



Resolution of Crohn's disease

Heike Schmitt¹ · Clemens Neufert¹ · Markus F. Neurath¹ · Raja Atreya¹

Received: 12 August 2019 / Accepted: 5 September 2019 / Published online: 24 September 2019
© Springer-Verlag GmbH Germany, part of Springer Nature 2019

Abstract

Crohn's disease (CD) is characterized by chronic inflammation of the gastrointestinal tract and represents one of the main inflammatory bowel disease (IBD) forms. The infiltration of immune cells into the mucosa and uncontrolled production of pro-inflammatory cytokines and other mediators trigger the chronic inflammatory reaction in the intestine [1]. The inflammatory setting consists of subsequent events that comprise an induction phase, the peak of inflammation which is subsequently followed by the resolution phase. The induction phase, which represents the first phase of inflammation, is important for the rapid and efficient activation of the immune system for sufficient host defense. The permanent sensing of exogenous or endogenous danger signals enables the fast initiation of the inflammatory reaction. The immune cell infiltrate initiates an inflammatory cascade where released lipid and protein mediators play an indispensable role [2, 3]. The last decades of research strongly suggest that resolution of inflammation is similarly a tightly coordinated and active process. The basic concept that resolution of inflammation has to be regarded as an active process has been thoroughly described by others [4–6]. The following review focuses on mechanisms, pathways, and specific mediators that are actively involved in the resolution of inflammation in CD.

Keywords Resolution of inflammation · Crohn's disease · Intestine · IL23 · Anti-TNF therapy · Apoptosis

Crohn's disease

The precise etiology of CD has not yet been fully elucidated, but current immunopathogenic concepts strongly indicate that mucosal inflammation is caused by an altered immune response in individuals with a genetic predisposition. This inflammatory reaction is believed to be a consequence of a multifaceted interaction among environmental factors, luminal antigens, and the respective mucosal immune system [7–9]. CD is characterized by segmental and transmural inflammation that can principally occur in all parts of the gastrointestinal tract, with the terminal ileum and colon representing the most often affected segments. Clinical symptoms are defined by disease location and behavior as well as severity of inflammation. They frequently comprise abdominal cramping, persistent diarrhea, severe fecal urgency, fatigue, anemia, and weight loss [10]. Up to 50% of patients are affected by skin,

joint, or eye extra-intestinal manifestations that often occur before the diagnosis of CD is made. Extra-intestinal manifestation can moreover also occur at other parts of the body (e.g., lung) [11]. Bowel damage and disability are also attendant symptoms in CD and reflect the progressive and debilitating course of disease in this IBD entity. The inflamed intestine may cause various complications such as formation of abscesses, stenoses, fistulae, and heightened incidence of colitis-associated neoplasias [10]. Therefore, CD patients often have a limited quality of life and a pronounced morbidity is furthermore linked with CD. Recently, it has been shown that CD is characterized by a rapidly increasing incidence worldwide, which is especially evident in newly industrialized countries, making CD a global disease. Europe (322 per 100,000 in Germany) and North America (319 per 100,000 in Canada) have shown the highest prevalence, but the steady increase in areas with previously low incidence is best reflected in Africa, South America, and Asia, where rising annual percentage changes have been recently recorded [12].

In CD, there is a dire need for rational and optimized anti-inflammatory therapy to prevent progression and subsequent complications of disease [13]. Earlier treatment goals mainly focused on the alleviation of clinical symptoms alone, but in the course of time and development of novel targeted therapies, clinical remission, steroid-free remission, and finally

This article is a contribution to the special issue on Resolution of Inflammation in Chronic Diseases - Guest Editor: Markus Neurath

✉ Raja Atreya
raja.atreya@uk-erlangen.de

¹ First Department of Medicine, Friedrich-Alexander-University Erlangen-Nürnberg, Ulmenweg 18, 91054 Erlangen, Germany

mucosal healing have become an integral part in the successful therapeutic management of CD. Transmural healing assessed in cross-sectional imaging and the inclusion of a validated histological scoring instrument into the definition of mucosal healing could determine future therapeutic goals [14]. The most widely used therapies in CD consisted of corticosteroids and immunosuppressants (thiopurines and methotrexate). A major breakthrough was achieved by the development of biological therapies that are directed against specific mediators of the inflammatory process. The first substance class to be approved for therapy of CD were anti-TNF antibodies (infliximab, adalimumab, and certolizumab pegol). Antibodies against the integrin alpha4beta7 (vedolizumab) and interleukin 12 and interleukin 23 through their common p40 subunit (ustekinumab) have been approved for CD therapy in the following years [15, 16]. Despite the large clinical success of the mentioned targeted therapies, it was found that only subgroups of CD patients will benefit from these treatments, while a substantial proportion of patients do not show signs of clinical response. Furthermore, there are currently no clinically compatible predictive biomarkers for an individual guidance of drug therapy. It is therefore of utmost clinical importance to gain a deeper understanding of the respective mode of action of the respective therapies to guarantee the most efficient and suitable therapy for each single patient [17].

The impact of anti-TNF therapy in the resolution of inflammation

IL-23 and TNF are among the main mediators produced by CD14⁺ intestinal macrophages which infiltrate into the inflamed intestine of CD patients [18, 19]. In particular, mucosal immune cells like macrophages and T cells have been linked with being actively involved in the maintenance of intestinal inflammation in CD. The involved T cells are characterized by enhanced proliferation, resistance against apoptosis, and excessive pro-inflammatory cytokine production [8, 20, 21]. Resistance against apoptosis leads to the accumulation of pro-inflammatory cells that are thus involved in the initiation and perpetuation of the inflammatory reaction. Caspases are a protease family that are strongly involved in the induction of apoptosis [22, 23]. TNF is on the one hand able to induce survival and proliferation of cells via the NF- κ B signaling pathway, but on the other hand is also able to initiate the induction of apoptosis via regulation of caspases [24] (Fig. 1).

Anti-TNF agents are commonly used as a highly efficient therapy in CD, underlining the pivotal importance of cytokines like TNF in the immunopathogenesis of the disease [25–27]. Although they are often effective in the resolution of CD, leading to endoscopic remission (Fig. 2a), they however fail in a relevant subgroup of patients, resulting in ongoing inflammation with signs of endoscopic disease activity

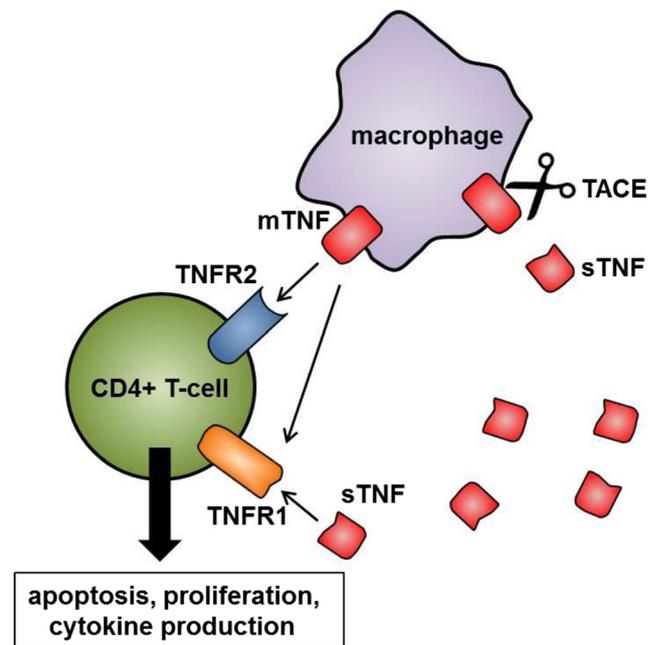


Fig. 1 TNF signaling mechanism. TNF is initially expressed as a membrane bound form (mTNF) by immune cells like macrophages. mTNF can be processed by the TNF alpha converting enzyme (TACE) into a soluble form (sTNF) which is able to activate both TNFR1 and TNFR2, whereas sTNF mainly activates TNFR1 on immune cells like CD4⁺ T cells. The binding of mTNF or sTNF to TNFR1 or TNFR2 can lead to apoptosis, proliferation, and different cytokine production

(Fig. 2b). About one third of CD patients do not demonstrate therapeutic efficacy upon initiated anti-TNF therapy (primary non-response); moreover, 30%–50% lose response to therapy in the course of treatment (secondary non-response) [28, 29].

Secondary non-response to anti-TNF therapy has so far primarily been associated with immunogenicity. Interestingly, the application of another anti-TNF antibody in these patients often also fails to show therapeutic efficacy [30, 31]. Besides neutralizing antibodies against the anti-TNF agents, insufficient levels of the drug in the serum has also been considered to explain inefficient therapeutic response [30]. In contrast, a recently published study demonstrated that a large cohort of patients who lose response to anti-TNF therapy exhibit sufficient levels of anti-TNF antibody in the serum and do not develop relevant levels of neutralizing antibodies against TNF [32]. These results strongly suggest that in addition to pharmacodynamic factors, existence of TNF-independent forms of CD that circumvent resolution of inflammation might similarly be involved in failure of anti-TNF agent therapy. Although some reasons for therapeutic non-response have been characterized in the last years, the concept of immune-related mechanisms that drive resistance to biologic therapy in IBD and thus blocking the resolution of inflammation in CD is still poorly understood. Here, studies in recent years have demonstrated that expression of the target molecule in the inflamed tissue may have a pivotal influence on the response to anti-TNF therapy in IBD. It could

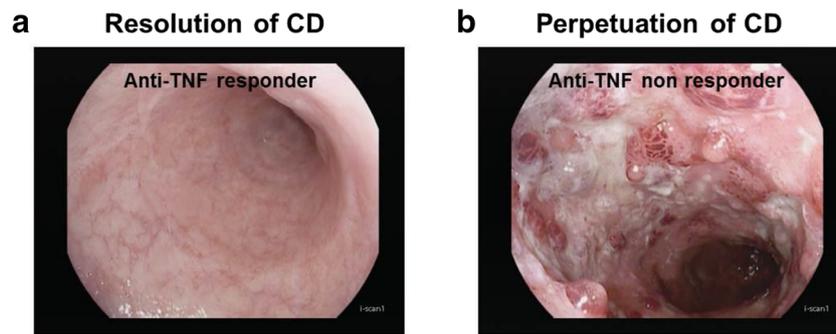


Fig. 2 Endoscopic resolution and perpetuation of mucosal inflammation in CD patients treated with an anti-TNF agent. (A) Patient with Crohn's colitis demonstrating resolution of inflammation after initiated anti-TNF

therapy. (B) Anti-TNF antibody treated patient with Crohn's colitis demonstrating signs of severe mucosal inflammation during ongoing anti-TNF therapy

be shown that anti-TNF antibodies are able to bind to membrane-bound TNF (mTNF) expressing macrophages, which leads to the induction of apoptosis in TNFR2 expressing mucosal T cells that lack the mTNF-dependent co-stimulation [33]. The interplay between mTNF (macrophages) and TNFR2 (T cells) therefore represents a potential component in the development of resistance to apoptosis in mucosal T cells in CD, which critically contributes to disease pathogenesis in CD. The importance of this mechanism is supported by the finding of an *in vivo* molecular endoscopy study in 25 CD patients, where high mucosal expression of mTNF before initiation of anti-TNF therapy was linked to higher clinical and endoscopic response rates to following anti-TNF therapy [34, 35]. Furthermore, other findings have similarly implicated that immune signaling pathways may predict and control responsiveness to anti-TNF therapy in IBD [17]. The release of cytokines and resulting different cytokine levels in the inflamed mucosa of CD patients has thus been moved into the focus as a possible reason for insufficient response rates to anti-TNF therapy. Besides elevated levels of IL1B, IL17A, and S100A8 in CD patients who do not respond to anti-TNF therapy, altered levels of IL-6 and IL-23p19 have similarly been found in CD non-responders [36]. Another study elegantly demonstrated that elevated pretreatment expression of intestinal oncostatin-M and mucosal plasma cells, as well as inflammatory macrophages were also linked to lack of efficient anti-TNF therapy [37, 38]. These findings indicate that immune cells and the interplay of cytokines in the inflamed mucosa of CD patients have to be regarded as critical components for the resolution of inflammation process.

Regulatory T cells as mediators for the resolution of CD

Regulatory T cells are mainly divided into classically activated regulatory T cells (Tregs) and type 1 regulatory T cells (Tr1). Tregs can be identified via the transcription factor FoxP3, while Tr1 cells are characterized by the production of high levels of IL-10 [39, 40]. The last years of research have

focused on the role of Tregs and Tr1 cells in tolerance and immune balance, but little is known about the role of these cells in the resolution of inflammation process [41]. Elevated levels of IL-10 and diminished IL-17A and ROR γ t levels can influence the differentiation of pro-inflammatory Th17 into anti-inflammatory Tr1 cells and therefore have an impact on the resolution of inflammation cascade [42, 43]. Alternative immunotherapeutic approaches represent a novel part of therapeutic research in CD. In a pilot study, antigen specific T cells, which were generated *ex vivo*, were administered intravenously and could reach a dose-dependent therapeutic effect in CD patients, which was accompanied by a favorable safety profile [44]. These findings suggest that changes in T-cell plasticity play a central role in the resolution of inflammation and might be an interesting target for future CD therapy.

Regulatory innate lymphoid cells (ILCreg) are important for intestinal barrier function

Different types of immune cells contribute to the resolution of inflammation process in CD. Activated ILCreg cells are characterized by the secretion of IL-10 and TGF beta while non-activated ILCregs are normally found as resting cells in the lamina propria of the murine and human intestine. The secretion of anti-inflammatory cytokines by ILCregs influences the release of IFN- γ and IL-17 but does not alter the production of IL22, a cytokine which together with IL-10 was shown to be an integral part in the resolution of inflammation conveyed by T cells. The role of IL-10 in CD is controversially discussed. Early-onset CD is linked to genetic variants in the IL-10 receptor, whereas therapeutic administration of IL-10 was not able to induce clinical efficacy [45–47]. The cytokine IL-22 is involved in the induction of proliferative and anti-apoptotic signals and thereby contributes to the restoration of the barrier function in the adaptive and innate immune system [48, 49]. Further cytokines that show similar effects are IL-28 and IL-36 and together with IL-22, they are able to foster resolution of inflammation [50, 51].

Monocytes/macrophages and neutrophils are key players in the innate immune response

T cells as well as monocytes/macrophages are centrally involved in the pathogenesis of CD, but especially tissue macrophages also influence resolution of inflammation [52–54]. M2-like macrophages produce anti-inflammatory cytokines like IL-10 and TGF- β and are therefore important for the homeostasis in the healthy mucosa. Regarding the inflamed mucosa of IBD patients, elevated cytokine levels of IL-23, IL-6, TNF- α , or IL-1 β are found, which are mainly produced by pro-inflammatory M1 like macrophages [55, 56]. These macrophages are responsible for the phagocytosis of apoptotic cells like neutrophils which accumulate in the inflamed tissue and trigger, if not properly cleared, the perpetuation of inflammation and tissue damage [57]. Phosphatidylserine (PS) is normally located in the inner layer of the membrane of immune cells, but in apoptotic cells, PS can be found outside the cell, where it interacts with different receptors to mediate efferocytosis [58, 59]. A study where mucosal barrier function was mechanically broken revealed a diminished infiltration of neutrophils in CD patients [60]. Missing sensor signals and diminished migration of neutrophils might have an impact on the recruitment of phagocytic cells and therefore contribute to the perpetuation of inflammation in CD.

IL-23 causes perpetuation of inflammation in CD

Different studies revealed that besides TNF, other cytokines are also important players in the development of CD. It has been demonstrated that especially IL-23 is centrally involved in the pathogenesis of CD [1, 61–63]. Polymorphisms in genes encoding the IL-23R have been analyzed in genome-wide association studies (GWAS) and were linked to IBD pathogenesis, indicating a central role of IL-23 in mucosal inflammation [64, 65]. Moreover, elevated levels of IL-23 in the mucosa of CD patients further emphasize its critical role in IBD pathogenesis [66]. Recently, published data demonstrate that treatment with the selective IL-23p19 inhibitors risankizumab or brazikumab result in high response rates in the cohort of CD patients that did not respond to previous anti-TNF therapy [67, 68]. These findings strongly indicate involvement of IL-23 in anti-TNF-resistant gut inflammation. This is supported by a recently published study, where mucosal TNFR2-expressing CD4⁺ T cells circumvent anti-TNF induced apoptosis through co-expression of the IL-23R, which is activated by upregulated IL-23 production of mucosal CD14⁺ macrophages. Here, IL-23 caused activation of pSTAT3 in mucosal CD4⁺ T cells which results in resistance to apoptotic signals. The activated T cells are characterized by the release of high amounts of Th1 and Th17 cytokines. These

TNFR2⁺IL-23R⁺ T cells expand and accumulate in the mucosa of anti-TNF refractory CD patients where they perpetuate chronic intestinal inflammation (Fig. 3) [69].

However, the mechanisms by which the IL-23 may influence effector functions and pathways contributing to resolution of inflammation in CD are currently not fully understood.

Pro-resolving lipid molecules (PLMs)

In contrast to the initiation of inflammation, which has been meticulously investigated during the past four decades, resolution of inflammation is far less well understood [53, 70]. Although concepts describing resolution of inflammation as a passive process have been largely banished, the characterization of pathways that are activated during the resolution of inflammation process, and thereby orchestrate programs which terminate inflammation, has remained incomplete [71]. Resolution of inflammation includes several active processes rather than a stepwise disappearance of pro-inflammatory factors and mediators or dilution of chemokine gradients [72–74]. Serhan and others have highlighted the importance of “pro-resolving” factors in the context of resolution of inflammation [6, 75]. Pro-resolving lipid mediators are produced by a concerted action of arachidonate lipoxygenases (ALOX), mediating the production of pro-resolving lipid mediators. These are mainly found in alternatively activated macrophages and eosinophils [76, 77]. Among these pro-resolving molecules, those derived from essential polyunsaturated fatty acids (PUFAs) have been shown to induce favorable effects on a plethora of human inflammatory disorders, including CD. Although numerous pre-clinical and clinical studies tried to establish the therapeutic application of PUFAs in intestinal inflammation, either as fatty acid precursors or single metabolites, contradictory results have so far precluded final assessment of this treatment approach in the IBD field [78–80].

Microbiota

The gut microbiota represents another key component in the pathogenesis of IBD [81]. It comprises bacteria, fungi, viruses, and other microorganisms. The composition of the microbiota can be influenced by environmental factors, diet, and hygiene during childhood, whereas in adult persons a more stable and defined composition is evident [82, 83]. In healthy conditions, there is a homeostasis between the intestinal microbiome, mucosal barrier function, and the gut immune system. In CD, this homeostasis is altered, reflected by microbial imbalance, disrupted barrier function, and immune system activation [81]. The concept of a perturbed gut microbiota composition is

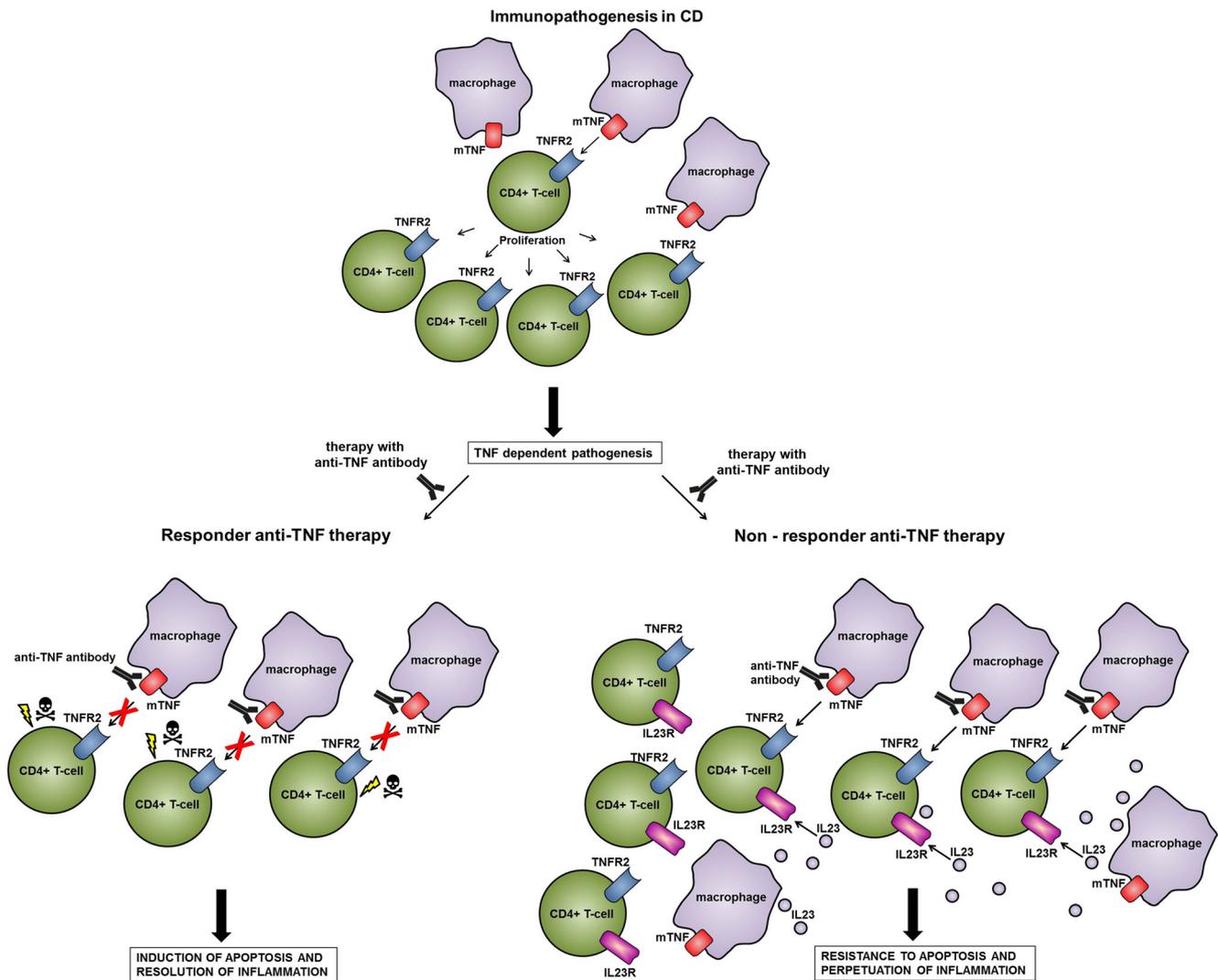


Fig. 3 IL-23 mediates resistance to apoptosis in CD. Model of IL-23-mediated resistance to apoptosis of mucosal CD4⁺ T cells in anti-TNF refractory CD patients. In CD, tissue-infiltrating CD14⁺ intestinal macrophages have been suggested to dominate the inflamed mucosa. Here, mTNF expressing CD14⁺ macrophages activate CD4⁺ T cells via the TNFR2, leading to mucosal inflammation in CD (upper image). In responders to initiated anti-TNF therapy, anti-TNF antibodies bind to mTNF-expressing CD14⁺ macrophages, thereby inhibiting activation of the TNFR2 on mucosal CD4⁺ T cells. The resulting blockade of TNFR2-dependent signaling pathways finally leads to the indirect induction of

apoptosis in intestinal CD4⁺ T cells in CD and resolution of inflammation (lower left image). In anti-TNF refractory patients, TNFR2 bearing gut CD4⁺ T cells additionally express the IL-23R. Heightened production of IL-23 from CD14⁺ macrophages leads to binding to the IL-23R on CD4⁺TNFR2⁺ T cells and induction of STAT3 activation. This activation leads to the expansion of CD4⁺IL-23R⁺TNFR2⁺ T cells that are resistant to apoptosis induction by anti-TNF antibodies, resulting in the perpetuation of mucosal inflammation (lower right image). Adapted from Schmitt et al. 2019 [69]

one of the striking findings in recent IBD research. Some changes of the microbiota seem to be characteristic for either CD or UC, while others appear to be attributed more generally to IBD. The most consistent changes can here be found regarding biodiversity [84–86]. In contrast to healthy controls, a reduced diversity in the fecal microbiome can be found in CD patients [87]. Even within the same patient, a reduction of diversity and a diminished bacterial load can be found in the inflamed tissue compared to unaffected tissue [88]. A multicenter study with samples collected from multiple concurrent

gastrointestinal locations revealed that changes in the bacterial composition (increased findings of Veillonellaceae, Pasteurellaceae, Enterobacteriaceae, and Fusobacteriaceae and decreased findings of Bacteroidales, Erysipelotrichales, and Clostridiales) strongly correlated with the level of mucosal inflammation of the disease. This study also revealed that rectal mucosa-associated microbiome profiling offered a reliable biomarker for the diagnosis of CD at the early stage of disease [89]. Antibiotics, probiotics, and prebiotics have been developed for the treatment of IBD with different results.

Whereas antibiotics at least achieved low effects in CD for a short time-span, studies for probiotics and prebiotics are generally disappointing and did not result in clinically relevant efficacy [90]. The use of single antibiotics can show a short-lived and modest therapeutic effect in CD at best, but has not been shown to be effective at all in patients with UC. Combinations of different antibiotics might slightly improve outcomes, but the long-term use of antibiotics may also lead to the development of antibiotic resistance in gut microbes [91, 92] and can altogether not be recommended for CD therapy. Studies have demonstrated that prognostic biomarkers from microbial profiling might be instrumental for personalized therapy. For example, microbial structure data in combination with apolipoprotein A1 levels can be used to predict steroid-free remission in children newly diagnosed with CD [93]. It was shown that ileal levels of *F. prausnitzii* increase the risk for postoperative recurrence of CD [94]. Furthermore, there is a correlation between microbial signatures and responsiveness to therapy and microbial imbalance is linked with relapse in patients after cessation of infliximab [95, 96]. The effect of an immunomodulatory therapy on the microbiota and its role in response to therapy is currently insufficiently studied and several studies showed controversial results. Sensing for pathogens which enter the gut due to a disrupted intestinal barrier is one important function of the mucosal immune system. A permanent influx of bacteria and bacterial antigens may contribute to the lack of resolution of inflammation and chronification of CD [97, 98].

Genetic risk factors and environmental influence on the resolution of CD

Although the pathophysiology of CD has not been fully clarified yet, it is currently assumed that a complex interaction between environmental, genetic, and intestinal microbial factors leads to a pathological auto-inflammatory response directed toward the intestine [99]. GWAS has become an integral part in the research field of multifactorial diseases like CD. Analysis of polymorphism or disease-susceptible genes with GWAS offers new possibilities to treat CD and to avoid the side effect of medications. There are several risk haplotypes which contribute to loss of efficacy. The HLA-DRB1*03 allele for example has also been associated with a sevenfold higher risk for the development of antibodies against infliximab [100]. Certain genes of the IL-23 pathway are IBD susceptibility genes, suggesting a critical role for this pathway in maintaining intestinal immune homeostasis [101]. Another important cytokine is IL-10 which germ-line variants have been linked to UC and CD [102, 103]. The

composition of intestinal bacteria and its critical role in the pathogenesis of IBD has come to the fore in the last years of research. CD-specific genes like NOD2, ATG16L1, and IRGM have focused attention on innate immunity in general and more specifically on microbial recognition and autophagy [60, 104, 105]. Important links have been elicited between NOD2 and autophagy, a natural regulated mechanism of the cell that disassembles dysfunctional components. NOD2 initiates autophagy by recruiting ATG16L1 to the cell membrane at the site of bacterial entry. Dendritic cells from CD patients, with NOD2 or ATG16L1 mutation, are defective in autophagy, bacterial handling, and antigen presentation [106, 107]. Environmental factors are also key components in the development of CD and can influence the disease outcome [108]. Higher social and hygiene status (with most likely less acute intestinal infections and subsequent acute inflammatory responses) is associated with a higher risk to suffer from IBD [109]. Environmental factors such as titanium dioxide, aluminum, or dietary emulsifiers may directly or indirectly activate pro-inflammatory pathways and thereby disturb the natural balance of the immune system which is an important part in the resolution of CD [110–112].

Conclusions

Resolution of inflammation is a strictly regulated process depending on cells, lipid, and protein mediators that maintain tissue homeostasis and prevent the development of chronic inflammatory diseases. In addition to universal mechanisms, specific mechanisms and cell interactions have been identified that control the resolution of inflammation in CD. Molecular mechanisms like resistance to apoptosis during inefficient anti-TNF therapy, altered cytokine secretion, or microbial imbalance in the gut of CD patients trigger the perpetuation of inflammation. Several pathways have to be considered as therapeutically relevant in this setting as their induction may provide new insights to control inflammation in CD without affecting the overall inflammatory response. First examples like antibodies targeting IL-23 in anti-TNF resistant patients give hope for therapeutic approaches that might be based on insights to the molecular phenotype of CD in specific situations. These approaches are therapeutically needed to enable sought-after resolution of inflammation based on a sound immunopathogenic basis.

Funding information CRC1181 Project C02 (R.A., C.N.) and DFG-SFB/TRR241 Project No. C02 (R.A.) are funded by the German Research Council DFG. The German Research Council DFG funds the Heisenberg Professorship of R.A.

Compliance with ethical standards

Conflict of interest The authors declare that there is no conflict of interest.

References

1. Strober W, Fuss I, Mannon P (2007) The fundamental basis of inflammatory bowel disease. *J Clin Invest* 117(3):514–521
2. Netea MG, Balkwill F, Chonchol M, Cominelli F, Donath MY, Giamarellos-Bourboulis EJ, Golenbock D, Gresnigt MS, Heneka MT, Hoffman HM, Hotchkiss R, Joosten LAB, Kastner DL, Korte M, Latz E, Libby P, Mandrup-Poulsen T, Mantovani A, Mills KHG, Nowak KL, O'Neill LA, Pickkers P, van der Poll T, Ridker PM, Schalkwijk J, Schwartz DA, Siegmund B, Steer CJ, Tilg H, van der Meer JWM, van de Veerdonk FL, Dinarello CA (2017) A guiding map for inflammation. *Nat Immunol* 18(8):826–831
3. Sanchez-Munoz F, Dominguez-Lopez A, Yamamoto-Furusho JK (2008) Role of cytokines in inflammatory bowel disease. *World J Gastroenterol* 14(27):4280–4288
4. Alessandri AL, Sousa LP, Lucas CD, Rossi AG, Pinho V, Teixeira MM (2013) Resolution of inflammation: mechanisms and opportunity for drug development. *Pharmacol Ther* 139(2):189–212
5. Chiang N, Fierro IM, Gronert K, Serhan CN (2000) Activation of lipoxin A(4) receptors by aspirin-triggered lipoxins and select peptides evokes ligand-specific responses in inflammation. *J Exp Med* 191(7):1197–1208
6. Serhan CN, Chiang N, Dalli J (2015) The resolution code of acute inflammation: novel pro-resolving lipid mediators in resolution. *Semin Immunol* 27(3):200–215
7. Hugot JP, Chamaillard M, Zouali H, Lesage S, Cézard JP, Belaiche J, Almer S, Tysk C, O'Morain CA, Gassull M, Binder V, Finkel Y, Cortot A, Modigliani R, Laurent-Puig P, Gower-Rousseau C, Macry J, Colombel JF, Sahbatou M, Thomas G (2001) Association of NOD2 leucine-rich repeat variants with susceptibility to Crohn's disease. *Nature* 411(6837):599–603
8. Neurath MF (2014) Cytokines in inflammatory bowel disease. *Nat Rev Immunol* 14(5):329–342
9. Atreya R, Neurath MF (2015) IBD pathogenesis in 2014: molecular pathways controlling barrier function in IBD. *Nat Rev Gastroenterol Hepatol* 12(2):67–68
10. Torres J, Mehandru S, Colombel JF, Peyrin-Biroulet L (2017) Crohn's disease. *Lancet* 389(10080):1741–1755
11. Harbord M, Annesse V, Vavricka SR, Allez M, Barreiro-de Acosta M, Boberg KM, Burisch J, de Vos M, de Vries AM, Dick AD, Juillerat P, Karlens TH, Koutroubakis I, Lakatos PL, Orchard T, Papay P, Raine T, Reinshagen M, Thaci D, Tilg H, Carbonnel F, for the European Crohn's and Colitis Organisation [ECCO] (2016) The first European evidence-based consensus on extra-intestinal manifestations in inflammatory bowel disease. *J Crohns Colitis* 10(3):239–254
12. Ng SC et al (2018) Worldwide incidence and prevalence of inflammatory bowel disease in the 21st century: a systematic review of population-based studies. *Lancet* 390(10114):2769–2778
13. Gomollon F et al (2017) 3rd European evidence-based consensus on the diagnosis and management of Crohn's disease 2016: part 1: diagnosis and medical management. *J Crohns Colitis* 11(1):3–25
14. Klenske E et al (2019) Targeting mucosal healing in Crohn's disease: what the clinician needs to know. *Ther Adv Gastroenterol* 12:1756284819856865
15. Sandborn WJ, Feagan BG, Rutgeerts P, Hanauer S, Colombel JF, Sands BE, Lukas M, Fedorak RN, Lee S, Bressler B, Fox I, Rosario M, Sankoh S, Xu J, Stephens K, Milch C, Parikh A, GEMINI 2 Study Group (2013) Vedolizumab as induction and maintenance therapy for Crohn's disease. *N Engl J Med* 369(8):711–721
16. Feagan BG, Sandborn WJ, Gasink C, Jacobstein D, Lang Y, Friedman JR, Blank MA, Johanns J, Gao LL, Miao Y, Adedokun OJ, Sands BE, Hanauer SB, Vermeire S, Targan S, Ghosh S, de Villiers WJ, Colombel JF, Tulassay Z, Seidler U, Salzberg BA, Desreumaux P, Lee SD, Loftus EV Jr, Dieleman LA, Katz S, Rutgeerts P (2016) Ustekinumab as induction and maintenance therapy for Crohn's disease. *N Engl J Med* 375(20):1946–1960
17. Atreya R, Neurath MF (2018) Mechanisms of molecular resistance and predictors of response to biological therapy in inflammatory bowel disease. *Lancet Gastroenterol Hepatol* 3(11):790–802
18. Kamada N et al (2008) Unique CD14 intestinal macrophages contribute to the pathogenesis of Crohn disease via IL-23/IFN-gamma axis. *J Clin Invest* 118(6):2269–2280
19. Tan ZY, Bealgey KW, Fang Y, Gong YM, Bao S (2009) Interleukin-23: immunological roles and clinical implications. *Int J Biochem Cell Biol* 41(4):733–735
20. Neurath MF, Finotto S, Fuss I, Boirivant M, Galle PR, Strober W (2001) Regulation of T cell apoptosis in inflammatory bowel disease: to die or not to die, that is the mucosal question. *Trends Immunol* 22(1):21–26
21. Luger A et al (2001) Infliximab induces apoptosis in monocytes from patients with chronic active Crohn's disease by using a caspase-dependent pathway. *Gastroenterology* 121(5):1145–1157
22. Broker LE, Kruyt FA, Giaccone G (2005) Cell death independent of caspases: a review. *Clin Cancer Res* 11(9):3155–3162
23. Nunes T, Bernardazzi C, de Souza HS (2014) Cell death and inflammatory bowel diseases: apoptosis, necrosis, and autophagy in the intestinal epithelium. *Biomed Res Int* 2014:218493
24. Rath PC, Aggarwal BB (1999) TNF-induced signaling in apoptosis. *J Clin Immunol* 19(6):350–364
25. Adegbola SO, Sahnun K, Warusavitame J, Hart A, Tozer P (2018) Anti-TNF Therapy in Crohn's Disease. *Int J Mol Sci* 19(8):2244.
26. Danese S (2012) New therapies for inflammatory bowel disease: from the bench to the bedside. *Gut* 61(6):918–932
27. Hanauer SB, Sandborn WJ, Rutgeerts P, Fedorak RN, Lukas M, MacIntosh D, Panaccione R, Wolf D, Pollack P (2006) Human anti-tumor necrosis factor monoclonal antibody (adalimumab) in Crohn's disease: the CLASSIC-I trial. *Gastroenterology* 130(2):323–333 **quiz 591**
28. Ben-Horin S, Kopylov U, Chowers Y (2014) Optimizing anti-TNF treatments in inflammatory bowel disease. *Autoimmun Rev* 13(1):24–30
29. Hanauer SB, Feagan BG, Lichtenstein GR, Mayer LF, Schreiber S, Colombel JF, Rachmilewitz D, Wolf DC, Olson A, Bao W, Rutgeerts P (2002) Maintenance infliximab for Crohn's disease: the ACCENT I randomised trial. *Lancet* 359(9317):1541–1549
30. Ben-Horin S, Chowers Y (2014) Tailoring anti-TNF therapy in IBD: drug levels and disease activity. *Nat Rev Gastroenterol Hepatol* 11(4):243–255
31. Sandborn WJ, Rutgeerts P, Enns R, Hanauer SB, Colombel JF, Panaccione R, D'Haens G, Li J, Rosenfeld MR, Kent JD, Pollack PF (2007) Adalimumab induction therapy for Crohn disease previously treated with infliximab: a randomized trial. *Ann Intern Med* 146(12):829–838
32. Steenholdt C, Brynskov J, Thomsen OØ, Munck LK, Fallingborg J, Christensen LA, Pedersen G, Kjeldsen J, Jacobsen BA, Oxholm AS, Kjellberg J, Bendtsen K, Ainsworth MA (2014) Individualised therapy is more cost-effective than dose intensification in patients with Crohn's disease who lose response to anti-TNF treatment: a randomised, controlled trial. *Gut* 63(6):919–927

33. Atreya R, Zimmer M, Bartsch B, Waldner MJ, Atreya I, Neumann H, Hildner K, Hoffman A, Kiesslich R, Rink AD, Rau TT, Rose-John S, Kessler H, Schmidt J, Neurath MF (2011) Antibodies against tumor necrosis factor (TNF) induce T cell apoptosis in patients with inflammatory bowel diseases via TNF receptor 2 and intestinal CD14(+) macrophages. *Gastroenterology* 141(6):2026–2038
34. Atreya R, Neumann H, Neufert C, Waldner MJ, Billmeier U, Zopf Y, Willma M, App C, Münster T, Kessler H, Maas S, Gebhardt B, Heimke-Brinck R, Reuter E, Dörje F, Rau TT, Uter W, Wang TD, Kiesslich R, Vieth M, Hannappel E, Neurath MF (2014) In vivo imaging using fluorescent antibodies to tumor necrosis factor predicts therapeutic response in Crohn's disease. *Nat Med* 20(3):313–318
35. Atreya R, Goetz M (2013) Molecular imaging in gastroenterology. *Nat Rev Gastroenterol Hepatol* 10(12):704–712
36. Leal RF, Planell N, Kajekar R, Lozano JJ, Ordás I, Dotti I, Esteller M, Masamunt MC, Parmar H, Ricart E, Panés J, Salas A (2015) Identification of inflammatory mediators in patients with Crohn's disease unresponsive to anti-TNF α therapy. *Gut* 64(2):233–242
37. West NR et al (2017) Oncostatin M drives intestinal inflammation and predicts response to tumor necrosis factor-neutralizing therapy in patients with inflammatory bowel disease. *Nat Med* 23(5):579–589
38. Gaujoux R, Starosvetsky E, Maimon N, Vallania F, Bar-Yoseph H, Pressman S, Weisshof R, Goren I, Rabinowitz K, Waterman M, Yanai H, Dotan I, Sabo E, Chowder Y, Khatri P, Shen-Orr SS (2019) Cell-centred meta-analysis reveals baseline predictors of anti-TNF α non-response in biopsy and blood of patients with IBD. *Gut* 68(4):604–614
39. Curotto de Lafaille MA, Lafaille JJ (2009) Natural and adaptive foxp3+ regulatory T cells: more of the same or a division of labor? *Immunity* 30(5):626–635
40. Gregori S, Goudy KS, Roncarolo MG (2012) The cellular and molecular mechanisms of immuno-suppression by human type 1 regulatory T cells. *Front Immunol* 3:30
41. Mottet C, Uhlig HH, Powrie F (2003) Cutting edge: cure of colitis by CD4+CD25+ regulatory T cells. *J Immunol* 170(8):3939–3943
42. Gagliani N, Vesely MCA, Iseppon A, Brockmann L, Xu H, Palm NW, de Zoete MR, Licona-Limón P, Paiva RS, Ching T, Weaver C, Zi X, Pan X, Fan R, Garmire LX, Cotton MJ, Drier Y, Bernstein B, Geginat J, Stockinger B, Esplugues E, Huber S, Flavell RA (2015) Th17 cells transdifferentiate into regulatory T cells during resolution of inflammation. *Nature* 523(7559):221–225
43. Schett G, Neurath MF (2018) Resolution of chronic inflammatory disease: universal and tissue-specific concepts. *Nat Commun* 9(1):3261
44. Desreumaux P, Foussat A, Allez M, Beaugerie L, Hébuterne X, Bouhnik Y, Nachury M, Brun V, Bastian H, Belmonte N, Ticchioni M, Duchange A, Morel-Mandrin P, Neveu V, Clerget-Chossat N, Forte M, Colombel J-F (2012) Safety and efficacy of antigen-specific regulatory T cell therapy for patients with refractory Crohn's disease. *Gastroenterology* 143(5):1207–1217 e2
45. Glocker EO, Kotlarz D, Boztug K, Gertz EM, Schäffer AA, Noyan F, Perro M, Diestelhorst J, Allroth A, Murugan D, Hätscher N, Pfeifer D, Sykora KW, Sauer M, Kreipe H, Lacher M, Nustede R, Woellner C, Baumann U, Salzer U, Koletzko S, Shah N, Segal AW, Sauerbrey A, Buderus S, Snapper SB, Grimbacher B, Klein C (2009) Inflammatory bowel disease and mutations affecting the interleukin-10 receptor. *N Engl J Med* 361(21):2033–2045
46. Herfarth H, Scholmerich J (2002) IL-10 therapy in Crohn's disease: at the crossroads. Treatment of Crohn's disease with the anti-inflammatory cytokine interleukin 10. *Gut* 50(2):146–147
47. Schreiber S, Fedorak RN, Nielsen OH, Wild G, Williams CN, Nikolaus S, Jacyna M, Lashner BA, Gangl A, Rutgeerts P, Isaacs K, van Deventer SJH, Koningsberger JC, Cohard M, LeBeaut A, Hanauer SB (2000) Safety and efficacy of recombinant human interleukin 10 in chronic active Crohn's disease. Crohn's disease IL-10 cooperative study group. *Gastroenterology* 119(6):1461–1472
48. Mizoguchi A (2012) Healing of intestinal inflammation by IL-22. *Inflamm Bowel Dis* 18(9):1777–1784
49. Sabat R, Ouyang W, Wolk K (2014) Therapeutic opportunities of the IL-22-IL-22R1 system. *Nat Rev Drug Discov* 13(1):21–38
50. Chiriac MT, Buchen B, Wandersee A, Hundorfean G, Günther C, Bourjau Y, Doyle SE, Frey B, Ekici AB, Büttner C, Weigmann B, Atreya R, Wirtz S, Becker C, Siebler J, Neurath MF (2017) Activation of epithelial signal transducer and activator of transcription 1 by interleukin 28 controls mucosal healing in mice with colitis and is increased in mucosa of patients with inflammatory bowel disease. *Gastroenterology* 153(1):123–138 e8
51. Medina-Contreras O, Harusato A, Nishio H, Flannigan KL, Ngo V, Leoni G, Neumann PA, Geem D, Lili LN, Ramadas RA, Chassaing B, Gewirtz AT, Kohlmeier JE, Parkos CA, Towne JE, Nusrat A, Denning TL (2016) Cutting edge: IL-36 receptor promotes resolution of intestinal damage. *J Immunol* 196(1):34–38
52. Bain CC, Mowat AM (2014) Macrophages in intestinal homeostasis and inflammation. *Immunol Rev* 260(1):102–117
53. Steinbach EC, Plevy SE (2014) The role of macrophages and dendritic cells in the initiation of inflammation in IBD. *Inflamm Bowel Dis* 20(1):166–175
54. Parihar A, Eubank TD, Doseff AI (2010) Monocytes and macrophages regulate immunity through dynamic networks of survival and cell death. *J Innate Immun* 2(3):204–215
55. Lissner D et al (2015) Monocyte and M1 macrophage-induced barrier defect contributes to chronic intestinal inflammation in IBD. *Inflamm Bowel Dis* 21(6):1297–1305
56. Zhu W, Yu J, Nie Y, Shi XK, Liu Y, Li F, Zhang XL (2014) Disequilibrium of M1 and M2 macrophages correlates with the development of experimental inflammatory bowel diseases. *Immunol Investig* 43(7):638–652
57. Elliott MR, Ravichandran KS (2016) The dynamics of apoptotic cell clearance. *Dev Cell* 38(2):147–160
58. Elliott MR, Koster KM, Murphy PS (2017) Efferocytosis signaling in the regulation of macrophage inflammatory responses. *J Immunol* 198(4):1387–1394
59. Onali S, A Favale, and MC Fantini (2019) The Resolution of Intestinal Inflammation: The Peace-Keeper's Perspective. *Cells* 8(4):34
60. Marks DJ, Harbord MWN, MacAllister R, Rahman FZ, Young J, al-Lazikani B, Lees W, Novelli M, Bloom S, Segal AW (2006) Defective acute inflammation in Crohn's disease: a clinical investigation. *Lancet* 367(9511):668–678
61. Iwakura Y, Ishigame H (2006) The IL-23/IL-17 axis in inflammation. *J Clin Invest* 116(5):1218–1222
62. Geremia A, Arancibia-Carcamo CV, Fleming MPP, Rust N, Singh B, Mortensen NJ, Travis SPL, Powrie F (2011) IL-23-responsive innate lymphoid cells are increased in inflammatory bowel disease. *J Exp Med* 208(6):1127–1133
63. McGovern D, Powrie F (2007) The IL-23 axis plays a key role in the pathogenesis of IBD. *Gut* 56(10):1333–1336
64. Duerr RH, Taylor KD, Brant SR, Rioux JD, Silverberg MS, Daly MJ, Steinhart AH, Abraham C, Regueiro M, Griffiths A, Dassopoulos T, Bitton A, Yang H, Targan S, Datta LW, Kistner EO, Schumm LP, Lee AT, Gregersen PK, Barnada MM, Rotter JL, Nicolae DL, Cho JH (2006) A genome-wide association study identifies IL-23R as an inflammatory bowel disease gene. *Science* 314(5804):1461–1463

65. Ahern PP, Schiering C, Buonocore S, McGeachy MJ, Cua DJ, Maloy KJ, Powrie F (2010) Interleukin-23 drives intestinal inflammation through direct activity on T cells. *Immunity* 33(2): 279–288
66. Kobayashi T, Okamoto S, Hisamatsu T, Kamada N, Chinen H, Saito R, Kitazume MT, Nakazawa A, Sugita A, Koganei K, Isobe K, Hibi T (2008) IL-23 differentially regulates the Th1/Th17 balance in ulcerative colitis and Crohn's disease. *Gut* 57(12):1682–1689
67. Feagan BG, Sandborn WJ, D'Haens G, Panés J, Kaser A, Ferrante M, Louis E, Franchimont D, Dewit O, Seidler U, Kim KJ, Neurath MF, Schreiber S, Scholl P, Pamulapati C, Lalovic B, Visvanathan S, Padula SJ, Herichova I, Soaita A, Hall DB, Böcher WO (2017) Induction therapy with the selective interleukin-23 inhibitor risankizumab in patients with moderate-to-severe Crohn's disease: a randomised, double-blind, placebo-controlled phase 2 study. *Lancet* 389(10080):1699–1709
68. Sands BE, Chen J, Feagan BG, Penney M, Rees WA, Danese S, Higgins PDR, Newbold P, Faggioni R, Patra K, Li J, Klekotka P, Morehouse C, Pulkstenis E, Drappa J, van der Merwe R, Gasser RA Jr (2017) Efficacy and safety of MEDI2070, an antibody against interleukin 23, in patients with moderate to severe Crohn's disease: a phase 2a study. *Gastroenterology* 153(1):77–86 **e6**
69. Schmitt H, Billmeier U, Dieterich W, Rath T, Sonnenwald S, Reid S, Hirschmann S, Hildner K, Waldner MJ, Mudter J, Hartmann A, Grützmann R, Neufert C, Münster T, Neurath MF, Atreya R (2019) Expansion of IL-23 receptor bearing TNFR2+ T cells is associated with molecular resistance to anti-TNF therapy in Crohn's disease. *Gut* 68(5):814–828
70. Medzhitov R (2008) Origin and physiological roles of inflammation. *Nature* 454(7203):428–435
71. Serhan CN (2017) Treating inflammation and infection in the 21st century: new hints from decoding resolution mediators and mechanisms. *FASEB J* 31(4):1273–1288
72. Fullerton JN, Gilroy DW (2016) Resolution of inflammation: a new therapeutic frontier. *Nat Rev Drug Discov* 15(8):551–567
73. Headland SE, Norling LV (2015) The resolution of inflammation: principles and challenges. *Semin Immunol* 27(3):149–160
74. Gilroy D, De Maeyer R (2015) New insights into the resolution of inflammation. *Semin Immunol* 27(3):161–168
75. Buckley CD, Gilroy DW, Serhan CN (2014) Proresolving lipid mediators and mechanisms in the resolution of acute inflammation. *Immunity* 40(3):315–327
76. Uderhardt S, Herrmann M, Oskolkova OV, Aschermann S, Bicker W, Ipseiz N, Sarter K, Frey B, Rothe T, Voll R, Nimmerjahn F, Bochkov VN, Schett G, Krönke G (2012) 12/15-lipoxygenase orchestrates the clearance of apoptotic cells and maintains immunologic tolerance. *Immunity* 36(5):834–846
77. Uderhardt S, Ackermann JA, Fillep T, Hammond VJ, Willeit J, Santer P, Mayr M, Biburger M, Miller M, Zellner KR, Stark K, Zarbock A, Rossaint J, Schubert I, Mielenz D, Dietel B, Raaz-Schrauder D, Ay C, Gremmel T, Thaler J, Heim C, Herrmann M, Collins PW, Schabbauer G, Mackman N, Voehringer D, Nadler JL, Lee JJ, Massberg S, Rauh M, Kiechl S, Schett G, O'Donnell VB, Krönke G (2017) Enzymatic lipid oxidation by eosinophils propagates coagulation, hemostasis, and thrombotic disease. *J Exp Med* 214(7):2121–2138
78. Masoodi M, Pearl DS, Eiden M, Shute JK, Brown JF, Calder PC, Trebble TM (2013) Altered colonic mucosal polyunsaturated fatty acid (PUFA) derived lipid mediators in ulcerative colitis: new insight into relationship with disease activity and pathophysiology. *PLoS One* 8(10):e76532
79. Pearl DS, Masoodi M, Eiden M, Brümmer J, Gullick D, Mckeever TM, Whittaker MA, Nitch-Smith H, Brown JF, Shute JK, Mills G, Calder PC, Trebble TM (2014) Altered colonic mucosal availability of n-3 and n-6 polyunsaturated fatty acids in ulcerative colitis and the relationship to disease activity. *J Crohns Colitis* 8(1):70–79
80. Rogler G (2017) Resolution of inflammation in inflammatory bowel disease. *Lancet Gastroenterol Hepatol* 2(7):521–530
81. Vindigni SM, Zisman TL, Suskind DL, Damman CJ (2016) The intestinal microbiome, barrier function, and immune system in inflammatory bowel disease: a tripartite pathophysiological circuit with implications for new therapeutic directions. *Ther Adv Gastroenterol* 9(4):606–625
82. Carroll IM, Threadgill DW, Threadgill DS (2009) The gastrointestinal microbiome: a malleable, third genome of mammals. *Mamm Genome* 20(7):395–403
83. Spor A, Koren O, Ley R (2011) Unravelling the effects of the environment and host genotype on the gut microbiome. *Nat Rev Microbiol* 9(4):279–290
84. Lane ER, Zisman TL, Suskind DL (2017) The microbiota in inflammatory bowel disease: current and therapeutic insights. *J Inflamm Res* 10:63–73
85. Zuo T, Ng SC (2018) The gut microbiota in the pathogenesis and therapeutics of inflammatory bowel disease. *Front Microbiol* 9: 2247
86. Sun Y, Li L, Xia Y, Li W, Wang K, Wang L, Miao Y, Ma S (2019) The gut microbiota heterogeneity and assembly changes associated with the IBD. *Sci Rep* 9(1):440
87. Manichanh C et al (2006) Reduced diversity of faecal microbiota in Crohn's disease revealed by a metagenomic approach. *Gut* 55(2):205–211
88. Sepeshri S, Kotlowski R, Bernstein CN, Krause DO (2007) Microbial diversity of inflamed and noninflamed gut biopsy tissues in inflammatory bowel disease. *Inflamm Bowel Dis* 13(6): 675–683
89. Gevers D, Kugathasan S, Denson LA, Vázquez-Baeza Y, van Treuren W, Ren B, Schwager E, Knights D, Song SJ, Yassour M, Morgan XC, Kostic AD, Luo C, González A, McDonald D, Haberman Y, Walters T, Baker S, Rosh J, Stephens M, Heyman M, Markowitz J, Baldassano R, Griffiths A, Sylvester F, Mack D, Kim S, Crandall W, Hyams J, Huttenhower C, Knight R, Xavier RJ (2014) The treatment-naïve microbiome in new-onset Crohn's disease. *Cell Host Microbe* 15(3):382–392
90. Sartor RB, Wu GD (2017) Roles for intestinal bacteria, viruses, and fungi in pathogenesis of inflammatory bowel diseases and therapeutic approaches. *Gastroenterology* 152(2):327–339 **e4**
91. Ohkusa T, Kato K, Terao S, Chiba T, Mabe K, Murakami K, Mizokami Y, Sugiyama T, Yanaka A, Takeuchi Y, Yamato S, Yokoyama T, Okayasu I, Watanabe S, Tajiri H, Sato N, Japan UC Antibiotic Therapy Study Group (2010) Newly developed antibiotic combination therapy for ulcerative colitis: a double-blind placebo-controlled multicenter trial. *Am J Gastroenterol* 105(8):1820–1829
92. Turner D, Levine A, Kolho KL, Shaoul R, Ledder O (2014) Combination of oral antibiotics may be effective in severe pediatric ulcerative colitis: a preliminary report. *J Crohns Colitis* 8(11): 1464–1470
93. Haberman Y, Tickle TL, Dexheimer PJ, Kim MO, Tang D, Kams R, Baldassano RN, Noe JD, Rosh J, Markowitz J, Heyman MB, Griffiths AM, Crandall WV, Mack DR, Baker SS, Huttenhower C, Keljo DJ, Hyams JS, Kugathasan S, Walters TD, Aronow B, Xavier RJ, Gevers D, Denson LA (2014) Pediatric Crohn disease patients exhibit specific ileal transcriptome and microbiome signature. *J Clin Invest* 124(8):3617–3633
94. Sokol H, Leducq V, Aschard H, Pham HP, Jegou S, Landman C, Cohen D, Liguori G, Bourrier A, Nion-Larmurier I, Cosnes J, Seksik P, Langella P, Skurnik D, Richard ML, Beaugerie L (2017) Fungal microbiota dysbiosis in IBD. *Gut* 66(6):1039–1048

95. Shaw KA, Bertha M, Hofmekler T, Chopra P, Vatanen T, Srivatsa A, Prince J, Kumar A, Sauer C, Zwick ME, Satten GA, Kostic AD, Mulle JG, Xavier RJ, Kugathasan S (2016) Dysbiosis, inflammation, and response to treatment: a longitudinal study of pediatric subjects with newly diagnosed inflammatory bowel disease. *Genome Med* 8(1):75
 96. Rajca S et al (2014) Alterations in the intestinal microbiome (dysbiosis) as a predictor of relapse after infliximab withdrawal in Crohn's disease. *Inflamm Bowel Dis* 20(6):978–986
 97. Antoni L et al (2014) Intestinal barrier in inflammatory bowel disease. *World J Gastroenterol* 20(5):1165–1179
 98. Jager S, Stange EF, Wehkamp J (2013) Inflammatory bowel disease: an impaired barrier disease. *Langenbeck's Arch Surg* 398(1):1–12
 99. de Souza HS, Fiocchi C (2016) Immunopathogenesis of IBD: current state of the art. *Nat Rev Gastroenterol Hepatol* 13(1):13–27
 100. Billiet T, Vande Casteele N, van Stappen T, Princen F, Singh S, Gils A, Ferrante M, van Assche G, Cleynen I, Vermeire S (2015) Immunogenicity to infliximab is associated with HLA-DRB1. *Gut* 64(8):1344–1345
 101. Brand S (2009) Crohn's disease: Th1, Th17 or both? The change of a paradigm: new immunological and genetic insights implicate Th17 cells in the pathogenesis of Crohn's disease. *Gut* 58(8):1152–1167
 102. Franke A et al (2008) Sequence variants in IL-10, ARPC2 and multiple other loci contribute to ulcerative colitis susceptibility. *Nat Genet* 40(11):1319–1323
 103. Franke A et al (2010) Genome-wide association study for ulcerative colitis identifies risk loci at 7q22 and 22q13 (IL17REL). *Nat Genet* 42(4):292–294
 104. Ogura Y, Bonen DK, Inohara N, Nicolae DL, Chen FF, Ramos R, Britton H, Moran T, Karaliuskas R, Duerr RH, Achkar JP, Brant SR, Bayless TM, Kirschner BS, Hanauer SB, Nuñez G, Cho JH (2001) A frameshift mutation in NOD2 associated with susceptibility to Crohn's disease. *Nature* 411(6837):603–606
 105. Hampe J, Franke A, Rosenstiel P, Till A, Teuber M, Huse K, Albrecht M, Mayr G, de la Vega FM, Briggs J, Günther S, Prescott NJ, Onnie CM, Häsler R, Sipos B, Fölsch UR, Lengauer T, Platzer M, Mathew CG, Krawczak M, Schreiber S (2007) A genome-wide association scan of nonsynonymous SNPs identifies a susceptibility variant for Crohn disease in ATG16L1. *Nat Genet* 39(2):207–211
 106. Cooney R, Baker J, Brain O, Danis B, Pichulik T, Allan P, Ferguson DJP, Campbell BJ, Jewell D, Simmons A (2010) NOD2 stimulation induces autophagy in dendritic cells influencing bacterial handling and antigen presentation. *Nat Med* 16(1):90–97
 107. Travassos LH, Carneiro LAM, Ramjeet M, Hussey S, Kim YG, Magalhães JG, Yuan L, Soares F, Chea E, le Bourhis L, Boneca IG, Allaoui A, Jones NL, Nuñez G, Girardin SE, Philpott DJ (2010) Nod1 and Nod2 direct autophagy by recruiting ATG16L1 to the plasma membrane at the site of bacterial entry. *Nat Immunol* 11(1):55–62
 108. Lees CW, Barrett JC, Parkes M, Satsangi J (2011) New IBD genetics: common pathways with other diseases. *Gut* 60(12):1739–1753
 109. Hafner S, Timmer A, Herfarth H, Rogler G, Schölmerich J, Schäffler A, Ehrenstein B, Jilg W, Ott C, Strauch UG, Obermeier F (2008) The role of domestic hygiene in inflammatory bowel diseases: hepatitis A and worm infestations. *Eur J Gastroenterol Hepatol* 20(6):561–566
 110. Ruiz PA, Morón B, Becker HM, Lang S, Atrott K, Spalinger MR, Scharl M, Wojtal KA, Fischbeck-Terhalle A, Frey-Wagner I, Hausmann M, Kraemer T, Rogler G (2017) Titanium dioxide nanoparticles exacerbate DSS-induced colitis: role of the NLRP3 inflammasome. *Gut* 66(7):1216–1224
 111. Pineton de Chambrun G et al (2014) Aluminum enhances inflammation and decreases mucosal healing in experimental colitis in mice. *Mucosal Immunol* 7(3):589–601
 112. Chassaing B, Koren O, Goodrich JK, Poole AC, Srinivasan S, Ley RE, Gewirtz AT (2015) Dietary emulsifiers impact the mouse gut microbiota promoting colitis and metabolic syndrome. *Nature* 519(7541):92–96
- Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.