keywords Catechol-O-methyltransferase · DNA methylation · Gout · Male · Promoter

Introduction

Gout is a common inflammatory disease that is featured with chronic deposition of monosodium urate (MSU) crystals [1]. Clinical features of gout include tophi, chronic gouty arthritis, destructive joint disease, and acute recurrence of severe inflammatory arthritis [2]. Gout can develop into a variety of chronic diseases, including hypertension, hyperlipidemia, obesity, kidney disease, and cardiovascular disease, but the direct causal relationship between uric acid and these diseases is unclear [3–5].

Previous studies have shown that there are at least two possible mechanisms for the development of gout. The first mechanism involves the dopaminergic system [6], which plays an important role in regulating blood pressure, sodium

balance, and renal function; the second one is associated with dopamine degradation. Dopamine regulates renal function [6], induces glomerular filtration [7], and subsequently reduces uric acid levels and excretion rates [7]. Dopamine degradation may increase the reabsorption of sodium and urate, eventually leading to gout [7–9].

Human *COMT* gene encodes catechol-O-methyltransferase (*COMT*), which is widely expressed and modulates renal dopamine activity [10]. *COMT* is most expressed in the liver and kidney, and high expression of *COMT* in renal epithelial cells is thought to indirectly regulate the metabolism of dopamine and other catecholamines [11]. Studies have shown that rs4680 (Val158Met) of *COMT* protects from gout and may affect gout by regulating dopamine levels [12]. However, in another study in Taiwan aboriginal Chinese, *COMT* Val158Met was found to be unrelated to gout [13].

COMT methylation has been reported to be associated with coronary heart disease [14] and schizophrenia [15]. *COMT* methylation was found to be inversely correlated with *COMT* mRNA expression in human cell lines [16]. *COMT* hypomethylation predicts an increase in *COMT* expression and decreases the complement level of frontal dopamine in the carriers of *COMT* Met158 allele [16]. The regulation of

XY and YC are co-first authors of this work

✉ Shiwei Duan
duanshiwei@nbu.edu.cn

¹ Medical Genetics Center, School of Medicine, Ningbo University, Ningbo, Zhejiang 315211, China

COMT expression is complex and may depend on a variety of factors and interactions, including the *COMT* Val158Met genotype, DNA methylation, and a range of other biological and environmental factors [16]. However, the relationship between *COMT* methylation and gout remains unknown.

The prevalence of gout in men is significantly higher than that in women in many countries, and the ratio of men to women is generally between 3 and 4:1 [17]. The large differences in the risk of men and women may be primarily related to the role of estrogen in promoting uric acid (UA) excretion, as studies have shown that the use of hormone replacement therapy can reduce the risk of gout during menopause [18]. In this study, we selected male individuals and investigated the relationship between *COMT* methylation and gout.

Materials and methods

Subjects

The study included 57 male gout patients and 103 healthy male patients from Ningbo Second Hospital. All gout patients were diagnosed according to the American College of Rheumatology criteria [19]. The blood samples were collected from patients before medical treatment. The age distribution of the cases was 51.52 ± 14.27 years, and the age distribution of the controls was 50.15 ± 11.94 ($p = 0.454$). Plasma levels of various biochemical factors were measured by the method described in the previous study [20, 21], including glutamic-pyruvic transaminase (ALT), glutamic oxalacetic transaminase (AST), creatinine (CRE), UA, blood glucose (Glu), cholesterol, high-density lipoprotein (HDL), low-density lipoprotein (LDL), and triglyceride (TG). The research protocol was approved by the Ethics Committee of Ningbo University. All participants signed an informed consent form.

DNA methylation assay

The details of blood DNA separation and subsequent bisulfite conversion were as previously described [22]. The methylation level of *COMT* was measured using a quantitative methylation-specific polymerase chain reaction (qMSP). The primer sequences of *COMT* were 5'-GGTGTAGTTAGTATAGTAGGATT-3' (forward primer) and 5'-CCACATACGACCTCTTA-3' (reverse primer), respectively. The forward primer sequence of the reference gene (ACTB) was 5'-TGGTGATGGAGGAGGTTTAGTAAGT-3', and its reverse primer sequence was 5'-AACCAATAAAACCTACTCCTCCCTTAA-3'. Details of qMSP can be found in our previous publication [23]. The percentage of methylation reference (PMR) was calculated as: $PMR = 2^{-(Ct_{\text{sample}} - Ct_{\text{internal reference}})} \times 100\%$ [24].

Bioinformatics data for *COMT* methylation and *COMT* expression

We extracted 413 *COMT* methylation and *COMT* expression data from TCGA database (<http://www.cbioportal.org/>) to test their correlation. We also downloaded *COMT* expression in LCA and C2C12 cell lines treated with and without demethylation reagents (5'-AZA-deoxycytidine) from the Gene Expression Synthesis (GEO) database (GSE30192) to check whether decreased *COMT* methylation was able to up-regulate *COMT* expression.

Statistical analysis

Nonparametric test was used to compare *COMT* methylation between cases and controls. The Spearman rank correlation coefficient was used to quantify the relationship of *COMT* methylation with metabolic characteristics and *COMT* expression. Gene expression in cell lines treated with and without the demethylating agent 5'-AZA-deoxycytidine was compared using a paired sample *T* test. $p < 0.05$ was considered statistically significant.

Results

In this study, we recruited 57 Chinese male gout patients and 103 healthy male controls to investigate the role of *COMT* promoter methylation in gout. Among the 11 clinical phenotypes, ALT, UA, cholesterol, TG, WBC, AST, and Glu were found to be significantly higher in gout cases than in controls (all $p < 0.05$, Table 1).

In the current study, we selected a fragment (hg19: chr22: 19,941,454–19,941,583, Fig. 1) for the methylation assay in the CpG island of the *COMT* promoter. Our results showed that *COMT* methylation levels were significantly lower in gout patients than in the control group (median PMR 9.50 vs 31.34, $p = 3E-5$, Fig. 2). The area under the curve (AUC) was 0.701 (95% CI 0.611–0.790, $p = 2.7E-5$, Fig. 3) with a sensitivity of 68% and a specificity of 68.4%.

The relationship between *COMT* methylation and baseline characteristics was tested by Spearman correlation test. In gout patients, our results showed a trend that *COMT* methylation levels were positively correlated with age and glucose levels (age $r = 0.206$; Glu $r = 0.203$), and *COMT* methylation levels were inversely correlated with LDL levels ($r = -0.225$). Aging is closely associated with increased risk of hyperuricemia and gout [25], and several studies have observed that the prevalence of gout or serum uric acid increases with age [26, 27]. Type 2 diabetes has been reported to be associated with an increased risk of gout [28–30]. Both population study and animal model study have shown that hyperuricemia may play a role in insulin and glucose disorders [31]. LDL is

Table 1 Baseline characteristics of all the subjects

Characteristics	Cases (n = 57)	Controls (n = 103)	p value
Age (years)	52.79 ± 14.56	50.15 ± 11.94	0.217
ALT (U/L)	40.40 (21.00, 44.50)	22.83 (17.00, 26.00)	< 0.001 ^{a*}
AST (U/L)	29.79 (20.00, 38.00)	22.70 (18.00, 27.00)	0.021 ^{b*}
CRE (μmol/L)	80.16 ± 15.37	77.83 ± 9.77	0.243
UA (μmol/L)	426.51 ± 144.93	344.41 ± 67.15	< 0.001 [*]
Glu (mmol/L)	5.70 (4.82, 5.79)	5.00 (4.72, 5.25)	0.005 ^{a*}
Cholesterol (mmol/L)	5.00 ± 1.34	4.42 ± 0.67	< 0.001 [*]
HDL (mmol/L)	1.21 ± 0.34	1.51 ± 0.36	0.997
LDL (mmol/L)	2.60 ± 1.05	2.60 ± 0.52	0.553
TG (mmol/L)	2.52 ± 1.46	1.15 ± 0.47	< 0.001 ^{b*}
WBC (10 ⁹ /L)	9.29 ± 3.31	6.40 ± 1.56	< 0.001 [*]

*p value less than 0.05 was in italic. ^a Log-transformation was used. ^b Nonparametric rank test was applied. *ALT*, glutamic-pyruvic transaminase; *AST*, glutamic oxalacetic transaminase; *CRE*, creatinine; *UA*, uric acid; *Glu*, blood glucose; *HDL*, high-density lipoprotein; *LDL*, low-density lipoprotein; *TG*, triglyceride; *WBC*, white blood cell

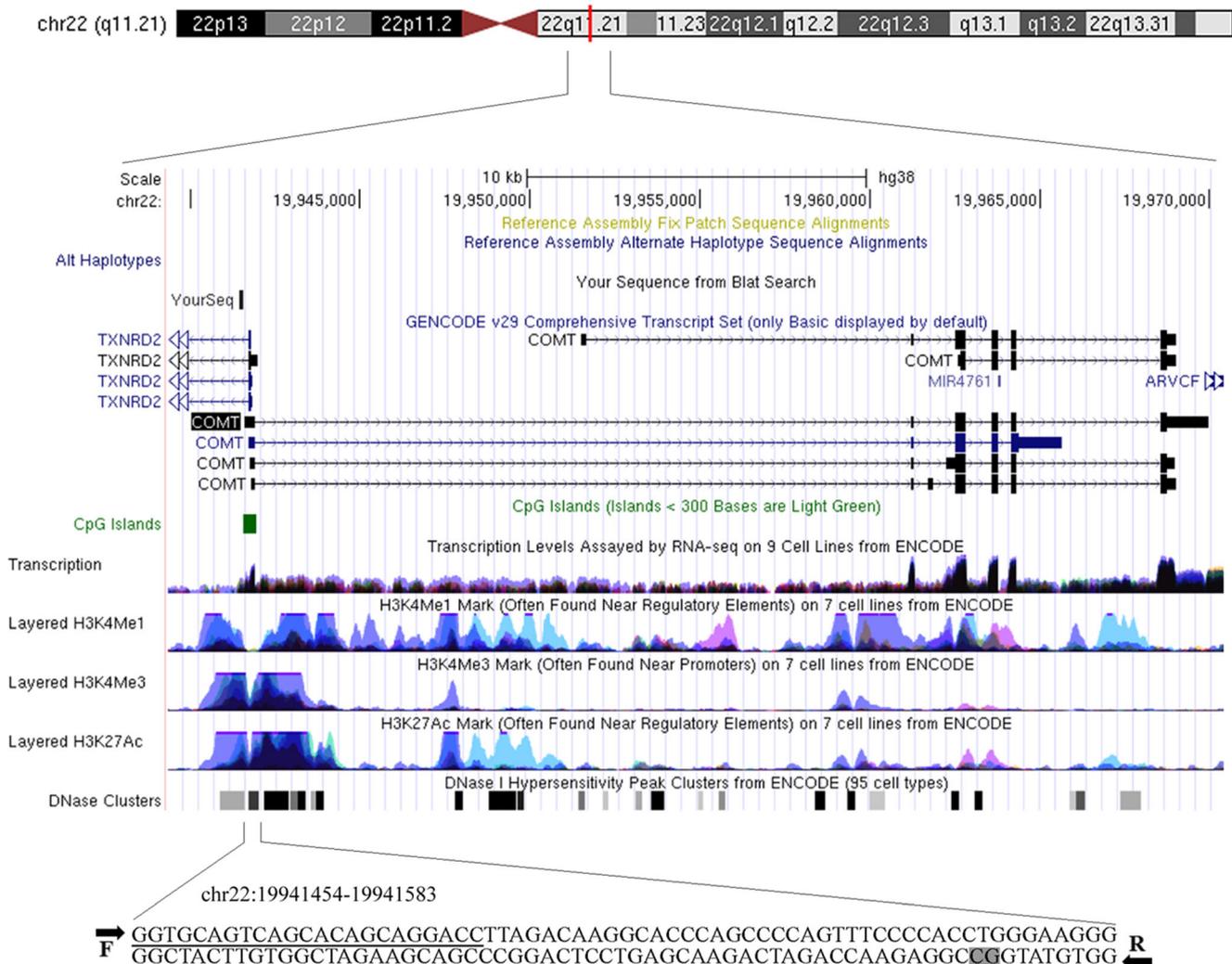


Fig. 1 The selected fragment of *COMT* in the methylation assay. Genomic positions and function annotations were obtained from UCSC genome browser according to human 2009 (GRCh37/hg19) assembly. F

and R were forward and reverse primers, respectively. The quantitative methylation-specific polymerase chain reaction (qMSP) primers were underlined, and CpG on the reverse primer was in gray

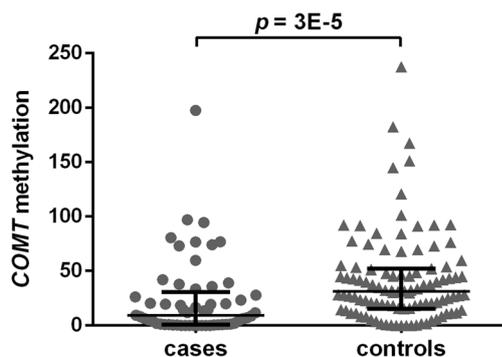


Fig. 2 Comparison of *COMT* methylation levels between gout cases and controls. *COMT* methylation was significantly lower in gout patients than in controls (median PMR 9.50 (0.93, 30.92) vs 31.34 (15.61, 51.92), $p = 3E-5$, Nonparametric rank test)

often oxidized to ox-LDL in a long-term oxidative environment [32]. Studies have found that ox-LDL levels in gout patients are significantly higher than healthy controls, and the changes in ox-LDL levels may be associated with increased inflammation in gout patients [33]. An increase in LDL levels and a decrease in HDL levels were also found in gout patients [34]. In addition, previous studies have shown that the Val158Met polymorphism of *COMT* is associated with LDL levels in plasma [35, 36]. Our results suggest that the level of *COMT* methylation is inversely correlated with LDL in gout patients, suggesting that LDL is a complex trait, and that both genetic and epigenetic factors are needed to study the relationship between *COMT* and LDL in the future. In the healthy controls, our results showed positive correlations of *COMT* promoter methylation with both age and TG

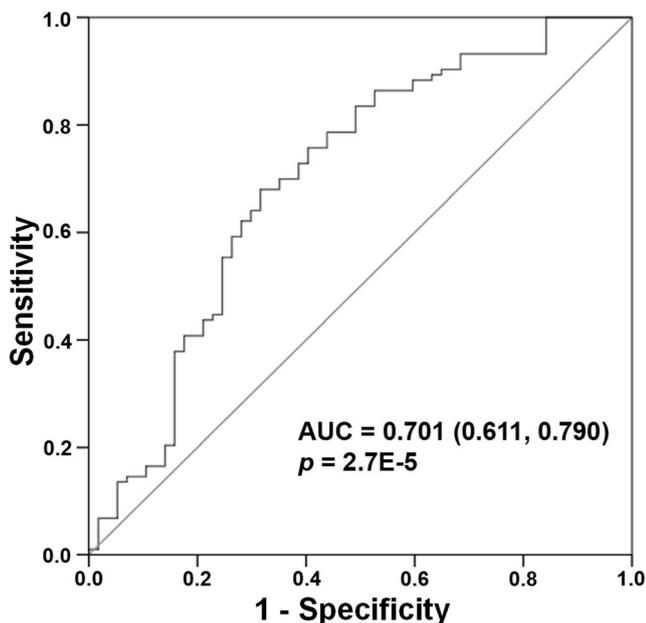


Fig. 3 Receiver operating characteristic (ROC) curve of *COMT* promoter hypomethylation for gout. The area under the curve (AUC) was 0.701 (95% CI 0.611–0.790, $p = 2.7E-5$) with a sensitivity of 68% and a specificity of 68.4%

levels (age $r = 0.249$, $p = 0.011$; TG $r = 0.244$, $p = 0.013$). In contrast, an inverse correlation was observed between *COMT* methylation and HDL levels ($r = -0.265$, $p = 0.007$). To note, we did not perform a correction for multiple comparisons, and thus there is a chance of false positive results for the above findings. In addition, *COMT* methylation was not significantly associated with other biochemical indicators (all $p > 0.05$, data not shown).

Our analysis of TCGA data showed a significant inverse correlation between methylation and expression of *COMT* (Spearman rank correlation test, $r = -0.521$, $p = 8E-30$, Fig. 4). Meanwhile, our analysis of GEO data showed that 5'-AZA-deoxycytidine treatment significantly increased *COMT* expression (mean fold change = 1.41, $p = 0.003$, Fig. 5), suggesting that *COMT* expression can be induced by demethylating agent.

Discussion

In this study, we found that *COMT* methylation in gout cases was significantly lower than in the matched controls. The ROC curve suggested that *COMT* hypomethylation was a potential biomarker for gout. In addition, there was a significant correlation between *COMT* methylation and several phenotypes such as HDL levels in controls. All the above findings suggested that *COMT* hypomethylation might be critical for gout development.

COMT is the major enzyme of catecholamine neurotransmitters (including dopamine, norepinephrine, and epinephrine), which degrades catecholamine neurotransmitters by enzymatic O-methylation of a single hydroxyl group in the catecholamine motif [37]. *COMT* is abundantly expressed in the

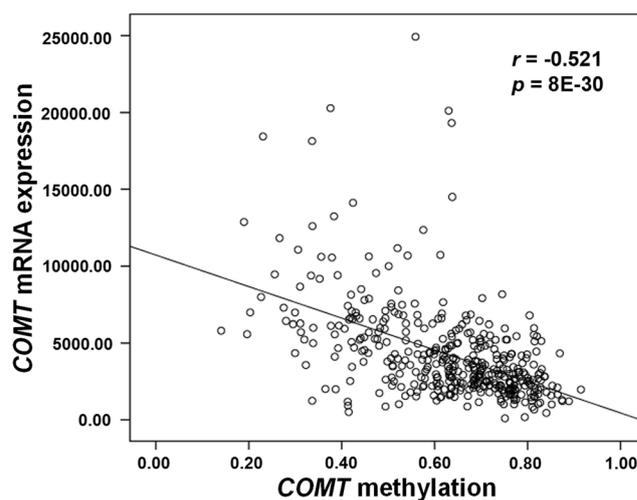


Fig. 4 Inverse correlation between *COMT* methylation and *COMT* expression in 413 individuals from TCGA dataset. The methylation and expression of *COMT* were significantly inversely correlated ($r = -0.521$, $p = 8E-30$, Spearman rank correlation test)

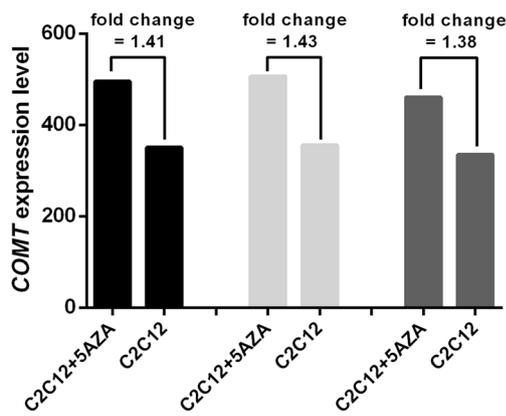


Fig. 5 5'-aza-deoxycytidine treatment increased *COMT* expression. Our results showed C2C12 cell lines with 5'-aza-deoxycytidine treatment had a significantly higher *COMT* expression than those without treatment (average fold change = 1.41, $p = 0.003$, paired-samples t test). The data of *COMT* expression was retrieved from the GEO database (GSE30192)

kidney and plays an important role in dopamine metabolism, especially in the action of the natriuretic peptide of the renal dopamine system [11]. Dopamine-induced glomerular filtration was inversely associated with uric acid concentration and uric acid excretion [7]. Renal uric acid transport plays an important role in the acquisition of factors affecting UA [38]. UA is above the saturation threshold and produces MSU crystals in and around the joints, which is the result of gout after long-term hyperuricemia [39].

Gout patients often have acute inflammatory reactions induced by MSU crystals, and the induced inflammation by gout were mainly mediated by macrophages and neutrophils [40]. In our study, we observed a significant increase in blood leukocytes of gout patients, suggesting a significantly high inflammatory response in our gout patients.

Gout is a complex disease that is often linked to the higher incidence of depression. It is reasonable that some biomarkers of gout might be also linked to other diseases or disorders. Our results showed that *COMT* methylation levels were significantly lower in gout patients than in the control group (median PMR 9.50 vs 31.34, $p = 3E-5$). The area under the curve (AUC) was 0.701 (95% CI 0.611–0.790, $p = 2.7E-5$) with a sensitivity of 68% and a specificity of 68.4%.

The moderate specificity of *COMT* methylation in the detection of gout might be due to the potential effect of diverse cellular distribution on gene methylation in whole blood. In acute gout, the natural killer cells emerge as mediator of inflammation [41]. In the blood of gout patients, shorter telomeres were found compared to those in healthy participants, indicating increased cellular senescence in the blood [42]. All the above evidence suggests that the distribution of cellular subsets in blood is likely to be different between gout patients and healthy controls. In the current study, we detected *COMT* methylation in peripheral blood DNA that comes from a

variety of cells, such as granulocytes and lymphocytes. Researchers suggested that DNA methylation based-association tests in whole blood should be adjusted by a set of methylation makers to avoid the impact of cellular composition [43]. DNA from a mixture of cells in the blood may affect the specificity of methylation detection to some extent.

Vitamin D levels were reduced in plasma from gout patients [44], and a significant reduction in catechol-*o*-methyltransferase (*COMT*) was observed in vitamin D-deficient neonatal rats [45]. There are three VDR binding regions in the *COMT* promoter region, which can be enriched with 1,25-dihydroxyvitamin D, the biologically active form of vitamin D [45]. And this strongly suggests that vitamin D directly regulates the expression of *COMT* [45]. Moreover, recent work has shown that the risk of depression in gout patients increased by 19% [46], especially in elderly gout patients, who had a 42% increased risk of depression [47]. It is known that the main symptoms of depression consist of decreased levels of extracellular dopamine [48]. *COMT* is an enzyme involved in the degradation of dopamine that affects dopamine levels especially in the prefrontal cortex [49]. The *COMT* Val158Met polymorphism (rs4680) has been reported to be associated with depression [50]. Individuals with the *COMT* Val allele have higher activity of *COMT* and lower baseline dopamine levels [51] compared to individuals with the *COMT* Met allele. However, the association results between depression and *COMT* Val158Met polymorphism are sometimes contradictory, and this may be due to ethnic differences [52]. *COMT* Val158Met was found to play a protective role in the development of gout in Chinese population [12], and this positive association was not validated in Taiwan's aborigines [13]. No studies have been found between *COMT* methylation and gout. Our study found that there was a significant correlation between *COMT* hypomethylation and the risk of gout in males, and this provides an epigenetic mechanism of *COMT* in gout. In the future, we need to further study whether there are interactions of the *COMT* promoter methylation with *COMT* Val158Met polymorphism and the regulation of vitamin D.

There are some shortcomings in our research. First of all, previous study showed that *COMT* polymorphisms were associated to gout in Chinese patients [12]. Due to the moderate sample size in our study, future work with more samples is needed to explore whether *COMT* polymorphisms may interact with *COMT* methylation in the contribution of *COMT* to gout. In addition, although TCGA and GEO data analysis confirmed that *COMT* hypomethylation was associated with high expression of *COMT*, future work needs to validate the association of *COMT* hypomethylation with gene expression. Meanwhile, though the CpG site in the current methylation study is likely to represent the CpGs in the surrounding region, it is necessary to further determine the methylation of other

regions of *COMT*. Lastly, but not least, we used peripheral blood in this study to identify molecular markers for in vitro diagnosis of gout risk. Future work on renal epithelial cells is needed to explore the relevant mechanisms of *COMT* methylation.

For the first time our study revealed the relationship between *COMT* hypomethylation and increased risk of gout in men. Our findings might also provide an epigenetic clue of *COMT* enzyme in gout.

Funding information The research is supported by K. C. Wong Magna Fund in Ningbo University.

Compliance with ethical standards

Disclosures None.

References

- McCarty DJ, Hollander JL (1961) Identification of urate crystals in gouty synovial fluid. *Ann Intern Med* 54:452–460
- Dalbeth N, Stamp L (2014) Hyperuricaemia and gout: time for a new staging system? *Ann Rheum Dis* 73(9):1598–1600. <https://doi.org/10.1136/annrheumdis-2014-205304>
- Hughes K, Flynn T, de Zoysa J, Dalbeth N, Merriman TR (2014) Mendelian randomization analysis associates increased serum urate, due to genetic variation in uric acid transporters, with improved renal function. *Kidney Int* 85(2):344–351. <https://doi.org/10.1038/ki.2013.353>
- Palmer TM, Nordestgaard BG, Benn M, Tybjaerg-Hansen A, Davey Smith G, Lawlor DA, Timpson NJ (2013) Association of plasma uric acid with ischaemic heart disease and blood pressure: mendelian randomisation analysis of two large cohorts. *Bmj* 347:f4262. <https://doi.org/10.1136/bmj.f4262>
- Kang EH, Lee EY, Lee YJ, Song YW, Lee EB (2008) Clinical features and risk factors of postsurgical gout. *Ann Rheum Dis* 67(9):1271–1275. <https://doi.org/10.1136/ard.2007.078683>
- Zeng C, Zhang M, Asico LD, Eisner GM, Jose PA (2007) The dopaminergic system in hypertension. *Clin Sci* 112(12):583–597. <https://doi.org/10.1042/CS20070018>
- Sulikowska B, Manitijs J, Odrowaz-Sypniewska G, Lysiak-Szydłowska W, Rutkowski B (2008) Uric acid excretion and dopamine-induced glomerular filtration response in patients with IgA glomerulonephritis. *Am J Nephrol* 28(3):391–396. <https://doi.org/10.1159/000112271>
- Pestana M, Jardim H, Correia F, Vieira-Coelho MA, Soares-da-Silva P (2001) Renal dopaminergic mechanisms in renal parenchymal diseases and hypertension. *Nephrology, dialysis, transplantation: official publication of the European Dialysis and transplant association. European Renal Association* 16(Suppl 1):53–59
- Jose PA, Raymond JR, Bates MD, Aperia A, Felder RA, Carey RM (1992) The renal dopamine receptors. *Journal of the American Society of Nephrology: JASN* 2(8):1265–1278
- Gogos JA, Morgan M, Luine V, Santha M, Ogawa S, Pfaff D, Karayiorgou M (1998) Catechol-O-methyltransferase-deficient mice exhibit sexually dimorphic changes in catecholamine levels and behavior. *Proc Natl Acad Sci U S A* 95(17):9991–9996
- Eklof AC, Holtback U, Sundelof M, Chen S, Aperia A (1997) Inhibition of COMT induces dopamine-dependent natriuresis and inhibition of proximal tubular Na⁺,K⁺-ATPase. *Kidney Int* 52(3):742–747
- Dong Z, Zhao D, Yang C, Zhou J, Qian Q, Ma Y, He H, Ji H, Yang Y, Wang X, Xu X, Pang Y, Zou H, Jin L, Wang J (2015) Common variants in LRP2 and COMT genes affect the susceptibility of gout in a Chinese population. *PLoS One* 10(7):e0131302. <https://doi.org/10.1371/journal.pone.0131302>
- Tu HP, Ko AM, Wang SJ, Lee CH, Lea RA, Chiang SL, Chiang HC, Wang TN, Huang MC, Ou TT, Lin GT, Ko YC (2010) Monoamine oxidase a gene polymorphisms and enzyme activity associated with risk of gout in Taiwan aborigines. *Hum Genet* 127(2):223–229. <https://doi.org/10.1007/s00439-009-0765-z>
- Zhong J, Chen X, Wu N, Shen C, Cui H, Du W, Zhang Z, Feng M, Liu J, Lin S, Zhang L, Wang J, Chen X, Duan S (2016) Catechol-O-methyltransferase promoter hypomethylation is associated with the risk of coronary heart disease. *Experimental and therapeutic medicine* 12(5):3445–3449. <https://doi.org/10.3892/etm.2016.3757>
- Gao S, Cheng J, Li G, Sun T, Xu Y, Wang Y, Du X, Xu G, Duan S (2017) Catechol-O-methyltransferase gene promoter methylation as a peripheral biomarker in male schizophrenia. *European psychiatry: the journal of the Association of European Psychiatrists* 44:39–46. <https://doi.org/10.1016/j.eurpsy.2017.03.002>
- Swift-Scanlan T, Smith CT, Bardowell SA, Boettiger CA (2014) Comprehensive interrogation of CpG island methylation in the gene encoding COMT, a key estrogen and catecholamine regulator. *BMC Med Genet* 7:5. <https://doi.org/10.1186/1755-8794-7-5>
- Kuo CF, Grainge MJ, Zhang W, Doherty M (2015) Global epidemiology of gout: prevalence, incidence and risk factors. *Nat Rev Rheumatol* 11(11):649–662. <https://doi.org/10.1038/nrrheum.2015.91>
- Hak AE, Curhan GC, Grodstein F, Choi HK (2010) Menopause, postmenopausal hormone use and risk of incident gout. *Ann Rheum Dis* 69(7):1305–1309. <https://doi.org/10.1136/ard.2009.109884>
- Wallace SL, Robinson H, Masi AT, Decker JL, McCarty DJ, Yu TF (1977) Preliminary criteria for the classification of the acute arthritis of primary gout. *Arthritis Rheum* 20(3):895–900
- Chang L, Wang Y, Ji H, Dai D, Xu X, Jiang D, Hong Q, Ye H, Zhang X, Zhou X, Liu Y, Li J, Chen Z, Li Y, Zhou D, Zhuo R, Zhang Y, Yin H, Mao C, Duan S, Wang Q (2014) Elevation of peripheral BDNF promoter methylation links to the risk of Alzheimer's disease. *PLoS One* 9(11):e110773. <https://doi.org/10.1371/journal.pone.0110773>
- Jiang D, Zheng D, Wang L, Huang Y, Liu H, Xu L, Liao Q, Liu P, Shi X, Wang Z, Sun L, Zhou Q, Li N, Xu L, Le Y, Ye M, Shao G, Duan S (2013) Elevated PLA2G7 gene promoter methylation as a gender-specific marker of aging increases the risk of coronary heart disease in females. *PLoS One* 8(3):e59752. <https://doi.org/10.1371/journal.pone.0059752>
- Hong Q, Chen X, Ye H, Zhou A, Gao Y, Jiang D, Wu X, Tian B, Chen Y, Wang M, Xie J, Xia Y, Duan S (2016) Association between the methylation status of the MGMT promoter in bone marrow specimens and chemotherapy outcomes of patients with acute myeloid leukemia. *Oncol Lett* 11(4):2851–2856. <https://doi.org/10.3892/ol.2016.4317>
- Chen R, Hong Q, Jiang J, Chen X, Jiang Z, Wang J, Liu S, Duan S, Shi S (2017) AGTR1 promoter hypermethylation in lung squamous cell carcinoma but not in lung adenocarcinoma. *Oncol Lett* 14(4):4989–4994. <https://doi.org/10.3892/ol.2017.6824>
- Kristensen LS, Mikeska T, Krypuy M, Dobrovic A (2008) Sensitive melting analysis after real time- methylation specific PCR (SMART-MSP): high-throughput and probe-free quantitative DNA methylation detection. *Nucleic Acids Res* 36(7):e42. <https://doi.org/10.1093/nar/gkn113>

25. MacFarlane LA, Kim SC (2014) Gout: a review of nonmodifiable and modifiable risk factors. *Rheumatic diseases clinics of North America* 40(4):581–604. <https://doi.org/10.1016/j.rdc.2014.07.002>
26. Wallace KL, Riedel AA, Joseph-Ridge N, Wortmann R (2004) Increasing prevalence of gout and hyperuricemia over 10 years among older adults in a managed care population. *J Rheumatol* 31(8):1582–1587
27. Fang J, Alderman MH (2000) Serum uric acid and cardiovascular mortality the NHANES I epidemiologic follow-up study, 1971–1992. *National Health and nutrition examination survey. Jama* 283(18):2404–2410
28. Li C, Hsieh MC, Chang SJ (2013) Metabolic syndrome, diabetes, and hyperuricemia. *Curr Opin Rheumatol* 25(2):210–216. <https://doi.org/10.1097/BOR.0b013e32835d951e>
29. Choi HK, De Vera MA, Krishnan E (2008) Gout and the risk of type 2 diabetes among men with a high cardiovascular risk profile. *Rheumatology* 47(10):1567–1570. <https://doi.org/10.1093/rheumatology/ken305>
30. Dehghan A, van Hoek M, Sijbrands EJ, Hofman A, Witteman JC (2008) High serum uric acid as a novel risk factor for type 2 diabetes. *Diabetes Care* 31(2):361–362. <https://doi.org/10.2337/dc07-1276>
31. Thottam GE, Krasnokutsky S, Pillinger MH (2017) Gout and metabolic syndrome: a tangled web. *Curr Rheumatol Rep* 19(10):60. <https://doi.org/10.1007/s11926-017-0688-y>
32. Kappelle PJ, Bijzet J, Hazenberg BP, Dullaart RP (2011) Lower serum paraoxonase-1 activity is related to higher serum amyloid A levels in metabolic syndrome. *Arch Med Res* 42(3):219–225. <https://doi.org/10.1016/j.arcmed.2011.05.002>
33. Jiang X, Li M, Yang Q, Du L, Du J, Zhou J (2014) Oxidized low density lipoprotein and inflammation in gout patients. *Cell Biochem Biophys* 69(1):65–69. <https://doi.org/10.1007/s12013-013-9767-5>
34. Takahashi S, Yamamoto T, Moriwaki Y, Tsutsumi Z, Higashino K (1994) Impaired lipoprotein metabolism in patients with primary gout—influence of alcohol intake and body weight. *Br J Rheumatol* 33(8):731–734
35. Ge L, Wu HY, Pan SL, Huang L, Sun P, Liang QH, Pang GF, Lv ZP, Hu CY, Liu CW, Zhou XL, Huang LJ, Yin RX, Peng JH (2015) COMT Val158Met polymorphism is associated with blood pressure and lipid levels in general families of Bama longevous area in China. *Int J Clin Exp Pathol* 8(11):15055–15064
36. Yoshida T, Kato K, Yokoi K, Oguri M, Watanabe S, Metoki N, Yoshida H, Satoh K, Aoyagi Y, Nishigaki Y, Nozawa Y, Yamada Y (2009) Association of genetic variants with chronic kidney disease in individuals with different lipid profiles. *Int J Mol Med* 24(2):233–246
37. Guldberg HC, Marsden CA (1975) Catechol-O-methyl transferase: pharmacological aspects and physiological role. *Pharmacol Rev* 27(2):135–206
38. Mount DB (2013) The kidney in hyperuricemia and gout. *Curr Opin Nephrol Hypertens* 22(2):216–223. <https://doi.org/10.1097/MNH.0b013e32835ddad2>
39. Perez-Ruiz F, Dalbeth N, Bardin T (2015) A review of uric acid, crystal deposition disease, and gout. *Adv Ther* 32(1):31–41. <https://doi.org/10.1007/s12325-014-0175-z>
40. So AK, Martinon F (2017) Inflammation in gout: mechanisms and therapeutic targets. *Nat Rev Rheumatol* 13(11):639–647. <https://doi.org/10.1038/nrrheum.2017.155>
41. Empson VG, McQueen FM, Dalbeth N (2010) The natural killer cell: a further innate mediator of gouty inflammation? *Immunol Cell Biol* 88(1):24–31. <https://doi.org/10.1038/icc.2009.91>
42. Vazirpanah N, Kienhorst LBE, Van Lochem E, Wichers C, Rossato M, Shiels PG, Dalbeth N, Stamp LK, Merriman TR, Janssen M, Radstake T, Broen JC (2017) Patients with gout have short telomeres compared with healthy participants: association of telomere length with flare frequency and cardiovascular disease in gout. *Ann Rheum Dis* 76(7):1313–1319. <https://doi.org/10.1136/annrheumdis-2016-210538>
43. Koestler DC, Usset J, Christensen BC, Marsit CJ, Karagas MR, Kelsey KT, Wiencke JK (2017) DNA methylation-derived neutrophil-to-lymphocyte ratio: an epigenetic tool to explore cancer inflammation and outcomes. *Cancer Epidemiol Biomark Prev* 26(3):328–338. <https://doi.org/10.1158/1055-9965.EPI-16-0461>
44. Zhou Q, Shao YC, Gan ZQ, Fang LS (2019) Lower vitamin D levels are associated with depression in patients with gout. *Neuropsychiatr Dis Treat* 15:227–231. <https://doi.org/10.2147/NDT.S193114>
45. Pertile RA, Cui X, Eyles DW (2016) Vitamin D signaling and the differentiation of developing dopamine systems. *Neuroscience* 333:193–203. <https://doi.org/10.1016/j.neuroscience.2016.07.020>
46. Lin S, Zhang H, Ma A (2018) Association of gout and depression: a systematic review and meta-analysis. *Int J Geriatr Psychiatry* 33(3):441–448. <https://doi.org/10.1002/gps.4789>
47. Singh JA, Cleveland JD (2018) Gout and the risk of incident depression in older adults. *Psychiatry Res* 270:842–844. <https://doi.org/10.1016/j.psychres.2018.10.067>
48. Lambert G, Johansson M, Agren H, Friberg P (2000) Reduced brain norepinephrine and dopamine release in treatment-refractory depressive illness: evidence in support of the catecholamine hypothesis of mood disorders. *Arch Gen Psychiatry* 57(8):787–793
49. Mannisto PT, Kaakkola S (1999) Catechol-O-methyltransferase (COMT): biochemistry, molecular biology, pharmacology, and clinical efficacy of the new selective COMT inhibitors. *Pharmacol Rev* 51(4):593–628
50. Gatt JM, Burton KL, Williams LM, Schofield PR (2015) Specific and common genes implicated across major mental disorders: a review of meta-analysis studies. *J Psychiatr Res* 60:1–13. <https://doi.org/10.1016/j.jpsychires.2014.09.014>
51. Lachman HM, Papolos DF, Saito T, Yu YM, Szumlanski CL, Weinshilboum RM (1996) Human catechol-O-methyltransferase pharmacogenetics: description of a functional polymorphism and its potential application to neuropsychiatric disorders. *Pharmacogenetics* 6(3):243–250
52. Wang M, Ma Y, Yuan W, Su K, Li MD (2016) Meta-analysis of the COMT Val158Met polymorphism in major depressive disorder: effect of ethnicity. *J NeuroImmune Pharmacol* 11(3):434–445. <https://doi.org/10.1007/s11481-016-9651-3>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.