



The expression of galectin-3 in breast cancer and its association with chemoresistance: a systematic review of the literature

Ioannis Boutas^{1,2} · Anastasios Potiris¹ · Walburgis Brenner² · Antje Lebrecht² · Annette Hasenburg² · Sophia Kalantaridou¹ · Marcus Schmidt²

Received: 21 April 2019 / Accepted: 4 September 2019 / Published online: 9 September 2019
© Springer-Verlag GmbH Germany, part of Springer Nature 2019

Abstract

Purpose Galectin-3 is a M_r 31,000 protein that belongs to a family of carbohydrate-binding proteins. Galectin-3 has already been associated with protection against apoptosis through cell to cell or cell to matrix adhesion processes. It seems that galectin-3 plays an important role in tumor progression, cell growth, invasion and metastasis. Galectin-3 is the only member of the chimeric galectins that has an N-terminal glycine and proline domain and a C-terminal carbohydrate recognition domain that allows galectin-3 to accommodate larger structures such as polylectosaminoglycans and intervene to DNA damage repair process. In this systematic review, our primary goal is to identify the effect of galectin-3 expression in association with drug resistance and apoptosis inhibition in breast cancer.

Materials and methods Scopus and PubMed databases were searched on 26 November 2018 using the following combination of keywords: (galectin-3 OR gal-3 OR LGALS3) AND (breast cancer) AND (chemoresistance OR (drug resistance) OR chemosensitivity). All the articles in English, regardless the time of publication, text availability and species included were initially accepted.

Results In the majority of the included studies, the expression of galectin-3 had a protective role in cell survival via different pathways such as the response to DNA damage and repair or the inhibition of apoptosis after treatment with a chemotherapeutic agent.

Conclusion Galectin-3 expression in breast tumors might be an important factor in the selection of the most suitable treatment.

Keywords Galectin-3 · Breast cancer · Chemoresistance · Chemosensitivity · Drug-induced apoptosis

Introduction

Galectin-3 is a M_r 31,000 protein that belongs to a family of carbohydrate-binding proteins which have affinity for b-galactoside containing glycoconjugates and conserved amino acid sequences in their carbohydrate-binding domains [1, 2]. It seems that galectin-3 plays an important role in tumor progression, cell growth, invasion and metastasis [3, 4]. Galectin-3 has already been associated with protection

against apoptosis through cell to cell or cell to matrix adhesion processes. This is mediated by the recognition of cell surface glycoconjugates and by activating human neutrophils, pre-mRNA splicing and raising interleukin-1 levels by human monocytes [4–7]. Another important aspect is that galectin-3 is involved in tumor progression and apoptosis by affecting the DNA damage repair response. DNA damage repair is a sophisticated process that includes injury detection and repair [8]. In that context, galectin-3 is the only member of the chimeric galectins that has an N-terminal glycine and proline domain and a C-terminal carbohydrate recognition domain that allows galectin-3 to accommodate larger structures such as polylectosaminoglycans and intervene to DNA damage repair process [9, 10]. By now, clinical research studies have correlated galectin-3 expression with several types of malignancies in head and neck, thyroid gland, gastric organs, colon and breast [11–15].

✉ Anastasios Potiris
apotiris@med.uoa.gr

¹ Third Department of Obstetrics and Gynecology, Attikon University Hospital, Medical School of the National and Kapodistrian University of Athens, Athens, Greece

² Department of Obstetrics and Women's Health, University Medical Center, Johannes Gutenberg-University, Mainz, Germany

Breast cancer is worldwide leading cancer in incidence and mortality for women [16]. In several studies, galectin-3 expression has been associated with breast cancer progression and metastasis using human breast cancer cell lines [17, 18]. Galectin-3 is the only known galectin with the capability to inhibit apoptosis [4]. This ability is a combined result of increased expression of anti-apoptotic proteins such as members of the Bcl-2 family and the ability to heterodimerize with Bcl-2 [5, 18–20]. Moreover, overexpression of galectin-3 has a protective impact on human breast cancer cells against tumor necrosis factor- α -induced apoptosis and nitric oxide-induced apoptosis [21]. Furthermore, it has been shown that galectin-3 levels are significantly higher in triple-negative breast carcinomas compared to non-triple-negative [22]. Ultimately, it has already been described that chemoresistance has been altered in galectin-3-silenced cancer cells [23].

Purpose of the review

In this systematic review, our primary goal is to identify the effect of galectin-3 expression in association with drug resistance and apoptosis inhibition.

Materials and methods

Scopus and PubMed databases were searched on February 26th, 2019 using the following combination of keywords: (galectin-3 OR gal-3 OR LGALS3) AND (breast cancer) AND (chemoresistance OR (drug resistance) OR

chemosensitivity). All the articles in English, regardless the time of publication, text availability and species included were initially accepted. Through the initial research, 55 publications were retrieved.

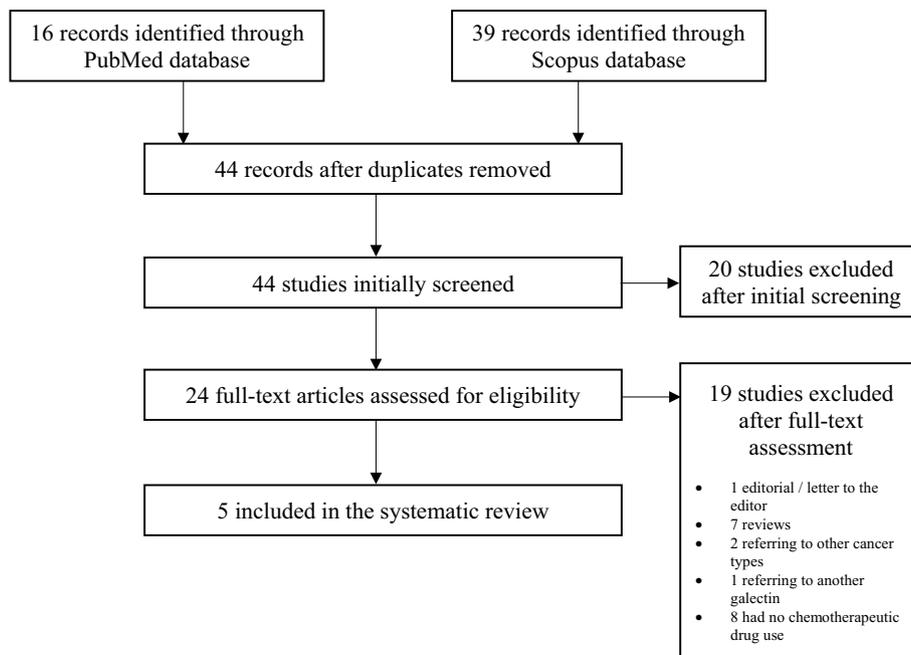
Title and abstract of all 55 publications were screened independently by two reviewers (I.B. and A.P.) and the full text of the eligible ones was screened subsequently. If there was a study selected only by one reviewer, the decision was taken by a third reviewer (S.K.).

Only original studies were selected that included data about the specimen (rats, mice and human cell lines), the levels of galectin-3 and the use of a chemotherapeutic drug. Reviews, editorials, letters to the editor and studies with no drug tested or different cancer types were excluded from the study.

From the initial 55 records identified, 11 duplicates were removed leading to 44 studies at initial screening. After the initial screening, 20 studies were excluded because they did not confront with the inclusion criteria of the present review and 24 studies were selected for full-text assessment. Through the full-text assessment, one study was an editorial, seven were reviews of galectin-3 in all cancers, one was referred to colorectal cancer, one to gastric cancer, one in galectin-7 and its association with galectin-3 and in eight, there was no chemotherapeutic drug use. After the comprehensive evaluation, five studies were selected for data extraction as Fig. 1 shows.

Data extraction was completed by the one reviewer (A.P.) and cross-checked by the second reviewer (I.B.). Data that were extracted was: author, year of publication, institution, specimen and chemotherapeutic drugs used. Concerning the

Fig. 1 Flow of the different phases of the systematic review



outcomes of each study, data about viability (time-dependent or not), apoptosis (time-dependent or not) and other associated protein levels were also extracted.

Results

cis-Diamminedichloroplatinum (CDDP)

Akahani et al. tested the significance of galectin-3 expression in cis-diamminedichloroplatinum cytotoxicity using sense and antisense (control) transfected, parental galectin-3-negative human breast carcinoma BT549 cell clones with confirmed and silence galectin-3 expression. After a 72-h exposure of 25 μM of CDDP, the viability of galectin-3-expressing clones was more than 60%, whereas in the silenced cell clones, was less than 30% [6]. Furthermore, to assess the apoptosis in the same CDDP-treated cells, they concluded that the apoptotic cells in the galectin-3 silenced clones were more than 30% in contrary to less than 20% of galectin-3-expressing clones. To further assess the morphologic features of apoptosis in the galectin-3-silenced population, more than 40% of the cells had morphologic features indicating apoptosis (DNA fragmentation and condensed chromatin), whereas the galectin-3-expressing cells had more visible nuclei without chromatin alterations [6]. With all the aforementioned facts, galectin-3 expression increases the chemoresistance of human breast carcinoma BT549 cell clones to CDDP and decreases the apoptotic effects of CDDP.

Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)

Mazurek et al. studied the effects of His64/Pro64 polymorphism of galectin-3 on its sensitivity to TRAIL. Nine breast cancer cell lines were analyzed concerning the sensitivity for TRAIL. After 4 h of treatment with 100 ng/mL of TRAIL, all the homozygous Pro64 galectin-3 cell lines were found not to be sensitive to TRAIL but all the homozygous His64 and one heterozygous were sensitive. Furthermore, they used BT549 cells for creating homozygous Pro64, homozygous His64 and heterozygous cell clones and found that TRAIL chemoresistance is significantly associated ($P < 0.05$) with Pro64 genotype [24]. Furthermore, when they tested galectin-3 null cell lines, after treatment with TRAIL, they found that they were also resistant to TRAIL [24].

Doxorubicin

In the previously described study, the authors additionally examined the doxorubicin sensitivity of BT549 cells with different polymorphism status after 16 h of treatment with 5 $\mu\text{g}/\text{mL}$ of doxorubicin. Among the different genotypes, the

sensitivity of doxorubicin was not found to alter. Moreover, the expression of His64 galectin-3 tends to render the cell lines more resistant to doxorubicin, whereas Pro64 galectin-3 has no effect [24].

Arsenic trioxide (ATO)

Zhang et al. used galectin-3-expressing MDA-MB-231 and MCF-7 breast cancer cell lines and showed that galectin-3 expression was significantly upregulated after 72 h of treatment with 2.5 mM of arsenic trioxide ($P < 0.01$) [25]. Regardless the higher levels of galectin-3, both cell lines showed a weak ATO-induced apoptosis. On the contrary, the galectin-3-knockdown MDA-MB-231 cell line showed a more than 20-fold increased apoptosis compared to control and a further two-fold increase when treated with ATO, suggesting that galectin-3 knockdown sensitizes these cells to ATO-induced apoptosis [25].

Etoposide, carboplatin and mitomycin C

Using silenced HEK293FT cell lines, Carvalho et al. performed cell viability assays after etoposide, carboplatin and mitomycin C treatment. Cells lacking galectin-3 expression showed an increased resistance to all three chemotherapeutic agents evaluated in all tested concentrations [26]. It is notable that *galectin-3*-silenced cells showed up to a 60% increase in viability when compared to control cells after treatment with 20 nM etoposide, suggesting that *galectin-3* has a major role in the cellular response to DNA damage and repair [26].

N-(4-Hydroxyphenyl)retinamide (4HPR)

Choi et al. tested the effects of 4HPR regarding viability and apoptosis on parental galectin-3-negative BT549 breast cancer cell clones. The three cell clones were a galectin-3-expressing line (BT549Gal-3Wt), an inactive galectin-3-expressing line (BT549Gal-3Mu) and an empty vector transfected, silenced galectin-3 line (BT549Vec). After a 3-day treatment with 5 μM of 4HPR, the inactive and silenced cell lines showed a marked reduction in survival. Namely, these cell lines had a 70–80% reduction in cell population whereas the reduction in galectin-3-expressing cell clones was about 25%, implying an association between galectin-3 expression and chemoresistance to 4HPR [27]. By assessing the apoptotic status of these cell lines with DNA fragmentation assays, they showed that after 2 days of treatment, about 40% of the cells were apoptotic in the inactive galectin-3 cell line and also apoptosis was observed in the galectin-3-negative group, whereas the galectin-3-expressing group was resistant to 4HPR even after 4 days of treatment [27] (Table 1).

Table 1 Results

Study	Year	Institution	Specimen	Drug	Outcomes in sensitivity and resistance
Shiro Akahani et al. [6]	1997	Wayne State University School of Medicine, Detroit, Michigan	Transfected human breast cancer cell line BT549	cis-Diamminedichloroplatinum (CDDP)	Galectin-3-expressing cell clones have increased viability after 72-h exposure to CDDP Galectin-3 inhibits apoptosis after 72-h exposure to CDDP Galectin-3-null and -positive cell clones showed no difference in the level of Bcl-2, Bcl-X _L and Bax protein expressions
Nachman Mazurek et al. [24]	2011	The University of Texas MD Anderson Cancer Center, Houston, Texas	Transfected human breast cancer cell line BT549 (Pro64His polymorphism)	Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) and doxorubicin	TRAIL resistance is associated with the homozygous Pro64 genotype ($P < 0.05$) Sensitivity to doxorubicin is not dependent on galectin-3 genotype Galectin-3 expression significantly increased in MDA-MB-231 cell lines after treatment with ATO ($P < 0.01$)
Hao Zhang et al. [25]	2014	Department of Breast Surgery, the Second Hospital of Dalian Medical University, Dalian, China,	Human breast cancer cell line MDA-MB-231 and MCF7	Arsenic trioxide (ATO)	Galectin-3 knockdown in MDA-MB-231 cell lines increases apoptosis compared to control ($P < 0.05$) Galectin-3 inhibition sensitizes MDA-MB-231 cell lines to ATO-induced apoptosis
Renato Carvalho et al. [26]	2014	Instituto de Biofísica Carlos Chagas Filho, Universidade Federal do Rio de Janeiro, Brazil	Human HEK293FT cell line shGAL3: silenced gal3 cell lines shSCRB: control scramble sequence cell lines	Etoposide, carboplatin, mitomycin C	shGAL3 cells had statistically significant increase in viability when treated with etoposide or carboplatin or mitomycin c in all tested concentrations shGAL3 cells showed up to a 60% increase in viability comparing to controls after treatment with 20 nM of etoposide

Table 1 (continued)

Study	Year	Institution	Specimen	Drug	Outcomes in sensitivity and resistance
Jin-Hyuk Choi et al. [27]	2004	Department of Thoracic/Head and Neck Medical Oncology; The University of Texas M.D. Anderson Cancer Center, Houston, Texas USA	Human breast cancer cell line BT549 BT549Vec: neomycin-resistant control vector BT549Gal-3Mu: BT549 cells expressing mutant galectin-3 BT549Gal-3Wt: BT549 cells expressing wild-type human galectin-3	N-(4-Hydroxyphenyl)retinamide (4HPR)	5 μ M of 4HPR after 24 to 72 h is leading to a time-dependent decrease in survival (70–80% reduction) of controls and those expressing inactive galectin-3. In contrast, cell lines expressing active galectin-3 showed only 25% reduction assuming that galectin-3 was conferring resistance to 4HPR In both BT549Vec and BT549Gal-3Mu, apoptotic cells comprised most of dead cells. On the other hand, most BT549Gal-3Wt cells were viable Both BT549Vec and BT549Gal-3Mu showed a four-fold increase of Intracellular ROS after the first 6 h of exposure to 4HPR. The later applies not to BT549Gal-3Wt cells After a 24-h exposure to 4HPR in both BT549Vec and BT549Gal-3Mu cell lines the levels of Cytochrome c increased implying mitochondrial apoptosis. In BT549Gal-3Wt cell lines 4HPR cytochrome c levels were stable 4HPR treatment led to decreased level of procaspase-3 or PARP cleavage in the control and mutant galectin-3 cell lines whereas in BT549Gal-3Wt cell lines the levels of both proteins remain unchanged

Discussion

cis-Diamminedichloroplatinum (CDDP)

Shiro Akahani et al. showed that galectin-3 expression increases the chemoresistance of human breast carcinoma BT549 cell clones to CDDP and decreases the apoptotic effects of CDDP. The authors concluded that galectin-3 inhibits apoptosis via a cysteine protease pathway and its affinity to BH1 domain of the Bcl-2 protein family [6]. The same results are observed through inhibition of galectin-3 in breast cancer cell lines as well as clear cell ovarian carcinoma and prostate cancer [28, 29].

Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL)

The results of Mazurek et al. are in line with their previously published study showing that expression of galectin-3 in breast cancer cell lines alters sensitivity to TRAIL [30]. Since TRAIL transmits death signals, through death domain-containing receptors, it contributes to apoptosis but also against tumor growth and metastasis [31–33]. On their previous hypothesis, the alteration on TRAIL sensitivity is via upregulation of phosphatase and tensin analog (PTEN) which inactivates the phosphatidylinositol 3-kinase/Akt survival pathway [30]. Based on recent results, the His64/Pro64 polymorphism may alter the galectin-3 apoptotic effects. Furthermore, the His64 genotype but not the Pro64 is associated with decreased sensitivity to cisplatin and doxorubicin [24, 31].

Arsenic trioxide (ATO)

Hao Zhang et al. showed that galectin-3 expression reduces the sensitivity of breast cancer cell lines to treatment with arsenic trioxide and subsequently galectin-3 inhibition sensitizes the same cell lines to ATO-induced apoptosis. Arsenic trioxide is an inorganic compound that can inhibit tumor growth and induce apoptosis in many cancers such as acute promyelocytic leukemia and breast [34, 35]. In this research study, the authors showed that galectin-3 was located in the cytoplasm and was significantly higher expressed in breast cancer cells compared to non-cancer cells. Furthermore, triple-negative breast cancer (TNBC) was expressing predominantly galectin-3 as compared to other subtypes. This finding is in line with other studies in the literature [22, 25]. ATO-induced apoptosis has been proven in many preclinical studies for brain, liver, prostate, renal and breast cancers [36, 37]. It is supported by Hao

Zhang et al. that the mechanism of the apoptotic effects of ATO may include the generation of reactive oxygen species (ROS) based on multiple myeloma studies [38].

Etoposide, carboplatin and mitomycin C

Carvalho et al. showed that silenced galectin-3-expressing cells had an increased resistance and delayed DNA damage repair response when treated with either etoposide, carboplatin or mitomycin C suggesting that galectin-3 has a notable role in DNA damage repair [26]. The authors suggest that galectin-3 is participating in complexes with BARD1 and BRCA1 and that it might regulate the BRCA1/BARD1 heterodimer recruitment to DNA damage sites [26]. This finding is in line with other observations suggesting that DNA damage repair involves G₂/M cell cycle arrest allowing mitosis after DNA repair [39]. The same events have also been described in BRCA1-deficient cells [40]. It is noteworthy that in the majority of the studies, galectin-3 has been associated with its anti-apoptotic characteristics and its induced chemoresistance. However, it seems that galectin-3 has also pro-apoptotic functions similar to BARD1 [41, 42].

N-(4-Hydroxyphenyl)retinamide (4HPR)

In their study, Jin-Hyuk Choi et al. showed that galectin-3 is associated with increased viability and protection against apoptosis in cell lined treated with 4HPR. 4HPR is a synthetic retinoid which has a proven ability to induce apoptosis in a variety of cell lines with hematological cancers, neuroblastomas, prostate, bladder, head and neck and lung cancer [43–48]. Moreover, its beneficial act has been seen and evaluated in clinical studies for breast and ovarian cancer [49, 50]. To identify the cause of this reduction in apoptosis, they investigated different known apoptotic pathways. Regarding the intracellular reactive oxygen species (ROS), they found that 4HPR caused a linear increase of ROS compared to silenced and inactive galectin-3 cell lines. On the contrary, the galectin-3 expressing group only showed a rather small increase in ROS generation [27]. To further investigate whether galectin-3 expression is the cause of the decreased ROS-induced apoptosis, they treated all three cell lines with H₂O₂ and reached the conclusion that galectin-3 expression does not lead to resistance to ROS-induced apoptosis but alters the ability of 4HPR to increase ROS generation. The same impact of galectin-3 expression to ROS generation has been also described after treatment with TNF α [51].

Another pathway investigated by the same group was the Bcl-2 family protein expression which has a well-known association with apoptosis [19]. After treatment with 4HPR, there was a reduction in Bcl-2, Bcl-x_L and Bax in both active and inactive galectin-3-expressing groups. The impact of 4HPR in these proteins in the inactive galectin-3

cell lines was more pronounced than in the silenced lines and the appearance of Bax fragments was only noticed in the silenced group. Since the exact impact of 4HPR to these proteins is not yet identified, the authors suggested that ROS created by 4HPR lead to decreased Bcl-2 expression and Bax fragments. This hypothesis is also supported by other studies suggesting that ROS generation can directly modify the Bcl expression [20, 52, 53].

Finally, they also measured the levels of cytochrome c and activated caspase-3. Since the release of cytochrome c into the cytosol is a sign of mitochondrial apoptosis, they described that levels of cytochrome c increased after the first day of treatment with 4HPR in both silenced and inactive galectin-3 cell lines, whereas in galectin-3-expressing group, cytochrome c levels were not elevated [27, 54]. Regarding the activated caspase-3 levels, the authors found that in both inactive and silenced lines, a decrease in pro-caspase-3 indicates the production of activated caspase-3. In contrast, these results were not confirmed in the galectin-3-expressing group [27]

Conclusion

In the present review, we have listed the published data regarding galectin-3 expression and its association with chemosensitivity and chemoresistance in breast cancer. In the majority of the included studies, the expression of galectin-3 had a protective role in cell survival via different pathways such as the response to DNA damage and repair or the inhibition of apoptosis after treatment with a chemotherapeutic agent. Hence, galectin-3 expression in breast tumors might be an important factor in the selection of the most suitable treatment.

This is, to the best of our knowledge, that first study thoroughly reviewing the effects of galectin-3 to chemosensitivity and chemoresistance in breast cancer. Moreover, the potential clinical impact in treatment selection is considered a strength. The limitations of our review are the relatively small amount of studies included, the lack of clinical studies and the lack of published data for long-term treatment.

However, galectin-3 expression and its impact is a very interesting field for future studies. There are not any published studies thus far regarding the effects of galectin-3 in cell lines treated with taxanes, anthracyclines and cyclophosphamide. This is a potential field for future research.

Author contributions IB: data collection and manuscript writing; AP: data collection and manuscript writing; WB: data management; AL: data management; AH: data review and manuscript editing; SK: data review and manuscript editing; and MS: project development and manuscript editing.

Funding No funding has been given for this study.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

Ethical approval This article does not contain any studies with human participants or animals performed by any of the authors.

References

1. Liu FT, Rabinovich GA (2005) Galectins as modulators of tumour progression. *Nat Rev Cancer* 5(1):29–41
2. Barondes SH, Castronovo V, Cooper DN, Cummings RD, Drickamer K, Feizi T et al (1994) Galectins: a family of animal beta-galactoside-binding lectins. *Cell* 76(4):597–598
3. Takenaka Y, Fukumori T, Raz A (2002) Galectin-3 and metastasis. *Glycoconj J* 19(7–9):543–549
4. Yang RY, Liu FT (2003) Galectins in cell growth and apoptosis. *Cell Mol Life Sci* 60(2):267–276
5. Kim HR, Lin HM, Biliran H, Raz A (1999) Cell cycle arrest and inhibition of anoikis by galectin-3 in human breast epithelial cells. *Cancer Res* 59(16):4148–4154
6. Akahani S, Nangia-Makker P, Inohara H, Kim HR, Raz A (1997) Galectin-3: a novel antiapoptotic molecule with a functional BH1 (NWGR) domain of Bcl-2 family. *Cancer Res* 57(23):5272–5276
7. Hughes RC (2001) Galectins as modulators of cell adhesion. *Biochimie* 83(7):667–676
8. Lord CJ, Ashworth A (2012) The DNA damage response and cancer therapy. *Nature* 481(7381):287–294
9. Knibbs RN, Agrwal N, Wang JL, Goldstein IJ (1993) Carbohydrate-binding protein 35. II. Analysis of the interaction of the recombinant polypeptide with saccharides. *J Biol Chem* 268(20):14940–14947
10. Dumić J, Dabelić S, Flogel M (2006) Galectin-3: an open-ended story. *Biochim Biophys Acta* 1760(4):616–635
11. Gillenwater A, Xu XC, el-Naggar AK, Clayman GL, Lotan R (1996) Expression of galectins in head and neck squamous cell carcinoma. *Head Neck* 18(5):422–432
12. Tsuboi K, Shimura T, Masuda N, Ide M, Tsutsumi S, Yamaguchi S et al (2007) Galectin-3 expression in colorectal cancer: relation to invasion and metastasis. *Anticancer Res* 27(4B):2289–2296
13. Turkoz HK, Oksuz H, Yurdakul Z, Ozcan D (2008) Galectin-3 expression in tumor progression and metastasis of papillary thyroid carcinoma. *Endocr Pathol* 19(2):92–96
14. Kim SJ, Shin JY, Cheong TC, Choi IJ, Lee YS, Park SH et al (2011) Galectin-3 germline variant at position 191 enhances nuclear accumulation and activation of beta-catenin in gastric cancer. *Clin Exp Metastasis* 28(8):743–750
15. Chen YR, Juan HF, Huang HC, Huang HH, Lee YJ, Liao MY et al (2006) Quantitative proteomic and genomic profiling reveals metastasis-related protein expression patterns in gastric cancer cells. *J Proteome Res* 5(10):2727–2742
16. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 68:394–424
17. Honjo Y, Nangia-Makker P, Inohara H, Raz A (2001) Down-regulation of galectin-3 suppresses tumorigenicity of human breast carcinoma cells. *Clin Cancer Res* 7(3):661–668

18. Song YK, Billiar TR, Lee YJ (2002) Role of galectin-3 in breast cancer metastasis: involvement of nitric oxide. *Am J Pathol* 160(3):1069–1075
19. Cory S, Adams JM (2002) The Bcl2 family: regulators of the cellular life-or-death switch. *Nat Rev Cancer* 2(9):647–656
20. Boya P, Morales MC, Gonzalez-Polo RA, Andreau K, Gourdier I, Perfettini JL et al (2003) The chemopreventive agent N-(4-hydroxyphenyl)retinamide induces apoptosis through a mitochondrial pathway regulated by proteins from the Bcl-2 family. *Oncogene* 22(40):6220–6230
21. Moon BK, Lee YJ, Battle P, Jessup JM, Raz A, Kim HR (2001) Galectin-3 protects human breast carcinoma cells against nitric oxide-induced apoptosis: implication of galectin-3 function during metastasis. *Am J Pathol* 159(3):1055–1060
22. Koo JS, Jung W (2011) Clinicopathologic and immunohistochemical characteristics of triple negative invasive lobular carcinoma. *Yonsei Med J* 52(1):89–97
23. Cheong TC, Shin JY, Chun KH (2010) Silencing of galectin-3 changes the gene expression and augments the sensitivity of gastric cancer cells to chemotherapeutic agents. *Cancer Sci* 101(1):94–102
24. Mazurek N, Byrd JC, Sun Y, Ueno S, Bresalier RS (2011) A galectin-3 sequence polymorphism confers TRAIL sensitivity to human breast cancer cells. *Cancer* 117(19):4375–4380
25. Zhang H, Luo M, Liang X, Wang D, Gu X, Duan C et al (2014) Galectin-3 as a marker and potential therapeutic target in breast cancer. *PLoS ONE* 9(9):e103482
26. Carvalho RS, Fernandes VC, Nepomuceno TC, Rodrigues DC, Woods NT, Suarez-Kurtz G et al (2014) Characterization of LGALS3 (galectin-3) as a player in DNA damage response. *Cancer Biol Ther* 15(7):840–850
27. Choi JH, Chun KH, Raz A, Lotan R (2004) Inhibition of N-(4-hydroxyphenyl)retinamide-induced apoptosis in breast cancer cells by galectin-3. *Cancer Biol Ther* 3(5):447–452
28. Wang Y, Nangia-Makker P, Balan V, Hogan V, Raz A (2010) Calcipain activation through galectin-3 inhibition sensitizes prostate cancer cells to cisplatin treatment. *Cell Death Dis* 1:e101
29. Oishi T, Itamochi H, Kigawa J, Kanamori Y, Shimada M, Takahashi M et al (2007) Galectin-3 may contribute to Cisplatin resistance in clear cell carcinoma of the ovary. *Int J Gynecol Cancer* 17(5):1040–1046
30. Mazurek N, Sun YJ, Liu KF, Gilcrease MZ, Schober W, Nangia-Makker P et al (2007) Phosphorylated galectin-3 mediates tumor necrosis factor-related apoptosis-inducing ligand signaling by regulating phosphatase and tensin homologue deleted on chromosome 10 in human breast carcinoma cells. *J Biol Chem* 282(29):21337–21348
31. Balan V, Nangia-Makker P, Schwartz AG, Jung YS, Tait L, Hogan V et al (2008) Racial disparity in breast cancer and functional germ line mutation in galectin-3 (rs4644): a pilot study. *Cancer Res* 68(24):10045–10050
32. Ashkenazi A, Dixit VM (1998) Death receptors: signaling and modulation. *Science* 281(5381):1305–1308
33. Cretney E, Takeda K, Smyth MJ (2007) Cancer: novel therapeutic strategies that exploit the TNF-related apoptosis-inducing ligand (TRAIL)/TRAIL receptor pathway. *Int J Biochem Cell Biol* 39(2):280–286
34. Baj G, Arnulfo A, Deaglio S, Mallone R, Vigone A, De Cesaris MG et al (2002) Arsenic trioxide and breast cancer: analysis of the apoptotic, differentiative and immunomodulatory effects. *Breast Cancer Res Treat* 73(1):61–73
35. Lo-Coco F, Avvisati G, Vignetti M, Thiede C, Orlando SM, Iacobelli S et al (2013) Retinoic acid and arsenic trioxide for acute promyelocytic leukemia. *N Engl J Med* 369(2):111–121
36. Dilda PJ, Hogg PJ (2007) Arsenical-based cancer drugs. *Cancer Treat Rev* 33(6):542–564
37. Chen Z, Chen GQ, Shen ZX, Sun GL, Tong JH, Wang ZY et al (2002) Expanding the use of arsenic trioxide: leukemias and beyond. *Semin Hematol* 39(2 Suppl 1):22–26
38. Berenson JR, Yeh HS (2006) Arsenic compounds in the treatment of multiple myeloma: a new role for a historical remedy. *Clin Lymphoma Myeloma* 7(3):192–198
39. Poehlmann A, Roessner A (2010) Importance of DNA damage checkpoints in the pathogenesis of human cancers. *Pathol Res Pract* 206(9):591–601
40. Yarden RI, Pardo-Reoyo S, Sgagias M, Cowan KH, Brody LC (2002) BRCA1 regulates the G2/M checkpoint by activating Chk1 kinase upon DNA damage. *Nat Genet* 30(3):285–289
41. Tembe V, Henderson BR (2007) BARD1 translocation to mitochondria correlates with Bax oligomerization, loss of mitochondrial membrane potential, and apoptosis. *J Biol Chem* 282(28):20513–20522
42. Yu F, Finley RL Jr, Raz A, Kim HR (2002) Galectin-3 translocates to the perinuclear membranes and inhibits cytochrome c release from the mitochondria. A role for synexin in galectin-3 translocation. *J Biol Chem* 277(18):15819–15827
43. Oridate N, Lotan D, Xu XC, Hong WK, Lotan R (1996) Differential induction of apoptosis by all-trans-retinoic acid and N-(4-hydroxyphenyl) retinamide in human head and neck squamous cell carcinoma cell lines. *Clin Cancer Res* 2(5):855–863
44. Zou CP, Kurie JM, Lotan D, Zou CC, Hong WK, Lotan R (1998) Higher potency of N-(4-hydroxyphenyl) retinamide than all-trans-retinoic acid in induction of apoptosis in non-small cell lung cancer cell lines. *Clin Cancer Res* 4(5):1345–1355
45. Zou C, Liebert M, Zou C, Grossman HB, Lotan R (2001) Identification of effective retinoids for inhibiting growth and inducing apoptosis in bladder cancer cells. *J Urol* 165(3):986–992
46. Maurer BJ, Metelitsa LS, Seeger RC, Cabot MC, Reynolds CP (1999) Increase of ceramide and induction of mixed apoptosis/necrosis by N-(4-hydroxyphenyl)—retinamide in neuroblastoma cell lines. *J Natl Cancer Inst* 91(13):1138–1146
47. Sun SY, Yue P, Lotan R (1999) Induction of apoptosis by N-(4-hydroxyphenyl) retinamide and its association with reactive oxygen species, nuclear retinoic acid receptors, and apoptosis-related genes in human prostate carcinoma cells. *Mol Pharmacol* 55(3):403–410
48. Delia D, Aiello A, Lombardi L, Pelicci PG, Grignani F, Grignani F et al (1993) N-(4-hydroxyphenyl) retinamide induces apoptosis of malignant hemopoietic cell lines including those unresponsive to retinoic acid. *Cancer Res* 53(24):6036–6041
49. De Palo G, Mariani L, Camerini T, Marubini E, Formelli F, Pasini B et al (2002) Effect of fenretinide on ovarian carcinoma occurrence. *Gynecol Oncol* 86(1):24–27
50. Wang TT, Phang JM (1996) Effect of N-(4-hydroxyphenyl) retinamide on apoptosis in human breast cancer cells. *Cancer Lett* 107(1):65–71
51. Matarrese P, Tinari N, Semeraro ML, Natoli C, Iacobelli S, Malorni W (2000) Galectin-3 overexpression protects from cell damage and death by influencing mitochondrial homeostasis. *FEBS Lett* 473(3):311–315
52. Zhou LJ, Zhu XZ (2000) Reactive oxygen species-induced apoptosis in PC12 cells and protective effect of bilobalide. *J Pharmacol Exp Ther* 293(3):982–988
53. Lu HF, Hsueh SC, Ho YT, Kao MC, Yang JS, Chiu TH et al (2007) ROS mediates baicalin-induced apoptosis in human promyelocytic leukemia HL-60 cells through the expression of the Gadd153 and mitochondrial-dependent pathway. *Anticancer Res* 27(1A):117–125
54. Hail N Jr, Lotan R (2001) Mitochondrial respiration is uniquely associated with the prooxidant and apoptotic effects of N-(4-hydroxyphenyl) retinamide. *J Biol Chem* 276(49):45614–45621

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.