



Review

Prospective oncotarget for gynecological cancer: Opioid growth factor (OGF) - opioid growth factor receptor (OGFr) axis

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ABSTRACT

The standard treatments for neoplasia include surgery, chemotherapy, hormone antagonists and radiotherapy, which can prolong survival, but rarely cure the tumors of gynecological cancer patients. OGF - OGFr expression, in various gynecologic cells and tissues, is an intersection point between cell development, neuroendocrine function and immune modulation. It has been identified that OGF and OGFr expression differs between gynecological tumor and normal cells. Further, exogenous or endogenous OGF and OGFr antagonists have been known to have a role in regulating cell viability and apoptosis. Moreover, the expression of proteins in the OGF - OGFr axis modulate differentiation and membrane expression of immune cells, which can enhance the immune response. In vivo and in vitro assays have shown that OGF and OGFr antagonists inhibit mitosis as well as induce apoptosis in gynecologic cancer cells. Although immune augmentation combination therapies can intensify cytotoxic activity, OGF or OGFr antagonists do not increase toxicities associated with dual-immune regulation. In conclusion, the OGF - OGFr axis provides significant strategies for antitumor efficiency in gynecological cancer.

1. Introduction

Globally, gynecological cancers have increased in incidence and mortality during the past decade [1]. Cervical cancer, uterine cancer and ovarian cancer form the most frequent gynecologic carcinomas and severely threaten the health of women. On an annual basis, these tumors include about 569,847 newly diagnosed and 311,365 deaths from cervical cancer, 382,069 newly diagnosed and 89,929 deaths from uterine cancer, and 295,414 newly diagnosed and 184,799 deaths from ovarian cancer, respectively [2]. Additional gynecological cancers include the less commonly occurring vaginal and vulvar cancers, and gestational trophoblastic neoplasia-like choriocarcinomas [3]. Despite traditional cytoreductive surgery, platinum-based chemotherapy and three-dimensional conformal or intensity-modulated radiotherapy provide the most common treatments for gynecologic tumors to delay tumor progression and improve quality of life. Without an efficient screening strategy or increased therapeutic activity, most of these patients are diagnosed at a terminal stage or are resistant to traditional

therapeutics, which results in a poor prognosis [4–6]. The use of oncotargeted molecular therapies are expanding, including PD (programmed cell death protein)-1/PD-L1 monoclonal antibodies [7], Poly (ADP-Ribose) Polymerase inhibitor (PARPi) [8], as well as adoptive cellular immunotherapy, including chimeric antigen receptor (CAR) T-cells and dendritic cell (DC) vaccines [9], which have the potential to provide curative effect and prolong overall survival (OS). However, low levels of efficacy [10] and increased side effects [11] have restricted clinical applications supporting a need for additional research and development.

There is a linked relationship between tumor induction, disease progression, neuroendocrine interaction and immunity. It is generally acknowledged that neoplasms originate following genetic mutations, cellular damage and regeneration [12]; however, the tumor itself is rarely the direct cause of death. In the clinic, the majority of cancer patients die from the toxic side effects of chemotherapy, associated neutropenia, tumor related cachexia and corresponding complications [13,14]. The benefit patients derive from standard of care is limited and

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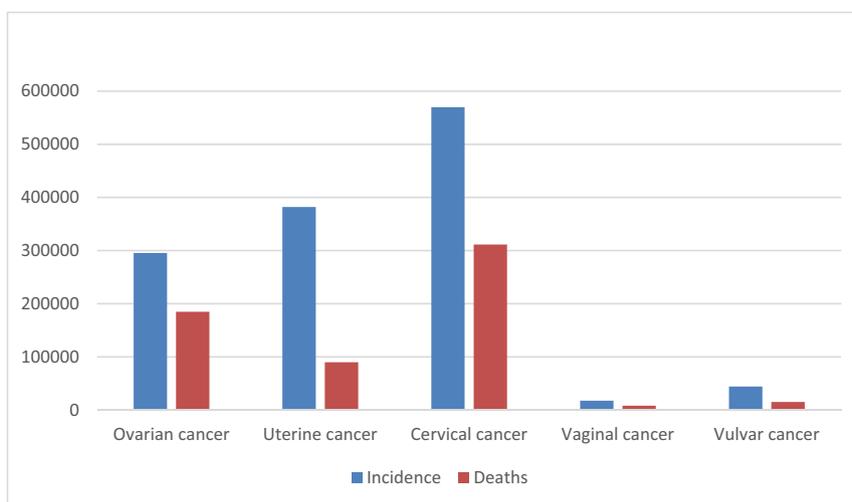


Fig. 1. General case of gynecological oncology.

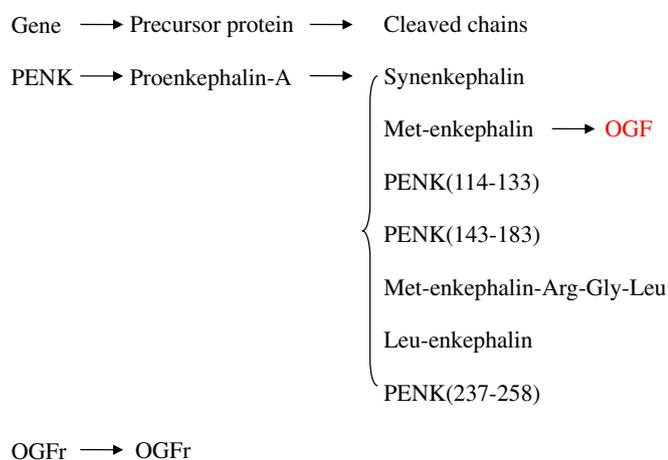


Fig. 2. Taxonomy of OGF and OGFr.

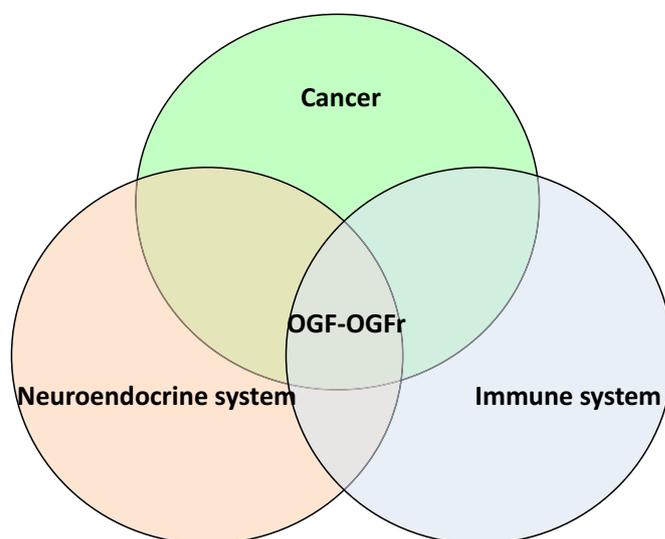


Fig. 4. The role of OGF - OGFr axis.

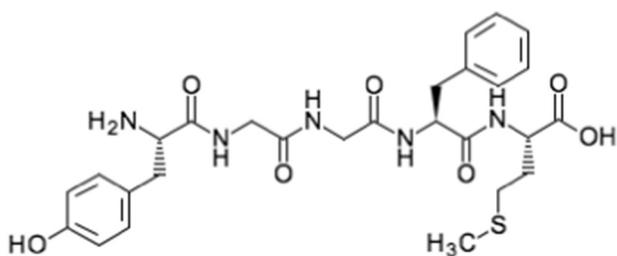


Fig. 3. Structure of OGF.

may reduce the quality of life. We have accepted the viewpoint that cancer is a chronic inflammatory disease, such that neuroendocrine and immunity regulates cancer occurrence and progression. In contrast, cancer may cause immunodeficiency and neuroendocrine and metabolic imbalances, leading to fatigue, anorexia and cachexia [15]. Consequently, understanding the interplay between the tumor, neuroendocrine interactions and immunity may provide clinicians an improved approach to control cancer and extend survival as opposed to traditional therapeutic strategies (Figs. 1–4).

2. OGF-OGFr axis

The endogenous opioid family is comprised of opioid peptides and opioid receptors that are ordinarily generated by the adrenal and

pituitary glands, and distributed through an autocrine or paracrine distribution pattern [16,17]. OGF, also termed methionine-enkephalin (MENK), is a native opiate pentapeptide derived from preproenkephalin and pro-opiomelanocortin genes, which is located at the 8q12.1 chromosome, and contains the structure H-Tyr-Gly-Phe-Met-OH [18,19]. As a natural ligand for OGFr, OGF provides a series of biological functions as a neuroprotective ingredient, including tissue development and organ maturation, DNA synthesis and cellular renewal, vascular generation and wound healing via binding to OGFr [20–22]. The OGFr is located at the 20q13.33 chromosome [23] and is characterized as a zeta-opioid receptor distributed to the cytoplasm and nucleus [24,25]. Three classic opiate receptors, mu-opioid receptor, delta-opioid receptor and kappa-opioid receptor contribute to the analgesic efficiency of opioids [26–28]. The OGF - OGFr axis is widely distributed on central and peripheral systems, especially the cerebrum and gastric-intestine (GI) tract [29]. OGFr is an integral membrane protein that is associated with the nucleus and following binding to OGF initiates a signaling cascade related to DNA synthesis and growth, and modulation of the immune and neuroendocrine systems following injury [23,30,31]. The action of OGF - OGFr is nonspecific, reversible, and dose-dependent, which is attributed to the duration of OGFr inhibition [32]. OGF or OGFr antagonists and low doses of naltrexone (LDN) have potential for anti-inflammation [28], antinociception [33], multiple sclerosis [34],

autoimmune encephalomyelitis [35], metabolism [36] and energy homeostasis [37], chronic hepatitis [38], anti-obesity and diabetic management [39].

3. OGF - OGF α r axis and gynecological cancer

The OGF - OGF α r axis has been identified in dozens of tumor cells, tissues and intraluminal secretions. Furthermore, natural or exogenous OGF depresses tumor proliferation by inducing cell cycle arrest and/or apoptosis while LDN inhibits neoplasm progression by blocking OGF α r or up-regulating OGF - OGF α r concentrations [40–44]. OGF and OGF α r possess the property of dual-immune modulation for innate and adaptive competent cells and cytokines which could remodel tumor micro-environment (TME) by enhancing anti-tumor immunoactivity and remitting the immuno-suppressive state [45,46].

3.1. *In vitro* assay

OGF - OGF α r are present in the cytoplasm and nucleocytoplasm of OVCAR-3, SKOV-3, SW626, CAOV-3, and HEY ovarian cancer cell lines and can block tumor growth in a dose-dependent way. A proliferation inhibition of 20% was observed with OVCAR-3 cells, following a 120 h co-culture and 23% on SKOV-3, 25% on SW626, 18% on CAOV-3, and 24% on HEY following 72 h of co-culture [47]. The antitumor effect was lost as OGF α r was silenced, and further analysis showed p16 and/or p21 levels were up-regulated 4.1-fold and/or 2.3-fold after exposure to OGF for 9 h. In ovarian cancer cell lines SH-6 and SH-16 transfected with OGF α r shRNA, cell numbers increased by 48–132% and 33–88% within 96 h without influencing the OGF level [48]. SH-6 and SH-16 cells with low OGF α r expression had hyperactive proliferation as measured by BrdU incorporation; however, normal SH-6 and SH-16 clonal cells showed little apoptosis or necrosis. Similarly, proliferation of human squamous carcinoma cells, SCC-1 were depressed by 41% after exposure to OGF, whereas the cell number was increased by 54% after transfection with OGF α r siRNA, and 42% after exposure to specific antibodies against OGF [43]. Thus, the OGF - OGF α r axis may suppress tumor proliferation by stabilization of the cell cycle in the G0/G1 phase. The process is mediated by cyclin-dependent kinase inhibitory (CKI) pathways in which p16 and/or p21 are predominant and reduces phospho-Rb, which has a phosphorylated reaction in G1 phase via down-regulation of cyclin-dependent kinase protein (Cdk) 2 [49,50]. Other mechanisms include RNA and protein synthesis inhibition by an OGF or OGF α r antagonist, in a non-specific, dose-dependent, reversible manner [51].

At present, our unpublished research data has revealed that human papillomavirus (HPV) \pm human cervical cancer Hela (HPV 16 +), Siha (HPV 18 +), C33A (HPV -) cell lines have a depressed proliferation, but also induced apoptosis by co-culture with an exogenous OGF or OGF α r antagonist (Fig. 6). Flow cytometry has verified the arrest of tumor cell mitosis and increased apoptosis by drug exposure for 48 h. Similarly, light microscopy, immunofluorescence and electron microscopy have revealed morphological pathologic, including irregular or rounder cells, lower cell density and adherence, intensive blebbing, hyperchromatic nuclei, excessive karyopyknosis and karyolysis in drug treatment group. Quantitative real time polymerase chain reaction (qRT-PCR) and Western blot analyses have confirmed cell-cycle related gene inhibition, as well as pro-apoptotic molecule expression, including caspase 3 and Bax activation. One mechanism of cellular apoptosis promotion by exogenous OGF may be the activity of the PI3K/AKT/mTOR pathway through OGF α r [52,53]. Intermittent or short exposure to an OGF α r antagonist would boost OGF presentation in the tissue which reacts analogously with OGF on tumor proliferation modulation [54]. In addition, OGF α r antagonist transduces extracellular signals to the cytoplasm and nucleus via regulating extracellular regulated kinase (ERK), Ca²⁺ channels and synergistically activating protein kinase C (PKC), increases integrin α 7 level, and therefore influences tumor cells

malignant biological behavior [55–57].

3.2. *In vivo* assay

Mice with a xenograft of the human ovarian cancer cell line SKOV-3 had an association with the expression of OGF and OGF α r in the cytoplasm and nucleus of tumor tissue. Further, the severity of disease progression was negatively correlated with OGF and OGF α r concentrations [58]. In a female mouse model, the number of intraperitoneal metastatic nodes of SKOV-3 ovarian carcinoma were reduced by 65% with a dosage of 10 mg/mL OGF given by intraperitoneal injection and 51% with a dosage of 0.1 mg/kg LDN. OGF α r expression was reduced by 29% in the OGF administration group and increased by 63% following LDN administration group. To assess the mechanism of tumor growth retardation, tumor cell BrdU labeling was found to be decreased 61% with exposure to OGF and 52% with exposure to LDN. Furthermore, the vascular density of tumor tissue with OGF and LDN treatment was reduced 89% and 73%, respectively. Established mouse models with under-expression of OGF α r have a delayed tumor induction in the low OGF α r group that was 21–22 days and 7–14 days shorter with 1–2 \times 10⁶ SH-6 and SH-16 cells [48]. Following injection with 10 mg/kg OGF, there was a 100% macroscopic SH-6 tumor incidence in the under-expressing OGF α r group by day 7, in the WT group by day 19 and in the EV group by day 21, respectively. Tumor volumes were enlarged by 41–267%, 4–14 days later comparative to controls. Exposure to OGF in WT and EV groups showed notable tumor protective effects including a decrease of 38–60% in volume and 32–55% in tumor weights relative to saline, which suggests that OGF α r stimulated tumor development and OGF slowed tumor growth by binding to OGF α r.

Tissue specimens from ovarian sites and omental metastases in 53 terminal ovarian cancer patients, and a comparable ovarian cyst were assessed by Fanner, et al., using semi-quantitative immunohistochemistry [59]. In these studies, the distribution of OGF and OGF α r in cytoplasm and nucleus was similar among ovarian cancer, ovarian cyst and HOSE cells. However, the expression of OGF was reduced by 29% in ovarian cyst and 58% in ovarian cancer cells, and the expression of OGF α r decreased 34% in ovarian cyst and 48% in ovarian cancer cells, respectively. Although OGF exhibited a specific binding to nuclear OGF α r, the binding capacity in ovarian cancer was 81% lower compared with ovarian cyst without OGF - OGF α r affinity discrepancy.

OGF and OGF α r have also been proven to occur on human squamous epithelium and endometrium [60]. In the tissue of human squamous cell carcinoma, OGF α r levels were down-regulated by 79% compared with that in normal tissue, and 42% compared with that in para-cancerous tissue [61]. The binding ability to nuclear OGF α r of OGF had a nine-fold reduction in comparison with that of normal tissue in which OGF α r was evenly expressed. Nonetheless, OGF α r mRNA displayed no marked variation among epithelial mucosa, tumor margin and tumor tissue, prompting OGF α r protein reduction to be triggered in the process of post-transcription. Studies of uterine membranes revealed that OGF accelerates cellular apoptosis via the Fas/FasL and/or NFAT1 signal pathway [62,63]. In addition, except for this antineoplastic mechanism of decreased cellular Ca²⁺ uptake, OGF and estradiol hormone, which are co-factors to promote cell growth and tissue development, and may also play a significant role in hormone-dependent uterine carcinoma [64,65].

3.3. Immuno-regulation

In view of immune modulation by opioids via opioid receptors [66,67], investigators have explored the role of the OGF - OGF α r axis on the immune system. Generally speaking, the function of the OGF - OGF α r axis is more like a double-edged sword. On the one side, OGF and OGF α r antagonists suppress immune reactivity, providing a functional remedy for autoimmune disease [32,68]. On the other side, these two molecules could enhance the antineoplastic response of immune cells and

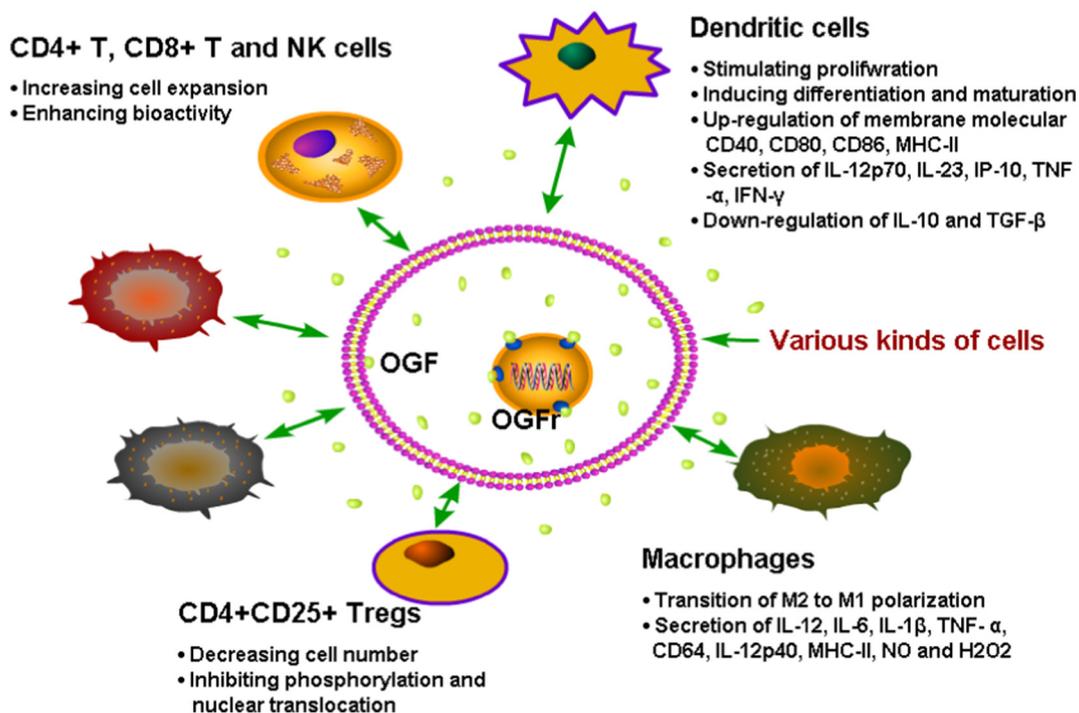


Fig. 5. Diagrammatical illustration of OGF - OGFr axis.

molecules [52,69]. The OGF - OGFr axis acts on toll-like receptors (TLRs) that are vital in initiating innate and adaptive immunoactivities to defeat inflammation and neoplasm [70–72]. TLRs belong to the family of pattern-recognition receptors and activate cytoplasmic downstream signal pathways to regulate released cytokines and chemokines [73]. However, dysfunctions are induced during induction and development, resulting in immunologic tolerance, which is closely related to the TME [74]. The professional antigen presenting cells (APCs), dendritic cells (DCs) can be stimulated to proliferation, differentiation and maturation by OGF and OGFr antagonist (Fig. 5), with larger cell bodies, more pseudopods, and up-regulation of membrane molecular CD40, CD80, CD86, MHC-II and secretion of IL-12p70, IL-23, IP-10, TNF- α , IFN- γ while down-regulation of DC function is via the TLR4/MyD88/NF- κ B molecular pathway [46]. OGF and OGFr antagonists can promote tumor association macrophages (TAMs) transition from M2 to M1 and production of IL-12, IL-6, IL-1 β , TNF- α , CD64, IL-12p40, MHC-II, NO and H₂O₂ [52,77,78]. In addition, OGF significantly activates lymphocyte subtypes of CD4 + T cells, CD8 + T cells (CTL) and natural killer cells (NK) while depressing negatively CD4 + CD25 + Tregs [79]. OGF decreases the number of CD4 + CD25 + Tregs as well as phosphorylation and nuclear translocation of Smad2/3 that participates in the TGF- β pathway [80]. In regard to CTL, OGF matures phenotype and competence via increasing FasL, CD28, Prf levels and cytotoxicity [81].

3.4. Additive antitumor potency

To compare the oncotherapeutic effect of OGFr antagonists, researchers have cultured SKOV-3 tumor with LDN, cisplatin, taxol and LDN plus cisplatin or taxol, respectively [82]. The number of tumor cells with LDN (10⁻⁵ mol/L) and taxol (10⁻¹⁰ mol/L) decreased by 61% and 31%, compared with LDN plus taxol (Fig. 7), respectively, while combinations of the two had identical tumor inhibition efficacy related to higher concentrations of taxol (10⁻⁹ mol/L). The number of cells within the cultures with LDN plus cisplatin were reduced 42% and 32% in contrast with LDN and cisplatin, respectively. A reduction in tumor duration has been observed in mice with subcutaneous transplanted SKOV-3 tumor that was 21–48% with LND, 21–54% with

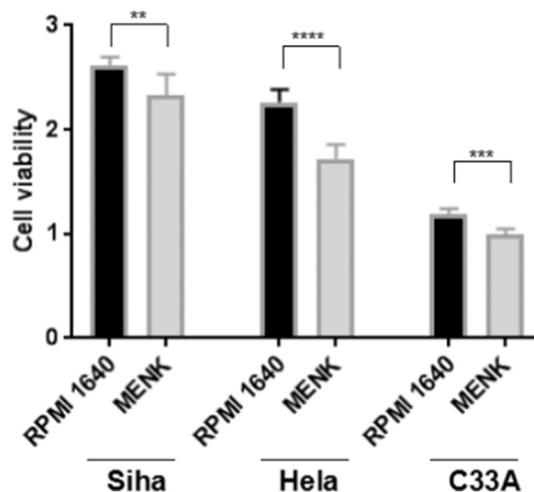


Fig. 6. Cell viability of cervical cancer cell lines after OGF exposure by MTS.

taxol, 24–54% with cisplatin, 39–60% with LDN plus taxol, and 16–60% with LDN and cisplatin, respectively. In addition, vascular density was depressed 52–73%, compared in single or combined reagent administration. OGFr presentation increased 46–61% in animals given LDN alone and PDL plus taxol or cisplatin. In clinical trials, patients who experienced chemotherapy failure could gain varying degrees of benefit ranging from delayed development to complete remission by LDN and chemotherapeutics [83–86]. At present, we suggest that OGF enhances the antitumor effect of molecular oncotargeting medicines in ovarian and cervical cancer cell lines. We conclude that there is clinical value via antitumor activity and reduced toxicities occur without affecting antitumor effect.

4. Prospection

The OGF - OGFr axis is involved in biochemical/hormonal reactions and in the maintenance of homeostasis. In the face of neoantigens or

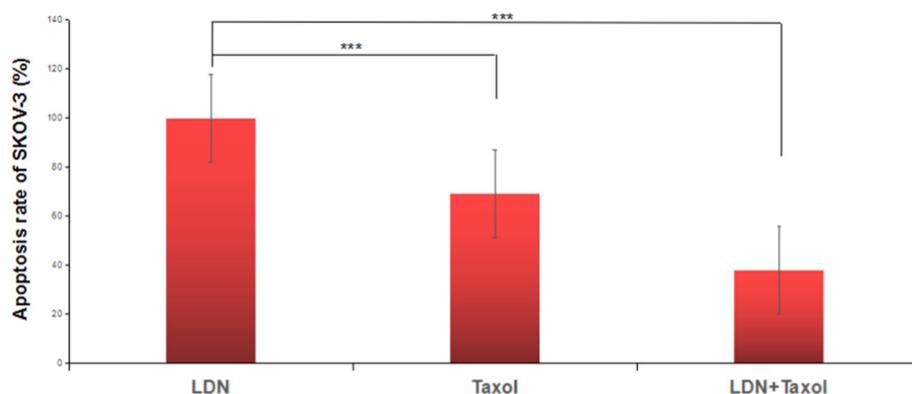


Fig. 7. The tumoricidal efficiency of LDN (10^{-6} mol/L) and taxol in vitro.

tissue toxicity, OGF or OGF α antagonist can activate natural immunity and specific killing to delete detrimental responses, while it averts an inflammatory over-response that may evoke further damage by immune cells and molecules via inhibitory cell multiplication or accelerated apoptosis, co-efficiently with neuro and endocrine system. Opioids have long been used for analgesia and chronic pain [87]. Limited by the low survival rate of traditional surgery, chemo- and radio-therapy against advanced cancers, intensive studies on immunologic escape and the TME search for a more suitable regimen for improving outcome [88]. There is increasing evidence that support an increase in associated toxicities with standard therapy [89]. Until now, the veil of OGF - OGF α axis on antitumor competence has been obscure, accounting for an essential part of immune modulation and tumoricidal activity. Future studies need to probe into tumor cytotoxicity and evaluate clinical effects of exogenous OGF and OGF α antagonists in gynecological cancer.

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References

- [1] S.A. Sullivan, E. Stringer, L. Van Le, A review of gynecologic oncology in the global setting: educating and training the next generation of women's health providers, *Obstet Gynecol Surv* 74 (1) (2019) 40–49.
- [2] F. Bray, J. Ferlay, I. Soerjomataram, R.L. Siegel, L.A. Torre, A. Jemal, Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries, *CA Cancer J. Clin.* 68 (6) (2018) 394–424.
- [3] P. Wreczycka-Cegielnny, T. Cegielnny, M. Oplawski, W. Sawicki, Z. Kojs, Current treatment options for advanced choriocarcinoma on the basis of own case and review of the literature, *Ginekol. Pol.* 89 (12) (2018) 711–715.
- [4] N. Arora, A. Talhouk, J.N. McAlpine, M.R. Law, G.E. Hanley, Causes of death among women with epithelial ovarian cancer by length of survival post-diagnosis: a population-based study in British Columbia, Canada, *Int. J. Gynecol. Cancer* 29 (3) (2019) 593–598.
- [5] H. Sahin, F.C. Sarioglu, M. Bagci, T. Karadeniz, H. Uluer, M. Sancı, Preoperative magnetic resonance volumetry in predicting myometrial invasion, lymphovascular space invasion, and tumor grade: is it valuable in International Federation of Gynecology and Obstetrics Stage I Endometrial Cancer? *Int. J. Gynecol. Cancer* 28 (4) (2018) 666–674.
- [6] A. Braga, P. Mora, A.C. de Melo, et al., Challenges in the diagnosis and treatment of gestational trophoblastic neoplasia worldwide, *World J Clin Oncol* 10 (2) (2019) 28–37.
- [7] S. Bellone, N. Buza, J. Choi, et al., Exceptional response to pembrolizumab in a metastatic, chemotherapy/radiation-resistant ovarian cancer patient harboring a PD-L1-genetic rearrangement, *Clin. Cancer Res.* 24 (14) (2018) 3282–3291.
- [8] R.W. Naumann, R.L. Coleman, J. Brown, K.N. Moore, Phase III trials in ovarian cancer: the evolving landscape of front line therapy, *Gynecol. Oncol.* 153 (2) (2019) 436–444.
- [9] M. Ramachandran, A. Dimberg, M. Essand, The cancer-immunity cycle as rational design for synthetic cancer drugs: novel DC vaccines and CAR T-cells, *Semin. Cancer Biol.* 45 (2017) 23–35.
- [10] J.M. Zaretsky, A. Garcia-Diaz, D.S. Shin, et al., Mutations associated with acquired resistance to PD-1 blockade in melanoma, *N. Engl. J. Med.* 375 (9) (2016) 819–829.
- [11] C. Johnson, A.A. Jazaeri, Diagnosis and management of immune checkpoint inhibitor-related toxicities in ovarian cancer: a series of case vignettes, *Clin. Ther.* 40

- (3) (2018) 389–394.
- [12] R. Anandakrishnan, R.T. Varghese, N.A. Kinney, H.R. Garner, Estimating the number of genetic mutations (hits) required for carcinogenesis based on the distribution of somatic mutations, *PLoS Comput. Biol.* 15 (3) (2019) e1006881.
- [13] S. Lai, Cancer related fatigue and cancer cachexia are the consequence of endocrine failure caused by persistent stress, *Med. Hypotheses* 123 (2019) 60–62.
- [14] K. Suresh, K.R. Voong, B. Shankar, et al., Pneumonitis in non-small cell lung cancer patients receiving immune checkpoint immunotherapy: incidence and risk factors, *J. Thorac. Oncol.* 13 (12) (2018) 1930–1939.
- [15] M.G. Hristova, Neuroendocrine and immune disequilibrium as a probable link between metabolic syndrome and carcinogenesis, *Med. Hypotheses* 118 (2018) 1–5.
- [16] R. Sinova, J. Kudova, K. Nesporova, et al., Opioid receptors and opioid peptides in the cardiomyogenesis of mouse embryonic stem cells, *J. Cell. Physiol.* 234 (8) (2019) 13209–13219.
- [17] M. Ferdousi, D.P. Finn, Stress-induced modulation of pain: role of the endogenous opioid system, *Prog. Brain Res.* 239 (2018) 121–177.
- [18] J. Hughes, T.W. Smith, H.W. Kosterlitz, L.A. Fothergill, B.A. Morgan, H.R. Morris, Identification of two related pentapeptides from the brain with potent opiate agonist activity, *Nature* 258 (5536) (1975) 577–580.
- [19] S.J. Watson, H. Akil, Recent studies on dynorphin and enkephalin precursor fragments in central nervous system, *Adv. Biochem. Psychopharmacol.* 33 (1982) 35–42.
- [20] P.J. McLaughlin, J.D. Wylie, G. Bloom, J.W. Griffith, I.S. Zagon, Chronic exposure to the opioid growth factor, [Met5]-enkephalin, during pregnancy: maternal and preweaning effects, *Pharmacol. Biochem. Behav.* 71 (1–2) (2002) 171–181.
- [21] I.S. Zagon, Y. Wu, P.J. McLaughlin, The opioid growth factor, [Met5]-enkephalin, and the zeta opioid receptor are present in human and mouse skin and tonically act to inhibit DNA synthesis in the epidermis, *J. Invest. Dermatol.* 106 (3) (1996) 490–497.
- [22] I.S. Zagon, J.W. Sassani, P.J. McLaughlin, Re-epithelialization of the rat cornea is accelerated by blockade of opioid receptors, *Brain Res.* 798 (1–2) (1998) 254–260.
- [23] I.S. Zagon, M.F. Verderame, S.S. Allen, P.J. McLaughlin, Cloning, sequencing, chromosomal location, and function of cDNAs encoding an opioid growth factor receptor (OGFr) in humans, *Brain Res.* 856 (1–2) (2000) 75–83.
- [24] I.S. Zagon, S.R. Goodman, P.J. McLaughlin, Demonstration and characterization of zeta (zeta), a growth-related opioid receptor, in a neuroblastoma cell line, *Brain Res.* 511 (2) (1990) 181–186.
- [25] I.S. Zagon, T.B. Ruth, P.J. McLaughlin, Nucleocytoplasmic distribution of opioid growth factor and its receptor in tongue epithelium, *Anat Rec A Discov Mol Cell Evol Biol* 282 (1) (2005) 24–37.
- [26] H. Yamashita, L. Shuman, J.I. Warrick, J.D. Raman, D.J. Degraff, Androgen represses opioid growth factor receptor (OGFR) in human prostate cancer LNCaP cells and OGFR expression in human prostate cancer tissue, *Am J Clin Exp Urol* 6 (4) (2018) 164–171.
- [27] J. Wan, Z. Qiu, Y. Ding, S. Nan, M. Ding, The expressing patterns of opioid peptides, anti-opioid peptides and their receptors in the central nervous system are involved in electroacupuncture tolerance in goats, *Front. Neurosci.* 12 (2018) 902.
- [28] D.A. Aykan, M. Kesim, B. Ayan, A. Kurt, Anti-inflammatory and antinociceptive activities of glucagon-like peptides: evaluation of their actions on serotonergic, nitric, and opioidergic systems, *Psychopharmacology* 236 (6) (2019) 1717–1728.
- [29] I.S. Zagon, R.E. Rhodes, P.J. McLaughlin, Localization of enkephalin immunoreactivity in diverse tissues and cells of the developing and adult rat, *Cell Tissue Res.* 246 (3) (1986) 561–565.
- [30] I.S. Zagon, M.F. Verderame, W.E. Zimmer, P.J. McLaughlin, Molecular characterization and distribution of the opioid growth factor receptor (OGFr) in mouse, *Brain Res. Mol. Brain Res.* 84 (1–2) (2000) 106–114.
- [31] K. Kido, Y. Shindo, S. Toda, E. Masaki, Expression of beta-endorphin in peripheral tissues after systemic administration of lipopolysaccharide as a model of endotoxic shock in mice, *Ann Endocrinol (Paris)* 80 (2) (2019) 117–121.
- [32] M.D. Ludwig, I.S. Zagon, P.J. McLaughlin, Featured article: serum [Met(5)]-enkephalin levels are reduced in multiple sclerosis and restored by low-dose naloxone, *Exp. Biol. Med.* (Maywood) 242 (15) (2017) 1524–1533.
- [33] Z. Oaks, A. Stage, B. Middleton, S. Faraone, B. Johnson, Clinical utility of the cold

- pressor test: evaluation of pain patients, treatment of opioid-induced hyperalgesia and fibromyalgia with low dose naltrexone, *Discov. Med.* 26 (144) (2018) 197–206.
- [34] M.D. Ludwig, I.S. Zagon, P.J. McLaughlin, Featured article: modulation of the OGF-OGFr pathway alters cytokine profiles in experimental autoimmune encephalomyelitis and multiple sclerosis, *Exp. Biol. Med.* (Maywood) 243 (4) (2018) 361–369.
- [35] M.D. Ludwig, I.S. Zagon, P.J. McLaughlin, Elevated serum [met(5)]-enkephalin levels correlate with improved clinical and behavioral outcomes in experimental autoimmune encephalomyelitis, *Brain Res. Bull.* 134 (2017) 1–9.
- [36] P. Kanagala, I.B. Squire, D.J.L. Jones, et al., Proenkephalin and prognosis in heart failure with preserved ejection fraction: a GREAT network study, *Clin. Res. Cardiol.* 108 (8) (2019) 940–949.
- [37] S.K. Panigrahi, K. Meece, S.L. Wardlaw, Effects of naltrexone on energy balance and hypothalamic melanocortin peptides in male mice fed a high-fat diet, *J Endocr Soc* 3 (3) (2019) 590–601.
- [38] M.M. Dull, A.E. Kremer, Management of Chronic Hepatic Itch, *Dermatol. Clin.* 36 (3) (2018) 293–300.
- [39] M. Carey, R. Gospin, A. Goyal, et al., Opioid receptor activation impairs hypoglycemic counterregulation in humans, *Diabetes* 66 (11) (2017) 2764–2773.
- [40] D.M. Wang, G.C. Wang, J. Yang, et al., Inhibition of the growth of human melanoma cells by methionine enkephalin, *Mol. Med. Rep.* 14 (6) (2016) 5521–5527.
- [41] I.S. Zagon, N.K. Porterfield, P.J. McLaughlin, Opioid growth factor - opioid growth factor receptor axis inhibits proliferation of triple negative breast cancer, *Exp. Biol. Med.* (Maywood) 238 (6) (2013) 589–599.
- [42] I.S. Zagon, P.J. McLaughlin, Opioid growth factor and the treatment of human pancreatic cancer: a review, *World J. Gastroenterol.* 20 (9) (2014) 2218–2223.
- [43] I.S. Zagon, R.N. Donahue, P.J. McLaughlin, Opioid growth factor-opioid growth factor receptor axis is a physiological determinant of cell proliferation in diverse human cancers, *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 297 (4) (2009) R1154–R1161.
- [44] F. Petraglia, F. Facchinetti, K. M'Futa, et al., Endogenous opioid peptides in uterine fluid, *Fertil. Steril.* 46 (2) (1986) 247–251.
- [45] P.J. McLaughlin, D.P. McHugh, M.J. Magister, I.S. Zagon, Endogenous opioid inhibition of proliferation of T and B cell subpopulations in response to immunization for experimental autoimmune encephalomyelitis, *BMC Immunol.* 16 (2015) 24.
- [46] Y. Meng, X. Gao, W. Chen, et al., Methionine enkephalin (MENK) mounts antitumor effect via regulating dendritic cells (DCs), *Int. Immunopharmacol.* 44 (2017) 61–71.
- [47] R.N. Donahue, P.J. McLaughlin, I.S. Zagon, Cell proliferation of human ovarian cancer is regulated by the opioid growth factor-opioid growth factor receptor axis, *Am J Physiol Regul Integr Comp Physiol* 296 (6) (2009) R1716–R1725.
- [48] R.N. Donahue, P.J. McLaughlin, I.S. Zagon, Under-expression of the opioid growth factor receptor promotes progression of human ovarian cancer, *Exp. Biol. Med.* (Maywood) 237 (2) (2012) 167–177.
- [49] F. Cheng, P.J. McLaughlin, M.F. Verderame, I.S. Zagon, The OGF-OGFr axis utilizes the p16INK4a and p21WAF1/CIP1 pathways to restrict normal cell proliferation, *Mol. Biol. Cell* 20 (1) (2009) 319–327.
- [50] F. Cheng, P.J. McLaughlin, M.F. Verderame, I.S. Zagon, The OGF-OGFr axis utilizes the p21 pathway to restrict progression of human pancreatic cancer, *Mol. Cancer* 7 (2008) 5.
- [51] Y. Kikuchi, T. Kita, M. Miyauchi, I. Iwano, K. Kato, Inhibition of human ovarian cancer cell proliferation in vitro by neuroendocrine hormones, *Gynecol. Oncol.* 32 (1) (1989) 60–64.
- [52] X. Wang, X. Jiao, Y. Meng, et al., Methionine enkephalin (MENK) inhibits human gastric cancer through regulating tumor associated macrophages (TAMs) and PI3K/AKT/mTOR signaling pathway inside cancer cells, *Int. Immunopharmacol.* 65 (2018) 312–322.
- [53] F.E. Lennon, T. Mirzapioazova, B. Mambetsariev, R. Salgia, J. Moss, P.A. Singleton, Overexpression of the mu-opioid receptor in human non-small cell lung cancer promotes Akt and mTOR activation, tumor growth, and metastasis, *Anesthesiology* 116 (4) (2012) 857–867.
- [54] I.S. Zagon, M.F. Verderame, P.J. McLaughlin, The biology of the opioid growth factor receptor (OGFr), *Brain Res. Brain Res. Rev.* 38 (3) (2002) 351–376.
- [55] E. Carrasco-Garcia, J. Auzmendi-Iriarte, A. Matheu, Integrin alpha7: a novel promising target in glioblastoma stem cells, *Stem Cell Investig* 5 (2018) 2.
- [56] D.Y. Oh, K.H. Jung, B.H. Yang, J.S. Lee, I.G. Choi, Y.G. Chai, Naltrexone influences protein kinase C and integrin alpha7 activity in SH-SY5Y neuroblastoma cells, *Exp. Mol. Med.* 38 (1) (2006) 100–106.
- [57] Q. Wu, X. Chen, J. Wang, et al., Nalmefene attenuates malignant potential in colorectal cancer cell via inhibition of opioid receptor, *Acta Biochim. Biophys. Sin. Shanghai* 50 (2) (2018) 156–163.
- [58] R.N. Donahue, P.J. McLaughlin, I.S. Zagon, The opioid growth factor (OGF) and low dose naltrexone (LDN) suppress human ovarian cancer progression in mice, *Gynecol. Oncol.* 122 (2) (2011) 382–388.
- [59] J. Fanning, C.A. Hossler, J.P. Kesterson, R.N. Donahue, P.J. McLaughlin, I.S. Zagon, Expression of the opioid growth factor-opioid growth factor receptor axis in human ovarian cancer, *Gynecol. Oncol.* 124 (2) (2012) 319–324.
- [60] F. Cheng, I.S. Zagon, M.F. Verderame, P.J. McLaughlin, The opioid growth factor (OGF)-OGF receptor axis uses the p16 pathway to inhibit head and neck cancer, *Cancer Res.* 67 (21) (2007) 10511–10518.
- [61] P.J. McLaughlin, B.C. Stack Jr., R.J. Levin, F. Fedok, I.S. Zagon, Defects in the opioid growth factor receptor in human squamous cell carcinoma of the head and neck, *Cancer* 97 (7) (2003) 1701–1710.
- [62] E. Chatzaki, A. Makrigiannakis, A.N. Margioris, E. Kouimtzoğlu, A. Gravanis, The Fas/FasL apoptotic pathway is involved in kappa-opioid-induced apoptosis of human endometrial stromal cells, *Mol. Hum. Reprod.* 7 (9) (2001) 867–874.
- [63] W.C. Lu, H. Xie, X.X. Tie, R. Wang, A.H. Wu, F.P. Shan, NFAT-1 hyper-activation by methionine enkephalin (MENK) significantly induces cell apoptosis of rats C6 glioma in vivo and in vitro, *Int. Immunopharmacol.* 56 (2018) 1–8.
- [64] Z. Vertes, J.L. Kornyei, S. Kovacs, M. Vertes, Opioids regulate cell proliferation in the developing rat uterus: effects during the period of sexual maturation, *J. Steroid Biochem. Mol. Biol.* 59 (2) (1996) 173–178.
- [65] A. Faletti, D. Bassi, A.L. Gimeno, M.A. Gimeno, Effects of beta-endorphin on spontaneous uterine contractions. Prostaglandins production and 45Ca^{2+} uptake in uterine strips from ovariectomized rats, *Prostaglandins Leukot Essent Fatty Acids* 47 (1) (1992) 29–33.
- [66] M. Aceves, M.N. Termino, A. Okoreeh, et al., Morphine increases macrophages at the lesion site following spinal cord injury: protective effects of minocycline, *Brain Behav. Immun.* 79 (2019) 125–138.
- [67] K.J. Levins, S. Prendeville, S. Conlon, D.J. Buggy, The effect of anesthetic technique on micro-opioid receptor expression and immune cell infiltration in breast cancer, *J. Anesth.* 32 (6) (2018) 792–796.
- [68] L.A. Hammer, H. Waldner, I.S. Zagon, P.J. McLaughlin, Opioid growth factor and low-dose naltrexone impair central nervous system infiltration by CD4 + T lymphocytes in established experimental autoimmune encephalomyelitis, a model of multiple sclerosis, *Exp. Biol. Med.* (Maywood) 241 (1) (2016) 71–78.
- [69] D.M. Wang, X. Jiao, N.P. Plotnikoff, et al., Killing effect of methionine enkephalin on melanoma in vivo and in vitro, *Oncol. Rep.* 38 (4) (2017) 2132–2140.
- [70] P. Dimitrova, K. Alipieva, K. Stojanov, V. Milanova, M.I. Georgiev, Plant-derived verbascoside and isoverbasoside regulate toll-like receptor 2 and 4-driven neutrophils priming and activation, *Phytomedicine* 55 (2018) 105–118.
- [71] Q. Zeng, C.M. Jewell, Directing toll-like receptor signaling in macrophages to enhance tumor immunotherapy, *Curr. Opin. Biotechnol.* 60 (2019) 138–145.
- [72] J. Tian, X. Jiao, X. Wang, et al., Novel effect of methionine enkephalin against influenza A virus infection through inhibiting TLR7-MyD88-TRAF6-NF-kappaB p65 signaling pathway, *Int. Immunopharmacol.* 55 (2018) 38–48.
- [73] N.J. Gay, Role of self-organising myddosome oligomers in inflammatory signalling by toll-like receptors, *BMC Biol.* 17 (1) (2019) 15.
- [74] H. Seo, I. Jeon, B.S. Kim, et al., IL-21-mediated reversal of NK cell exhaustion facilitates anti-tumour immunity in MHC class I-deficient tumours, *Nat. Commun.* 8 (2017) 15776.
- [75] Y. Meng, Q. Wang, Z. Zhang, E. Wang, N.P. Plotnikoff, F. Shan, Synergistic effect of methionine enkephalin (MENK) combined with pidotimod (PTD) on the maturation of murine dendritic cells (DCs), *Hum. Vaccin. Immunother.* 9 (4) (2013) 773–783.
- [76] J. Meng, Y. Meng, N.P. Plotnikoff, G. Youkilis, N. Griffin, F. Shan, Low dose naltrexone (LDN) enhances maturation of bone marrow dendritic cells (BMDCs), *Int. Immunopharmacol.* 17 (4) (2013) 1084–1089.
- [77] Z. Yi, S. Guo, X. Hu, et al., Functional modulation on macrophage by low dose naltrexone (LDN), *Int. Immunopharmacol.* 39 (2016) 397–402.
- [78] W. Chen, J. Liu, J. Meng, et al., Macrophage polarization induced by neuropeptide methionine enkephalin (MENK) promotes tumoricidal responses, *Cancer Immunol. Immunother.* 61 (10) (2012) 1755–1768.
- [79] Q. Wang, X. Gao, Z. Yuan, et al., Methionine enkephalin (MENK) improves lymphocyte subpopulations in human peripheral blood of 50 cancer patients by inhibiting regulatory T cells (Tregs), *Hum Vaccin Immunother* 10 (7) (2014) 1836–1840.
- [80] X. Li, Y. Meng, N.P. Plotnikoff, et al., Methionine enkephalin (MENK) inhibits tumor growth through regulating CD4+ Foxp3+ regulatory T cells (Tregs) in mice, *Cancer Biol. Ther.* 16 (3) (2015) 450–459.
- [81] W. Li, J. Meng, X. Li, et al., Methionine enkephalin (MENK) improved the functions of bone marrow-derived dendritic cells (BMDCs) loaded with antigen, *Hum Vaccin Immunother* 8 (9) (2012) 1236–1242.
- [82] R.N. Donahue, P.J. McLaughlin, I.S. Zagon, Low-dose naltrexone suppresses ovarian cancer and exhibits enhanced inhibition in combination with cisplatin, *Exp. Biol. Med.* (Maywood) 236 (7) (2011) 883–895.
- [83] L. Schwartz, L. Buhler, P. Icard, H. Lincet, J.M. Steyaert, Metabolic treatment of cancer: intermediate results of a prospective case series, *Anticancer Res.* 34 (2) (2014) 973–980.
- [84] B.M. Berkson, D.M. Rubin, A.J. Berkson, Reversal of signs and symptoms of a B-cell lymphoma in a patient using only low-dose naltrexone, *Integr Cancer Ther* 6 (3) (2007) 293–296.
- [85] M. Rogosnitzky, M.J. Finegold, P.J. McLaughlin, I.S. Zagon, Opioid growth factor (OGF) for hepatoblastoma: a novel non-toxic treatment, *Investig. New Drugs* 31 (4) (2013) 1066–1070.
- [86] A. Khan, Long-term remission of adenoid cystic tongue carcinoma with low dose naltrexone and vitamin D3—a case report, *Oral Health Dent Manag* 13 (3) (2014) 721–724.
- [87] C. Lefkowitz, M.K. Buss, A.A. Ramzan, et al., Opioid use in gynecologic oncology in the age of the opioid epidemic: part I - effective opioid use across clinical settings, a society of gynecologic oncology evidence-based review, *Gynecol. Oncol.* 149 (2) (2018) 394–400.
- [88] R.C. Bast Jr., U.A. Matulonis, A.K. Sood, et al., Critical questions in ovarian cancer research and treatment: report of an American Association for cancer research special conference, *Cancer* 125 (12) (2019) 1963–1972.
- [89] N.F. Pistamaltzian, V. Georgoulas, A. Kotsakis, The role of immune checkpoint inhibitors in advanced non-small cell lung cancer, *Expert Rev. Respir. Med.* 13 (5) (2019) 435–447.