



Prodrugs as drug delivery system in oncology

J. Delahousse^{1,2} · C. Skarbak¹ · A. Paci^{1,2,3}

Received: 27 February 2019 / Accepted: 5 July 2019 / Published online: 7 August 2019
© Springer-Verlag GmbH Germany, part of Springer Nature 2019

Abstract

The use of conventional chemotherapy in the treatment of cancer has been restricted by the lack of cell specificity, which causes toxicity regarding healthy cells resulting in limiting side effects responsible for low therapeutic efficiency. To overcome these drawbacks, the design of prodrugs has evolved and improved by covalently linking the drug through a degradable spacer. The use of these prodrugs as drug delivery systems, which are able to inactivate the drug during its biodistribution to specifically deliver the drug to its target, is an important breakthrough in cancer therapy. This strategy consisting in the covalent binding of a promoiety to daily used therapeutic compounds has been clinically proven in the design of targeted prodrugs leading enhanced therapeutic efficacy and increase of the therapeutic index. This review summarizes and compares several strategies that improve the therapeutic index of chemotherapy (i.e. conventional drugs) by their chemical transformation into prodrugs improving pharmacokinetic profiles and optimizing administration routes in comparison to the initial drug. This review provides an overview of the methods used to control the structure and function of prodrugs and, ultimately, their current and future potential in increasing the therapeutic index of daily used anticancer drugs. First, prodrugs' design and their activation within the tumor microenvironment or within the tumor cell will be exposed. Then, the different strategies used leading to these prodrugs will be presented.

Keywords Cancer therapy · Drug delivery systems · Microenvironment · Prodrugs · Targeting

Targeting for efficient care

Prodrug strategy

Chemotherapy (CT) has demonstrated important curative rates; nevertheless, the lack of specificity is still an issue as CT is responsible for side-effects especially towards fast dividing healthy cells. CT drugs act through an antiproliferative mechanism or by disruption of the cell cycle at a

specific phase. Hence, these drugs, due to poor specificity, affect rapidly proliferating and dividing cells such as red blood cells, hair follicles, gut epithelial cells, bone marrow, and the lymphatic system, leading to numerous side-effects which limit their uses. High-dose CT is generally required to effectively inhibit tumor proliferation, especially in resistant solid tumors. The lack of specificity of CT could lead to lethal damages of the adjacent normal proliferating cells, resulting in a discontinuity of therapy. Therefore, improving the specificity of CT drugs used daily is a vital step to increase their therapeutic index. Many strategies have been developed for this purpose. Among them, a relevant strategy is based on the development of prodrugs. Initially, a prodrug is an inactive drug designed to be activated through a spontaneous process (e.g. hydrolytic degradation) or through a biocatalytic mechanism, near the pharmacological target allowing a specific release of the active entity at its target site [1].

Their design is commonly based on three components (Fig. 1): (1) the active moiety for anticancer activity (i.e. drug); (2) a cleavable labile chemical moiety (i.e. linker) which is conjugated to the active moiety through a functional

J. Delahousse and C. Skarbak have contributed equally to the present study.

✉ C. Skarbak
charles.skarbak@gmail.com

¹ Vectorologie des anticancéreux et des acides nucléiques, UMR 8203, CNRS, Université Paris-Saclay, Gustave Roussy Cancer Campus, Villejuif, France

² Service de Pharmacologie, Département de Biologie et Pathologie médicales, Gustave Roussy Cancer Campus, Villejuif, France

³ University Paris-Saclay, Faculté de Pharmacie, Chatenay-Malabry, France

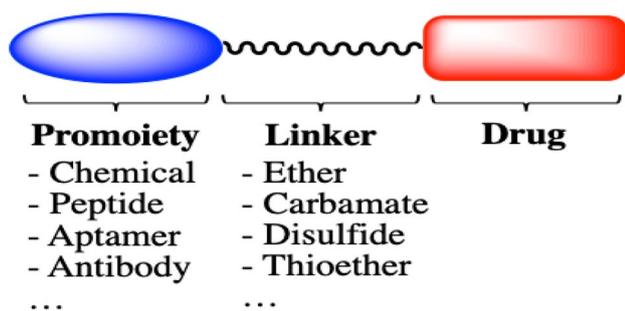


Fig. 1 Common structure of a prodrug

group (hydroxyl, carboxylic, amine, carbonyl, and phosphate groups, etc.); and (3) a targeting moiety for specific delivery to tumor cells such as polypeptides or monoclonal antibodies (mAb) (i.e. promoiety) [2].

The design of such compounds depends on two major principles: (1) the structural features of the active moiety with the ability of the targeted moiety to mask the pharmacodynamic activity of the anticancer drug. (2) The bioconversion mechanism of the drug release which can either gradually or immediately degrade the linker chosen for the conjugation [3]. The design of prodrugs provides a remarkable tool to improve the pharmacological properties of the active moiety. Conventional prodrug design aims mostly to: improve solubility in water or lipid membrane, improve chemical stability, increase oral or local absorption and brain permeability, mask unacceptable taste, reduce irritation or pain, modify pre-systemic metabolism, and reduce toxicity [4].

Paracelsus said some 500 years ago: “All things are poison and nothing is without poison, only the dose permits something not to be poisonous”. In the sixteenth century, Paracelsus initiated the shift from conventional medicine to modern medicine based on chemical knowledge for a better development of pharmaceuticals. For a long time, the galenic development of drugs has been limited to physicochemical stability concerns to be stored and administered. Better understandings on the fate and behavior of the drugs in the body (pharmacokinetics) have allowed designing “clever” drugs. In the beginning of the twentieth century,

a German scientist named Paul Ehrlich thought that a compound could target selectively a disease-causing organism without harming the body itself and called the hypothetical agents “Zauberkegel”, the “Magic Bullet” [5]. His idea has been extended to many drugs used in various diseases including malignant diseases. The dream of Paul Ehrlich is becoming true towards new vectorization concepts, with the development of new physicochemical concepts, new therapeutic targets, and new biomaterials. We are now faced with more than 110,000 publications (Pubmed) in the field of drug delivery systems. Today, controlling the duration and the localization of the drug release for optimized exposure is a reality (Fig. 2).

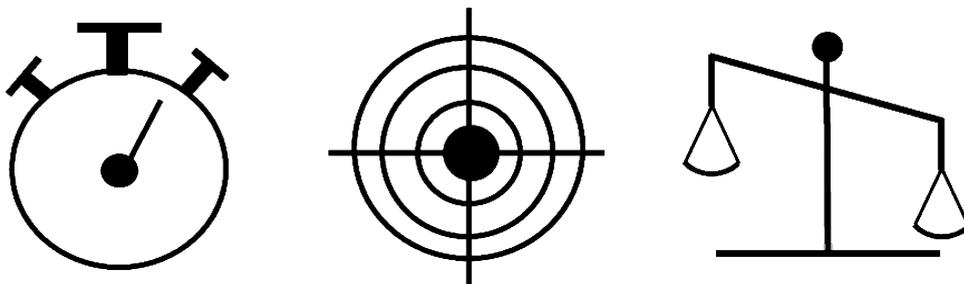
Temporally controlled drug delivery systems (TCDDS)

Controlling the chronology of drug release displays endpoints: (1) delay the drug release in order to reach the right tissue based on the pharmacokinetic and (2) increase the drug exposure by controlling its release (diffusion through a structure, erosion, etc.) and degradation/elimination process (decreasing the renal or hepatic clearance, protecting the drug from biologic events). Nowadays, drug release can be modulated through innovative structure, such as lipidic or polymeric scaffolds, sensible to biologic or external stimuli, and benefit from new knowledge of the specific microenvironment of the target.

Spatially controlled drug delivery systems (SCDDS)

In oncology, CT uses are limited because of their off-target toxicities; CT agents act by inhibiting the cellular division and, by consequence, have drastic effects on healthy cells, especially on cells with rapid cell divisions such as bone marrow cells, cells from the digestive tract and hair follicles. In addition, some cytotoxic agents have a specific toxicity profile such as platinum (Pt) with nephro-/ototoxicity, doxorubicin (DOX) with cardiotoxicity, leading to interruptions or cessation of treatment. DDS are now able to control the drug release to target a specific physiologic compartment

Fig. 2 Control of duration and localization for improved quantity release to the target



which could be a tissue involved in the disease. This strategy hugely improves the risk/benefit balance.

The strategy aiming to design new spatially controlled drug delivery systems (SCDDS) can be of interest in the targeting of specific organs or compartment and can be divided into: (1) the passive targeting with an indirect targeting focused on tumor microenvironment and (2) the active targeting by a direct targeting of the specific proteins or surface marker of tumor cells. It is necessary to keep in mind two characteristics of cancer cells related to targeting. First, the majority of cancer cells express surface molecular targets such as ligand, also present in healthy cells at lower levels. Secondly, the microenvironment of the cancer cells represents significant limitations for drug penetration especially in large solid tumors.

Quantity release-controlled drug delivery systems (QRCDDS)

Galenic developments play a role in the drug release at the right place and at the right time as well as the right delivered quantity. Therapeutic concentrations are a major concern especially in anticancer drugs. High cytotoxic exposure can cause collateral damages and poor exposure can lead to treatment failure. DDS provides the opportunity to control and optimize blood and/or tumor concentration and thus an increased therapeutic index (efficacy/toxicity balance). Besides, unfavorable physicochemical characteristics of some CT limited the effective doses, for example low water solubility for paclitaxel. DDS strategies allow developing structures to improve physicochemical properties.

Prodrugs design

The term “prodrug” was first introduced by Albert in 1951; he defined a prodrug as “a molecule which does not have any intrinsic biological activity but which is capable, during the different phases of its metabolism, to generate a biologically active drug” [6]. Ideally, the prodrug should convert to the active drug as soon as the target is reached concomitantly with the release of the non-cytotoxic moiety followed by its subsequent elimination. This conversion can be obtained

after an enzymatic or chemical degradation *in vivo*. According to the linked moiety, self-assembling properties can be observed giving access to a passive targeting. This new property leads to an increased specificity for the target site decreasing the exposure regarding normal cells and tissue which leads to an improved therapeutic index of the parent active drug.

Functional linker for an optimal release

The design of prodrug relies on the choice of the right functional group present between the drug and the promoiety. This functional group should be self-immolative or enzymatically/chemically cleavable so that the active drug can be liberated spontaneously or under triggerable condition. This process takes place either in the tumor microenvironment: such as in the presence of an overexpressed enzyme, a change in the pH, the presence of a characteristic stimulus, or at the surface of the tumor cell using a specific ligand–receptor combination. The promoiety linked to the active moiety (drug) plays a key role in the improvement of the drug-like properties and in overcoming physiological barriers or in the guidance of the drug to its target cells. The linker between the active moiety and the promoiety should be stable enough to remain intact in the blood circulation until it reaches the target cells. Optimization of the conjugation is highly suggested during the prodrug design to achieve the optimal stability and efficacy. Commonly, only a few functional groups are used for the design of prodrugs (Table 1).

Ester functional groups are among the most used as they are most easily cleaved by a large number of esterase in biological environments. They are usually introduced by the conjugation using charged groups such as hemi-esters, sulfates, phosphates or their salts, thus creating a variety of links. These functional groups can be readily hydrolyzed to release effective molecules by ubiquitous esterase, phosphatases, and sulfatases in the blood, liver, and other organs and tissues [7]. Depending on the differences in the structure of the prodrug or of the environmental conditions, the half-life of the ester bond can vary from several minutes to several hours according to accessibility of the bond [8].

Table 1 Commonly found linkers in prodrug conjugation strategy

| Ester | | | | | Amide | | Other linkers | | |
|----------------|-----------------|-----------------|-----------------|---------------|--------------|-------|---------------|-----------|-----------|
| Carboxyl ester | Carbamate ester | Carbonate ester | Phosphate ester | Sulfate ester | Peptide bond | Oxime | Imine | Disulfide | Thioether |
| | | | | | | | | | |

Amide group is usually designed to enhance oral absorption of drugs with amine and carboxylic functionalities. It has a relatively higher enzymatic stability than ester bond. Most of the amide bonds are stable for several hours or even several days in the plasma in the absence of specific enzymes [9]. However, the use of an amide linker in prodrugs is designed to be cleavable by a specific enzyme to increase the target-ability or reduce the toxicity.

In addition to ester and amide bonds, several other types of linkers including oximes/imines, disulfide bond or uncleavable thioether and ether bond, have also been used as functional groups [10].

Other functional groups have recently been proposed. Self-immolative linkers have significantly broadened the synthetic scope with regard to chemical functionalities amenable for prodrug design. These new types of linker are designed to rely on the cleavage of the promoiety and the active moiety with the activation of a precursor. This chemical activation generates a cascade of disassembling reaction leading to the spontaneous and fast release of the active moiety drug (Fig. 3) [11].

Self-assembling concept

Chemical modifications can also provide new physicochemical properties to the compound such as self-assembling property leading to prodrug-based nanoparticle drug delivery systems (PBN-DDS) [13]. These PBN-DDS could lead to a dual targeting property, by integrating prodrug strategy and nanomedicine strategy into one same drug delivering system. PBN-DDS have many advantages; on the one hand, prodrugs are inactive drugs and become active only after a specific metabolization or after a chemical reaction at the active site. On the other hand, the nanoparticle system plays the role of cargo resulting in the protection and guidance of the prodrug to the active site. As for today, PBN-DDS can be divided into two major groups according to the characteristic of the material used for its design which can be: (1) high weight polymeric

structures; or (2) small molecular weight PBN-DDS. Each category corresponds to the conjugation of the drug to a polymeric block or a small molecular weight compound, respectively. The physicochemical property of the moiety enables the PBN-DDS to self-assemble into nano-object in aqueous media.

These PBN-DDS refer to amphiphilic conjugates in which the carrier material and the drug play an important role in the hydrophilic–lipophilic balance of the prodrug. These prodrugs are designed by the conjugation of the carrier material directly to the anticancer drug by a covalent bond. The resulting nano-object provides the protection of the prodrug from degradation and diminishes the premature burst release. In addition, the nanoparticulate form provides an increased internalization of the drug leading to an increase of the intracellular drug concentration [14]. The first generation of PBN-DDS describes a drug delivery system that targets the lesion through passive mechanism using the so-called “Enhanced Permeability Retention” (EPR) effect proposed by Maeda [15]. The EPR effect is a phenomenon due to the anatomical and physiological characteristics of tumors. In contrast to normal tissues and organs, most tumors show a higher vascular density, such as in the case of angiogenesis which is one of the most important features of tumors to sustain their uncontrolled and rapid growth. Important differences in the structure between tumor vessels and normal blood vasculature are observed as normal cell vascularization is characterized by joined cells. On the contrary, tumor vessels are formed by non-binding cells leading to small gaps where PBN-DDS auto-agglutinate; this phenomenon characterizes the EPR effect [16]. Even if the EPR phenomenon is widely documented, controversies are discussed. The tumor physiology outlined three points: (1) heterogeneous blood vasculature which can be totally disorderly with loops and shunts leading to tight supplies, (2) elevated interstitial fluid pressure due to the plasmatic protein leakage as well as macromolecules which can block the drug uptake or promote its exclusion, and (3) distance into tumor interstitium for the macromolecules transport [17].

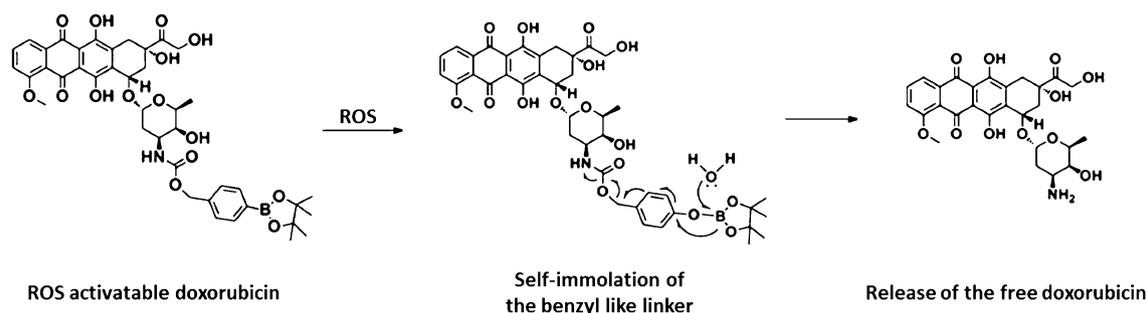


Fig. 3 Example of a ROS activatable self-immolative doxorubicin prodrug [12]

Surface modulation of this PBN-DDS is frequently observed by the use of poly(ethylene) glycol chains (PEG) which leads to “stealth” PBN-DDS with substantially increased circulation time and more likely an increased passive tumor activity (second generation) [18, 19]. The evolutions towards the third generation of PBN-DDS correspond to a dual PBN-DDS with the compilation of different functional moieties such as the drug and a ligand moiety specific for a tumor antigen/receptor (Fig. 4) [20].

Polymer PBN-DDS

This class corresponds to the covalent conjugation of drugs to polymer blocks. In the field of anticancer therapeutics, various polymer blocks have been used to design polymer PBN-DDS, such as poly-lactic-*co*-glycolic acid (PLGA), poly-lactic acid (PLA), poly-glutamic acid (PGA), polycaprolactone (PCL), *N*-hydroxypropyl-methacrylate copolymers (HPMA), and poly-amino acids. Among them, the mostly used are PLA, PLGA and PGA due to their favorable biocompatibility and biodegradability properties [21]. The use of these polymer blocks led to the development of various polymer PBN-DDS which afford different properties, such as water solubility optimization and self-assembling properties. Moreover, the choice of the optimal polymer is essential as it drives the final structure of the skeleton which plays an important role in the final physicochemical properties of the obtained nanosystem. Linear polymer blocks are usually used to synthesize self-assembling PBN-DDS polymer due to their low drug loading. Regarding the use of branched polymer blocks, they offer more functional

groups and enable to increase the conjugation of numerous drugs on the same polymer but leads to an increase of the drug loading. However, the increase of the drug conjugation leads to an important modification of the skeleton which can result in the loss of the self-assembling property. In addition, branched polymers exhibit high steric hindrance which can be critical for cellular internalization and might often result in a delayed cellular entry [22]. The conjugation of paclitaxel to PLA (CT-2103) led to the development of a polymer PBN-DDS which has been studied in phase II trial for the treatment of ovarian, breast and lung cancer. Unfortunately, the study showed no increased overall survival in terms of efficacy compared to the control treatment [23]. More complex polymer can also be used such as polysaccharides which are interesting regarding their physicochemical and biological properties. Chitosan, for example, is an important polysaccharide which is biodegradable, biocompatible, showing antibacterial, antifungal and antioxidant properties. Nevertheless, the conjugation of drugs to chitosan is mainly fulfilled in the antimicrobial or antifungal research area and not in the cancer research [24, 25].

Small molecular weight PBN-DDS

Unlike polymer PBN-DDS, small molecular weight PBN-DDS use small molecules. Some of the most commonly used are polyethylene glycol, hyperbranched poly(ether-ester) (< 5 kD) or squalene [26]. The main advantage of these conjugates is their small size (lower than 250 nm) and their interesting higher drug loading efficiency. Among them, the ones using squalene or polyisoprenoid moieties have generated expanding attention in recent years [27]. Furthermore, to deceive the short circulation time in the blood stream, poor solubility and structural stability in time issues, the co-precipitation in the presence of amphiphilic long-chain PEG copolymers can be fulfilled to achieve an increased circulation time and a higher structural stability [28]. The liberation of the drug can be triggered by a variety of stimuli and many researchers have studied approaches to design stimuli-sensitive nanosystems by applying various specific and/or sensitive linkers. These stimuli can be endogenous redox, enzyme and pH or exogenous, such as light, ultrasound, magnetic fields or temperature [29].

The stimuli-responsive behavior can be achieved by grafting these moieties to the appropriate anticancer drug through cleavable bonds or using gatekeepers, which are ruptured in response to the specific stimuli. In this case, the nanosystem transports the drug to the target tissue and, once there, the presence of a particular stimulus will trigger the release of the trapped drug achieving better control over the administered dose and bringing specificity for the target [30].

To increase the specificity, different target can be defined. Among them, we have chosen to present the strategies used

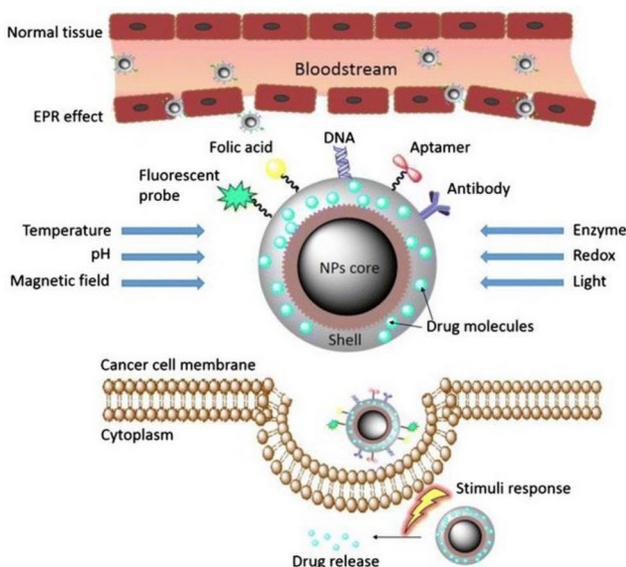


Fig. 4 Illustration underlining examples and targets of third-generation PBN-DDS taken from [20]

to target either (1) the tumor microenvironment taking into account the general surrounding of the tumor (passive targeting), and, (2) the tumor cell more specifically by taking advantage of the ligand receptor interaction (active targeting) (Fig. 5).

Targeting strategies using prodrugs

Enzyme-specific release

One of the targeting strategies takes advantage in the elevated amounts of specific enzymes in the tumor tissues. The most common elevated enzymes are from the proteases family including lysosomal proteases, such as cathepsins, and proteases present in the extracellular matrix, such as matrix metalloproteases (MMP) (Table 2).

Matrix metalloproteinase

A methotrexate (MTX) prodrug has been developed by conjugation with a MMP-2 and MMP-9-cleavable peptide (PVGLIG). This prodrug was combined also with dextran in order to take advantage of the EPR effect. This strategy has demonstrated a better therapeutic index for the prodrug

compared to MTX alone in fibrosarcoma and glioblastoma xenografted mouse models [31]. Other research teams have investigated this strategy, i.e. PVGLIG peptide with paclitaxel [32]. However, to date, no prodrug with this linker has reached clinical stage.

Prostate-specific antigen

Prostate-specific antigen (PSA) is a chymotrypsin-like protease overexpressed and secreted in the extracellular fluid of the prostate cancer microenvironment. Taking advantage of the high PSA level is one of the targeting strategies in prostate cancer research.

Denmeade et al. have synthesized a prodrug of DOX conjugated to a peptide (L-377202) specifically cleavable by PSA in prostate tumor tissue. The release of the active metabolites Leu-DOX and DOX in the prostate gland led to a localized antitumor action [33]. L-377202 has shown a good tolerance in patients at equivalent dose on a molar basis (NCT00987753) [34]. Nevertheless, the clinical phases II and III of the present clinical trial have never raised. Other cytotoxic drugs (paclitaxel, nitrogen mustard, vinblastine and methotrexate) have also been conjugated with PSA-activated peptide [35]. Among them, PSA-activated vinblastine prodrug has shown an excellent antitumor efficacy

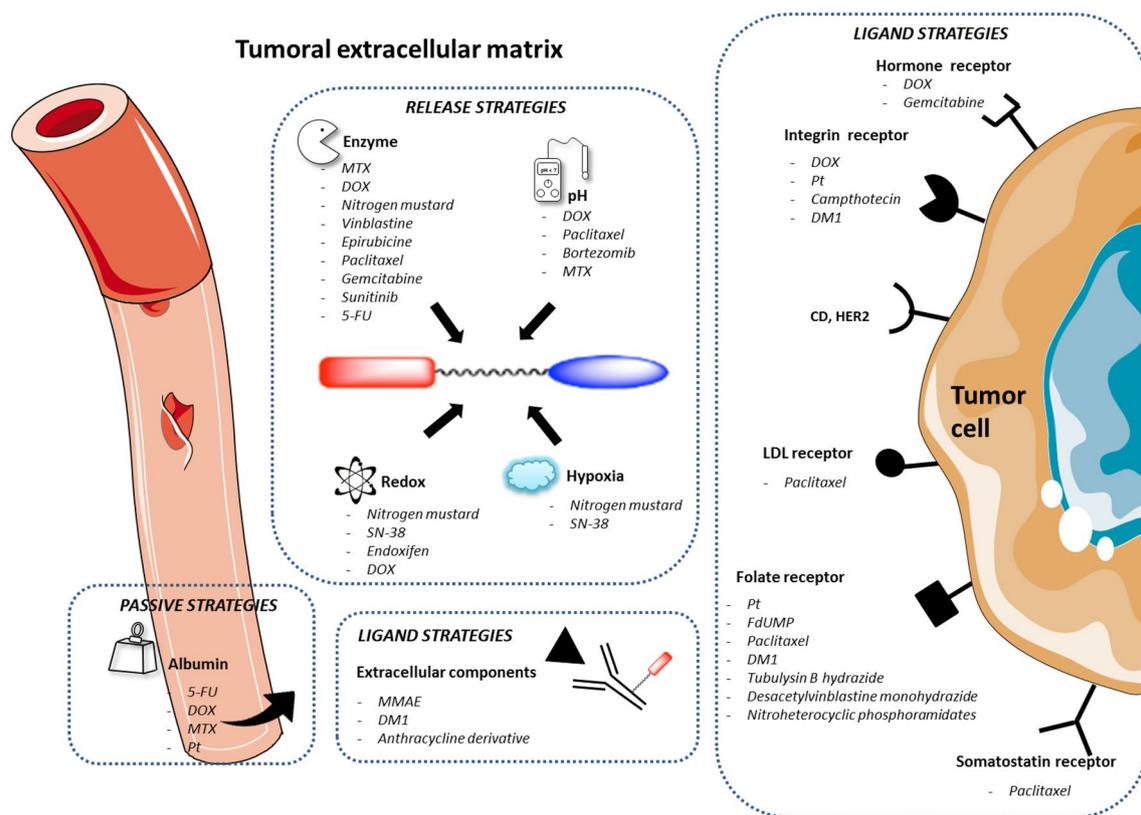


Fig. 5 Use of the tumor microenvironment or the tumor cell as a targeting strategy (Servier Medical Art)

Table 2 Enzyme-specific prodrugs

| Target | Parent drug | Linker | Activity | References |
|-------------------------------|---|---|---|------------|
| Matrix-metalloproteinase | MTX | Matrix-metalloproteinase specific peptide: Pro-Val-Gly-Leu-Ile-Gly (PVGLIG) | In vivo efficacy on Glioblastoma, fibrosarcoma and bladder carcinoma xenografted tumor | [31] |
| Prostate specific antigen | DOX | Gly-Ile-Val-Gly-Pro-Leu (GIVGPL) PSA-specific peptide: His-Ser-Ser-Lys-Leu-Gln-Leu | Enzymatic activation validation, in vitro activity on a panel of prostate cancer cell line | [33] |
| Cancer associated fibroblasts | Nitrogen mustards Vinblastine Vinblastine | PSA-specific peptide: Glu-Hyp-Ala-Ser-Chg-Gln | Validation of in vitro proof of concept | [35] |
| | | PSA-specific peptide: Hyp-Ser-Ser-Chg-Gln-Ser-Ser-Pro | In vivo proof of concept on tumor bearing mice | [36] |
| | | Fibroblast protein specific peptide: Gly-Pro | In vivo efficacy on a panel of xenografted tumor (lung, liver and breast cancer) | [38] |
| Cathepsin B | DOX | Fibroblast protein specific peptide: Gly-Pro | Evaluation of the toxicological profile and the safety pharmacological property of FAP α -targeting prodrug of doxorubicin in vitro and in vivo | [39] |
| | Epirubicin | Fibroblast protein specific peptide: Gly-Pro | Similar in vivo efficacy and no visible cardiotoxicity side effects compared to free EPI | [40] |
| Cathepsin B | DOX | Ac-Phe-Lys: dipeptide specific for cathepsin B Cathepsin B specific peptide: Gly-Phe-Leu-Gly | In vivo efficacy and toxicity study | [42] |
| | Paclitaxel | Polyglutamate residue specifically digested by Cathepsin B | Phase II study in the treatment of breast, non-small cell lung and colorectal cancer | [44] |
| Thymidine phosphorylase | Gemcitabine | Cathepsin B-sensitive maleimide linker | Phase III trial in non-small cell lung and ovarian cancer | [45] |
| | Sunitinib | Cathepsin B specific peptide: Phe-Lys | In vivo efficacy superiority compared to gemcitabine | [67] |
| | Vedotin | cathepsin B specific peptide: Val-Cit | In vitro and ex vivo Validation cathepsin B cleavage | [48] |
| | 5-FU | Fluoropyrimidine carbamate moiety | Approved in refractory or recurrent Hodgkin's lymphoma and large cell anaplastic lymphoma, primary cutaneous large cell anaplastic lymphoma, Hodgkin's lymphoma stage III or IV | [47] |
| Endopeptidase | DOX | Conjugation of albumin | Increased safety and efficacy compared to free 5-FU in more several tumor grafted models | [56] |
| | | Endopeptidase specific peptide: Glu-Hyp-Ala-Ser-Chg-Gln-Ser-Leu | Improvement of the circulation in blood stream and better PK/PD properties | [59] |
| Legumain | DOX | Legumain specific peptide: Ala-Ala-Asp-Leu peptide | Phase I trial led to showing a good tolerable and safe prodrug with partial response in patient with soft-tissue sarcoma A phase II is recommended | [61] |
| | | | In vivo efficacy and improved therapeutic index were observed | [68] |

with slight side-effects on prostate human tumor xenograft and dogs [36].

Fibroblast activation protein α

Targeting tumor cancer-associated fibroblasts (CAFs) is also a strategy in order to release the cytotoxic drug into the tumor environment. Fibroblast activation protein α (FAP α) is an enzyme overexpressed by CAFs as well as in pericytes in human epithelial cancers [37]. Recently, Chen et al. have developed a FAP α -activated vinblastine prodrug by conjugation of the peptide Gly-Pro (Z-Gly-Pro), in order to overcome resistance to vascular disrupting agents through pericytes targeting, which has shown efficacy in tumor xenografts of carcinomas [38]. DOX was also tested in combination with the FAP α -cleaved peptide Z-Gly-Pro; the solubility of DOX was enhanced and the first clinical results in mouse and dog models have shown a reduced toxicity compared to DOX with a comparable antitumor efficacy [39]. The prodrug Z-Gly-Pro epirubicin also has shown a better therapeutic index in mouse model [40]. However, no clinical trials are ongoing for this kind of prodrugs.

Cathepsin B

Cathepsin B is a cysteine protease present in the lysosome as well as in the extracellular matrix. Its overexpression is correlated with invasive and metastatic cancers [41]. Regarding cathepsin B-cleavable tumor-targeting prodrug, Shao et al. have designed a cathepsin B-activated DOX prodrug, Ac-Phe-Lys-PABC-ADM [42], enhancing efficacy and reduced side-effects in gastric cancer peritoneal carcinomatosis mouse model. Other cathepsin B-activated prodrugs of DOX (PDOX) have been developed with a different linker, Gly-Phe-Leu-Gly. PDOX has shown a similar antitumor activity but reduced toxicities in carcinoma-bearing rabbits [43]. Another DOX prodrug made of [*N*-(2-hydroxypropyl) methacrylamide] copolymer and DOX linked by a tetrapeptide spacer cleavable by lysosomal cathepsins (FCE28068 or PK1) has been evaluated in patients with non-small cell lung, breast and colorectal cancer. PK1 takes advantage of the EPR effect and specific cleavage. Moreover, PK1 has shown a good tolerance and efficacy in phase II clinical trial [44]. Paclitaxel prodrug was also developed with the dual strategy “EPR and cathepsin B-cleavable linker” with combination of paclitaxel between degradable polymers made of L-glutamic acid residues (PPX for Paclitaxel polyglumex). PPX has shown in phase III in patients with non-small cell lung cancer a similar efficacy compared docetaxel but an increased neurotoxicity and lower neutropenia and alopecia [45]. Under the trade name Opaxio[®], PPX has been withdrawn of the application for marketing authorization by the

European Medicines Agency (EMA) in 2009 because of the neurotoxicity.

More recently, a multifunctional tumor-targeting cathepsin B-sensitive gemcitabine prodrug has been developed to take advantage of the specific cleavage by cathepsin B in the tumor cell as well as the *in vivo* covalently bound with the circulating albumin. This dual strategy has shown a better antitumor efficacy in mouse breast tumor xenografted model [46]. Finally, a new ADC obtained by the conjugation of Brentuximab, a mAb targeting specifically CD30 surface receptors on the surface of activated B and T cells, to the antimetabolic agent vedotin was designed. The conjugation is fulfilled through a valine–citrulline linker specifically cleaved inside the lysosome by lysosomal cysteine proteases such as cathepsin B. This ADC is approved in refractory or recurrent Hodgkin’s lymphoma and large cell anaplastic lymphoma, primary cutaneous large cell anaplastic lymphoma, Hodgkin’s lymphoma stage III or IV and is commercialized under the trade name Adcetris[®] [47].

In addition to the work on CT, developments are also being made on tyrosine kinase inhibitors, such as sunitinib, the VEGFR inhibitor. Sunitinib prodrug was prepared by conjugation of sunitinib to a cathepsin B cleavable dipeptide. Preliminary *in vitro* data have shown similar efficacy compared to sunitinib alone; *in vivo* investigations are needed to appreciate the potential increased therapeutic index [48].

This strategy seems to be promising but no prodrugs have gone through the phase III of clinical trial.

Thymidine phosphorylase and others

The enzyme thymidine phosphorylase is necessary to convert fluoropyrimidines-based prodrug such as 5-fluorouracil (5-FU) into the antimetabolite agent. This enzyme is overexpressed in tumor cells, which leads to an increase of the fluoropyrimidine-based prodrug concentration into the tumor tissue [49]. 5-FU is a cytotoxic agent that has been widely investigated in order to improve its therapeutic index. Briefly, 5-FU is administered by intravenous injection (IV) due to an unpredictable gastrointestinal absorption and a fast degradation. To overcome the drawbacks of IV injection, such as infection risks, oral prodrugs of 5-FU have been developed. The first oral 5-FU prodrug, tegafur, was developed in 1967 [50] leading to the release of 5-FU after hepatic biotransformation (mainly by CYP2A6), but has shown neurotoxicities and is used in association with uracil to overcome these side-effects [51]. Tegafur has also been integrated in a therapeutic cocktail (S-1 cocktail) with an inhibitor of dihydropyrimidine dehydrogenase (DPD) to increase the 5-FU half-life, and, an inhibitor of 5-FU phosphorylation in the gastrointestinal tract to decrease the gastrointestinal toxicities. S-1 cocktail has shown a high efficacy but also high incidence of side-effects during the

phase II trial [52]; nevertheless it has been approved for the treatment of advanced gastric cancer by the EMA in 2011 (Teysuno®) but not by the Food and Drug Administration (FDA). Another oral prodrug has been developed taking advantage of metabolic pathway enabling the liberation of 5-FU to decrease the dose limiting due to hematotoxicity, the doxifluridine. The pyrimidine phosphorylase required for the transformation of doxifluridine into 5-FU is practically absent in the bone marrow [53]. A Japanese study did not show any significant advantages of doxifluridine over 5-FU in rectal cancer in clinic [54]. Besides, doxifluridine has been also studied with intravenous injections. Nevertheless, the phase II clinical trial of doxifluridine in patients with advanced ovarian cancer has shown toxic deaths and severe neurotoxicities; doxifluridine could pass through the blood–brain barrier more easily than 5-FU [55]. Capecitabine (Xeloda®), a derivative of doxifluridine, is not metabolized by intestinal cells with a nearly complete absorption and concentrates mostly into tumor cell to give 5-FU by metabolism through a cascade reaction of three enzymes with the last step by thymidine phosphorylase [56]. Nowadays, capecitabine is widely used in antitumor treatments. However, a recent meta-analysis has concluded no difference in overall survival and progression-free survival between 5-FU, capecitabine and S-1 in advanced gastric cancer, while lower toxicities with S-1 were observed [57]. Fukushima et al. have developed DFP-11207, which is a combination with precursor form of 5-FU (1-ethoxymethyl-5-FU), an inhibitor of 5-FU degradation (5-chloro-2,4-dihydroxypyridine) and an inhibitor of 5-FU phosphorylation (citrazinic acid). Pharmacokinetic study on rats has shown, after an oral administration, a lower C_{max} and longer half-life compared to the other 5-FU prodrugs, and a similar efficacy of 5-FU but lower toxicities [58]. A phase I is ongoing in solid tumors (NCT02171221).

Other developments aim to increase the half-life and the biodistribution of 5-FU, such as an albumin-bound 5-FU prodrug, which forms a drug–albumin conjugate nanoparticle after administration [59].

Endopeptidase

More recently, a first-in-man phase I study investigated the safety and pharmacokinetic parameters of DOX prodrug DTS-201 (ALAL-DOX). This prodrug is cleaved into tumor tissue by extracellular tumor-specific endopeptidases, neprilysin and thimetoligopeptidase. Preclinical evaluations have shown clearly difference in pharmacokinetic and tissue distribution between DOX and DTS-201 after equimolar administration: with DTS-201, normal tissue were much less exposed to DOX (i.e. area under the curve (AUC) of DOX in the heart normal tissue is decreased of 93% after DTS-201

administration), whereas the exposure of the xenografted tumors was almost doubled [60].

It has shown a better cardiac safety profile at doses of DOX higher than those usually used (threefold increase) [61] and a phase II trial is ongoing. A second generation of tetrapeptidic enzyme-activated DOX prodrug, ALGP-DOX, has demonstrated a tolerated dose 30- to 40-fold higher compared to DOX in patient-derived soft tissue sarcoma xenograft model. The modification of the tetrapeptidic sequence could offer a better prodrug stability and tumor specificity [62].

Legumain

Legumain is a lysosomal protease elevated in several solid tumors such as breast, ovarian, colon and prostate cancer as well as in tumor-associated macrophages and intra tumoral blood vessels [63]. Wu et al. have developed a legumain-activated DOX prodrug (LEG-3) which has shown a better therapeutic index (lower toxicity and better efficacy) in CT26 mouse model [64]. More recently, DOX was conjugated to legumain-cleavable carboxyl-terminated poly(ethylene glycol) via the Ala-Ala-Asp-Leu peptide (4-arm PEG-AANL-DOX) which is known to be a good legumain substrate. Besides, this prodrug could self-assemble into nanoparticles and by consequence takes advantage of the so-called EPR effect. The 4-arm PEG-AANL-DOX has shown in vivo efficacy in human melanoma model comparable to DOX without toxicities [65]. The etoposide agent was also conjugated to legumain-cleavable peptide, Ala-Ala-Asp-ethylenediamine and this prodrug has shown a better cytotoxicity activity in vitro on tumor cell models [66]. Recent studies established on neuroblastoma mostly in children using high-dose of therapeutic treatment were fulfilled, but their use are so far limited. The authors synthesized 6-maleimidocaproyl-AANL-DOX (EMA-AANL-DOX) which has shown lower toxicities in vivo neuroblastoma human cell than DOX. The advantage of EMA-AANL-DOX over previous legumain-cleavable prodrugs is the presence of 6-maleimidocaproyl which is protective, as well as it can bind to serum albumin (SA) and by consequence benefit from the EPR effect [46]. The therapeutic strategy with legumain-cleavable based prodrug is recent and no clinical trial is yet on going.

pH-specific release

Abnormal structure of tumor vasculature along with abnormal blood flow lead to a heterogeneity of oxygen concentration which is lower in the microenvironment of the tumor. By consequence, the low level of oxygen triggers an anaerobic metabolism for the tumor cell and the production of lactic acid and CO_2 . This phenomenon promotes an acidic microenvironment in tumor between pH 6.0 and 6.8 and

in endosome/lysosome ($\text{pH} < 5.5$) which is different from the physiological pH observed in healthy cells ($\text{pH} \approx 7.4$). Among the design of prodrug strategies, pH-cleavable prodrugs are investigated to specifically target the acidity of tumors (Table 3). Different linkers have been developed using polymers responsive to tumoral low pH [polymers having imidazole groups or poly(β -amino ester)]. Paclitaxel has been linked to an imidazole derivative and has shown in vivo increased efficacy on carcinoma mouse model compared to paclitaxel alone [69]. On the other hand, linkers have been synthesized using polymers responsive to intracellular pH (mostly prodrugs with hydrazone bond) [70] with a main focus on the use of DOX as active entity [71–73]. Besides, one of the most interesting challenges of vectorized treatment is to target the bone tissue where metastases are often found. Recently, researchers have studied a bortezomib (BTZ) pH-based prodrug made of co-encapsulated BTZ with aryl boronate group as acidity-labile linker, and alendronate as bone-targeting ligand. This strategy takes advantage of the EPR effect, pH-cleavable linker and active targeting. This prodrug has shown a good efficacy in vivo in bone metastasis human breast adenocarcinoma model; decreased bone destruction was also underlined [74]. MTX was also studied as pH-sensible prodrug; it was coupled to 1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine-*N*-[aldehyde-(polyethyleneglycol)-2000] (DSPE-PEG-Imine-MTX) and the obtained complex showed the ability to self-assemble into micellar nanoparticles. Combining the EPR effect and the imine-pH cleavable linker, DSPE-PEG-Imine-MTX has

shown promising efficacy in vivo in human cervical carcinoma model [75].

Nevertheless, none of these pH-responsive prodrugs is involved on clinical trial for now.

Hypoxic-sensitive release

The formation of abnormal physiopathology of tumor tissue leads to hypoxia. Hypoxia is widely described as a biomarker of malignancy and an obstacle in drug therapy. Taking advantage of the hypoxic environment, several DDS have been developed with a specific hypoxic sensitivity. Among them, hypoxia-activated prodrugs (HAPs), which are specifically cleaved by reductive enzymes with a bioreductive group, have been designed (Table 4). Their developments involve the conjugation of the 2-nitroimidazole moiety to different anticancer drugs. One of the most clinically advanced hypoxia-activated prodrugs, evofosfamide (TH-302), was obtained by conjugation with the DNA cross-linking toxin bromo-isophosphoramidate mustard (Br-IPM). However, TH-302 did not show effective results in phase III trials in locally advanced, unresectable or metastatic soft-tissue sarcoma [76]. Nevertheless, TH-302 is widely studied in others malignancies tumors; such as in i) relapse/refractory multiple myeloma with a positive phase I/II trial [77], ii) patients with aggressive human papillomavirus-negative head and neck squamous cell carcinoma [78], iii) combination with sunitinib in neuroblastoma [79], iv) melanoma xenograft models [80], v) nasopharyngeal carcinoma

Table 3 Acidic pH specific prodrugs

| Target | Parent drug | Linker | Activity | References |
|-----------|-------------|-----------------------------------|--|------------|
| Acidic pH | Paclitaxel | pH sensitive imidazole moiety | Similar cell toxicity and improved efficacy in vivo compared to free paclitaxel | [69] |
| | DOX | pH sensitive hydrazone moiety | Similar cell toxicity and improved efficacy in vivo compared to free DOX | [71] |
| | Bortezomib | pH sensitive aryl boronate moiety | Reduced systemic toxicity and improved therapeutic effects compared to free bortezomib | [74] |
| | MTX | pH sensitive imine moiety | Increased of the in vitro and in vivo anticancer activity | [75] |

Table 4 Hypoxic environment specific prodrugs

| Target | Parent drug | Linker | Activity | References |
|---------------------|---|---|---|------------|
| Hypoxic environment | Nitrogen mustard | Hypoxia sensitive nitroimidazole moiety | Promising phase II results in multiple myeloma patients | [77] |
| | | | Failed in phase III trial in soft tissue sarcoma | [76] |
| | Nitrogen mustard | Hypoxia sensitive nitroaromatic moiety | Validated a phase I clinical trial but increased hematotoxicity | [86] |
| | | | Phase I/II this prodrug is unlikely to be successful for long-term leukemia control because of on-target toxicity | [88] |
| SN-38 | Hypoxia sensitive nitroimidazole moiety | The proof of concept was validated in vitro | [85] | |

xenograft models [81], vi) recurrent bevacizumab-refractory glioblastoma with positive phase I trial [82], vii) osteosarcoma xenograft model [83], and viii) combination with radiotherapy in human orthotopic pancreatic tumor model [84]. This strategy was then applied to design a new prodrug by conjugation of the nitro-imidazole moiety to SN-38, the principal metabolite of irinotecan, showing the capacity of being a promising hypoxia-selective antitumor agent [85]. Besides, PR-104 is a phosphate ester that is hydrolyzed through hypoxia-activated pathway releasing nitrogen mustard. Nevertheless, a clinical trial phase I in advanced solid tumors has shown severe side-effects such as myelotoxicity [86]. Indeed, Guise et al. have demonstrated a second mechanism for PR104 activation through hypoxia-independent aldo-keto reductase 1C3 enzyme [87], which is highly expressed in acute myeloid leukemia (AML) blast [78]. For this reason, PR-104 has been investigated in a phase I/II trial (NCT 01037556) in relapsed or refractory AML and has shown a threefold-higher AUCs due to overexpression of this enzyme into AML blasts compared to other solid tumors. Nevertheless, severe gastro-intestinal toxicities and myelosuppression have been reported and PR-104 did not provide efficacy evidence [88].

Reactive oxygen species-sensitive release

It has now been demonstrated that cancer cells exhibit increased intrinsic oxidative stress leading to an increased amount of reactive oxygen species (ROS) compared to normal healthy cells, such as superoxide anion, hydrogen peroxide and the hydroxyl radicals. These high levels of ROS in cancer cells contribute to cancer-cell proliferation, DNA alterations, apoptosis, metastasis, angiogenesis and alteration in the cellular sensitivity to anticancer agents [89]. Consequently, ROS production has been used as a target to design new ROS-activable prodrug which would be activated in the presence of high amounts of ROS (Table 5). As described by Saravanakumar et al., numerous ROS-responsive linkers can be used to design these prodrugs [90]. However, boron-based linkers have been widely developed in the past decade by designing several prodrugs by conjugation of numerous anticancer compounds to boronate moieties.

Among the anticancer drugs used with this strategy, we can find nitrogen mustards [91], SN-38 [92], endoxifen (non-steroidal selective estrogen receptor modulator) [93] and more recently DOX [12]. All these new prodrugs showed interesting results as the proof of concept of ROS activation was validated in vitro. Nevertheless, the type of linker, its sensitivity to be cleaved and its placement in the molecule structure govern the kinetic of the drug release as each linker has its own advantages and limitations. In regards to arylboronate linkers, there is the generation of harmful quinone methide upon degradation of the arylboronate, which remains a main concern for the clinical development [90].

High weight molecule-conjugate prodrug: serum albumin

The abnormality of the blood vasculature and lymphatic drainage lead to an accumulation of plasmatic protein such as SA into the tumor tissues. Moreover, SA has a biological half-life of 19 days, mostly due to its recognition by FcRn. Researchers have investigated the binding between SA and chemotherapy agents (Table 6) in order to take advantage of EPR effects and they have shown enhanced pharmacokinetic parameters of the active moiety. Since 1994, researchers have investigated albumin–drug conjugates; Gabor et al. have shown promising in vitro results of albumin–DOX conjugates against Ewing’s sarcoma [94]. DOXO-EMCH (also called INNO-206 or aldoxorubicin) binds to the circulating albumin and accumulates into the tumor. Moreover, this DOX-derivative contains an acid-sensitive hydrazone linker that releases the cytotoxic drugs in acidic microenvironment [95]. The phase I and phase II in soft tissue sarcoma have shown a better antitumor activity compared to DOX with a well-tolerated cumulative dose (more than fivefold the median cumulative dose of DOX) [96]. Pharmacokinetic study has shown a narrow volume of distribution (0.3 L/kg versus 72 L/kg at DOX equivalent dose) which means that aldoxorubicin remains into the blood stream and does not accumulate in the body [95]. Then a small concentration of free-DOX and DOX metabolite has been found into the systemic compartment and urine that means aldoxorubicin has bounded to albumin and few DOX was released from

Table 5 ROS specific prodrugs

| Target | Parent drug | Linker | Activity | References |
|-------------------------------|-------------------|---|---|------------|
| Reactive oxygen species (ROS) | Nitrogen mustards | ROS sensitive arylboronate or boronic acid moieties | The proof of concept was validated in vitro | [91] |
| | SN-38 | ROS sensitive arylboronate or boronic acid moieties | The proof of concept was validated in vitro | [92] |
| | Endoxifen | ROS sensitive arylboronate or boronic acid moieties | validated in vitro | [93] |
| | DOX | ROS sensitive arylboronate or boronic acid moieties | and in vivo | [12] |

Table 6 Albumin targeting prodrugs

| Target | Parent drug | Linker | Activity | References |
|---------|-------------|---|---|------------|
| Albumin | DOX | Acid-sensitive hydrazone linker | Clinical trials in sarcomas, small-cell lung cancer, glioblastoma | [95] |
| | DOX | Ile-Ala-Gly-Gln (MMP2-sensitive) | In vivo: melanoma model | [98] |
| | DOX | Ala-Leu-Ala-Leu (Cathepsin B-sensitive) | In vivo: colon model | [99] |
| | DOX | 6-Maleimidocaproic acid-Arg-Arg-Ser-Ser-Tyr-Tyr—Ser-Gly (PSA-sensitive) | In vivo: prostate adenocarcinoma model | [100] |
| | MTX | No linker: simple Amide bond | Phase II in RCC: failed | [103] |
| | Oxaliplatin | Mono-maleimide moiety | In vivo in CT-26 model: tumor regression | [104] |

RCC renal cell carcinoma

aldoxorubicin [97]. Phase III is being investigated in soft tissue sarcomas (NCT02049905), a phase II in recurrent small cell lung cancer (NCT02200757), a phase II in glioblastoma (NCT2014844) and a phase II in HIV-related Kaposi's sarcoma (NCT02029430). Albumin-binding prodrugs of DOX have been investigated with other enzyme-sensible linkers such as Ile-Ala-Gly-Gln for a cleavage by MMP-2 [98], Ala-Leu-Ala-Leu for a cleavage by cathepsin B [99], 6-maleimidocaproic acid-Arg-Arg-Ser-Ser-Tyr-Tyr—Ser-Gly for a cleavage by PSA [100].

Other cytotoxic agents have been investigated such as MTX, drug with a short half-life due to renal elimination leading also to nephrotoxicity. MTX has been bounded to SA in order to limit the renal elimination and promote the tumor uptake rate. The bioconjugate has shown therapeutic activity in various cancers without toxic side-effects [101]. A phase I in cancer patients confirmed a well-tolerated treatment [102]. Unfortunately, no objective responses have been demonstrated in phase II in renal cell carcinoma [103]. Platinum complexes (oxaliplatin and cisplatin) bounded to mono-maleimide as promoiety have been studied; only oxaliplatin derivatives that have shown outstanding antitumor activity are presented, which will now be further developed towards clinical phase I trials [104]. The difference in antitumor activity between oxaliplatin and cisplatin derivatives might be explained by different extravasation potency or different affinity for glycoprotein 18 and 30, which are involved in the recognition of conformationally modified albumin molecules.

Tumor-specific ligand

The active targeting strategy takes advantage of the cellular difference between healthy cells and tumor cells. The targeting ligands can be classified into three groups according to the place where tumor-specific or overexpressed antigen is presented: (1) the tumor cell surface, (2) the endothelial cell surface or (3) the extracellular matrix cells surface. Proteins (monoclonal antibody), peptides (Arg-Gly-Asp (RGD)

peptides, poly-Arg peptides) and small molecules have been investigated to target selectively cancer cells (Table 7).

Immuno-driven DDS

Antibody–drug conjugates The coupling of a cytotoxic agent to a monoclonal antibody (mAb) is called immuno-conjugate or antibody–drug conjugate (ADC). ADC offers a strategy to improve the tolerance/efficacy balance for highly cytotoxic molecules. Indeed, mAb brings target specificity and, consequently, improves the pharmacokinetics (PK) and pharmacodynamics (PD) of these molecules and some of them have obtained their approval from either the FDA or the EMA (Table 8). There are two types of release of the agents: ADC can be cleaved externally via a cleavable linker (non-internalizing mAb) or inside the cell after endocytosis with a non-cleavable linker (internalizing mAb). The highest advantage of non-cleavable linkers compared to cleavable linkers is their plasma stability and by consequence, an optimized therapeutic window mostly due to reduced off-target toxicity. The PK of ADC is complex because it takes into account the PK characteristics of the cytotoxic molecule, the mAb, as well as the physicochemical properties of the linker. It is nevertheless greatly influenced by the PK of the mAb as the latter represents more than 90% of the molecular weight. Besides, sizes ranging of ADC are from 10 to 100 nm and can benefit of EPR effect as well. The first ADC to receive marketing authorization (MA) was Mylotarg[®], conjugation of ozogamicin with gemtuzumab (mean drug:Ab ratio (DAR) 1.5) bounded by an acid-cleavable hydrazine [105]. Ozogamicin is a derivative of calicheamicin, a highly cytotoxic antibiotic, and gemtuzumab is an IgG4 anti-CD33, a surface marker expressed on cells of the myeloid line. Mylotarg[®] received the MA in 2000 from the FDA for the indication of relapsed AML but was withdrawn in 2010 and then reinstated in 2017 with a dosage regimen optimization including a lower dose [106]. Its withdrawal in 2010 was due to off-target reactions, particularly in hepatic tissue where K upffer cells also express

Table 7 Immuno-driven prodrugs

| Target | Parent drug | Ligand | Linker | Activity | References |
|------------------------|--|--|---|--|------------|
| Integrin receptor | DOX | RGD | Polyethylene glycol chain | Increased in vivo efficacy | [143] |
| | Pt | RGD | Amide bond | In vitro: greater the number of bounded RGD, greater is the internalization and efficiency | [120] |
| Hormone receptor | Camptothecin | RGD | Dipeptide alanine–citrulline or phenylalanine–lysine + glycol chain | In vitro: shorter is the glycol chain, lower are the solubility and the affinity for receptor | [121] |
| | DM1 | RGD | PEGP (TMC- <i>co</i> -Poly(pyridyldisulfide cyclic carbonate) [PDSC]) and Mal-PEG-P (TMC- <i>co</i> -PDSC) copolymers | In vivo: increase efficacy in human breast model | [123] |
| LDL receptor | DOX | Small peptide agonist to LHRH receptor | Peptidic bond | Phase II ovarian cancer, prostate cancer), III (endometrial cancer) | [125–127] |
| | Gemcitabine | Gonadotrophin releasing hormone receptor ligand peptide | Succinate linker | Efficacy in human prostate model | [128] |
| Somato-statin receptor | Paclitaxel | Angiopep-2 | Cleavable ester linkage | Phase I in recurrent glioma, phase II for brain metastasis from breast or lung cancer | [130, 144] |
| | Paclitaxel | Octreotide | Polyethylene glycol chain | Efficacy in non-small cell lung cancer model | [132] |
| Folate receptor | Nitroheterocyclic phosphoramidates | Pteric acid | Lysine | In vitro: no satisfactory cytotoxicity | [133] |
| | Pt | Folic acid | Polyethylene glycol | In vitro: no increased cytotoxicity compared PEG-Pt | [134] |
| Folate receptor | N 5-fluoro2'-deoxyuridine-5'-O-monophosphate (FdUMP) | Folic acid | Phosphodiester linkage | In vitro: increased cytotoxicity on 5-FU resistant cells | [135] |
| | Paclitaxel | Folic acid | Ester linker | In vivo: no increased efficacy | [136] |
| Folate receptor | DM1 | Folic acid | Disulfide linker | In vitro: increased cytotoxicity | [137] |
| | Desacetylvinblastine monohydrate | Folic acid | Endosome-cleavable disulfide bond | Phase III in Pt-resistant ovarian cancer: failure in improvement progression-free survival | [139] |
| Folate receptor | Tubulysin B hydrazide | Folic acid | Water soluble saccharo-peptide spacer | Phase I in advanced solid tumors | [145] |
| | Paclitaxel | Folic acid + evans blue (derivative for albumin bonding) | Ester linker | In vivo: Increased circulation half-life, more delivered paclitaxel into tumor and better efficacy | [141] |
| DOX | <i>trans</i> -Activating factor | Ala-Ala-Asp | Breast cancer model | [142] | |

Table 8 Approved ADC by FDA and EMA

| Target | Parent drug | Ligand | Linker | Activity | References |
|--------|-------------|-------------|--|---|-------------------------------|
| CD33 | Ozogamicin | Gemtuzumab | Acid-cleavable hydrazone | Approved drug in relapsed AML | SmPC Mylotarg [®] |
| CD30 | Vedotin | Brentuximab | Maleimide group-valine-citrulline-para-aminobenzyl carbamate | Approved in refractory or recurrent Hodgkin's lymphoma and large cell anaplastic lymphoma, primary cutaneous large cell anaplastic lymphoma, Hodgkin's lymphoma stage III or IV | SmPC Adcetris [®] |
| CD22 | Ozogamicin | Inotuzumab | Acid-labile acetyl butyrate | Relapsed or refractory acute lymphoblastic leukemia with B precursor | SmPC Besponsa [®] |
| HER2 | DM1 | Trastuzumab | Amide bond | HER2 positive, metastatic or locally advanced, unresectable breast cancer | SmPC Trastuzumab [®] |

SmPC summary of product characteristics

CD33 [107], leading to patient deaths (study SWOG S0106). The second ADC to receive MA by FDA (2011) and EMA (2012) was Adcetris[®] for refractory or recurrent Hodgkin's lymphoma and large cell anaplastic lymphoma. This ADC has also received MA in primary cutaneous large cell anaplastic lymphoma (FDA-2017 and EMA-2017) and more recently in first-line treatment against Hodgkin's lymphoma stage III or IV (FDA-2018) [47]. Adcetris[®] is composed of the antimetabolic monomethylauristatin E (MMAE or vedotin) and the IgG1 brentuximab. MMAE which belongs to the antimetabolic and antimicrotubule family is 50- to 200-fold more active than vinca-alkaloid. Brentuximab specifically targets CD30 surface receptors on the surface of activated B and T cells. The binding between the mAb and the cytotoxic agent (mean DAR 3.9) is made of a maleimide group, protease (cathepsin)-cleavable linker (valine-citrulline) and para-aminobenzyl carbamate spacers. This peptide-based linker is stable under physiological conditions, but, after internalization, is cleaved inside the lysosome by lysosomal cysteine proteases such as cathepsin B. Recently, the ADC, named Besponsa[®], received MA in 2017 for in acute lymphoblastic leukemia (ALL) with B precursor, CD22 positive, relapsed or refractory. This ADC is made of ozogamicin and IgG4 inotuzumab (mean DAR 5.1) linked by an acid-labile acetyl butyrate linker, more stable in bloodstream than the linker used in Mylotarg[®] [108]. Inotuzumab specifically targets the CD22 surface protein present on B cells.

Regarding treatment of solid tumors, the PK biodisponibility is more complex because ADC has to diffuse to the targeting tissue. The parent drug maytansine—or DM1—is widely investigated. It is a vinca-alkaloid derivative, a potent inhibitor of microtubule assembly, 25–400 times more active than paclitaxel and 100–5000 times more active than DOX [109]. However, it failed as an anticancer agent in human clinical trials because of its lack of tumor specificity associated with an important systemic toxicity. DM1 has been linked to trastuzumab to afford T-DM1 with a mean DAR 3.4 via succinimidyl-*trans*-4(maleimidylmethyl) cyclohexane-1-carboxylate, a stable thioether linker that binds the

mAb lysine residues [110]. Trastuzumab (Herceptin[®]) is a IgG1 mAb targeting extracellular domain of human epidermal growth factor receptor 2 (HER2 receptor, also known as neu, ErbB2, *p185^{HER2}*) and is widely used in the treatment of HER2 receptor positive breast cancer since 1998 in the United States and 2000 in Europe. This new ADC showed a similar drug release compared to Cantuzumab-DM1. Unlike Cantuzumab-DM1, the bond that connects the two entities is not cleavable by enzymes or reducing agents (e.g. glutathione) but the release of the cytotoxic agent is done by catabolism of the antibody after internalization in the targeted cancer cell. T-DM1 conjugate design led to favorable pharmacokinetic properties with an increased half-life from 2 to 44 h and an increased area under the curve by 60-fold [111]. T-DM1 (Kadcyla[®]) received MA in 2013 (FDA) and 2014 (EMA) in HER2 positive, metastatic or locally advanced, unresectable breast cancer previously treated with trastuzumab and a taxane. However, many DM1 or DM4-coupled mAbs failed as anticancer agents in clinical trials [112]. The reasons for discontinued ADCs are due to off-target effect with unacceptable toxicities (for example Bivatuzumab-DM1 has shown a fatal dermal toxicity in phase I leading to stop the development; Bivatuzumab targets CD44v6, receptor abundantly present in head and neck squamous cell carcinomas but also on normal keratinocytes [113]). It was discontinued also for the lack evidence of clinical activity [112].

The previously mentioned ADC target a receptor present on the surface of the tumor cell. A recent strategy aims to develop non-internalizing ADC targeting antigens overexpressed in the tumor extracellular matrix such as collagen IV, fibrin, fibronectin and tenascin-C [114]. Indeed Neri's team is investigating DM1 coupled with an antibody directed against fibronectin [115], MMAE and anthracycline derivative coupled with an antibody directed against tenascin-C through a valine-citrulline linker [114]. These ADCs have shown promising results in vivo in teratocarcinoma and human epidermoid carcinoma. The notable advantages are the accessibility of the extracellular matrix components and

the release of cytotoxic into the tumoral microenvironment may facilitate the diffusion and internalization in neighboring cells.

Approximately 222 clinical trials have been reported describing 82 different new ADCs that are listed on the clinicaltrials.gov website, 123 of them are currently under clinical evaluation.

Cancer-targeting ADC and more generally mAbs' uses are limited by the pharmacokinetic properties: Cellular up-take clearance due tumor tissue infiltration and reticulo-endothelial elimination via Fc fragments is the major parameter of the distribution of ADC.

Fragment-based antibody Over the past decades, the use of antibody-based therapeutics has grown exponentially leading to a revolution in biological drugs and has revolutionized targeted cancer therapies. However, the use of such therapeutics has limitation due to the large size of the scaffolds leading to the following: (1) poor penetration in the tumor, (2) important systemic accumulation and (3) slow clearance profiles. As the targeting capacity of Ab is achieved by a small variable loop, the selection of these small fragments has led to the design of small fragment-based prodrugs [116]. Aubrey et al. have recently developed two promising auristatin prodrugs targeting specifically HER2-positive breast cancer using this strategy. Their work was fulfilled by conjugation of auristatin to an engineered scFv fragment through maleimide linker. Their strategy was validated in vitro on HER2-positive breast cancer cells with activities in the subnanomolar range and no activity on HER2-negative breast cancer cells [117].

Peptides and small molecule-driven DDS

Investigation of peptide as drug delivery system conjugate offers some advantages compared to mAbs. Indeed peptides are easier to use (no tertiary structure, control of the drug ratio, lower immunogenicity), easier and less expensive to produce in large quantities. Upon peptide-receptor binding, the complex can undergo receptor-mediated endocytosis.

Integrin receptor targeting $\alpha_v\beta_3$ integrin is a protein involved in cell adhesion playing a major role in angiogenesis and tumor metastasis. This protein is overexpressed on the surface of tumor cell in glioma, melanoma, breast cancer and ovarian cancer [118]. RGD peptide has been shown to exhibit excellent binding affinity and selectivity for $\alpha_v\beta_3$ integrin [119]. Synthesis of RGD-derivative DOX, Pt and camptothecin has been investigated in order to give specificity and decrease toxicity, particularly cardiotoxicity for DOX and nephrotoxicity for Pt. Nevertheless, only promising in vitro data are available and need more investigations [120, 121]. Moreover, many studies are investigating

in the development of integrin-targeting DDS [122]. Recent study on cyclic RGD-functionalized micellar DM1 prodrug; compared to T-DM1, the cytotoxic loading is higher as well as the stability and the efficacy has been proved in vivo in human breast model [123].

Hormone receptor targeting Luteinizing hormone releasing hormone (LHRH) receptors is over-expressed in prostate and gynecological cancers [124]. A small peptide agonist to LHRH receptor has been conjugated to the DOX (AEZS-108). AEZS-108 has shown positive clinical results in ovarian cancer (phase II [125]), in castration- and taxane-resistant prostate cancer (phase I: [126]; phase II: [127]) and in endometrial cancer but the phase III did not improve efficacy or safety compared to DOX alone (NCT01767155). Besides, gemcitabine was coupled with gonadotrophin releasing hormone receptor ligand peptide with a succinate linker. The complex has shown efficacy in vivo in a castration resistant prostate cancer model [128].

LDL receptor targeting ANG1005, called GRN1005 or paclitaxel trevatide, is a prodrug designed from three molecules of paclitaxel coupled with angiopep-2. Angiopep-2 targets low-density lipoprotein receptors which are overexpressed on solid tumor and enhances the uptake through the blood–brain-barrier [129]. Its half-life is shorter compared to nab-paclitaxel (3.6 h vs 21.6 h, respectively) and paclitaxel alone (20.5 h) [129]. Several trials have been investigated with this prodrug in recurrent malignant glioma [130] (NCT01967810), in non-small cell lung cancer patients with brain metastases (NCT01497665; NCT02048059) and in breast or lung cancer patients with brain metastases (NCT01679743-ended in 2018).

Somatostatin receptor targeting Somatostatin receptor are G-protein-coupled receptors overexpressed in various cancer, therefore targeting somatostatin receptor has been proposed for anticancer strategy. Octreotide, an octapeptide that mimics natural somatostatin pharmacologically, has been used to develop drug delivery system. Huo et al. have studied the somatostatin receptor as a target for the delivery of paclitaxel. Paclitaxel presents a poor water solubility (0.3 $\mu\text{g}/\text{mL}$); the currently formulation mixes Cremophor[®]EL and ethanol (Taxol[®]) in order to solubilize paclitaxel for intravenous injection. However, Cremophor[®]EL can lead to serious acute hypersensitivity reactions and other toxicities [131], therefore researchers have investigated new paclitaxel analog in order to increase specificity and especially solubility. The authors have developed a paclitaxel prodrug made of taxane and octreotide linked by polyethylene glycol chain (OCT-PEG-PTX). The paclitaxel prodrug solubility was hugely increased (30,000-fold) compared paclitaxel, and has shown a higher antitumor efficacy in somatostatin receptor

overexpressing non-small cell lung cancer cells [132] but no clinical data are currently available.

Folate receptor targeting The overexpression of folate receptor in many human cancers leads to developed folate-targeting prodrugs. DNA alkylating agents [133], platinum derivatives [134], 5-fluorouracil [135], paclitaxel [136] conjugates have been investigated for this purpose. Nevertheless, all cited folate receptor-specific prodrugs have shown no increased cytotoxicity in vitro and in vivo (for paclitaxel prodrug) compared to the parent drug except for folic acid (FA)-derivated 5FU which has proven enhanced efficacy on 5FU-resistant cells. FA has been linked to FdUMP and this conjugate is 10- to 25-fold more cytotoxic than 5-FU on human colorectal tumor cell lines. Ladino et al. have investigated the synthesis of DM1 bound to FA which has demonstrated high cytotoxicity on folate receptor positive carcinoma (KB), ovarian cancer (SK-OV-3) and colon cancer (Lo Vo, SW620) cell lines [137]; 10 years later Reddy et al. has confirmed the high specificity of folate-conjugated DM1 and the optimized therapeutic window. However, to our knowledge, this was only demonstrated in preclinical experiments [138].

EC145 (vintafolide), a conjugate of folic acid and a vinca-alkaloid compound (desacetyl vinblastine hydrazone) through an endosome-cleavable disulfide bond, has been evaluated in a phase I and II trial in advanced non-small lung cancer, ovarian and endometrial cancers [139]. The drug received orphan drug status in Europe in 2012 for the treatment of folate-positive Pt-resistant ovarian cancer in combination with pegylated liposomal DOX, but in 2014, Merck and Endocyte stopped its commercialization because of drug failure in improvement progression-free survival. Guertin et al. proposed the hypothesis that this failure is due to the implication of P-glycoprotein pump that trigger a folate-prodrug resistance [140]. A second-generation FA drug conjugate linked to tubulysin B hydrazone is under investigation in patients with advanced solid tumors (NCT01999738). EC1456) delivers higher doses than EC145,

Recently, Shan et al. have developed a dual-targeting prodrug of paclitaxel with bifunctional folate and albumin binding moieties for both passive and active targeted cancer therapy [141]. This prodrug has shown a prolonged blood circulation, enhanced drug accumulation in tumors and higher therapeutic index in vivo on human breast adenocarcinoma model.

Others Liu et al. have designed a DOX derivative conjugated to cell-penetrating peptides (TAF, *trans*-activating factor) with a legumain-sensible linker (alanine-alanine-asparagine), and loaded on liposomes (using PEG 2000) in order to combine different targeting ways [142]. TAF is known to promote the cell internalization capability of lipo-

somal NPs. The addition of the legumain-sensible linker to TAT has shown, in vitro, a decrease of 72.65% for its transmembrane transport capacity. Therefore legumain-sensible targeting has been used in vivo in breast cancer model to effectively recover the internalization capacity of TAF in tumor cells and tumor-associated macrophages which both upregulate legumain expression.

This complex DDS has shown a better therapeutic window on breast cancer mouse model.

Most of these the studies fulfilled with peptide-associated prodrug were carried out in the last decades. Taking into account the relative recentness of these studies the proof of concept in clinics is hard to establish as for today. Thus, none of the peptide-associated prodrug has successfully reached the therapeutic market, yet.

Conclusion

The intratumoral chemotherapies, surgery and radiotherapy, are widely used but their success is guaranteed only if the tumor presents a well-defined localization. In the field of cancer chemotherapy, the main goal is to facilitate the accumulation of active drugs in tumor tissues or in its microenvironment compared to normal tissues. The therapeutic effect of daily used chemotherapeutic agents is far from being optimal due to their poor specificity and/or insufficient access to the tumor. Indeed, non-local administration of chemotherapy needs the delivery of anticancer drugs from the blood to the tumor cell through a complex transport process. The antitumor drug vectorization (i.e. targeted drug delivery systems), allows, on one hand, to target the malignant cells, and on the other hand to avoid toxicities on non-malignant cells. Prodrug design has become an established concept and a powerful tool in optimizing daily used chemotherapy agents, overcoming their physicochemical and pharmaceutical issues. These modifications aim to improve tumor targeting, as the active compound is released only after an activation process is triggered near the target. Targeted prodrug drug delivery systems should be designed to facilitate the targeted delivery of anticancer agents to the tumor. In the present review, we have reported that several PBN-DDS strategies achieve this goal by combining the advantages of both prodrugs and nanotechnology. Nowadays, numerous PBN-DDS are under clinical trials suggesting that these new targeted compounds/DDS could have great potential to be used in clinical practice in the future. These prodrug delivery strategies are novel. Most of the investigations are from academic laboratories and are currently in preclinical or early stages clinical studies. The way to FDA or EMA approval is long and numerous prodrugs fail to obtain this invaluable sesame, the market authorization. However, these failures pave the way for future pharmaceutical development as in oncology,

and in other therapeutic areas, medicine will be always more precision (i.e. specific) medicine.

Funding This review work was not supported by any grant.

Compliance with ethical standards

Conflict of interest All authors declare that they have no conflict of interest.

Ethical approval This article does not contain any studies with human participants or animals performed by any of the authors.

References

- Lesniewska-Kowiel MA, Muszalska I (2017) Strategies in the designing of prodrugs, taking into account the antiviral and anticancer compounds. *Eur J Med Chem* 129:53–71. <https://doi.org/10.1016/j.ejmech.2017.02.011>
- Singh Y, Palombo M, Sinko PJ (2008) Recent trends in targeted anticancer prodrug and conjugate design. *Curr Med Chem* 15(18):1802–1826
- Soyez H, Schacht E, Vanderkerken S (1996) The crucial role of spacer groups in macromolecular prodrug design. *Adv Drug Deliv Rev* 21(2):81–106. [https://doi.org/10.1016/s0169-409x\(96\)00400-0](https://doi.org/10.1016/s0169-409x(96)00400-0)
- Muller CE (2009) Prodrug approaches for enhancing the bio-availability of drugs with low solubility. *Chem Biodivers* 6(11):2071–2083. <https://doi.org/10.1002/cbdv.200900114>
- Heynick F (2009) The original ‘magic bullet’ is 100 years old—extra. *Br J Psychiatry* 195(5):456. <https://doi.org/10.1192/bjp.195.5.456>
- Albert A (1958) Chemical aspects of selective toxicity. *Nature* 182(4633):421–422. <https://doi.org/10.1038/182421a0>
- Rautio J, Kumpulainen H, Heimbach T, Oliyai R, Oh D, Jarvinen T, Savolainen J (2008) Prodrugs: design and clinical applications. *Nat Rev Drug Discov* 7(3):255–270. <https://doi.org/10.1038/nrd2468>
- Mahato R, Tai W, Cheng K (2011) Prodrugs for improving tumor targetability and efficiency. *Adv Drug Deliv Rev* 63(8):659–670. <https://doi.org/10.1016/j.addr.2011.02.002>
- Gupta D, Gupta SV, Lee KD, Amidon GL (2009) Chemical and enzymatic stability of amino acid prodrugs containing methoxy, ethoxy and propylene glycol linkers. *Mol Pharm* 6(5):1604–1611. <https://doi.org/10.1021/mp900084v>
- Walther R, Rautio J, Zelikin AN (2017) Prodrugs in medicinal chemistry and enzyme prodrug therapies. *Adv Drug Deliv Rev* 118:65–77. <https://doi.org/10.1016/j.addr.2017.06.013>
- Alouane A, Labruere R, Le Saux T, Schmidt F, Jullien L (2015) Self-immolative spacers: kinetic aspects, structure–property relationships, and applications. *Angew Chem Int Ed Engl* 54(26):7492–7509. <https://doi.org/10.1002/anie.201500088>
- Ye M, Han Y, Tang J, Piao Y, Liu X, Zhou Z, Gao J, Rao J, Shen Y (2017) A tumor-specific cascade amplification drug release nanoparticle for overcoming multidrug resistance in cancers. *Adv Mater* 29(38):1702342. <https://doi.org/10.1002/adma.201702342>
- Bildstein L, Pili B, Marsaud V, Wack S, Meneau F, Lepetre-Mouelhi S, Desmaele D, Bourgaux C, Couvreur P, Dubernet C (2011) Interaction of an amphiphilic squalenoyl prodrug of gemcitabine with cellular membranes. *Eur J Pharm Biopharm* 79(3):612–620. <https://doi.org/10.1016/j.ejpb.2011.07.003>
- Blanco E, Shen H, Ferrari M (2015) Principles of nanoparticle design for overcoming biological barriers to drug delivery. *Nat Biotechnol* 33(9):941–951. <https://doi.org/10.1038/nbt.3330>
- Maeda H, Nakamura H, Fang J (2013) The EPR effect for macromolecular drug delivery to solid tumors: improvement of tumor uptake, lowering of systemic toxicity, and distinct tumor imaging in vivo. *Adv Drug Deliv Rev* 65(1):71–79. <https://doi.org/10.1016/j.addr.2012.10.002>
- Greish K (2010) enhanced permeability and retention (EPR) effect for anticancer nanomedicine drug targeting. In: Grobmyer SR, Moudgil BM (eds) *Cancer nanotechnology: methods and protocols*. Humana Press, Totowa, pp 25–37. https://doi.org/10.1007/978-1-60761-609-2_3
- Nichols JW, Bae YH (2014) EPR: evidence and fallacy. *J Control Release* 190:451–464. <https://doi.org/10.1016/j.jconrel.2014.03.057>
- Bansal R, Post E, Proost JH, de Jager-Krikken A, Poelstra K, Prakash J (2011) PEGylation improves pharmacokinetic profile, liver uptake and efficacy of interferon gamma in liver fibrosis. *J Control Release* 154(3):233–240. <https://doi.org/10.1016/j.jconrel.2011.05.027>
- Choi KY, Min KH, Yoon HY, Kim K, Park JH, Kwon IC, Choi K, Jeong SY (2011) PEGylation of hyaluronic acid nanoparticles improves tumor targetability in vivo. *Biomaterials* 32(7):1880–1889. <https://doi.org/10.1016/j.biomaterials.2010.11.010>
- Piktel E, Niemirowicz K, Watek M, Wollny T, Deptula P, Bucki R (2016) Recent insights in nanotechnology-based drugs and formulations designed for effective anti-cancer therapy. *J Nanobiotechnol* 14(1):39. <https://doi.org/10.1186/s12951-016-0193-x>
- Kamaly N, Xiao Z, Valencia PM, Radovic-Moreno AF, Farokhzad OC (2012) Targeted polymeric therapeutic nanoparticles: design, development and clinical translation. *Chem Soc Rev* 41(7):2971–3010. <https://doi.org/10.1039/c2cs15344k>
- Khandare JJ, Jayant S, Singh A, Chandna P, Wang Y, Vorsa N, Minko T (2006) Dendrimer versus linear conjugate: influence of polymeric architecture on the delivery and anticancer effect of paclitaxel. *Bioconjug Chem* 17(6):1464–1472. <https://doi.org/10.1021/bc060240p>
- Langer CJ, O’Byrne KJ, Socinski MA, Mikhailov SM, Lesniewski-Kmak K, Smakal M, Ciuleanu TE, Orlov SV, Dediu M, Heigener D, Eisenfeld AJ, Sandalic L, Oldham FB, Singer JW, Ross HJ (2008) Phase III trial comparing paclitaxel polyglumex (CT-2103, PPX) in combination with carboplatin versus standard paclitaxel and carboplatin in the treatment of PS 2 patients with chemotherapy-naive advanced non-small cell lung cancer. *J Thorac Oncol* 3(6):623–630. <https://doi.org/10.1097/JTO.0b013e3181753b4b>
- Cheng C, Jiang-Ling Z, Xue H, Fei S, Xiu-Li W, Yu-Zhong W (2014) A prodrug strategy based on chitosan for efficient intracellular anticancer drug delivery. *Nanotechnology* 25(25):255101
- Dragojevic S, Ryu JS, Raucher D (2015) Polymer-based prodrugs: improving tumor targeting and the solubility of small molecule drugs in cancer therapy. *Molecules* 20(12):21750–21769. <https://doi.org/10.3390/molecules201219804>
- Fumagalli G, Marucci C, Christodoulou MS, Stella B, Dosio F, Passarella D (2016) Self-assembly drug conjugates for anticancer treatment. *Drug Discov Today* 21(8):1321–1329. <https://doi.org/10.1016/j.drudis.2016.06.018>
- Zhou M, Zhang RH, Wang M, Xu GB, Liao SG (2017) Prodrugs of triterpenoids and their derivatives. *Eur J Med Chem* 131:222–236. <https://doi.org/10.1016/j.ejmech.2017.03.005>
- Maksimenko A, Dosio F, Mougín J, Ferrero A, Wack S, Reddy LH, Weyn AA, Lepeltier E, Bourgaux C, Stella B, Cattel L, Couvreur P (2014) A unique squalenoylated and nonpegylated doxorubicin nanomedicine with systemic long-circulating

- properties and anticancer activity. *Proc Natl Acad Sci USA* 111(2):E217–E226. <https://doi.org/10.1073/pnas.1313459110>
29. Mura S, Nicolas J, Couvreur P (2013) Stimuli-responsive nano-carriers for drug delivery. *Nat Mater* 12(11):991–1003. <https://doi.org/10.1038/nmat3776>
 30. Siafaka PI, Ustundag Okur N, Karavas E, Bikiaris DN (2016) Surface modified multifunctional and stimuli responsive nanoparticles for drug targeting: current status and uses. *Int J Mol Sci*. <https://doi.org/10.3390/ijms17091440>
 31. Chau Y, Padera RF, Dang NM, Langer R (2006) Antitumor efficacy of a novel polymer-peptide-drug conjugate in human tumor xenograft models. *Int J Cancer* 118(6):1519–1526. <https://doi.org/10.1002/ijc.21495>
 32. Huang C, Yi X, Kong D, Chen L, Min G (2016) Controlled release strategy of paclitaxel by conjugating to matrix metalloproteinases-2 sensitive peptide. *Oncotarget* 7(32):52230–52238. <https://doi.org/10.18632/oncotarget.10735>
 33. Denmeade SR, Nagy A, Gao J, Lilja H, Schally AV, Isaacs JT (1998) Enzymatic activation of a doxorubicin-peptide prodrug by prostate-specific antigen. *Cancer Res* 58(12):2537–2540
 34. DiPaola RS, Rinehart J, Nemunaitis J, Ebbinghaus S, Rubin E, Capanna T, Ciardella M, Doyle-Lindrud S, Goodwin S, Fontaine M, Adams N, Williams A, Schwartz M, Winchell G, Wickersham K, Deutsch P, Yao SL (2002) Characterization of a novel prostate-specific antigen-activated peptide-doxorubicin conjugate in patients with prostate cancer. *J Clin Oncol* 20(7):1874–1879. <https://doi.org/10.1200/JCO.2002.07.001>
 35. Wu X, Hu L (2016) Design and synthesis of peptide conjugates of phosphoramidate mustard as prodrugs activated by prostate-specific antigen. *Bioorg Med Chem* 24(12):2697–2706. <https://doi.org/10.1016/j.bmc.2016.04.035>
 36. DeFeo-Jones D, Brady SF, Feng DM, Wong BK, Bolyar T, Haskell K, Kiefer DM, Leander K, McAvoy E, Lumma P, Pawluczyk JM, Wai J, Motzel SL, Keenan K, Van Zwieten M, Lin JH, Garsky VM, Freidinger R, Oliff A, Jones RE (2002) A prostate-specific antigen (PSA)-activated vinblastine prodrug selectively kills PSA-secreting cells in vivo. *Mol Cancer Ther* 1(7):451–459
 37. Brennen WN, Rosen DM, Wang H, Isaacs JT, Denmeade SR (2012) Targeting carcinoma-associated fibroblasts within the tumor stroma with a fibroblast activation protein-activated prodrug. *J Natl Cancer Inst* 104(17):1320–1334. <https://doi.org/10.1093/jnci/djs336>
 38. Chen M, Lei X, Shi C, Huang M, Li X, Wu B, Li Z, Han W, Du B, Hu J, Nie Q, Mai W, Ma N, Xu N, Zhang X, Fan C, Hong A, Xia M, Luo L, Ma A, Li H, Yu Q, Chen H, Zhang D, Ye W (2017) Pericyte-targeting prodrug overcomes tumor resistance to vascular disrupting agents. *J Clin Investig* 127(10):3689–3701. <https://doi.org/10.1172/JCI94258>
 39. Huang S, Zhang Y, Zhong J, Pan Y, Cai S, Xu J (2018) Toxicological profile and safety pharmacology of a single dose of fibroblast activation protein- α -based doxorubicin prodrug: in vitro and in vivo evaluation. *Anticancer Drugs* 29(3):253–261. <https://doi.org/10.1097/CAD.0000000000000593>
 40. Wang J, Li Q, Li X, Yuan W, Huang S, Cai S, Xu J (2017) A novel FAP α -based Z-Gly-Pro epirubicin prodrug for improving tumor-targeting chemotherapy. *Eur J Pharmacol* 815:166–172. <https://doi.org/10.1016/j.ejphar.2017.09.016>
 41. Aggarwal N, Sloane BF (2014) Cathepsin B: multiple roles in cancer. *Proteom Clin Appl* 8(5–6):427–437. <https://doi.org/10.1002/prca.201300105>
 42. Shao LH, Liu SP, Hou JX, Zhang YH, Peng CW, Zhong YJ, Liu X, Liu XL, Hong YP, Firestone RA, Li Y (2012) Cathepsin B cleavable novel prodrug Ac-Phe-Lys-PABC-ADM enhances efficacy at reduced toxicity in treating gastric cancer peritoneal carcinomatosis: an experimental study. *Cancer* 118(11):2986–2996. <https://doi.org/10.1002/cncr.26596>
 43. Zhong YJ, Shao LH, Li Y (2013) Cathepsin B-cleavable doxorubicin prodrugs for targeted cancer therapy (review). *Int J Oncol* 42(2):373–383. <https://doi.org/10.3892/ijo.2012.1754>
 44. Seymour LW, Ferry DR, Kerr DJ, Rea D, Whitlock M, Poyner R, Boivin C, Hesslewood S, Twelves C, Blackie R, Schatzlein A, Jodrell D, Bissett D, Calvert H, Lind M, Robbins A, Burtles S, Duncan R, Cassidy J (2009) Phase II studies of polymer-doxorubicin (PK1, FCE28068) in the treatment of breast, lung and colorectal cancer. *Int J Oncol* 34(6):1629–1636. https://doi.org/10.3892/ijo_00000293
 45. Paz-Ares L, Ross H, O'Brien M, Riviere A, Gatzemeier U, Von Pawel J, Kaukel E, Freitag L, Digel W, Bischoff H, Garcia-Campelo R, Iannotti N, Reiterer P, Bover I, Prendiville J, Eisenfeld AJ, Oldham FB, Bandstra B, Singer JW, Bonomi P (2008) Phase III trial comparing paclitaxel poliglumex vs docetaxel in the second-line treatment of non-small-cell lung cancer. *Br J Cancer* 98(10):1608–1613. <https://doi.org/10.1038/sj.bjc.6604372>
 46. Zhang M, Jiang Z, Chen S, Wu Z, Chen K, Wu Y (2018) Legumain correlates with neuroblastoma differentiation and can be used in prodrug design. *Chem Biol Drug Des* 91(2):534–544. <https://doi.org/10.1111/cbdd.13116>
 47. Connors JMRJ (2018) Brentuximab vedotin for stage III or IV Hodgkin's lymphoma. *N Engl J Med* 378(16):1558–1561. <https://doi.org/10.1056/NEJMc1802363>
 48. Karthaler-Benbakka C, Koblmüller B, Mathuber M, Holste K, Berger W, Heffeter P, Kowol CR, Keppler BK (2018) Synthesis, characterization and in vitro studies of a Cathepsin B-cleavable prodrug of the VEGFR inhibitor sunitinib. *Chem Biodivers*. <https://doi.org/10.1002/cbdv.201800520>
 49. Bollag W (1965) Hartmann HR (1980) tumor inhibitory effects of a new fluorouracil derivative: 5'-deoxy-5-fluorouridine. *Eur J Cancer* 16(4):427–432. [https://doi.org/10.1016/0014-2964\(80\)90221-2](https://doi.org/10.1016/0014-2964(80)90221-2)
 50. Hiller SA, Lidak MY, Zhuk RA et al (1969) Analogs of pyrimidine nucleosides. *Chem Heterocycl Compd* 5(2):283–285. <https://doi.org/10.1007/BF00943946>
 51. Koukourakis GV, Kouloulis V, Koukourakis MJ, Zacharias GA, Zabatis H, Kouvaris J (2008) Efficacy of the oral fluorouracil pro-drug capecitabine in cancer treatment: a review. *Molecules* 13(8):1897–1922. <https://doi.org/10.3390/molecules13081897>
 52. Sakata Y, Ohtsu A, Horikoshi N, Sugimachi K, Mitachi Y, Taguchi T (1998) Late phase II study of novel oral fluoropyrimidine anticancer drug S-1 (1 M tegafur–0.4 M gimestat–1 M otastat potassium) in advanced gastric cancer patients. *Eur J Cancer* 34(11):1715–1720. [https://doi.org/10.1016/s0959-8049\(98\)00211-1](https://doi.org/10.1016/s0959-8049(98)00211-1)
 53. Alberto P, Winkelmann JJ, Paschoud N, Peytremann R, Bruyere A, Righetti A, Decoster G, Holdener EE (1989) Phase I study of oral doxifluridine using two schedules. *Eur J Cancer Clin Oncol* 25(5):905–908. [https://doi.org/10.1016/0277-5379\(89\)90139-9](https://doi.org/10.1016/0277-5379(89)90139-9)
 54. Kim NK, Min JS, Park JK, Yun SH, Sung JS, Jung HC, Roh JK (2001) Intravenous 5-fluorouracil versus oral doxifluridine as preoperative concurrent chemoradiation for locally advanced rectal cancer: prospective randomized trials. *Jpn J Clin Oncol* 31(1):25–29. <https://doi.org/10.1093/jjco/hye009>
 55. van Oosterom AT, ten Bokkel Huinink WW, van der Burg ME, Vermorken JB, Willemse PH, Neijt JP (1991) Phase II clinical trial of doxifluridine in patients with advanced ovarian cancer. *Eur J Cancer* 27(6):747–749. [https://doi.org/10.1016/0277-5379\(91\)90180-L](https://doi.org/10.1016/0277-5379(91)90180-L)
 56. Miwa M, Ura M, Nishida M, Sawada N, Ishikawa T, Mori K, Shimma N, Umeda I, Ishitsuka H (1998) Design of a novel oral fluoropyrimidine carbamate, capecitabine, which generates

- 5-fluorouracil selectively in tumours by enzymes concentrated in human liver and cancer tissue. *Eur J Cancer* 34(8):1274–1281. [https://doi.org/10.1016/s0959-8049\(98\)00058-6](https://doi.org/10.1016/s0959-8049(98)00058-6)
57. Ter Veer E, Ngai LL, Valkenhoeft GV, Mohammad NH, Anderegg MCJ, van Oijen MGH, van Laarhoven HWM (2017) Capecitabine, 5-fluorouracil and S-1 based regimens for previously untreated advanced oesophagogastric cancer: a network meta-analysis. *Sci Rep* 7(1):7142. <https://doi.org/10.1038/s41598-017-07750-3>
 58. Fukushima M, Iizuka K, Jin C, Zhang C, Hong M, Eshima K (2017) Development of new promising antimetabolite, DFP-11207 with self-controlled toxicity in rodents. *Drug Des Dev Ther* 11:1693–1705. <https://doi.org/10.2147/DDDT.S128420>
 59. Zhao D, Zhang H, Tao W, Wei W, Sun J, He Z (2017) A rapid albumin-binding 5-fluorouracil prodrug with a prolonged circulation time and enhanced antitumor activity. *Biomater Sci* 5(3):502–510. <https://doi.org/10.1039/c6bm00884d>
 60. Dubois V, Dasnois L, Lebtahi K, Collot F, Heylen N, Havaux N, Fernandez AM, Lobl TJ, Oliyai C, Nieder M, Shochat D, Yarranton GT, Trouet A (2002) CPI-0004Na, a new extracellularly tumor-activated prodrug of doxorubicin: in vivo toxicity, activity, and tissue distribution confirm tumor cell selectivity. *Cancer Res* 62(8):2327–2331
 61. Schoffski P, Delord JP, Brain E, Robert J, Dumez H, Gasmi J, Trouet A (2017) First-in-man phase I study assessing the safety and pharmacokinetics of a 1-h intravenous infusion of the doxorubicin prodrug DTS-201 every 3 weeks in patients with advanced or metastatic solid tumours. *Eur J Cancer* 86:240–247. <https://doi.org/10.1016/j.ejca.2017.09.009>
 62. Cornillie J, Wozniak A, Pokreisz P, Casazza A, Vreys L, Wellens J, Vanleeuw U, Gebreyohannes YK, Debiec-Rychter M, Sciort R, Hompes D, Schoffski P (2017) In vivo antitumoral efficacy of PhAc-ALGP-doxorubicin, an enzyme-activated doxorubicin prodrug, in patient-derived soft tissue sarcoma xenograft models. *Mol Cancer Ther* 16(8):1566–1575. <https://doi.org/10.1158/1535-7163.MCT-16-0832>
 63. Vandooren J, Opendakker G, Loadman PM, Edwards DR (2016) Proteases in cancer drug delivery. *Adv Drug Deliv Rev* 97:144–155. <https://doi.org/10.1016/j.addr.2015.12.020>
 64. Wu W, Luo Y, Sun C, Liu Y, Kuo P, Varga J, Xiang R, Reisfeld R, Janda KD, Edgington TS, Liu C (2006) Targeting cell-impermeable prodrug activation to tumor microenvironment eradicates multiple drug-resistant neoplasms. *Cancer Res* 66(2):970–980. <https://doi.org/10.1158/0008-5472.CAN-05-2591>
 65. Zhou H, Sun H, Lv S, Zhang D, Zhang X, Tang Z, Chen X (2017) Legumain-cleavable 4-arm poly(ethylene glycol)-doxorubicin conjugate for tumor specific delivery and release. *Acta Biomater* 54:227–238. <https://doi.org/10.1016/j.actbio.2017.03.019>
 66. Stern L, Perry R, Ofek P, Many A, Shabat D, Satchi-Fainaro R (2009) A novel antitumor prodrug platform designed to be cleaved by the endoprotease legumain. *Bioconjug Chem* 20(3):500–510. <https://doi.org/10.1021/bc800448u>
 67. Zhang H, Sun Z, Wang K, Li N, Chen H, Tan X, Li L, He Z, Sun J (2018) Multifunctional tumor-targeting cathepsin B-sensitive gemcitabine prodrug covalently targets albumin in situ and improves cancer therapy. *Bioconjug Chem* 29(6):1852–1858. <https://doi.org/10.1021/acs.bioconjchem.8b00223>
 68. Wu W, Luo Y, Sun C, Liu Y, Kuo P, Varga J, Xiang R, Reisfeld R, Janda KD, Edgington TS, Liu C (2006) Targeting cell-impermeable prodrug activation to tumor microenvironment eradicates multiple drug-resistant neoplasms. *Can Res* 66(2):970. <https://doi.org/10.1158/0008-5472.CAN-05-2591>
 69. Seo K, Chung SW, Byun Y, Kim D (2012) Paclitaxel loaded nano-aggregates based on pH sensitive polyaspartamide amphiphilic graft copolymers. *Int J Pharm* 424(1–2):26–32. <https://doi.org/10.1016/j.ijpharm.2011.12.047>
 70. Yoshida T, Lai TC, Kwon GS, Sako K (2013) pH- and ion-sensitive polymers for drug delivery. *Expert Opin Drug Deliv* 10(11):1497–1513. <https://doi.org/10.1517/17425247.2013.821978>
 71. Xiong S, Wang Z, Liu J, Deng X, Xiong R, Cao X, Xie Z, Lei X, Chen Y, Tang G (2019) A pH-sensitive prodrug strategy to co-deliver DOX and TOS in TPGS nanomicelles for tumor therapy. *Colloids Surf B Biointerfaces* 173:346–355. <https://doi.org/10.1016/j.colsurfb.2018.10.012>
 72. Huang X, Liao W, Xie Z, Chen D, Zhang CY (2018) A pH-responsive prodrug delivery system self-assembled from acid-labile doxorubicin-conjugated amphiphilic pH-sensitive block copolymers. *Mater Sci Eng C Mater Biol Appl* 90:27–37. <https://doi.org/10.1016/j.msec.2018.04.036>
 73. Rahoui N, Jiang B, Taloub N, Hegazy M, Huang YD (2018) Synthesis and evaluation of water soluble pH sensitive poly(vinyl alcohol)-doxorubicin conjugates. *J Biomater Sci Polym Ed* 29(12):1482–1497. <https://doi.org/10.1080/09205063.2018.1466470>
 74. Zhu J, Huo Q, Xu M, Yang F, Li Y, Shi H, Niu Y, Liu Y (2018) Bortezomib-catechol conjugated prodrug micelles: combining bone targeting and aryl boronate-based pH-responsive drug release for cancer bone-metastasis therapy. *Nanoscale* 10(38):18387–18397. <https://doi.org/10.1039/c8nr03899f>
 75. Xie J, Fan Z, Li Y, Zhang Y, Yu F, Su G, Xie L, Hou Z (2018) Design of pH-sensitive methotrexate prodrug-targeted curcumin nanoparticles for efficient dual-drug delivery and combination cancer therapy. *Int J Nanomed* 13:1381–1398. <https://doi.org/10.2147/IJN.S152312>
 76. Tap WD, Papai Z, Van Tine BA, Attia S, Ganjoo KN, Jones RL, Schuetz S, Reed D, Chawla SP, Riedel RF, Krarup-Hansen A, Toulmonde M, Ray-Coquard I, Hohenberger P, Grignani G, Cranmer LD, Okuno S, Agulnik M, Read W, Ryan CW, Alcindor T, del Muro XFG, Budd GT, Tawbi H, Pearce T, Kroll S, Reinke DK, Schöffski P (2017) Doxorubicin plus evofosfamide versus doxorubicin alone in locally advanced, unresectable or metastatic soft-tissue sarcoma (TH CR-406/SARC021): an international, multicentre, open-label, randomised phase 3 trial. *Lancet Oncol* 18(8):1089–1103. [https://doi.org/10.1016/s1470-2045\(17\)30381-9](https://doi.org/10.1016/s1470-2045(17)30381-9)
 77. Laubach JP, Liu CJ, Raje NS, Yee AJ, Armand P, Schlossman RL, Rosenblatt J, Hedlund J, Martin M, Reynolds C, Shain KH, Zackon I, Stampleman L, Henrick P, Rivotto B, Hornburg KTV, Dumke HJ, Chuma S, Savell A, Handisides DR, Kroll S, Anderson KC, Richardson PG, Ghobrial IM (2018) A phase I/II study of evofosfamide, a hypoxia-activated prodrug with or without bortezomib in subjects with relapsed/refractory multiple myeloma. *Clin Cancer Res*. <https://doi.org/10.1158/1078-0432.ccr-18-1325>
 78. Jamieson SM, Tsai P, Kondratyev MK, Budhani P, Liu A, Senzer NN, Chiorean EG, Jalal SI, Nemunaitis JJ, Kee D, Shome A, Wong WW, Li D, Poonawala-Lohani N, Kakadia PM, Knowlton NS, Lynch CR, Hong CR, Lee TW, Grenman RA, Caporiccio L, McKee TD, Zaidi M, Butt S, Macann AM, McIvor NP, Chaplin JM, Hicks KO, Bohlander SK, Wouters BG, Hart CP, Print CG, Wilson WR, Curran MA, Hunter FW (2018) Evofosfamide for the treatment of human papillomavirus-negative head and neck squamous cell carcinoma. *JCI Insight*. <https://doi.org/10.1172/jci.insight.122204>
 79. Kumar S, Sun JD, Zhang L, Mokhtari RB, Wu B, Meng F, Liu Q, Bhupathi D, Wang Y, Yeger H, Hart C, Baruchel S (2018) Hypoxia-targeting drug evofosfamide (TH-302) enhances sunitinib activity in neuroblastoma xenograft models. *Transl Oncol* 11(4):911–919. <https://doi.org/10.1016/j.tranon.2018.05.004>
 80. Liu S, Tetzlaff M, Wang T, Chen X, Yang R, Kumar SM, Xu X (2017) Hypoxia-activated prodrug enhances therapeutic effect

- of sunitinib in melanoma. *Oncotarget* 8(70):115140–115152. <https://doi.org/10.18632/oncotarget.22944>
81. Huang Y, Tian Y, Zhao Y, Xue C, Zhan J, Liu L, He X, Zhang L (2018) Efficacy of the hypoxia-activated prodrug evofosfamide (TH-302) in nasopharyngeal carcinoma in vitro and in vivo. *Cancer Commun (Lond)* 38(1):15. <https://doi.org/10.1186/s40880-018-0285-0>
 82. Brenner A, Zuniga R, Sun JD, Floyd J, Hart CP, Kroll S, Fichtel L, Cavazos D, Caffisch L, Gruslova A, Huang S, Liu Y, Lodi A, Tiziani S (2018) Hypoxia-activated evofosfamide for treatment of recurrent bevacizumab-refractory glioblastoma: a phase I surgical study. *Neurooncology* 20(9):1231–1239. <https://doi.org/10.1093/neuonc/noy015>
 83. Liapis V, Zysk A, DeNichilo M, Zinonos I, Hay S, Panagopoulos V, Shoubridge A, Difelice C, Ponomarev V, Ingman W, Atkins GJ, Findlay DM, Zannettino ACW, Evdokiou A (2017) Anticancer efficacy of the hypoxia-activated prodrug evofosfamide is enhanced in combination with proapoptotic receptor agonists against osteosarcoma. *Cancer Med* 6(9):2164–2176. <https://doi.org/10.1002/cam4.1115>
 84. Hajj C, Russell J, Hart CP, Goodman KA, Lowery MA, Haimovitz-Friedman A, Deasy JO, Humm JL (2017) A combination of radiation and the hypoxia-activated prodrug evofosfamide (TH-302) is efficacious against a human orthotopic pancreatic tumor model. *Transl Oncol* 10(5):760–765. <https://doi.org/10.1016/j.tranon.2017.06.010>
 85. Jin C, Zhang Q, Lu W (2017) Synthesis and biological evaluation of hypoxia-activated prodrugs of SN-38. *Eur J Med Chem* 132:135–141. <https://doi.org/10.1016/j.ejmech.2017.03.040>
 86. McKeage MJ, Gu Y, Wilson WR, Hill A, Amies K, Melink TJ, Jameson MB (2011) A phase I trial of PR-104, a prodrug of the bioreductive prodrug PR-104A, given weekly to solid tumour patients. *BMC Cancer* 11:432. <https://doi.org/10.1186/1471-2407-11-432>
 87. Guise CP, Abbattista MR, Singleton RS, Holford SD, Connolly J, Dachs GU, Fox SB, Pollock R, Harvey J, Guilford P, Donate F, Wilson WR, Patterson AV (2010) The bioreductive prodrug PR-104A is activated under aerobic conditions by human aldo-keto reductase 1C3. *Cancer Res* 70(4):1573–1584. <https://doi.org/10.1158/0008-5472.CAN-09-3237>
 88. Konopleva M, Thall PF, Yi CA, Borthakur G, Coveler A, Bueso-Ramos C, Benito J, Konoplev S, Gu Y, Ravandi F, Jabbour E, Faderl S, Thomas D, Cortes J, Kadia T, Kornblau S, Daver N, Pemmaraju N, Nguyen HQ, Feliu J, Lu H, Wei C, Wilson WR, Melink TJ, Gutheil JC, Andreeff M, Estey EH, Kantarjian H (2015) Phase I/II study of the hypoxia-activated prodrug PR104 in refractory/relapsed acute myeloid leukemia and acute lymphoblastic leukemia. *Haematologica* 100(7):927–934. <https://doi.org/10.3324/haematol.2014.118455>
 89. Peng X, Gandhi V (2012) ROS-activated anticancer prodrugs: a new strategy for tumor-specific damage. *Ther Deliv* 3(7):823–833. <https://doi.org/10.4155/tde.12.61>
 90. Saravanakumar G, Kim J, Kim Won J (2016) Reactive-oxygen-species-responsive drug delivery systems: promises and challenges. *Adv Sci* 4(1):1600124. <https://doi.org/10.1002/advs.201600124>
 91. Kuang Y, Balakrishnan K, Gandhi V, Peng X (2011) Hydrogen peroxide inducible DNA cross-linking agents: targeted anticancer prodrugs. *J Am Chem Soc* 133(48):19278–19281. <https://doi.org/10.1021/ja2073824>
 92. Wang L, Xie S, Ma L, Chen Y, Lu W (2016) 10-Boronic acid substituted camptothecin as prodrug of SN-38. *Eur J Med Chem* 116:84–89. <https://doi.org/10.1016/j.ejmech.2016.03.063>
 93. Zhang C, Zhong Q, Zhang Q, Zheng S, Miele L, Wang G (2015) Boronic prodrug of endoxifen as an effective hormone therapy for breast cancer. *Breast Cancer Res Treat* 152(2):283–291. <https://doi.org/10.1007/s10549-015-3461-9>
 94. Gabor F, Wollmann K, Theyer G, Haberl I, Hamilton G (1994) In vitro antiproliferative effects of albumin-doxorubicin conjugates against Ewing's sarcoma and peripheral neuroectodermal tumor cells. *Anticancer Res* 14(5A):1943–1950
 95. Kratz F (2007) DOXO-EMCH (INNO-206): the first albumin-binding prodrug of doxorubicin to enter clinical trials. *Expert Opin Investig Drugs* 16(6):855–866. <https://doi.org/10.1517/13543784.16.6.855>
 96. Chawla SP, Papai Z, Mukhametshina G, Sankhala K, Vasylyev L, Fedenko A, Khamly K, Ganjoo K, Nagarkar R, Wieland S, Levitt DJ (2015) First-line aldorubicin vs doxorubicin in metastatic or locally advanced unresectable soft-tissue sarcoma: a phase 2b randomized clinical trial. *JAMA Oncol* 1(9):1272–1280. <https://doi.org/10.1001/jamaoncol.2015.3101>
 97. Mita MM, Natale RB, Wolin EM, Laabs B, Dinh H, Wieland S, Levitt DJ, Mita AC (2015) Pharmacokinetic study of aldorubicin in patients with solid tumors. *Investig New Drugs* 33(2):341–348. <https://doi.org/10.1007/s10637-014-0183-5>
 98. Mansour AM, Dreves J, Esser N, Hamada FM, Badary OA, Unger C, Fichtner I, Kratz F (2003) A new approach for the treatment of malignant melanoma: enhanced antitumor efficacy of an albumin-binding doxorubicin prodrug that is cleaved by matrix metalloproteinase 2. *Can Res* 63(14):4062
 99. Schmid B, Chung DE, Warnecke A, Fichtner I, Kratz F (2007) Albumin-binding prodrugs of camptothecin and doxorubicin with an Ala-Leu-Ala-Leu-linker that are cleaved by cathepsin B: synthesis and antitumor efficacy. *Bioconjug Chem* 18(3):702–716. <https://doi.org/10.1021/bc0602735>
 100. Graeser R, Chung DE, Esser N, Moor S, Schachtele C, Unger C, Kratz F (2008) Synthesis and biological evaluation of an albumin-binding prodrug of doxorubicin that is cleaved by prostate-specific antigen (PSA) in a PSA-positive orthotopic prostate carcinoma model (LNCaP). *Int J Cancer* 122(5):1145–1154. <https://doi.org/10.1002/ijc.23050>
 101. Wunder A, Stehle G, Schrenk HH, Hartung G, Heene DL, Maier-Borst W, Sinn H (1998) Antitumor activity of methotrexate-albumin conjugates in rats bearing a Walker-256 carcinoma. *Int J Cancer* 76(6):884–890. [https://doi.org/10.1002/\(SICI\)1097-0215\(19980610\)76:6%3c884::AID-IJC19%3e3.0.CO;2-2](https://doi.org/10.1002/(SICI)1097-0215(19980610)76:6%3c884::AID-IJC19%3e3.0.CO;2-2)
 102. Hartung G, Stehle G, Sinn H, Wunder A, Schrenk HH, Heeger S, Kranzle M, Edler L, Frei E, Fiebig HH, Heene DL, Maier-Borst W, Queisser W (1999) Phase I trial of methotrexate-albumin in a weekly intravenous bolus regimen in cancer patients. Phase I Study Group of the Association for Medical Oncology of the German Cancer Society. *Clin Cancer Res* 5(4):753–759
 103. Vis AN, van der Gaast A, van Rhijn BW, Catsburg TK, Schmidt C, Mickisch GH (2002) A phase II trial of methotrexate-human serum albumin (MTX-HSA) in patients with metastatic renal cell carcinoma who progressed under immunotherapy. *Cancer Chemother Pharmacol* 49(4):342–345. <https://doi.org/10.1007/s00280-001-0417-z>
 104. Mayr J, Heffeter P, Groza D, Galvez L, Koellensperger G, Roller A, Alte B, Haider M, Berger W, Kowol CR, Keppler BK (2017) An albumin-based tumor-targeted oxaliplatin prodrug with distinctly improved anticancer activity in vivo. *Chem Sci* 8(3):2241–2250. <https://doi.org/10.1039/c6sc03862j>
 105. Beck A, D'Atri V, Ehkirch A, Fekete S, Hernandez-Alba O, Gahoual R, Leize-Wagner E, Francois Y, Guillarme D, Cianferani S (2019) Cutting-edge multi-level analytical and structural characterization of antibody–drug conjugates: present and future. *Expert Rev Proteom* 16(4):337–362. <https://doi.org/10.1080/14789450.2019.1578215>
 106. Jen EY, Ko CW, Lee JE, Del Valle PL, Aydanian A, Jewell C, Norsworthy KJ, Przepiorcka D, Nie L, Liu J, Sheth CM,

- Shapiro M, Farrell AT, Pazdur R (2018) FDA approval: gemtuzumab ozogamicin for the treatment of adults with newly diagnosed CD33-positive acute myeloid leukemia. *Clin Cancer Res* 24(14):3242–3246. <https://doi.org/10.1158/1078-0432.CCR-17-3179>
107. Tack DK, Letendre L, Kamath PS, Tefferi A (2001) Development of hepatic veno-occlusive disease after Mylotarg infusion for relapsed acute myeloid leukemia. *Bone Marrow Transplant* 28(9):895–897. <https://doi.org/10.1038/sj.bmt.1703242>
 108. Lu J, Jiang F, Lu A, Zhang G (2016) Linkers having a crucial role in antibody–drug conjugates. *Int J Mol Sci* 17(4):561. <https://doi.org/10.3390/ijms17040561>
 109. Junttila TT, Li G, Parsons K, Phillips GL, Sliwkowski MX (2011) Trastuzumab-DM1 (T-DM1) retains all the mechanisms of action of trastuzumab and efficiently inhibits growth of lapatinib insensitive breast cancer. *Breast Cancer Res Treat* 128(2):347–356. <https://doi.org/10.1007/s10549-010-1090-x>
 110. Girish S, Gupta M, Wang B, Lu D, Krop IE, Vogel CL, Burris Iii HA, LoRusso PM, Yi J-H, Saad O, Tong B, Chu Y-W, Holden S, Joshi A (2012) Clinical pharmacology of trastuzumab emtansine (T-DM1): an antibody–drug conjugate in development for the treatment of HER2-positive cancer. *Cancer Chemother Pharmacol* 69(5):1229–1240. <https://doi.org/10.1007/s00280-011-1817-3>
 111. Joubert N, Denevault-Sabourin C, Bryden F, Viaud-Massuard MC (2017) Towards antibody–drug conjugates and prodrug strategies with extracellular stimuli-responsive drug delivery in the tumor microenvironment for cancer therapy. *Eur J Med Chem* 142:393–415. <https://doi.org/10.1016/j.ejmech.2017.08.049>
 112. Chen H, Lin Z, Arnst KE, Miller DD, Li W (2017) Tubulin inhibitor-based antibody–drug conjugates for cancer therapy. *Molecules*. <https://doi.org/10.3390/molecules22081281>
 113. Riechelmann H, Sauter A, Golze W, Hanft G, Schroen C, Hoermann K, Erhardt T, Gronau S (2008) Phase I trial with the CD44v6-targeting immunoconjugate bivatuzumab mertansine in head and neck squamous cell carcinoma. *Oral Oncol* 44(9):823–829. <https://doi.org/10.1016/j.oraloncology.2007.10.009>
 114. Dal Corso A, Cazzamalli S, Gebleux R, Mattarella M, Neri D (2017) Protease-cleavable linkers modulate the anticancer activity of noninternalizing antibody–drug conjugates. *Bioconjug Chem* 28(7):1826–1833. <https://doi.org/10.1021/acs.bioconjchem.7b00304>
 115. Perrino E, Steiner M, Krall N, Bernardes GJ, Pretto F, Cusi G, Neri D (2014) Curative properties of noninternalizing antibody–drug conjugates based on maytansinoids. *Cancer Res* 74(9):2569–2578. <https://doi.org/10.1158/0008-5472.CAN-13-2990>
 116. Richards DA (2018) Exploring alternative antibody scaffolds: antibody fragments and antibody mimics for targeted drug delivery. *Drug Discov Today Technol* 30:35–46. <https://doi.org/10.1016/j.ddtec.2018.10.005>
 117. Aubrey N, Allard-Vannier E, Martin C, Bryden F, Letast S, Colas C, Lakhfir Z, Collinet N, Dimier-Poisson I, Chourpa I, Viaud-Massuard MC, Joubert N (2018) Site-specific conjugation of auristatins onto engineered scFv using second generation maleimide to target HER2-positive breast cancer in vitro. *Bioconjug Chem* 29(11):3516–3521. <https://doi.org/10.1021/acs.bioconjchem.8b00668>
 118. Desgrosellier JS, Cheresh DA (2010) Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer* 10(1):9–22. <https://doi.org/10.1038/nrc2748>
 119. Dechantsreiter MA, Planker E, Matha B, Lohof E, Holzemann G, Jonczyk A, Goodman SL, Kessler H (1999) *N*-Methylated cyclic RGD peptides as highly active and selective alpha(V) beta(3) integrin antagonists. *J Med Chem* 42(16):3033–3040. <https://doi.org/10.1021/jm970832g>
 120. Massaguer A, Gonzalez-Canto A, Escribano E, Barrabes S, Artigas G, Moreno V, Marchan V (2015) Integrin-targeted delivery into cancer cells of a Pt(IV) pro-drug through conjugation to RGD-containing peptides. *Dalton Trans* 44(1):202–212. <https://doi.org/10.1039/c4dt02710h>
 121. Dal Pozzo A, Esposito E, Ni M, Muzi L, Pisano C, Bucci F, Vesce L, Castorina M, Penco S (2010) Conjugates of a novel 7-substituted camptothecin with RGD-peptides as alpha(v)beta(3) integrin ligands: an approach to tumor-targeted therapy. *Bioconjug Chem* 21(11):1956–1967. <https://doi.org/10.1021/bc100097r>
 122. Arosio D, Casagrande C (2016) Advancement in integrin facilitated drug delivery. *Adv Drug Deliv Rev* 97:111–143. <https://doi.org/10.1016/j.addr.2015.12.001>
 123. Zhong P, Gu X, Cheng R, Deng C, Meng F, Zhong Z (2017) alpha(v)beta(3) integrin-targeted micellar mertansine prodrug effectively inhibits triple-negative breast cancer in vivo. *Int J Nanomed* 12:7913–7921. <https://doi.org/10.2147/IJN.S146505>
 124. Engel J, Emons G, Pinski J, Schally AV (2012) AEZS-108: a targeted cytotoxic analog of LHRH for the treatment of cancers positive for LHRH receptors. *Expert Opin Investig Drugs* 21(6):891–899. <https://doi.org/10.1517/13543784.2012.685128>
 125. Emons G, Gorchev G, Sehouli J, Wimberger P, Stahle A, Hanke L, Hilpert F, Sindermann H, Grundker C, Harter P (2014) Efficacy and safety of AEZS-108 (INN: zoptarelin doxorubicin acetate) an LHRH agonist linked to doxorubicin in women with platinum refractory or resistant ovarian cancer expressing LHRH receptors: a multicenter phase II trial of the ago-study group (AGO GYN 5). *Gynecol Oncol* 133(3):427–432. <https://doi.org/10.1016/j.ygyno.2014.03.576>
 126. Liu SV, Tsao-Wei DD, Xiong S, Groshen S, Dorff TB, Quinn DI, Tai YC, Engel J, Hawes D, Schally AV, Pinski JK (2014) Phase I, dose-escalation study of the targeted cytotoxic LHRH analog AEZS-108 in patients with castration- and taxane-resistant prostate cancer. *Clin Cancer Res* 20(24):6277–6283. <https://doi.org/10.1158/1078-0432.CCR-14-0489>
 127. Yu SS, Athreya K, Liu SV, Schally AV, Tsao-Wei D, Groshen S, Quinn DI, Dorff TB, Xiong S, Engel J, Pinski J (2017) A phase II trial of AEZS-108 in castration- and taxane-resistant prostate cancer. *Clin Genitourin Cancer* 15(6):742–749. <https://doi.org/10.1016/j.clgc.2017.06.002>
 128. Karampelas T, Skavatsou E, Argyros O, Fokas D, Tamvakopoulos C (2017) Gemcitabine based peptide conjugate with improved metabolic properties and dual mode of efficacy. *Mol Pharm* 14(3):674–685. <https://doi.org/10.1021/acs.molpharmaceut.6b00961>
 129. Kurzrock R, Gabrail N, Chandhasin C, Moulder S, Smith C, Brenner A, Sankhala K, Mita A, Elian K, Bouchard D, Sarantopoulos J (2012) Safety, pharmacokinetics, and activity of GRN1005, a novel conjugate of angiopoep-2, a peptide facilitating brain penetration, and paclitaxel, in patients with advanced solid tumors. *Mol Cancer Ther* 11(2):308–316. <https://doi.org/10.1158/1535-7163.MCT-11-0566>
 130. Drappatz J, Brenner A, Wong ET, Eichler A, Schiff D, Groves MD, Mikkelsen T, Rosenfeld S, Sarantopoulos J, Meyers CA, Fielding RM, Elian K, Wang X, Lawrence B, Shing M, Kelsey S, Castaigne JP, Wen PY (2013) Phase I study of GRN1005 in recurrent malignant glioma. *Clin Cancer Res* 19(6):1567–1576. <https://doi.org/10.1158/1078-0432.CCR-12-2481>
 131. Gelderblom H, Verweij J, Nooter K, Sparreboom A (2001) Cremophor EL: the drawbacks and advantages of vehicle selection for drug formulation. *Eur J Cancer* 37(13):1590–1598. [https://doi.org/10.1016/S0959-8049\(01\)00171-X](https://doi.org/10.1016/S0959-8049(01)00171-X)
 132. Huo M, Zhu Q, Wu Q, Yin T, Wang L, Yin L, Zhou J (2015) Somatostatin receptor-mediated specific delivery of paclitaxel prodrugs for efficient cancer therapy. *J Pharm Sci* 104(6):2018–2028. <https://doi.org/10.1002/jps.24438>

133. Steinberg G, Borch RF (2001) Synthesis and evaluation of ptericoic acid-conjugated nitroheterocyclic phosphoramidates as folate receptor-targeted alkylating agents. *J Med Chem* 44(1):69–73. <https://doi.org/10.1021/jm000306g>
134. Aronov O, Horowitz AT, Gabizon A, Gibson D (2003) Folate-targeted PEG as a potential carrier for carboplatin analogs. Synthesis and in vitro studies. *Bioconjug Chem* 14(3):563–574. <https://doi.org/10.1021/bc025642i>
135. Liu J, Kolar C, Lawson TA, Gmeiner WH (2002) Targeted drug delivery to chemoresistant cells: folic acid derivatization of FdUMP[10] enhances cytotoxicity toward 5-FU-Resistant human colorectal tumor cells. *J Org Chem* 67(8):2734. <https://doi.org/10.1021/jo01619-6>
136. Lee JW, Lu JY, Low PS, Fuchs PL (2002) Synthesis and evaluation of taxol–folic acid conjugates as targeted antineoplastics. *Bioorg Med Chem* 10(7):2397–2414. [https://doi.org/10.1016/S0968-0896\(02\)00019-6](https://doi.org/10.1016/S0968-0896(02)00019-6)
137. Ladino CA, Chari RV, Bourret LA, Kedersha NL, Goldmacher VS (1997) Folate-maytansinoids: target-selective drugs of low molecular weight. *Int J Cancer* 73(6):859–864. [https://doi.org/10.1002/\(SICI\)1097-0215\(19971210\)73:6%3c859:AID-IJC16%3e3.0.CO;2-%23](https://doi.org/10.1002/(SICI)1097-0215(19971210)73:6%3c859:AID-IJC16%3e3.0.CO;2-%23)
138. Reddy JA, Westrick E, Santhapuram HK, Howard SJ, Miller ML, Vetzal M, Vlahov I, Chari RV, Goldmacher VS, Leamon CP (2007) Folate receptor-specific antitumor activity of EC131, a folate-maytansinoid conjugate. *Cancer Res* 67(13):6376–6382. <https://doi.org/10.1158/0008-5472.CAN-06-3894>
139. Li J, Sausville EA, Klein PJ, Morgenstern D, Leamon CP, Messmann RA, LoRusso P (2009) Clinical pharmacokinetics and exposure-toxicity relationship of a folate-Vinca alkaloid conjugate EC145 in cancer patients. *J Clin Pharmacol* 49(12):1467–1476. <https://doi.org/10.1177/0091270009339740>
140. Guertin AD, O’Neil J, Stoeck A, Reddy JA, Cristescu R, Haines BB, Hinton MC, Dorton R, Bloomfield A, Nelson M, Vetzal M, Lejnine S, Nebozhyn M, Zhang T, Loboda A, Picard KL, Schmidt EV, Dussault I, Leamon CP (2016) High levels of expression of p-glycoprotein/multidrug resistance protein result in resistance to vintafolide. *Mol Cancer Ther* 15(8):1998–2008. <https://doi.org/10.1158/1535-7163.MCT-15-0950>
141. Shan L, Zhuo X, Zhang F, Dai Y, Zhu G, Yung BC, Fan W, Zhai K, Jacobson O, Kiesewetter DO, Ma Y, Gao G, Chen X (2018) A paclitaxel prodrug with bifunctional folate and albumin binding moieties for both passive and active targeted cancer therapy. *Theranostics* 8(7):2018–2030. <https://doi.org/10.7150/thno.24382>
142. Liu Z, Xiong M, Gong J, Zhang Y, Bai N, Luo Y, Li L, Wei Y, Liu Y, Tan X, Xiang R (2014) Legumain protease-activated TAT-liposome cargo for targeting tumours and their microenvironment. *Nat Commun* 5:4280. <https://doi.org/10.1038/ncomms5280>. <https://www.nature.com/articles/ncomms5280#supplementary-information>
143. Zhang Y, Huang F, Ren C, Yang L, Liu J, Cheng Z, Chu L, Liu J (2017) Targeted chemo-photodynamic combination platform based on the DOX prodrug nanoparticles for enhanced cancer therapy. *ACS Appl Mater Interfaces* 9(15):13016–13028. <https://doi.org/10.1021/acsami.7b00927>
144. Regina A, Demeule M, Che C, Lavallee I, Poirier J, Gabathuler R, Beliveau R, Castaigne JP (2008) Antitumour activity of ANG1005, a conjugate between paclitaxel and the new brain delivery vector Angiopep-2. *Br J Pharmacol* 155(2):185–197. <https://doi.org/10.1038/bjp.2008.260>
145. Reddy JA, Dorton R, Bloomfield A, Nelson M, Dircksen C, Vetzal M, Kleindl P, Santhapuram H, Vlahov IR, Leamon CP (2018) Pre-clinical evaluation of EC1456, a folate-tubulysin anti-cancer therapeutic. *Sci Rep* 8(1):8943. <https://doi.org/10.1038/s41598-018-27320-5>

Publisher’s Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.